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Citation for published version:

ONDRI Investigators, Dillott, AA, Berberian, SA, Sunderland, KM, Binns, MA, Zimmer, J, Ozzoude, M, Scott, CJM, Gao, F, Lang, AE, Breen, DP, Tartaglia, MC, Tan, B, Swartz, RH, Rogaeva, E, Borrie, M, Finger, E, Fischer, CE, Frank, A, Freedman, M, Kumar, S, Pasternak, S, Pollock, BG, Rajji, TK, Tang-Wai, DF, Abrahao, A, Turnbull, J, Zinman, L, Casaubon, L, Dowlatshahi, D, Hassan, A, Mandzia, J, Sahlas, D, Saposnik, G, Grimes, D, Marras, C, Steeves, T, Masellis, M, Farhan, SMK, Bartha, R, Symons, S, Hegele, RA, Black, SE & Ramirez, J 2023, 'Rare neurovascular genetic and imaging markers across neurodegenerative diseases', *Alzheimer's & Dementia: The Journal of the Alzheimer's Association*. https://doi.org/10.1002/alz.13316

Digital Object Identifier (DOI):

10.1002/alz.13316

Link:

Link to publication record in Edinburgh Research Explorer

Document Version: Publisher's PDF, also known as Version of record

Published In: Alzheimer's & Dementia: The Journal of the Alzheimer's Association

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RESEARCH ARTICLE

Alzheimer's & Dementia® THE JOURNAL OF THE ALZHEIMER'S ASSOCIATION

Rare neurovascular genetic and imaging markers across neurodegenerative diseases

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Abstract

Introduction: Cerebral small vessel disease (SVD) is common in patients with cognitive impairment and neurodegenerative diseases such as Alzheimer's and Parkinson's. This study investigated the burden of magnetic resonance imaging (MRI)-based markers of SVD in patients with neurodegenerative diseases as a function of rare genetic variant carrier status.

Methods: The Ontario Neurodegenerative Disease Research Initiative study included 520 participants, recruited from 14 tertiary care centers, diagnosed with various neurodegenerative diseases and determined the carrier status of rare non-synonymous variants in five genes (*ABCC6, COL4A1/COL4A2, NOTCH3/HTRA1*).

Results: NOTCH3/HTRA1 were found to significantly influence SVD neuroimaging outcomes; however, the mechanisms by which these variants contribute to disease progression or worsen clinical correlates are not yet understood.

Discussion: Further studies are needed to develop genetic and imaging neurovascular markers to enhance our understanding of their potential contribution to neurodegenerative diseases.

KEYWORDS HTRA1, neurodegeneration, NOTCH3, ONDRI, small vessel disease, WMH

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1 | INTRODUCTION

Cerebral small vessel disease (SVD) represents a group of pathologies with heterogeneous etiology that affect the small arteries, veins, and capillaries of the brain's vascular system.¹ Notably, SVD most commonly leads to vascular cognitive impairment and dementia in an aging population. Additionally, SVD is associated with several markers that are quantifiable on structural magnetic resonance imaging (MRI), including white matter hyperintensities (WMH). MRI-visible perivascular spaces (PVS), and lacunes.² These observable neuroimaging features have been reported in various neurodegenerative diseases, including Alzheimer's disease (AD), mild cognitive impairment (MCI), amyotrophic lateral sclerosis (ALS). Parkinson's disease (PD), and frontotemporal dementia (FTD), as well as in cerebrovascular disease (CVD),³⁻¹⁰ all of which are represented in the Ontario Neurodegenerative Disease Research Initiative (ONDRI). These diagnoses were selected based on their high prevalence within the aging population as well as the high rates of neuropsychological dysfunction reported in these patient populations. As the cases of dementia increase worldwide, one of the key objectives of ONDRI is to accurately identify the individuals at risk of developing dementia, with and without comorbidities with other neurodegenerative and/or neurovascular disorders, at an earlier stage. Therefore, these diagnoses were selected to align with this objective.

Previously, several rare nonsynonymous variants had been associated with vascular neuroimaging markers in various neurodegenerative diseases via linkage analyses and candidate gene sequencing of patient cohorts.^{11–17} Specifically, five genes that were previously associated with SVD and vasculopathy include ATP Binding Cassette Subfamily C Member 6 (ABCC6), Collagen Type IV Alpha-I (COL4A1), Collagen Type IV Alpha-II (COL4A2), High-Temperature Requirement A Serine Peptidase 1 (HTRA1), and Notch Receptor 3 (NOTCH3). Under normal conditions, ABCC6 encodes a protein that aids in the transportation of molecules across the cell membrane; however, when mutated, it can cause arterial calcification and pseudoxanthoma elasticum, an autosomal recessive disease primarily affecting the connective tissue in the skin, retina, and cardiovascular system, and more recently the gene has been associated with increased ischemic strokes.^{18,19} COL4A1 and COL4A2 encode for collagen chain proteins that combine and together play a critical role in supporting the basement membranes of human vasculature.²⁰ Autosomal dominant COL4A1 mutations cause COL4A1-related SVD, which presents as early strokes and brain cysts,²¹ whereas autosomal dominant COL4A2 mutations cause intracerebral hemorrhage and SVD, which result in neural degeneration.²² NOTCH3 encodes a transmembrane protein receptor critical for the survival and function of vascular smooth muscle cells; however, when mutated it is associated with a monogenic form of SVD commonly known as cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy (CADASIL).¹⁷ Similarly, HTRA1 encodes a protein involved with cell signaling, musculoskeletal development, vascular maturation, and protein degradation; however, when mutated it is associated with a phenotype similar to CADASIL, known as cerebral autosomal recessive arteriopathy with

RESEARCH IN CONTEXT

- Systematic Review: Burden of white matter hyperintensities, perivascular spaces, and lacunes were quantified in a study examining 520 participants diagnosed with Alzheimer's disease, mild cognitive impairment, amyotrophic lateral sclerosis, Parkinson's disease, frontotemporal dementia, or cerebrovascular disease. Carrier status of rare non-synonymous variants in ABCC6, COL4A1, COL4A2, HTRA1, and NOTCH3 was determined for each study participant, and multiple regression models were used to estimate effects across the neuroimaging markers as a function of variant carrier status.
- Interpretation: We observed a significant influence of rare variants in *HTRA1* and *NOTCH3* on neuroimaging markers of cerebral small vessel disease in a variety of neurodegenerative disease patient cohorts.
- 3. Future directions: Our study highlights the importance of genetic analysis in understanding the development of vascular injury and the potential contribution to neurodegenerative disease, yet future work must determine the biological mechanisms of these associations and their influence on downstream clinical outcomes.

subcortical infarcts and leukoencephalopathy (CARASIL).²³ Although the compendium of SVD-associated genes continues to grow, these five largely remain a consensus of the most well-established genes of interest.²⁴

The primary aim of this study was to examine the burden of MRIbased SVD markers (WMH, PVS, and lacunes) as a function of carrier status of non-synonymous, rare genetic variants of interest in a sample of patients with neurodegenerative diseases that are represented in the ONDRI study. In light of these previous findings in genetics and neuroimaging, we hypothesized that carrying rare genetic variation in SVD-associated genes described would contribute to greater burdens of SVD markers in both neurodegenerative disease and CVD patients.

