Check for updates

OPEN ACCESS

EDITED BY Navonil De Sarkar, Fred Hutchinson Cancer Research Center, United States

REVIEWED BY

Takeshi Yuasa, Japanese Foundation For Cancer Research, Japan Emilie Lalonde, London Health Sciences Centre, Canada Deepak Kilari, Medical College of Wisconsin, United States

*CORRESPONDENCE Natalia Lukashchuk Matalia.lukashchuk@astrazeneca.com

RECEIVED 09 February 2023 ACCEPTED 01 June 2023 PUBLISHED 26 June 2023

CITATION

Lukashchuk N, Barnicle A, Adelman CA, Armenia J, Kang J, Barrett JC and Harrington EA (2023) Impact of DNA damage repair alterations on prostate cancer progression and metastasis. *Front. Oncol.* 13:1162644. doi: 10.3389/fonc.2023.1162644

COPYRIGHT

© 2023 Lukashchuk, Barnicle, Adelman, Armenia, Kang, Barrett and Harrington. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Impact of DNA damage repair alterations on prostate cancer progression and metastasis

Natalia Lukashchuk^{1*}, Alan Barnicle¹, Carrie A. Adelman¹, Joshua Armenia², Jinyu Kang³, J. Carl Barrett⁴ and Elizabeth A. Harrington¹

¹Translational Medicine, Oncology Research and Development (R&D), AstraZeneca, Cambridge, United Kingdom, ²Oncology Data Science, Oncology Research and Development (R&D), AstraZeneca, Cambridge, United Kingdom, ³Global Medicines Development, Oncology Research and Development (R&D), AstraZeneca, Gaithersburg, MD, United States, ⁴Translational Medicine, Oncology Research and Development (R&D), AstraZeneca, Waltham, MA, United States

Prostate cancer is among the most common diseases worldwide. Despite recent progress with treatments, patients with advanced prostate cancer have poor outcomes and there is a high unmet need in this population. Understanding molecular determinants underlying prostate cancer and the aggressive phenotype of disease can help with design of better clinical trials and improve treatments for these patients. One of the pathways often altered in advanced prostate cancer is DNA damage response (DDR), including alterations in *BRCA1/2* and other homologous recombination repair (HRR) genes. Alterations in the DDR pathway are particularly prevalent in metastatic prostate cancer. In this review, we summarise the prevalence of DDR alterations in the DDR pathway on aggressive disease phenotype, prognosis and the association of germline pathogenic¹ alterations in DDR genes with risk of developing prostate cancer.

KEYWORDS

DNA damage response, homologous recombination repair, alterations, prognosis, prostate cancer

Introduction: prostate cancer and treatment landscape

Prostate cancer is the third most common cancer worldwide, with 1,414,259 new cases and 375,304 deaths in 2020, and the 5th leading cause of cancer deaths worldwide (1, 2). Approximately 80% of men with prostate cancer are diagnosed with localised prostate cancer, and their 10-year survival is up to 99% if diagnosed early. Approximately 10–20% of men with advanced prostate cancer will develop castration-resistant prostate cancer

¹ Throughout the text, germline or somatic alterations mean pathogenic/likely pathogenic, or deleterious/likely deleterious, loss-of-function alterations.

(CRPC) within five years, and at least 84% of these men will have metastases at the time of CRPC diagnosis. Men with metastatic CRPC have poor outcomes (3–5).

Metastatic prostate cancer is a broad term used to describe a range of advanced disease states that are no longer organ-confined. This group includes *de novo* metastatic castration-sensitive prostate cancer (mCSPC), as well as cancers that progress during or after androgen deprivation therapy (ADT), termed metastatic castration-resistant prostate cancer (mCRPC) (6). Based on clinical trials, median overall survival (OS) for patients with mCRPC is approximately three years, and is even less in the real-world setting (7). Approximately half of patients with mCRPC may receive only one line of active treatment, with diminishing benefit observed with the use of subsequent therapies (5, 8–11).

A variety of life-prolonging agents are approved for the mCRPC population overall, however the most used medicines for patients with mCRPC remain mainly chemotherapy (docetaxel/cabazitaxel) and new hormonal agents (NHAs; e.g abiraterone and enzalutamide) (8, 12-14). Once patients with mCRPC have failed NHA, the benefit from approved therapeutic options appears substantially diminished (12, 15-22). As such, there is a high unmet medical need for patients who have progressed after NHA treatment, and many efforts to find more effective treatment options have failed in the past decade. Recently, poly(ADP-ribose) polymerase inhibitors (PARPi) treatment as monotherapy demonstrated radiographic progression-free survival (rPFS) and OS improvement in biomarker-selected patient populations with an underlying alteration in the homologous recombination repair (HRR) pathway (HRRm, or BRCA1/2 alterations) (23-29) and have been approved as monotherapy for patients with HRR mutant (HRRm) mCRPC post NHA (25, 30, 31). The mechanism of action of PARP inhibitors as monotherapy is described in Figure 1. The androgen receptor, in addition to its role in binding androgen and stimulating prostate cancer cell growth (32), also contributes towards the general repair of DNA damage, including damage not normally repaired by HRR (33-35). This demonstrates that androgen receptor and PARP are both important for the repair of DNA damage in prostate cancer cells and highlights the importance of crosstalk (interaction) between both mechanisms. This evidence provided the rationale for the clinical development of co-administration of NHAs and PARP inhibitors (36). The clinical benefit of PARPi in combination with NHA has been demonstrated in biomarker-selected and biomarker-unselected populations in first-line mCRPC (Table 1) (36-39) and approvals have been granted in some regions (40).

Despite recent progress in treatments for metastatic disease, there is a high unmet need in this patient population (5, 41). Understanding the biology underlying primary and metastatic prostate cancer, and the differences in prognosis, can help improve prostate cancer treatment and patient outcomes.

Prostate cancer genomic landscape

Comprehensive molecular characterisation through genetic profiling has revealed a complex and heterogenous genomic landscape of prostate cancer. Multiple landmark genomics studies have identified some of the most recurrent altered genes and pathways in advanced prostate cancer, including genes involved in androgen signalling (50%), PI3K signalling (40%), the cell cycle (24%), WNT/beta-catenin signalling (19%), as well as genes involved in DNA damage response (DDR; 27%), with significant enrichment of all pathways observed in mCRPC (42-46). Alterations in the RAS pathway, including hotspots in BRAF or deleterious alterations in NF1 or RASA1 are detected at lower prevalence than other genes, at around 8% (43). These studies have also highlighted the presence of distinct genomic subtypes defined by rearrangements involving the ERG transcription factor (46%), or hotspot mutations in SPOP (8-11%) and FOXA1 (3%) (44). Due to the highly complex nature of this disease, patients with prostate cancer could greatly benefit from better means of molecular stratification to better select appropriate anticancer therapies.

Prevalence of alterations in the HRR/DDR pathway in prostate cancer

Here we discuss alterations in the DDR pathway, which are frequent in prostate cancer and particularly in advanced stages of disease. DDR is a tightly coordinated pathway that enables cells to control and regulate DNA damage that arises every day. Accumulating damage can lead to mutations and promote genomic instability, which is one of the hallmarks of cancer development (47, 48). Alterations in DDR genes are found to be frequently mutated in many types of cancer, including prostate cancer, where around 23–31% of patients with advanced prostate cancer have been reported to have alterations in DDR genes (43, 45, 46, 49).

As reviewed recently by Morgado & Mateo (50), DDR mutant cancer has been a term used broadly to describe genomic alterations in any gene involved in DDR, including HRR or mismatch repair (MMR) alterations, which have different implications and therapeutic targets in primary and metastatic prostate cancer. Within DDR genes, alterations in genes involved in HRR are most prevalent (23–28%) in mCRPC, with alterations in other pathways of DDR, such as MMR (3–4%) or Fanconi anaemia (FA; around 5%), found at lower prevalence (42, 43, 49, 51, 52).

DNA repair is a complex process that involves sensing DNA damage and downstream signalling cascades to promote DNA repair by recruiting DDR factors and triggering cell cycle checkpoints to allow cells to repair DNA (53, 54). Cancer cells often deregulate the DDR pathway *via* genomic alterations or epigenetic silencing of DDR genes, which can lead to genomic instability – one of the hallmarks of tumourigenesis (47). Alterations in some of the DDR genes, particularly genes involved in HRR in prostate cancer, are associated with worse prognosis and a higher likelihood of developing metastatic disease (55). The HRR pathway is an accurate pathway that regulates the repair of DNA damage, such as double-strand breaks (DSBs). This pathway relies on the presence of sister chromatid, and therefore only operates in S and G2 stages of the cell cycle when the homologous chromatid is

available, whereas the NHEJ pathway takes place in all stages of the cell cycle or in quiescent cells. The HRR pathway is also required for repair of DSBs arising during inter-strand crosslink repair, a process that includes FA factors and other pathways such as nucleotide excision repair and translesion synthesis (47, 48, 56). There are multiple DDR factors that have a direct and indirect role in the HRR pathway: DNA damage sensors (i.e. MRN complex and ATM), signal mediator proteins (i.e. BARD1, BRCA1, PALB2, BRCA2, FANCL, RAD54L), effector proteins directly involved in DNA repair (i.e. RAD51, RAD54L) *via* strand invasion and replication fork stabilisation, downstream signalling to trigger cell cycle checkpoints (ATM, CHEK2, CHEK1) or regulating transcription of HRR genes (i.e. CDK12); (Figure 1E) (57, 58).

Multiple genes in the HRR pathway are altered in several types of cancers, including in prostate cancer; these include: *BRCA1*, *BRCA2*, *ATM*, *CDK12*, *PALB2*, *BRIP1*, *CHEK1*, *CHEK2*, *RAD51B*, *RAD51C*, *RAD51D*, *RAD54L*, *BARD1* and *FANCL* (Table 2).

The most well characterised genes involved in HRR are tumour suppressor genes *BRCA1* and *BRCA2* (also referred to as 'BRCA' genes), alterations in which have been associated with prostate cancer as well as breast, ovarian and pancreatic cancer (61). BRCA1 and BRCA2 proteins play a key role in HRR, where BRCA1 plays a role in the early step of determining DSB repair pathway choice to

promote resection and channel it to HRR, for which interaction with BARD1 is important. Later, BRCA1 interacts with PALB2 to bring BRCA2, which promotes RAD51 filament formation and strand invasion, a key step in HRR (62, 63). RAD54L plays a supportive role by promoting RAD51 filament stabilisation (58) BARD1 and BRIP1 are important BRCA1 interacting partners, promoting repair pathway choice, DNA repair and DDR checkpoints (64, 65). There are other important HRR genes that play a direct or indirect role in HRR, such as a kinase, ATM, which is involved in response to DSBs by phosphorylating key DDR proteins to propagate signalling to promote repair or to arrest the cell cycle. ATM directs repair of DSBs associated with replication to HRR by promoting efficient DNA resection (66). One of the key downstream targets of ATM is CHEK2 kinase, the phosphorylation of which leads to activation of the G1 checkpoint (67-70). On the other hand, CHEK1 kinase is phosphorylated by ATR and is involved in triggering G2/M as well as intra-S checkpoints (68, 69, 71-73). CDK12, a kinase with an indirect role in HRR, regulates transcription of HRR genes (74, 75). Alterations in CDK12 lead to a unique tandem duplication genotype (76).

In prostate cancer, alterations in genes involved in HRR are enriched in later/advanced stages of disease compared with primary prostate cancer (46, 55).

