

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,400

Open access books available

174,000

International authors and editors

190M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Chapter

Introductory Chapter: The Significance of *Toxoplasma gondii* in Humans, and in Domestic and Wild Animals

*Saeed El-Ashram, Gungor Cagdas Dincel,
Mohammed Hamdy Farouk, Abdulaziz Alouffi,
Hany M.R. Abdel-Latif, Shagufta Iqbal, Guillermo Tellez-Isaias,
Billy M. Hargis, Víctor Manuel Petrone-García,
Gabriela Aguilar-Tipacamú and Inkar Castellanos-Huerta*

1. Introduction

1.1 *Toxoplasma gondii* and life cycle

Since 1908, when *Toxoplasma gondii* was isolated in a rodent (*Ctenodactylus gundi*), from which the parasite gained its name [1], more than a century has passed. Because of the crescent-shaped form of the parasite, the genus name was taken from the Greek word toxin, which means bow. It was not until 1970 that the entire life cycle of the parasite was revealed, considering cat (*Felis catus*) the only definitive host (DH) in which the parasite could complete its entire sexual life cycle [2]. *T. gondii* belongs to the phylum *Ampicomplexa*, which contains intercellular parasites with a polarized cell structure and an apical complex [3]. *T. gondii* life cycle is considered sexual and asexual with a characteristic complexity; it reproduces sexually in the DH and asexually in the intermediate hosts (IH) [4] with three distinct stages of reproduction: tachyzoites (replicating fast), bradyzoites (replicating slowly), and sporozoites (in sporulated oocysts). As described briefly, after consuming any of the three stages, wild and domestic cats are infected [5]; prepatent period typically lasts between 3 and 10 days until 19 days or more [6]. Digestive enzymes break down the cyst wall, releasing the bradyzoites [7] and allowing the infection of intestinal epithelial cells through asexual reproduction (schizonts). Merozoites are discharged and reinvade the intestinal epithelium when the schizont reaches maturity. After this stage, sexual differentiation is observed [5, 8], rising to flagellated microgametes and macrogametes and beginning fertilization [6], resulting in the production of a zygote (diploid) and growing into an unsporulated oocyst, which is discharged into the intestinal lumen and expelled from the DH. Sporulation of the oocysts takes from 1 to 5 days, depending on environmental variables such as temperature and humidity [9, 10]. Oocysts

have impermeable walls and are resistant to physical and chemical stimuli, living in damp soil and staying infectious for months [11]. The asexual cycle of *T. gondii* begins when an IH consumes contaminated water or food. In the gut, the sporozoites are released and infect the intestinal epithelial cells [5]. The IH may also gain *Toxoplasma* infection by consuming raw or undercooked meat, harboring bradyzoite-bearing tissue cysts. The freshly released bradyzoites convert into tachyzoites after entering the intestinal epithelium after the rupture of the cyst wall [12]. After successful cell invasion, intracellular proliferation takes place until the host cell can no longer sustain its development [13]. Eventually, the tachyzoites egress from the host cell and disperse through the circulatory system to organs such as the liver, lungs, brain, eye, placenta, and heart, completing the asexual life cycle [14]. Once the host's immunological response is elicited, the multiplying tachyzoites develop into bradyzoites, form tissue cysts, and stay dormant in the host [15, 16]. In healthy individuals, tissue cysts may survive throughout life without eliciting an immune response from the host. However, when the immune system is compromised or weakened, bradyzoites may revert quickly to replicating and invasive tachyzoites, leading to infection [13].

