

Calculating the Minimum Inhibitory Concentration (MIC) of Gentamycin on Staphylococcus epidermidis Grown Under Simulated Microgravity Sofia Saldarriaga, Janelle Hicks, Collin Topolski, Hugo Castillo Department of Human Factors and Behavioral Neurobiology, Embry-Riddle Aeronautical University, Daytona Beach FL 32114

Introduction

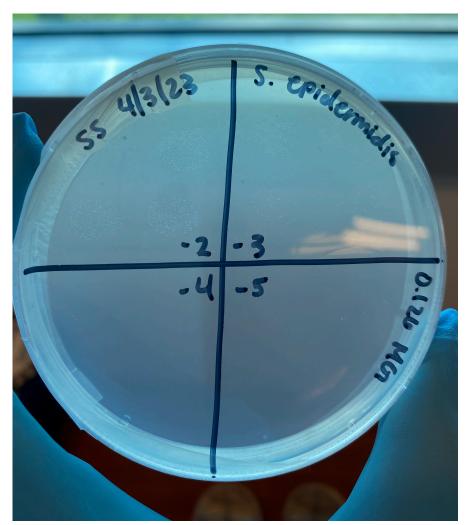
- With space travel becoming more prominent, scientists have been searching for ways to allow for a thriving environment while on long-duration space missions.
- One of the ways researchers are investigating ways to maintain optimal health in astronauts is through antibiotic resistance.
- astronaut's immune system functions differently in ■ An microgravity than it does on Earth due to metabolic changes.
- Therefore, a certain dosage of an antibiotic may used to treat a certain infection may be affected in these conditions.
- This study aims to determine the MIC (minimum inhibitory) concentration) of Gentamicin and measure its effect on S. epidermidis under simulated microgravity.

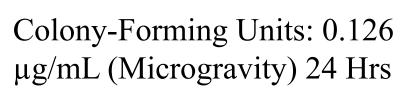
Experimental Design

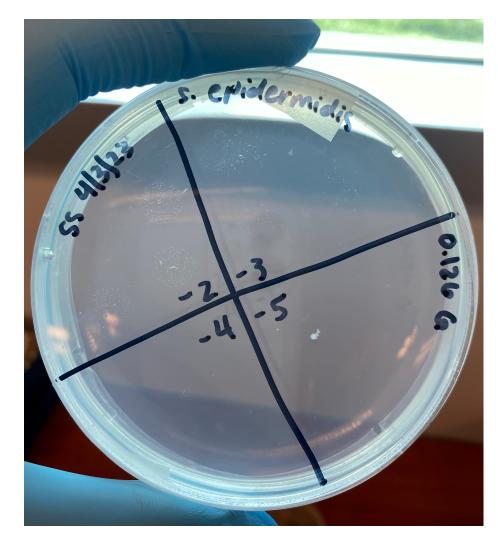
- epidermidis was cultured from ■ S. glycerol stocks for short term exposure experiments.
- S. epidermidis was cultured in Nutrient Broth (NB) media overnight.
- These cultures were then grown in epitubes on the EAGLESTAT within different antibiotic concentrations.
- After growth occurred, the cultures were transferred to a 94well plate and their optical density was analyzed. Drop plates were also made to count colonies.
- Data from this growth was analyzed and used to decide which antibiotic concentration was significant enough to use for the growth curve.

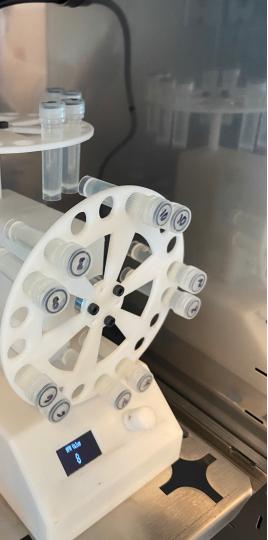
Growth Curve Procedure

- S. epidermidis was grown in a 24 well plate with concentrations of Gentamicin ranging from 0.5 to 0.126 μg/ml.
- Plate was placed on a shaker and incubated at 30°C.
- Absorbance was read on a spectrophotometer to observe the changes in biomass over a period of 8 hours.
- Dilutions were made to measure numbers of colonies at a countable quantity that were able to grow after being exposed to the antibiotic and microgravity.
- Colony-forming units were obtained by counting the number of colonies that were grown on the plates.



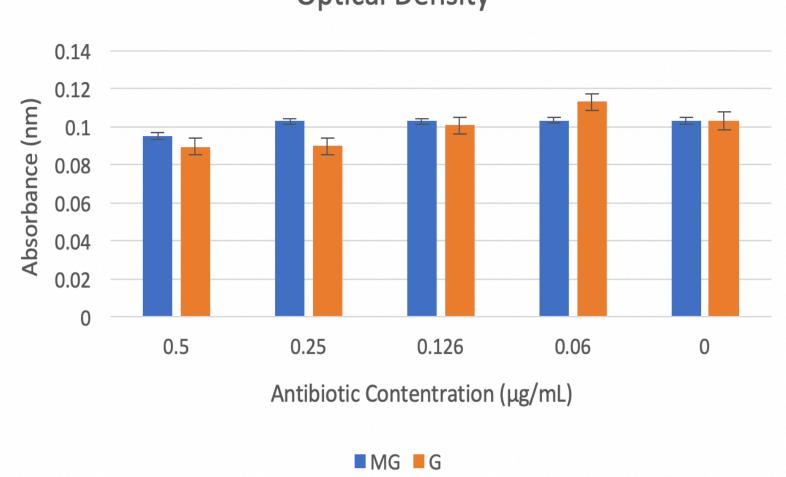






Optical Density

- epidermidis S. was grown in 2 mL epitubes on the EAGLESTAT and incubated for 30 hours.
- Epitubes had range of antibiotic concentrations.



