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Induction of silicon defences in wheat landraces is local not systemic and driven by mobilisation of soluble silicon to damaged leaves

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1 1. Highlight

Silicon is an important defence in crops. Here, a new mechanism involving the movement of
 soluble silicon in the phloem is proposed to explain localised induction of silicon defences.

4 2. Abstract

5	•	In response to herbivory, many grasses, including crops such as wheat, accumulate
6		significant levels of silicon (Si) as an antiherbivore defence. Damage-induced increases in
7		Si can be localised in damaged leaves or more systemic, but the mechanisms leading to
8		these differences in Si distribution remain untested.
9	•	Ten genetically diverse wheat landraces (Triticum aestivum) were used to assess
10		genotypic variation in Si induction in response to mechanical damage and how this was
11		affected by exogenous Si supply. Total and soluble Si levels were measured in damaged
12		and undamaged leaves, as were Si levels in the phloem, to test how Si was allocated to
13		different parts of the plant after damage.
14	•	Localised, but not systemic, induction of Si defences occurred, more pronounced when
15		plants had supplemental Si. Damaged plants had significant increases in Si concentration
16		in their damaged leaves, while the Si concentration in undamaged leaves decreased,
17		such that there was no difference in the average Si concentration of damaged and
18		undamaged plants.
19	•	The increased Si in damaged leaves was due to the redirection of soluble Si present in

The increased Si in damaged leaves was due to the redirection of soluble Si, present in
 the phloem, from undamaged to damaged plant parts, potentially a more cost-effective
 defence mechanism for plants than increased Si uptake.

Keywords: damage, genotypic variation, localised response, phloem, reallocation, silicon, wheat
 (*Triticum aestivum*)

24 **3. Introduction**

25 Plants have many different types of defences against herbivores, including both physical and 26 chemical defences (Howe and Jander, 2007). Particularly in grasses, which include many cereal 27 crops, high silicon (Si) accumulation is an effective antiherbivore defence (Massey et al., 2009; Han 28 et al., 2015; Singh et al., 2020; Waterman et al., 2021). Grasses deposit Si in structures such as 29 phytoliths and silicified spines (Hartley et al., 2015). Phytoliths increase leaf abrasiveness making 30 tissues less palatable (Hall et al., 2020), which deters both insect and mammalian herbivores. 31 Furthermore, phytoliths can reduce herbivore digestive efficiencies and hence limit their growth and 32 development (Massey and Hartley, 2006, 2009; Massey et al., 2008, 2009). In addition, Si deposited

in the apoplast may act as a physical barrier, potentially preventing the release of insect oral
secretions and oviposition fluids, known as effectors, which are used by herbivores to recognise
compatible host plants (Coskun *et al.*, 2019; Singh *et al.*, 2020).

36 At least in grasses, Si is an inducible defence (Massey et al., 2007): in response to herbivory, overall 37 Si accumulation increases, and this is correlated with reduced herbivory (Reynolds et al., 2012; 38 Hartley et al., 2015; Hall et al., 2020). However, the Si response to herbivory varies significantly 39 between both plant species and genotypes (Hartley and DeGabriel, 2016). For example, different 40 patterns of Si accumulation and deposition were found in three species of *Festuca* in response to 41 artificial damage and Si supply (Hartley et al., 2015), and similar genotypic variation in Si uptake and deposition have been found in wheat (Triticum aestivum; Thorne et al., 2021) and rice (Talukdar et 42 43 al., 2019). Soininen et al. (2013) reported both within and between species variation in Si 44 accumulation among grasses in response to damage.

45 Plants accumulate Si in the form of silicic acid from the soil using a pathway that is relatively well 46 characterised in rice. Si is transported through the roots by the serial action of two transporters, Lsi1 47 and Lsi2 (Ma and Yamaji, 2015). Silicic acid is then loaded into the xylem by the action of a third 48 transporter, Lsi3 (Huang et al., 2022). Subsequently, Si is translocated to the shoots via the transpiration stream (Ma and Yamaji, 2015). A fourth transporter, Lsi6, is required for xylem 49 50 unloading of silicic acid (Yamaji et al., 2008). Lsi6, Lsi3, and Lsi2 are also highly expressed in the node 51 where they are involved in intervascular transfer to direct Si distribution within the plant (Yamaji et 52 al., 2015). High levels of silicic acid result in its autopolymerisation into immobile silica, which is the 53 predominant form of Si in plants (Yoshida et al., 1962). Thus, plant Si accumulation is affected by 54 both the transpiration rate and the activity of Si transporters. Genotypic variation in Si accumulation 55 has been linked to both differences in transpiration (McLarnon et al., 2017) and differences in the 56 abundance of Si transporters (Ma et al., 2007b). However, a range of biotic and abiotic factors have 57 been shown to influence genotypic variation in Si accumulation including damage (Soininen et al., 58 2013), nitrogen availability and plant competition (de Tombeur et al., 2022), and climatic factors 59 (Johnson et al., 2023).

Although damage-induced localised increases in tissue Si have been reported in several grasses (Hartley *et al.*, 2015; McLarnon *et al.*, 2017) and in cucumber (Islam *et al.*, 2020), the mechanism underpinning this localised increase in Si remains to be determined. One hypothesis is that the uptake of Si from the soil increases, and this additional Si is then directed specifically to damaged tissue. It is unlikely that differences in transpiration rate explain increased Si accumulation in

- 65 damaged leaves, as a localised damage response can be observed even in plants where transpiration
- rates were greatly reduced (McLarnon *et al.*, 2017). Instead, changes in Si transporter abundance
- 67 and activity may be responsible, as has been suggested for rice (Ye *et al.*, 2013), though other
- 68 studies have not found evidence for this mechanism (McLarnon *et al.*, 2017).