1.2 *Toxoplasma* genotypes

T. gondii genome is composed of a 65 Mb nuclear genome with 14 chromosomes [17]. The genome of *T. gondii* is bigger than that of other *apicomplexans* [18], which might be attributed to the large number of IHs of *T. gondii* [19]. Online database (ToxoDB (<http://toxodb.org/toxo/>)) contains complete information on the genome of *T. gondii* [20], including the three main types of *T. gondii*: GT-1 (type I), Me49 (type II), and VEG (type III) [21, 22]. Genotypes vary substantially across different geographic locations; Asia, Europe, and North and South Americas present type I, and all the continents present types II and III [23]. Thus, diversity in pathogenicity and virulence is directly correlated with genetic differences [24]. A median lethal dose (LD₅₀) to mice in laboratory settings is a common way to describe virulence strain. LD₅₀, for as little as one parasite, in type I strain of the parasite may kill mice. Types II and III, in contrast, are less virulent and only result in LD₅₀ following infection with several parasites. Multi-locus restriction fragment length polymorphism (RFLP) is used to detect strains with distinct genotypes compared to types I, II, and III, which were first found in isolated populations of exotic animals, but they have lately been reported in chickens, pigs, and sheep [25]. Human primary infections of ocular illness are associated with atypical toxoplasmosis strains [26]. These atypical strains are characterized primarily by unique alleles and allele-shuffled combinations from the three major types [27]. Microsatellite genotyping of immunocompromised patients with toxoplasmosis revealed three novel genotypes, Africa I, Africa II, and Caribbean I, and subsequently determined a different Africa III genotype [28, 29]. The impact of secreted *Toxoplasma* virulence effectors and genetic diversity of *T. gondii* vary in encystment rates and acute pathogenicity in mice [30]. *T. gondii* strains grouped lineages (types I, II, and III) are based on variations in acute virulence in mice and genetic markers, RFLPs, and single-nucleotide polymorphisms (SNPs) [31, 32]. Highly pathogenic type I strain presents a LD₁₀₀ of 1 [31], in contrast to strain type II with a LD₅₀ of 10³–10⁴, and type III strain with a LD₅₀ of 10⁴–10⁵ [33]. Advances in sequencing capabilities resulted in a broader diversity of *T. gondii* strains, giving rise to a new classification scheme, which separates strains into 15 genetic haplotypes, with types I, II, and III being referred to as haplotype 1, 2, and 3 and clade A, D, and C, respectively [33]. Strain-specific alleles of two proteins, ROP5 and ROP18, were

found to explain the differential in virulence [34, 35]. The virulent alleles of these proteins operate together to inhibit families of interferon-inducible gene families [35, 36].

1.3 Cats' susceptibility to *Toxoplasma* infection: determinants

Even though DH presents more than 180 parasite species [37], *T. gondii* appears more often than others in literature because of its effects on the health of humans, livestock, and wildlife [38]. Interactions between the three essential “ecological components,” (a) DH, (b) environment (Env), and (c) IH influence the sustainability of these parasites, resulting in the fecal-oral route as the most common mode of transmission. A rise in parasite shedding and environmental contamination should be positively associated with the number of DHs. The likelihood of oocysts contributing to parasite transmission may also be influenced by specific host behaviors, and by physically separating the DH and IH niches, it may be possible to reduce parasite transmission. Env conditions may impact *Toxoplasma* oocyst survival, while the behavior of IH (such as grazing) may affect transmission [39]. Oocysts have the potential to survive in the environment at 4°C or room temperature for 6 months, resulting in oocysts sporulating and living for an extended period of time [40]. Oocysts are resistant to freezing [41] but are destroyed by prolonged desiccation [42], high temperatures (60°C), and exposure to UV light [43]. Therefore, any Env parameters, DH, and IH influence the completion of *T. gondii* life cycle. Since cats only shed it once, seropositive cats no longer shed *T. gondii* after their first infection; the presence of IHs encourages the transmission of parasites throughout an environment if the IHs provide a scarce source of food for the DH.