2 | METHODS

2.1 | Participants

ONDRI recruited 520 participants from 14 tertiary care centers across Ontario, Canada, each of whom had passed preliminary screening. Every participant was previously diagnosed with one of five neurodegenerative diseases: (1) AD; (2) amnestic MCI; (3) ALS; (4) FTD; (5) Parkinson's disease (PD); or (6) cerebrovascular disease (CVD) with or without cognitive impairment. Detailed inclusion/exclusion criteria, demographics, and cohort characteristics of the ONDRI study were published previously.^{25,26} Briefly, the prevailing consensus-based clinical diagnostic criteria at the time of enrollment for each ONDRIfocused disease were implemented at each tertiary clinic: AD/MCI patients met National Institute on Aging-Alzheimer's Association criteria for probable or possible AD or MCI, where amnestic MCI patients were tracked annually to ensure accurate characterization of the cohort²⁵; PD patients met the criteria for idiopathic PD defined by the UK's Parkinson's Disease Society Brain Bank clinical diagnostic criteria; ALS patients met El Escorial World Federation of Neurology diagnostic criteria for possible, probable, or definite familial or sporadic ALS; FTD patients included possible or probable behavioral variants of frontotemporal degeneration, agrammatic/non-fluent and semantic variants of primary progressive aphasia, and possible or probable progressive supranuclear palsy. The CVD cohort comprised of individuals who experienced mild to moderate acute ischemic stroke, transient ischemic attack, or subcortical infarction, at least 3 months prior to enrollment, in compliance with the National Institute of Neurological Disorders and Stroke and the Canadian Stroke Network vascular cognitive impairment harmonization standards. Evidence of stroke required clinical imaging confirmation; however, individuals with large vessel occlusive infarction causing severe neurological deficits were excluded. Additionally, the AD/MCI patients' MRI scans were assessed by a neuroradiologist to exclude individuals with non-AD-related causes for cognitive impairment. Ethics approval was obtained from the Research Ethics Board at each participating site. All participants provided written, informed consent.

2.2 Gene sequencing and variant prioritization

Of the 520 participants recruited by ONDRI, 519 had a blood sample collected, and genomic DNA was extracted from them as previously described.²⁷ All participant DNA samples were then sequenced with the custom-designed next-generation sequencing (NGS) gene panel, ONDRISeq, which covers the exonic regions of 80 genes, each previously associated with a neurodegenerative disease or SVD phenotype.²⁸ Briefly, all genomic DNA samples were sequenced using ONDRISeq on the Illumina MiSeq Personal Genome Sequencer (Illumina, San Diego, CA, USA). A custom bioinformatics pipeline was subsequently used to process the FASTQ files and produce a variant calling format (VCF) file and binary alignment map (BAM) file for each participant.^{27,29}

Generated VCF files for each ONDRI participant were annotated using VarSeq® (Golden Helix, Bozeman, MT, USA). Annotations included sequence ontologies, minor allele frequencies (MAFs) from the Genome Aggregation Database (gnomAD; version 3.1), in silico prediction scores from Combined Annotation Dependent Depletion (CADD, version 1.3), and ClinVar pathogenicity classifications. The variant data were then filtered to include only those within genes on ONDRISeq that had been previously associated with SVD or neurovasculopathy, namely, *ABCC6*, *COL4A1*, *COL4A2*, *HTRA1*, *NOTCH3*, *SAMHD1*, and *TREX1*. Of the variants in these genes, we identified those considered non-synonymous, including missense variants, frameshift insertions or deletions, non-frameshift insertions or dele-

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tions, nonsense variants, and splicing variants. Variants were further prioritized to identify those most likely to be deleterious, first by identifying those considered rare in the general population (MAF < 0.01, gnomAD version 3.1), then by selecting variants with a CADD score \geq 20 (top 1% of deleterious variants in the human genome)³⁰ or variants that had been classified as pathogenic or likely pathogenic in ClinVar. All prioritized variants are hereafter referred to as "non-synonymous, rare variants of interest." The genes SAMHD1 and TREX1 were eliminated from further analysis, as no non-synonymous, rare variants of interest were identified in SAMHD1 and only two non-synonymous, rare variants of interest were identified in TREX1, each carried by only a single ONDRI participant.

Based on the similar molecular pathways and disease associations of the genes under study and to preserve statistical power due to our modest sample size, rare, non-synonymous variants of interest identified in *COL4A1* or *COL4A2* were binned, as were variants of interest identified in *NOTCH3* or *HTRA1*. Participants carrying a non-synonymous, rare variant of interest in at least one of *ABCC6*, *COL4A1/COL4A2*, or *NOTCH3/HTRA1* were considered variant positive, while those not carrying a variant in at least one a gene bin were considered variant negative.

All ONDRI participants were also assessed for the APOE genotype using the ONDRISeq data by extracting calls for the APOE variants rs429358(CT):p.Cys130Arg and rs7412(CT):p.Arg176Cys and mapping to the respective genotype, as previously described.³¹ Any participant carrying at least one copy of either the ε 2 or ε 4 genotype were considered APOE variant positive, as both variants have been previously shown to increase the presence of SVD markers, including WMH burden.³²⁻³⁴

2.3 | Neuroimaging

Of the 520 ONDRI study participants, 513 had usable 3 Tesla MRI that included the following sequences: three-dimensional (3D) T1-weighted (T1), interleaved proton density and T2-weighted (T2), and T2-fluid attenuated inversion recovery (FLAIR). Imaging protocol details were published previously.³⁵ All MRI images were evaluated to ensure excellent imaging quality by a medical biophysicist (R.B.) and for clinical incidental findings by a licensed neuroradiologist (S.S.). Neuroimaging-based markers for cerebral SVD (WMH, PVS, lacunes) were quantified using ONDRI's standardized image-processing pipeline³⁶ in compliance with the STRIVE criteria.² Following initial segmentation, PVS and lacunar counts were generated using a 3D, six-connected voxel connectivity contour segmentation algorithm.³⁷

2.4 Statistical analyses

Of the 513 ONDRI participants with usable 3 Tesla MRI and 519 ONDRI participants that underwent genetic testing, 512 had both imaging and genetics data available. However, two participants' imaging segmentation failed quality control of the aforementioned TABLE 1 Participants in ONDRI carrying non-synonymous, rare variants of interest in CVD-associated genes.

Cohort	Sample size	ABCC6 variant positive	COL4A1/COL4A2 variant positive	HTRA1/NOTCH3 variant positive
ONDRI	510	27 (5.3%)	28 (5.5%)	42 (8.2%)
AD/MCI	126	5 (4.0%)	8 (6.3%)	8 (6.3%)
ALS	38	1 (2.6%)	2 (5.3%)	3 (7.9%)
FTD	52	1 (1.9%)	3 (5.8%)	3 (5.8%)
PD	139	6 (4.3%)	10 (7.2%)	15 (10.8%)
CVD	155	14 (9.0%)	5 (3.2%)	13 (7.9%)

Abbreviations: AD/MCI, Alzheimer's disease/mild cognitive impairment; ALS, amyotrophic lateral sclerosis; CVD, cerebrovascular disease with or without cognitive impairment; FTD, frontotemporal dementia; ONDRI, Ontario Neurodegenerative Disease Research Initiative; PD, Parkinson's disease.

image-processing pipeline, resulting in a final ONDRI total cohort size of 510 used in our analyses.