A second hypothesis proposes that part or all the "extra" Si gets relocated from undamaged tissue to 69 70 damaged tissue. Si accumulation is an active process which involves significant energetic costs (de 71 Tombeur et al., 2023) and thus the mobilisation of the Si already present in the plant towards the 72 sites where it is most needed to repel attack may be a more beneficial strategy than increased 73 uptake from the soil. However, once Si is deposited, it cannot be remobilised (Samuels et al., 1991), 74 and thus this hypothesis could only pertain to the soluble Si fraction in plants. It has not yet been 75 experimentally tested, nor has the interaction between genetic variation in patterns of Si 76 accumulation, damage, and Si supply.

77 Here, the effect of mechanical damage on Si accumulation in a number of genetically diverse

- 78 landraces of wheat (*Triticum aestivum*) was investigated and the following questions were79 addressed:
- Is the induction of silicon-based defences in wheat a localised or systemic response, and how is the magnitude and pattern of induction affected by Si supply?
 If the response is localised, by what mechanism is this achieved? Is the source of the increased Si in damaged leaves due to *de novo* uptake, or to reallocation of Si, in soluble form, to damaged leaves from other parts of the plant?
- 85

4. Methods

4.1. Genotype selection, growth conditions, and experimental treatments

88 To investigate whether there is genotypic variation in the effect of damage on Si accumulation, ten 89 wheat landraces that differed significantly in their Si accumulation were selected: five high and five 90 low Si-accumulating landraces (Thorne et al., 2021, Supplementary Table S1). A balanced factorial 91 experimental design was used with three plants per landrace per damage versus undamaged 92 treatment. Two seeds were planted in 1 L pots filled with a 2:1 mix of sand and terragreen. One 93 week after germination, seedlings were thinned to one plant per pot. After thinning, all plants were 94 fed twice weekly with 200 mL half-strength Hoagland's solution supplemented with 1.8 mM 95 dissolved sodium metasilicate (Na₂SiO₃.9H₂O). Plants were watered as required.

96 Two of the high and two of the low Si-accumulating landraces were then selected to examine the 97 effect of Si availability on Si accumulation in response to damage. To control Si supply effectively, 98 plants were grown hydroponically. Seeds were germinated in sand for 10-11 days, and then 99 seedlings were transferred to 9 L plastic hydroponics boxes, filled with half-strength Hoagland's 100 solution. The pH was adjusted to 5.6-6.0 using 1 M HCl or 0.1 M KOH. The nutrient solution was 101 changed every 3-4 days. The hydroponics solutions were aerated throughout the experiment. A 102 balanced factorial experimental design was used with plants either damaged or not and 103 supplemented with Si (+Si) or not (-Si). Plants supplemented with Si received Hoagland's solution 104 containing 1.8 mM Si. Sodium chloride (NaCl) was used to balance sodium levels for plants not 105 supplemented with Si. Three plants per landrace, per Si fertilisation level, and per damage treatment 106 were used.

To identify the source of the additional Si leading to the localised induction in damaged leaves, one
landrace (L1) was selected. A balanced factorial experimental design was used with plants either
damaged or not. All plants were grown hydroponically with 1.8 mM Si for the first three weeks.
Immediately prior to damage, half the plants were moved to a medium without Si supplementation
(+/- Si) while the remaining plants continued to be grown with Si supplementation (+/+ Si). Plants
were harvested three days after the first, second, and fourth damage. Three plants per treatment
were used.

All plants were grown under controlled glasshouse conditions (16 h daylight; 20 °C /15 °C day/night).
At harvest, roots were washed in deionised water and excess water was removed. Leaf, stem, and
root fresh weight was recorded. Plants were oven-dried at 70 °C until constant mass was achieved,
then dry weight was recorded.

118 4.2. Damage Treatment

A damage treatment was started four weeks after germination and continued for three weeks. In the case of the ten landraces grown in soil, plants were damaged three times a week, whereas plants grown hydroponically were damaged twice weekly. A damage treatment involved removing approximately half of a newly produced leaf along the midrib. Plants were harvested one day after the final damage event, seven weeks after germination unless otherwise stated. Plants that were not mechanically damaged were labelled as undamaged plants. The weight and Si concentration of damaged and undamaged leaves of damaged plants were analysed separately.

126

127 4.3. Si Measurements

Portable X-ray fluorescence spectroscopy (P-XRF) was used to measure the leaf Si concentration of 128 129 all plants and the root Si concentration of plants grown hydroponically (Reidinger et al., 2012). Dried 130 leaf material was ball-milled (Retsch MM400 Mixer mill, Haan, Germany) and ground material was 131 pressed at 10 tons into pellets using a manual hydraulic press with a 13 mm die (Specac, Orpington, 132 UK). Si analysis (% Si dry weight) was performed using a commercial P-XRF instrument (Nitron 133 XL3t900 GOLDD analyser: Thermo Scientific Winchester, UK) held in a test stand (SmartStand, 134 Thermo Scientific, Winchester, UK). The P-XRF machine was calibrated using Si-spiked synthetic 135 methyl cellulose (Sigma-Aldrich, product no. 274429) and validated using Certified Reference 136 Materials of NCS DC73349 'Bush branches and leaves' obtained from China National Analysis Center 137 for Iron and Steel. To avoid signal loss by air absorption, the analyses were performed under a 138 helium atmosphere (Reidinger et al., 2012). A reading of each side of the pellet was taken, 139 approximately one hour apart, to account for u-drift in the instrument (i.e. variation in readings 140 between consecutive runs using identical parameters; Johnson, 2014). The two readings were averaged to obtain the Si concentration (%). The Si concentration of damaged and undamaged 141 142 leaves of damaged plants was analysed separately. Si content (mg) was calculated as:

143
$$Si \ content \ (mg) = \frac{Si \ concentration \ (\%)}{100} \times dry \ weight \ (mg)$$

144

4.4. Measuring soluble Si

To measure the amount of soluble Si present in the leaves, one landrace (L1) was grown hydroponically with 1.8 mM Si for seven weeks. Four leaves were damaged simultaneously (~17 % of the leaves) and plants were harvested five days after the damage. The soluble Si concentration was measured for ten damaged plants and six undamaged plants. Damaged and undamaged leaves of damaged plants were analysed separately.

