Check for updates

#### **OPEN ACCESS**

EDITED BY Peter Korsten, University Medical Center Göttingen, Germany

REVIEWED BY Peizeng Yang, First Affiliated Hospital of Chongqing Medical University, China Takayuki Katsuyama, Okayama University, Japan Tadashi Hosoya, Tokyo Medical and Dental University, Japan

\*CORRESPONDENCE Yoshishige Miyabe ⊠ yoshishige.miyabe@marianna-u.ac.jp

RECEIVED 28 September 2022 ACCEPTED 12 June 2023 PUBLISHED 26 June 2023

#### CITATION

Shimizu J, Murayama MA, Mizukami Y, Arimitsu N, Takai K and Miyabe Y (2023) Innate immune responses in Behçet disease and relapsing polychondritis. *Front. Med.* 10:1055753. doi: 10.3389/fmed.2023.1055753

#### COPYRIGHT

© 2023 Shimizu, Murayama, Mizukami, Arimitsu, Takai and Miyabe. This is an openaccess article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Innate immune responses in Behçet disease and relapsing polychondritis

Jun Shimizu<sup>1</sup>, Masanori A. Murayama<sup>2</sup>, Yoshihisa Mizukami<sup>1</sup>, Nagisa Arimitsu<sup>1</sup>, Kenji Takai<sup>1</sup> and Yoshishige Miyabe<sup>1\*</sup>

<sup>1</sup>Department of Immunology and Parasitology, St. Marianna University of School of Medicine, Kawasaki, Kanagawa, Japan, <sup>2</sup>Department of Animal Models for Human Diseases, Institute of Biomedical Science, Kansai Medical University, Hirakata, Osaka, Japan

Behçet disease (BD) and relapsing polychondritis (RP) are chronic multisystem disorders characterized by recurrent flare-ups of tissue inflammation. Major clinical manifestations of BD are oral aphthae, genital aphthous ulcers, skin lesions, arthritis, and uveitis. Patients with BD may develop rare but serious neural, intestinal, and vascular complications, with high relapse rates. Meanwhile, RP is characterized by the inflammation of the cartilaginous tissues of the ears, nose, peripheral joints, and tracheobronchial tree. Additionally, it affects the proteoglycan-rich structures in the eyes, inner ear, heart, blood vessels, and kidneys. The mouth and genital ulcers with inflamed cartilage (MAGIC) syndrome is a common characteristic of BD and RP. The immunopathology of these two diseases may be closely related. It is established that the genetic predisposition to BD is related to the human leukocyte antigen (HLA)-B51 gene. Skin histopathology demonstrates the overactivation of innate immunity, such as neutrophilic dermatitis/panniculitis, in patients with BD. Monocytes and neutrophils frequently infiltrate cartilaginous tissues of patients with RP. Somatic mutations in UBA1, which encodes a ubiquitylation-related enzyme, cause vacuoles, E1 enzyme, X-linked, autoinflammatory, somatic syndrome (VEXAS) with severe systemic inflammation and activation of myeloid cells. VEXAS prompts auricular and/or nasal chondritis, with neutrophilic infiltration around the cartilage in 52-60% of patients. Thus, innate immune cells may play an important role in the initiation of inflammatory processes underlying both diseases. This review summarizes the recent advances in our understanding of the innate cell-mediated immunopathology of BD and RP, with a focus on the common and distinct features of these mechanisms.

#### KEYWORDS

Behçet disease, relapsing polychondritis, neutrophils, monocytes, macrophages, cytokines, autoinflammatory disease, autoimmune disease

## 1. Introduction

Behçet disease (BD) is an inflammatory disorder characterized by the frequent occurrence of oral ulcers, genital aphthous ulcers, and uveitis, with clinical manifestations involving the skin, cardiovascular, intestinal, and central nervous system (CNS) (1). These manifestations are important for diagnosis as there are no clinical or laboratory findings specific to BD. In 1985, an international study group developed diagnostic criteria based on the major symptoms of BD; a diagnosis is made when an individual has developed recurrent oral ulceration (at least three times over the past 12 months) with at least two of the following symptoms: persistent genital ulceration; eye lesions, such as uveitis and retinal vasculitis; skin involvement, such as erythema nodosum and thrombophlebitis; and a positive pathergy test (2). The clinical diagnostic criteria must be followed by the exclusion criteria for patients with other immune disorders presenting with common symptoms of BD. For example, chronic oral ulcerations are frequently observed in Crohn disease too (3). Additionally, Vogt-Koyanagi-Harada and Cogan syndromes should be considered during the differential diagnosis of BD in patients with uveitis (4, 5). CNS and gastrointestinal involvement are indicators of a poor prognosis in patients with neuro-BD and intestinal BD, respectively. Patients in the BD subgroup are difficult to distinguish from those with multiple sclerosis (6) and Crohn disease (7).

Relapsing polychondritis (RP) is a chronic inflammatory disorder characterized by chondritis of the auricular, nasal, joint, and tracheal cartilaginous tissues (8). In addition, it affects the proteoglycan-rich structures in the eyes, inner ear, heart, blood vessels, and kidneys. In the clinical features, respiratory involvement is associated with poor prognosis through the severe pulmonary complications such as tracheobronchomalacia and/or pulmonary infection (9). The diagnosis is usually made based on clinical symptoms using McAdam's (10), Damiani's (11), and/or Michet's (12) criteria, because there are no pathognomonic clinical and laboratory features, similar to patients with BD. Approximately 20-35% of RP cases are further complicated by other immune disorders such as systemic vasculitis, rheumatoid arthritis, and systemic lupus erythematosus (10-13). In most instances, coexisting diseases precede the onset of RP (10), and in some patients, complications occur as consequent symptoms of RP (8). It is possible that the clinical course of the disease makes it difficult to obtain an early and accurate diagnosis.

Identifying new diagnostic biomarkers for human inflammatory diseases require further studies. Generally, human autoinflammatory diseases, such as familial Mediterranean fever (FMF) and tumor necrosis factor (TNF) receptor-associated periodic fever syndrome (TRAPS), are thought to be caused by abnormalities in phagocytes against pathogenic elements. In contrast, human autoimmune diseases are characterized by the overactivation of lymphocytes in response to autoantigens. In these studies, most immune disorders were suggested to be caused by a combination of autoimmune and autoinflammatory mechanisms in the disease spectrum, based on the genetic and cellular basis (14). In the immune disease spectrum, BD is associated with a mixed pattern of autoinflammatory and autoimmune diseases (Figure 1) (14).

In this review, we summarize current knowledge of innate cell mediated immunopathology in BD and RP to identify accurate positions of the immune disorders in the disease spectrum to facilitate the development of new therapeutic strategies.

## 2. Behçet disease

## 2.1. Epidemiology of BD

#### 2.1.1. Environmental factors in BD

BD is prevalent along the ancient Silk Road between the Mediterranean Basin and East Asia (1). The human leukocyte antigen (HLA)-B51 gene is established as a major BD susceptibility gene, especially in the patients with ocular involvement (1). Additionally, a

geological association was observed between the prevalence of BD and HLA-B51 (15). These data demonstrated that BD inflammation may be triggered by innate immunity as well as environmental factors, such as bacterial and viral agents.

Oral plaque index scores are associated with the presence of oral ulcers and BD severity (16). Dental plaque bacteria (*Streptococcus sanguinis*) are frequently observed in the oral cavity of patients (17). Mouthwashes containing soluble betamethasone, doxycycline, and nystatin improve oral ulcer severity scores in patients with BD (18).

Recent studies have revealed perturbation of oral and gut microbiota, especially increases in lactate-producing bacteria such as *Lactobacillus* and *Bifidobacterium*, in patients with BD compared with those in healthy individuals. Researchers have suggested pathological relationships between microbiota and immunological dysfunction in BD (Figure 2A) (19–22). In contrast to these clinical and laboratory findings, HLA-B51 transgenic mice demonstrate no obvious clinical phenotypes of BD, although stimulated neutrophils produce high levels of superoxide (23).

### 2.1.2. Genetic variations in BD

Genome-wide profiling analyses revealed that, adding to HLA-B51, myeloid immune cell-related molecules, such as endoplasmic reticulum aminopeptidase-1 (ERAP1), major histocompatibility complex (MHC) class I polypeptide-related sequence-A (MICA), familial Mediterranean (MEFV) gene products, toll-like receptor-4 (TLR-4), c-c motif chemokine receptors CCR1-CCR3, interleukin (IL)-1β, IL-10, interferon (IFN)-y receptor (IFNGR)-1, IL-23R, and IL-12RB, were risk factors of BD (24-29). These findings suggest that innate immune functions and bacterial identification systems play crucial roles in the pathogenesis of BD (Figure 2A). Lymphocytes obtained from BD patients react with human and/or mycobacterial heat shock protein peptides (30, 31). TLR-1, 2, and 4 are expressed more abundantly on neutrophils, monocytes, and lymphocytes derived from patients with BD than on those derived from healthy individuals (32).

The tumor necrosis factor- $\alpha$ -induced protein-3 (TNFAIP3) gene encodes A20 which regulates negatively TNF $\alpha$  pathway through the ubiquitin ligase activity (33). Patients with A20 haploinsufficiency develop BD phenotypes with an onset in childhood and young adulthood. Peripheral blood mononuclear cells (PBMCs) of the patients produced higher amounts of proinflammatory cytokines, such as IL-1 $\beta$ , TNF $\alpha$ , IL-17, and IL-18, in the presence of lipopolysaccharide (LPS), than those of healthy individuals.

#### 2.1.3. Clinical phenotypes in BD

Recent clustering analyses have demonstrated that patients with BD can be divided into several subgroups according to their clinical symptoms to simplify and increase the accuracy of the clinical assessment (34, 35). For example, patients with mucocutaneous manifestations belonged to one subgroup, those with vascular manifestations belonged to another, and those with eye and/or CNS involvement belonged to another (35).

Interestingly, HLA-B51 positivity was relatively low in patients with BD with intestinal involvement, and male and female patients had eye and mucocutaneous involvement, respectively (Figure 2A) (34).