Multiple regression models were used to estimate effects across the three neuroimaging-based markers of SVD (PVS, lacunes, and WMH) as a function of carrier status of rare, non-synonymous variants of interest in each of the gene bins (ABCC6, COL4A1/COL4A2, and HTRA1/NOTCH3) and of disease cohort (AD/MCI, ALS, FTD, PD, CVD), while also accounting for interactions between the gene bins and disease cohort.

The individual coefficient estimates were used to determine the extent to which predictor variables contributed to each neuroimaging metric. We applied negative binomial generalized linear models for the outcome variables that were counts, namely, PVS counts and lacune counts, whereas we applied Gamma log distribution generalized linear models for the continuous variable of WMH volume. All multiple regression models were adjusted for age, sex, supratentorial total intracranial volume (head size), smoking history, and *APOE* carrier status.

To maximize statistical power based on our modest sample sizes, participants diagnosed with AD and participants diagnosed with MCI were binned into a single AD/MCI cohort. Participant disease cohorts were transformed using a weighted effect coding with the wec R package version 0.4-1,³⁸ which adjusts the point of reference for the dataset to be the sample mean of all participants, regardless of cohort. However, one cohort needed to be excluded from the model coding to avoid statistical redundancy. As the ALS cohort was reported to have the least amount of SVD pathology anecdotally, it was chosen as the remainder cohort. The model was rerun using FTD as the omitted cohort to obtain coefficient estimates for the ALS cohort that could be included in the visualizations of the results.³⁹

To adjust for multiple comparisons, corrected p values were calculated using the Benjamini–Hochberg false discovery rate (FDR) method.⁴⁰ Significance for the multiple regression models was then measured at an alpha level of 0.05.

Statistical estimates were calculated using R statistical software version $3.6.0^{41}$ in R Studio 1.1.463, and data visualization was performed using the ggplot2 R package version $3.3.s.^{42}$

3 RESULTS

3.1 Study participants and variant prioritization

In total, after quality control evaluation, genomic and neuroimaging data were available for 510 ONDRI participants (AD/MCI = 126, ALS = 38, FTD = 52, PD = 139, CVD = 155), of which 104 carried a non-synonymous, rare variant of interest in at least one of the genes: ABCC6, COL4A1, COL4A2, HTRA2, NOTCH3 (Table 1). Of the genes examined in this study, 14 unique variants were found in ABCC6, 11 unique variants were found in COL4A1, nine unique variants were found in COL4A2, six unique variants were found in HTRA1, and 21 unique variants were found in NOTCH3 (Supplemental Table S1). All variants identified were of heterozygous zygosity. HTRA1/NOTCH3 non-synonymous, rare variants of interest were found at the greatest frequency in the full ONDRI cohort, with the highest proportion of carriers in the PD cohort. Four participants were found to carry rare variants in multiple gene groupings, including two participants diagnosed with CVD carrying rare variants in ABCC6 and NOTCH3, one participant diagnosed with PD carrying rare variants in ABCC6 and HTRA1, and one participant diagnosed with FTD carrying rare variants in COL4A2 and NOTCH3. Demographic and whole-brain volumetric comparisons of the ONDRI participants carrying a non-synonymous, rare variant of interest in a SVD-associated gene to non-carriers are displayed in Table 2.

Using multiple regression models, we assessed the contribution of variant carrier status in *ABCC6*, *COL4A1/COL4A2*, or *NOTCH3/HTRA1*; disease cohort; and the two predictor variables to the neuroimaging volumetrics PVS count, lacune count, and WMH volume. Including an interaction term between disease cohort and variant carrier status within the model allowed us to determine the influence of carrying a non-synonymous, rare variant of interest on participant neuroimaging volumetrics in each individual ONDRI disease cohort. Supplementary Figure S1 displays the magnitude of difference in the neuroimaging volumetrics of participants carrying non-synonymous, rare variants of interest in *ABCC6*, *COL4A1/COL4A2*, and *NOTCH3/HTRA1* in comparison to non-carriers across the full ONDRI cohort, as well as in each

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TABLE 2 Demographics and whole-brain volumetrics from magnetic resonance imaging comparison of ONDRI participants carrying and not carrying non-synonymous, rare variants of interest in a CVD-associated gene.

Descriptor	Total cohort	Variant carriers	Variant non-carriers	Effect size
Sample size	510	95	415	
Demographics				
Sex (male:female)	341:169	61:34	280:135	2.7e-2
Age (yr)	68.6 (7.7)	69.0 (7.2)	68.6 (7.9)	4.0e-4
Education (yr)	14.9 (2.9)	15.0 (2.8)	14.9 (3.0)	4.0e-4
MoCA	24.4 (3.4)	24.9 (3.2)	24.3 (3.4)	4.1e-3
Modified Rankin score	1.4 (0.9)	1.3 (0.8)	1.5 (0.9)	4.9e-3
Whole-brain volumetrics				
Supratentorial intracranial volume (cc)	1.3e6 (1.4e5)	1.2e6 (1.5e5)	1.3e6 (1.4e5)	3.0e-4
Normal-appearing white matter (cc)	4.1e5 (6.6e4)	4.0e5 (6.7e4)	4.1e5 (6.6e4)	2.2e-3
Normal-appearing gray matter (cc)	5.5e5 (5.5e4)	5.4e5 (6.0e4)	5.5e5 (5.4e4)	2.0e-4
Sulcal cerebrospinal fluid (cc)	2.5e5 (6.0e4)	2.5e5 (6.3e4)	2.5e5 (6.0e4)	2.0e-5
Ventricular cerebrospinal fluid (cc)	4.1e4 (2.3e4)	4.0e4 (2.2e4)	4.1e4 (2.3e4)	0.0
Periventricular WMH (mm ³)	5.6e3 (8.4e3)	7.5e3 (1.1e4)	5.2e3 (7.5e3)	1.2e-2
Deep WMH (mm ³)	6.5e2 (9.8e2)	7.3e2 (1.0e3)	6.3e2 (9.7e2)	1.4e-3
APOE ε 2 or ε 4 carriers	222	44	178	2.7e-2

Note: Values in table represent mean (standard deviation). Effect sizes were calculated using eta squared of an ANOVA model. A large effect size is defined as >0.8, whereas a small effect size is defined as <0.2. All effect sizes reported were considered small.

Abbreviations: APOE, apolipoprotein E; MoCA, Montreal Cognitive Assessment; WMH, white matter hyperintensities; yr, years.

individual disease cohort. To best illustrate these comparisons, disease effects were removed.

3.2 Associations between ABCC6, COL4A1, and COL4A2 and SVD burden

Carrying a non-synonymous, rare variant of interest in ABCC6 or COL4A1/COL4A2 was not found to significantly influence ONDRI participants' PVS counts, lacune counts, or WMH volume across the full ONDRI cohort (Supplemental Tables S2 and S3). Similarly, carrier statuses of rare variants of interest in ABCC6 or COL4A1/COL4A2 were not significantly associated with SVD markers in either the individual neurodegenerative disease cohorts or the CVD cohort.