TABLE 1	Clinical trials involving PARP inhibitor	s in mCRPC evaluating the predictive value	e of BRCA mutation/HRR mutation/HRD status.
---------	--	--	---

Study	Phase	Treatments	Target population	DDRm pro- spective selection	Primary endpoint	Findings	Reference
PARPi (single a	agent)				, 		
PROfound	III	Two arms (2:1): Olaparib 300 mg BID vs physician's choice of enzalutamide or abiraterone	mCRPC post NHA	Yes	rPFS in Cohort A (<i>BRCA1/2</i> m and/or <i>ATM</i> m) by BICR	HR 0.34 (95% CI 0.25– 0.47), median 7.4 vs 3.6 months	(25)
TRITON-3	III	Two arms (2:1): Rucaparib 600 mg BID vs physician's choice of docetaxel, enzalutamide or abiraterone	mCRPC post NHA	Yes	rPFS by BICR	BRCA subgroup: HR 0.50 (95% CI 0.36–0.69), 11.2 vs 6.4 months; in the total population: HR 0.61; (95% CI 0.47– 0.80), median 10.2 vs 6.4 months	(28)
TALAPRO-1	п	Single arm: Talazoparib 1 mg/day	mCRPC post 1-2 cycles TBC and NHA	Yes	Best ORR (RECIST)	ORR: 29.8% (95% CI 21.2–39.6)	(26)
TRITON-2	п	Single arm: Rucaparib 600 mg BID	mCRPC post 1 cycle TBC and NHA	Yes	Confirmed ORR (RECIST)	ORR: 43.5% (95% CI 31.0–56.7)	(29)
GALAHAD	II	Single arm: Niraparib 300 mg/day	mCRPC post 1 cycle TBC and NHA	Yes	Confirmed ORR in BRCAm (RECIST)	ORR: 34.2% (95% CI 23.7–46.0)	(27)
TOPARP-A	II	Single arm: Olaparib 400 mg BID	mCRPC post docetaxel	No	Composite RR (PSA + RECIST + CTC)	ORR in HRRm: 33% (95% CI 20–48)	(23)
TOPARP-B	II	Two arms (1:1): Olaparib 400 mg BID vs olaparib 300 mg BID	mCRPC post TBC	Yes	Composite RR (PSA + RECIST + CTC)	400 mg: ORR 54% (95% CI 39–69); 300 mg: 37% (95% CI 23–53)	(24)

(Continued)

TABLE 1 Continued

Study	Phase	Treatments	Target population	DDRm pro- spective selection	Primary endpoint	Findings	Reference
PARPi + NHA co	mbination	5					
PROpel	III	Two arms (1:1): Olaparib 300 mg BID + abiraterone (1000 mg) + prednisolne or prednisolone (5 mg) once daily vs placebo + abiraterone (1000 mg) + prednisolone or prednisolone (5 mg) once daily	1L mCRPC (no prior chemo or NHA in metastatic setting)	No	rPFS by investigator	HR: 0.66 (95% CI 0.54– 0.81), median 24.8 vs 16.6 months	(37)
Magnitude	ш	Two arms (1:1): Niraparib 200 mg + abiraterone (1000 mg) + prednisone or prednisolone (10 mg) once daily versus placebo + abiraterone (1000 mg) + prednisone or prednisolone (10 mg) once daily	1L mCRPC (no prior chemo or <4 months prior NHA in metastatic setting)	Yes	rPFS in patients with HRRm, by BICR	HR: 0.73 (95% CI 0.56– 0.96), median 16.5 vs 13.7 months	(38)
TALAPRO-2	ш	Two arms (1:1): Talazoparib 0.5 mg/day + enzalutamide 160 mg once daily vs placebo + enzalutamide 160 mg once daily	1L mCRPC	No	rPFS by BICR	In ITT (all comers) HR: 0.63 (95% CI 0.51– 0.78), median NR vs 21.9 months	(39)
Study 8	п	Two arms (1:1): Olaparib 300 mg BID + abiraterone (1000 mg) + prednisolone (5 mg) versus once daily placebo + abiraterone (1000 mg) + prednisone or prednisolone (5 mg) once daily	mCRPC post docetaxel and no prior NHA in metastatic setting	No	rPFS by investigator	HR: 0.65 (95% CI 0.44– 0.97), median 13.8 vs 8.2 months	(36)

¹L, first line; BICR, blinded independent central review; BID, twice daily; BRCA1/2m, mutations in BRCA1 and BRCA2; CI, confidence interval; CTC, Circulating Tumour Cells; DDRm, DNA Damage Repair gene mutations; HR, hazard ratio; HRRm, homologous recombination repair gene mutations; mCRPC, metastatic castration-resistant prostate cancer; NHA, new hormonal agent; NR, not reached; ORR, objective response rate; PARPi, poly(ADP-ribose) polymerase inhibitors; PSA, prostate-specific antigen; rPFS, radiological progression-free survival; RR, response rate; TBC, Taxane Based Chemotherapy.

Around 10–11% of patients with primary cancer harbour HRR alterations, whereas between 23% and 28% of patients with mCRPCs have loss-of-function mutations in genes involved in the HRR pathway of DDR in the tumour (Table 2).

In the PROfound clinical trial (a randomised, open-label, Phase III trial evaluating the PARP inhibitor olaparib in men with mCRPC who had disease progression while receiving a new hormonal agent, e.g. enzalutamide or abiraterone), an alteration in one or more of 15^2 prespecified HRR genes was detected in 28% of over 4000 patients screened (25). Very similar prevalence of HRRm was also observed in the Phase III PROpel trial (double-blind, Phase III trial of abiraterone)

and olaparib versus abiraterone and placebo in patients with mCRPC in the first-line setting), where patients were enrolled regardless of HRR status; 28% of patients enrolled had alterations in HRR genes (37). Comparable prevalence of HRR/DDR genomic alterations were also observed in other clinical trials and datasets involving advanced prostate cancer tumours. In the TOPARP-A clinical trial (Phase II trial in which patients with mCRPC were treated with olaparib), 33% of patients harboured genomic alterations specific to DDR genes (23). Chung et al. used real-world data from routine prospective genomic profiling and showed that 23% of patients had alterations in genes involved in the HRR pathway (49) and a similar prevalence of HRR gene mutations was also demonstrated by Abida et al. in 429 patients with mCRPC (42).

Mutations in the BRCA genes (*BRCA1* and/or *BRCA2*) are the most prevalent HRR gene mutations in mCRPC (with *BRCA2* more prevalent than *BRCA1*), with *ATM* being the second most

² Although PPP2R2A was originally included in 15 HRR genes, no benefit of olaparib over control therapy was noted in patients with alterations in PPP2R2A in the PROfound trial (84)

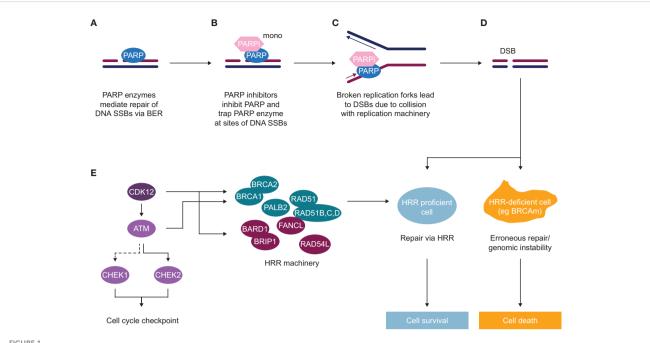


FIGURE 1

Mechanism of action of PARP inhibitors (as monotherapy) in HRR deficient background (A) Poly(ADP-ribose) polymerase (PARP) enzymes are recruited to sites of DNA single-strand breaks (SSBs) and mediate their repair *via* base excision repair (BER). (B) PARP inhibitors inhibit PARP and trap PARP enzyme at sites of SSBs. (C) In replicating cells, due to collision of trapped PARP with replication machinery, broken replication forks occur, which lead to double-strand breaks (DSBs). (D) In normal cells or HRR proficient cells, such DSBs get repaired *via* an accurate DNA repair process of HRR, with some of the key HRR factors depicted in panel (E) In HRR-deficient cells, such as BRCA mutant cells or with alteration in another HRR gene, lack of efficient HRR repair leads to erroneous DNA repair and subsequent genomic instability and cell death. (E) HRR is an accurate DNA repair pathway, which operates in S/G2 cells where the sister chromatid is present. HRR factors include DNA damage sensors and factors (BARD1, BRIP1, FANCL, RAD54L) or regulating HRR gene expression (CDK12), as well as key kinases inducing cell cycle arrest to allow repair to occur (ATM, CHEK1).

frequently mutated (23, 43, 46, 49). In mCRPC, prevalence of BRCAm ranges from around 11% to 13%, and *ATMm* from 4% to 6% (Table 2). The next most prevalent mutations in HRR genes in mCRPC are *CDK12* (1.3–8%), *CHEK2* (1.4–2%), *PALB2* (0.3–3%), and *CHEK1* (0.9–2%) (Table 2). The prevalence of alterations in other HRR genes in prostate cancer is low and ranges between 0% and 1.8% (Table 2).

In summary, between around 23–28% of patients have deleterious alterations in HRR/DDR genes in metastatic prostate cancer, with most studies reporting alterations in the tumour (Table 2) (25, 37, 42, 49)

The origin of alterations in DDR genes found in tumour can be either germline or somatic. The relative ratio of pathogenic germline to somatic mutation events in DDR is highly dependent on the HRR genes interrogated. Lai et al. (52) identified that the ratio of germline to somatic alterations in *BRCA1*/2 genes was roughly 1:1, whereas other HRR genes such as *ATM* and *CDK12* had a much higher ratio of HRRm that were somatic in origin relative to germline with 70% of *ATM* alterations and 89% of *CDK12* alterations being somatic, based on a validated computational algorithm (77). The prevalence of germline alterations in HRR genes in mCRPC is 12% (78), with 6% in *BRCA1/2, ATM* 1.6%, *CHEK2* 1.9% and other HRR genes below 1%. In primary prostate cancer the overall prevalence of germline DDR alterations is lower at 4.6%, with highest prevalence of 1% in *ATM*, and 0.6% in *BRCA1* and 0.2% in *BRCA2* (78).

Loss of function of both alleles is needed for inactivation of HRR gene function. Biallelic loss-of-function rate of HRR gene mutations in prostate cancer is high at 73%, with the highest rate in *BRCA2* (>90%) and *ATM* (around 75%); beyond these, the rate is variable (ranging between under 10% for *BRIP1* and over 60% for *CDK12*) (52), suggesting alterations in HRR genes are important drivers of tumourigenesis for prostate cancer. Biallelic inactivation rate was high for both germline and somatic BRCA alterations (52, 79), suggesting both play an important role in prostate cancer tumourigenesis.

Beyond HRR gene alterations described above, epigenetic silencing of another HRR gene – XRCC3 – has been reported in prostate adenocarcinoma (80). XRCC3 is one of the RAD51 paralogues and its role in HRR has been previously reported (81). Interestingly, XRCC3 alterations were mutually exclusive with alterations in BRCA genes in a pan-cancer dataset, similar to other HRR genes, which confirms its functional importance in the HRR pathway. Depletion of this gene sensitised cells to PARP inhibition preclinically (80), which warrants further investigation of loss of XRCC3 expression as a potential biomarker for PARPi sensitivity in clinic.

TABLE 2 Prevalence of HRRm in primary and metastatic prostate cancer.

