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The implications of socioeconomic factors on salivary bioscience methodological variables in a large pediatric multi-site study

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Introduction: Salivary bioscience has found increased utilization within pediatric research, given the non-invasive nature of self-collecting saliva for measuring biological markers. With this growth in pediatric utility, more understanding is needed of how social-contextual factors, such as socioeconomic factors or status (SES), influence salivary bioscience in large multi-site studies. Socioeconomic factors have been shown to influence non-salivary analyte levels across childhood and adolescent development. However, less is understood about relationships between these socioeconomic factors and salivary collection methodological variables (e.g., time of saliva collection from waking, time of day of saliva collection, physical activity prior to saliva collection, and caffeine intake prior to saliva collection). Variability in salivary methodological variables between participants may impact the levels of analytes measured in a salivary sample, thus serving as a potential mechanism for non-random systematic biases in analytes.

Methods: Our objective is to examine relationships between socioeconomic factors and salivary bioscience methodological variables within the Adolescent Brain Cognitive Development Study® cohort of children aged 9–10 years old ($n=10,567$ participants with saliva samples).

Results: We observed significant associations between household socioeconomic factors (poverty status, education) and salivary collection methodological variables (time since waking, time of day of sampling, physical activity, and caffeine intake). Moreover, lower levels of household poverty and education were significantly associated with more sources of potential bias in salivary collection methodological variables (e.g., longer times since waking, collections later in the day, higher odds of caffeine consumption, and lower odds of physical activity). Consistent associations were not observed with neighborhood socioeconomic factors and salivary methodological variables.

Discussion: Previous literature demonstrates associations between collection methodological variables and measurements of salivary analyte levels, particularly with analytes that are more sensitive to circadian rhythms, pH levels, or rigorous physical activity. Our novel findings suggest that unintended distortions in measured salivary analyte values, potentially resulting from the non-random systematic biases in salivary methodology, need to be intentionally incorporated into analyses and interpretation of results. This is particularly salient for future studies interested in examining underlying mechanisms of childhood socioeconomic health inequities in future analyses.

KEYWORDS

socioeconomic status, salivary bioscience, child and adolescent development, methodology, health inequities

1. Introduction

Socioeconomic factors or status (SES) that drive health inequities are well established (1–3). However, a thorough understanding of SES-driven health inequities is needed within pediatric populations to elucidate early-life biological antecedents of adult health inequities. Previous studies among pediatric populations demonstrate multiple salivary biomarkers implicated in associations between the broader social environment and physiology, including neuroendocrine markers (e.g., alpha-amylase, cortisol, DHEA), metabolic markers (e.g., insulin, glucose), and immune markers (e.g., c-reactive protein, cytokines) (4–7). However, a number of these biomarkers rely on invasive sampling techniques, particularly blood draws, risking harm to participant-researcher rapport and overall willingness of communities to participate in biomedical research, particularly among pediatric populations. One approach to address this research gap in biological measures among pediatric studies is the use of salivary biosciences.

Salivary biospecimen technologies have grown in popularity over the last decade within research studies and clinical testing to non-invasively measure levels of analytes within diverse human populations (8). This utility is primarily due to its contextual practicality, allowing for sample collection outside of laboratory or clinical settings, as well as the non-invasiveness and feasibility of saliva sampling relative to more invasive techniques, such as phlebotomy (9–11). The many advantages of collecting salivary samples over other types of biospecimens in research include (1) being a low-cost option particularly for studies requiring multiple samples, (2) the ability for a participant to self-sample, and (3) adaptability to various field settings (10, 12–14). This method offers increased feasibility to measure physiological correlates of SES and related factors given the non-invasive nature, ease of collection of salivary samples, and reduced cost of sampling (10). These cost-saving benefits afford strengthening of study design such as sampling from more participants, increased number of collections within participants, or increased number of biomarkers assayed from each saliva sample. Further, salivary bioscience demonstrates great potential for diagnostic capability including pediatric endocrine dysfunction, cardiometabolic disease (15), monitoring lithium levels for psychiatric disorders (16), and diagnosing COVID-19 at home (17).

Additional methodological strengths of salivary sampling allow for the inclusion of communities that have been traditionally underrepresented in research and eases the burden of participation for families, improving adherence (13, 18). Certainly, a history of scientific injustices exists, disproportionately affecting low socioeconomic status and racially/ethnically minoritized communities, and driving historical and current-day underrepresentation in biomedical research that has often resulted in varying degrees of distrust of researchers (19–21). These historical and current injustices often occur when the cultural appropriateness of biological sample collection is not adequately considered (19, 22). Salivary collection is a tool that can minimize cultural insensitivities inherent in the collection of biological data, given its acceptance among diverse adolescent communities (23, 24). However, it is important to note that any biological collection can be precarious and warrants culturally and equity guided investigations. Some potential examples include: (a) some cultures or communities may feel averse to producing a saliva sample, particularly when observed by an

experimenter, and may prefer other biological methods over saliva; (b) age of study sample matters, with children generally exhibiting aversion to blood sampling but willingness to produce saliva; and (c) certain cultures may perceive discarding unused saliva into waste as disrespectful. It is our recommendation that the community preferences for or against saliva collection be well understood before leveraging salivary biosciences.

Given these advantages, feasibility, and promising diagnostic future of salivary biosciences it is essential to first understand how the experimental design and saliva collection methodology should be standardized to ensure precision of measured analyte levels, particularly for the investigation of health inequities, and for increased application within pediatric research or clinical utility. Without this deeper methodological understanding, spurious differences in experimental design and methodological implementation of salivary biosciences may undermine the interpretability, accuracy, and utility of salivary analytes.

Several decisions in the experimental design can directly influence the methodology of salivary sample collections. For example, a design that rigorously standardizes collection of salivary samples can reduce or eliminate unintentional biases due to variations in collection methodological variables. These methodological decisions include how much time should be allowed between a participant's waking time to their saliva collection time, the time of day the saliva sample is collected, the amount of physical activity allowed prior to sampling, if caffeine is consumed prior to sampling, or other oral considerations that can impact measured analyte levels (5, 8, 12, 25, 26). Standardized collection practices help eliminate unintended experimental noise, where non-biological factors may influence the composition or volume of whole unstimulated saliva (27). Without stringent standardized collection practices of how and when saliva samples are collected, leveraging salivary biosciences on a large scale may result in unintended methodological variations, which can impact the analyte levels measured in the collected saliva sample and thus take a detour from true biological levels, warranting caution (28).

Many adrenal steroid analytes demonstrate diurnal/circadian or seasonal rhythms, marked by patterns of varying levels over an extended period of time. For example, cortisol, a marker of psychological stress, fluctuates throughout the day, peaking approximately 30–45 min after waking followed by tapering levels in the evening (e.g., 3–12 h after waking) (11, 29). In addition, the amount of sunlight at various points of the day drives circadian rhythms (30). Waking later in the day when sunlight is different than morning light may shift circadian phases and thus alter typical patterns of analytes.