3.3 Associations between HTRA1 or NOTCH3 and SVD burden

In contrast, carrying a non-synonymous, rare variant of interest in NOTCH3/HTRA1 was found to significantly influence brain imaging outcomes both across the full ONDRI cohort and within specific diagnostic cohorts (Table 3). More specifically, carriers of non-synonymous, rare variants of interest in NOTCH3/HTRA1 displayed significantly higher WMH volumes, agnostic of participant diagnosis (p = 4.92e-03, FDR: p = 1.82e-02; Figure 1A). Further, in participants diagnosed with CVD, carrying a rare variant of interest in NOTCH3/HTRA1 was

associated with significantly increased WMH volume, although the result was no longer significant following multiple testing corrections (p = 4.94e-02, FDR: p = 1.30e-01; Figure 1B). No associations of interest were observed between *NOTCH3/HTRA1* variant status and WMH volume in any of the individual neurodegenerative disease cohorts.

Interestingly, the directions of association of non-synonymous, rare variant of interest carrier status in NOTCH3/HTRA1 were not uniform across the disease cohorts with respect to lacune count and PVS count. Although variant status was not associated with lacune counts or PVS counts agnostic of disease cohort compared to variant non-carriers (Figures 2A and 3A, respectively), variant carriers diagnosed with CVD displayed a significantly greater number of lacunes (p = 9.93e-03, FDR: p = 3.29e-02; Figure 2B) and PVS (p = 5.93e-04, FDR: p = 2.77e-03; Figure 3B). Similarly, NOTCH3/HTRA1 variant carriers diagnosed with AD/MCI displayed significantly greater number of lacunes than non-carriers, but the result was not significant following multiple testing corrections (p = 4.35e-02, FDR: p = 1.25e-01; Figure 2B). In contrast, NOTCH3/HTRA1 variant carriers diagnosed with PD displayed significantly fewer lacunes than non-carriers (p = 1.38e-02, FDR: p = 4.45e-02; Figure 2B), while carriers diagnosed with FTD displayed significantly fewer PVS, but again this result was not significant following multiple testing corrections (p = 2.35e-02, FDR: p = 7.04e-02; Figure 3B). No additional significant associations were identified between non-synonymous, rare variant of interest carrier status in NOTCH3/HTRA1 and brain volumetric outcomes in any of the individual disease cohorts.

	Lacune count				PVS count				WMH volume	a		
		Standard		FDR p		Standard		FDR p		Standard		FDR p
	Estimate	deviation	P value	value	Estimate	deviation	P value	value	Estimate	deviation	P value	value
Intercept	-3.9	0.9	8.5e-06	7.7e-05	6.6e-02	0.5	8.9e-01	9.6e-01	1.6	0.6	4.9e-03	1.8e-02
NOTCH3/HTRA1 Pos	-4.1e-02	0.3	8.9e-01	9.6e-01	0.2	0.1	9.3e-02	2.1e-01	0.5	0.2	4.3e-04	2.1e-03
AD/MCI	-0.2	0.1	1.5e-01	3.2e-01	9.3e-02	6.7e-02	1.7e-01	3.5e-01	-0.3	8.3e-02	1.2e-04	8.2e-04
NOTCH3/HTRA1 Pos × AD/MCI	1.0	0.5	4.4e-02	1.3e-01	-0.3	0.3	3.2e-01	5.4e-01	8.6e-02	0.3	8.0e-01	8.9e-01
CVD	0.3	0.1	6.4e-03	2.4e-02	0.2	5.8e-02	4.4e-03	1.7e-02	0.6	7.2e-02	4.5e-14	9.4e-13
NOTCH3/HTRA1 Pos × CVD	1.0	0.4	9.9e-03	3.3e-02	0.7	0.2	5.9e-04	2.8e-03	0.5	0.2	4.9e-02	1.3e-01
FTD	-8.6e-02	0.2	6.9e-01	8.4e-01	-3.1e-02	0.1	7.8e-01	9.1e-01	-0.1	0.1	4.3e-01	6.2e-01
NOTCH3/HTRA1 Pos × FTD	-0.9	1.2	4.5e-01	6.3e-01	-1.2	0.5	2.4e-02	7.0e-02	-0.6	0.6	3.2e-01	5.2e-01
PD	-0.4	0.1	4.3e-03	1.7e-02	-0.1	6.6e-02	9.4e-02	2.1e-01	-0.2	8.1e-02	1.3e-02	4.0e-02
NOTCH3/HTRA1 Pos × PD	-1.1	0.4	1.4e-02	4.5e-02	-0.2	0.2	3.0e-01	5.2e-01	0.1	0.2	6.1e-01	8.0e-01
ALS	0.9	0.2	ı	ı	-0.5	0.1		ı	-0.3	0.2	ı	ī
NOTCH3/HTRA1 Pos × ALS	-0.5	0.9	I	ı	-6.9e-02	0.5	ı	ı	-0.7	0.6	I	ī
<i>lote</i> : Participant diseas	se cohorts were tree treet	ransformed using the ALS cohort, a	g weighted eff as displayed be	ect coding usinנ slow the line. Foו	g the ALS cohor r the outcome v	t as the cohort th ariables that wen	at was not cod e counts, namel	ed in the mode y PVS counts a	el. The model wa nd lacune count:	s rerun using the s, negative binom	ETD cohort a ial generalized	s the exclu linear mo

TABLE 3 Results of multiple regression analyses used to analyze the contribution of variant carrier status in NOTCH3/HTRA1 and disease cohort to neuroimaging metrics of PVS count, lacune cou ed els were applied. For the continuous variable of WMH volume, Gamma log distribution generalized linear models were applied. Not moc

Significance for the multiple regression models was measured at an alpha level of 0.05, and significant results are indicated in bold.

Abbreviations: AD/MCI, Alzheimer's disease and mild cognitive impairment; ALS, amyotrophic lateral sclerosis; CVD, cerebrovascular disease with or without cognitive impairment; Dev., deviation; FTD, frontotemporal dementia; PD, Parkinson's disease; Pos, variant positive; PVS, MRI-visible perivascular spaces; WMH, white matter hyperintensities.

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FIGURE 1 White matter hyperintensity (WMH) volume in the ONDRI participants carrying *NOTCH3/HTRA1* rare, non-synonymous variants compared to variant negative participants. (A) WMH volumes compared to head size volume in ONDRI participants that carried a rare, non-synonymous variant in either *NOTCH3* or *HTRA1* and those that did not. (B) WMH volumes compared to head size volume of participants within each disease cohort of ONDRI that carried a rare, non-synonymous variant in either *NOTCH3* or *HTRA1* and those that did not. (B) WMH volumes compared to head size volume of participants within each disease cohort of ONDRI that carried a rare, non-synonymous variant in either *NOTCH3* or *HTRA1* compared to those that did not. *P* values are presented for the interaction term of disease status and variant status from the multivariate regression analysis that modeled WMH volume as a function *NOTCH3* or *HTRA1* variant carrier status, disease cohort, and interactions between the two predictor variables, as well as adjustment for head size, age, smoking history, and *APOE* $\varepsilon 2$ or $\varepsilon 4$ variant status. To avoid statistical redundancy, the ALS cohort was not coded in the regression model. ST_TIV, supratentorial total intracranial volume.