1.4 *T. gondii* in humans and animals

Humans, as IH, present a third or more people who have been exposed to infection with *T. gondii*. The prevalence ranges widely (10–85%) between nations, geographical areas within a country [7], socioeconomic conditions, climate (warm, humid tropical areas, cold-temperate, or desert regions), poor sanitation, a lack of clean water, and poor hygiene [44]. Toxoplasmosis is classified into an acute acquired infection (horizontal transmission) and a congenital infection (CT) [45] with an incubation period from 10 to 23 days after consuming tissue cysts and from 5 to 10 days following the consumption of oocysts [10, 46]. CT to the fetus (transplacental) may occur if the mother gets a primary infection or the chronically infected mother becomes immunocompromised during pregnancy [13]. Also, blood transfusion, solid organ transplant, or bone marrow transplant from a seropositive donor might be considered for infection [47]. Sexual transmission is established in many animal species, and circumstantial evidence implies that it may also occur in humans [48–50]. Because most infections are subclinical or misdiagnosed, it is challenging to establish the human toxoplasmosis and welfare effects [42]. Moderate flu-like symptoms, such as fever, muscle discomfort, and fatigue, may occur in some people [51]. Cases of severe toxoplasmosis and ocular disease are documented [52]. The most common cause is thought to be consuming raw or undercooked meat from an infected animal [53]. Unpasteurized milk; contaminated meat, vegetables, and fruits; and contaminated water might be a source of infection [54]. Infection may be caused by consuming water or food contaminated with feline excrement [55]. Disease transmission can be prevented by adequately boiling or freezing; temperatures over 67°C or below –12°C render tissue cysts nonviable [56, 57], and salted and cured meat often weakens the

viability. However, *Toxoplasma* tissue cysts seem refractory to these approaches [58]. The efficiency of thermal, nonthermal, and chemical/biochemical treatments for inactivating *T. gondii* in foods destined for human consumption has been reviewed [59]. When these individuals have a healthy immune system, the infection may not cause any symptoms. However, if the immune system is suppressed, the parasite can reactivate and cause potentially fatal complications [60]. Clinical signs of the disease include encephalitis, schizophrenia, bipolar disorders, depression, obsessive-compulsive disorders, retinochoroiditis, myocarditis, and fetal abnormalities following transplacental infection in immunocompromised people [61]. Both immunocompetent and immunocompromised pregnant women are at risk of transmitting *T. gondii* to their unborn children if they become infected with the parasite during their pregnancies [62]. The prevalence of CT varies by nation and location. Pregnancy-related fever, eating unwashed vegetables, and diagnosis in the third trimester were all linked to CT [63]. It is believed that the high genetic diversity of *T. gondii* seen in animals from this country correlates with illness severity. Infections after birth are usually undetectable, and toxoplasmosis predominantly affects immunocompromised patients where the clinical course of the illness is frequently considerably more severe [51]. Infected newborns might have nonspecific symptoms [64]. However, most infections are asymptomatic because the organism has developed several strategies to prevent significant inflammatory and immune responses. In the case of infections in wildlife populations, *T. gondii* infects any warm-blooded wildlife by ingesting environmental oocysts or, in omnivores and carnivores, both oocysts and tissue cysts [65, 66]. Production species are also affected by *T. gondii*, such as poultry [67]. Like chickens, *T. gondii* is the IH in livestock [68, 69], sheep [70, 71], and swine [72]. During infection of IH, first contact is made when parasites infiltrate enterocytes or cross the epithelial barrier to infect resident macrophages, dendritic cells (DCs), and lymphocytes [73] and rapidly multiply, infecting any cell with a nucleus; the resulting cell lysis may trigger rapid disease onset. A solid first line of defense is encouraged by the innate immune response. Within a few days of infection, the body develops an adaptive immune response that limits the infection and strengthens resistance to the disease. In response, the parasite undergoes a transition into the bradyzoite, a latent stage that develops inside the parasite cysts within the host. The immune system cannot combat these tissue cysts [16]. The complex relationship between *T. gondii* and innate immune system cells has been the subject of several researches and serves as a model for various microbial illnesses [74].