Not only is the time since waking important, the time of day when the sample is collected is also a source of experimental variation. For example, salivary dehydroepiandrosterone (DHEA) and testosterone levels are typically highest in morning samples and drop continuously throughout the day to produce lower levels in evening samples (31–34). In addition, DHEA is implicated in physiological responses to acute stress (35, 36). Thus, saliva sampled later in the day may represent different hormonal profiles compared to morning collections given fluctuating levels with circadian patterns, or greater opportunity to experience acute stressors as the day goes on. Given these considerations, minimizing variations in collection practices or pre-collection exposures are important for making accurate conclusions about the source of differences in analyte levels. Variations in methodological factors may

become increasingly problematic for obtaining precise measured analyte levels in maturing adolescent populations, especially where pubertal maturation is underlying the biological systems producing the analytes of interest.

Further, methodological variables related to lifestyle such as rigorous physical activity and caffeine intake prior to salivary sample collection may introduce bias in analyte levels by altering physiological states or the integrity of the saliva sample. Rigorous (>20 min) physical activity can alter levels of DHEA or testosterone (37), particularly in saliva samples taken during early stages of pubertal maturation when hormone levels are very low (38). Salivary DHEA levels among adolescent males have been documented to increase post-exercise, yet with varying slopes according to pubertal development (35). Caffeine intake prior to saliva sampling can impact analyte levels through a few different mechanisms, including shifting the salivary pH, increasing sample acidity, and therefore impacting the performance of certain pH-sensitive assays (5, 39), or promote bacterial growth, thereby compromising the integrity of salivary fluid (40). In addition, caffeine intake may risk dehydration in the participant that would reduce salivary flow rate, and/or activation of physiological pathways that overlap with origins of the analyte of interest, such as caffeine activating the adrenergic pathway and increasing urine concentrations of metanephrine (41–43). Although these observations are in serum or urine samples, unclear evidence on correlations of serum/urine metanephrine with salivary levels as a function of caffeine intake warrants consideration of caffeine exposure in salivary collections.

Standardized collection practices can minimize differences between and within participants in these methodological variables by regulating time of day when the saliva sample is collected, prohibiting participants from consuming caffeine or performing rigorous exercise beforehand, and standardizing the duration of saliva sampling between and within participant sampling (25). Analytes closely connected to circadian patterns may be particularly sensitive to variability in sampling times, or alterations in pH levels due to caffeine consumption. The present analysis examined relationships with several salivary methodological collection variables in a large US-based, representative pediatric cohort participating in the Adolescent Brain Cognitive Development Study® (44). In the ABCD Study, detailed data was collected on methodological variables mentioned above, but was not standardized in the collection protocol allowing for our evaluation of potential non-random methodological variation relating to saliva collection and key socioeconomic factors.

Socioeconomic factors have been of central focus for understanding health inequities. Socioeconomic factors reflect access to economic or social resources and are often represented by individual or composite measures of household income level, poverty status, parental education attainment, or occupation (45). These factors have been described in the literature to influence child developmental outcomes. Low SES has been associated with poor school readiness and academic achievement, more frequent adverse experiences, structural brain differences, and altered executive functioning (46–50). Studies investigating the relationship with SES using salivary samples among children from low SES households have noted higher baseline neuroendocrine profiles and steeper neuroendocrine trajectories over time relative to children from high SES households (51, 52).

SES has been purported to operate as a function of resource availability for a study participant (53). If collected at the home,

participants may have limited access to freezers to store salivary samples, mailing resources to mail collected saliva, technology, such as text messages or phone, that would facilitate reminders to collect samples at consistent timings or more accurate collection time records without the aid of digital tools (25). Possible limited availability and access to social and economic resources may influence salivary sample collection variables when participants self-schedule throughout the day when to come into the laboratory for sampling. Thus, collections performed at a laboratory or at a study site issue the question whether collection methods differ as a function of participant resource availability.

Relationships between SES and other variables important in salivary collection, namely physical activity and caffeine consumption, have been demonstrated. Positive relationships between SES and the amount of physical activity performed among adolescents have been reported, such that low SES tends to be associated with less physical activity compared to those with a high SES (3, 54, 55). However, variations in the measurement of both SES (e.g., income-to-needs ratio, household income, parental occupation, parental education) and amount of physical activity (e.g., time or duration, frequency, school-based or extracurricular) may contribute to some null findings (55). Despite overall reductions in the amount of caffeine consumption among children and adolescents since 2000, those living at 0–99% and 100–199% of the federal poverty level have consistently consumed caffeine at higher rates compared to those living at greater than 200% of the federal poverty level (56). Particularly among children ages 6–11 years old, rates of caffeine consumption in households with low or very low food security and income-to-poverty ratios below 2.0 are significantly higher compared to households with income-to-poverty ratios above 2.0 (57). Thus, child/adolescent physical activity and caffeine consumption are a possible source of methodological variation in saliva collection when not standardized in the collection design.

Given that many analyte levels fluctuate on a circadian rhythm, patterns of saliva collections earlier or later in the day among one socioeconomic context relative to others in the study sample would suggest potential non-random systematic errors in salivary analyte values due to broader social determinants. Similarly, socioeconomic-related differences in physical activity or caffeine consumption prior to salivary sampling may serve as another mechanism for non-random systematic errors in salivary analyte levels. Without disentangling these contributors, the inclusion of these salivary analyte values in analyses would bias conclusions regarding differences in biological outcomes. Thus, it remains important to capture a greater understanding of socioeconomic influences on salivary bioscience methodology before leveraging salivary data for accurate investigation of health inequities. The present analyses will inform how special considerations need to be made when leveraging salivary analyte levels from large multi-site studies in childhood, a critical period of development when inequities during early life developmental periods, “get under the skin.”

Investigations of the relationship between salivary collection methodological variables and socioeconomic factors among child populations are limited. However, with the emergence of salivary technology we are observing widespread utilization of salivary biosciences in large cohort studies. The objective of this study was to examine the association between key socioeconomic factors (e.g., poverty status, household education, neighborhood deprivation) and

key salivary sample collection methodological variables (e.g., time since waking, collection time of day, and caffeine intake and physical activity within 24 h of sampling) among a diverse and large sample of US-based children aged 9–10 years old.

2. Materials and methods

2.1. Background on study sample and sample characteristics

This analysis was performed using a sample of children aged 9–10 years at enrollment participating in a 21-site study in the United States from the Adolescent Brain Cognitive Development (ABCD) Study© Release 3.0. This dataset was selected given that it is a large-scale longitudinal (e.g., annually over the course of 10 years) pediatric collection of whole saliva via passive drool for analysis of several hormonal analytes (e.g., estradiol among females only, DHEA and testosterone among males and females). Although there have been three collection timepoints to-date in this dataset (e.g., enrollment/baseline, year 1, and year 2), this current analysis focuses on baseline measures collected in 2016–2018 only. Longitudinal change was not the focus of the *a priori* aims, and any existing methodological variation observed at baseline are most likely repeated and similar in future waves of saliva collection in this cohort.