4 DISCUSSION

Here we investigated the burden of MRI-based SVD markers, including lacunes, PVS, and WMH, in neurodegenerative disease patients carrying non-synonymous, rare variants in genes previously associated with cerebrovascular neuroimaging features – *ABCC6*, *COL4A1*, *COL4A2*, *HTRA1*, and *NOTCH3*. It is well established that neurodegenerative diseases, which are characterized by neuronal cell loss, display strong genetic components likely driving disease pathology.^{43,44} However, in recent years it has become increasingly more accepted that the course and manifestation of neurodegenerative diseases are also influenced by concurrent/comorbid cerebrovascular injury. In some cases, SVD may drive a neurodegenerative phenotype, as is the case with vascular dementia, whereas in other cases SVD can exist as a concurrent pathology or act as one of many contributors to disease presentation, such as in AD and PD.⁴⁵⁻⁴⁷ In either scenario, it must be recognized that there may be unique genetic factors that can drive SVD markers, which therefore may also have relevance in neurodegenerative disease risk or features.

We determined that across all neurodegenerative disease and CVD patients, non-synonymous, rare variant carrier status in ABCC6 or COL4A1/COL4A2 did not significantly influence lacune count, PVS



FIGURE 2 Lacune counts in ONDRI participants carrying *NOTCH3/HTRA1* rare, non-synonymous variants compared to variant negative participants. (A) Distribution of lacune counts in ONDRI participants that carried a rare, non-synonymous variant in either *NOTCH3* or *HTRA1* compared to those that did not. (B) Distribution of lacune counts in each of the participants within each disease cohort of ONDRI that carried a rare, non-synonymous variant in either *NOTCH3* or *HTRA1* compared to those that did not. (P) Distribution of lacune counts in each of the participants within each disease cohort of ONDRI that carried a rare, non-synonymous variant in either *NOTCH3* or *HTRA1* compared to those that did not. *P* values are presented for the interaction term of disease status and variant status from the multivariate regression analysis that modeled lacune count as a function *NOTCH3* or *HTRA1* variant carrier status, disease cohort, and interactions between the two predictor variables, as well as adjustment for head size, age, smoking history, and *APOE ε*2 or *ε*4 variant status. To avoid statistical redundancy, the ALS cohort was not coded in the regression model. Indicated *P* values are uncorrected for multiple testing. Neg, variant negative; Pos, variant positive.

count, or WMH volume. In contrast, across all neurodegenerative diseases, non-synonymous, rare variant carrier status in *NOTCH3/HTRA1* was associated with higher WMH volumes. Yet the influence of variants in these genes had varying effects across the neurodegenerative diagnoses with respect to lacune and PVS counts. While carriers of *NOTCH3/HTRA1* variants diagnosed with CVD displayed a greater number of lacunes and PVS than non-carriers, carriers diagnosed with PD displayed significantly fewer lacunes than non-carriers, which is discussed further in what follows. Altogether, our results suggest that genetic influence on SVD markers in neurodegenerative disease patients is not only gene-specific but potentially disease-specific as well.

In our neurodegenerative disease cohort, rare genetic variants in *HTRA1* or *NOTCH3* exerted the greatest influence on markers of SVD. As previously described, heterozygous pathogenic variants in *NOTCH3* are causative for CADASIL, while homozygous pathogenic variants in *HTRA1* are causative for CARASIL, with both phenotypes

sharing similar features, such as recurrent ischemic attacks, migraines with aura, cognitive decline and dementia, and hallmark WMH.^{17,16,48} Within our cohort, there were more rare, non-synonymous variants reported in NOTCH3 than in HTRA1 (n = 21 and n = 6, respectively), and indeed, all HTRA1 variants reported were in the heterozygous state. Yet, heterozygous variants in HTRA1 had been previously associated with cerebral SVD but of lesser severity than CARASIL, accounting for their inclusion in our analyses.⁴⁹⁻⁵¹ Similarly, many of the variants in NOTCH3 that we identified were not considered typical of CADASIL pathogenicity, as they were not cysteine-modifying or located within an epidermal growth factor-like repeat; however, previous studies suggested that these atypical NOTCH3 variants might contribute to a CADASIL phenotype of milder severity or to more generalized SVD.52-54 Therefore, while we are not suggesting that the HTRA1 or NOTCH3 rare variants identified herein are causing CARASIL or CADASIL, respectively, as confirmed by clinical assessment of our study patients, our results do suggest that the

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FIGURE 3 Enlarged PVS counts in ONDRI participants carrying *NOTCH3/HTRA1* rare, non-synonymous variants compared to variant negative participants. (A) Distribution of PVS counts in ONDRI participants that carried a rare, non-synonymous variant in either *NOTCH3* or *HTRA1* compared to those that did not. (B) Distribution of PVS counts in each of the participants within each disease cohort of ONDRI that carried a rare, non-synonymous variant in either *NOTCH3* or *HTRA1* compared to those that did not. P values are presented for the interaction term of disease status and variant status from the multivariate regression analysis that modeled PVS count as a function *NOTCH3* or *HTRA1* variant carrier status, disease cohort, and interactions between the two predictor variables, as well as adjustment for head size, age, smoking history, and APOE *ε*2 or *ε*4 variant status. To avoid statistical redundancy, the ALS cohort was not coded in the regression model. Indicated *P* values are uncorrected for multiple testing. Neg, variant negative; Pos, variant positive.

identified non-synonymous, rare variants identified may contribute to cerebrovascular injury in subjects, potentially acting as a contributing factor to neurodegenerative disease risk or modifying clinical presentation and progression.

Specifically, we observed an association between *NOTCH3/HTRA1* non-synonymous, rare variant carrier status and greater WMH volume across the entire neurodegenerative disease cohort. The association was unsurprising based on the striking hallmark of WMH in both CADASIL and CARASIL. Additionally, a recent analysis of >200,000 participants who self-reported as healthy from the UK Biobank identified a significant association between *NOTCH3* genetic variation and increased WMH volume, as well as ultrastructural damage to white matter.⁵⁵ Notably, an association was also observed between *HTRA1* or *NOTCH3* variants and increased WMH volume in the CVD cohort; however, this result did not pass multiple testing corrections, and therefore, we believe it is unlikely for it to fully account for the association observed across the entire cohort and that the variation is still

influencing SVD markers across the neurodegenerative diseases as well. What remains to be determined is how the influence of *HTRA1* or *NOTCH3* variants on WMH volume may contribute to neurodegenerative disease risk, presentation, and progression, particularly due to the complex relationship between cerebrovascular injury and neurodegeneration.

In contrast to the WMH analysis, NOTCH3/HTRA1 variant carrier status had variable effects on lacune and PVS count across the neurodegenerative disease cohorts. Interestingly, only the CVD cohort displayed an association between genetic variation in NOTCH3/HTRA1 and PVS count, demonstrating a significant positive association, suggesting that the genes have an influence on PVS only in individuals with a classical CVD phenotype.

