

MICROSPOROGENESIS, MICROGAMETOGENESIS, AND POLLEN GRAIN MORPHOLOGY
OF *FOUQUIERIA FASCICULATA* (FOUQUIERIACEAE, ERICALES)
MICROSPOROGÉNESIS, MICROGAMETOGÉNESIS Y MORFOLOGÍA DEL GRANO DE POLEN
DE *FOUQUIERIA FASCICULATA* (FOUQUIERIACEAE, ERICALES)

FABIOLA SOTO-TREJO^{1*}, SERGIO ZAMUDIO-RUIZ² AND SILVIA ESPINOSA-MATÍAS³

¹ Unidad de Biotecnología y Prototipos, Facultad de Estudios Superiores Iztacala, Universidad Nacional Autónoma de México, Tlalnepantla, Estado de México, Mexico.

² Investigador independiente, Pátzcuaro, Michoacán, México.

³ Laboratorio de Microscopía Electrónica de Barrido, Facultad de Ciencias, Universidad Nacional Autónoma de México, Ciudad de México, Mexico.

*Author for correspondence: fabiolasototrejo@gmail.com

Abstract

Background: Embryology of the male reproductive structures in Fouquieriaceae has been poorly studied. The ontogeny of the anther wall, microsporogenesis, microgametogenesis, and pollen morphology were described in detail for *Fouquieria fasciculata*.

Questions: How microsporogenesis and microgametogenesis are carried out in *Fouquieria fasciculata*? How is the morphology of mature pollen grain?

Study species: *Fouquieria fasciculata* (Fouquieriaceae).

Study site and date: Río Estórax Canyon, near the El Plátano, Querétaro, México.

Methods: Floral buds and flowers at different stages of development were collected and processed for further analysis with light and scanning electron microscopy.

Results: The development of the anther wall is of Dicotyledonous type. The anther wall consists of a single-layered epidermis covered by a thick cuticle, an endothecium with U-shaped fibrous thickenings, one middle layer and a secretory tapetum with uni or binucleate cell. Two orbicules types were characterized: doughnut-shape orbicules and aggregated orbicules with ruminant appearance. Microsporogenesis is successive, the microspore tetrads are tetrahedral. Pollen grains are prolate, tricolporate with reticulate-heterobrochate exine, and are shed in the two-celled stage.

Conclusions: This research is the first comprehensive study of male structures ontogeny of Fouquieriaceae. Remarkable features were described in *Fouquieria fasciculata*, such as the differentiation of a secretory tapetum and the presence of orbicules, which are described for the first time and contribute to characterize the embryology of Fouquieriaceae.

Key words: male gametophyte, orbicules, pollen grain, secretory tapetum.

Resumen

Antecedentes: La embriología de las estructuras reproductivas masculinas en Fouquieriaceae ha sido poco estudiada. La ontogenia de la pared de la antera, la microsporogénesis, la microgametogénesis y la morfología del polen fueron descritas en detalle para *Fouquieria fasciculata*.

Preguntas: ¿Cómo se llevan a cabo los procesos de microsporogénesis y microgametogénesis en *Fouquieria fasciculata*? ¿Cómo es la morfología del grano de polen maduro?

Especie de estudio: *Fouquieria fasciculata* (Fouquieriaceae).

Sitio de estudio y fecha: Cañón del Río Estórax cerca de El Plátano, Querétaro, México.

Métodos: Botones florales y flores fueron recolectadas, procesadas y analizadas con microscopía de luz y microscopía electrónica de barrido.

Resultados: El desarrollo de la pared de la antera es de tipo Dicotiledóneo. La pared de la antera consiste en una epidermis con cutícula gruesa, un endotecio con engrosamientos fibrosos en forma de U, una capa media y un tapete secretor con células uni o binucleadas. Se caracterizaron dos tipos de orbículas: orbículas con forma de dona y orbículas agregadas con apariencia ruminada. La microsporogénesis es sucesiva, las tetradas de microsporas son tetraédricas. Los granos de polen son prolados, tricolporados con exina reticulada-heterobrocada y son dispersados en etapa bicelular.

Conclusiones: Esta investigación es el primer estudio embriológico detallado de las estructuras reproductivas masculinas en Fouquieriaceae. Características novedosas fueron descritas en *Fouquieria fasciculata*, tales como la diferenciación de un tapete secretor y la presencia de orbículas, descritas por primera vez y contribuyen a caracterizar la embriología de Fouquieriaceae.

Palabras clave: gametofito masculino, granos de polen, orbículas, tapete secretor.

This is an open access article distributed under the terms of the Creative Commons Attribution License CCBY-NC (4.0) international.

<https://creativecommons.org/licenses/by-nc/4.0/>



Fouquieriaceae (Ericales) is a small family of eleven species of trees and shrubs endemic to mostly dry environments of Mexico and southwestern United States (Henrickson 1972, Kubitzki 2004). A recent study undertaken by Soto-Trejo *et al.* (2022) suggested that the family was originated in dry habitats in central-southern Mexico during the Late Miocene. Fouquieriaceae present a series of anatomical adaptations to live in seasonal arid areas, such as presence of a water-storage tissue by means of expansion rays and axial parenchyma bands (Carlquist 2000). The flowers are usually pentamerous, actinomorphic and bisexual with petals fused into a tube; the androecium consists of approximately 10-23 stamens, and the gynoecium is syncarpous with three fused carpels (Henrickson 1972, Waser 1979, Schönenberger & Grenhagen 2005, Schönenberger 2009).

The first embryological studies in Fouquieriaceae were conducted by Johansen (1936), Mauritzon (1936), Khan (1943), Amlie (1965) and Govil (1970) who provided characters associated mainly with the development of ovules and seeds. Recently, Soto-Trejo *et al.* (2021) described in detail the development of the female gametophyte and the ovule in *Fouquieria fasciculata* (Roem. & Schult.) Nash, showing the differentiation of an integumentary tapetum, a *Polygonum*-type embryo sac and the formation of a lateral haustorial arm in the female gametophyte. The androecium is organized in two-level with all the stamens arranged in a single series (Henrickson 1972, Schönenberger & Grenhagen 2005). In a comprehensive study of the pollen morphology of the family, Henrickson (1967) showed tricolporate pollen grains and analyzed the exine sculpturing, showing that size variation correlates directly with flower size. However, features related to the anther development, microsporogenesis and microgametogenesis have not been described in detail or are incomplete for Fouquieriaceae (Davis 1966, Govil 1970, Johri *et al.* 1992).