Stratification of human autoinflammatory and autoimmune diseases by evaluating immune conditions (14, McGonagle and McDermotte. PLoS Med. 2006; 3:e297. Modified). Innate immune overactivation via inflammatory cytokine signaling, pathogen sensing, and/or disruption of local tissue homeostasis are the proposed main causes of autoinflammatory diseases, while autoimmune diseases are associated with self-reactive lymphocytes through impaired immune tolerance. Behçet disease (BD) and relapsing polychondritis (RP) may be allocated to distinct clusters of the stratification. FMF: Familial Mediterranean fever; TRAPS: TNF receptor-associated periodic fever syndrome; CD: Crohn disease; UC: Ulcerative colitis; AS: Ankylosing spondylitis; RA: Rheumatoid arthritis; SLE: Systemic lupus erythematosus; ALPS: autoimmune lymphoproliferative syndrome; IPEX: immune dysrequlation, polyendocrinopathy, enteropathy, X-linked.

## 2.2. Histopathology of BD

Erythema nodosum (EN)-like lesions and papulopustular lesions are common in patients with BD (36). Histopathological examination of EN-like lesions in BD demonstrates panniculitis with vasculitis, leukocytoclastic and lymphocytic vasculitis, or phlebitis of the dermis. BD skin lesions are occasionally associated with thrombosis. The infiltrating immune cells consist mainly of neutrophils and lymphocytes (37). Similarly, in papulopustular lesions, neutrophilic infiltration was observed in the epidermis and around hair follicles. Lymphocytic infiltration has been suggested to occur as a late-stage inflammation in the neutrophilic reaction (38). Indeed, an investigation of patients with EN-like lesions revealed that neutrophilic dermatitis/panniculitis was more frequently observed in patients with BD with EN-like lesions than in patients with nodular vasculitis or EN of other immune disorders (39).

Sterile needle pricks often form inflammatory papules or pustules on the skin in patients with BD, with infiltration of neutrophils and lymphocytes as a positive pathergy test (40). Higher response rates were observed for pricks with larger gauge and/or blunt needles in patients with BD (41). A similar procedure using saliva pricks increased positive rates and was associated with disease activity (42). These data suggest that pathergy tests with needle pricks lead to the overactivation of immune cells against pathogen- and damage-associated molecular patterns in patients with BD (40).

In the pathergy test, lymphocyte and monocyte infiltrations were persistent up to 48 h after the needle prick in patients with BD compared to healthy individuals (43). Small clusters of elastasepositive neutrophils have been observed at needle prick sites in the relatively early phases of the test until 24 h after the prick (44, 45).

In the oral and genital ulcer lesions, leukocytoclastic vasculitis and lymphocyte infiltration were frequently observed in the lamina propria of the lesions (46). Intestinal ulceration is commonly found in the ileocecal region and is histologically characterized by neutrophilic and lymphocytic cell infiltration around lesions (47–49). The postmortem brain tissues of patients with neuro-BD demonstrate perivascular cuffing of macrophages and T cells in the parenchyma (50).



FIGURE 2

Stratification of human autoinflammatory and autoimmune diseases by evaluating immune conditions (14, McGonagle and McDermotte. PLoS Med. 2006; 3:e297. Modified).

## 2.3. Peripheral blood cells in BD

#### 2.3.1. Neutrophils in BD

Neutrophils produce reactive oxygen species (ROS) as a first-line defense against infectious pathogens (51). HLA-B51-positive neutrophils produce excessive superoxide compared to those without HLA in patients with BD and healthy individuals (23).

Neutrophil migration in patients with BD was enriched in an *in vivo* assay compared to that in healthy individuals, and the titers were significantly reduced in the disease remission phases (52). No significant differences were observed in superoxide production or adhesion capabilities between patients with BD and healthy individuals (52).

Neutrophil oxidative burst responses in patients with severe BD and organ involvement, such as complications of the eye, intestines, central nervous system, and cardiovascular system, were significantly higher than those in patients with mild BD (53).

#### 2.3.2. Neutrophil extracellular traps in BD

Neutrophils activated by phorbol myristate acetate, IL-8, or LPS become flat and produce extracellular structures called NETs, which contain myeloperoxidase, neutrophil elastase, and cathepsin G (54). The nuclear enzyme protein-arginine deiminase type 4 (PAD4) citrullinates histones and promotes chromatin decondensation (55). NETs degrade virulence factors and inhibit the growth of bacteria such as *Staphylococcus aureus*, *Salmonella typhimurium*, and *Shigella flexneri* (54). On the other hand, damage-associated molecular patterns, such as cholesterol crystals, induced NET release of neutrophils and the NETs with cholesterol crystals promoted IL-1 $\beta$  production of macrophages (56). PAD4 deficient mice demonstrated reduced NET formation and a lower degree of thrombosis (57).

NETs may increase and induce thrombosis in patients with BD. Neutrophil NET release and serum levels of DNA components were significantly increased in patients with active BD compared to those in patients with inactive BD and healthy individuals (58). Neutrophil PAD4 expression level is significantly higher in patients with BD than in healthy individuals (59). Thrombin generation parameters in platelet-poor plasma obtained from patients with BD were significantly higher than those in healthy individuals, and correlated well with DNA component levels (58). NETs obtained from BD patients effectively promoted IL-8 and TNF $\alpha$  production of monocytes/macrophages compared with healthy individuals (60). Diffuse elastase-producing neutrophils were observed in BD skin panniculitis and vasculitis (59). NETs play a crucial role in the pathogenesis of BD.

Low-density neutrophils are immature or degranulated and recognized in human diseases (61). The frequencies of low-density neutrophils and NET production by the stimulated cells were increased in patients with BD compared to healthy individuals, but the cells exhibited decreased phagocytic capacities (62). Determining the mechanisms underlying these associations warrant further studies.

#### 2.3.3. Monocytes/macrophages in BD

In *in vitro* experiments, bone marrow cells were differentiated into classically activated M1 macrophages in the presence of IFN $\gamma$ and LPS and promoted production of proinflammatory cytokines, such as IL-1 $\beta$ , IL-6, and TNF $\alpha$  (63). Cells differentiate into alternatively activated M2 macrophages with IL-4 and increased IL-10 expression levels (63). M2 macrophages have been suggested to play distinct roles in lesions by reducing inflammation and promoting tissue remodeling.

Monocyte-derived macrophages treated with BD sera produced more effectively IL-12 and TNF $\alpha$  than those treated with sera of healthy individuals, suggesting M1 macrophage prevalence in peripheral blood of patients with BD (64). Stimulated M1 macrophages from patients with BD exhibit higher CCR1 expression levels than those from healthy individuals (65). Similarly, a gene expression profiling study demonstrated that, compared with healthy individuals, expression levels of proinflammatory monocyte-associated molecules, such as IL-1 $\beta$  and a CCR1 ligand CCL3, were elevated in patients with BD (66).

### 2.3.4. Inflammasome components in BD

The inflammasome complex consists of a cytosolic nucleotidebinding domain, leucine-rich-repeat-containing (NLR) proteins, AIM2-like receptor (ALR) proteins, adaptor apoptosis-associated speck-like protein containing a CARD (ASC), and pro-caspase-1 (67). The well-studied inflammasome NLRP3 responds to and is activated by bacterial, fungal, and viral pathogen-associated molecular patterns and damage-associated molecular patterns such as ATP and uric acid crystals (68). Activated caspase-1 processes pro-IL-1 $\beta$  and pro-IL-18 and biologically active cytokines are secreted (68). An autoinflammatory disease, cryopyrin-associated periodic syndrome, has been suggested to be associated with NLRP3 gene mutations (69).

In the PBMCs of patients with BD, the protein and mRNA levels of NLRP3, ASC, and caspase-1 were significantly increased compared with healthy individuals (70, 71). Activated PBMCs with LPS and ATP induced significantly higher levels of IL-1 $\beta$  compared with cells without the stimulation (70). NLRP3 levels of cerebrospinal fluids of BD patients with CNS involvement are positively correlated with serum C-reactive protein concentrations and erythrocyte sedimentation rates (71). Patients with BD share common clinical features, at least among those with autoinflammatory diseases.

#### 2.3.5. Eosinophils in BD

Serum immunoglobulin E (IgE) and eosinophil counts are significantly reduced in patients with BD (72). Similarly, serum eosinophil cationic protein levels are significantly lower in active patients with BD than in inactive patients (73), suggesting a role for helper type 1 (Th1)-skewed cytokine responses in the pathogenesis of BD.

### 2.4. Humoral mediators in BD

#### 2.4.1. Cytokines/chemokines in BD

A literature-based meta-analysis ascertained that serum IL-1 $\beta$ , IL-6, and TNF $\alpha$  were significantly increased in patients with BD compared with healthy individuals (Figure 2A) (74). High levels of Th1 and Th17 related cytokines, such as IL-1 $\beta$ , IL-6, IL-12, IL-17, IL-23, IFN $\gamma$ , and TNF $\alpha$ , were identified in an array analysis (75). BD shares skewed IL-17/IL-23 pathways and several clinical features with spondyloarthritis and Vogt-Koyanagi-Harada disease (76, 77). Serum and plasma levels of CCL2, CCL3, and CXCL10 are higher in patients with BD compared with healthy individuals (78–80). Aqueous humor CXCL16 and CX3CL1 levels are higher in patients

with BD than in healthy individuals and patients with Vogt-Koyanagi-Harada disease, suggesting an enhancement of Th1 responses in BD uveitis (81).

#### 2.4.2. Matrix metalloproteinases in BD

Serum MMP-2 and MMP-9 levels were significantly higher in patients with vasculo-BD than in healthy individuals (Figure 2A) (82), especially in patients with aneurysms, similar to patients with abdominal aortic aneurysms (83). Synovial fluid concentrations of MMP-3 are significantly lower in patients with BD than in patients with rheumatoid arthritis and are comparable to those in patients with osteoarthritis (84).

#### 2.4.3. Autoantibodies in BD

Autoantibodies were observed in patients with BD and reacted with an endothelial cell antigen,  $\alpha$ -enolase (a positive rate of 38% by an ELISA) (85), a ubiquitously expressed membrane protein, prohibitin (28% by an ELISA) (86),  $\alpha$ -tropomyosin (22% by an ELISA) (87), the nuclear mitotic apparatus protein (NuMA; 28% by an ELISA) (88), a riboflavin-containing flavoprotein (41% by an ELISA) (89), a membrane protein annexin A2 (34% by an ELISA) (90), a microtubule-related protein, kinectin (23% by an immunoprecipitation assay) (91), and an actin-binding protein, cofilin-1 (13% by western blotting; Figure 2A) (92). Thus, BD demonstrates a mixed pattern of autoinflammatory and autoimmune diseases within the spectrum (14).