1.5 Conclusion

Since the first isolation over a century ago, *Toxoplasma gondii* has been extensively studied to understand its biology, transmission, life cycle, and implications for the health of its definitive hosts and intermediate hosts, including the human being. The life cycle of this protozoan of the phylum Apicomplexa presents different stages (sexual and asexual), which, in combination with a capacity for resistance to the environment and multiple hosts, makes this organism highly adaptable and easy to transmit and spread. Sequencing of *T. gondii* genome allowed the development of databases applied to the study, diagnosis, and typification, as well as the analysis of pathogenic factors associated with the capacity. Particularly, *T. gondii* is mainly mentioned in the literature due to its potential effect on various species, including humans. The DH-environment-IH interactions form the essential ecological components for its dissemination, mainly by fecal material/oral cycle. Human infection is

highly impacted worldwide since it is presumed that more than a third of the human population has been infected by this protozoan. Due to the immune system being actively involved in controlling infection, clinical signs range from a simple cold to severe illness in the case of an immunodeficient situation (eye lesions, lymphopathy, embryonic damage, and mental disorders). Its study demonstrated its presence in many foods of animal and vegetable origin, placing this infection as one of the main risks of food-borne diseases. Its presence in wildlife is also an intrinsically related factor in its worldwide distribution. Due to this, *T. gondii* represents a high risk for humans and a great variety of IHs that can be affected by this parasite; therefore, the study is highly relevant for its control and research worldwide.

IntechOpen

Author details

Saeed El-Ashram^{1,2*}, Gungor Cagdas Dincel³, Mohammed Hamdy Farouk⁴, Abdulaziz Alouffi⁵, Hany M.R. Abdel-Latif⁶, Shagufta Iqbal⁷, Guillermo Tellez-Isaias⁸, Billy M. Hargis⁹, Víctor Manuel Petrone-García¹⁰, Gabriela Aguilar-Tipacamú¹¹ and Inkar Castellanos-Huerta¹²

1 Faculty of Science, Kafrelsheikh University, Kafr El-Sheikh, Egypt

2 College of Life Science and Engineering, Foshan University, Foshan, China

3 Eskil Vocational School, Laboratory and Veterinary Science, Aksaray University, Aksaray, Turkey

4 Animal Production Department, Faculty of Agriculture, Al-Azhar University, Nasr City, Cairo, Egypt

5 King Abdulaziz City for Science and Technology, Riyadh, Saudi Arabia

6 Department of Poultry and Fish Diseases, Faculty of Veterinary Medicine, Alexandria University, Alexandria, Egypt

7 Department of Zoology, University of Kashmir, India

8 Department of Poultry Science, University of Arkansas, Fayetteville, USA

9 Department of Poultry Science, University of Arkansas Agricultural Experiment Station, Fayetteville, United States


10 Petrone-García Departamento de Ciencias Pecuarias, Facultad de Estudios Superiores Cuautitlán UNAM, Cuautitlán, Mexico

11 Licenciatura en Medicina Veterinaria y Zootecnia, Facultad de Ciencias Naturales, Universidad Autónoma de Querétaro, Querétaro, México

12 Programa de Maestría y Doctorado en Ciencias de la Producción y de la Salud Animal, Facultad de Medicina Veterinaria y Zootecnia, Universidad Nacional Autónoma de México, Ciudad Universitaria, Ciudad de Mexico, Mexico

*Address all correspondence to: saeed_elashram@yahoo.com

IntechOpen

© 2023 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. 