Participants reported to the study site for salivary sample collection, where one salivary sample was collected via passive drool from each participant at each annual timepoint (58). Participants and their guardian/parent did not receive prior instruction to prepare for the saliva collection during the study visit (e.g., participants were not instructed to abstain from eating, caffeine, or vigorous exercise prior to study visit). Upon arrival at the study site, a minimum of 30 min time passed between participants' arrival and starting the saliva collection. During this time, participants were instructed to not eat or drink anything other than water (including no mints/gum), then asked to rinse their mouth out with water 10 min prior to providing the saliva sample. If participants were given a lunch break, or arrived immediately after lunch, the protocol allowed for minimum of 60 min before sampling. Thus, the majority of saliva samples occurred ~60 min after a large meal (38, 58). Participants and their guardian/parent arrived at the study site for collection based on when the study site and participant schedules aligned. Current guidelines for optimal utilization of salivary bioscience recommend the notation of time of recent meal, oral health or injuries, braces, or recent loss of deciduous teeth (5). However, many of these variables were not controlled or collected in the ABCD Study given considerations for reducing participant burden, and experimentally prioritize the central aims of the ABCD study including multi-modal MRI, comprehensive profiles of adolescent substance use, and mental health assessments.

When present at the study site, a research assistant (RA) documented the arrival time of the participant, presence of parent or guardian, and the time the participant reported waking. After the RA instructed the participant to passively drool into a sample collection tube, the RA then documented the timing of the salivary sample, duration of sample collection, discoloration, or visible imperfections, as well as duration from collection to placement into a -20°C to -80°C freezer. Guardians/parents were compensated for their participation

in the ABCD study, with the level of compensation being varied between study sites to account for differences in cost of living (44). Salivary samples were then shipped from study sites on dry ice, confirmed for frozen state upon arrival, and assayed by an external laboratory (59).

To reduce statistical noise within the analytic sample unrelated to sampling methodological variables, we removed participants whose biological sex at birth was not collected ($n=7$), reported unable to complete ($n=59$), and refused ($n=19$) from analyses. We further cross-referenced each participant's biological sex at birth with the biological sex reported at the time of salivary sample collection and removed those with mismatched sex ($n=23$). We adopted this decision to cross-reference reported sex at birth with biological sex reported at Baseline collections because early ABCD protocol indicated that a participant's sex at birth would determine which hormone panel (e.g., being inclusive or exclusive of estradiol) would be analyzed at the study visit. Only 2 participants were marked as male at birth but had missing entries at salivary sample collection. Those 2 participants were reclassified as male for analyses. We also reclassified the 4 participants reported as intersex (I) at birth (Figure 1) with the sex reported at salivary sample collection. In addition, participants with a gestational age less than 28 weeks and a reported birthweight less than 1,200 grams were removed from the analytic sample. These participants were erroneously included in the study given that the exclusion criteria required gestational age to be 28 weeks or greater. The final analytic sample consisted of $n=10,567$, of which 5,534 were male and 5,033 were female at baseline (Figure 1).

2.2. Measures

2.2.1. Demographic and socioeconomic variables

The inclusion of child age in statistical analyses (in months) was informed by evidence of differential sleep habits, caffeine intake, and physical activity habits between children ages 7 to 10 years old. Sleep habits including sleep duration, which may inform waking time before salivary collection, is significantly associated with child age around 9- and 10-year-olds (e.g., sleep duration decreases as child age increases) (60, 61). Further, documented significant declines in physical activity with increases in child age between ages 9 and 15 years (62–64) demonstrates a need to control for child age as a precision variable due to independent relationships with the outcome in these analyses. Regarding caffeine intake, inconsistent relationships in the literature warrant investigation in our analyses. While previous evidence demonstrates general increases in caffeine intake with increases in age, several studies (65) observed lower caffeine intake between 9- and 10-year-olds, while other studies observed similar caffeine intake among 9–10-year-olds (66). Given these existing associations, bivariate relationships were examined between child age and salivary methodological variables. After observing significant bivariate relationships (Table 1), multivariate models were adjusted for child age as a precision variable to isolate effects due to independent relationships between each predictor and the outcomes.

To examine relationships between salivary collection methods with socioeconomic factors, we constructed the following measures.

Poverty status represents the household's socioeconomic position relative to the federal poverty level (FPL). This was indexed according