## 3. Relapsing polychondritis

## 3.1. Epidemiology of RP

#### 3.1.1. Genetic variations in RP

Epidemiological studies on patients with RP have identified that the incidence rates are approximately the same in several regions of the world (93–95). HLA-DR4 appears to be a susceptibility allele for RP (96). A recent genetic study demonstrated that HLA-DRB1\*16:02, HLA-DQB1\*05:02, and HLA-B\*67:01 are associated with RP (Figure 2B) (97). Based on our data, the authors concluded that RP may be a distinct disease from other rheumatic diseases such as rheumatoid arthritis, systemic lupus erythematosus, Takayasu arthritis, and BD.

#### 3.1.2. Environmental factors in RP

Based on the global incidence rates mentioned above, few environmental factors have been reported to be associated with RP pathogenesis. Interestingly, similar to the data of patients with BD, a metagenomic analysis demonstrated characteristic alterations in the gut microbiota composition, such as an increase in the abundance of *Eubacterium*, *Ruminococcus*, *Bacteroides*, and *Veillonella*, in patients with RP compared with that in healthy individuals. Here, we suggest an association between gut microbes and RP immunopathogenesis (98).

#### 3.1.3. Clinical phenotypes of RP

In the clinical manifestations, respiratory and auricular involvement, which are two key hallmark features of RP, are recognized in 40–67% and 85–90% of patients with RP, respectively, at the latest follow-up (10-13). Tracheobronchial chondritis is increasingly recognized as distinct from other pathogenic complications (Figure 2B) (99, 100). Certainly, patients with RP with respiratory involvement have progressive disease compared to those with auricular involvement (101). Similar to the ocular involvement in BD, posterior segment inflammation is associated with a weak response to treatment (102).

## 3.2. Histopathology of RP

In the initial stages of the disease, mononuclear cells and neutrophils infiltrate the perichondrium beside the normal cartilage tissue (103, 104). Among the inflammatory cells in granulation tissues, CD4+ Th cells and CD68+ monocytes/macrophages are prevalent (105). Damaged chondrocytes produce MMP-3 and cathepsins, and the number of proteolytic enzyme-expressing cells correlates with that of apoptotic chondrocytes. Interestingly, MMP-3 was observed in the cartilage and perichondrium, whereas MMP-8 and MMP-9 were detected only in perichondrium granulation tissues. Cartilage tissues are progressively destroyed and finally replaced by fibrous connective tissues.

Notably, 1.6–38% of patients with RP showed skin involvement (9, 10, 12, 13, 95, 106); mucosal aphthosis, nodules on the limbs, purpura, and sterile pustules were the most common dermatological manifestations (107). Skin biopsy specimens revealed leukocytoclastic vasculitis, thrombosis of the skin vessels, septal panniculitis, neutrophil infiltration, and lymphocytic vasculitis as their histological findings. About 0–12% of patients with RP develop neurological manifestations, mainly confusion, seizures, delusions, amnesia, and/ or dementia (108). Histopathology of the CNS exhibited perivascular cuffs of monocytes/lymphocytes and lymphocytic infiltration in the meninges and the cerebral parenchyma of patients with RP (109–111). In contrast to BD, gastrointestinal involvement is not generally identified in patients with RP (8–13, 93, 95, 106).

## 3.3. Peripheral blood cells in RP

#### 3.3.1. Neutrophils in RP

As mentioned previously, neutrophil infiltration into cartilage tissues has been recognized since the early stages of chondritis (103, 104). Leukocyte clastic vasculitis and neutrophil infiltration are frequently observed (40%) in skin biopsy specimens (107). These results suggested that neutrophil activation plays a crucial role in the initiation of chondritis in patients with RP.

#### 3.3.2. Monocytes/macrophages in RP

Gene expression level of IL-10, a major effector cytokine of regulatory T (Treg) cells, was significantly higher in freshly isolated PBMCs from patients with RP than in those from healthy individuals (112). After the initiation of cell culture with mitogen stimulation, IL-10 gene expression level was significantly decreased in patients with RP compared to that in healthy individuals. The researchers suggested that the gene expression analysis of PBMCs revealed Treg cell exhaustion or anergy of patients with RP and the skewed T cell function associated with innate cell overactivation (Figure 2B) (113).

### 3.3.3. Eosinophils in RP

Similar to neutrophil infiltration in RP lesions, eosinophils have been identified in specimens from the conjunctiva (114), nasal septum (104), and skin (115), probably indicating their early involvement in the process of chondritis in patients with RP.

## 3.4. Humoral mediators in RP

#### 3.4.1. Cytokines/chemokines in RP

In Th cell-related cytokines, IFN $\gamma$  and IL-10 were increased in the sera of patients with RP compared with healthy individuals (116, 117). Serum levels of innate cytokines and chemokines, such as IL-8, CCL2, and CCL4, were higher in patients with RP than in healthy individuals (Figure 2B) (116, 117).

### 3.4.2. MMPs in RP

Serum MMP-3 levels were higher in patients with RP than in healthy individuals, likely corresponding to histopathological changes in the patients (101, 116). Lymphocytes, monocytes/macrophages, and MMP-3 positive chondrocytes were simultaneously observed in RP lesions, suggesting that these cells aggravate chondritis (105). When the concentrations were compared between RP patients with and without respiratory involvement based on the epidemiological data mentioned above, MMP-3 levels increased significantly in patients with respiratory involvement compared to those without respiratory involvement (118).

In an *in vitro* assay, RP PBMCs upregulated mRNA expression of inflammatory cytokines IL-1 $\beta$  and IL-6 against stimulation compared with those of healthy individuals (118). Expression positively correlated with serum MMP-3 only in patients with RP and respiratory involvement. These data suggested that mononuclear cells with innate cytokines play a crucial role in the inflammatory processes of RP lesions, especially in patients with RP and respiratory involvement (Figure 2B).

# 3.4.3. Triggering receptor expressed on myeloid cell in RP

As the molecule name, neutrophils and monocytes/macrophages express TREM-1 and promote inflammation partly through TLR-4 pathway activation (119). Soluble TREM-1 is increased in the sera of RP patients with active disease compared with those with inactive disease, suggesting its possible role as a biomarker (116).

#### 3.4.4. Autoantibodies in RP

Several cartilage elements were identified as potential autoantigens for RP (Figure 2B). An initial report of circulating autoantibodies in patients with RP revealed that, using indirect immunofluorescence, 33% of patients had autoantibodies against type 2 collagen, and the titers increased when acute symptoms were exhibited (120). Type 2 collagen-immunized rats develop auricular chondritis in the presence of type 2 collagen-reactive antibodies (121).

Matrilin-1 is a cartilage-specific protein, and its serum concentrations were found to be significantly elevated in an RP patient with tracheal chondritis who was monitored for 2 years (122). Autoantibodies for matrilin-1 were detected using ELISA in 13% of 97 patients with RP (123). In this study, researchers ascertained that

sera from RP patients with positive anti-matrilin-1 antibodies reacted with newborn mouse tracheolaryngeal cartilage, whereas sera from patients with rheumatoid arthritis did not.

Certainly, several cartilage components are associated with phenotypic differences between patients with RP with and without respiratory involvement.

## 4. VEXAS and RP

A cutting-edge analysis demonstrated that patients with somatic mutations in UBA1, a gene encoding the ubiquitin activating enzyme E1, developed treatment-refractory severe autoinflammatory conditions in late middle age, such as vacuole, E1 enzyme, X-linked, autoinflammatory, and somatic syndrome (VEXAS) (124). It is characterized by refractory constitutional symptoms, ear and nose chondritis, and inflammatory arthritis. Patients with VEXAS often develop hematological disorders, such as myelodysplastic syndrome (MDS) and multiple myeloma, with a poor prognosis. Hypercellular bone marrow, vacuolization of erythroid and myeloid precursors, and spontaneously activated peripheral blood myeloid cells are common laboratory findings in patients with VEXAS.

When the symptoms were compared between RP patients with and without VEXAS, fever, ear chondritis, skin involvement (leukocytoclastic vasculitis and neutrophilic dermatosis), and periorbital edema were frequently observed in the patients with the syndrome (125). Notably, RP patients with VEXAS do not develop tracheobronchial chondritis during their clinical course. These data support the hypothesis that local interactions between inflammatory myeloid cells and chondrocytes/extracellular matrix are important for the initiation of chondritis in patients with RP.

## 5. Myelodysplastic syndrome in BD

A recent case report demonstrated that a 60-year-old man with somatic variants of UBA1 developed BD phenotypes with MDS and was resistant to aggressive treatments (126). This report demonstrates the possibility that the clinical spectrum of VEXAS can expand to BD manifestations.

In epidemiological studies, 10–20% of patients with MDS developed autoimmune manifestations (127, 128); conversely, autoimmune manifestations proceeded with the onset of MDS in 30% of patients (129). The prevalent autoimmune manifestations in a retrospective cohort study were neutrophilic dermatoses, such as Sweet syndrome, pyoderma gangrenosum, and BD (127). In this study, the deletion of 5q and trisomy 8 were associated with neutrophilic dermatosis and BD, respectively.

# 6. Mouth and genital ulcers with inflamed cartilage syndrome

Patients with MAGIC syndrome exhibit clinical features of both BD and RP. A prospective cohort study demonstrated good sensitivity in classifying patients according to McAdam's or Damiani's Criteria for RP and the International Criteria for BD (130). Interestingly, in this study, RP patients with MAGIC syndrome demonstrated higher frequency of anti-type 2 collagen autoantibodies than in those without MAGIC syndrome. This finding suggests differences in the underlying molecular mechanisms between the two respective groups of patients.

# 7. Innate immune responses in treatment of BD and RP

Tables 1, 2 show previously reported data on innate immune responses to therapeutic treatment in BD and RP, respectively (59, 131–149).

Colchicine reduced neutrophil and monocyte infiltration into lesions by decreasing the expression levels of adhesion molecules, such as selectin P ligand (SELPLG) and platelet endothelial cell adhesion molecule-1 (PECAM-1) (150). In a study using BD neutrophils, colchicine reduced NET release to an extent similar to that of methylprednisolone, a PAD inhibitor (Cl-amidine), and an ROS inhibitor (N-acetyl cysteine) (59). Biological agents are recommended for the treatment of patients with refractory BD and RP (76, 151).

A phosphodiesterase (PDE)-4 inhibitor, apremilast, reduces degradation of cyclic adenosine monophosphate and inhibits production of proinflammatory cytokines such as IL-12, IL-23, and TNF $\alpha$  from PBMCs (152). Additionally, apremilast decreased the total number of oral ulcers during a 12-week placebo-controlled clinical trial (153).