References

- [1] Dubey J. History of the discovery of the life cycle of *Toxoplasma gondii*. *International Journal for Parasitology*. 2009;**39**:877-882
- [2] Sharifpour MF. Artificial regulation of transcription in *Toxoplasma gondii*, the model apicomplexan [PhD thesis]. 2021
- [3] Dubey J. Advances in the life cycle of *Toxoplasma gondii*. *International Journal for Parasitology*. 1998;**28**:1019-1024
- [4] Weiss LM. *Toxoplasma Gondii: The Model Apicomplexan-Perspectives and Methods*. 2nd ed. Amsterdam, Netherlands: Elsevier's Science and Technology; 2013
- [5] Yan C, Liang L-J, Zheng K-Y, Zhu X-Q. Impact of environmental factors on the emergence, transmission and distribution of *Toxoplasma gondii*. *Parasites & Vectors*. 2016;**9**:1-7
- [6] Dumètre A, Dardé M-L. How to detect *Toxoplasma gondii* oocysts in environmental samples? *FEMS Microbiology Reviews*. 2003;**27**:651-661
- [7] Robert-Gangneux F, Dardé M-L. Epidemiology of and diagnostic strategies for toxoplasmosis. *Clinical Microbiology Reviews*. 2012;**25**:264-296
- [8] Ferguson DJ, Dubremetz J-F. The ultrastructure of *Toxoplasma gondii*. *Toxoplasma Gondii*. 2014:19-59
- [9] Kieffer F, Wallon M. Congenital toxoplasmosis. *Handbook of Clinical Neurology*. 2013;**112**:1099-1101
- [10] Hampton MM. Congenital toxoplasmosis: A review. *Neonatal Network*. 2015;**34**:274-278
- [11] Simon A, Poulin MB, Rousseau AN, Ogden NH. Fate and transport of *Toxoplasma gondii* oocysts in seasonally snow covered watersheds: A conceptual framework from a melting snowpack to the Canadian arctic coasts. *International Journal of Environmental Research and Public Health*. 2013;**10**:994-1005
- [12] Muñoz-Zanzi CA, Fry P, Lesina B, Hill D. *Toxoplasma gondii* oocyst-specific antibodies and source of infection. *Emerging Infectious Diseases*. 2010;**16**:1591
- [13] Harker K, Ueno N, Lodoen M. *Toxoplasma gondii* dissemination: A parasite's journey through the infected host. *Parasite Immunology*. 2015;**37**:141-149
- [14] Randall L, Hunter C. Parasite dissemination and the pathogenesis of toxoplasmosis. *European Journal of Microbiology & Immunology*. 2011;**1**:3-9
- [15] Tomita T, Bzik DJ, Ma YF, Fox BA, Markillie LM, Taylor RC, et al. The *Toxoplasma gondii* cyst wall protein CST1 is critical for cyst wall integrity and promotes bradyzoite persistence. *PLoS Pathogens*. 2013;**9**:e1003823
- [16] Sullivan WJ Jr, Jeffers V. Mechanisms of *Toxoplasma gondii* persistence and latency. *FEMS Microbiology Reviews*. 2012;**36**:717-733
- [17] Walochnik J, Duchêne M. *Molecular Parasitology: Protozoan Parasites and Their Molecules*. Heidelberg Platz 3, Berlin: Springer; 2016
- [18] Khan A, Taylor S, Su C, Sibley LD, Paulsen I, Ajioka JW. Genetics and genome organization of *Toxoplasma gondii*. In: *Toxoplasma: Molecular and Cellular Biology*. Norfolk, England: Horizon Bioscience; 2007. pp. 193-207