150 Leaf soluble Si was measured using an adapted version of the molybdenum method of measuring 151 plant available Si in the soil (Sauer et al., 2006). Briefly, samples were oven-dried at 70 °C and ground 152 in a ball mill. A total of 2 g of sample was added to a 50 mL polyethylene tube with 20 mL 0.01 M 153 CaCl₂. Samples were shaken slowly at 30 rpm for 16 h then centrifuged at 2000 rpm for 10 min. The 154 supernatant was passed through filter paper and the resulting sample was used for Si determination by the molybdenum assay. For the assay, 1 mL sample, 30 mL 20 % acetic acid, and 10 mL 155 156 ammonium molybdate (54 g L⁻¹) were mixed in a 50 mL polyethylene tube. Samples were mixed and 157 then left to stand for 5 minutes. 5 mL 20 % (w/v) tartaric acid and 1 mL of reducing solution were

added. The reducing solution comprised 8 g L⁻¹ Na₂SO₃, 1.6 g L⁻¹ 1-amino-2-napthol-4-sulfonic acid,

- and 100 g L^{-1} NaHSO₃ dissolved in deionised water. A final volume of 50 mL was made using 20 %
- acetic acid. Samples were left at room temperature for 30 min for colour development then the
- absorbance at 810 nm was measured. A standard curve was created using 0, 0.1, 0.5, 1, 5, and 10 μ g
- 162 Si mL⁻¹ to determine the Si concentration of the samples. The amount of soluble Si measured using
- 163 the molybdenum assay was compared to the total Si concentration measured using XRF to estimate
- the total soluble Si present.

165 **4.5.** Measuring soluble Si in phloem exudate

166 Phloem exudate was collected using the ETDA-mediated method as described in Xu et al. (2019) 167 using plants grown hydroponically with 1.8 mM supplementary Si for three weeks. Three leaves per 168 plant were damaged, then half the plants were moved to medium without supplementary Si. 169 Phloem exudate was collected 24 h later using three leaves per plant. Phloem exudate was collected 170 separately for damaged and undamaged leaves of damaged plants, and for undamaged plants. As a 171 negative control, phloem exudate was collected in water (no EDTA) from undamaged plants. Three 172 plants per treatment were used. The Si concentration of the phloem exudate was then analysed 173 using the molybdenum method as described above.

174 **4.6.** Measuring expression of Si transporters

175 To investigate whether damage affected Si transporter gene expression, the expression levels of the 176 Si transporters Lsi3 and Lsi6 were determined using reverse transcriptase quantitative polymerase 177 chain reaction (RT-qPCR). These transporters were chosen as they are known to be expressed in the 178 shoots, whereas the other Si transporters, Lsi1 and Lsi2, are expressed predominantly in the roots 179 (Yamaji et al., 2015). Initial experiments on the leaves of wheat landraces confirmed this with only 180 very low levels of expression of Lsi1 and Lsi2, but significant expression of Lsi3 and Lsi6, found. Plants 181 of the landrace L1 were grown hydroponically with 1.8 mM Si for four weeks. Plants were then 182 moved to – Si medium immediately prior to the first damage event and then subject to either one or 183 two damage events (with the second damage event occurring 24 hours after the first damage 184 event). Leaf samples were taken from the middle of the newest expanded leaf 4, 24, and 48 h after one or two damages. Initially two samples were analysed for each time point and subsequently up to 185 186 five biological replicates from a given timepoint were analysed once the most relevant timepoints had been identified. 187

Primers were designed to match all homoeologs, based on existing wheat sequences where
available, or on homology to the barley sequence (Supplementary Table 2). Leaf tissue was collected

190 and ground under liquid nitrogen using a mortar and pestle and RNA extracted using a RNeasy kit 191 with DNase treatment (Qiagen), according to the manufacturer's instructions. cDNA synthesis was 192 performed using Reverse Transcriptase Superscript II M-MLV (Invitrogen) on 500 ng RNA. RT-qPCR 193 was performed using a QuantiNova SYBR green PCR kit (Qiagen) with 2 µL cDNA (diluted 1:10) and 194 700 nM primer on a Rotor-Gene Q PCR machine (Qiagen). Amplification was performed as follows: 195 95 °C for 2 min; 40 cycles of 95 °C 5 s and 60 °C for 10 s; determination of melt curve. Actin and TEF1 196 were used as reference genes. The qPCR results were analysed using a variation of the 2^{DDCt} method 197 as described in Muller et al. (2002) and an adapted version of the Q-gene excel software (Simon, 198 2003).

199 4.7. Statistical Analysis

200 All statistical analyses were performed using R software (version 4.2.0, R Core Team, 2022). 201 Summary statistics were calculated using the Rmisc package (Hope, 2013) and graphs were 202 produced using the ggplot2 package (Wickham, 2016). Two-way and three-way analyses of variance 203 (ANOVA) were used to test the effect of Si supply, damage, number of damage events, and landrace 204 on Si concentration, as relevant to each experiment. Due to the lack of independence between 205 damaged and undamaged leaves from damaged plants, ANOVAs were performed separately 206 comparing undamaged plants to either damaged or undamaged leaves of damaged plants. 207 Additionally, the average leaf Si concentration of damaged plants was calculated by averaging the Si concentration of damaged and undamaged leaves, accounting for differences in the proportion of 208 209 the leaves that were damaged or undamaged. ANOVA was then used to compare the average leaf Si 210 concentration of damaged plants to undamaged plants.