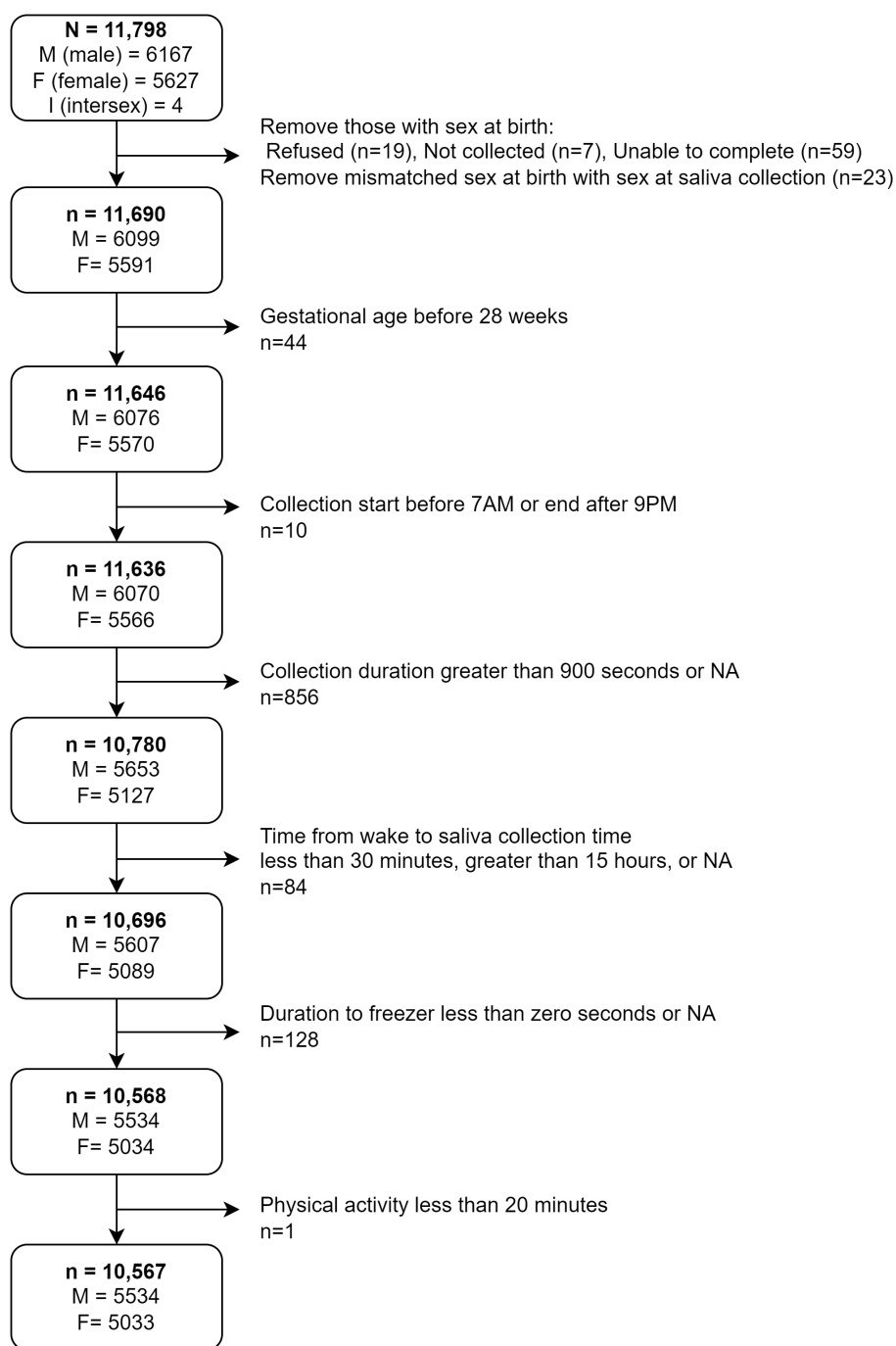


FIGURE 1
Depiction of decision tree to obtain final analytical sample. M, Male; F, Female; I, Intersex.

to the reported combined household income and the reported number of family members living from that combined income in the household. Poverty status was categorized relative to the FPL according to the following: Deep Poverty (<50%), Poverty (50–<100%), Near Poverty (100–<200%), Mid Income (200–<400%), High Income (≥400%). Although group membership across poverty status levels is imbalanced (Table 1), we made an evidence-informed decision to distinguish Deep Poverty from Poverty. From 1996 to 2011 the percentage of households living in Deep Poverty has grown 129.6% while the percentage of households in Poverty has grown

80.4% (67). Children living in Deep Poverty are at greater risk of adverse physical health and intellectual outcomes compared to children in poverty but who are not deeply poor, and children not living in poverty (68–70). Therefore, Deep Poverty is an important, unique construct of experienced poverty.

The participant’s guardian/parent self-reported their level of education, and if partnered, also reported the partner’s level of education. Household education in our analyses represents the highest level of education in the household reported by the parent. If the parent reported having a partner, then the highest level of education

TABLE 1 Descriptive statistics between socioeconomic variables and salivary methodological variables.

	Mean (SD)	Time since waking (hours)		Collection time of day (hours since midnight)		Physical activity			Caffeine intake		
		r_s	P^A	r_s	P^A	<20	>20	P^B	Yes	No	P^B
		Range: 0.60–14.63		Range: 7.02–20.70							
						Mean (SD)	Mean (SD)		Mean (SD)	Mean (SD)	
Child age (months)	118.9 (7.5)	−0.047	<0.001	−0.041	<0.001	119 (7.5)	118.8 (7.7)	ns	119.3 (7.5)	118.9 (7.5)	ns

Household poverty status	N	Time since waking (hours)		Collection time of day (hours since midnight)		Physical activity			Caffeine intake		
		Mean (SE)	P^B	Mean (SE)	P^B	<20	>20	P^C	Yes	No	P^C
						N (%)	N (%)		N (%)	N (%)	
Deep poverty	798	5.98 (0.11)	0.01	13.05 (0.11)	<0.001	732 (7.8)	64 (5.4)	0.03	84 (12.1)	713 (7.3)	<0.001
Poverty	616	5.89 (0.13)		13.04 (0.13)		545 (5.8)	69 (5.8)		57 (8.2)	557 (5.7)	
Near poverty	1,541	5.89 (0.08)		12.99 (0.08)		1,369 (14.7)	169 (14.2)		127 (18.4)	1,413 (14.4)	
Mid income	2,358	5.84 (0.06)		12.96 (0.06)		2083 (22.3)	270 (22.8)		163 (23.6)	2,192 (22.3)	
High income	4,165	5.62 (0.05)		12.69 (0.04)		3,654 (39.1)	499 (42.0)		185 (26.7)	3,973 (40.4)	
Household education											
Less than HS	523	6.05 (0.13)	0.28	13.26 (0.13)	0.02	473 (5.1)	47 (4.0)	<0.001	59 (5.8)	461 (4.7)	<0.001
HS graduate	1,004	5.78 (0.10)		12.96 (0.10)		897 (9.6)	105 (8.8)		111 (16.0)	891 (9.1)	
Some college or associate	2,748	5.78 (0.06)		12.88 (0.06)		2,466 (26.4)	273 (23.0)		241 (34.8)	2,503 (25.5)	
College graduate	2,660	5.78 (0.06)		12.87 (0.06)		2,364 (25.3)	293 (24.7)		144 (20.8)	2,514 (25.6)	
Graduate or professional	3,596	5.76 (0.05)		12.85 (0.05)		3,118 (33.4)	469 (39.5)		137 (19.8)	3,454 (35.1)	
Household marital status											
Yes	7,082	5.87 (0.04)	0.78	12.86 (0.03)	0.05	6,216 (66.6)	848 (71.4)	0.001	392 (56.7)	6,680 (68.0)	<0.001
No	3,377	5.81 (0.05)		12.96 (0.05)		3,037 (32.6)	332 (28.0)		292 (42.2)	3,079 (31.3)	
Area deprivation index											
Quartile 1 (least deprived)	2,488	5.57 (0.06)	0.003	12.73 (0.06)	0.10	2,190 (23.5)	292 (24.6)	0.004	117 (16.9)	2,368 (24.1)	<0.001
Quartile 2	2,484	5.87 (0.06)		12.96 (0.06)		2,147 (23.0)	328 (27.6)		131 (18.9)	2,347 (23.9)	
Quartile 3	2,482	5.87 (0.06)		12.97 (0.06)		2,215 (23.7)	260 (21.9)		177 (25.6)	2,303 (23.4)	
Quartile 4 (most deprived)	2,485	5.83 (0.06)		12.87 (0.06)		2,225 (23.9)	257 (21.7)		223 (32.2)	2,258 (23.0)	

Bivariate relationships between salivary collection variables and socioeconomic variables are represented in Table 2.