Zinc plays a crucial role in innate and adaptive immune function and its depletion has led to IL-1 $\beta$  secretion increase of

TABLE 1 Innate immune responses in vivo and in vitro induced by	immunosuppressants in patients with Behçet disease.
---	---

References	Patient numbers	Medication	Study protocols	Laboratory findings		
(131)	80	Colchicine	Peripheral blood samples were obtained before and 1, 3 months after the initiation	Neutrophil-lymphocyte and monocyte-lymphocyte is ratios decreases		
(132)	61	Colchicine, methyl-PSL, PSL	Peripheral blood samples were obtained from active patients with and without the treatment before and 1, 3 months after the initiation	Neutrophil-lymphocyte ratio decrease, plasma IFNγ, IL-4 decease		
(59)	31	Colchicine, dexamethasone, Cl-amidine, N-Acetyl cysteine	Neutrophils were obtained and incubated with compounds ( <i>ex</i> <i>vivo</i> )	NETS release decrease (all compounds)		
(133)	10	Colchicine	Neutrophils and monocytes were obtained and incubated with colchicine ( <i>ex vivo</i> )	Oxidative burst decrease, ROS production decrease		
(134)	N/A	Colchicine	Neutrophils were obtained and incubated with colchicine ( <i>in vitro</i> )	NET release decrease, intracellular ROS levels not change		
(135)	35	Colchicine, PSL	Peripheral blood samples were obtained from active patients with and without treatment	Neutrophil CXCR2 expression decrease only by corticosteroid		
(136)	8	Infliximab	Peripheral blood samples were obtained before and 1 week after infliximab infusion	IFN $\gamma$ , IL-6, TNF $\alpha$ production not change in active patients		
(137)	5	Infliximab	CSF and sera were obtained before, 1 day, and 3, 7, 18 weeks after infliximab infusion	CSF IL-6 decrease, CSF TNF $\alpha$ and serum IL-6 not change		
(138)	18	Infliximab	Peripheral blood samples were obtained before and 1 day after infliximab infusion ( <i>ex vivo</i> )	$TNF\alpha$ decease and IFN $\gamma,$ IL-12R increase		
(139)	7	Gevokizumab	Peripheral blood samples were obtained before and 7 days after infliximab infusion ( <i>ex vivo</i> )	$IL\text{-}1\beta$ decrease and IL-1ra not change		
(140)	12	Colchicine, apremilast	Neutrophils were collected before and 12 weeks after treatment and incubated with colchicine and apremilast ( <i>ex</i> <i>vivo</i> )	NET release and CD11b, CD64, CD66b-expressing cells decrease by both		

#### TABLE 1 (Continued)

References	Patient numbers	Medication	Study protocols	Laboratory findings
(141)	10	Azithromycin	Peripheral blood samples were obtained and incubated with azithromycin ( <i>ex vivo</i> )	IFNγ production decrease
(142)	50	Zinc supplementation	Patients were randomly allocated into zinc gluconate or placebo groups for 12 weeks	Caspase-1/NLRP3 expressions, and serum IL-1 $\beta$ decrease

PSL, prednisolone. Cl-amidine, an inhibitor for PAD4 (a key enzyme of NET formation). N-Acetyl cysteine, a reactive oxygen species (ROS) inhibitor. N/A: not applicable. Infliximab: anti-TNF antibody, Gevokizumab: anti-IL-1β antibody.

TABLE 2 Induction of innate immune responses by administration of immunosuppressants in patients with relapsing polychondritis.

References	Age/ gender	Clinical phenotypes	Medications	Previous medications	Clinical responses	Laboratory findings
(143)	29/f	Respiratory	Tocilizumab	PSL, CsA, TAC, CP, infliximab	PSL reduction to 10 mg/day	Serum MMP3 decrease
(143)	52/m	Respiratory	Tocilizumab	PSL, MTX	PSL reduction to 10 mg/day	Serum MMP3 decrease
(144)	65/f	Respiratory	Tocilizumab	PSL, CP	PSL reduction to 0 mg/day	Serum IL-6 decrease
(145)	55/m	Respiratory	Adalimumab	PSL, MTX, AZA	PSL reduction to 8 mg/day	Serum MMP3 decrease
(146)	68/m	Auricular	Adalimumab	PSL	PSL reduction to 20 mg/day	Serum IL-6, Anti- type 2 collagen antibody titer decrease
(147)	18/m	Respiratory	Infliximab, Etanercept	PSL, MTX, CsA	MTX resumption after biologic therapy cessation	Serum IFNγ, IL-2, IL-12 decrease
(148)	61/m	Auricular + meningoencephalitis	Methyl-PSL	None	Follow-up with PSL and MTX	CSF IL-6 decrease
(149)	66/m	VEXAS	Tocilizumab	PSL	PSL dose before 9 mg/day to 3 mg/ day for 8 months	High levels of IL-6, IL-1β, IFNγ, TNFα sustained
(149)	74/m	VEXAS	Tocilizumab	PSL, MTX	PSL dose before 22.5 mg/day to 13.5 mg/day for 5 months	High levels of IL-6, IL-1β, IFNγ sustained
(149)	67/m	VEXAS	Tocilizumab	PSL, AZA, colchicine	PSL dose before 30 mg/day to 30 mg/ day for 5 months	High levels of IL-6, IL-1β, IFNγ, TNFα sustained

PSL, prednisolone; CsA, cyclosporine; TAC, tacrolimus; CP, cyclophosphamide; MMP, matrix metalloproteinase; MTX, methotrexate; AZA, azathioprine; CSF, cerebrospinal fluid; VEXAS, vacuole, E1 enzyme, X-linked, autoinflammatory, somatic syndrome. Tocilizumab: anti-IL-6R antibody, Adalimumab: anti-TNF antibody, Infliximab: anti-TNF antibody, Etanercept: anti-TNF antibody.

stimulated macrophages through induction of NLRP3 inflammasome (154). Zinc gluconate supplementation reduced the expression levels of serum IL-1 $\beta$  and white blood cell NLRP3, while decreasing the incidence of genital ulcer in patients with BD within 3 months (142).

Sustained high concentrations of inflammatory cytokines have been observed in patients with VEXAS, indicating the refractory nature of the disease, even after the initiation of an anti-IL-6 agent, tocilizumab, administration (149).

## 8. Conclusion

This review collates and summarizes the recent advances in our understanding of the innate cell mediated immunopathology of BD and RP. These studies suggest that innate immune cells are crucial in the direct initiation of local inflammation under dysregulated lymphocyte function in inflammatory diseases (Figure 1). Certain susceptibility genes characterizing BD are associated with several rare monogenic autoinflammatory diseases, such as FMF and TRAPS. Meanwhile, RP is characterized by distinct clinical phenotypes that are associated with several autoantigens in humans and mice. Adaptive immune cells and genetic/environmental factors simultaneously enhance innate immune responses in BD and RP. The identification of controllable active elements is important for the development of effective and safe treatment approaches.

## Author contributions

JS and MM conceived and prepared the manuscript. YMiz, NA, KT, and YMiy prepared and edited the manuscript. JS, MM, YMiz, NA, KT, and YMiy approved the published version of the manuscript.

## Funding

The work of YMiy is supported by the Japanese Society for the Promotion of Science (JSPS) KAKENHI grant number JP22K08531 and AMED under Grant Number 22jm0210069h004 and 22jm0610070h0001, Kato Memorial Bioscience Foundation, The Naito Foundation and The Uehara Memorial Foundation, MM is

## References

1. Sakane T, Takeno M, Suzuki N, Inaba G. Behcet's disease. N Engl J Med. (1999) 341:1284–91.

2. Criteria for diagnosis of Behcet's disease. International study group for Behcet's disease. *Lancet*. (1990) 335:1078-80.

3. Mays JW, Sarmadi M, Moutsopoulos NM. Oral manifestations of systemic autoimmune and inflammatory diseases: diagnosis and clinical management. *J Evid Based Dent Pract.* (2012) 12:265–82. doi: 10.1016/S1532-3382(12)70051-9

4. Hou S, Li N, Liao X, Kijlstra A, Yang P. Uveitis genetics. *Exp Eye Res.* (2020) 190:107853. doi: 10.1016/j.exer.2019.107853

5. Singer O. Cogan and Behcet syndromes. *Rheum Dis Clin N Am.* (2015) 41:75–91.viii. doi: 10.1016/j.rdc.2014.09.007

6. Borhani-Haghighi A, Kardeh B, Banerjee S, Yadollahikhales G, Safari A, Sahraian MA, et al. Neuro-Behcet's disease: an update on diagnosis, differential diagnoses, and treatment. *Mult Scler Relat Disord*. (2019) 39:101906. doi: 10.1016/j.msard.2019.101906

7. Skef W, Hamilton MJ, Arayssi T. Gastrointestinal Behcet's disease: a review. *World J Gastroenterol.* (2015) 21:3801–12. doi: 10.3748/wjg.v21.i13.3801

 Letko E, Zafirakis P, Baltatzis S, Voudouri A, Livir-Rallatos C, Foster CS. Relapsing polychondritis: a clinical review. Semin Arthritis Rheum. (2002) 31:384–95. doi: 10.1053/ sarh.2002.32586

9. Shimizu J, Yamano Y, Kawahata K, Suzuki N. Elucidation of predictors of disease progression in patients with relapsing polychondritis at the onset: potential impact on patient monitoring. *BMC Rheumatol.* (2020) 4:41. doi: 10.1186/s41927-020-00141-8

10. McAdam LP, O'Hanlan MA, Bluestone R, Pearson CM. Relapsing polychondritis: prospective study of 23 patients and a review of the literature. *Medicine (Baltimore)*. (1976) 55:193–215.

11. Damiani JM, Levine HL. Relapsing polychondritis--report of ten cases. Laryngoscope. (1979) 89:929-46.

12. Michet CJ Jr, McKenna CH, Luthra HS, O'Fallon WM. Relapsing polychondritis. Survival and predictive role of early disease manifestations. *Ann Intern Med.* (1986) 104:74–8.