Data normality was checked using Shapiro tests and homogeneity of variance was tested using
Levene's tests. To satisfy the test assumptions, Si concentration was logit transformed and Si content
was log transformed. No transformation was applied to gene expression or soluble Si data. Paired *t*tests were used to test for localised induction of Si defences between damaged and undamaged
leaves of damaged plants. A significance level of *P* < 0.05 was used for all analyses. Significant results
were analysed by performing Tukey's Honest Significance Difference (HSD) *post-hoc* tests using the
emmeans package (Lenth, 2021).

218 **5. Results**

219 5.1. Damage results in a localised increase in Si

Averaged across all ten landraces, repeated damage significantly increased Si accumulation in
 damaged leaves of damaged plants when compared to undamaged plants (Figure 1; F_{1,40} = 67.9, *P* <

222 0.001). The biggest percentage increase was 107.6 \pm 28.4 % in the L3 landrace, compared to an 223 increase of only 34.2 \pm 9.3 % in H5. The localised induction in damaged leaves was significant in all 224 landraces except landraces H5 and L2, though there was still a trend towards significance even in 225 these landraces (*P* = 0.055 and 0.052 respectively). Overall, the interaction between landrace and 226 damage was not significant (F_{9.40} = 1.5, *P* = 0.171).

227 Si accumulation was lower in the undamaged leaves of the damaged plants, averaged across all ten 228 landraces, when compared to the levels in undamaged plants (Figure 1; $F_{1,40} = 9.2$, P = 0.004). 229 However, post-hoc testing revealed that the decrease in undamaged leaves was specific to the low 230 Si-accumulating landraces L1 (P = 0.013) and L5 (P = 0.026), with L3 and L4 showing a similar trend albeit marginally non-significant (P = 0.051 and 0.054 respectively). The lack of increase in Si in 231 232 undamaged leaves on damaged plants, or even decreases in some landraces, means there was no 233 overall increase in leaf Si in damaged plants compared to undamaged ones for any landrace 234 (Supplementary Figure 1). Thus, the induction of Si defences occurred only locally and was not

235 systemic.

On average, high Si-accumulating landraces had higher shoot Si concentrations compared to low Si accumulating landraces, with an average Si concentration of 2.97 ± 0.17 for undamaged high Si landraces compared to 1.89 ± 0.09 for low Si landraces. There was no significant correlation between leaf Si in undamaged plants and the relative increase in leaf Si due to damage (r = -0.32, P = 0.364).

5.2. The effect of damage on Si accumulation depends on Si availability

241Repeated damage significantly increased the leaf Si accumulation in the damaged leaves of the four242landraces tested (Figure 2; $F_{1,47} = 216.0$, P < 0.001). This was the case in both the +Si (1.8 mM Si243supplied to the hydroponic medium) and -Si (no exogenous Si supply) treatments, although this244increase was significantly greater in the +Si treatment (Figure 2; Si availability x damage interaction:245 $F_{1,47} = 23.1$, P < 0.001).

246

As with the ten landraces, damage resulted in only a localised induction of Si defences. Damage significantly decreased leaf Si accumulation in the undamaged leaves of damaged plants compared to undamaged plants in the +Si treatment ($F_{1,24} = 18.2$, P < 0.001), but this did not occur in the –Si plants, although leaf Si in this treatment was already an order of magnitude lower ($F_{1,23} = 0.5$, P =0.503). There was no significant variation in the Si response to damage among landraces in either the –Si or +Si treatments. No significant effect of damage on root Si accumulation was found,

although, as in the leaf tissue, there was a positive effect of Si supply on Si concentrations

- 254 (Supplementary Figure 2).
- 255

256

5.3. Si is redirected towards damaged leaves in soluble form

257 To determine whether the observed increase in Si accumulation in damaged leaves was the result of 258 de novo Si uptake or due to reallocation of Si from undamaged leaves, the exogenous Si supply was 259 removed from plants from the L1 landrace prior to a series of damage events. Increasing the number 260 of damage events significantly increased Si accumulation in damaged leaves of damaged plants, 261 although this response was significantly higher in +/+Si (continuous Si supply) plants compared to 262 +/–Si (Si removed before damage) plants (Figure 3; Si x number of damage events interaction: $F_{2,24} =$ 263 21.6, P < 0.001). For +/+Si plants, a greater number of damage events led to a more pronounced Si 264 increase in damaged leaves. However, in the case of +/–Si plants, the opposite trend occurred: each 265 damage event was associated with a reduced localised induction, possibly because the total Si concentration in these plants was "diluted" down by to continuing growth of plant tissue. Thus, after 266 four damage events, the increase in Si in damaged leaves in +/- Si plants was reduced to 81 % of the 267 268 increase occurring after a single damage, whereas for +/+ Si plants, the increase in Si after four 269 damage events was over three times as great (302 %) of that after a single damage.

270 The distribution of Si in the root, stem, and leaf tissues of damaged and undamaged plants, after 271 one, two, or four damage treatments, was calculated for both +/+Si and +/-Si plants. The total 272 amount of Si accumulated in the plants did not differ significantly between damaged and 273 undamaged plants (Figure 4; $F_{1,23} = 0.1$, P = 0.778), regardless of the number of damage events or 274 the nature of the Si supply. Si content increased with an increasing number of damage events only 275 for +/+ Si plants, as would be expected in the case of continuous Si supply. There was no significant 276 difference in root or stem Si concentration between damaged and undamaged plants, although Si 277 supply significantly increased both root and stem Si concentration (Supplementary Figure 3). For +/-278 Si plants, increasing the number of damage events significantly decreased the stem Si concentration $(F_{2,24} = 20.5, P < 0.001).$ 279