As described, the PD cohort displayed the highest proportion of *HTRA1/NOTCH3* variant carriers. In fact, a previous gene-based rare variant association analysis by our group identified a significant enrichment of *NOTCH3* rare variants in the PD cohort,²⁹ which led to an initial

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investigation of the contribution of these variants to SVD features in only the PD patients.⁵⁶ Using a Bayesian approach, we identified a doubling of WMH volume in PD patients carrying NOTCH3 variants compared to variant negative patients. Additionally, NOTCH3 rare variation was associated with significantly increased lacune volumes. Yet, we did not observe an association between HTRA1/NOTCH3 variant status and lacune *count* in the PD patients in the current analysis that passed multiple testing corrections. These seemingly contradictory findings highlight a current gap in the clinical-scientific literature regarding the wide variation in definition of lacunes and the differences between lacunar counts versus volumes, size criterion, etiology of infarction, and future cavitation.^{57,58} Given these issues and the difficulty of assessing the precise etiological origins (eg, lipohyalinosis vs ostium/branch disease), the ONDRI study's neuroimaging group implemented a multimodal imaging-based criterion for the segmentation of subcortical lacunar infarcts, defined as regions that are hypointense on T1, hyperintense on T2, and appearing with a hypointense central core with a hyperintense rim on T2 FLAIR.³⁶ They are differentiated from PVS, which are isointense to gray matter on proton density, tend to be smaller in diameter, and are more linear depending on the orientation of the vasculature.² Together with our previous results, this suggests that, although the variant carriers do not have a greater number of lacune counts than non-carriers, the lacunar volumes that are present in the carriers tend to be much more substantial in size, potentially resulting in greater clinical consequences.

Rare genetic variants in ABCC6, COL4A1, and COL4A2 did not account for SVD markers in neurodegenerative disease patients in our study. Of note, only heterozygous variants were observed in the studied genes across our participants. Typically, a homozygous variant is required to cause the phenotype associated with ABCC6, namely, pseudoxanthoma elasticum, although there have been reports of heterozygous variants in the gene causing a milder vascular disease.^{59,60} Yet these cases are rather uncommon, and, due to both the subtlety of the influence of heterozygous ABCC6 variation and our modest sample sizes, we might not have been able to detect their phenotypic influence. Further, pathogenic variants within COL4A1 and COL4A2 are most associated with hemorrhagic stroke⁶¹⁻⁶⁴; however, individuals in the ONDRI cohort were documented to have experienced mild to moderate ischemic stroke, and those with hemorrhagic stroke were excluded. Again, this likely limited the number of variant carriers within our study and, therefore, the ability to detect associations between the collagen chain genes and markers of SVD.

In addition to the limitations imposed by our modest sample sizes described earlier, which might have increased the incidence of results that did not survive multiple testing corrections, the presented work was also inherently limited by focusing upon rare variation, which naturally resulted in fewer variant positive patients to analyze. However, it is generally accepted that variants that are less common in the general population will have a greater likelihood of higher phenotypic effects. Further, our study lacked a normative sample to determine whether the observed associations were specific to neurodegenerative and CVD cohorts, or whether similar results would have been observed in a healthy elderly population. Finally, we did not further analyze the effect of the SVD-associated genes on clinical outcomes or co-morbid vascular risk, again due to the limited power within our study. These analyses will be important for future investigations using larger sample sizes.

The mechanisms by which rare variants in SVD-associated genes – such as *HTRA1* and *NOTCH3* – may contribute to cerebrovascular injury in neurodegenerative disease patients and how that injury may influence progression of neurodegeneration remain to be determined. Yet, our findings highlight the importance of comprehensive genetic study in understanding the development of SVD markers and the potential contribution to neurodegenerative disease, particularly in light of recent literature highlighting the value of clinical genetic testing in cases of cerebral SVD and neurological phenotypes that present with features of SVD. Our results suggest that variants in these genes indeed alter the burden of MRI-based SVD markers in a variety of neurodegenerative disease patients and demonstrate that potential downstream effects on clinical correlates, pathogenesis, and disease progression must be further explored.

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ACKNOWLEDGMENTS

We thank the ONDRI study participants and their respective caregivers for their time and significant contribution to our study. Thank you to the ONDRI investigators (www.ONDRI.ca/people) - the governing, executive, steering, publication, recruiting, assessment, and project management teams. The authors and investigators would like to acknowledge ONDRI's project management team, past and present: Susan Boyd, Lisa Desalaiz, Catarina Downey, Stephanie Hetherington, Heather Hink, Ruth Kruger, Sean Lucas, Donna McBain, Andrea Richter, Michelle Vanderspank, Joanna White, Ashley Wilcox, and the coordinators at each site. Thank you to the L.C. Campbell Foundation and the brainlab.ca neuroimaging analysis team. M.F. received funding support from the Saul A. Silverman Family Foundation as a Canadian International Scientific Exchange Program and the Morris Kerzner Memorial Fund. J.R. acknowledges support from the Ontario Brain Institute, the Sandra Black Centre for Brain Resilience & Recovery, the Weston-Selfridges UK Brain Institute, and Fondation Leducq Transatlantic Network of Excellence. A.A.D. is supported by the Canadian Institute of Health Research Banting Postdoctoral Fellowship program. This research was conducted with the support of the OBI, an independent not-for-profit corporation funded partially by the Ontario government. The opinions, results, and conclusions are those of the authors, and no endorsement by the OBI is intended or should be inferred. Matching funds were provided by participant hospital and research foundations, including the Baycrest Foundation, Bruyere Research Institute, Centre for Addiction and Mental Health Foundation, London Health Sciences Foundation, McMaster University Faculty of Health Sciences, Ottawa Brain and Mind Research Institute, Queen's University Faculty of Health Sciences, the Thunder Bay Regional Health Sciences Centre, the University of Ottawa Faculty of Medicine, and the Windsor/Essex County ALS Association. The Temerty Family Foundation provided the major infrastructure matching funds.

CONFLICT OF INTEREST STATEMENT

The authors report no competing interests. Author disclosures are available in the supporting information.

CONSENT STATEMENT

All study participants provided written, informed consent.