- [19] Roos DS. Themes and variations in apicomplexan parasite biology. *Science*. 2005;**309**:72-73
- [20] Gajria B, Bahl A, Brestelli J, Dommer J, Fischer S, Gao X, et al. ToxoDB: An integrated toxoplasma gondii database resource. *Nucleic Acids Research*. 2007;**36**:D553-D556
- [21] Sibley LD, Boothroyd JC. Virulent strains of toxoplasma gondii comprise a single clonal lineage. *Nature*. 1992;**359**:82-85
- [22] Ajioka JW, Boothroyd JC, Brunk BP, Hehl A, Hillier L, Manger ID, et al. Gene discovery by EST sequencing in toxoplasma gondii reveals sequences restricted to the apicomplexa. *Genome Research*. 1998;**8**:18-28
- [23] Galal L, Ariey F, Gouilh MA, Dardé M-L, Hamidović A, Letourneur F, et al. A unique toxoplasma gondii haplotype accompanied the global expansion of cats. *Nature Communications*. 2022;**13**:5778
- [24] Saeij JP, Boyle JP, Boothroyd JC. Differences among the three major strains of toxoplasma gondii and their specific interactions with the infected host. *Trends in Parasitology*. 2005;**21**:476-481
- [25] Frazão-Teixeira E, Sundar N, Dubey J, Grigg M, De Oliveira F. Multi-locus DNA sequencing of toxoplasma gondii isolated from Brazilian pigs identifies genetically divergent strains. *Veterinary Parasitology*. 2011;**175**:33-39
- [26] Vaudaux JD, Muccioli C, James ER, Silveira C, Magargal SL, Jung C, et al. Identification of an atypical strain of toxoplasma gondii as the cause of a waterborne outbreak of toxoplasmosis in Santa Isabel do Ivaí, Brazil. *The Journal of Infectious Diseases*. 2010;**202**:1226-1233
- [27] Grigg ME, Suzuki Y. Sexual recombination and clonal evolution of virulence in toxoplasma. *Microbes and Infection*. 2003;**5**:685-690
- [28] Ajzenberg D, Yera H, Marty P, Paris L, Dalle F, Menotti J, et al. Genotype of 88 toxoplasma gondii isolates associated with toxoplasmosis in immunocompromised patients and correlation with clinical findings. *The Journal of Infectious Diseases*. 2009;**199**:1155-1167
- [29] Mercier A, Devillard S, Ngoubangoye B, Bonnabau H, Bañuls A-L, Durand P, et al. Additional haplogroups of toxoplasma gondii out of Africa: Population structure and mouse-virulence of strains from Gabon. *PLoS Neglected Tropical Diseases*. 2010;**4**:e876
- [30] Suzuki Y, Orellana MA, Schreiber RD, Remington JS. Interferon- γ : The major mediator of resistance against toxoplasma gondii. *Science*. 1988;**240**:516-518
- [31] Howe DK, Sibley LD. Toxoplasma gondii comprises three clonal lineages: Correlation of parasite genotype with human disease. *The Journal of Infectious Diseases*. 1995;**172**:1561-1566
- [32] Su C, Khan A, Zhou P, Majumdar D, Ajzenberg D, Dardé M-L, et al. Globally diverse toxoplasma gondii isolates comprise six major clades originating from a small number of distinct ancestral lineages. *Proceedings of the National Academy of Sciences*. 2012;**109**:5844-5849
- [33] Silva NM, Rodrigues CV, Santoro MM, Reis LF, Alvarez-Leite JI, Gazzinelli RT. Expression of indoleamine 2, 3-dioxygenase, tryptophan