The systematic importance of embryological characters, such as the tapetum type and the presence of orbicules (or Ubisch bodies), has been particularly studied in angiosperms (Huysmans *et al.* 1998, Vinckier *et al.* 2000, Vinckier & Smets 2002a, b, Janssens *et al.* 2005, Verstraete *et al.* 2014, Song *et al.* 2017, Moon 2018, Ruggiero & Bedini 2020, Shamrov *et al.* 2021). The tapetum is innermost layer of microsporangium wall and its cells often have dense cytoplasm with one or several nuclei (El-Ghazaly 1999, Shamrov *et al.* 2021). The tapetal cells provides protection and essential nutrients (*e.g.*, polysaccharides, enzymes, hormones, lipids) to pollen grain development and supplies precursors for pollen exine formation (El-Ghazaly 1999, Pacini 2010, Shamrov *et al.* 2021). Two main tapetum types are recognized: a) secretory type, where a layer of cells remains intact around the anther locule, and b) plasmodial type, where a multinucleate tapetal plasmodium is formed in the anther locule by fusion of tapetal protoplasts (Pacini *et al.* 1985, Pacini 1997, Furness & Rudall 1998, 2001, Shamrov *et al.* 2021). In angiosperms, a secretory tapetum has been associate with orbicules, although they are also formed from a plasmodial tapetum (Huysmans *et al.* 1998, El-Ghazaly 1999, Galati *et al.* 2007, Verstraete *et al.* 2014). The orbicules are sporopollenin particles that are found in the walls of the tapetal cells, which have been widely mentioned and described in species of the ANA grade, magnoliids, and monocots, varying in abundance, size, shape, and ornamentation (Vinckier & Smets 2002a, b, Verstraete *et al.* 2014, Ruggiero & Bedini 2020).

As mentioned previously, features related to the development of male structures have not been described in detail for Fouquieriaceae. Therefore, we study the development of the anther wall, microsporogenesis, microgametogenesis, and pollen grain morphology of *Fouquieria fasciculata*, to understand the embryology of the family. This species is a deciduous shrub restricted to the dry environments of the states of Hidalgo and Querétaro in central Mexico, and is characterized by a large fat swollen stem (Henrickson 1972, Zamudio 1995).

Materials and methods

Specimens of *Fouquieria fasciculata* were collected in the Río Estórax Canyon near the El Plátano, Querétaro, Mexico (21° 01' 22" N; 99° 30' 60" W) at the altitude of 910 m asl. Vouchers were deposited at the Herbarium IEB, Centro Regional del Bajío, Instituto de Ecología, A.C.

Floral buds and flowers in pre-anthesis, anthesis and post-anthesis were fixed *in situ* in FAA (10 % formaldehyde, 5 % glacial acetic acid, 50 % ethanol 96 %, and 35 % distilled water) for 24 hours and stored in 70 % ethanol (Johansen 1940). After that, samples were dehydrated in an ethanol series (70, 85, 96 and 100 %) and embedded in Paraplast

wax following Johansen (1940) and sectioned using an American Optical 810 rotary microtome. Further, samples were also embedded in LR-W resin (London resin white) following Ruzin (1999). Sections (~2 µm) were cut using an ultramicrotome RMC-MT990, stained with 0.05 % Toluidine Blue (O'Brien *et al.* 1964) and mounted in Entellan (Merck). The images were obtained using an Olympus Provis AX70 optical microscope.

Scanning electron microscopy was used to describe the morphology and development of anthers and mature pollen grain. Samples were dehydrated in a graded ethanol series and critical-point dried using a CPD 030 critical-point dryer (Bal-Tec AG, Liechtenstein). Later, the samples were mounted on aluminum stubs using carbon double-sided tape and coated with gold by means of a Denton Vacuum Desk II sputter coater (Denton Vacuum, Moorestown, NJ, USA) (Bozzola & Russell 1992). Then, samples were analyzed and photographed on a JEOL JSM5310-LV scanning electron microscope at 15 kV.

Results

Androecium. The androecium of *Fouquieria fasciculata* consist of 10 stamens (Figure 1) organized in a single series. In early stages, the stamen primordia become enlarged starting to differentiate into anther and filament (Figure 2A, B). Then, stamens are arranged at two levels due to differences in filament length (Figure 2C, D). In the flower at anthesis, the anthers are of equal sizes but are arranged at different levels due to unequal length of filaments. The filaments present simple and unicellular trichomes on the basal portion above at the level the ovary (Figure 2E, F). Anthers are oblong-lanceolate, cordate with two lobes at the base and cuspidate at the apex (Figure 2C, D, G). The mature anthers are bilobed, dorsifixed with introrse dehiscence along two lines (Figure 2G).



Figure 1. Flowers in anthesis of *Fouquieria fasciculata*.