13. Trentham DE, Le CH. Relapsing polychondritis. Ann Intern Med. (1998) 129:114-22. doi: 10.7326/0003-4819-129-2-199807150-00011

14. McGonagle D, McDermott MF. A proposed classification of the immunological diseases. *PLoS Med.* (2006) 3:e297. doi: 10.1371/journal.pmed.0030297

15. Verity DH, Marr JE, Ohno S, Wallace GR, Stanford MR. Behcet's disease, the silk road and HLA-B51: historical and geographical perspectives. *Tissue Antigens*. (1999) 54:213–20.

16. Mumcu G, Ergun T, Inanc N, Fresko I, Atalay T, Hayran O, et al. Oral health is impaired in Behcet's disease and is associated with disease severity. *Rheumatology* (*Oxford*). (2004) 43:1028–33. doi: 10.1093/rheumatology/keh236

supported by JSPS KAKENHI (JP21K06955 and JP21H02394), SRF foundation (2022Y003), Kansai Medical University alumni association (Katano Prize) and Kansai Medical University Molecular Imaging Center of Diseases. The work of JS and MM is supported by JSPS KAKENHI (JP22K08532 and JP23K05618). The work of NA and MM is supported by JSPS KAKENHI (JP21K07379).

## **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

17. Yokota K, Hayashi S, Araki Y, Isogai E, Kotake S, Yoshikawa K, et al. Characterization of *Streptococcus sanguis* isolated from patients with Behçet's disease. *Microbiol Immunol.* (1995) 39:729–32. doi: 10.1111/j.1348-0421.1995.tb03249.x

18. Senusi A, Kang A, Buchanan JAG, Adesanya A, Aloraini G, Stanford M, et al. New mouthwash: an efficacious intervention for oral ulceration associated with Behçet's disease. *Br J Oral Maxillofac Surg*. (2020) 58:1034–9. doi: 10.1016/j.bjoms.2020.07.027

19. Seoudi N, Bergmeier LA, Drobniewski F, Paster B, Fortune F. The oral mucosal and salivary microbial community of Behcet's syndrome and recurrent aphthous stomatitis. *J Oral Microbiol.* (2015) 7:27150. doi: 10.3402/jom.v7.27150

20. Coit P, Mumcu G, Ture-Ozdemir F, Unal AU, Alpar U, Bostanci N, et al. Sequencing of 16S rRNA reveals a distinct salivary microbiome signature in Behcet's disease. *Clin Immunol.* (2016) 169:28–35. doi: 10.1016/j.clim.2016.06.002

21. Consolandi C, Turroni S, Emmi G, Severgnini M, Fiori J, Peano C, et al. Behçet's syndrome patients exhibit specific microbiome signature. *Autoimmun Rev.* (2015) 14:269–76. doi: 10.1016/j.autrev.2014.11.009

22. Shimizu J, Kubota T, Takada E, Takai K, Fujiwara N, Arimitsu N, et al. Bifidobacteria abundance-featured gut microbiota compositional change in patients with Behcet's disease. *PLoS One.* (2016) 11:e0153746. doi: 10.1371/journal. pone.0153746

23. Takeno M, Kariyone A, Yamashita N, Takiguchi M, Mizushima Y, Kaneoka H, et al. Excessive function of peripheral blood neutrophils from patients with Behcet's disease and from HLA-B51 transgenic mice. *Arthritis Rheum*. (1995) 38:426–33. doi: 10.1002/art.1780380321

24. Takeuchi M, Mizuki N, Meguro A, Ombrello MJ, Kirino Y, Satorius C, et al. Dense genotyping of immune-related loci implicates host responses to microbial exposure in Behçet's disease susceptibility. *Nat Genet.* (2017) 49:438–43. doi: 10.1038/ng.3786

25. Kirino Y, Bertsias G, Ishigatsubo Y, Mizuki N, Tugal-Tutkun I, Seyahi E, et al. Genome-wide association analysis identifies new susceptibility loci for Behçet's disease and epistasis between HLA-B\*51 and ERAP1. *Nat Genet.* (2013) 45:202–7. doi: 10.1038/ng.2520

26. Kirino Y, Zhou Q, Ishigatsubo Y, Mizuki N, Tugal-Tutkun I, Seyahi E, et al. Targeted resequencing implicates the familial Mediterranean fever gene MEFV and the toll-like receptor 4 gene TLR4 in Behçet disease. *Proc Natl Acad Sci U S A*. (2013) 110:8134–9. doi: 10.1073/pnas.1306352110

27. Hughes T, Coit P, Adler A, Yilmaz V, Aksu K, Düzgün N, et al. Identification of multiple independent susceptibility loci in the HLA region in Behçet's disease. *Nat Genet.* (2013) 45:319–24. doi: 10.1038/ng.2551

28. Mizuki N, Meguro A, Ota M, Ohno S, Shiota T, Kawagoe T, et al. Genome-wide association studies identify IL23R-IL12RB2 and IL10 as Behçet's disease susceptibility loci. *Nat Genet*. (2010) 42:703–6. doi: 10.1038/ng.624

29. Remmers EF, Cosan F, Kirino Y, Ombrello MJ, Abaci N, Satorius C, et al. Genomewide association study identifies variants in the MHC class I, IL10, and IL23R-IL12RB2 regions associated with Behçet's disease. Nat Genet. (2010) 42:698–702. doi: 10.1038/ ng.625

30. Pervin K, Childerstone A, Shinnick T, Mizushima Y, van der Zee R, Hasan A, et al. T cell epitope expression of mycobacterial and homologous human 65-kilodalton heat shock protein peptides in short term cell lines from patients with Behçet's disease. *J Immunol.* (1993) 151:2273–82. doi: 10.4049/jimmunol.151.4.2273

31. Filleron A, Tran TA, Hubert A, Letierce A, Churlaud G, Koné-Paut I, et al. Regulatory T cell/Th17 balance in the pathogenesis of paediatric Behçet disease. *Rheumatology (Oxford).* (2021) 61:422–9. doi: 10.1093/rheumatology/keab253

32. van der Houwen TB, Dik WA, Goeijenbier M, Hayat M, Nagtzaam NMA, van Hagen M, et al. Leukocyte toll-like receptor expression in pathergy positive and negative Behçet's disease patients. *Rheumatology (Oxford)*. (2020) 59:3971–9. doi: 10.1093/rheumatology/keaa251

33. Zhou Q, Wang H, Schwartz DM, Stoffels M, Park YH, Zhang Y, et al. Loss-offunction mutations in TNFAIP3 leading to A20 haploinsufficiency cause an early-onset autoinflammatory disease. *Nat Genet*. (2016) 48:67–73. doi: 10.1038/ng.3459

34. Soejima Y, Kirino Y, Takeno M, Kurosawa M, Takeuchi M, Yoshimi R, et al. Changes in the proportion of clinical clusters contribute to the phenotypic evolution of Behçet's disease in Japan. *Arthritis Res Ther.* (2021) 23:49. doi: 10.1186/s13075-020-02406-6

35. Bettiol A, Hatemi G, Vannozzi L, Barilaro A, Prisco D, Emmi G. Treating the different phenotypes of Behçet's syndrome. *Front Immunol.* (2019) 10:2830. doi: 10.3389/fimmu.2019.02830

36. Nakamura K, Tsunemi Y, Kaneko F, Alpsoy E. Mucocutaneous manifestations of Behcet's disease. *Front Med.* (2020) 7:613432. doi: 10.3389/fmed.2020.613432

37. Misago N, Tada Y, Koarada S, Narisawa Y. Erythema nodosum-like lesions in Behcet's disease: a clinicopathological study of 26 cases. *Acta Derm Venereol.* (2012) 92:681–6. doi: 10.2340/00015555-1349

38. Jorizzo JL, Abernethy JL, White WL, Mangelsdorf HC, Zouboulis CC, Sarica R, et al. Mucocutaneous criteria for the diagnosis of Behçet's disease: an analysis of clinicopathologic data from multiple international centers. *J Am Acad Dermatol.* (1995) 32:968–76. doi: 10.1016/0190-9622(95)91333-5

39. Demirkesen C, Tüzüner N, Mat C, Senocak M, Büyükbabani N, Tüzün Y, et al. Clinicopathologic evaluation of nodular cutaneous lesions of Behçet syndrome. *Am J Clin Pathol.* (2001) 116:341–6. doi: 10.1309/GCTH-0060-55K8-XCTT

40. Ergun T. Pathergy phenomenon. Front Med. (2021) 8:639404. doi: 10.3389/fmed.2021.639404

41. Dilsen N, Konice M, Aral O, Ocal L, Inanc M, Gul A. Comparative study of the skin pathergy test with blunt and sharp needles in Behcet's disease: confirmed specificity but decreased sensitivity with sharp needles. *Ann Rheum Dis.* (1993) 52:823–5.

42. Shenavandeh S, Sadeghi SMK, Aflaki E. Pathergy test with a 23G needle with and without self-saliva in patients with Behcet's disease, recurrent aphthous stomatitis and control group compared to the 20G test. *Reumatologia*. (2021) 59:302–8. doi: 10.5114/ reum.2021.110567

43. Melikoglu M, Uysal S, Krueger JG, Kaplan G, Gogus F, Yazici H, et al. Characterization of the divergent wound-healing responses occurring in the pathergy reaction and normal healthy volunteers. *J Immunol.* (2006) 177:6415–21. doi: 10.4049/jimmunol.177.9.6415

44. Ergun T, Gürbüz O, Harvell J, Jorizzo J, White W. The histopathology of pathergy: a chronologic study of skin hyperreactivity in Behçet's disease. *Int J Dermatol.* (1998) 37:929–33. doi: 10.1046/j.1365-4362.1998.00474.x

45. Gül A, Esin S, Dilsen N, Koniçe M, Wigzell H, Biberfeld P. Immunohistology of skin pathergy reaction in Behçet's disease. *Br J Dermatol.* (1995) 132:901–7. doi: 10.1111/j.1365-2133.1995.tb16946.x

46. Chun SI, Su WP, Lee S. Histopathologic study of cutaneous lesions in Behçet's syndrome. J Dermatol. (1990) 17:333–41.