Gene	Prevalence range primary PC (%)	Prevalence range in mCRPC (%)	References
BRCA1/ BRCA2	2.5-6.5	11-13	
ATM	0.5-3	4-6	
BRCA1/ 2/ATM	6-7	13–19	
CDK12	2-3	1.3-8	
CHEK2	0-1	1.4–2	
PALB2	0-1	0.3–3	
RAD51B	0	0-0.7	(23, 43, 45, 46, 49, 51)
RAD51C	0-0.9	0-1.8	
RAD51D	0-0.6	0-0.6	
RAD54L	0	0-0.6	
BRIP1	0-0.3	0-0.3	
CHEK1	0-1	0.9–2	
BARD1	0-0.4	1.2–1.4	
FANCL	0-0.7	1.2	
HRRm	10-11	23–28	

Pathogenic/likely pathogenic or deleterious/likely deleterious alterations were included, and variants of uncertain significance were excluded where possible. Data from available public sources may differ in the determination of pathogenicity of mutations. Some of the datasets (42, 43, 59) have been reanalysed by AstraZeneca using cBioPortal (60). Co-occurring HRR gene mutations were excluded from analysis of total HRRm prevalence where possible. Some of the germline variants were excluded in MSK-IMPACT and MSK-DFCI datasets (43, 59) due to patients consent.

HRRm, homologous recombination repair gene mutations; mCRPC, metastatic castrationresistant prostate cancer; PC, prostate cancer.

Genomic instability/HRD in prostate cancer

A consequence of deficiency in the HRR pathway (HRD) is the accumulation of DNA damage leading to genomic instability signatures, or scars, over time. In addition to alterations in HRR genes, HRD/genomic instability can be another way to identify patients who might benefit from PARP inhibitors. HRD has been associated with a clinical benefit for PARP inhibitor treatment in ovarian cancer (82–85) and to platinum-based chemotherapy in breast cancer (86); however, there is lack of evidence of clinical utility of HRD in other tumour types.

In prostate cancer, genomic instability/HRD (as measured by genome-wide loss of heterozygosity [gLOH]) is generally lower than in ovarian, breast or pancreatic cancers (52, 79). Biallelic alterations in BRCA genes or HRR genes are associated with higher gLOH/HRD scores compared with BRCA wild type (wt) or HRR wt prostate cancer tumours, respectively, which is consistent with these alterations leading to deficiency in the HRR pathway (52, 79). However, there are no data on the clinical utility of gLOH/HRD scores in prostate cancer. The distinction between HRD-positive and HRD-negative tumours, based on genomic instability in prostate cancer, is not clear relative to ovarian and breast cancer. The cut-off to identify HRD-positive tumours in prostate cancer

would need to be robustly defined, and this is likely to be more challenging than in ovarian cancer due to a lower dynamic range in prostate cancer. Interestingly, Zurita et al. assessed the relationship between genome instability and clinical features and identified that higher genomic instability was associated with higher risk of disease progression to CRPC (87). Recently, a functional biomarker of HRR has been developed based on measuring nuclear foci formed by the key HRR factor, RAD51, at DNA damage sites, which is currently being implemented in clinical trials (88).

Beyond HRR, alterations in other genes in the DDR pathway are relatively low, however some are potentially actionable. Deficiency in MMR is the underlying cause of the microsatellite instability-high (MSI-H) phenotype, which is a biomarker of response to immune checkpoint blockade therapy (89). Prevalence of alterations in the MMR pathway (*MLH1, MSH2, MSH6, PMS2*) or MSI-H in prostate cancer is much lower than HRRm, at around 3–5% (49). Interestingly, mutual exclusivity between genomic instability/HRD and MSI-H has been reported across tumours (79, 90). In prostate cancer, 12.8% of *BRCA1* and 3.4% of *BRCA2* alterations co-occurred with MSI-H, and 46.3% of MSI-H had at least one HRR gene mutation; however, most *BRCA* mutations in the MSI-H segment were monoallelic (90, 91).

Increased prevalence of HRR alterations in metastatic vs primary/early prostate cancer

Enrichment of BRCA1/2, ATM and CDK12 mutations in advanced prostate cancer has been documented in the literature by several studies (43, 45, 49, 92). Armenia et al. performed a large study that analysed 680 primary tumours and 333 metastatic samples and identified HRR defects in 10% and 27% of the primary and metastatic samples, respectively (43). Similarly, Abida et al. (45) observed an increase in HRR alterations according to disease progression, with 10% of HRR alterations detected in primary tumours, 14% in castration-sensitive prostate cancer and 27% in CRPC. The high representation of BRCA2 mutations in advanced/metastatic prostate cancer is considered to be a consequence of BRCA2 mutations being associated with a particularly aggressive phenotype (49, 93-95) rather than these mutations (e.g. androgen receptor mutations and amplifications) being acquired under treatment with standard therapies (96). An increase in the prevalence of DDR alterations in metastatic compared with primary prostate cancer could either be due to disease progression or therapy exposure, or could be due to a worse prognosis for patients with mutations/alterations in DDR genes (DDRm) prostate cancer. Recent analysis of paired tumour samples from patients with prostate cancer (treatment-naïve and metastatic tumour samples) showed that in most cases, alterations in DDR genes were already present in the primary prostate cancer sample, suggesting that this is an early event in tumourigenesis (97). On the contrary, alterations of AR/TP53/RB1 are enriched at later disease stages (97). These data suggest that genomic instability associated with alterations in HRR genes leads to a more aggressive disease, which is more likely to metastasise, highlighting the need to treat those patients early.

Increased prostate cancer risk for germline alterations in DDR genes

Family history is an important risk factor to be considered for development of prostate cancer. Germline alterations in MMR genes (*MLH1, MSH2, MSH6, PMS2*) and the HRR pathway (*BRCA1/2, ATM, PALB2, CHEK2*), particularly *BRCA* and *ATM*, are associated with increased risk of hereditary prostate cancer, as reported by National Comprehensive Cancer Network (NCCN) guidelines (98). The proportion of patients with prostate cancer with germline mutations in DDR genes increases from around 5% in primary cancer to 12–16% in mCRPC, indicating a more aggressive nature of disease with germline DDR alterations (46, 78, 99).

Men with germline pathogenic *BRCA1/2* mutations have an increased risk of prostate cancer and the relative increase in risk of prostate cancer in men <65 years ranges from 1.8-fold to 3.8-fold for germline *BRCA1m* carriers (100, 101) and from 2.5-fold to 8.6-fold for germline *BRCA2m* carriers (102–105) compared with non-carriers. A large meta-analysis of 8 cohort, 7 case control, 4 case series, 28 frequency and 11 survival studies found that being a *BRCA* mutation carrier (*BRCA1* and/or *BRCA2*) was associated with a significant increase in prostate cancer risk (odds ratio [OR] =1.90; 95% CI 1.58, 2.29), with *BRCA2* mutations being associated with a greater risk of prostate cancer than *BRCA1* (106) (Table 3). Lifetime risk of cancer ranged between 19–61%

for BRCA2 carriers, and 7-26% for BRCA1 carriers (108). A prospective study, IMPACT, which is evaluating targeted screening using prostate-specific antigen (PSA) in men with germline pathogenic BRCA1/2 mutations, has reported a higher incidence of prostate cancer in BRCA2 carriers compared with controls at interim analysis (114). Furthermore, germline pathogenic BRCA2 mutations are associated with a particularly aggressive phenotype and poor outcomes (93, 94). Germline pathogenic mutation status of BRCA and ATM distinguishes risk for lethal and indolent prostate cancer and is associated with earlier age at death and shorter survival time (115). Germline pathogenic variants in ATM lead to an approximate fourfold elevated risk of developing prostate cancer, and in addition, they are more likely to develop the disease earlier (109, 116). Beyond BRCA genes, there is evidence of elevated prostate cancer risk for CHEK2 heterozygotes (110, 111) and increased incidence of germline pathogenic CHEK2, BRIP1 and PALB2 mutations in familial cases of prostate cancer (Table 3) (64, 112, 117).

In addition to prostate cancer, HRR genes are associated with increased risk of breast, ovarian and pancreatic cancer (*BRCA1*, *BRCA2*, *ATM*, *BRIP1*, *PALB2*, *BARD1*, *CHEK2*, *RAD51C* and *RAD51D*). Alterations in some of the DDR genes are associated with cancer predisposition syndromes (e.g. *BRCA1*, *BRCA2*, *ATM*, *FANCL*, *PALB2*, *RAD51C*, *BRIP1*, FA genes, MMR genes) (64, 100, 101, 110, 115, 116, 118–127). Germline pathogenic mutations in MMR genes (*MLH1*, *MSH2*, *MSH6* and *PMS2*) are the key cause of

TABLE 3 Hereditary	cancer	risk for	HRR	genes.
--------------------	--------	----------	-----	--------

Gene name	Prostate cancer risk					
	RR or OR (95% CI)	References				
BRCA1	RR 1.8–3.8	(101, 108, 100)				
	OR 1.35 (1.03–1.76) OR 1.83 (0.35–9.51)	(106, 107)				
BRCA2	RR 2.5–8.6	(102–105, 108)				
	OR 2.64 (2.03–3.47) OR 3.92 (1.34–11.47)	(106, 107)				
ATM	OR 4.4 (2.0-9.5) OR 3.83 (1.09-13.41)	(107, 109)				
CHEK2	Higher risk suggested (OR 1.837) increased incidence of <i>CHEK2</i> mutations in family cases compared with unselected cases and the general population OR 0.47 (0.15–1.45)	(107, 110, 111)				
PALB2	Family segregation case	(112)				
BRIP1	Increased prevalence in familial and young-onset cases compared with controls OR 2.4 (0.25, 23.4)	(105, 113)				
RAD51C	Unknown					
RAD51D	Unknown					
BARD1	OR 0.32 (0.03–3.55)	(107)				
CDK12 ^a , CHEK1, RAD54L, RAD51B, FANCL ^b	Unknown					

^aCDK12, not likely to be associated as very few/no germline mutations; most are somatic; ^bFANCL, associated with cancer predisposition syndrome Fanconi anaemia. CI, confidence interval; HRR, homologous recombination repair; OR, odds ratio; RR, relative risk, SRR, standardised relative risk.

Lynch syndrome (LS) – an inherited cancer predisposition syndrome leading to increased risk of particularly colorectal cancer and other LS-associated cancers. Patients with LS have a twofold elevated risk of incidence of prostate cancer compared with the general population (128).

Prognosis for HRRm prostate cancer

Among HRR alterations, germline pathogenic BRCA mutations in particular are associated with a more aggressive prostate cancer and worse outcomes for patients with localised prostate cancer and mCRPC compared to non-carriers. The presence of a germline pathogenic BRCA mutation in prostate cancer is associated with a more aggressive phenotype, such as higher Gleason scores, nodal involvement and presence of metastases at diagnosis, as shown in large retrospective analyses (93, 94). BRCA2 was reported to be an independent prognostic factor in a multivariate analysis, where patients with germline pathogenic BRCA mutations had overall worse outcomes than non-BRCAm patients in a cohort with localised disease as well as in the overall cohort (93, 94). Another study reported that the combined rate of germline BRCA/ATM alterations was significantly higher in patients with lethal prostate cancer than in patients with localised prostate cancer, and patients with germline BRCA/ATM alterations with either localised disease or a diagnosis with metastases, had a shorter prostate-cancerspecific survival compared with non-carriers (129).