^AP-values of Spearman Correlation tests reflect correlations between continuous salivary collection variables and continuous child age. ^BP-values of Kruskal-Wallis tests reflect associations between child age and categorical physical activity and caffeine intake. value of ps of Kruskal-Wallis tests also reflect associations between continuous salivary collection variables and categorical household poverty status, household education, household marital status, and Area Deprivation Index (ADI). ^CP-values of Chi-Square test of independent reflect associations between categorical salivary collection variables and categorical socioeconomic variables. r_s , Spearman correlation coefficient; SD, standard deviation; P , p -value; ns, non-significant.

TABLE 2 Bivariate correlations between socioeconomic variables.

	Household poverty status	Household education	Household marital status	ADI
Household poverty status	–	–	–	–
Household education	0.63**	–	–	–
Household marital status	0.46**	0.42**	–	–
ADI	–0.46**	–0.39**	–0.26**	–

Spearman rank correlations coefficients (r_s) between ordinal categorical socioeconomic variables are represented below. Values range from -1 to $+1$ reflecting strong negative to strong positive correlations, respectively. ADI, Area Deprivation Index.

** $p < 0.001$.

by either the reporting parent or the partner was included in the analyses. Otherwise, if the reporting parent did not have a partner, the single-caregiver's reported education level was used. Previous evidence demonstrates strong positive correlations between reports of maternal education, paternal education, and the highest education level of either parent in household (71, 72). Thus, to leverage a single operationalization of household education and to reflect inclusivity in gender-neutral terminology (73), we used the highest level of education in the household reported by the parent.

Household marital status was categorized as, “yes,” if the parent reported being married. Otherwise, marital status was categorized as, “no,” if the parent reported being widowed, divorced, separated, never married, or living with partner.

Area deprivation index (ADI) was calculated as the scaled weighted sum of 17 neighborhood-level characteristics within the participant's reported census block group. A detailed list of census variables has been summarized in Kind et al. and adapted for use in ABCD (74, 75). This includes proportion of population aged ≥ 25 years with < 9 years of education; proportion of population aged ≥ 25 years with less than high school diploma; proportion of employed persons age 16+ in a “white collar” occupation; median household income; income disparity; median home value; median gross rent; median monthly mortgage; percent owner-occupied housing; percent of population age 16+ unemployed; percent of families below poverty line; percent of population below 138% of poverty line; percent of single-parent households with children < 18 years; percent occupied housing units without vehicle; percent occupied units without telephone; percent occupied units without complete plumbing; percent occupied units with more than 1 person per room (74). Higher ADI scores, and thus upper quartile categorization, refer to higher levels of area deprivation, while lower quartile categorization refers to lower levels of area deprivation. Similar assessments of ADI have been widely applied in pediatric developmental research and support the validity of ADI for predicting child and family well-being (76–78). Specifically, within the ABCD cohort, many childhood outcomes such as brain structure and function, as well as body mass index, are associated with the ADI measure used in this analysis (79–81).

2.2.2. Methodological variables for salivary collection

The following salivary collection variables were analyzed.

Time since waking reflects the duration of time from the participant's self-reported time of waking to the start of the salivary sample collection documented by the RA. If a participant's time since waking was calculated to be less than 30 min, greater than 15 h, or was missing, values were assumed to be erroneous data, and therefore were

excluded from the analyses ($n = 84$). Samples with time since waking less than 30 min were removed because due to ABCD protocol, it is highly unlikely that saliva sampling occurred within this time frame. Specifically, after participants arrived at the study site, the research assistant preformed a series of pre-collection assessments, including obtaining consent/assent, explanation of saliva sampling, and conducting demographic and pubertal questionnaires before soliciting a saliva sample (82). Given that the estimated time to complete these steps was at least 30 min, samples documented to be collected within 30 min of waking are likely erroneous.

Collection time of day refers to the time of day the salivary sample collection took place at the local study site laboratory. Collections that were reported before 06:00 a.m. and after 9:00 p.m. were assumed to be erroneous data, and therefore excluded from the analyses ($n = 10$).

Physical activity was categorized dichotomously, reflecting whether the participant was vigorously physically active (sweating, breathing heavy) for at least 20 min within the 12 h prior to sampling. Participants were classified into less than 20 min of physical activity, or greater than 20 min of physical activity.

Caffeine intake was categorized dichotomously as a yes or no response, referring to whether the participant reported consuming caffeine from drink within the 12 h prior to sampling. We categorized affirmative responses coinciding with reports of non-zero milligrams of caffeine as, “yes,” and denial responses coinciding with reports of zero milligrams of caffeine as, “no,” for these analyses.

2.3. Statistical analyses

Associations between socioeconomic variables and salivary collection variables were examined through a series of bivariate tests. A Spearman test of correlation (r_s) was performed to examine correlations between ordinally coded socioeconomic variables (Table 2). Given that neither the participant's age in months nor the continuous salivary collection variables were normally distributed, a Spearman test of correlation (r_s) was performed to examine the strength and direction of their relationship (Table 1). A Kruskal-Wallis non-parametric test of equality (H test statistic) was performed to identify differences in continuous salivary collection variables between levels of categorical socioeconomic variables (Table 1). A Chi-square test of independence (X^2) was performed to identify associations between categorical salivary collection variables and categorical socioeconomic variables (Table 1).

A series of univariate and multivariate multi-level linear or logistic mixed effects models were performed to examine potential confounding effects among socioeconomic factors determining salivary collection outcomes. To account for clustering effects by study

site, random intercepts were specified according to study site as level 2 and subject ID as level 1 random intercepts. Time since waking (skew = 0.5, kurtosis = -0.81) and collection time of day (skew = 0.40, kurtosis = -0.92) were log-transformed due to non-normality prior to analyses. Post-transformation skew and kurtosis for time since waking (skew = -0.30 and kurtosis = -0.76) and collection time of day (skew = 0.08, kurtosis = -1.05) were improved. Due to log transformation of continuous outcomes, beta coefficients in regression models were exponentiated to improve interpretability. The Bonferroni-corrected significance level was set to $\alpha = 0.0125$ for 4 outcomes. Bonferroni corrected p -values are reported.

All tests were performed in R Statistical Software Studio version 1.3.1073 utilizing the following packages: nlme (83), car (84), piecewiseSEM (85), lubridate (86), Hmisc (87).

3. Results

3.1. Descriptive statistics

Within the entire sample, the mean number of hours between participant waking and time of collection was 5.79 h, and the average time of collection was approximately 12 h and 53 min after midnight local time (not pictured). Time since waking and collection time of day were significantly strongly positively correlated ($r_s = 0.93$, $p < 0.05$). No significant associations were observed between physical activity and caffeine intake ($X^2 = 0.25$, $df = 1$, $p = 0.61$). A descriptive summary of salivary collection methods for the entire analytical sample, according to income group, is presented in Figure 2. Correlations between socioeconomic variables for the entire analytic sample are reflected in Table 2. All socioeconomic variables were significantly correlated with each other, albeit with ranging direction and strengths (p -values < 0.05 ,

Table 2). Household poverty status was strongly positively correlated with household education ($r_s = 0.63$, $p < 0.05$), yet moderately positively correlated with household marital status ($r_s = 0.46$, $p < 0.05$). Household education was also moderately positively correlated with household marital status ($r_s = 0.42$, $p < 0.05$). ADI was negatively correlated with household poverty status ($r_s = -0.46$, $p < 0.05$), education ($r_s = -0.39$, $p < 0.05$), and marital status ($r_s = -0.26$, $p < 0.05$).