47. Köklü S, Yüksel O, Onur I, Ünverdi S, Bıyıkoğlu İ, Akbal E, et al. Ileocolonic involvement in Behçet's disease: endoscopic and histological evaluation. *Digestion*. (2010) 81:214–7. doi: 10.1159/000264643

48. Hayasaki N, Ito M, Suzuki T, Ina K, Ando T, Kusugami K, et al. Neutrophilic phlebitis is characteristic of intestinal Behçet's disease and simple ulcer syndrome. *Histopathology*. (2004) 45:377–83. doi: 10.1111/j.1365-2559.2004.01954.x

49. Takada Y, Fujita Y, Igarashi M, Katsumata T, Okabe H, Saigenji K, et al. Intestinal Behçet's disease--pathognomonic changes in intramucosal lymphoid tissues and effect of a "rest cure" on intestinal lesions. *J Gastroenterol.* (1997) 32:598–604. doi: 10.1007/BF02934108

50. Hirohata S, Kikuchi H, Sawada T, Nagafuchi H, Kuwana M, Takeno M, et al. Clinical characteristics of neuro-Behcet's disease in Japan: a multicenter retrospective analysis. *Mod Rheumatol.* (2012) 22:405–13. doi: 10.3109/s10165-011-0533-5

51. Burn GL, Foti A, Marsman G, Patel DF, Zychlinsky A. The neutrophil. *Immunity*. (2021) 54:1377–91. doi: 10.1016/j.immuni.2021.06.006

52. Carletto A, Pacor ML, Biasi D, Caramaschi P, Zeminian S, Bellavite P, et al. Changes of neutrophil migration without modification of in vitro metabolism and adhesion in Behçet's disease. *J Rheumatol.* (1997) 24:1332–6.

53. Perazzio SF, Soeiro-Pereira PV, de Souza AW, Condino-Neto A, Andrade LE. Behçet's disease heterogeneity: cytokine production and oxidative burst of phagocytes are altered in patients with severe manifestations. *Clin Exp Rheumatol.* (2015) 33:S85–95.

54. Brinkmann V, Reichard U, Goosmann C, Fauler B, Uhlemann Y, Weiss DS, et al. Neutrophil extracellular traps kill bacteria. *Science*. (2004) 303:1532–5. doi: 10.1126/ science.1092385

55. Wang Y, Wysocka J, Sayegh J, Lee YH, Perlin JR, Leonelli L, et al. Human PAD4 regulates histone arginine methylation levels via demethylimination. *Science*. (2004) 306:279–83. doi: 10.1126/science.1101400

56. Warnatsch A, Ioannou M, Wang Q, Papayannopoulos V. Inflammation. Neutrophil extracellular traps license macrophages for cytokine production in atherosclerosis. *Science.* (2015) 349:316–20. doi: 10.1126/science.aaa8064

57. Martinod K, Demers M, Fuchs TA, Wong SL, Brill A, Gallant M, et al. Neutrophil histone modification by peptidylarginine deiminase 4 is critical for deep vein thrombosis in mice. *Proc Natl Acad Sci U S A*. (2013) 110:8674–9. doi: 10.1073/pnas.1301059110

58. le Joncour A, Martos R, Loyau S, Lelay N, Dossier A, Cazes A, et al. Critical role of neutrophil extracellular traps (NETs) in patients with Behcet's disease. *Ann Rheum Dis.* (2019) 78:1274–82. doi: 10.1136/annrheumdis-2018-214335

59. Safi R, Kallas R, Bardawil T, Mehanna CJ, Abbas O, Hamam R, et al. Neutrophils contribute to vasculitis by increased release of neutrophil extracellular traps in Behçet's disease. *J Dermatol Sci.* (2018) 92:143–50. doi: 10.1016/j.jdermsci.2018.08.010

60. Li L, Yu X, Liu J, Wang Z, Li C, Shi J, et al. Neutrophil extracellular traps promote aberrant macrophages activation in Behcet's disease. *Front Immunol.* (2020) 11:590622. doi: 10.3389/fimmu.2020.590622

61. Mistry P, Nakabo S, O'Neil L, Goel RR, Jiang K, Carmona-Rivera C, et al. Transcriptomic, epigenetic, and functional analyses implicate neutrophil diversity in the pathogenesis of systemic lupus erythematosus. *Proc Natl Acad Sci U S A*. (2019) 116:25222–8. doi: 10.1073/pnas.1908576116

62. Murad M, Low L, Davidson M, Murray PI, Rauz S, Wallace GR. Low density neutrophils are increased in patients with Behcet's disease but do not explain differences in neutrophil function. *J Inflamm*. (2022) 19:5. doi: 10.1186/s12950-022-00302-1

63. Jablonski KA, Amici SA, Webb LM, Ruiz-Rosado JD, Popovich PG, Partida-Sanchez S, et al. Novel markers to delineate murine M1 and M2 macrophages. *PLoS One*. (2015) 10:e0145342. doi: 10.1371/journal.pone.0145342

64. Wu X, Wang Z, Shi J, Yu X, Li C, Liu J, et al. Macrophage polarization toward M1 phenotype through NF-κB signaling in patients with Behçet's disease. *Arthritis Res Ther.* (2022) 24:249. doi: 10.1186/s13075-022-02938-z

65. Nakano H, Kirino Y, Takeno M, Higashitani K, Nagai H, Yoshimi R, et al. GWASidentified CCR1 and IL10 loci contribute to M1 macrophage-predominant inflammation in Behçet's disease. *Arthritis Res Ther.* (2018) 20:124. doi: 10.1186/s13075-018-1613-0

66. Verrou KM, Vlachogiannis NI, Ampatziadis-Michailidis G, Moulos P, Pavlopoulos GA, Hatzis P, et al. Distinct transcriptional profile of blood mononuclear cells in Behçet's disease: insights into the central role of neutrophil chemotaxis. *Rheumatology (Oxford)*. (2021) 60:4910–9. doi: 10.1093/rheumatology/keab052

67. Rathinam VA, Fitzgerald KA. Inflammasome complexes: emerging mechanisms and effector functions. *Cells.* (2016) 165:792–800. doi: 10.1016/j.cell.2016.03.046

68. Lamkanfi M, Dixit VM. Inflammasomes and their roles in health and disease. Annu Rev Cell Dev Biol. (2012) 28:137–61. doi: 10.1146/annurev-cellbio-101011-155745

69. Booshehri LM, Hoffman HM. CAPS and NLRP3. J Clin Immunol. (2019) 39:277–86. doi: 10.1007/s10875-019-00638-z

70. Kim EH, Park MJ, Park S, Lee ES. Increased expression of the NLRP3 inflammasome components in patients with Behçet's disease. *J Inflamm*. (2015) 12:41. doi: 10.1186/s12950-015-0086-z

71. Hamzaoui K, Borhani-Haghighi A, Dhifallah IB, Hamzaoui A. Elevated levels of IL-32 in cerebrospinal fluid of neuro-Behcet disease: correlation with NLRP3 inflammasome. *J Neuroimmunol.* (2022) 365:577820. doi: 10.1016/j. jneuroim.2022.577820

72. Chang HK, Lee SS, Kim JW, Jee YK, Kim JU, Lee YW, et al. The prevalence of atopy and atopic diseases in Behçet's disease. *Clin Exp Rheumatol.* (2003) 21:S31-4.

73. Tas DA, Ozer HT, Erken E. Serum eosinophil cationic protein levels in Behçet's disease and its relation to clinical activity. *Asian Pac J Allergy Immunol.* (2013) 31:67–72.

74. Hirahara L, Takase-Minegishi K, Kirino Y, Iizuka-Iribe Y, Soejima Y, Yoshimi R, et al. The roles of monocytes and macrophages in Behçet's disease with focus on M1 and M2 polarization. *Front Immunol.* (2022) 13:852297. doi: 10.3389/fimmu.2022.852297

75. Gholijani N, Ataollahi MR, Samiei A, Aflaki E, Shenavandeh S, Kamali-Sarvestani E. An elevated pro-inflammatory cytokines profile in Behcet's disease: a multiplex analysis. *Immunol Lett.* (2017) 186:46–51. doi: 10.1016/j.imlet.2016.12.001

76. Yazici H, Seyahi E, Hatemi G, Yazici Y. Behçet syndrome: a contemporary view. *Nat Rev Rheumatol.* (2018) 14:119. doi: 10.1038/nrrheum.2018.3

77. Zhong Z, Su G, Kijlstra A, Yang P. Activation of the interleukin-23/interleukin-17 signalling pathway in autoinflammatory and autoimmune uveitis. *Prog Retin Eye Res.* (2021) 80:100866. doi: 10.1016/j.preteyeres.2020.100866

78. Kaburaki T, Fujino Y, Kawashima H, Merino G, Numaga J, Chen J, et al. Plasma and whole-blood chemokine levels in patients with Behcet's disease. *Graefes Arch Clin Exp Ophthalmol.* (2003) 241:353–8. doi: 10.1007/s00417-003-0668-y

79. Ozer HT, Erken E, Gunesacar R, Kara O. Serum RANTES, MIP-1alpha, and MCP-1 levels in Behçet's disease. *Rheumatol Int.* (2005) 25:487–8. doi: 10.1007/s00296-004-0519-0

80. Lee SJ, Kang SE, Kang EH, Choi BY, Masek-Hammerman K, Syed J, et al. CXCL10/ CXCR3 axis is associated with disease activity and the development of mucocutaneous lesions in patients with Behçet's disease. *Sci Rep.* (2017) 7:14720. doi: 10.1038/ s41598-017-15189-9

81. el-Asrar AMA, Berghmans N, al-Obeidan SA, Gikandi PW, Opdenakker G, van Damme J, et al. Differential CXC and CX3C chemokine expression profiles in aqueous humor of patients with specific endogenous Uveitic entities. *Invest Ophthalmol Vis Sci.* (2018) 59:2222–8. doi: 10.1167/iovs.17-23225

82. Pay S, Abbasov T, Erdem H, Musabak U, Simsek I, Pekel A, et al. Serum MMP-2 and MMP-9 in patients with Behçet's disease: do their higher levels correlate to vasculo-Behçet's disease associated with aneurysm formation? *Clin Exp Rheumatol.* (2007) 25:S70–5.