Once prostate cancer becomes castration-resistant and progresses to a metastatic stage (mCRPC) the disease is not curable and treatment must focus on extending life, delaying disease progression and improving quality of life (130, 131). Germline mutations in HRRm have been found in around 12% of patients with mCRPC (78). There has been conflicting evidence reported for the association of HRRm and prognosis for patients with mCRPC. Annala et al. reported worse outcomes for germline DDR carriers (17/22 were gBRCAm) compared with non-carriers when treated with first-line androgen receptor signaling inhibitors (132). However, more recently Antonarakis et al. reported that patients with BRCA/ATM mutations (n=9) do better on first-line NHA than those without mutations in these genes (133). These findings are based on a relatively small number of patients with HRRm. A retrospective analysis showed similar OS outcomes for patients with metastatic prostate cancer and with germline mutations in DDR genes compared with those without; however, somatic alterations in DDR genes were not assessed, and a significant proportion of patients were treated with PARPi or platinum-based chemotherapy, which might have contributed to better outcomes for patients with DDRm (50, 96). A prospective study, PRO-REPAIR, evaluated prevalence of germline DDR mutations and their impact on outcomes for patients with mCRPC (99). This trial enrolled unselected patients with mCRPC and screened for germline alterations in 107 DDR genes with the primary objective to assess the impact of germline alterations in ATM/BRCA1/BRCA2/PALB2 on cause-specific survival (CSS) from diagnosis of mCRPC. In PRO-REPAIR, 16% of patients had a germline mutation in a HRR gene (most commonly BRCA and *ATM* alterations). Although numerically the CSS was shorter in patients with *ATM/BRCA1/BRCA2*, the difference was not statistically significant; however, patients with germline *BRCA2* mutation had considerably shorter CSS than non-carriers and *BRCA2* was an independent prognostic factor, where sequence and type of treatment may impact the outcomes (99). Castro et al. reported that treatment sequence is important for prognosis, with patients with mCRPC and DDR mutations having worse outcomes overall but better outcomes following first-line NHA treatment (99). These observations might explain conflicting observations in different studies. In addition to alterations in HRR genes, genomic instability might also be associated with worse outcomes, such as with risk of biochemical recurrence and metastases (134, 135).

Treatment options for patients with DDRm prostate cancer

High unmet clinical need and poor prognosis for patients with prostate cancer and DDR has triggered an active development of targeted treatment options for these patients (136). As described above, based on the principle of synthetic lethality, cells deficient in the HRR pathway are sensitive to PARP inhibitors (137, 138) (Figure 1). PARP inhibitors as monotherapy have demonstrated efficacy in patients with evidence of deficiency in the HRR pathway in the tumour and have been approved for treatment of HRRm mCRPC (136). The clinical benefit of PARPi in combination with NHA has been demonstrated in biomarker-selected and biomarker-unselected populations in first-line mCRPC (Table 1) (36–39), with approval in some regions, providing additional efficacious treatment options for these patients.

Platinum-based chemotherapy also leads to DNA damage, which is repaired by HRR pathway. Increased platinum sensitivity in tumours deficient in HRR pathway (ie BRCAm) has been reported in other tumour types, such as breast or ovarian cancer (139, 140) Retrospective analyses indicated encouraging antitumour activity of platinum-based chemotherapy in advanced prostate cancer patients with BRCA alterations or some DDR alterations with higher PSA response rates in patients with DDR/ BRCA alterations compared to patients without (141–143), although these findings need to be validated in a prospective setting.

ATM-deficient cells are dependent on ATR activity, which leads to sensitivity to ATR inhibition preclinically; clinical trials with ATR inhibitors as monotherapy or in combinations are ongoing, including in biomarker-selected patients with ATM- or DDRdeficiency in advanced solid tumours, including prostate cancer, as reviewed recently by Ngoi et al. (144).

Anti-PD1 antibody (Pembrolizumab) is approved by the Food and Drug Administration (FDA) to treat cancers with MMR mutations or MSI-H, including prostate cancer (145, 146). *CDK12* inactivation results in tandem duplications in the genome leading to increased fusions and mutations and might lead to increased antigens. Clinical trials with immune checkpoint inhibitors are ongoing for patients with *CDK12*m and advanced prostate cancer (147–149).

Conclusions/takeaway

Prostate cancer is the third most common cancer worldwide, and metastatic prostate cancer is associated with poor outcomes and high mortality. There are various genomic alterations commonly associated with prostate cancer, and alterations in the HRR pathway of DDR are prevalent in prostate cancer, ranging from 23-28% (25, 42, 43, 46, 78). Germline alterations in several HRR genes, such as BRCA, ATM and others, are associated with increased prostate cancer risk, and are generally associated with worse prognosis for patients with prostate cancer. Alterations in DDR genes in tumours tend to be an early event in prostate tumourigenesis and are associated with more aggressive disease and likelihood of metastasis (97). With current breakthroughs regarding targeted treatments, PARP inhibitors as monotherapy are an option for patients with HRR alterations in mCRPC, who have progressed on NHA (136) and in some countries, are available in combination with NHA in a broad population of 1L mCRPC (36, 55, 99). Clinical trials are ongoing that are evaluating ATR inhibitors in all solid tumours, including prostate cancer, as reviewed by Ngoi et al. (144). Given the association of some DDR genes with worse prognosis, and DDR alterations being an early event in prostate cancer, future clinical trials for patients with DDRm and an earlier stage of disease are important to improve outcomes for these patients.

Author contributions

NL and AB developed the initial draft of the manuscript. NL, AB, EH, CA and JCB made substantial contributions to the concept

References

 Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin (2021) 71:209–49. doi: 10.3322/caac.21660

2. Rawla P. Epidemiology of prostate cancer. World J Oncol (2019) 10:63–89. doi: 10.14740/wjon1191

3. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2018. CA Cancer J Clin (2018) 68:7-30. doi: 10.3322/caac.21442

4. Rebello RJ, Oing C, Knudsen KE, Loeb S, Johnson DC, Reiter RE, et al. Prostate cancer. *Nat Rev Dis Primers* (2021) 7:9. doi: 10.1038/s41572-020-00243-0

5. Kirby M, Hirst C, Crawford ED. Characterising the castration-resistant prostate cancer population: a systematic review. *Int J Clin Pract* (2011) 65:1180–92. doi: 10.1111/j.1742-1241.2011.02799.x

6. Ku S-Y, Gleave ME, Beltran H. Towards precision oncology in advanced prostate cancer. *Nat Rev Urol* (2019) 16:645–54. doi: 10.1038/s41585-019-0237-8

7. Ng K, Smith S, Shamash J. Metastatic hormone-sensitive prostate cancer (mHSPC): advances and treatment strategies in the first-line setting. *Oncol Ther* (2020) 8:209–30. doi: 10.1007/s40487-020-00119-z

8. Beer TM, Armstrong AJ, Rathkopf D, Loriot Y, Sternberg CN, Higano CS, et al. Enzalutamide in men with chemotherapy-naïve metastatic castration-resistant prostate cancer: extended analysis of the phase 3 PREVAIL study. *Eur Urol* (2017) 71:151–4. doi: 10.1016/j.eururo.2016.07.032

9. Ryan CJ, Smith MR, Fizazi K, Saad F, Mulders PFA, Sternberg CN, et al. Abiraterone acetate plus prednisone versus placebo plus prednisone in chemotherapynaive men with metastatic castration-resistant prostate cancer (COU-AA-302): final overall survival analysis of a randomised, double-blind, placebo-controlled phase 3 study. *Lancet Oncol* (2015) 16:152–60. doi: 10.1016/S1470-2045(14)71205-7 and content of review. JA, JK and CA contributed to writing parts of the manuscript. All authors contributed to the article and approved the submitted version.

Funding

This manuscript was funded by AstraZeneca.

Acknowledgments

We thank Mudskipper Business LTD medical writing agency for editing support and review of the manuscript. This support was funded by AstraZeneca

Conflict of interest

NL, AB, CA, JA, JK, JCB and EH are employees of, and hold stock at AstraZeneca.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

10. de Bono JS, Smith MR, Saad F, Rathkopf DE, Mulders PFA, Small EJ, et al. Subsequent chemotherapy and treatment patterns after abiraterone acetate in patients with metastatic castration-resistant prostate cancer: *Post hoc* analysis of COU-AA-302. *Eur Urol* (2017) 71:656–64. doi: 10.1016/j.eururo.2016.06.033

11. George DJ, Sartor O, Miller K, Saad F, Tombal B, Kalinovský J, et al. Treatment patterns and outcomes in patients with metastatic castration-resistant prostate cancer in a real-world clinical practice setting in the united states. *Clin Genitourin Cancer* (2020) 18:284–94. doi: 10.1016/j.clgc.2019.12.019

12. Flaig TW, Potluri RC, Ng Y, Todd MB, Mehra M. Treatment evolution for metastatic castration-resistant prostate cancer with recent introduction of novel agents: retrospective analysis of real-world data. *Cancer Med* (2016) 5:182–91. doi: 10.1002/cam4.576

13. Ryan CJ, Smith MR, de Bono JS, Molina A, Logothetis CJ, de Souza P, et al. Abiraterone in metastatic prostate cancer without previous chemotherapy. *New Engl J Med* (2013) 368:138–48. doi: 10.1056/NEJMoa1209096

14. Beer TM, Armstrong AJ, Rathkopf DE, Loriot Y, Sternberg CN, Higano CS, et al. Enzalutamide in metastatic prostate cancer before chemotherapy. *New Engl J Med* (2014) 371:424–33. doi: 10.1056/NEJMoa1405095

15. Mezynski J, Pezaro C, Bianchini D, Zivi A, Sandhu S, Thompson E, et al. Antitumour activity of docetaxel following treatment with the CYP17A1 inhibitor abiraterone: clinical evidence for cross-resistance? *Ann Oncol* (2012) 23:2943–7. doi: 10.1093/annonc/mds119

16. Azad AA, Eigl BJ, Leibowitz-Amit R, Lester R, Kollmannsberger C, Murray N, et al. Outcomes with abiraterone acetate in metastatic castration-resistant prostate cancer patients who have poor performance status. *Eur Urol* (2015) 67:441–7. doi: 10.1016/j.eururo.2014.01.030

17. Schweizer MT, Zhou XC, Wang H, Bassi S, Carducci MA, Eisenberger MA, et al. The influence of prior abiraterone treatment on the clinical activity of docetaxel in men

with metastatic castration-resistant prostate cancer. Eur Urol (2014) 66:646-52. doi: 10.1016/j.eururo.2014.01.018

18. Cheng HH, Gulati R, Azad A, Nadal R, Twardowski P, Vaishampayan UN, et al. Activity of enzalutamide in men with metastatic castration-resistant prostate cancer is affected by prior treatment with abiraterone and/or docetaxel. *Prostate Cancer Prostatic Dis* (2015) 18:122–7. doi: 10.1038/pcan.2014.53

19. Suzman DL, Luber B, Schweizer MT, Nadal R, Antonarakis ES. Clinical activity of enzalutamide versus docetaxel in men with castration-resistant prostate cancer progressing after abiraterone. *Prostate* (2014) 74:1278–85. doi: 10.1002/pros.22844

20. Moreira RB, Debiasi M, Francini E, Nuzzo PV, Velasco G, Maluf FC, et al. Differential side effects profile in patients with mCRPC treated with abiraterone or enzalutamide: a meta-analysis of randomized controlled trials. *Oncotarget* (2017) 8:84572–8. doi: 10.18632/oncotarget.20028

21. Parker C, Heidenreich A, Nilsson S, Shore N. Current approaches to incorporation of radium-223 in clinical practice. *Prostate Cancer Prostatic Dis* (2018) 21:37–47. doi: 10.1038/s41391-017-0020-y

22. Den RB, George D, Pieczonka C, McNamara M. Ra-223 treatment for bone metastases in castrate-resistant prostate cancer. *Am J Clin Oncol* (2019) 42:399–406. doi: 10.1097/COC.00000000000528

23. Mateo J, Carreira S, Sandhu S, Miranda S, Mossop H, Perez-Lopez R, et al. DNA-Repair defects and olaparib in metastatic prostate cancer. *New Engl J Med* (2015) 373:1697–708. doi: 10.1056/NEJMoa1506859

24. Mateo J, Porta N, McGovern UB, Elliott T, Jones RJ, Syndikus I, et al. TOPARPb: a phase II randomized trial of the poly(ADP)-ribose polymerase (PARP) inhibitor olaparib for metastatic castration resistant prostate cancers (mCRPC) with DNA damage repair (DDR) alterations. *J Clin Oncol* (2019) 37:5005–5. doi: 10.1200/ JCO.2019.37.15_suppl.5005