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REFERENCES

- Pantoni L. Cerebral small vessel disease: from pathogenesis and clinical characteristics to therapeutic challenges. *Lancet Neurol.* 2010;9(1474-4465):689-701. Electronic.
- 2. Wardlaw JM, Smith EE, Biessels GJ, et al. Neuroimaging standards for research into small vessel disease and its contribution to ageing and neurodegeneration. *Lancet Neurol.* 2013;12(8):822-838.
- Cai Z, Wang C, He W, et al. Cerebral small vessel disease and Alzheimer's disease. *Clin Interv Aging*. 2015;10:1695-1704. doi:10. 2147/CIA.S90871
- Yoshita M, Fletcher E, Harvey D, et al. Extent and distribution of white matter hyperintensities in normal aging, MCI, and AD. *Neurology*. 2006;67(12):2192-2198.
- Ramirez J, Berezuk C, McNeely AA, Scott CJM, Gao F, Black SE. Visible Virchow-Robin spaces on magnetic resonance imaging of Alzheimer's disease patients and normal elderly from the Sunnybrook dementia study. J Alzheimer's Dis 2015;43(2):415-424. doi:10.3233/JAD-132528
- Wirth AM, Johannesen S, Khomenko A, et al. Value of fluid-attenuated inversion recovery MRI data analyzed by the lesion segmentation toolbox in amyotrophic lateral sclerosis. J Magn Reson Imaging. 2019;50(2):552-559. doi:10.1002/jmri.26577
- Chen H, Wan H, Zhang M, et al. Cerebral small vessel disease may worsen motor function, cognition, and mood in Parkinson's disease. *Parkinsonism Relat Disord*. 2021;83:86-92. doi:10.1016/j.parkreldis. 2020.12.025
- Patankar TF, Mitra D, Varma A, Snowden J, Neary D, Jackson A. Dilatation of the Virchow-Robin space is a sensitive indicator of cerebral microvascular disease: study in elderly patients with dementia. *AJNR Am J Neuroradiol*. 2005;26(0195-6108):1512-1520. Print.
- Desmarais P, Gao AF, Lanctôt K, et al. White matter hyperintensities in autopsy-confirmed frontotemporal lobar degeneration and Alzheimer's disease. *Alzheimers Res Ther.* 2021;13(1):129. doi:10. 1186/s13195-021-00869-6
- Gorelick PB, Scuteri A, Black SE, et al. Vascular contributions to cognitive impairment and dementia: a statement for healthcare professionals from the American Heart Association/American Stroke Association. *Stroke*. 2011;42(9):2672-2713. doi:10.1161/STR. 0b013e3182299496
- Le Saux O, Urban Z, Tschuch C, et al. Mutations in a gene encoding an ABC transporter cause pseudoxanthoma elasticum. *Nat Genet*. 2000;25(2):223-227. doi:10.1038/76102
- Bergen AAB, Plomp AS, Schuurman EJ, et al. Mutations in ABCC6 cause pseudoxanthoma elasticum. *Nat Genet*. 2000;25(2):228-231. doi:10.1038/76109
- Gould DB, Phalan FC, Breedveld GJ, et al. Mutations in Col4a1 cause perinatal cerebral hemorrhage and porencephaly. *Science*. 2005;308(5725):1167-1171. doi:10.1126/SCIENCE.1109418
- Yoneda Y, Haginoya K, Arai H, et al. De novo and inherited mutations in COL4A2, encoding the type IV collagen α2 chain cause porencephaly. *Am J Hum Genet.* 2012;90(1):86-90. doi:10.1016/J.AJHG.2011.11.016
- Weng YC, Sonni A, Labelle-Dumais C, et al. COL4A1 mutations in patients with sporadic late-onset intracerebral hemorrhage. *Ann Neurol.* 2012;71(4):470-477. doi:10.1002/ANA.22682
- Hara K, Shiga A, Fukutake T, et al. Association of HTRA1 mutations and familial ischemic cerebral small-vessel disease. N Engl J Med. 2009;360(17):1729-1739. doi:10.1056/NEJMOA0801560

OURNAL OF THE ALZHEIMER'S ASSOCIATION

- 17. Joutel A. Corpechot C. Ducros A. et al. Notch3 mutations in CADASIL. a hereditary adult-onset condition causing stroke and dementia. Nature. 1996:383(0028-0836):707-710. Print.
- 18. Chassaing N, Martin L, Calvas P, Le Bert M, Hovnanian A. Pseudoxanthoma elasticum: a clinical, pathophysiological and genetic update including 11 novel ABCC6 mutations. J Med Genet. 2005;42(12):881-892. doi:10.1136/jmg.2004.030171
- 19. De Vilder EYG, Cardoen S, Hosen MJ, et al. Pathogenic variants in the ABCC6 gene are associated with an increased risk for ischemic stroke. Brain Pathology. 2018;28(6):822-831. doi:10.1111/bpa.12620
- 20. Khoshnoodi J, Pedchenko V, Hudson BG. Mammalian collagen IV. Microsc Res Tech. 2008;71(5):357-370. doi:10.1002/jemt.20564
- 21. Alamowitch S, Plaisier E, Favrole P, et al. Cerebrovascular disease related to COL4A1 mutations in HANAC syndrome. Neurology. 2009;73(22):1873-1882. doi:10.1212/WNL.0b013e3181c3fd12
- 22. Verbeek E, Meuwissen MEC, Verheijen FW, et al. COL4A2 mutation associated with familial porencephaly and small-vessel disease. Eur J Hum Genet. 2012;20(8):844-851. doi:10.1038/ejhg.2012.20
- 23. Fukutake T. Cerebral autosomal recessive arteriopathy with subcortical infarcts and leukoencephalopathy (CARASIL): from discovery to gene identification. J Stroke Cerebrovasc Dis. 2011;20(2):85-93. doi:10. 1016/j.jstrokecerebrovasdis.2010.11.008
- 24. Manini A, Pantoni L. Genetic causes of cerebral small vessel diseases: a practical guide for neurologists. Neurology. 2023;100(16). doi:10. 1212/WNL.000000000201720
- 25. Farhan SMK, Bartha R, Black SE, et al. The Ontario neurodegenerative disease research initiative (ONDRI). Can J Neurol Sci. 2017;44(2):196-202. doi:10.1017/cjn.2016.415
- 26. Sunderland KM, Beaton D, Arnott SR, et al. Characteristics of the Ontario neurodegenerative disease research initiative cohort. Alzheimer's Dis. 2023;19(1):226-243. doi:10.1002/alz.12632. Published online March 30.
- 27. Dilliott AA, Farhan SMK, Ghani M, et al. Targeted next-generation sequencing and bioinformatics pipeline to evaluate genetic determinants of constitutional disease. J Vis Exp. 2018;134(e57266):1-10. doi:10.3791/57266
- 28. Farhan SMK, Dilliott AA, Ghani M, et al. The ONDRISeq panel: customdesigned next-generation sequencing of genes related to neurodegeneration. NPJ Genom Med. 2016;1(1):16032. doi:10.1038/npjgenmed. 2016.32
- 29. Dilliott AA, Abdelhady A, Sunderland KM, et al. Contribution of rare variant associations to neurodegenerative disease presentation. NPJ Genom Med. 2021;6(1):80. doi:10.1038/S41525-021-00243-3
- 30. Kircher M, Witten DM, Jain P, O'roak BJ, Cooper GM, Shendure J. A general framework for estimating the relative pathogenicity of human genetic variants. Nat Genet. 2014;46(3):310-315. doi:10.1038/ NG.2892
- 31. Dilliott AA, Evans EC, Farhan SMK, et al. Genetic variation in the Ontario neurodegenerative disease research initiative. Can J Neurol Sci. 2019;46(5):491-498. doi:10.1017/cjn.2019.228
- 32. Gesierich B, Opherk C, Rosand J, et al. APOE $\varepsilon 2$ is associated with white matter hyperintensity volume in CADASIL. J Cereb Blood Flow Metab. 2016;36(1):199-203. doi:10.1038/JCBFM.2015.85
- 33. Lyall DM, Cox SR, Lyall LM, et al. Association between APOE e4 and white matter hyperintensity volume, but not total brain volume or white matter integrity. Brain Imaging Behav. 2020;14(5):1468-1476. doi:10.1007/S11682-019-00069-9
- 34. Mirza SS, Saeed U, Knight J, et al. APOE £4, white matter hyperintensities, and cognition in Alzheimer and Lewy body dementia. Neurology. 2019;93(19):e1807-e1819. doi:10.1212/WNL.00000000008377
- 35. Scott CJM, Arnott SR, Chemparathy A, et al. An overview of the quality assurance and quality control of magnetic resonance imaging data for the Ontario Neurodegenerative Disease Research Initiative (ONDRI): pipeline development and neuroinformatics. bioRxiv. 2020. Published online.