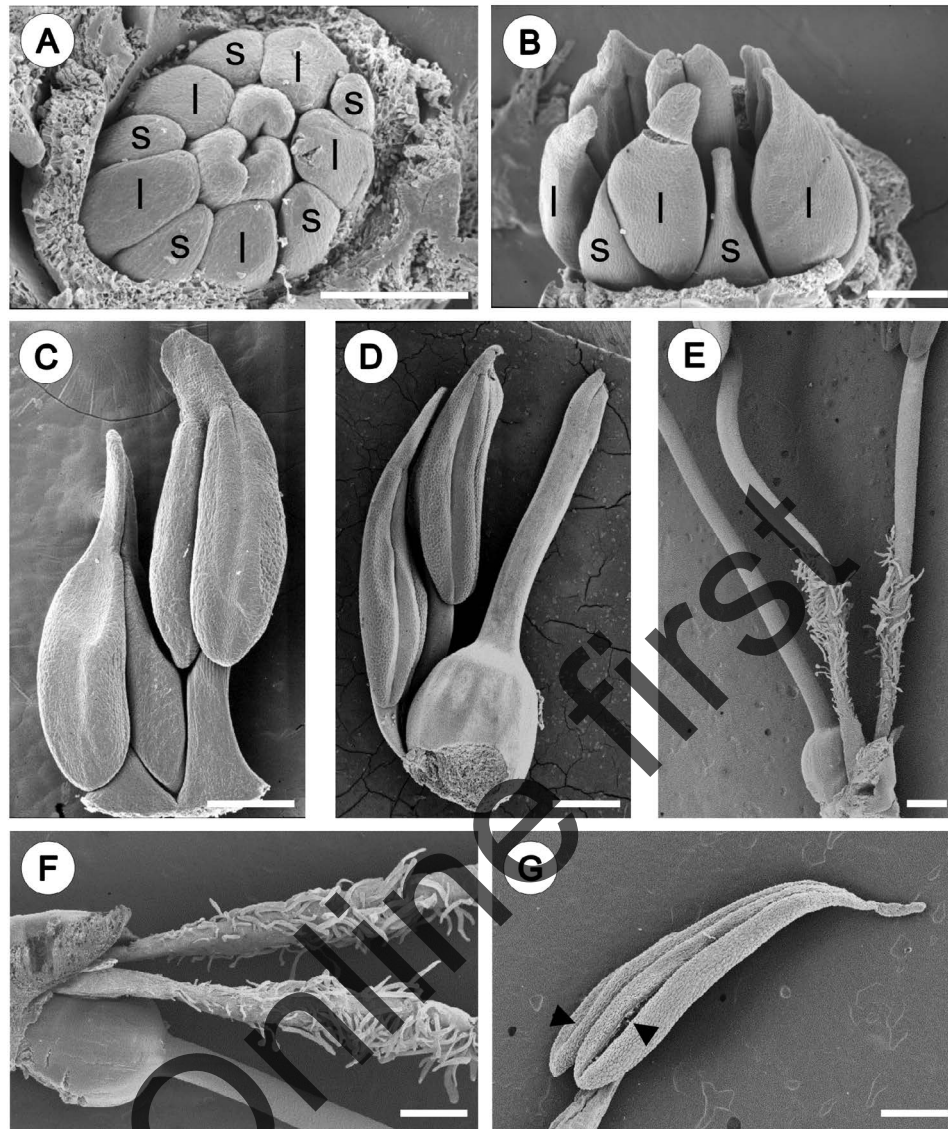


Figure 2. Androecium morphology of *Fouquieria fasciculata*. A-B. Young floral bud showing the stamens starting to differentiate into anther with five shorter filaments (s) and five longer filaments (l). C-D. Pre-anthesis flower, stamens are arranged at two levels due to differences in filament length. E. Flower at anthesis showing filaments with simple and unicellular trichomes on the basal portion. F. Close up of the trichomes. G. Mature anther showing two lines of dehiscence (black arrow heads). Scale bars = 200 μ m (A-D) and 500 μ m (E-G).

Development of the anther wall. The anther primordia develop as small hemispherical bumps surrounded by protodermis in transection (Figure 3A). Each anther primordium consisted of meristematic cells with a voluminous nucleus, the subepidermal layer increased by the number of divisions giving rise to the four anther lobes or sporangia (Figure 3B). The subepidermal cells of sporangia are larger (Figure 3B), and become differentiated into archesporial tissue, which characterize by isodiametric cells with dense cytoplasm and conspicuous nuclei (Figure 3C). Then, the archesporial cells are divided periclinally, originating a primary parietal layer and the sporogenous tissue. Periclinial divisions in the primary parietal layer generate two secondary parietal layers: external and internal (Figure 3D). The external secondary parietal layer was divided periclinally, giving rise to the endothecium and a middle layer, while the inner secondary parietal layer develops into the tapetum. Therefore, the anther wall consists of a single-layered epidermis covered by a thick cuticle, an endothecium with fibrous thickenings, one middle layer, and 1-2 nucleate

secretory tapetum (Figure 3E, Table 1). According to our observations, the development of the anther wall corresponds to the Dicotyledonous type. The mature anther is bithecal and tetrasporangiate, the four lobes are separated by connective tissue (Figure 3F).

Microsporogenesis and microgametogenesis. The sporogenous tissue consists of cells with conspicuous nuclei and dense cytoplasm (Figure 4A). Then, mitotic divisions give rise to microspore mother cells, which are large and spherical with a thickened callose wall (Figure 4B). The microspore mother cells undergo meiotic divisions and give rise to microspore tetrads with tetrahedral arrangement surrounded by a thick and uniform wall of callose (Figure 4C), and the tapetal cells are rectangular with thin walls, uni- or bi-nucleate (Figure 4D). Later, the callose walls of the tetrads are dissolved and the microspores are free in the anther locule, and the microspore cytoplasm presents a large central vacuole and the nucleus is displaced to a parietal position (Figure 4E). During this stage, tapetal cells increase considerably their volume and present extremely dense cytoplasm with prominent nuclei (Figure 4E). Furthermore, scanning electron micrographs of the radial and inner tangential surfaces of tapetal cells showed orbicules that vary in shape and size (Figure 4F). Two orbicules types were observed: a) doughnut-shape orbicules, where a single central perforation is present (Figure 4G), and b) aggregated orbicules, that are irregular bodies with grooves and wavy margin giving ruminant appearance (Figure 4H). The orbicules surface is granular and resemble the exine of the pollen grain.

In mature anthers, the endothecium develops thickenings on the radial and inner tangential wall, and the cells of the middle layer and tapetum show signs of degradation. Notably, the tapetal cell persists for a long time and disintegrates just before anther dehiscence. During anther dehiscence, the parenchymal septum that separates both locules of an anther theca breaks down, and consequently a single locule is formed (Figure 5A). The endothecium cells are rectangular and developed U-shaped fibrous thickenings on their radial and inner tangential walls (Figure 5B). Finally, microspores undergo a mitotic division, forming the generative cell and the vegetative cell; thus the pollen grains are shed in the two-celled stage (Figure 5C).

Morphology of the pollen grain. The mature pollen grain is prolate, tricolporate with reticulate-heterobrochate exine (Figure 5D-F).

Table 1. Embryological characters of *Fouquieria fasciculata*.

Characters	State
Number of sporangia per anther	Four
Type of anther wall development	Dicotyledonous
Epidermis	Persistent
Endothecium	Develops U-shaped fibrous thickenings
Middle layer	Not persistent
Tapetum	Secretory
Number of nuclei in a tapetal cell	One or two
Orbicules	Present
Orbicules morphology	Doughnut-shape, ruminant
Type of microspore tetrads	Tetrahedral
Number of cells in a mature pollen	Two

Discussion

In *Fouquieria fasciculata*, the androecium consists of ten stamens organized in a single series. During early stages, stamens are arranged at two levels due to differences in filament length. In flowers at anthesis and post-anthesis, the anthers are of equal sizes but are arranged at different levels. Our results are consistent with reported in *F. columnaris* (Kellogg) Kellogg ex Curran and *F. splendens* Engelm. by Schöenberger & Grenhagen (2005), who described in detail the androecium development.

The stamens present filaments with trichomes on basal portion above at the ovary level covering the nectary surface. According to Henrickson (1972), these trichomes reduce evaporation and present the nectar to flower visitors. Most of Fouquieriaceae species possess flowers with tubular red corollas pollinated by hummingbirds (Waser 1979, Schultheis & Baldwin 1999), thus the nectar could be obtained by these birds. But, *F. fasciculata* present white