In the absence of external Si supply (+/- Si plants) and given the similarity of total Si content
between damaged and undamaged plants, any increase in Si in damaged leaves most likely
originates from the relocation of Si from undamaged leaves. The soluble Si fraction of damaged and
undamaged leaves was measured in plants of the landrace L1 grown hydroponically for 7 weeks and
damaged once, in order to assess whether it is sufficiently large to account for the extra Si found in
damaged leaves, without the need for additional uptake. Undamaged plants had a soluble Si content

- 286 of 2.2 \pm 0.1 mg g⁻¹, compared to 4.0 \pm 0.1 mg g⁻¹ for damaged leaves and 1.8 \pm 0.1 mg g⁻¹ for 287 undamaged leaves of damaged plants. Based on the difference between undamaged leaves of damaged plants and undamaged plants, this means that on average there will be 0.4 mg g⁻¹ soluble 288 Si available to move from undamaged leaves to damaged leaves of damaged plants. The soluble Si 289 from undamaged leaves needs to provide the 1.8 mg g⁻¹ increase in soluble Si observed in damaged 290 291 leaves compared to undamaged plants. Thus, as long as the amount of undamaged leaves is 4.5 292 times as the weight of damaged leaves on a plant, there is sufficient soluble Si to relocate from 293 undamaged to damaged leaves and account for all the localised induction in Si defences observed.
- 294 It was hypothesised that the redistribution of Si from undamaged to damaged leaves is the result of
- soluble Si being moved *via* the phloem. Supporting this idea, significant concentrations of soluble Si
- were measured in the phloem exudate. For +/+ Si plants, soluble Si in the phloem was found to be:
- 297 1.9 ± 0.6 mM in undamaged plants, 1.1 ± 0.5 mM in undamaged leaves of damaged plants, and $1.9 \pm$
- 298 0.4 mM in damaged leaves. The phloem Si of +/- Si plants was lower at: 1.1 ± 0.5 mM in undamaged
- plants, 0.9 ± 0.7 mM in undamaged leaves of damaged plants, and 1.0 ± 0.4 mM in damaged leaves.
- The Si in the phloem of damaged leaves was significantly higher in the +/+ Si plants than in the +/- Si
- 301 plants ($F_{1,8}$ = 9.4, P = 0.016) but there was no difference between the phloem Si levels in the
- undamaged leaves of damaged plants from the Si two treatments ($F_{1,8} = 2.0$, P = 0.196). This suggests
- that soluble Si is being directed from undamaged to damaged leaves, particularly in plants with
- 304 continuous Si supply, supporting the hypothesis that Si is being mobilised from undamaged to
- 305 damaged leaves where Si can be deposited as a physical defence.

Soluble Si may be loaded into the phloem by the action of Si transporters. Specifically, the activity of *Lsi3* and *Lsi6*, which are known to be expressed in the leaves, may be upregulated in damaged
plants. To test this hypothesis, RT-qPCR was applied to determine the expression levels of the *Lsi3*and *Lsi6* genes. However, no consistent significant differences in *Lsi3* or *Lsi6* gene expression
between damaged and undamaged plants were found, irrespective of the time point (4, 24, or 48 h)
or the number of damage events (one or two; Supplementary Figure 4).

312 **6. Discussion**

313 314

6.1. Is the induction of silicon-based defences a localised or systemic response, and how is this affected by Si supply?

Damage resulted in localised, but not systemic, induction of silicon defences. This response was
 observed across a range of landraces and at different levels of Si availability, although providing
 supplementary Si significantly increased the magnitude of the response. In contrast to the prevailing

hypothesis of Si being immediately deposited and thus rendered immobile, this study provides
evidence that soluble Si can be moved from undamaged to damaged leaves to increase Si defences
close to the site of wounding, where they are most needed. This appears to be the result of soluble
Si being transported in the phloem sap to the stem, where it is then loaded into the xylem and
transported to damaged leaves *via* the transpiration stream. However, this study found no evidence
to suggest changes in Si transporter gene expression are involved.

In this study, artificial damage was used in place of herbivory to separate the effects of damage to
tissue caused by the herbivore from the effects of molecules in the saliva and other excretions of the
herbivore (Waterman *et al.*, 2019). Damage was found to significantly increase Si accumulation, a
conclusion supported by previous studies which have found that mechanical damage is sufficient to
significantly increase plant Si-based defences (McNaughton *et al.*, 1985; Kim *et al.*, 2014; Ryalls *et al.*, 2018), though it is also clear that actual herbivory can produce greater induction (Massey *et al.*,
2007).

331 This study found significant variation in Si accumulation among wheat landraces. Eight out of ten 332 landraces responded to damage by significantly increasing Si accumulation in damaged leaves, and 333 this increase ranged from 34.2 % to 107.6 %. Similar genotypic variation has been found in other species: Bañuelos and Obeso (2000) reported significant genotypic variation in response to damage 334 335 in the grass species Agrostis tenuis. Likewise, França et al. (2019) reported genotype-specific effects 336 of Si in rice, such that Si reduced stem damage by stink bugs in only two out of three genotypes 337 investigated. Using six genotypes for each of four grass species, Soininen et al. (2013) found 338 significant genotypic variation in Si induction in response to damage in only two of the species 339 examined.

340 This study found only a localised, and not systemic, Si response to damage, with the Si concentration 341 increasing only in the damaged leaves of damaged plants. Few previous studies have made a 342 distinction between the Si concentration of damaged and undamaged leaves of damaged plants. In 343 agreement with the results of this study, insect herbivory significantly increased Si accumulation in 344 attacked leaves of two genotypes of cucumber (Islam et al., 2020) and damage resulted in localised 345 induction of Si defences in three grass species (Hartley et al., 2015). However, although damaged 346 leaves had higher Si compared to undamaged plants for all genotypes, McLarnon et al. (2017) found 347 significantly increased Si in damaged leaves compared to undamaged leaves in only one out of three 348 tall fescue genotypes investigated.

349 6.2. By what mechanism is localised induction of Si defences achieved?