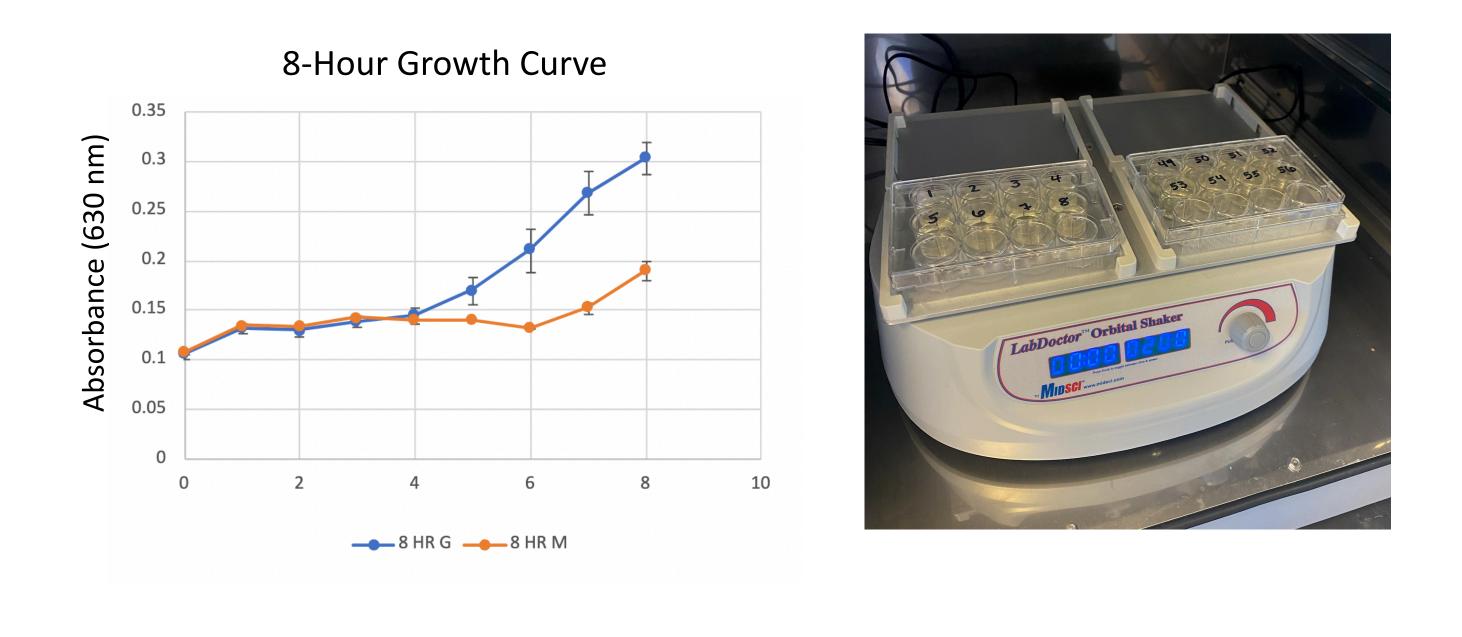
• After the 30-hr incubation period, absorbance was read at 630 nm on the spectrophotometer to determine biomass.

Colony-Forming Units: 0.126 µg/mL (Gravity) 24 Hrs



Optical Density

- time points.



References: Tirumalai, M. R., Karouia, F., Tran, Q., Stepanov, V. G., Bruce, R. J., Ott, C. M., ... & Fox, G. E. (2019). Evaluation of acquired antibiotic resistance in Escherichia coli exposed to long-term low-shear modeled microgravity and background antibiotic exposure. Mbio, 10(1), e02637-18.

Cira, N. J., Ho, J. Y., Dueck, M. E., & Weibel, D. B. (2012). A self-loading microfluidic device for determining the minimum inhibitory concentration of antibiotics. Lab on a Chip, 12(6), 1052-1059.

Credito, K., Lin, G., & Appelbaum, P. C. (2007). Activity of daptomycin alone and in combination with rifampin and gentamicin against Staphylococcus aureus assessed by time-kill methodology. Antimicrobial agents and chemotherapy, 51(4), 1504-1507.





Growth Curve

Samples were collected to test their viability at hourly

These samples were incubated for 8 hours and their absorbance was measured every hour.

Results indicate a major difference between the two time points, with gravity being more viable at 8 hours and microgravity being more viable at 24 hours.

Future Research

Expand screening of antibiotics and their concentrations.

Expand the time period of the growth curve.

Observe growth of post-antibiotic exposure.

Observe antibiotic mechanisms of action.

Study changes in gene expression of cultures.