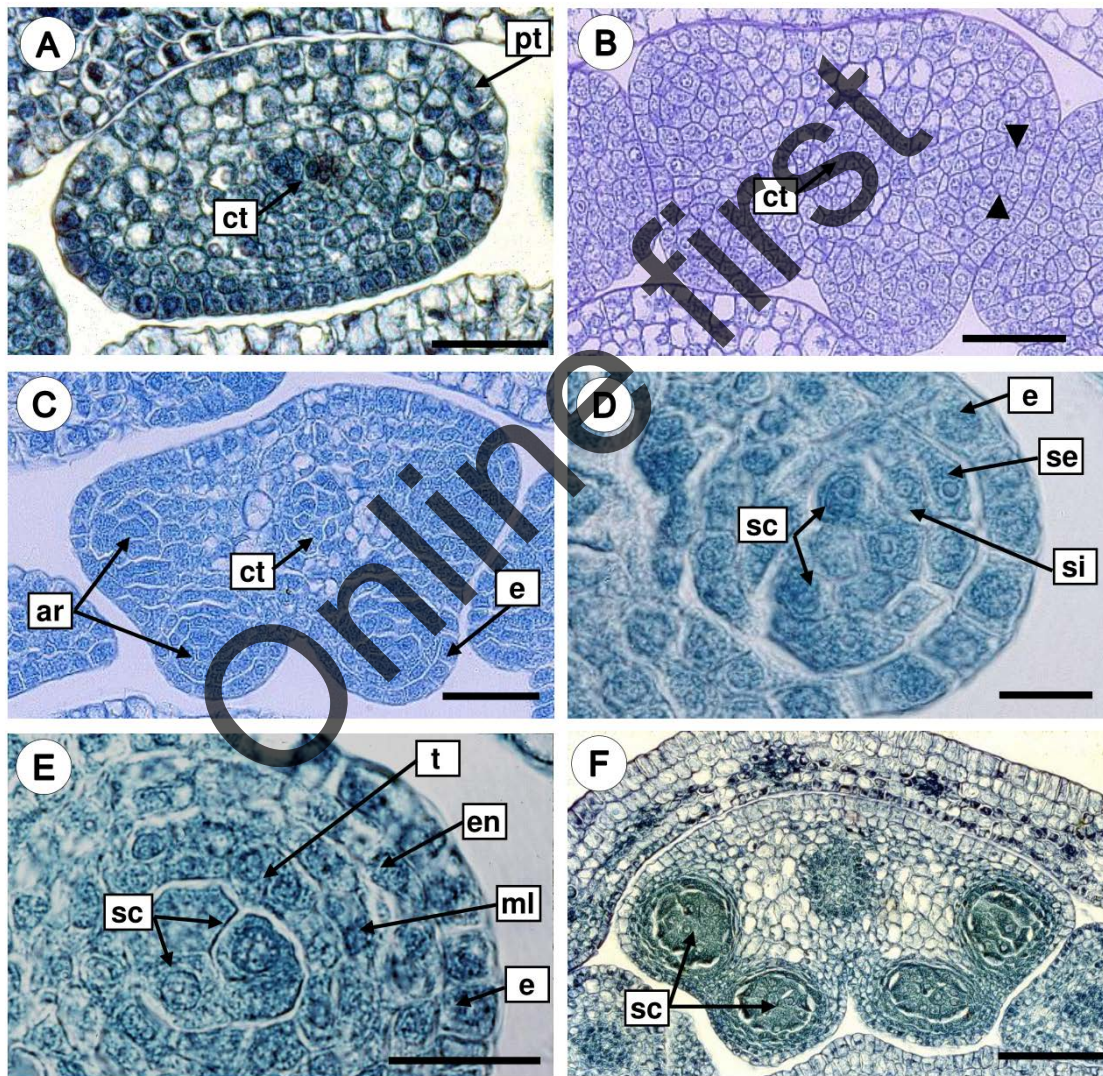


Figure 3. Development of the anther wall in *Fouquieria fasciculata*. A. Anther primordium surrounded by protodermis. B. Periclinal divisions in the subepidermal layer (black arrow heads) gave rise to the four anther lobes. C. Tetrasporangiate anther with arquesporial cells. D. Anther wall with external and internal secondary parietal layers and sporogenous tissue. E. Anther young wall with the four layers. F. Bilobate and tetrasporangiate anther with sporogenous cells. ar, arquesporial cells; ct, connective tissue; e, epidermis; en, endothecium; ml, middle layer; pt, protodermis; sc, sporogenous cells; se, secondary external parietal layer; si, secondary internal parietal layer; t, tapetum. Scale bars = 30 μ m (A-C), 10 μ m (D-E), 100 μ m (F).