Child age (mean \pm SD = 118.9 \pm 7.5 months) was significantly negatively correlated with time since waking and collection time of day, albeit weakly ($p_s < 0.05$, Table 1). No significant bivariate associations were observed between child age in months and physical activity nor caffeine intake.

Significant bivariate associations were observed between household poverty status and all salivary collection measures, but varying relationships between other SES factors and salivary collection measures. Mean time since waking was significantly different between levels of household poverty status (Table 1; $H = 12.4$, $df = 4$, $p = 0.01$), yet it was not significantly associated with household education (Table 1; $H = 5.04$, $df = 4$, $p = 0.28$). Household marital status was also not significantly associated with time since waking (Table 1; $H = 0.08$, $df = 1$, $p = 0.78$). Regarding bivariate associations at the neighborhood-level with ADI, mean time since waking ($H = 13.9$, $df = 3$, $p = 0.003$) was significantly different between quartiles of neighborhood deprivation (Table 1).

Additionally, while mean collection time of day was significantly different between levels of household poverty status (Table 1; $H = 25.8$, $df = 4$, $p < 0.001$) and household education (Table 1; $H = 11.6$, $df = 4$, $p = 0.02$), it was not significantly associated with household marital status (Table 1; $H = 3.8$, $df = 1$, $p = 0.05$) nor ADI (Table 1; $H = 6.2$, $df = 3$, $p = 0.10$).

Lastly, categories of physical activity and caffeine intake were not significantly independent (e.g., reject null hypothesis) of household

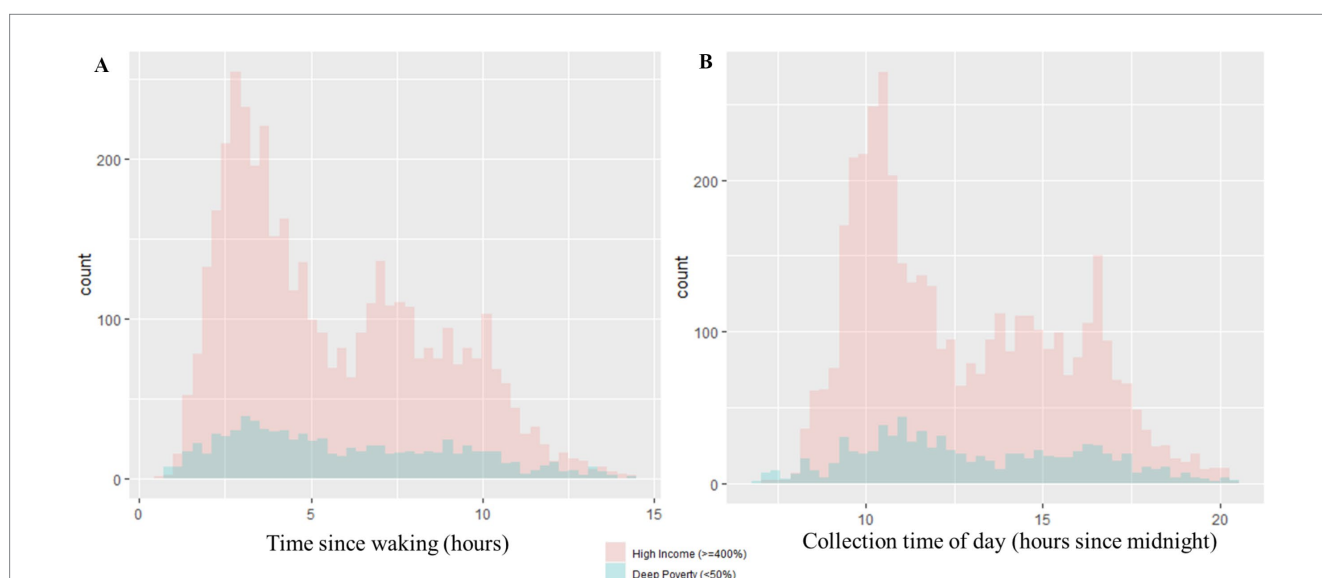


FIGURE 2

Distributions of salivary collection variables. (A) The distribution of time since waking in hours is represented by (A). The distributions are further depicted according to poverty status (Deep Poverty versus High Income groups). Within the Deep Poverty group, the range of time since waking is 0.77–14.38h and the median is 5.32h. Within the High Income group, the range of time since waking is 0.60–14.33h and the median is 4.85h. (B) The distribution of collection time of day in hours since midnight is represented by (B). The distributions are further depicted according to poverty status (Deep Poverty versus High Income groups). Within the Deep Poverty group, the range of collection time of day is 7.02–20.42h and the median is 12.46h. Within the High Income group, the range of collection time of day is 7.27–20.53h and the median is 11.92h.

poverty status, household education, marital status, nor ADI (Table 1). Whether or not a participant engaged in physical activity prior to sampling appeared to be significantly associated with household poverty status ($X^2 = 10.8$, $df = 4$, $p = 0.03$), household education ($X^2 = 19.7$, $df = 4$, $p < 0.001$), marital status ($X^2 = 10.3$, $df = 1$, $p = 0.001$) and ADI ($X^2 = 13.3$, $df = 3$, $p = 0.004$). In addition, caffeine consumption prior to sampling was significantly associated with household poverty status ($X^2 = 66.6$, $df = 4$, $p < 0.001$), household education ($X^2 = 124.3$, $df = 4$, $p < 0.001$), marital status ($X^2 = 35.8$, $df = 1$, $p < 0.001$), and ADI ($X^2 = 45.8$, $df = 3$, $p < 0.001$).

3.2. Child age

In univariate models, no significant independent relationships were observed between child age in months and time since waking and collection time of day. However, because of significant bivariate associations between child age and these salivary collection methods (Table 1), child age (months) was adjusted for in multivariate models predicting the outcomes described below.

3.3. Time since waking

Time since waking refers to the timeframe between the participant’s waking time and subsequent start of saliva

collection. Univariate analyses demonstrated a significant 5.34% longer time since waking among deep poverty households compared to high income households (Table 3; $\beta = 0.05$; $p < 0.0125$). ADI was not significantly associated with time since waking (Table 3).