25. de Bono J, Mateo J, Fizazi K, Saad F, Shore N, Sandhu S, et al. Olaparib for metastatic castration-resistant prostate cancer. *New Engl J Med* (2020) 382:2091–102. doi: 10.1056/NEJMoa1911440

26. de Bono JS, Mehra N, Scagliotti GV, Castro E, Dorff T, Stirling A, et al. Talazoparib monotherapy in metastatic castration-resistant prostate cancer with DNA repair alterations (TALAPRO-1): an open-label, phase 2 trial. *Lancet Oncol* (2021) 22:1250–64. doi: 10.1016/S1470-2045(21)00376-4

27. Smith MR, Scher HI, Sandhu S, Efstathiou E, Lara PN, Yu EY, et al. Niraparib in patients with metastatic castration-resistant prostate cancer and DNA repair gene defects (GALAHAD): a multicentre, open-label, phase 2 trial. *Lancet Oncol* (2022) 23:362–73. doi: 10.1016/S1470-2045(21)00757-9

28. Fizazi K, Piulats JM, Reaume MN, Ostler P, McDermott R, Gingerich JR, et al. Rucaparib or physician's choice in metastatic prostate cancer. *New Engl J Med* (2023) 388:719–32. doi: 10.1056/NEJMoa2214676

29. Abida W, Patnaik A, Campbell D, Shapiro J, Bryce AH, McDermott R, et al. Rucaparib in men with metastatic castration-resistant prostate cancer harboring a *BRCA1* or *BRCA2* gene alteration. *J Clin Oncol* (2020) 38:3763–72. doi: 10.1200/ JCO.20.01035

30. Clovis Oncology, Inc. TRITON3 phase 3 trial of rubraca[®] (rucaparib) achieves primary endpoint in men with metastatic castration-resistant prostate cancer with BRCA or ATM mutations. Available at: https://ir.clovisoncology.com/investors-and-news/news-releases/press-release-details/2022/TRITON3-Phase-3-Trial-of-Rubraca-rucaparib-Achieves-Primary-Endpoint-in-Men-with-Metastatic-Castration-Resistant-Prostate-Cancer-with-BRCA-or-ATM-Mutations/default.aspx (Accessed December 21, 2022).

31. Hussain M, Mateo J, Fizazi K, Saad F, Shore N, Sandhu S, et al. Survival with olaparib in metastatic castration-resistant prostate cancer. *New Engl J Med* (2020) 383:2345–57. doi: 10.1056/NEJMoa2022485

32. Westaby D, Fenor de la Maza M de LD, Paschalis A, Jimenez-Vacas JM, Welti J, de Bono J, et al. A new old target: androgen receptor signaling and advanced prostate cancer. *Annu Rev Pharmacol Toxicol* (2022) 62:131–53. doi: 10.1146/annurev-pharmtox-052220-015912

33. Tarish FL, Schultz N, Tanoglidi A, Hamberg H, Letocha H, Karaszi K, et al. Castration radiosensitizes prostate cancer tissue by impairing DNA double-strand break repair. *Sci Transl Med* (2015) 7:312re11. doi: 10.1126/scitranslmed.aac5671

34. Goodwin JF, Schiewer MJ, Dean JL, Schrecengost RS, de Leeuw R, Han S, et al. A hormone–DNA repair circuit governs the response to genotoxic insult. *Cancer Discovery* (2013) 3:1254–71. doi: 10.1158/2159-8290.CD-13-0108

35. Polkinghorn WR, Parker JS, Lee MX, Kass EM, Spratt DE, Iaquinta PJ, et al. Androgen receptor signaling regulates DNA repair in prostate cancers. *Cancer Discovery* (2013) 3:1245–53. doi: 10.1158/2159-8290.CD-13-0172

36. Clarke N, Wiechno P, Alekseev B, Sala N, Jones R, Kocak I, et al. Olaparib combined with abiraterone in patients with metastatic castration-resistant prostate cancer: a randomised, double-blind, placebo-controlled, phase 2 trial. *Lancet Oncol* (2018) 19:975–86. doi: 10.1016/S1470-2045(18)30365-6

37. Clarke NW, Armstrong AJ, Thiery-Vuillemin A, Oya M, Shore N, Loredo E, et al. Abiraterone and olaparib for metastatic castration-resistant prostate cancer. *NEJM Evidence* (2022) 1(9). doi: 10.1056/EVIDOA2200043

38. Chi KN, Rathkopf DE, Smith MR, Efstathiou E, Attard G, Olmos D, et al. Phase 3 MAGNITUDE study: first results of niraparib (NIRA) with abiraterone acetate and prednisone (AAP) as first-line therapy in patients (pts) with metastatic castrationresistant prostate cancer (mCRPC) with and without homologous recombination repair (HRR) gene alterations. J Clin Oncol (2022) 40:12-2. doi: 10.1200/ JCO.2022.40.6_suppl.012

39. Agarwal N, Azad AA, Carles J, Fay AP, Matsubara N, Heinrich D, et al. Talazoparib plus enzalutamide in men with first-line metastatic castration-resistant prostate cancer (TALAPRO-2): a randomised, placebo-controlled, phase 3 trial. *Lancet* (2023). doi: 10.1016/S0140-6736(23)01055-3

40. Lynparza in combination with abiraterone approved in the EU as 1st-line treatment for patients with metastatic castration-resistant prostate cancer . Available at: https://www.astrazeneca.com/media-centre/press-releases/2022/lynparza-approved-in-eu-for-prostate-cancer.html (Accessed March 21, 2023).

41. Liu JM, Lin CC, Liu KL, Lin CF, Chen BY, Chen TH, et al. Second-line hormonal therapy for the management of metastatic castration-resistant prostate cancer: a real-world data study using a claims database. *Sci Rep 2020 10:1* (2020) 10:1–7. doi: 10.1038/ s41598-020-61235-4

42. Abida W, Cyrta J, Heller G, Prandi D, Armenia J, Coleman I, et al. Genomic correlates of clinical outcome in advanced prostate cancer. *Proc Natl Acad Sci* (2019) 116:11428–36. doi: 10.1073/pnas.1902651116

43. Armenia J, Wankowicz SAM, Liu D, Gao J, Kundra R, Reznik E, et al. The long tail of oncogenic drivers in prostate cancer. *Nat Genet* (2018) 50:645–51. doi: 10.1038/s41588-018-0078-z

44. Abeshouse A, Ahn J, Akbani R, Ally A, Amin S, Andry CD, et al. The molecular taxonomy of primary prostate cancer. *Cell* (2015) 163:1011–25. doi: 10.1016/j.cell.2015.10.025

45. Abida W, Armenia J, Gopalan A, Brennan R, Walsh M, Barron D, et al. Prospective genomic profiling of prostate cancer across disease states reveals germline and somatic alterations that may affect clinical decision making. *JCO Precis Oncol* (2017) 1:1–16. doi: 10.1200/PO.17.00029

46. Robinson D, Van Allen EM, Wu Y-M, Schultz N, Lonigro RJ, Mosquera J-M, et al. Integrative clinical genomics of advanced prostate cancer. *Cell* (2015) 161:1215–28. doi: 10.1016/j.cell.2015.05.001

47. Jiang M, Jia K, Wang L, Li W, Chen B, Liu Y, et al. Alterations of DNA damage repair in cancer: from mechanisms to applications. *Ann Transl Med* (2020) 8:1685–5. doi: 10.21037/atm-20-2920

48. Lord CJ, Ashworth A. The DNA damage response and cancer therapy. *Nature* (2012) 481:287–94. doi: 10.1038/nature10760

49. Chung JH, Dewal N, Sokol E, Mathew P, Whitehead R, Millis SZ, et al. Prospective comprehensive genomic profiling of primary and metastatic prostate tumors. *JCO Precis Oncol* (2019) 3:1-23. doi: 10.1200/PO.18.00283

50. Cresta Morgado P, Mateo J. Clinical implications of homologous recombination repair mutations in prostate cancer. *Prostate* (2022) 82:S45–59. doi: 10.1002/pros.24352

51. de Bono JS, Fizazi K, Saad F, Shore N, Sandhu SK, Mehra N, et al. Central, prospective detection of homologous recombination repair gene mutations (HRRm) in tumour tissue from >4000 men with metastatic castration-resistant prostate cancer (mCRPC) screened for the PROfound study. *Ann Oncol* (2019) 30:v328–9. doi: 10.1093/ANNONC/MDZ248.004

52. Lai Z, Brosnan M, Sokol ES, Xie M, Dry JR, Harrington EA, et al. Landscape of homologous recombination deficiencies in solid tumours: analyses of two independent genomic datasets. *BMC Cancer* (2022) 22:13. doi: 10.1186/s12885-021-09082-y

53. Blackford A, Jackson SP. ATM, ATR, and DNA-PK: The trinity at the heart of the DNA damage response. *Mol Cell* (2017) 66:801–17. doi: 10.1016/j.molcel.2017.05.015

54. Pilié PG, Tang C, Mills GB, Yap TA. State-of-the-art strategies for targeting the DNA damage response in cancer. *Nat Rev Clin Oncol* (2019) 16:81–104. doi: 10.1038/ s41571-018-0114-z

55. Lang SH, Swift SL, White H, Misso K, Kleijnen J, Quek RGW. A systematic review of the prevalence of DNA damage response gene mutations in prostate cancer. *Int J Oncol* (2019) 55:597–616. doi: 10.3892/IJO.2019.4842/HTML

56. Thompson LH, Schild D. Homologous recombinational repair of DNA ensures mammalian chromosome stability. *Mutat Research/Fundamental Mol Mech Mutagenesis* (2001) 477:131-53. doi: 10.1016/S0027-5107(01)00115-4

57. Jackson SP, Bartek J. The DNA-damage response in human biology and disease. *Nature* (2009) 461:1071–8. doi: 10.1038/nature08467

58. Mazin AV, Mazina OM, Bugreev DV, Rossi MJ. Rad54, the motor of homologous recombination. DNA Repair (Amst) (2010) 9:286-302. doi: 10.1016/j.dnarep.2009.12.006

59. Abida W, Armenia J, Gopalan A, Brennan R, Walsh M, Barron D, et al. Prospective genomic profiling of prostate cancer across disease states reveals germline and somatic alterations that may affect clinical decision making. *JCO Precis Oncol* (2017) 1:1–16. doi: 10.1200/PO.17.00029

60. Gao J, Aksoy BA, Dogrusoz U, Dresdner G, Gross B, Sumer SO, et al. Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. *Sci Signal* (2013) 6:11. doi: 10.1126/scisignal.2004088

61. Mersch J, Jackson MA, Park M, Nebgen D, Peterson SK, Singletary C, et al. Cancers associated with *BRCA 1* and *BRCA 2* mutations other than breast and ovarian. *Cancer* (2015) 121:269–75. doi: 10.1002/cncr.29041

62. Taylor MRG, Špírek M, Chaurasiya KR, Ward JD, Carzaniga R, Yu X, et al. Rad51 paralogs remodel pre-synaptic Rad51 filaments to stimulate homologous recombination. *Cell* (2015) 162:271–86. doi: 10.1016/j.cell.2015.06.015 63. Bonilla B, Hengel SR, Grundy MK, Bernstein KA. *RAD51* gene family structure and function. *Annu Rev Genet* (2020) 54:25–46. doi: 10.1146/annurev-genet-021920-092410

64. Ramus SJ, Song H, Dicks E, Tyrer JP, Rosenthal AN, Intermaggio MP, et al. Germline mutations in the BRIP1, BARD1, PALB2, and NBN genes in women with ovarian cancer. *JNCI: J Natl Cancer Institute* (2015) 107(11):djv214. doi: 10.1093/jnci/djv214

65. Christou C, Kyriacou K. BRCA1 and its network of interacting partners. *Biol* (*Basel*) (2013) 2:40–63. doi: 10.3390/biology2010040