Overall, the Si concentration of damaged plants was not significantly different to that of undamaged plants. It was hypothesised that this was a result of the redirection of Si into damaged leaves, with the Si content of undamaged leaves of damaged plants being significantly lower than that of undamaged leaves of undamaged plants. To test for this hypothesis, plants were grown with Si and then moved to –Si medium prior to damage. Increased Si accumulation in damaged leaves was still observed in these plants, as well as decreased Si accumulation in undamaged leaves, suggesting that Si redirection had occurred.

It has widely been reported in the literature that once deposited as silica, Si cannot be remobilised (Yoshida *et al.*, 1962; Jones and Handreck, 1967; Samuels *et al.*, 1991; Liu *et al.*, 2019; Frick *et al.*, 2020). However, it is likely that the Si was redirected into damaged leaves prior to deposition as silica, when it was still in a soluble form. Earlier studies have indicated that there may be relatively high levels of soluble Si present in the cytoplasm (Gartner *et al.*, 1984; Hodson and Evans, 1995), which could be used to increase the levels of Si in damaged leaves even after the removal of Si from the growth medium.

To examine whether there was sufficient soluble Si to explain the increased Si localised in damaged 364 365 leaves, leaf soluble Si was measured. The soluble Si concentration of undamaged plants was taken as a baseline for the amount of soluble Si predicted to be present in plants before damage. The 366 367 undamaged plants had an average total leaf Si concentration of 1.4 %, of which 15.7 % was soluble 368 Si. Thus, undamaged plants comprised approximately 0.22 % soluble Si. However, this is likely to be 369 an underestimate as the formation of silicomolybdate complexes in the molybdenum-based assay 370 depends on the size of the silicates present. While monomeric and dimeric silicates react quickly, 371 higher oligomers may not have fully reacted over the course of the 30-minute assay (Coradin et al., 372 2004). Despite this, it was calculated that there is sufficient soluble Si present in undamaged leaves 373 to explain the increase in Si in damaged leaves. Thus, it was concluded that the increase in Si in 374 damaged leaves can be explained by the movement of soluble Si from undamaged leaves into 375 damaged leaves of damaged plants (Figure 5).

It was hypothesised that the increase in Si in damaged leaves would be the result of differences in Si transporter gene expression. In rice, Si transporters are used to preferentially allocate Si to the panicle and away from the flag leaf (Yamaji and Ma, 2009; Yamaji *et al.*, 2015) and it is possible that a similar mechanism results in the preferential allocation of Si to damaged leaves. To test for this, several time points, after one and two damage events, were used to investigate whether the

381 localised increase in Si in response to damage was the result of changes in Si transporter gene 382 expression. However, no significant differences in Si transporter gene expression were found, 383 despite the numerous timepoints tested meaning that it is unlikely any increase in gene expression 384 was overlooked. Differences in Si transporter gene expression have been found in rice (Ye et al., 385 2013) but not in other species (McLarnon et al., 2017). It remains possible that post-transcriptional 386 processes affect the activity of Si transporters as gene expression is not always indicative of protein activity. Post-translational regulation has been found to be important for aquaporins (Verdoucq et 387 388 al., 2014). Alternatively, yet to be identified Si transporters may be involved, or the activity of 389 transcriptional regulators may be involved, as has been found in rice (Wang et al., 2017).

390 To the best of our knowledge, no previous studies have been carried out to directly measure Si in 391 the phloem, and it is generally suggested that Si may not be phloem-mobile (Raven, 1983). However, 392 xylem-to-phloem Si transfer has been suggested to occur during grain filling in rice, though this was 393 not actually tested (Zhou et al., 2021). Likewise, Yang et al. (2017) speculated that Si in the phloem 394 may deter aphid feeding in rice but could not rule out the possibility that their results were due to Si 395 affecting phloem sap composition. Both boron and arsenic, which share many chemical similarities 396 to Si, have been observed in the phloem (Carey et al., 2011; Zhao et al., 2012; Umemura and 397 Takenaka, 2014), although previous studies have not investigated whether this is also the case for Si. 398 In the case of boron, it must bind to sugars before being transported in the phloem (Umemura and 399 Takenaka, 2014) and it is possible that a similar mechanism is involved in Si redirection, with in vitro 400 studies supporting the idea that various forms of Si can bind to biologically relevant polymers 401 (Annenkov et al., 2017). Our study is the first to report significant levels of soluble Si present in the 402 phloem exudate.

403 The data presented here suggest a model of soluble Si being redirected from undamaged to 404 damaged leaves via the phloem. While it is noted that contamination is often an issue associated 405 with ETDA-facilitated phloem exudate collection, alternative methods including aphid stylectomy 406 would not provide sufficient yield for soluble Si analysis (Gaupels et al., 2008). Another issue with 407 EDTA-facilitated phloem exudate collection is that the amount of exudate released is unknown, 408 meaning that quantitative comparisons between samples are not possible. Nevertheless, the 409 presence of Si in the phloem exudate of +/- Si plants which have been grown without exogenous Si 410 for 24 hr strongly supports the hypothesis that Si is being transported in the phloem as the presence 411 of soluble Si in such +/- Si plants cannot be explained by *de novo* uptake from the hydroponic 412 medium. Furthermore, as localised Si induction was observed even nine days after the removal of

external Si supply, it is unlikely that there is still enough Si present in the xylem to explain theobserved induction of Si.