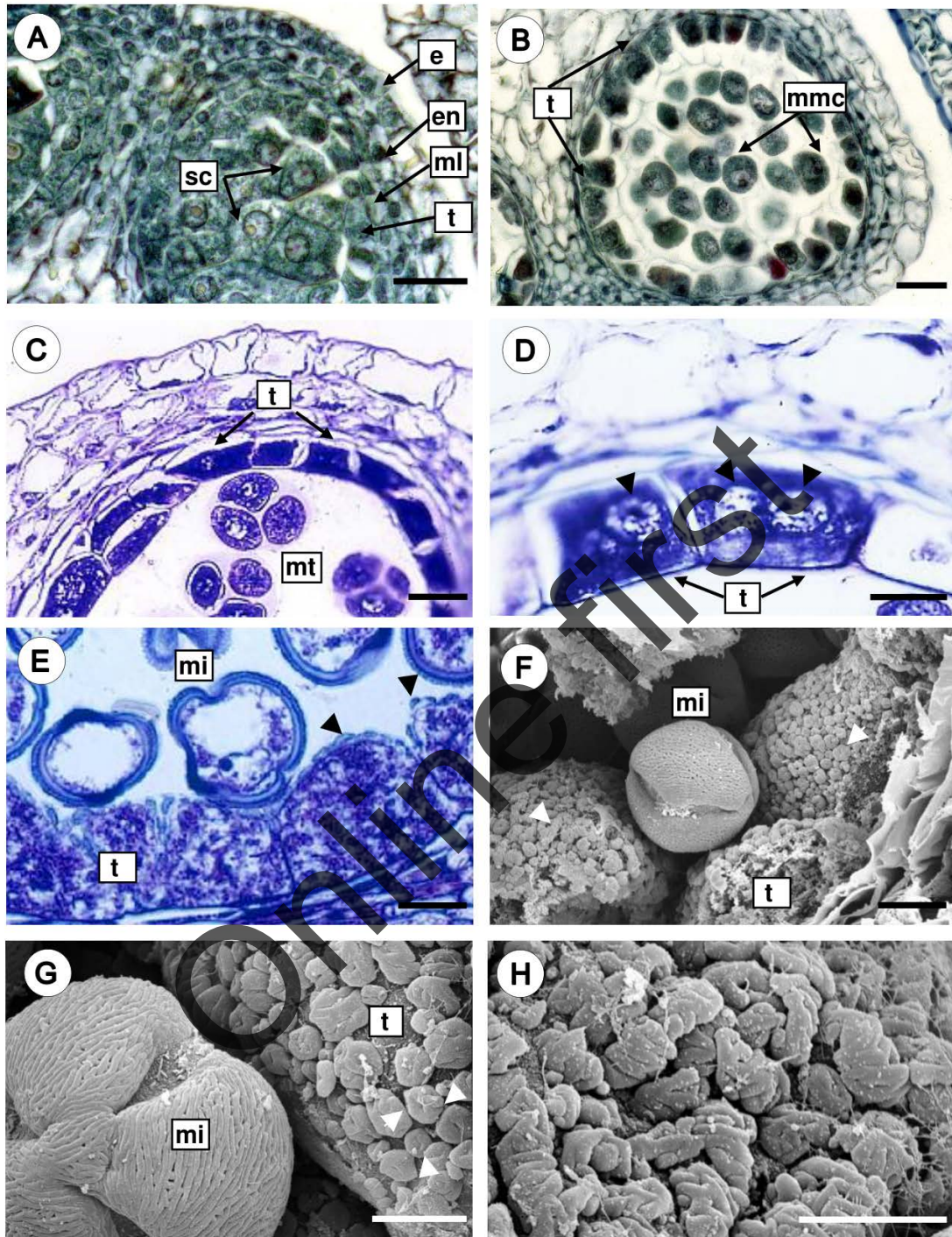


Figure 4. Microsporogenesis and microgametogenesis in *Fouquieria fasciculata*. A. Anther with sporogenous cells. B. Anther with microspore mother cells surrounded by the callose wall. C. Microspore tetrahedral tetrads still enclosed in callose wall. D. Tapetal cells uni- or bi-nucleate (black arrow heads) with dense cytoplasm. E-F. Free microspores with a conspicuous nucleus and a large central vacuole, and orbicules are observed on the tapetal cell walls (white and black arrow heads). G. Doughnut-shape orbicules (white arrow heads). H. Close up of the aggregate orbicules of ruminant appearance. e, epidermis; en, endothecium; mi, microspore; ml, middle layer; mmc, microspore mother cell; mt, microspore tetrahedral tetrads; sc, sporogenous cells; t, tapetum. Scale bars = 25 μm (A-D), 10 μm (E-F), 5 μm (G-H).

flowers and the pollinators are not known, thus more research is needed to determine the type of pollinators in this specie. Anther morphology is relatively uniform throughout the family Fouquieriaceae (Henrickson 1972). Anthers are oblong-lanceolate, cordate with two lobes at the base and cuspidate apex. Further, the mature anthers are tetrasporangiate, dorsifixed with introrse dehiscence along two lines, features that coincide with what has been observed by Henrickson (1972) and Johri *et al.* (1992).

Anther wall development is of Dicotyledonous type formed by the epidermis, fibrous endothecium, one middle layer, and a secretory tapetum with uni-binucleate cells as described by Davis (1966) for Fouquieriaceae. During the anther wall development, the tapetum presents the most important changes. Our results showed that during free microspore stage, the tapetal cells increase considerably their physiological-biochemical activity for developing microspores, pollen grains and orbicules; thus, the tapetum persist for a long time during of pollen grains forma-

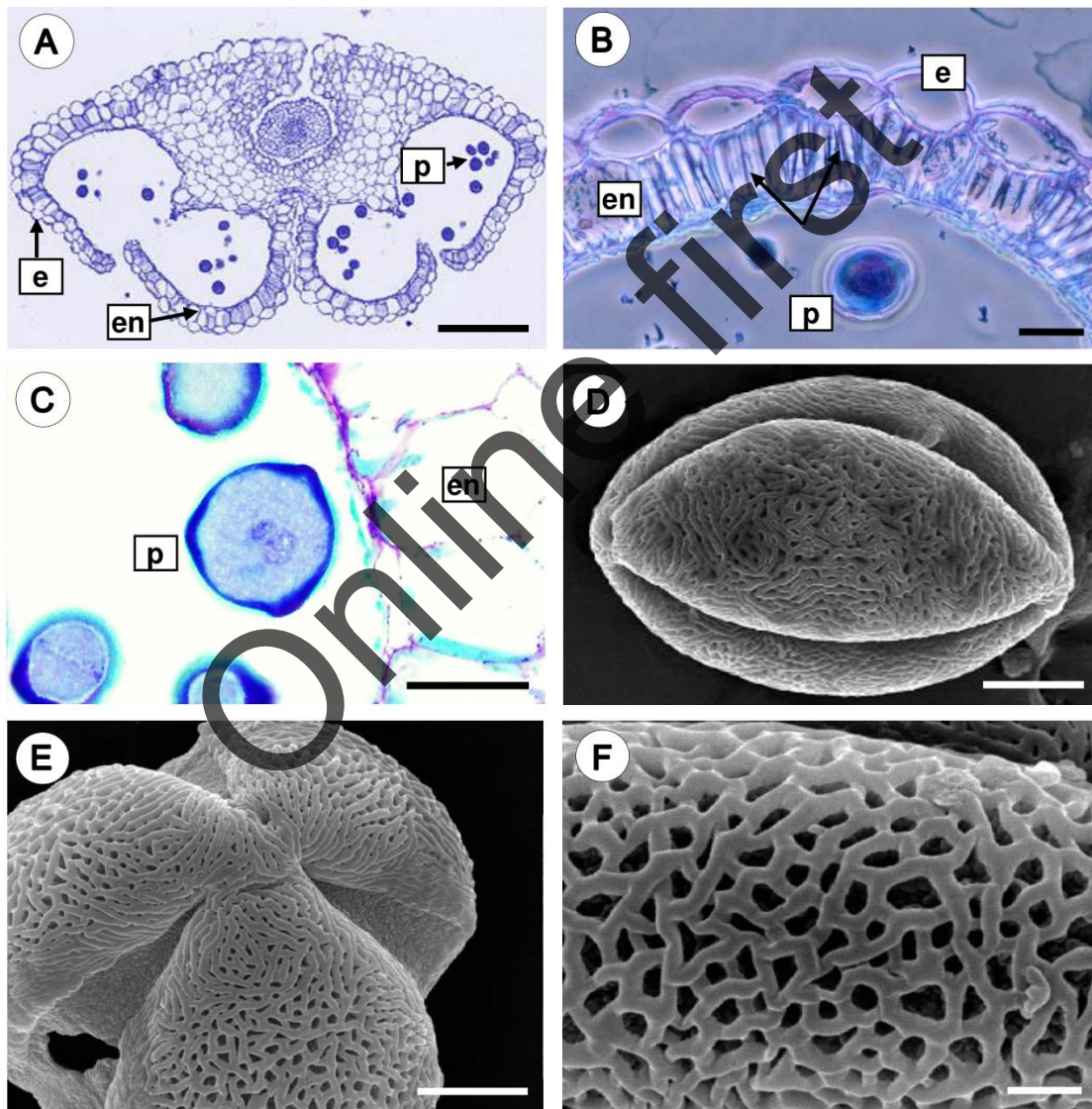


Figure 5. Mature anther and pollen grain of *Fouquieria fasciculata*. A. Dehiscent anther showing epidermal cells, endothecium with wall thickenings, and pollen grains. B. Detail of the endothecium cells with U-shaped fibrous thickenings (black arrows). C. The mature anther with two-celled pollen grain. D. Equatorial view of a 3-colpate pollen grain. E. Polar view of pollen grain. F. Close up of reticulate-heterobrochate exine. e, epidermis; en, endothecium; p, pollen grains. Scale bars = 200 μm (A), 20 μm (B-C), 5 μm (D-E), 1 μm (F).