83. Cai D, Sun C, Zhang G, Que X, Fujise K, Weintraub NL, et al. A novel mechanism underlying inflammatory smooth muscle phenotype in abdominal aortic aneurysm. *Circ Res.* (2021) 129:e202–14. doi: 10.1161/CIRCRESAHA.121.319374

84. Pay S, Erdem H, Pekel A, Simsek I, Musabak U, Sengul A, et al. Synovial proinflammatory cytokines and their correlation with matrix metalloproteinase-3 expression in Behçet's disease. Does interleukin-1beta play a major role in Behçet's synovitis? *Rheumatol Int.* (2006) 26:608–13. doi: 10.1007/s00296-005-0040-0

85. Lee KH, Chung HS, Kim HS, Oh SH, Ha MK, Baik JH, et al. Human ?-enolase from endothelial cells as a target antigen of anti-endothelial cell antibody in Behçet's disease. *Arthritis Rheum*. (2003) 48:2025–35. doi: 10.1002/art.11074

86. Xun Y, Chen P, Yan H, Yang W, Shi L, Chen G, et al. Identification of prohibitin as an antigen in Behcet's disease. *Biochem Biophys Res Commun.* (2014) 451:389–93. doi: 10.1016/j.bbrc.2014.07.126

87. Mahesh SP, Li Z, Buggage R, Mor F, Cohen IR, Chew EY, et al. Alpha tropomyosin as a self-antigen in patients with Behçet's disease. *Clin Exp Immunol.* (2005) 140:368–75. doi: 10.1111/j.1365-2249.2005.02760.x

88. Hussain M, Ma F, Chen P, Tian Y, Du H. Circulation autoantibodies against C-terminus of NuMA in patients with Behcet's disease. *Cent Eur J Immunol.* (2020) 45:86–92. doi: 10.5114/ceji.2020.94710

89. Chen P, Yang W, Tian Y, Sun S, Chen G, Zhang C, et al. Electron transfer flavoprotein subunit beta is a candidate endothelial cell autoantigen in Behçet's disease. *PLoS One.* (2015) 10:e0124760. doi: 10.1371/journal.pone.0124760

90. Chen P, Yan H, Tian Y, Xun Y, Shi L, Bao R, et al. Annexin A2 as a target endothelial cell membrane autoantigen in Behçet's disease. *Sci Rep.* (2015) 5:8162. doi: 10.1038/srep08162

91. Lu Y, Ye P, Chen SL, Tan EM, Chan EK. Identification of kinectin as a novel Behcet's disease autoantigen. *Arthritis Res Ther.* (2005) 7:R1133-9. doi: 10.1186/ar1798

92. Ooka S, Nakano H, Matsuda T, Okamoto K, Suematsu N, Kurokawa MS, et al. Proteomic surveillance of autoantigens in patients with Behcet's disease by a proteomic approach. *Microbiol Immunol.* (2010) 54:354–61. doi: 10.1111/j.1348-0421.2010.00215.x

93. Mathew SD, Battafarano DF, Morris MJ. Relapsing polychondritis in the Department of Defense population and review of the literature. *Semin Arthritis Rheum.* (2012) 42:70–83. doi: 10.1016/j.semarthrit.2011.12.007

94. Horvath A, Pall N, Molnar K, Kovats T, Surjan G, Vicsek T, et al. A nationwide study of the epidemiology of relapsing polychondritis. *Clin Epidemiol.* (2016) 8:211–30. doi: 10.2147/CLEP.S91439

95. Hazra N, Dregan A, Charlton J, Gulliford MC, D'Cruz DP. Incidence and mortality of relapsing polychondritis in the UK: a population-based cohort study. *Rheumatology* (*Oxford*). (2015) 54:2181–7. doi: 10.1093/rheumatology/kev240

96. Lang B, Rothenfusser A, Lanchbury JS, Rauh G, Breedveld FC, Urlacher A, et al. Susceptibility to relapsing polychondritis is associated with HLA-DR4. *Arthritis Rheum*. (1993) 36:660–4. doi: 10.1002/art.1780360513

97. Terao C, Yoshifuji H, Yamano Y, Kojima H, Yurugi K, Miura Y, et al. Genotyping of relapsing polychondritis identified novel susceptibility HLA alleles and distinct genetic characteristics from other rheumatic diseases. *Rheumatology (Oxford)*. (2016) 55:1686–92. doi: 10.1093/rheumatology/kew233

98. Shimizu J, Takai K, Takada E, Fujiwara N, Arimitsu N, Ueda Y, et al. Possible association of proinflammatory cytokines including IL1beta and TNFalpha with enhanced Th17 cell differentiation in patients with Behcet's disease. *Clin Rheumatol.* (2016) 35:1857–63. doi: 10.1007/s10067-015-2966-2

99. de Montmollin N, Dusser D, Lorut C, Dion J, Costedoat-Chalumeau N, Mouthon L, et al. Tracheobronchial involvement of relapsing polychondritis. *Autoimmun Rev.* (2019) 18:102353. doi: 10.1016/j.autrev.2019.102353

100. Liu Y, Li X, Cheng L, Zhan H, Huang Y, Li H, et al. Progress and challenges in the use of blood biomarkers in relapsing polychondritis. *Clin Exp Immunol.* (2023) 212:199–211. doi: 10.1093/cei/uxad014

101. Shimizu J, Yamano Y, Kawahata K, Suzuki N. Relapsing polychondritis patients were divided into three subgroups: patients with respiratory involvement (R subgroup), patients with auricular involvement (A subgroup), and overlapping patients with both involvements (O subgroup), and each group had distinctive clinical characteristics. *Medicine (Baltimore).* (2018) 97:e12837. doi: 10.1097/MD.00000000012837

102. Yang P, Yuan W, du L, Zhou Q, Wang C, Ye Z, et al. Clinical features of Chinese patients with relapsing polychondritis. *Br J Ophthalmol.* (2019) 103:1129–32. doi: 10.1136/bjophthalmol-2018-312660

103. Kumakiri K, Sakamoto T, Karahashi T, Mineta H, Takebayashi S. A case of relapsing polychondritis preceded by inner ear involvement. *Auris Nasus Larynx.* (2005) 32:71–6. doi: 10.1016/j.anl.2004.09.012

104. Kobayashi T, Moody S, Komori M, Jibatake A, Yaegashi M. Early stage relapsing polychondritis diagnosed by nasal septum biopsy. *Case Rep Med.* (2015) 2015:307868. doi: 10.1155/2015/307868

105. Ouchi N, Uzuki M, Kamataki A, Miura Y, Sawai T. Cartilage destruction is partly induced by the internal proteolytic enzymes and apoptotic phenomenon of chondrocytes in relapsing polychondritis. *J Rheumatol.* (2011) 38:730–7. doi: 10.3899/jrheum.101044

106. Zhang L, Yun S, Wu T, He Y, Guo J, Han L, et al. Clinical patterns and the evolution of relapsing polychondritis based on organ involvement: a Chinese retrospective cohort study. *Orphanet J Rare Dis.* (2021) 16:225. doi: 10.1186/s13023-021-01861-x

107. Francès C, el Rassi R, Laporte JL, Rybojad M, Papo T, Piette JC. Dermatologic manifestations of relapsing polychondritis. A study of 200 cases at a single center. *Medicine (Baltimore)*. (2001) 80:173–9. doi: 10.1097/00005792-200105000-00003

108. Kondo T, Fukuta M, Takemoto A, Takami Y, Sato M, Takahashi N, et al. Limbic encephalitis associated with relapsing polychondritis responded to infliximab and maintained its condition without recurrence after discontinuation: a case report and review of the literature. *Nagoya J Med Sci.* (2014) 76:361–8.

109. Fujiki F, Tsuboi Y, Hashimoto K, Nakajima M, Yamada T. Non-herpetic limbic encephalitis associated with relapsing polychondritis. *J Neurol Neurosurg Psychiatry*. (2004) 75:1646–7. doi: 10.1136/jnnp.2003.035170

110. Storey K, Matěj R, Rusina R. Unusual association of seronegative, nonparaneoplastic limbic encephalitis and relapsing polychondritis in a patient with history of thymectomy for myasthenia: a case study. *J Neurol.* (2011) 258:159–61. doi: 10.1007/s00415-010-5691-4

111. Yan M, Cooper W, Harper C, Schwartz R. Dementia in a patient with nonparaneoplastic limbic encephalitis associated with relapsing polychondritis. *Pathology*. (2006) 38:596–9. doi: 10.1080/00313020601023989

112. Shimizu J, Kubota T, Takada E, Takai K, Fujiwara N, Arimitsu N, et al. Propionate-producing bacteria in the intestine may associate with skewed responses of IL10-producing regulatory T cells in patients with relapsing polychondritis. *PLoS One*. (2018) 13:e0203657. doi: 10.1371/journal.pone.0203657

113. Shimizu J, Suzuki N. Mechanical model of steady-state and inflammatory conditions in patients with relapsing polychondritis: a review. *Medicine (Baltimore)*. (2022) 101:e28852. doi: 10.1097/MD.00000000028852

114. Yu EN, Jurkunas U, Rubin PA, Baltatzis S, Foster CS. Obliterative microangiopathy presenting as chronic conjunctivitis in a patient with relapsing polychondritis. *Cornea*. (2006) 25:621–2. doi: 10.1097/01.ico.0000227886.26747.a9

115. Khan JH, Ahmed I. A case of relapsing polychondritis involving the tragal and the Conchal bowl areas with sparing of the helix and the antihelix. *J Am Acad Dermatol.* (1999) 41:299–302.

116. Sato T, Yamano Y, Tomaru U, Shimizu Y, Ando H, Okazaki T, et al. Serum level of soluble triggering receptor expressed on myeloid cells-1 as a biomarker of disease activity in relapsing polychondritis. *Mod Rheumatol.* (2014) 24:129–36. doi: 10.3109/14397595.2013.852854

117. Stabler T, Piette JC, Chevalier X, Marini-Portugal A, Kraus VB. Serum cytokine profiles in relapsing polychondritis suggest monocyte/macrophage activation. *Arthritis Rheum*. (2004) 50:3663–7. doi: 10.1002/art.20613

118. Shimizu J, Wakisaka S, Suzuki T, Suzuki N. Serum MMP3 correlated with IL1beta messenger RNA expressions of peripheral blood mononuclear cells in patients with relapsing Polychondritis with respiratory involvement. *ACR Open Rheumatol.* (2021) 3:636–41. doi: 10.1002/acr2.11301

119. Tammaro A, Stroo I, Rampanelli E, Blank F, Butter LM, Claessen N, et al. Role of TREM1-DAP12 in renal inflammation during obstructive nephropathy. *PLoS One*. (2013) 8:e82498. doi: 10.1371/journal.pone.0082498

120. Foidart JM, Abe S, Martin GR, Zizic TM, Barnett EV, Lawley TJ, et al. Antibodies to type II collagen in relapsing polychondritis. *N Engl J Med.* (1978) 299:1203–7. doi: 10.1056/NEJM197811302992202

121. Cremer MA, Pitcock JA, Stuart JM, Kang AH, Townes AS. Auricular chondritis in rats. An experimental model of relapsing polychondritis induced with type II collagen. *J Exp Med.* (1981) 154:535–40.