3.3.1. Multivariate

When adjusting for child age or ADI in multivariate analyses, significant relationships were observed between household poverty status and a longer time since waking (Table 3; Model 1 and Model 2). Deep poverty households demonstrated a significant 2.06% longer time since waking compared to high income households, adjusting for only child age (Table 3; $\beta = 0.02$; $p < 0.0125$). Moreover, when adjusting for both child age and ADI, time since waking was significantly 5.88% longer among deep poverty households compared to high income households (Table 3; $\beta = 0.057$; $p < 0.0125$).

3.4. Collection time of day

Collection time of day refers to the local time of day of the salivary sample collection. In univariate analyses, deep poverty households significantly demonstrated collection start times 2.43% later in the day compared to high income households (Table 4; $\beta = 0.024$; $p < 0.0125$). No significant differences were observed between marital status, levels

TABLE 3 Univariate and multivariate multilevel linear models of log transformed time since waking.

	Time since waking (log)								
	Univariate			Multivariate					
	% inc or dec	Beta (SE)	P	Model 1			Model 2		
				% inc or dec	Beta (SE)	P	% inc or dec	Beta (SE)	P
Intercept					1.684 (0.104)	<0.001		1.684 (0.107)	<0.001
Household poverty status									
(Intercept)	374.85	1.558 (0.062)	<0.001						
Deep poverty	5.34	0.052 (0.02)	0.011	2.06	0.020 (0.020)	0.012	5.88	0.057 (0.022)	0.011
Poverty	-2.07	-0.021 (0.022)	0.352	2.27	0.022 (0.022)	0.349	-1.12	-0.011 (0.024)	0.645
Near poverty	0.36	0.004 (0.016)	0.820	1.58	0.016 (0.016)	0.823	0.60	0.006 (0.017)	0.725
Mid income	0.42	0.004 (0.013)	0.750	1.34	0.013 (0.013)	0.780	0.52	0.005 (0.014)	0.713
High income	Ref			Ref			Ref		
Area deprivation index									
Quartile 1 (least deprived)	Ref			Ref			Ref		
(Intercept)	377.10	1.563 (0.062)	<0.001						
Quartile 2	1.35	0.013 (0.015)	0.385	-	-	-	1.30	0.013 (0.016)	0.431
Quartile 3	-0.40	-0.004 (0.016)	0.796	-	-	-	-1.06	-0.011 (0.017)	0.542
Quartile 4 (most deprived)	0.55	0.005 (0.017)	0.755	-	-	-	-1.28	-0.013 (0.02)	0.530

All models adjusted for child age (months). % inc or dec refers to exponentiated beta coefficient and reflect percent increase or decrease from reference group. Due to log transformation of the outcome, beta coefficients were log transformed to improve interpretability. SE, standard error; P, p-value.

TABLE 4 Univariate and multivariate multilevel linear models of log transformed collection time of day.

	Collection time of day (log)								
	Univariate			Multivariate					
	% inc or dec	Beta (SE)	P	Model 3			Model 4		
				% inc or dec	Beta (SE)	P	% inc or dec	Beta (SE)	P
Intercept				1,143	2.52 (0.042)	<0.001	1,143	2.52 (0.042)	<0.001
Household marital status									
Intercept	1134.60	2.5133 (0.0259)	<0.001						
No	0.77	0.0076 (0.0043)	0.073	0.28	0.0028 (0.0052)	0.582	0.28	0.0028 (0.0053)	0.600
Yes		Ref			Ref			Ref	
Household poverty status									
(Intercept)	1129.54	2.509 (0.026)	<0.001						
Deep poverty	2.43	0.024 (0.008)	0.003	2.41	0.024 (0.01)	0.016	2.60	0.026 (0.01)	0.013
Poverty	0.02	0.0002 (0.009)	0.985	0.08	0.001 (0.01)	0.940	0.50	0.005 (0.011)	0.633
Near poverty	0.54	0.005 (0.006)	0.375	0.77	0.008 (0.007)	0.274	0.90	0.009 (0.007)	0.222
Mid income	0.66	0.007 (0.005)	0.202	0.75	0.008 (0.006)	0.173	0.89	0.009 (0.006)	0.125
High income		Ref			Ref			Ref	
Household education									
(Intercept)	1133.21	2.512 (0.026)	<0.001						
Less than HS	0.39	0.004 (0.01)	0.685	-1.07	-0.011 (0.013)	0.409	-1.42	-0.014 (0.014)	0.299
HS graduate	1.23	0.012 (0.007)	0.090	0.64	0.006 (0.009)	0.493	0.77	0.008 (0.01)	0.434
Some college or associate	0.37	0.004 (0.005)	0.465	-0.75	-0.007 (0.006)	0.236	-0.79	-0.008 (0.007)	0.229
College graduate	0.39	0.004 (0.005)	0.438	0.23	0.002 (0.005)	0.664	0.19	0.002 (0.006)	0.729
Graduate or professional		Ref			Ref			Ref	
Area deprivation index									
Quartile 1 (least deprived)		Ref			Ref			Ref	
(Intercept)	1134.21	2.513 (0.026)	<0.001						
Quartile 2	0.75	0.008 (0.006)	0.210	-	-	-	0.64	0.015 (0.016)	0.320
Quartile 3	0.00	0.00002 (0.006)	0.997	-	-	-	-0.52	-0.007 (0.018)	0.446
Quartile 4 (most deprived)	0.14	0.001 (0.007)	0.831	-	-	-	-0.88	-0.01 (0.021)	0.277

All models adjusted for child age (months). % inc or dec refers to exponentiated beta coefficient and reflect percent increase or decrease from reference group. Due to log transformation of the outcome, beta coefficients were exponentiated to improve interpretability. SE, standard error; P, p-value.

of household education, nor ADI and collection time of day in univariate analyses.

3.4.1. Multivariate

In multivariate analyses adjusting for child age, marital status, and household education, significant relationships between household poverty status and collection time of day

were maintained (Table 4; Model 3). Collection start times among deep poverty households were 2.41% significantly (marginal) later in the day compared to high income households (Table 4; $\beta = 0.024$; $p = 0.016$). When including ADI in multivariate analyses, marginal significant relationships between household poverty status and collection time of day were still maintained (Table 4; Model 4).

3.5. Physical activity

Physical activity refers to any rigorous physical activity for 20 or more minutes in the 12h prior to providing a saliva sample. In univariate analyses, significant increases were observed in the odds of physical activity with decreasing levels of poverty. Deep poverty households demonstrated 42% lower odds of physical activity within 12h of salivary sampling compared to high income households (Table 5; OR=0.58, 95% CI [0.44–0.77]; $p < 0.0125$). Despite a stepwise increase in odds of physical activity with lesser impoverished households, these households were still less likely to engage in physical activity relative to high income households, albeit not significantly.