tion and disintegrates just before anther dehiscence. Tapetum plays a main role in the development of the pollen grains, and some of its main functions are the production and release of callase enzyme, the formation of exine precursors, and the production and release of orbicules (Pacini *et al.* 1985, Pacini & Franchi 1993, Pacini 1997, Huysmans *et al.* 1998, Bhojwani & Bhatnagar 1999, Lei & Liu 2020, Shamrov *et al.* 2021). In angiosperms, the distribution of tapetum types have evolved several times through the phylogeny. Most of basal angiosperms present a secretory tapetum, whereas plasmodial tapetum is relatively consistent in monocotyledons (Furness & Rudall 2001, Shamrov *et al.* 2021); and in eudicots the secretory tapetum is predominant (Furness 2008). Based on previous studies in Ericales, embryological characters related the development of male structures vary among the families and might be informative for understanding evolution in the order (Janssens *et al.* 2005, Schönenberger & Grenhagen 2005, Schönenberger 2009). Most families in Ericales have secretory tapetum with binucleate cells including family Fouquieriaceae (Johri *et al.* 1992).

Although Fouquieriaceae is known to have a secretory tapetum (Henrickson 1972, Johri *et al.* 1992), orbicules have never been examined in this family. Recent studies have been focused orbicules morphological diversity in different plant species (*e.g.*, Song *et al.* 2017, Moon 2018, Galati *et al.* 2019, Ruggiero & Bedini 2020). In Ericales, *Pelliciera* (Pellicieraceae) presents aggregated orbicules with irregular shape and psilate surface (Janssens *et al.* 2005), while in Marcgraviaceae orbicules are mostly round or elliptic with smooth surface (Lens *et al.* 2005). *Fouquieria fasciculata* present two orbicules types: doughnut-shape orbicules and ruminant orbicules, both were abundantly distributed on the radial and inner tangential wall of tapetal cells. According to Galati *et al.* (2010), two or more units of orbicules could be grouped forming aggregates as in *F. fasciculata*. Thus, we suggested that doughnut-shape orbicules could be early stages of the ruminant orbicules; but further studies including the morphology and ultrastructure of the orbicules in Fouquieriaceae are needed to understanding their origin and development. Furthermore, the orbicules surface is granular and resemble the exine of the pollen grain, which could indicate a similar-patterned biosynthesis of sporopollenin (Ruggiero & Bedini 2020).

Studies on the structure and development of orbicules have suggested different functions, such as: a) lysis and degeneration of tapetal cells (Rowley & Erdtman 1967, Herich & Lux 1985), b) retain on their surface sporopollenin for the normal development of the microspores (El-Ghazaly & Jensen 1986, El-Ghazaly & Nilsson 1991), c) act as vectors of allergens and factors for why pollen causes allergic reactions (Vinckier & Smets 2001, Vinckier *et al.* 2005), d) associated to the pollen dispersion and the pollination mode (Heslop-Harrison 1968, Keijzer 1984, Pacini 1997, Galati *et al.* 2010, Verstraete *et al.* 2014). According to Galati *et al.* (2010), orbicules with an irregular surface and/or perforations would be common among of species pollinated by hummingbirds or long-tongued insects. Our observations in *F. fasciculata* showed that some orbicules have a single perforation giving them a doughnut-like appearance, which suggest that flowers could be primarily pollinated by either hummingbirds or long-tongued insects. As mentioned previously, most of Fouquieriaceae species with tubular red flowers are pollinated by hummingbirds, but the pollinators are not known for *F. fasciculata*, whose flowers are white. Further studies are needed to confirm the presence or absence of orbicules in other species of the family, and to understand the possible correlation between their morphology and pollinators.

Orbicules have phylogenetic and taxonomic value in several families of angiosperms, such as, Apocynaceae, Gentianaceae, Loganiaceae, Oxalidaceae, Rosaceae, and Rubiaceae (Huysmans *et al.* 1997, Vinckier *et al.* 2000, Vinckier & Smets 2002a, b, 2003, Rosenfeldt & Galati 2008, Verstraete *et al.* 2014, Song *et al.* 2017, Romero *et al.* 2017, Moon 2018, Ruggiero & Bedini 2020). This study showed the presence of orbicules in Fouquieriaceae for the first time, other families of order Ericales, such as Marcgraviaceae and Pellicieraceae presented consistently orbicules, while are completely absent in Balsaminaceae and Tetrameristaceae (Janssens *et al.* 2005). The presence of orbicules generally represents a plesiomorphic trait for angiosperms since orbicules are common in the basal clades (Verstraete *et al.* 2014, Ruggiero & Bedini 2020). Phylogenetic studies indicate that Fouquieriaceae is an ancient lineage that diverged from Polemoniaceae at approximately 75.5 Ma during the late Cretaceous (Schultheis & Baldwin 1999, Anderberg *et al.* 2002, De Nova *et al.* 2018); but orbicules have not been described in Polemoniaceae. More investigations are needed to clarify the systematic significance of orbicules in family Fouquieriaceae and its phylogenetic relationships in order Ericales.

Regarding the pollen morphology in Fouquieriaceae, Henrickson (1967) carried out a comprehensive study in the family and showed that the pollen grains are prolate, tricolporate, and the exine is reticulate, which is corroborated here for *F. fasciculata*, further the exine of this species is heterobrochate. According to Henrickson (1972), pollen grains are shed as single-celled monads, and our observations showed that are released in two-celled stage.

The present study carrying out detailed research of development of androecium in *F. fasciculata* and provide information on tapetum, pollen development and orbicules features that might be informative for understanding embryology and evolution in the family Fouquieriaceae. Further expanded studies about morphological and ultrastructural orbicules diversity in the Fouquieriaceae species are needed to understanding their correlation with the type of pollinators, as well as their taxonomic value at family level.

Acknowledgments

We thank the Plant Development Laboratory of the Faculty of Sciences, UNAM, for the technical support and supply of materials. We also thank A. Martínez Mena† and A. I. Bieler Antolin for microphotography, J. A. Hernández Gómez for digitization of photographs. We thank Ma. Eugenia Muñoz Díaz de León for the technical support. We would also like to thank the anonymous reviewers for their comments and suggestions that undoubtedly improved our manuscript.