122. Saxne T, Heinegard D. Involvement of nonarticular cartilage, as demonstrated by release of a cartilage-specific protein, in rheumatoid arthritis. *Arthritis Rheum*. (1989) 32:1080–6.

123. Hansson AS, Heinegard D, Piette JC, Burkhardt H, Holmdahl R. The occurrence of autoantibodies to matrilin 1 reflects a tissue-specific response to cartilage of the respiratory tract in patients with relapsing polychondritis. *Arthritis Rheum*. (2001) 44:2402–12. doi: 10.1002/1529-0131(200110)44:10<2402::aid-art405>3.0.co;2-l

124. Beck DB, Ferrada MA, Sikora KA, Ombrello AK, Collins JC, Pei W, et al. Somatic mutations in UBA1 and severe adult-onset autoinflammatory disease. *N Engl J Med.* (2020) 383:2628–38. doi: 10.1056/NEJMoa2026834

125. Ferrada MA, Sikora KA, Luo Y, Wells KV, Patel B, Groarke EM, et al. Somatic mutations in UBA1 define a distinct subset of relapsing polychondritis patients with VEXAS. *Arthritis Rheum.* (2021) 73:1886–95. doi: 10.1002/art.41743

126. Matsumoto H, Asano T, Tsuchida N, Maeda A, Yoshida S, Yokose K, et al. Behçet's disease with a somatic UBA1 variant: expanding spectrum of autoinflammatory phenotypes of VEXAS syndrome. *Clin Immunol.* (2022) 238:108996. doi: 10.1016/j. clim.2022.108996

127. Lee SJ, Park JK, Lee EY, Joo SH, Jung KC, Lee EB, et al. Certain autoimmune manifestations are associated with distinctive karyotypes and outcomes in patients with myelodysplastic syndrome: a retrospective cohort study. *Medicine (Baltimore)*. (2016) 95:e3091. doi: 10.1097/MD.000000000003091

128. Hochman MJ, DeZern AE. Myelodysplastic syndrome and autoimmune disorders: two sides of the same coin? *Lancet Haematol*. (2022) 9:e523–34. doi: 10.1016/S2352-3026(22)00138-7

129. Grignano E, Jachiet V, Fenaux P, Ades L, Fain O, Mekinian A. Autoimmune manifestations associated with myelodysplastic syndromes. *Ann Hematol.* (2018) 97:2015–23. doi: 10.1007/s00277-018-3472-9

130. Luo Y, Bolek EC, Quinn KA, Wells K, Rose E, Sikora K, et al. A prospective observational cohort study and systematic review of 40 patients with mouth and genital ulcers with inflamed cartilage (MAGIC) syndrome. *Semin Arthritis Rheum.* (2022) 52:151924. doi: 10.1016/j.semarthrit.2021.10.007

131. Demirbaş A, Kaya İslamoğlu ZG. Can decreased monocyte to HDL-cholesterol ratio be a marker indicating the anti-inflammatory effect of the colchicine in Behçet's disease? A preliminary study. *Dermatol Ther.* (2020) 33:e14013. doi: 10.1111/dth.14013

132. Djaballah-Ider F, Touil-Boukoffa C. Effect of combined colchicine-corticosteroid treatment on neutrophil/lymphocyte ratio: a predictive marker in Behçet disease activity. *Inflammopharmacology*. (2020) 28:819–29. doi: 10.1007/s10787-020-00701-x

133. Davtyan TK, Mkrtchyan NR, Manukyan HM, Avetisyan SA. Dexamethasone, colchicine and iodine-lithium-alpha-dextrin act differentially on the oxidative burst and endotoxin tolerance induction in vitro in patients with Behçet's disease. *Int Immunopharmacol.* (2006) 6:396–407. doi: 10.1016/j.intimp.2005.09.003

134. Bettiol A, Becatti M, Silvestri E, Argento FR, Fini E, Mannucci A, et al. Neutrophil-mediated mechanisms of damage and in-vitro protective effect of colchicine in non-vascular Behcet's syndrome. *Clin Exp Immunol.* (2021) 206:410–21. doi: 10.1111/cei.13664

135. Qiao H, Sonoda KH, Ariyama A, Kuratomi Y, Kawano Y, Ishibashi T. CXCR2 expression on neutrophils is upregulated during the relapsing phase of ocular Behcet disease. *Curr Eye Res.* (2005) 30:195–203. doi: 10.1080/02713680490904331

136. Takeuchi M, Karasawa Y, Harimoto K, Tanaka A, Shibata M, Sato T, et al. Analysis of Th cell-related cytokine production in Behçet disease patients with uveitis before and after infliximab treatment. *Ocul Immunol Inflamm.* (2017) 25:52–61. doi: 10.3109/09273948.2016.1158276

137. Kikuchi H, Aramaki K, Hirohata S. Effect of infliximab in progressive neuro-Behçet's syndrome. J Neurol Sci. (2008) 272:99–105. doi: 10.1016/j.jns.2008.05.002

138. Misumi M, Hagiwara E, Takeno M, Takeda Y, Inoue Y, Tsuji T, et al. Cytokine production profile in patients with Behcet's disease treated with infliximab. *Cytokine*. (2003) 24:210–8. doi: 10.1016/j.cyto.2003.09.003

139. Gül A, Tugal-Tutkun I, Dinarello CA, Reznikov L, Esen BA, Mirza A, et al. Interleukin-1 $\beta$ -regulating antibody XOMA 052 (gevokizumab) in the treatment of acute exacerbations of resistant uveitis of Behcet's disease: an open-label pilot study. *Ann Rheum Dis.* (2012) 71:563–6. doi: 10.1136/annrheumdis-2011-155143 140. le Joncour A, Régnier P, Maciejewski-Duval A, Charles E, Barete S, Fouret P, et al. Type-4 Phosphodiesterase (PDE4) blockade reduces neutrophil activation in Behçet's disease *Arthritis Rheum* (2023), doi: 10.1002/art.42486 [Epub ahead of print].

141. Mumcu G, İnanç N, Özdemir FT, Tulunay A, Ekşioğlu-Demiralp E, Ergun T, et al. Effects of azithromycin on intracellular cytokine responses and mucocutaneous manifestations in Behçet's disease. *Int J Dermatol.* (2013) 52:1561–6. doi: 10.1111/ ijd.12144

142. Faghfouri AH, Baradaran B, Khabbazi A, Abdoli Shadbad M, Papi S, Faghfuri E, et al. Regulation of NLRP3 inflammasome by zinc supplementation in Behçet's disease patients: a double-blind, randomized placebo-controlled clinical trial. *Int Immunopharmacol.* (2022) 109:108825. doi: 10.1016/j.intimp.2022.108825

143. Kawai M, Hagihara K, Hirano T, Shima Y, Kuwahara Y, Arimitsu J, et al. Sustained response to tocilizumab, anti-interleukin-6 receptor antibody, in two patients with refractory relapsing polychondritis. *Rheumatology (Oxford)*. (2009) 48:318–9. doi: 10.1093/rheumatology/ken468

144. Wallace ZS, Stone JH. Refractory relapsing polychondritis treated with serial success with interleukin 6 receptor blockade. *J Rheumatol.* (2013) 40:100–1. doi: 10.3899/jrheum.120381

145. Maekawa M, Yoshimura M, Kadowaki M, Nakano M, Moriwaki A, Ueda H, et al. Successful treatment of relapsing polychondritis with circumferential bronchial wall thickening including the tracheomembranous area with tumor necrosis factor- $\alpha$ inhibitor. *Mod Rheumatol Case Rep.* (2023) 7:197–201. doi: 10.1093/mrcr/rxac005

146. Nakamura H, Suzuki T, Nagaoka K, Yamasaki S, Tamai M, Hayashi T, et al. Efficacy of adalimumab for a refractory case of relapsing polychondritis with reduction of pro-inflammatory cytokines. *Mod Rheumatol.* (2011) 21:665–8. doi: 10.3109/ s10165-011-0453-4

147. Kraus VB, Stabler T, Le ET, Saltarelli M, Allen NB. Urinary type II collagen neoepitope as an outcome measure for relapsing polychondritis. *Arthritis Rheum*. (2003) 48:2942–8. doi: 10.1002/art.11281

148. Matsumoto H, Tokimura R, Fujita Y, Matsuoka N, Asano T, Sato S, et al. Meningoencephalitis in relapsing polychondritis: a case report. *Medicine (Baltimore)*. (2021) 100:e26315. doi: 10.1097/MD.000000000026315

149. Kirino Y, Takase-Minegishi K, Tsuchida N, Hirahara L, Kunishita Y, Yoshimi R, et al. Tocilizumab in VEXAS relapsing polychondritis: a single-center pilot study in Japan. *Ann Rheum Dis.* (2021) 80:1501–2. doi: 10.1136/annrheumdis-2021-220876

150. Meyer-Lindemann U, Mauersberger C, Schmidt AC, Moggio A, Hinterdobler J, Li X, et al. Colchicine impacts leukocyte trafficking in atherosclerosis and reduces vascular inflammation. *Front Immunol.* (2022) 13:898690. doi: 10.3389/ fimmu.2022.898690

151. Petitdemange A, Sztejkowski C, Damian L, Martin T, Mouthon L, Amoura Z, et al. Treatment of relapsing polychondritis: a systematic review. *Clin Exp Rheumatol.* (2022) 40:81–5. doi: 10.55563/clinexprheumatol/h9gq10

152. Schafer PH, Parton A, Gandhi AK, Capone L, Adams M, Wu L, et al. Apremilast, a cAMP phosphodiesterase-4 inhibitor, demonstrates anti-inflammatory activity in vitro and in a model of psoriasis. *Br J Pharmacol.* (2010) 159:842–55. doi: 10.1111/j.1476-5381.2009.00559.x

153. Hatemi G, Mahr A, Ishigatsubo Y, Song YW, Takeno M, Kim D, et al. Trial of apremilast for oral ulcers in Behçet's syndrome. *N Engl J Med.* (2019) 381:1918–28. doi: 10.1056/NEJMoa1816594

154. Summersgill H, England H, Lopez-Castejon G, Lawrence CB, Luheshi NM, Pahle J, et al. Zinc depletion regulates the processing and secretion of IL-1 $\beta$ . *Cell Death Dis.* (2014) 5:e1040. doi: 10.1038/cddis.2013.547