66. Balmus G, Pilger D, Coates J, Demir M, Sczaniecka-Clift M, Barros AC, et al. ATM Orchestrates the DNA-damage response to counter toxic non-homologous endjoining at broken replication forks. *Nat Commun* (2019) 10:87. doi: 10.1038/s41467-018-07729-2

67. Bartek J, Falck J, Lukas J. Chk2 kinase [[/amp]]mdash; a busy messenger. Nat Rev Mol Cell Biol (2001) 2:877-86. doi: 10.1038/35103059

68. Reinhardt HC, Yaffe MB. Kinases that control the cell cycle in response to DNA damage: Chk1, Chk2, and MK2. *Curr Opin Cell Biol* (2009) 21:245–55. doi: 10.1016/j.ceb.2009.01.018

69. Bartek J, Lukas J. Chk1 and Chk2 kinases in checkpoint control and cancer. *Cancer Cell* (2003) 3:421–9. doi: 10.1016/S1535-6108(03)00110-7

70. Zhang J, Willers H, Feng Z, Ghosh JC, Kim S, Weaver DT, et al. Chk2 phosphorylation of BRCA1 regulates DNA double-strand break repair. *Mol Cell Biol* (2004) 24:708–18. doi: 10.1128/MCB.24.2.708-718.2004

71. Sørensen CS, Hansen LT, Dziegielewski J, Syljuåsen RG, Lundin C, Bartek J, et al. The cell-cycle checkpoint kinase Chk1 is required for mammalian homologous recombination repair. *Nat Cell Biol* (2005) 7:195–201. doi: 10.1038/ncb1212

72. Booth L, Roberts J, Poklepovic A, Dent P. The CHK1 inhibitor SRA737 synergizes with PARP1 inhibitors to kill carcinoma cells. *Cancer Biol Ther* (2018) 19:786–96. doi: 10.1080/15384047.2018.1472189

73. Zhang Y, Hunter T. Roles of Chk1 in cell biology and cancer therapy. Int J Cancer (2014) 134:1013–23. doi: 10.1002/ijc.28226

74. Dubbury SJ, Boutz PL, Sharp PA. CDK12 regulates DNA repair genes by suppressing intronic polyadenylation. *Nature* (2018) 564:141–5. doi: 10.1038/s41586-018-0758-y

75. Bajrami I, Frankum JR, Konde A, Miller RE, Rehman FL, Brough R, et al. Genome-wide profiling of genetic synthetic lethality identifies CDK12 as a novel determinant of PARP1/2 inhibitor sensitivity. *Cancer Res* (2014) 74:287–97. doi: 10.1158/0008-5472.CAN-13-2541

76. Pan E, Cabal A, Javier-DesLoges J, Patel D, Panian J, Lee S, et al. Analysis of *CDK12* alterations in a pan-cancer database. *Cancer Med* (2022) 11:753-63. doi: 10.1002/cam4.4483

77. Sun JX, He Y, Sanford E, Montesion M, Frampton GM, Vignot S, et al. A computational approach to distinguish somatic vs. germline origin of genomic alterations from deep sequencing of cancer specimens without a matched normal. *PloS Comput Biol* (2018) 14:e1005965. doi: 10.1371/JOURNAL.PCBI.1005965

78. Pritchard CC, Mateo J, Walsh MF, De Sarkar N, Abida W, Beltran H, et al. Inherited DNA-repair gene mutations in men with metastatic prostate cancer. *New Engl J Med* (2016) 375:443–53. doi: 10.1056/NEJMoa1603144

79. Sokol ES, Pavlick D, Khiabanian H, Frampton GM, Ross JS, Gregg JP, et al. Pancancer analysis of BRCA1 and *BRCA2* genomic alterations and their association with genomic instability as measured by genome-wide loss of heterozygosity. *JCO Precis Oncol* (2020) 4:442–65. doi: 10.1200/PO.19.00345

80. Jamal K, Galbiati A, Armenia J, Illuzzi G, Hall J, Bentouati S, et al. Drug-gene interaction screens coupled to tumor data analyses identify the most clinically relevant cancer vulnerabilities driving sensitivity to PARP inhibition. *Cancer Res Commun* (2022) 2:1244–54. doi: 10.1158/2767-9764.CRC-22-0119

81. Garcin EB, Gon S, Sullivan MR, Brunette GJ, Cian A, Concordet J-P, et al. Differential requirements for the RAD51 paralogs in genome repair and maintenance in human cells. *PloS Genet* (2019) 15:e1008355. doi: 10.1371/journal.pgen.1008355

82. Ray-Coquard I, Pautier P, Pignata S, Pérol D, González-Martín A, Berger R, et al. Olaparib plus bevacizumab as first-line maintenance in ovarian cancer. *New Engl J Med* (2019) 381:2416–28. doi: 10.1056/NEJMoa1911361

83. Patel JN, Braicu I, Timms KM, Solimeno C, Tshiaba P, Reid J, et al. Characterisation of homologous recombination deficiency in paired primary and recurrent high-grade serous ovarian cancer. *Br J Cancer* (2018) 119:1060–6. doi: 10.1038/s41416-018-0268-6

84. Swisher EM, Lin KK, Oza AM, Scott CL, Giordano H, Sun J, et al. Rucaparib in relapsed, platinum-sensitive high-grade ovarian carcinoma (ARIEL2 part 1): an international, multicentre, open-label, phase 2 trial. *Lancet Oncol* (2017) 18:75–87. doi: 10.1016/S1470-2045(16)30559-9

85. Coleman RL, Oza AM, Lorusso D, Aghajanian C, Oaknin A, Dean A, et al. Rucaparib maintenance treatment for recurrent ovarian carcinoma after response to platinum therapy (ARIEL3): a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet* (2017) 390:1949–61. doi: 10.1016/S0140-6736(17)32440-6

86. Telli ML, Timms KM, Reid J, Hennessy B, Mills GB, Jensen KC, et al. Homologous recombination deficiency (HRD) score predicts response to platinumcontaining neoadjuvant chemotherapy in patients with triple-negative breast cancer. *Clin Cancer Res* (2016) 22:3764–73. doi: 10.1158/1078-0432.CCR-15-2477 87. Zurita AJ, Graf RP, Villacampa G, Raskina K, Sokol E, Jin D, et al. Genomic biomarkers and genome-wide loss-of-Heterozygosity scores in metastatic prostate cancer following progression on androgen-targeting therapies. *JCO Precis Oncol* (2022) 6:e2200195. doi: 10.1200/PO.22.00195

88. Carreira S, Porta N, Arce-Gallego S, Seed G, Llop-Guevara A, Bianchini D, et al. Biomarkers associating with PARP inhibitor benefit in prostate cancer in the TOPARPb trial. *Cancer Discovery* (2021) 11:2812–27. doi: 10.1158/2159-8290.CD-21-0007

89. Le DT, Durham JN, Smith KN, Wang H, Bartlett BR, Aulakh LK, et al. Mismatch repair deficiency predicts response of solid tumors to PD-1 blockade. *Science* (2017) 357:409–13. doi: 10.1126/science.aan6733

90. Westphalen CB, Fine AD, André F, Ganesan S, Heinemann V, Rouleau E, et al. Pan-cancer analysis of homologous recombination repair-associated gene alterations and genome-wide loss-of-Heterozygosity score. *Clin Cancer Res* (2022) 28:1412–21. doi: 10.1158/1078-0432.CCR-21-2096

91. Sokol ES, Jin DX, Fine A, Trabucco SE, Maund S, Frampton G, et al. PARP inhibitor insensitivity to BRCA1/2 monoallelic mutations in microsatellite instability-high cancers. *JCO Precis Oncol* (2022) 6:e2100531. doi: 10.1200/PO.21.00531

92. Mateo J, Boysen G, Barbieri CE, Bryant HE, Castro E, Nelson PS, et al. DNA Repair in prostate cancer: biology and clinical implications. *Eur Urol* (2017) 71:417–25. doi: 10.1016/j.eururo.2016.08.037

93. Castro E, Goh C, Leongamornlert D, Saunders E, Tymrakiewicz M, Dadaev T, et al. Effect of BRCA mutations on metastatic relapse and cause-specific survival after radical treatment for localised prostate cancer. *Eur Urol* (2015) 68:186–93. doi: 10.1016/ j.eururo.2014.10.022

94. Castro E, Goh C, Olmos D, Saunders E, Leongamornlert D, Tymrakiewicz M, et al. Germline *BRCA* mutations are associated with higher risk of nodal involvement, distant metastasis, and poor survival outcomes in prostate cancer. *J Clin Oncol* (2013) 31:1748–57. doi: 10.1200/JCO.2012.43.1882

95. Hatano K, Nonomura N. Genomic profiling of prostate cancer: an updated review. World J Mens Health (2022) 40:368. doi: 10.5534/wjmh.210072

96. Mateo J, Carreira S, Seed G, Chandler R, Dolling D, Figueiredo I, et al. Genomic profiling of primary prostate tumors from patients who develop metastatic castration-resistant prostate cancer (mCRPC). *J Clin Oncol* (2018) 36:5013–3. doi: 10.1200/JCO.2018.36.15_suppl.5013

97. Mateo J, Seed G, Bertan C, Rescigno P, Dolling D, Figueiredo I, et al. Genomics of lethal prostate cancer at diagnosis and castration resistance. *J Clin Invest* (2020) 130:1743–51. doi: 10.1172/JCI132031

98. Schaeffer E, Srinivas S, Antonarakis ES, Armstrong AJ, Bekelman JE, Cheng H, et al. NCCN guidelines insights: prostate cancer, version 1.2021. J Natl Compr Cancer Network (2021) 19:134–43. doi: 10.6004/jnccn.2021.0008

99. Castro E, Romero-Laorden N, Del Pozo A, Lozano R, Medina A, Puente J, et al. PROREPAIR-b: a prospective cohort study of the impact of germline DNA repair mutations on the outcomes of patients with metastatic castration-resistant prostate cancer. *J Clin Oncol* (2019) 37:490–503. doi: 10.1200/JCO.18.00358

100. Leongamornlert D, Mahmud N, Tymrakiewicz M, Saunders E, Dadaev T, Castro E, et al. Germline BRCA1 mutations increase prostate cancer risk. *Br J Cancer* (2012) 106:1697–701. doi: 10.1038/bjc.2012.146

101. Thompson D, Easton F. Cancer incidence in BRCA1 mutation carriers. CancerSpectrum Knowledge Environ (2002) 94:1358-65. doi: 10.1093/jnci/94.18.1358

102. van Asperen CJ. Cancer risks in BRCA2 families: estimates for sites other than breast and ovary. J Med Genet (2005) 42:711–9. doi: 10.1136/jmg.2004.028829

103. Breast cancer linkage consortium. Cancer risks in BRCA2 mutation carriers. JNCI J Natl Cancer Institute (1999) 91:1310–6. doi: 10.1093/jnci/91.15.1310

104. Gallagher DJ, Gaudet MM, Pal P, Kirchhoff T, Balistreri L, Vora K, et al. Germline *BRCA* mutations denote a clinicopathologic subset of prostate cancer. *Clin Cancer Res* (2010) 16:2115–21. doi: 10.1158/1078-0432.CCR-09-2871

105. Kote-Jarai Z, Leongamornlert D, Saunders E, Tymrakiewicz M, Castro E, Mahmud N, et al. BRCA2 is a moderate penetrance gene contributing to young-onset prostate cancer: implications for genetic testing in prostate cancer patients. *Br J Cancer* (2011) 105:1230–4. doi: 10.1038/bjc.2011.383

106. Oh M, Alkhushaym N, Fallatah S, Althagafi A, Aljadeed R, Alsowaida Y, et al. The association of *BRCA1* and *BRCA2* mutations with prostate cancer risk, frequency, and mortality: a meta-analysis. *Prostate* (2019) 79:880–95. doi: 10.1002/pros.23795

107. Matejcic M, Patel Y, Lilyquist J, Hu C, Lee KY, Gnanaolivu RD, et al. Pathogenic variants in cancer predisposition genes and prostate cancer risk in men of African ancestry. *JCO Precis Oncol* (2020) 4:32–43. doi: 10.1200/PO.19.00179

108. Lecarpentier J, Silvestri V, Kuchenbaecker KB, Barrowdale D, Dennis J, McGuffog L, et al. Prediction of breast and prostate cancer risks in Male *BRCA1* and *BRCA2* mutation carriers using polygenic risk scores. *J Clin Oncol* (2017) 35:2240–50. doi: 10.1200/JCO.2016.69.4935

109. Karlsson Q, Brook MN, Dadaev T, Wakerell S, Saunders EJ, Muir K, et al. Rare germline variants in ATM predispose to prostate cancer: a PRACTICAL consortium study. *Eur Urol Oncol* (2021) 4:570–9. doi: 10.1016/j.euo.2020.12.001

110. Wang Y, Dai B, Ye D. CHEK2 mutation and risk of prostate cancer: a systematic review and meta-analysis. Int J Clin Exp Med (2015) 8:15708-15.