Literature cited

- Amlie R. 1965. *The development of the embryo sac in the Fouquieriaceae*. MSc. Thesis, University of Arizona.
- Anderberg AA, Rydin C, Källersjö M. 2002. Phylogenetic relationships in the order Ericales *s.l.*: analyses of molecular data from five genes from the plastid and mitochondrial genomes. *American Journal of Botany* **89**: 677-687. DOI: <https://doi.org/10.3732/ajb.89.4.677>
- Bhojwani SS, Bhatnagar SP. 1999. *The embryology of angiosperms*. New Delhi: Vikas Publishing. ISBN 10: 8125923462
- Bozzola JL, Russell LD. 1992. *Electron microscopy: Principles and techniques for biologists*. London: Jones and Bartlett Publishers. ISBN 10: 0763701920
- Carlquist S. 2000. Wood anatomy of Fouquieriaceae in relation to habit, ecology, and systematics; nature of meristems in wood and bark. *Aliso: A Journal of Systematic and Floristic Botany* **19**: 137-163. DOI: <https://doi.org/10.5642/ALISO.20001902.03>
- Davis GL. 1966. *Systematic embryology of the angiosperms*. New York: John Wiley & Sons. ISBN 10: 0471198870
- De Nova JA, Sánchez-Reyes LL, Eguiarte LE, Magallón S. 2018. Recent radiation and dispersal of an ancient lineage: The case of *Fouquieria* (Fouquieriaceae, Ericales) in North American deserts. *Molecular Phylogenetics and Evolution* **126**: 92-104. DOI: <https://doi.org/10.1016/j.ympev.2018.03.026>
- El-Ghazaly G. 1999. Tapetum and orbicules (Ubisch bodies): development, morphology and role of pollen grains and tapetal orbicules in allergenicity. In: Cresti M, Cai G, Moscatelli A, eds. *Fertilization in Higher Plants: Molecular and Cytological Aspects*. Berlin: Springer-Verlag, pp. 157-173. DOI: https://doi.org/10.1007/978-3-642-59969-9_13
- El-Ghazaly G, Jensen WA. 1986. Studies of the development of wheat (*Triticum aestivum*) pollen. I. Formation of the pollen wall and Ubisch bodies. *Grana* **5**: 1-29. DOI: <https://doi.org/10.1080/00173138609429929>
- El-Ghazaly G, Nilsson S. 1991. Development of tapetum and orbicules of *Catharanthus roseus* (Apocynaceae). In: Blackmore S, Barnes SH, eds. *Pollen and Spores: Patterns of Diversification*. Systematics Association Special. Oxford: Clarendon Press, pp. 317-329.
- Furness CA. 2008. A review of the distribution of plasmodial and invasive tapeta in eudicots. *International Journal of Plant Sciences* **169**: 207-223. DOI: <https://doi.org/10.1086/524111>
- Furness CA, Rudall PJ. 1998. The tapetum and systematics in monocotyledons. *The Botanical Review* **64**: 201-239. DOI: <https://doi.org/10.1007/BF02856565>

- Furness CA, Rudall PJ. 2001. The tapetum in basal angiosperms: early diversity. *International Journal of Plant Sciences* **162**: 375-392. DOI: <https://doi.org/10.1086/319580>
- Galati BG, Gotelli MM, Dolinko AE, Rosenfeldt S. 2019. Could microechinate orbicules be related to the release of pollen in anemophilous and 'buzz pollination' species? *Australian Journal of Botany* **67**: 16-35. <https://doi.org/10.1071/BT18169>
- Galati BG, Gotelli MM, Rosenfeldt S, Torretta JP, Zarlavsky G. 2010. Orbicules in relation to the pollination modes. In: Kaiser BJ ed. *Pollen: Structure, Types and Effects* New York: Nova Science Publishers, pp. 1-15. ISBN: 978-1-61668-669-7
- Galati BG, Monaci F, Gotelli MM, Rosenfeldt S. 2007. Pollen, tapetum and orbicule development in *Mediolastrum malvifolium* (Malvaceae). *Annals of Botany* **99**: 755-763. DOI: <https://doi.org/10.1093/aob/mcm011>
- Govil CM. 1970. Fouquieriaceae. *Bulletin of the Indian National Science Academy* **41**: 244-245.
- Henrickson J. 1967. Pollen morphology of the Fouquieriaceae. *Aliso: A Journal of Systematic and Floristic Botany* **6**: 137-160. DOI: <https://doi.org/10.5642/aliso.19670603.08>
- Henrickson J. 1972. A taxonomic revision of Fouquieriaceae. *Aliso: A Journal of Systematic and Floristic Botany* **7**: 439-537.
- Herich R, Lux A. 1985. Lytic activity of Ubisch bodies (orbicules). *Cytologia* **50**: 563-569. DOI: <https://doi.org/10.1508/cytologia.50.563>
- Heslop-Harrison J. 1968. Pollen wall development: the succession of events in the growth of intricately patterned pollen walls is described and discussed. *Science* **16**: 30-237. DOI: <https://doi.org/10.1126/science.161.3838.230>
- Huysmans S, El-Ghazaly G, Smets E. 1998. Orbicules in angiosperms: morphology, function, distribution, and relation with tapetum types. *The Botanical Review* **64**: 240-272. DOI: <https://doi.org/10.1007/BF02856566>
- Huysmans S, Smets E, El-Ghazaly G, Nilsson S. 1997. Systematic value of tapetal orbicules: a preliminary survey of the Cinchonoideae (Rubiaceae). *Canadian Journal of Botany* **75**: 815-826. DOI: <https://doi.org/10.1139/b97-091>
- Janssens S, Lens F, Dressler S, Geuten K, Smets E, Vinckier S. 2005. Palynological variation in balsaminoid Ericales. II. Balsaminaceae, Tetrameristaceae, Pellicieraceae and general conclusions. *Annals of Botany* **96**: 1061-1073. DOI: <https://doi.org/10.1093/aob/mci257>
- Johansen DA. 1936. Morphology and embryology of the *Fouquieria*. *American Journal of Botany* **23**: 95-99. DOI: <https://doi.org/10.2307/2436301>
- Johansen DA. 1940. *Plant microtechnique*. New York: MacGraw-Hill. ISBN 10: 0070325405
- Johri B, Ambegaokar M, Srivastava P. 1992. *Comparative Embryology of Angiosperms*. Vol. II. New York: Springer-Verlag. ISBN 10: 3540536337
- Keijzer CJ. 1984. Pollen dispersal and function of the orbicules. *Acta Botanica Neerlandica* **2**: 244.
- Khan R. 1943. The ovules and embryo sac of *Fouquieria*. *Proceedings of the National Institute of Sciences in India* **9**: 253-256.
- Kubitzki K. 2004. Fouquieriaceae. In: *Flowering Plants: Dicotyledons*. Berlin: Springer Heidelberg. pp. 195-198. ISBN: 978-3-662-07257-8
- Lei X, Liu B. 2020. Tapetum-dependent male meiosis progression in plants: increasing evidence emerges. *Frontiers in Plant Science* **10**: 1667. DOI: <https://doi.org/10.3389/fpls.2019.01667>
- Lens F, Dressler S, Vinckier S, Janssens S, Dessein S, van Evelghem L, Smets E. 2005. Palynological variation in Balsaminoid Ericales. I. Marcgraviaceae. *Annals of Botany* **96**: 1047-1060. DOI: <https://doi.org/10.1093/aob/mci256>
- Mauritzon J. 1936. Zur embryologie einiger Parietales-Familien. *Svensk Botanisk Tidskrift* **30**: 79-113.
- Moon HK. 2018. The phylogenetic potential of orbicules in angiosperms. *Korean Journal of Plant Taxonomy* **48**: 9-23. DOI: <https://doi.org/10.11110/kjpt.2018.48.1.9>
- O'Brien T, Feder N, McCully ME. 1964. Polychromatic staining of plant cell walls by toluidine blue O. *Protoplasma* **59**: 368-373. DOI: <https://doi.org/10.1007/BF01248568>