- degradation, and kynurenine formation during in vivo infection with *Toxoplasma gondii*: Induction by endogenous gamma interferon and requirement of interferon regulatory factor 1. *Infection and Immunity*. 2002;**70**:859-868
- [34] Niedelman W, Gold DA, Rosowski EE, Sprockholt JK, Lim D, Farid Arenas A, et al. The rhoptry proteins ROP18 and ROP5 mediate *Toxoplasma gondii* evasion of the murine, but not the human, interferon-gamma response. *PLoS Pathogens*. 2012;**8**:e1002784
- [35] Etheridge RD, Alagunan A, Tang K, Lou HJ, Turk BE, Sibley LD. The *Toxoplasma* pseudokinase ROP5 forms complexes with ROP18 and ROP17 kinases that synergize to control acute virulence in mice. *Cell Host & Microbe*. 2014;**15**:537-550
- [36] Steinfeldt T, Könen-Waisman S, Tong L, Pawlowski N, Lamkemeyer T, Sibley LD, et al. Phosphorylation of mouse immunity-related GTPase (IRG) resistance proteins is an evasion strategy for virulent *Toxoplasma gondii*. *PLoS Biology*. 2010;**8**:e1000576
- [37] Taylor MA, Coop RL, Wall RL. *Veterinary Parasitology*. New York, NY: John Wiley & Sons; 2015
- [38] Doherty TS, Dickman CR, Johnson CN, Legge SM, Ritchie EG, Woinarski JC. Impacts and management of feral cats *felis catus* in Australia. *Mammal Review*. 2017;**47**:83-97
- [39] Frenkel J, Ruiz A, Chinchilla M. Soil survival of *Toxoplasma* oocysts in Kansas and Costa Rica. *The American Journal of Tropical Medicine and Hygiene*. 1975;**24**:439-443
- [40] Lindsay DS, Collins MV, Mitchell SM, Cole RA, Flick GJ, Wetch CN, et al. Sporulation and survival of *Toxoplasma gondii* oocysts in seawater. *The Journal of Eukaryotic Microbiology*. 2003;**50**:687-688
- [41] Frenkel J, Dubey J, et al. Effects of freezing on the viability of *Toxoplasma* oocysts. *The Journal of Parasitology*. 1973;**59**:587-588
- [42] Dubey JP. *Toxoplasmosis of Animals and Humans*. Boca Raton, Florida, USA: CRC Press; 2016
- [43] Dumètre A, Le Bras C, Baffet M, Meneceur P, Dubey J, Derouin F, et al. Effects of ozone and ultraviolet radiation treatments on the infectivity of *Toxoplasma gondii* oocysts. *Veterinary Parasitology*. 2008;**153**:209-213
- [44] Flegr J, Prandota J, Sovičková M, Israili ZH. Toxoplasmosis—A global threat. Correlation of latent toxoplasmosis with specific disease burden in a set of 88 countries. *PLoS One*. 2014;**9**:e90203
- [45] Hussain MA, Stitt V, Szabo EA, Nelan B. *Toxoplasma gondii* in the food supply. *Pathogens*. 2017;**6**:21
- [46] Dubey J, Murata F, Cerqueira-Cézar C, Kwok O, Villena I. Congenital toxoplasmosis in humans: An update of worldwide rate of congenital infections. *Parasitology*. 2021;**148**:1406-1416
- [47] Singh S. Congenital toxoplasmosis: Clinical features, outcomes, treatment, and prevention. *Tropical Parasitology*. 2016;**6**:113
- [48] Liu S, Qin C, Yao Z, Wang D. Study on the transmission of *Toxoplasma gondii* by semen in rabbits. *Zhongguo Ji Sheng Chong Xue Yu Ji Sheng Chong Bing Za Zhi*. 2006;**24**:166-170
- [49] Hlaváčová J, Flegr J, Řežábek K, Calda P, Kaňková Š. Male-to-female

- presumed transmission of toxoplasmosis between sexual partners. *American Journal of Epidemiology*. 2021;**190**:386-392
- [50] Arantes TP, Lopes WDZ, Ferreira RM, Pieroni JSP, Pinto VM, Sakamoto CA, et al. *Toxoplasma gondii*: Evidence for the transmission by semen in dogs. *Experimental Parasitology*. 2009;**123**:190-194
- [51] Jg M, Liesenfeld O. *Toxoplasmosis*. *Lancet*. 2004;**363**:1965-1976
- [52] Demar M, Hommel D, Djossou F, Peneau C, Boukhari R, Louvel D, et al. Acute toxoplasmosis in immunocompetent patients hospitalized in an intensive care unit in French Guiana. *Clinical Microbiology and Infection*. 2012;**18**:E221-E231
- [53] Sobanski V, Ajzenberg D, Delhaes L, Bautin N, Just N. Severe toxoplasmosis in immunocompetent hosts: Be aware of atypical strains. *American Journal of Respiratory and Critical Care Medicine*. 2013;**187**:1143-1145
- [54] Hill DE, Dubey JP. *Toxoplasma Gondii as a Parasite in Food: Analysis and Control*. *Microbiol Spectrum* 4: PFS-0011-2015. DOI: 10.1128/Microbiolspec.PFS-0011-2015. 2016
- [55] Organization, WH. *Foodborne Disease Outbreaks: Guidelines for Investigation and Control*. Geneva, Switzerland: World Health Organization; 2008
- [56] Kotula A, Dubey J, Sharar A, Andrews C, Shen S, Lindsay D. Effect of freezing on infectivity of *Toxoplasma gondii* tissue cysts in Pork. *Journal of Food Protection*. 1991;**54**:687-690
- [57] Dubey J, Kotula A, Sharar A, Andrews C, Lindsay D. Effect of high temperature on infectivity of *Toxoplasma gondii* tissue cysts in Pork. *The Journal of Parasitology*. 1990;**76**(2):201-204
- [58] Pott S, Koethe M, Bangoura B, Zöller B, Dausgchies A, Straubinger R, et al. Effects of PH, sodium chloride, and curing salt on the infectivity of *Toxoplasma gondii* tissue cysts. *Journal of Food Protection*. 2013;**76**:1056-1061
- [59] Mirza Alizadeh A, Jazaeri S, Shemshadi B, Hashempour-Baltork F, Sarlak Z, Pilevar Z, et al. A review on inactivation methods of *Toxoplasma gondii* in foods. *Pathogens and Global Health*. 2018;**112**:306-319
- [60] Porter SB, Sande MA. *Toxoplasmosis of the central nervous system in the acquired immunodeficiency syndrome*. *The New England Journal of Medicine*. 1992;**327**:1643-1648
- [61] Daher D, Shaghlil A, Sobh E, Hamie M, Hassan ME, Moumneh MB, et al. Comprehensive overview of *Toxoplasma gondii*-induced and associated diseases. *Pathogens*. 2021;**10**:1351
- [62] Dunn D, Wallon M, Peyron F, Petersen E, Peckham C, Gilbert R. Mother-to-child transmission of toxoplasmosis: Risk estimates for clinical counselling. *The Lancet*. 1999;**353**:1829-1833
- [63] Vivacqua DPF, Paz AB, Frota ACC, Penna CRR, Martins MG, Abreu TF, et al. Antenatal factors related to congenital toxoplasmosis in Rio De Janeiro, Brazil. *Journal of Maternal-fetal and Neonatal Medicine*. 2022;**35**:7200-7206
- [64] Andreoletti O, Budka H, Buncic S, Colin P, Collins JD, De A, et al. Surveillance and monitoring of *Toxoplasma* in humans, food and animals scientific opinion of the panel on biological hazards. *EFSA Journal*. 2007;**583**:1-64
- [65] Lopes AP, Sargo R, Rodrigues M, Cardoso L. High seroprevalence of