111. Cybulski C, Wokolorczyk D, Huzarski T, Byrski T, Gronwald J, Gorski B, et al. A large germline deletion in the Chek2 kinase gene is associated with an

increased risk of prostate cancer. J Med Genet (2006) 43:863-6. doi: 10.1136/ jmg.2006.044974

112. Erkko H, Xia B, Nikkilä J, Schleutker J, Syrjäkoski K, Mannermaa A, et al. A recurrent mutation in PALB2 in Finnish cancer families. *Nature* (2007) 446:316–9. doi: 10.1038/nature05609

113. Leongamornlert D, Saunders E, Dadaev T, Tymrakiewicz M, Goh C, Jugurnauth-Little S, et al. Frequent germline deleterious mutations in DNA repair genes in familial prostate cancer cases are associated with advanced disease. *Br J Cancer* (2014) 110:1663–72. doi: 10.1038/bjc.2014.30

114. Page EC, Bancroft EK, Brook MN, Assel M, Hassan Al Battat M, Thomas S, et al. Interim results from the IMPACT study: evidence for prostate-specific antigen screening in BRCA2 mutation carriers. *Eur Urol* (2019) 76:831–42. doi: 10.1016/j.eururo.2019.08.019

115. Na R, Zheng SL, Han M, Yu H, Jiang D, Shah S, et al. Germline mutations in ATM and BRCA1/2 distinguish risk for lethal and indolent prostate cancer and are associated with early age at death. *Eur Urol* (2017) 71:740–7. doi: 10.1016/ j.eururo.2016.11.033

116. Thompson D, Duedal S, Kirner J, McGuffog L, Last J, Reiman A, et al. Cancer risks and mortality in heterozygous ATM mutation carriers. *JNCI: J Natl Cancer Institute* (2005) 97:813–22. doi: 10.1093/jnci/dji141

117. Näslund-Koch C, Nordestgaard BG, Bojesen SE. Increased risk for other cancers in addition to breast cancer for *CHEK2* *1100delC heterozygotes estimated from the Copenhagen general population study. *J Clin Oncol* (2016) 34:1208–16. doi: 10.1200/JCO.2015.63.3594

118. Ford D. Risks of cancer in BRCA1-mutation carriers. *Lancet* (1994) 343:692–5. doi: 10.1016/S0140-6736(94)91578-4

119. Foo TK, Tischkowitz M, Simhadri S, Boshari T, Zayed N, Burke KA, et al. Compromised BRCA1-PALB2 interaction is associated with breast cancer risk. *Oncogene* (2017) 36:4161-70. doi: 10.1038/onc.2017.46

120. Risch HA, McLaughlin JR, Cole DEC, Rosen B, Bradley L, Kwan E, et al. Prevalence and penetrance of germline BRCA1 and BRCA2 mutations in a population series of 649 women with ovarian cancer. *Am J Hum Genet* (2001) 68:700–10. doi: 10.1086/318787

121. Antoniou A, Pharoah PDP, Narod S, Risch HA, Eyfjord JE, Hopper JL, et al. Average risks of breast and ovarian cancer associated with BRCA1 or BRCA2 mutations detected in case series unselected for family history: a combined analysis of 22 studies. *Am J Hum Genet* (2003) 72:1117–30. doi: 10.1086/375033

122. Kote-Jarai Z, Jugurnauth S, Mulholland S, Leongamornlert DA, Guy M, Edwards S, et al. A recurrent truncating germline mutation in the BRIP1/FANCJ gene and susceptibility to prostate cancer. *Br J Cancer* (2009) 100:426–30. doi: 10.1038/ sj.bjc.6604847

123. Nysom K, Leblond P, Frappaz D, Aerts I, Varlet P, Giangaspero F, et al. PALB2 reversion mutations in breast, prostate, and ovarian carcinomas. *Ann Oncol* (2017) 28: v23–4. doi: 10.1093/ANNONC/MDX363.004

124. CHEK2 mutation and risk of prostate cancer: a systematic review and metaanalysis - PubMed. Available at: https://ncbi.nlm.nih.gov/26629066/ (Accessed pril 13, 2022).

125. Mavrou A, Tsangaris GTH, Roma E, Kolialexi A. The ATM gene and ataxia telangiectasia. *Anticancer Res* (2008) 28:401–5.

126. McKinnon PJ. ATM And ataxia telangiectasia. *EMBO Rep* (2004) 5:772-6. doi: 10.1038/sj.embor.7400210

127. Che R, Zhang J, Nepal M, Han B, Fei P. Multifaceted fanconi anemia signaling. *Trends Genet* (2018) 34:171–83. doi: 10.1016/j.tig.2017.11.006

128. Raymond VM, Mukherjee B, Wang F, Huang S-C, Stoffel EM, Kastrinos F, et al. Elevated risk of prostate cancer among men with lynch syndrome. *J Clin Oncol* (2013) 31:1713–8. doi: 10.1200/JCO.2012.44.1238

129. Na R, Zheng SL, Han M, Yu H, Jiang D, Shah S, et al. Germline mutations in ATM and BRCA1/2 distinguish risk for lethal and indolent prostate cancer and are associated with early age at death [figure presented]. *Eur Urol* (2017) 71:740–7. doi: 10.1016/j.eururo.2016.11.033

130. Caffo O, De Giorgi U, Fratino L, Alesini D, Zagonel V, Facchini G, et al. Clinical outcomes of castration-resistant prostate cancer treatments administered as third or fourth line following failure of docetaxel and other second-line treatment: results of an Italian multicentre study. *Eur Urol* (2015) 68:147–53. doi: 10.1016/j.eururo.2014.10.014

131. Scher HI, Solo K, Valant J, Todd MB, Mehra M. Prevalence of prostate cancer clinical states and mortality in the united states: estimates using a dynamic progression model. *PloS One* (2015) 10:e0139440. doi: 10.1371/journal.pone.0139440

132. Annala M, Struss WJ, Warner EW, Beja K, Vandekerkhove G, Wong A, et al. Treatment outcomes and tumor loss of heterozygosity in germline DNA repairdeficient prostate cancer. *Eur Urol* (2017) 72:34–42. doi: 10.1016/j.eururo.2017.02.023

133. Antonarakis ES, Lu C, Luber B, Liang C, Wang H, Chen Y, et al. Germline DNA-repair gene mutations and outcomes in men with metastatic castration-resistant prostate cancer receiving first-line abiraterone and enzalutamide. *Eur Urol* (2018) 74:218–25. doi: 10.1016/j.eururo.2018.01.035

134. Lalonde E, Ishkanian AS, Sykes J, Fraser M, Ross-Adams H, Erho N, et al. Tumour genomic and microenvironmental heterogeneity for integrated prediction of 5-year biochemical recurrence of prostate cancer: a retrospective cohort study. *Lancet Oncol* (2014) 15:1521–32. doi: 10.1016/S1470-2045(14)71021-6

135. Lalonde E, Alkallas R, Chua MLK, Fraser M, Haider S, Meng A, et al. Translating a prognostic DNA genomic classifier into the clinic: retrospective validation in 563 localized prostate tumors. *Eur Urol* (2017) 72:22–31. doi: 10.1016/ j.eururo.2016.10.013

136. Herzog TJ, Vergote I, Gomella LG, Milenkova T, French T, Tonikian R, et al. Testing for homologous recombination repair or homologous recombination deficiency for poly (ADP-ribose) polymerase inhibitors: a current perspective. *Eur J Cancer* (2022) 179:136–46. doi: 10.1016/j.ejca.2022.10.021

137. Bryant HE, Schultz N, Thomas HD, Parker KM, Flower D, Lopez E, et al. Specific killing of BRCA2-deficient tumours with inhibitors of poly(ADP-ribose) polymerase. *Nature* (2005) 434:913–7. doi: 10.1038/nature03443

138. Farmer H, McCabe H, Lord CJ, Tutt AHJ, Johnson DA, Richardson TB, et al. Targeting the DNA repair defect in BRCA mutant cells as a therapeutic strategy. *Nat* 2005 434:7035 (2005) 434:917–21. doi: 10.1038/nature03445

139. Tutt A, Tovey H, Cheang MCU, Kernaghan S, Kilburn L, Gazinska P, et al. Carboplatin in BRCA1/2-mutated and triple-negative breast cancer BRCAness subgroups: the TNT trial. *Nat Med* (2018) 24:628–37. doi: 10.1038/s41591-018-0009-7

140. Tan DSP, Kaye SB. Chemotherapy for patients with BRCA1 and BRCA2 – mutated ovarian cancer: same or different? *American Society of clinical oncology educational book* (2015) 35:114–21. doi: 10.14694/EdBook_AM.2015.35.114

141. Mota JM, Barnett E, Nauseef JT, Nguyen B, Stopsack KH, Wibmer A, et al. Platinum-based chemotherapy in metastatic prostate cancer with DNA repair gene alterations. JCO Precis Oncol (2020) 4:355–66. doi: 10.1200/PO.19.00346

142. Pomerantz MM, Spisák S, Jia L, Cronin AM, Csabai I, Ledet E, et al. The association between germline *BRCA2* variants and sensitivity to platinum-based chemotherapy among men with metastatic prostate cancer. *Cancer* (2017) 123:3532–9. doi: 10.1002/cncr.30808

143. Schmid S, Omlin A, Higano C, Sweeney C, Martinez Chanza N, Mehra N, et al. Activity of platinum-based chemotherapy in patients with advanced prostate cancer with and without DNA repair gene aberrations. *JAMA Netw Open* (2020) 3:e2021692. doi: 10.1001/jamanetworkopen.2020.21692

144. Ngoi NYL, Pham MM, Tan DSP, Yap TA. Targeting the replication stress response through synthetic lethal strategies in cancer medicine. *Trends Cancer* (2021) 7:930–57. doi: 10.1016/j.trecan.2021.06.002

145. Boyiadzis MM, Kirkwood JM, Marshall JL, Pritchard CC, Azad NS, Gulley JL. Significance and implications of FDA approval of pembrolizumab for biomarkerdefined disease. *J Immunother Cancer* (2018) 6:35. doi: 10.1186/s40425-018-0342-x

146. Brahmer JR, Tykodi SS, Chow LQM, Hwu W-J, Topalian SL, Hwu P, et al. Safety and activity of anti–PD-L1 antibody in patients with advanced cancer. *New Engl J Med* (2012) 366:2455–65. doi: 10.1056/NEJMoa1200694