415 Due to the low levels of plant-available Si found in many soils, it has recently been suggested that 416 there are significant costs associated with high Si accumulation and that these are much higher than 417 previously recognised (de Tombeur et al., 2023). Negative correlations between Si accumulation and 418 biomass have been reported in several grass species, and in a number of studies (Simpson et al., 419 2017; Johnson and Hartley, 2018; de Tombeur et al., 2021), suggesting that there is a trade-off 420 between Si uptake and growth. This could reflect the fact that Si deposition is an active process 421 involving the use of active efflux transporters (Ma et al., 2007a; Ma and Yamaji, 2015), so there may be an energetic cost associated with high Si uptake (Simpson et al., 2017). Thus, redirection of 422 423 soluble Si from undamaged leaves to sites of damage might be a more energetically favourable way 424 of increasing defences against herbivores than the more costly mechanism of increasing Si uptake.

425 **6.3.** Conclusions

426 By separately measuring Si accumulation in damaged and undamaged leaves, this study has 427 demonstrated that damage results in only a localised, and not systemic, induction of Si defences. 428 This localised induction was observed in multiple landraces, although it varied between them, and 429 the response was stronger when plants were grown at high levels of Si availability. The evidence 430 presented here suggests that this localised induction is the result of the redirection of soluble Si 431 from undamaged to damaged leaves via the phloem. To the best of our knowledge, this is the first 432 demonstration of soluble Si being redirected towards the sites of wounding within damaged plants 433 to increase their defences, a mechanism which has implications for the cost-effectiveness of these 434 defences, as well as for wider understanding of the fitness benefits of Si accumulation.

- 435 **7. Supplementary data**
- 436 The following supplementary data are available at JXB online.
- 437 Table S1. List of landraces used for this study.
- 438 Table S2. List of primers used for this study.
- Table S3. ANOVA results for the effect of damage and Si supply on Si accumulation.
- 440 Fig. S1. Average leaf Si concentration for ten wheat landraces.
- 441 Fig. S2. Root Si concentration for four landraces grown hydroponically with and without Si
- 442 supplementation and subject to damage.
- 443 Fig. S3. Stem and root Si concentrations for plants grown without Si supplementation after damage.

444 Fig. S4. Si transporter gene expression after damage.

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449 **9.** Author contributions

- 450 Conceptualization, S.J.T., S.E.H. and F.J.M.M.; methodology, S.J.T., S.E.H. and F.J.M.M.; formal
- 451 analysis, S.J.T.; investigation, S.J.T.; data curation, S.J.T.; writing—original draft preparation, S.J.T.;
- 452 writing—review and editing, S.E.H. and F.J.M.M.; funding acquisition, S.E.H. and F.J.M.M. All authors
- 453 have read and agreed to the published version of the manuscript.

454 **10. Conflict of interest**

455 The authors declare no competing interests.

456 **11.Funding statement**

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459 **12. Data availability**

460 All data generated and analysed in this study are available upon request.

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14. Figures







Figure 2: Effect of Si supply and damage on Si accumulation. a) Leaf Si concentration of –Si plants. b) Leaf Si concentration of +Si plants. Note the different scales on the y-axis. Mean values \pm standard error (SE) are shown. N = 3. L4 and L5 are low Si-accumulating landrace. H1 and H3 are high Si-accumulating landraces. L indicates a low Si-accumulating landrace. ANOVA for Landrace, Si availability, Landrace x Si availability, and Damage treatment x Si availability at *P* < 0.001 for both damaged and undamaged leaves of damaged plants vs undamaged plants, see

Supplementary Table **3** for full results. Statistically significant differences between damaged and undamaged leaves of damaged plants, determined by paired *t*-tests, are indicated: *** P < 0.001, ** P < 0.01, and * P < 0.05.

a) Si removed prior to damage (+/- Si)



Figure 3: Effect of reducing Si availability on Si accumulation after damage. Leaf Si concentration for damaged and undamaged leaves of damaged plants, and undamaged plants. a) Plants moved to medium without Si supplementation when damage was started (+/–Si plants). b) Plants grown with continuous Si supplementation (+/+ Si plants). The low Si-accumulating landrace, L1, was used. Mean values \pm SE are shown. N = 3. ANOVA for Si availability, Damage treatment, and Number of damage events x Si availability at *P* < 0.01 for both damaged and undamaged leaves of damaged plants vs undamaged plants, see

Supplementary Table **3** for full results. Statistically significant differences between damaged and undamaged leaves of damaged plants, determined by paired *t*-tests, are indicated: *** P < 0.001, ** P < 0.01, and * P < 0.05.



Figure 4: Total leaf Si content was unaffected by damage. Allocation of total Si accumulated to different plant tissues in damaged and undamaged plants after successive damage events. a) +/– Si plants sampled after 1, 2, and 4 damage events. b) +/+ Si plants after 1, 2, and 4 damage. Mean values \pm SE are shown. N = 3. Note different scales of y-axis. Significant ANOVA effects: Number of damage events: $F_{2,23} = 36.0$, P < 0.001, Si availability $F_{1,23} = 13.8$, P = 0.001, Number of damage events x Si availability $F_{2,23} = 8.1$, P = 0.002.



Figure 5: The localised increase in Si in damaged leaves can be explained by the redirection of soluble Si from undamaged leaves of damaged plants. An undamaged plant weighing 1 g DW contains a total of 14 mg Si g⁻¹ leaf on average, of which 2.2 mg g⁻¹ is soluble. After damage, soluble Si is moved from undamaged leaves to damaged leaves. This movement of soluble Si increases the total Si concentration in damaged leaves to 17 mg Si g⁻¹, of which 4 mg g⁻¹ is soluble. Provided the weight of undamaged leaves is at least 4.5 times more than the weight of damaged leaves, the increase in Si in damaged leaves can be explained by the redirection of soluble Si from undamaged to damaged plants. This was the case for all the experiments presented here. *Figure created using BioRender (https://www.biorender.com)*.

Supplementary Information

Supplementary Table 1: List of landraces used in this study. H1-H5 are high Si-accumulating landraces while L1-L5 are low Si-accumulating landraces, as characterised in Thorne *et al.* (2021).