- Pacini E. 1997. Tapetum character states: analytical keys for tapetum types and activities. *Canadian Journal of Botany* **75**: 1448-1459. DOI: <https://doi.org/10.1139/b97-859>
- Pacini E. 2010. Relationships between tapetum, loculus, and pollen during development. *International Journal of Plant Sciences* **171**: 1-11. DOI: <https://doi.org/10.1086/647923>
- Pacini E, Franchi GG. 1993. Role of the tapetum in pollen and spore dispersal. *Plant Systematics and Evolution* **7**:1-11. DOI: https://doi.org/10.1007/978-3-7091-6661-1_1
- Pacini E, Franchi GG, Hesse M. 1985. The tapetum: its form, function, and possible phylogeny in Embryophyta. *Plant Systematics and Evolution* **149**: 155-185. DOI: <https://doi.org/10.1007/BF00983304>
- Romero MF, Salas R, Gonzalez AM. 2017. Pollen development and orbicule and pollen grain morphology in species of *Cephalanthus* (Rubiaceae-Naucleaeae) from the Americas. *Australian Journal of Botany* **65**: 233-247. DOI: <https://doi.org/10.1071/BT16238>
- Rosenfeldt S, Galati BG. 2008. Orbicules diversity in *Oxalis* species from the province of Buenos Aires (Argentina). *Biocell* **32**: 41-47. <https://doi.org/10.32604/biocell.2008.32.041>
- Rowley JR, Erdtman G. 1967. Sporoderm in *Populus* and *Salix*. *Grana* **7**: 517-567. DOI: <https://doi.org/10.1080/00173136709430048>
- Ruggiero F, Bedini G. 2020. Phylogenetic and morphologic survey of orbicules in angiosperms. *Taxon* **69**: 543-566. DOI: <https://doi.org/10.1002/tax.12281>
- Ruzin S. 1999. *Plant microtechnique and microscopy*. Oxford: Oxford University Press. ISBN 10: 0195089561
- Schönenberger J. 2009. Comparative floral structure and systematics of Fouquieriaceae and Polemoniaceae (Ericales). *International Journal of Plant Sciences* **170**: 1132-1167. DOI: <https://doi.org/10.1086/605875>
- Schönenberger J, Grenhagen A. 2005. Early floral development and androecium organization in Fouquieriaceae (Ericales). *Plant Systematics and Evolution* **254**: 233-249. DOI: <https://doi.org/10.1007/s00606-005-0331-7>
- Schultheis M, Baldwin G. 1999. Molecular phylogenetics of Fouquieriaceae: evidence from nuclear rDNA its studies. *American Journal of Botany* **86**: 578-589. DOI: <https://doi.org/10.2307/2656819>
- Shamrov II, Anisimova GM, Babro AA. 2021. Tapetum types and forms in angiosperms. *Proceedings of the Latvian Academy of Sciences* **75**: 167-179. DOI: <https://doi.org/10.2478/prolas-2021-0026>
- Song JH, Oak MK, Roh HS, Hong SP. 2017. Morphology of pollen and orbicules in the tribe Spiraeae (Rosaceae) and its systematic implications. *Grana* **56**: 351-367. DOI: <https://doi.org/10.1080/00173134.2016.1274334>
- Soto-Trejo F, Magallón S, De-Nova JA, Dávila P, Sánchez-González LA, Oyama K. 2022. The evolutionary history of Fouquieriaceae (Ericales): biogeography, growth habit, habitat colonization and chromosome evolution. *Plant Systematics and Evolution* **308**: 35. DOI: <https://doi.org/10.1007/s00606-022-01827-x>
- Soto-Trejo F, Zamudio-Ruiz S, Espinosa-Matías S. 2021. Ovule development, megasporogenesis, and megagametogenesis of *Fouquieria fasciculata* (Willd. ex Roem. & Schult.) Nash (Fouquieriaceae). *Botanical Sciences* **99**: 877-885. DOI: <https://doi.org/10.17129/botsci.2832>
- Verstraete B, Moon HK, Smets E, Huysmans S. 2014. Orbicules in flowering plants: A phylogenetic perspective on their form and function. *The Botanical Review* **80**: 107-134. DOI: <https://doi.org/10.1007/s12229-014-9135-1>
- Vinckier S, Cadot P, Smets E. 2005. The manifold characters of orbicules: structural diversity, systematic significance, and vectors for allergens. *Grana* **44**: 300-307. DOI: <https://doi.org/10.1080/00173130510031654>
- Vinckier S, Huysmans S, Smets E. 2000. Morphology and ultrastructure of orbicules in the subfamily Ixoroideae (Rubiaceae). *Review of Palaeobotany and Palynology* **108**: 151-174. DOI: [https://doi.org/10.1016/S0034-6667\(99\)00036-6](https://doi.org/10.1016/S0034-6667(99)00036-6)
- Vinckier S, Smets E. 2001. The potential role of orbicules as a vector of allergens. *Allergy* **56**: 1129-1136. DOI: <https://doi.org/10.1034/j.1398-9995.2001.00172.x>
- Vinckier S, Smets E. 2002a. Morphological and ultrastructural diversity of orbicules in relation to evolutionary tendencies in Apocynaceae s.l. *Annals of Botany* **90**: 647-662. DOI: <https://doi.org/10.1093/aob/mcf243>
- Vinckier S, Smets E. 2002b. Morphology, ultrastructure and typology of orbicules in Loganiaceae s.l. and related genera, in relation to systematics. *Review of Palaeobotany and Palynology* **119**: 161-189. DOI: [https://doi.org/10.1016/S0034-6667\(01\)00137-3](https://doi.org/10.1016/S0034-6667(01)00137-3)

- Vinckier S, Smets E. 2003. Morphological and ultrastructural diversity of orbicules in Gentianaceae. *Annals of Botany* **92**: 657-672. DOI: <https://doi.org/10.1093/aob/mcg187>
- Waser NM. 1979. Pollinator availability as a determinant of flowering time in ocotillo (*Fouquieria splendens*). *Oecologia* **39**: 107-121. DOI: <https://www.jstor.org/stable/4215803>
- Zamudio S. 1995. Familia Fouquieriaceae. *Flora del Bajío y de Regiones Adyacentes*. **36**: 1-7.

Editor de sección: Maria Silvia Ferrucci

Author contributions: FST, field work, serial microtome sectioning, analysis and writing. SZR, field work, specimen examination, writing. SEM, field work, scanning electron microscopy, analysis and writing.

Online first