147. de Almeida DVP, Fong L, Rettig MB, Autio KA. Immune checkpoint blockade for prostate cancer: niche role or next breakthrough? *American Society of clinical oncology educational book* (2020) 40:e89–e106. doi: 10.1200/EDBK_278853

148. Alva AS, Li J, Chou J, Reimers MA, McKay RR, Zhang J, et al. Phase 2 trial of immunotherapy in tumors with CDK12 inactivation (IMPACT): results from cohort a of patients (pts) with metastatic castration resistant prostate cancer (mCRPC) receiving dual immune checkpoint inhibition (ICI). *J Clin Oncol* (2022) 40:103–3. doi: 10.1200/ JCO.2022.40.6_suppl.103

149. Rebuzzi SE, Rescigno P, Catalano F, Mollica V, Vogl UM, Marandino L, et al. Immune checkpoint inhibitors in advanced prostate cancer: current data and future perspectives. *Cancers (Basel)* (2022) 14:1245. doi: 10.3390/cancers14051245

Glossary

AD1andregen deprivation metapyBERbase excision repairBRCAmmutations in <i>BRCA1</i> and <i>BRCA2</i> CIconfidence intervalCRPCcastration-resistant prostate cancerCSScause-specific survivalDDRDNA damage responseFAFanconi anaemiagLOHgenome-wide loss of heterozygosityHRDhomologous recombination deficiencyHRRloynch syndromemCRPCmetastatic castration-resistant prostate cancermCSPCmetastatic castration-sensitive prostate cancerMMRmismatch repairMSI-Hmiscrosatellite instability-highNCCNNational Comprehensive Cancer NetworkNHAnew hormonal agentORoverall survivalPARPPoly(ADP-ribose) polymerase inhibitorPARPiroly(ADP-ribose) polymerase inhibitorPSAgradiographic progression-free survivalRRstandardised relative riskSBBssingle-strand breaks.	ADT	androgen deprivation therapy
BRCAmmutations in <i>BRCA1</i> and <i>BRCA2</i> GIconfidence intervalCRPCcastration-resistant prostate cancerCSScause-specific survivalDDRDNA damage responseFAFanconi anaemiagLOHgenome-wide loss of heterozygosityHRDhomologous recombination deficiencyHRRbomologous recombination repairLSLynch syndromemCRPCmetastatic castration-resistant prostate cancerMMRmismatch repairMSI-Hmiscrosatellite instability-highNCCNAdional Comprehensive Cancer NetworkNHAodds ratioOSoverall survivalPARPiPoly(ADP-ribose) polymerasePARPiradiographic progression-free survivalRRrelative riskSRRkandardised relative risk		
CIconfidence intervalCRPCcastration-resistant prostate cancerCSScause-specific survivalDDRDNA damage responseFAFanconi anaemiagLOHgenome-wide loss of heterozygosityHRDhomologous recombination deficiencyHRRlomologous recombination repairLSLynch syndromemCSPCmetastatic castration-resistant prostate cancerMMRmismatch repairMSI-Hmiscosatellite instability-highNCCNNational Comprehensive Cancer NetworkNHAnev hormonal agentOSoverall survivalPARPiPoly(ADP-ribose) polymerasePARPirostate-specific antigenPARPirediographic progression-free survivalRRrelative riskSRRstandardised relative risk	BER	base excision repair
CRPCcastration-resistant prostate cancerCSScause-specific survivalDDRDNA damage responseFAFanconi anaemiagLOHgenome-wide loss of heterozygosityHRDhomologous recombination deficiencyHRRNomologous recombination repairLSLynch syndromemCRPCmetastatic castration-resistant prostate cancerMMRmismatch repairMMRnismatch repairMSI-Hniscrosatellite instability-highNCCNNational Comprehensive Cancer NetworkNHAnew hormonal agentOSoverall survivalPARPiPoly(ADP-ribose) polymerase inhibitorPSAradiographic progression-free survivalRRrelative riskSRRstandardised relative risk	BRCAm	mutations in BRCA1 and BRCA2
CSScause-specific survivalDDRDNA damage responseFAFanconi anaemiagLOHgenome-wide loss of heterozygosityHRDhomologous recombination deficiencyHRRhomologous recombination repairLSLynch syndromemCRPCmetastatic castration-resistant prostate cancermCSPCmetastatic castration-sensitive prostate cancerMMRmismatch repairMSI-HNational Comprehensive Cancer NetworkNHAnew hormonal agentORodds ratioOSoverall survivalPARPPoly(ADP-ribose) polymerasePARPiradiographic progression-free survivalRRrelative riskstandardised relative risk	CI	confidence interval
DDRDNA damage responseFAFanconi anaemiagLOHgenome-wide loss of heterozygosityHRDhomologous recombination deficiencyHRRhomologous recombination repairLSLynch syndromemCRPCmetastatic castration-resistant prostate cancerMCRPCmetastatic castration-sensitive prostate cancerMMRmismatch repairMSI-Hmiscrosatellite instability-highNCCNNational Comprehensive Cancer NetworkNHAnew hormonal agentORodds ratioOSoverall survivalPARPPoly(ADP-ribose) polymerasePARPiradiographic progression-free survivalRRrelative riskSRRstandardised relative risk	CRPC	castration-resistant prostate cancer
FAFanconi anaemiagLOHgenome-wide loss of heterozygosityHRDhomologous recombination deficiencyHRRhomologous recombination repairLSLynch syndromemCRPCmetastatic castration-resistant prostate cancermCSPCmetastatic castration-sensitive prostate cancerMMRmiscrosatellite instability-highNCCNNational Comprehensive Cancer NetworkNHAnew hormonal agentORodds ratioOSoverall survivalPARPPoly(ADP-ribose) polymerasePARPiprostate-specific antigenrPFSradiographic progression-free survivalRRrelative risk	CSS	cause-specific survival
FinalFunction matchinggLOHgenome-wide loss of heterozygosityHRDhomologous recombination deficiencyHRRhomologous recombination repairLSLynch syndromemCRPCmetastatic castration-resistant prostate cancermCSPCmetastatic castration-sensitive prostate cancerMMRmismatch repairMSI-Hmiscrosatellite instability-highNCCNNational Comprehensive Cancer NetworkNHAnew hormonal agentORodds ratioOSoverall survivalPARPPoly(ADP-ribose) polymerasePARPiprostate-specific antigenrPFSradiographic progression-free survivalRRrelative risk	DDR	DNA damage response
CCICHRDhomologous recombination deficiencyHRRhomologous recombination repairLSLynch syndromemCRPCmetastatic castration-resistant prostate cancermCSPCmetastatic castration-sensitive prostate cancerMMRmismatch repairMSI-Hmiscrosatellite instability-highNCCNNational Comprehensive Cancer NetworkNHAnew hormonal agentORodds ratioOSoverall survivalPARPPoly(ADP-ribose) polymerasePARPiprostate-specific antigenrPFSradiographic progression-free survivalRRrelative riskSRRstandardised relative risk	FA	Fanconi anaemia
HRRhomologous recombination repairLSLynch syndromemCRPCmetastatic castration-resistant prostate cancermCSPCmetastatic castration-sensitive prostate cancerMMRmismatch repairMSI-Hmiscrosatellite instability-highNCCNNational Comprehensive Cancer NetworkNHAnew hormonal agentORodds ratioOSoverall survivalPARPPoly(ADP-ribose) polymerasePARPiprostate-specific antigenrPFSradiographic progression-free survivalRRstandardised relative risk	gLOH	genome-wide loss of heterozygosity
LSLynch syndromemCRPCmetastatic castration-resistant prostate cancermCSPCmetastatic castration-sensitive prostate cancerMMRmismatch repairMSI-Hmiscrosatellite instability-highNCCNNational Comprehensive Cancer NetworkNHAnew hormonal agentORodds ratioOSoverall survivalPARPPoly(ADP-ribose) polymerasePARPiprostate-specific antigenrPFSradiographic progression-free survivalRRrelative risk	HRD	homologous recombination deficiency
mCRPCmetastatic castration-resistant prostate cancermCSPCmetastatic castration-sensitive prostate cancerMMRmismatch repairMSI-Hmiscrosatellite instability-highNCCNNational Comprehensive Cancer NetworkNHAnew hormonal agentORodds ratioOSoverall survivalPARPPoly(ADP-ribose) polymerasePARPiprostate-specific antigenrPFSradiographic progression-free survivalRRrelative risk	HRR	homologous recombination repair
mCSPCmetastatic castration-sensitive prostate cancerMMRmismatch repairMSI-Hmiscrosatellite instability-highNCCNNational Comprehensive Cancer NetworkNHAnew hormonal agentORodds ratioOSoverall survivalPARPPoly(ADP-ribose) polymerasePARPiPoly(ADP-ribose) polymerase inhibitorPSArelative riskRRrelative risk	LS	Lynch syndrome
MMRmismatch repairMSI-Hmiscrosatellite instability-highNCCNNational Comprehensive Cancer NetworkNHAnew hormonal agentORodds ratioOSoverall survivalPARPPoly(ADP-ribose) polymerasePARPiPoly(ADP-ribose) polymerase inhibitorPSAprostate-specific antigenrPFSradiographic progression-free survivalRRrelative risk	mCRPC	metastatic castration-resistant prostate cancer
MSI-Hmiscrosatellite instability-highNCCNNational Comprehensive Cancer NetworkNHAnew hormonal agentORodds ratioOSoverall survivalPARPPoly(ADP-ribose) polymerasePARPiPoly(ADP-ribose) polymerase inhibitorPSAprostate-specific antigenrPFSradiographic progression-free survivalRRstandardised relative risk	mCSPC	metastatic castration-sensitive prostate cancer
NCCNNational Comprehensive Cancer NetworkNHAnew hormonal agentORodds ratioOSoverall survivalPARPPoly(ADP-ribose) polymerasePARPiPoly(ADP-ribose) polymerase inhibitorPSAprostate-specific antigenrPFSradiographic progression-free survivalRRstandardised relative risk	MMR	mismatch repair
NHAnew hormonal agentORodds ratioOSoverall survivalPARPPoly(ADP-ribose) polymerasePARPiPoly(ADP-ribose) polymerase inhibitorPSAprostate-specific antigenrPFSradiographic progression-free survivalRRrelative riskSRRstandardised relative risk	MSI-H	miscrosatellite instability-high
ORodds ratioOSoverall survivalPARPPoly(ADP-ribose) polymerasePARPiPoly(ADP-ribose) polymerase inhibitorPSAprostate-specific antigenrPFSradiographic progression-free survivalRRrelative riskSRRstandardised relative risk	NCCN	National Comprehensive Cancer Network
OSoverall survivalPARPPoly(ADP-ribose) polymerasePARPiPoly(ADP-ribose) polymerase inhibitorPSAprostate-specific antigenrPFSradiographic progression-free survivalRRrelative riskSRRstandardised relative risk	NHA	new hormonal agent
PARPPoly(ADP-ribose) polymerasePARPiPoly(ADP-ribose) polymerase inhibitorPSAprostate-specific antigenrPFSradiographic progression-free survivalRRrelative riskSRRstandardised relative risk	OR	odds ratio
PARPi Poly(ADP-ribose) polymerase inhibitor PSA prostate-specific antigen rPFS radiographic progression-free survival RR relative risk SRR standardised relative risk	OS	overall survival
PSA prostate-specific antigen rPFS radiographic progression-free survival RR relative risk SRR standardised relative risk	PARP	Poly(ADP-ribose) polymerase
rPFS radiographic progression-free survival RR relative risk SRR standardised relative risk	PARPi	Poly(ADP-ribose) polymerase inhibitor
RR relative risk SRR standardised relative risk	PSA	prostate-specific antigen
SRR standardised relative risk	rPFS	radiographic progression-free survival
	RR	relative risk
SSBs single-strand breaks.	SRR	standardised relative risk
	SSBs	single-strand breaks.