Landraca	Collection	Diant ID	Blant Name	Origin
Lanurace	Conection			Ongin
H1	CIMMYT	CWI 2166	K7155.41	Kenya
H2	Watkins	1190195	Gahu (Nepali) or Kyo (Sikkimese)	India
H3	CIMMYT	CWI 3909	OUBAARD	South Africa
H4	Watkins	1190777	Finland 3	Finland
H5	Prague	01C0201531	Orchon	Mongolia
L1	Watkins	1190521	Dandi	India
L2	Watkins	1190568	China 19	China
L3	Watkins	1190605	Karabash	Greece
L4	Watkins	1190662	Samanta 1252	Romania
L5	Watkins	1190751	Armavir	USSR

Supplementary Table 2: List of primers used for RT-qPCR.

Target Gene	Forward Primer	Reverse Primer	Product size	Ensembl gene name (all homologues)
Actin	AATGGTCAAGGCTGGTTTCG	ATCACCGACATAGGCATCCTTC	124	TraesCS1A02G020500, TraesCS1B02G024500, TraesCS1D02G020000
Tef1	TTGGTGGCATTGGAACTGTG	TTGACCTCAGTTGTCAGACCAG	103	TraesCS5B02G418200 TraesCS5D02G423400
Lsi3	TGTTCAAGTACCTCGGCAAC	TTGAGGATGAACTCGGTGAGG	144	TraesCS4A02G412500, TraesCS4B02G312600, TraesCS4D02G310100
Lsi6	TACTCGAACGAGATCCACGAC	TCTCCGATATCACCTTCTTGCC	132	TraesCS7A02G187800, TraesCS7B02G092900, TraesCS7D02G188800

Supplementary Table 3: ANOVA results for Figures 2 and 3. Statistically significant results are highlighted in bold. N = 3 for each factor combination.

a) The effect of landrace, damage, and Si supply on Si concentration in damaged and undamaged leaves of damaged plants compared to undamaged plants.

	Leaf Si (%)							Root Si (%)		
	Undamaged leaves			Damaged leaves						
	df	F	Ρ	df	F	Ρ	df	F	Ρ	
Landrace	3	22.39	< 0.001	3	24.81	< 0.001	3	1.71	0.179	
Damage	1	0.09	0.764	1	215.99	< 0.001	1	0.79	0.378	
Si	1	6188.77	< 0.001	1	9076.47	< 0.001	1	88.54	< 0.001	
Landrace x Damage	3	0.20	0.897	3	0.54	0.66	3	0.71	0.550	
Landrace x Si	3	23.73	< 0.001	3	27.41	< 0.001	3	1.19	0.326	
Damage x Si	1	12.32	< 0.001	1	23.10	< 0.001	1	0.16	0.691	
Landrace x Damage x Si	3	0.16	0.923	3	1.39	0.26	3	0.86	0.466	

b) The effect of damage, Si supply, and number of damage events on total plant Si content and leaf Si concentration in damaged and undamaged leaves of damaged plants compared to undamaged plants.

	Leaf Si (%)						Plant Si content (mg)		
	Undamaged leaves			Damaged leaves					
	df	F	Ρ	df	F	Р	df	F	Р
Number of damage events	2	10.30	< 0.001	2	1.75	0.196	2	36.01	< 0.001
Damage	1	42.81	< 0.001	1	84.23	< 0.001	1	0.08	0.778
Si	1	95.99	< 0.001	1	108.24	< 0.001	1	13.80	0.001
Number x Damage	2	2.62	0.094	2	1.34	0.281	2	0.37	0.698
Number x Si	2	6.79	0.005	2	21.64	< 0.001	2	8.11	0.002
Damage x Si	1	0.15	0.701	1	0.07	0.800	1	0.16	0.694
Number x Damage x Si	2	1.21	0.316	2	1.87	0.176	2	0.35	0.709



Supplementary Figure 1: Variation in leaf Si accumulation between damaged and undamaged plants for ten wheat landraces. The average leaf Si concentration for damaged and undamaged leaves of damaged plants was calculated. L1-L5 are low Si-accumulating landraces; H1-H5 are high Si-accumulating landraces. Mean values \pm SE are shown. N = 3. Statistically significant ANOVA effect: Landrace: F_{9,40} = 18.7, *P* < 0.001.



Supplementary Figure 2: Effect of damage and Si supply on root Si accumulation. (A) Root Si of –Si plants. (B) Root Si of +Si plants. L4 and L5 are low Si-accumulating landraces; H1 and H3 are high Si-accumulating landraces. Mean values \pm standard error (SE) are shown. N = 3. Statistically significant ANOVA effect: Si: $F_{1,47}$ = 88.5, P < 0.001.



Supplementary Figure 3: Effect of reducing Si availability on Si accumulation after damage. (A) Stem Si concentration for plants moved to medium without Si supplementation when damage was started (+/– Si plants). (B) Stem Si concentration for plants grown continuously with Si supplementation (+/+ Si plants). (C) Root Si concentration for +/– Si plants. (D) Root Si concentration for +/+ Si plants. Mean values ± SE are shown. N = 3. The low Si accumulating landrace, L1, was used. Significant ANOVA effects for stem Si: Si availability: $F_{1,24} = 138.5$, P < 0.001; Number of damage events: $F_{2,24} = 12.2$, P < 0.001; Number of damage events x Si availability: $F_{2,24} = 20.5$, P < 0.001; for root Si: Si availability: $F_{1,23} = 12.7$, P = 0.002; Number of damage events: $F_{2,23} = 5.2$, P = 0.013.



Supplementary Figure 4: Si transporter gene expression at different time points. (A) *Lsi3* expression. (B) *Lsi6* expression. The low Si-accumulating landrace, L1, was used. Mean values ± SE are shown. N = 2-5. Dam: number of damage events. No significant differences between damaged and undamaged plants, and between damaged and undamaged leaves of damaged plants, were found.