antibodies to toxoplasma gondii in wild animals from Portugal. *Parasitology Research*. 2011;**108**:1163-1169

[66] Wendte JM, Gibson AK, Grigg ME. Population genetics of toxoplasma gondii: New perspectives from parasite genotypes in wildlife. *Veterinary Parasitology*. 2011;**182**:96-111

[67] Chumpolbanchorn K, Lymbery A, Pallant L, Pan S, Sukthana Y, Thompson R. A high prevalence of toxoplasma in Australian chickens. *Veterinary Parasitology*. 2013;**196**:209-211

[68] Dubey J. Toxoplasma gondii infections in chickens (gallus domesticus): Prevalence, clinical disease, diagnosis and public health significance. *Zoonoses and Public Health*. 2010;**57**:60-73

[69] van der Giessen J, Fonville M, Bouwknegt M, Langelaar M, Vollema A. Seroprevalence of trichinella spiralis and toxoplasma gondii in pigs from different housing systems in The Netherlands. *Veterinary Parasitology*. 2007;**148**:371-374

[70] Vesco G, Buffolano W, La Chiusa S, Mancuso G, Caracappa S, Chianca A, et al. Toxoplasma gondii infections in sheep in Sicily, Southern Italy. *Veterinary Parasitology*. 2007;**146**:3-8

[71] Innes EA, Bartley PM, Buxton D, Katzer F. Ovine toxoplasmosis. *Parasitology*. 2009;**136**:1887-1894

[72] Hill D, Dubey J, Baroch J, Swafford S, Fournet V, Hawkins-Cooper D, et al. Surveillance of feral swine for Trichinella spp. and Toxoplasma gondii in the USA and host-related factors associated with infection. *Veterinary Parasitology*. 2014;**205**:653-665

[73] Buzoni-Gatel D, Schulthess J, Menard LC, Kasper LH. Mucosal defences

against orally acquired protozoan parasites, emphasis on toxoplasma gondii infections. *Cellular Microbiology*. 2006;**8**:535-544

[74] David Sibley L. Invasion and intracellular survival by protozoan parasites. *Immunological Reviews*. 2011;**240**:72-91