- 36. Ramirez J. Holmes MF. Scott CJM, et al. Ontario neurodegenerative disease research initiative (ONDRI): structural MRI methods & outcome measures. Front Neurol. 2020;11:847. Published online.
- 37. Yushkevich PA, Piven J, Hazlett HC, et al. User-guided 3D active contour segmentation of anatomical structures: significantly improved efficiency and reliability. Neuroimage. 2006;31(1053-8119):1116-1128. Print.
- 38. Nieuwenhuis R, te Grotenhuis M, Pelzer B. Weighted effect coding for observational data with wec. R Journal. 2017;9(1):477-485.
- 39. te Grotenhuis M, Pelzer B, Eisinga R, Nieuwenhuis R, Schmidt-Catran A, Konig R. When size matters: advantages of weighted effect coding in observational studies. Int J Public Health. 2017;62(1):163-167. doi:10. 1007/S00038-016-0901-1/TABLES/2
- 40. Yekutieli D, Benjamini Y. Resampling-based false discovery rate controlling multiple test procedures for correlated test statistics. J Stat Plan Inference. 1999;82(1-2):171-196. doi:10.1016/s0378-3758(99) 00041-5
- 41. R Core Team. R: A Language for Statistical Computing. 2013
- 42. Wickham H, ggplot2. ggplot2. Published online 2009. doi:10.1007/ 978-0-387-98141-3
- 43. Bertram L, Tanzi RE. The genetic epidemiology of neurodegenerative disease. J Clin Invest. 2005;115(6):1449-1457. doi:10.1172/JCI24761
- 44. Pihlstrøm L, Wiethoff S, Houlden H. Genetics of neurodegenerative diseases: an overview. Handb Clin Neurol. 2017:145:309-323. doi:10. 1016/B978-0-12-802395-2.00022-5
- 45. Schneider JA, Arvanitakis Z, Bang W, Bennett DA. Mixed brain pathologies account for most dementia cases in community-dwelling older persons. Neurology. 2007;69(24):2197-2204.
- 46. Seidel GA, Giovannetti T, Libon DJ. Cerebrovascular disease and cognition in older adults. Curr Top Behav Neurosci. 2012;10:213-241.. Accessed October 24, 2015. http://www.ncbi.nlm.nih.gov/pubmed/ 21773915
- 47. Kummer BR, Diaz I, Wu X, et al. Associations between cerebrovascular risk factors and Parkinson disease. Ann Neurol. 2019;86(4):572-581. doi:10.1002/ANA.25564
- 48. Tikka S, Baumann M, Siitonen M, et al. CADASIL and CARASIL. Brain Pathol. 2014;24(5):525-544. doi:10.1111/BPA.12181
- 49. Mendioroz M, Fernández-Cadenas I, Del Río-Espinola A, et al. A missense HTRA1 mutation expands CARASIL syndrome to the Caucasian population. Neurology. 2010;75(22):2033-2035. doi:10.1212/ WNL.0B013E3181FF96AC
- 50. Chen Y, He Z, Meng S, Li L, Yang H, Zhang X. A novel mutation of the high-temperature requirement A serine peptidase 1 (HTRA1) gene in a Chinese family with cerebral autosomal recessive arteriopathy with subcortical infarcts and leukoencephalopathy (CARASIL). J Int Med Res. 2013;41(5):1445-1455. doi:10.1177/0300060513480926
- 51. Bianchi S, Di Palma C, Gallus GN, et al. Two novel HTRA1 mutations in a European CARASIL patient. Neurology. 2014;82(10):898-900. doi:10. 1212/WNL.00000000000202
- 52. Ungaro C, Mazzei R, Conforti FL, et al. CADASIL: extended polymorphisms and mutational analysis of the NOTCH3 gene. J Neurosci Res. 2009;87(5):1162-1167. doi:10.1002/jnr.21935
- 53. Ragno M, Berbellini A, Cacchiò G, et al. Parkinsonism is a late, not rare, feature of CADASIL a study on Italian patients carrying the R1006C mutation. Stroke. 2013;44(4):1147-1149. doi:10.1161/STROKEAHA. 111.000458
- 54. Muiño E, Gallego-Fabrega C, Cullell N, et al. Systematic review of cysteine-sparing NOTCH3 missense mutations in patients with clinical suspicion of CADASIL. Int J Mol Sci. 2017;18(9):1964. doi:10.3390/ ijms18091964
- 55. Cho BPH, Nannoni S, Harshfield EL, et al. NOTCH3 variants are more common than expected in the general population and associated with stroke and vascular dementia: an analysis of 200 000 participants. J Neurol Neurosurg Psychiatry. 2021;92(7):694-701. doi:10.1136/JNNP-2020-325838

- Ramirez J, Dilliott AA, Binns MA, et al. Parkinson's disease, NOTCH3 genetic variants, and white matter hyperintensities. *Mov Disord* 2020;35(11):2090-2095. doi:10.1002/mds.28171
- Potter GM, Marlborough FJ, Wardlaw JM. Wide variation in definition, detection, and description of lacunar lesions on imaging. *Stroke*. 2011;42(2):359-366. doi:10.1161/STROKEAHA.110.594754
- Potter GM, Doubal FN, Jackson CA, et al. Counting cavitating lacunes underestimates the burden of lacunar infarction. *Stroke*. 2010;41(2):267-272. doi:10.1161/STROKEAHA.109.566307
- Nollet L, Campens L, De Zaeytijd J, et al. Clinical and subclinical findings in heterozygous ABCC6 carriers: results from a Belgian cohort and clinical practice guidelines. J Med Genet. 2022;59(5). doi:10.1136/ JMEDGENET-2020-107565
- Hornstrup LS, Tybjærg-Hansen A, Haase CL, et al. Heterozygosity for R1141X in ABCC6 and risk of ischemic vascular disease. *Circ Cardiovasc Genet*. 2011;4(5):534-541. doi:10.1161/CIRCGENETICS.110. 958801
- Gould DB, Phalan FC, van Mil SE, et al. Role of COL4A1 in small-vessel disease and hemorrhagic stroke. *NEnglJ Med.* 2006;354(14):1489-1496.
- 62. Lemmens R, Maugeri A, Niessen HWM, et al. Novel COL4A1 mutations cause cerebral small vessel disease by haploinsufficiency. *Hum Mol Genet*. 2013;22(2):391-397. doi:10.1093/HMG/DDS436

- Jeanne M, Labelle-Dumais C, Jorgensen J, et al. COL4A2 mutations impair COL4A1 and COL4A2 secretion and cause hemorrhagic stroke. *Am J Hum Genet.* 2012;90(1):91-101. doi:10.1016/J.AJHG.2011.11. 022
- Meuwissen MEC, Halley DJJ, Smit LS, et al. The expanding phenotype of COL4A1 and COL4A2 mutations: clinical data on 13 newly identified families and a review of the literature. *Genet Med.* 2015;17(11):843-853. doi:10.1038/GIM.2014.210

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Dilliott AA, Berberian SA, Sunderland KM, et al. Rare neurovascular genetic and imaging markers across neurodegenerative diseases. *Alzheimer's Dement*. 2023;1-13. https://doi.org/10.1002/alz.13316