In univariate analyses, lower levels of household education demonstrated a significantly lower odds of physical activity compared

to households with Graduate/Professional educations (Table 5; $p < 0.05$). To note, univariate relationships between household education (e.g., HS graduate and College Graduate) and physical activity were not significant after Bonferroni correction. There was a pattern of increasing odds of physical activity with higher education levels. Households with a less than HS education demonstrated a 43% reduced odds of physical activity 12h prior to salivary sampling compared to Graduate/professional households (OR=0.57, 95% CI [0.41–0.79]; $p < 0.0125$). Households with a HS graduate, Some College/Associate, or College education demonstrated a respective 26, 27, and 16% reduced odds of physical activity compared to the reference group (Table 5).

ADI was not significantly associated with physical activity in univariate analyses (Table 5).

TABLE 5 Univariate and multivariate multilevel logistic models of physical activity.

	Physical activity								
	Univariate			Multivariate					
				Model 5			Model 6		
	OR	95% CI	P	OR	95% CI	P	OR	95% CI	P
Intercept				0.13	[0.09, 0.18]	<0.001	0.12	[0.09, 0.18]	<0.001
Household marital status									
Intercept	0.12	[0.08, 0.16]	<0.001						
No	0.78	[0.68, 0.90]	0.001	0.88	[0.75, 1.04]	0.140	0.85	[0.72, 1.01]	0.073
Yes	Ref			Ref			Ref		
Household poverty status									
(Intercept)	0.12	[0.09, 0.17]	<0.001						
Deep poverty	0.58	[0.44, 0.77]	<0.001	0.71	[0.50, 0.99]	0.046	0.76	[0.54, 1.09]	0.134
Poverty	0.82	[0.62, 1.08]	0.156	0.95	[0.69, 1.30]	0.735	0.97	[0.70, 1.36]	0.876
Near poverty	0.83	[0.69, 1.01]	0.066	0.96	[0.77, 1.20]	0.711	0.98	[0.77, 1.23]	0.848
Mid income	0.89	[0.76, 1.05]	0.175	0.98	[0.82, 1.16]	0.793	0.99	[0.83, 1.19]	0.943
High income	Ref			Ref			Ref		
Household education									
(Intercept)	0.13	[0.09, 0.17]	<0.001						
Less than HS	0.57	[0.41, 0.79]	0.001	0.66	[0.42, 1.04]	0.075	0.65	[0.40, 1.04]	0.074
HS graduate	0.74	[0.59, 0.94]	0.013	1.01	[0.75, 1.35]	0.969	1.01	[0.75, 1.38]	0.927
Some college or associate	0.73	[0.62, 0.86]	<0.001	0.79	[0.64, 0.97]	0.022	0.78	[0.63, 0.97]	0.024
College graduate	0.84	[0.72, 0.99]	0.034	0.87	[0.73, 1.03]	0.096	0.88	[0.74, 1.04]	0.131
Graduate or professional	Ref			Ref			Ref		
Area deprivation index									
Quartile 1 (least deprived)	Ref			Ref			Ref		
(Intercept)	0.11	[0.08, 0.16]	<0.001	–	–	–			
Quartile 2	1.12	[0.93, 1.35]	0.227	–	–	–	1.23	[1.00, 1.50]	0.045
Quartile 3	0.83	[0.68, 1.01]	0.060	–	–	–	0.92	[0.73, 1.15]	0.443
Quartile 4 (most deprived)	0.84	[0.67, 1.04]	0.111	–	–	–	1.04	[0.80, 1.35]	0.779

OR, odds ratio; CI, confidence interval; P, p-value.

3.5.1. Multivariate

In multivariate analyses adjusting for household socioeconomic factors and ADI, relationships between household poverty status and odds of physical activity became fully attenuated (Table 5; Model 5 and Model 6).

Relationships between household education and odds of physical activity became partially attenuated. Only households with Some college/Associate education demonstrated 21% lower odds of physical activity (OR=0.79, 95% CI [0.64–0.97]; $p < 0.05$) within 12h of salivary sampling compared to households with Graduate/Professional educations (Table 5). This result however is not significant after Bonferroni correction.

Despite univariate non-significance between ADI and physical activity, a marginally significant relationship between ADI and physical activity emerged in multivariate analyses adjusting for

household marital status, household poverty status, and household education. An ADI in quartile 2 (e.g., moderately deprived neighborhood) was significantly associated with 1.23 higher odds of physical activity compared to an ADI in quartile 1 (least deprived) (OR=1.23, 95% CI [1.00–1.50]; $p < 0.05$). These results are not significant after Bonferroni correction.

3.6. Caffeine intake

Caffeine intake refers to the child’s self-report of any caffeinated beverage during the 12h prior to providing a saliva sample. In univariate analyses, significantly higher odds of caffeine intake was observed among lower levels of household poverty compared to high income households (Table 6; $p < 0.0125$). Deep poverty households

TABLE 6 Univariate and multivariate multilevel logistic models of caffeine Intake.

	Caffeine intake								
	Univariate			Multivariate					
				Model 7			Model 8		
	OR	95% CI	P	OR	95% CI	P	OR	95% CI	P
Intercept				0.04	[0.03, 0.05]	<0.001	0.04	[0.03, 0.05]	<0.001
Household marital status									
Intercept	1.10	[1.10, 1.10]	<0.001						
No	1.09	[1.09, 1.09]	<0.001	1.01	[0.83, 1.23]	0.908	1.00	[0.82, 1.23]	0.984
Yes	Ref			Ref			Ref		
Household poverty status									
(Intercept)	0.05	[0.04, 0.06]	<0.001						
Deep poverty	2.15	[1.62, 2.85]	<0.001	1.14	[0.80, 1.64]	0.466	1.15	[0.78, 1.67]	0.483
Poverty	1.95	[1.42, 2.68]	<0.001	1.05	[0.72, 1.53]	0.799	1.00	[0.67, 1.48]	0.990
Near poverty	1.86	[1.46, 2.36]	<0.001	1.20	[0.90, 1.58]	0.213	1.15	[0.86, 1.55]	0.348
Mid income	1.57	[1.26, 1.96]	<0.001	1.21	[0.95, 1.53]	0.119	1.18	[0.92, 1.51]	0.197
High income	Ref			Ref			Ref		
Household education									
(Intercept)	0.04	[0.03, 0.05]	<0.001						
Less than HS	2.88	[2.07, 4.01]	<0.001	2.49	[1.58, 3.91]	<0.001	2.21	[1.36, 3.57]	0.001
HS graduate	2.79	[2.14, 3.64]	<0.001	2.78	[1.98, 3.90]	<0.001	2.61	[1.83, 3.73]	<0.001
Some college or associate	2.33	[1.87, 2.90]	<0.001	2.11	[1.62, 2.75]	<0.001	1.94	[1.47, 2.55]	<0.001
College graduate	1.50	[1.18, 1.90]	<0.001	1.39	[1.08, 1.79]	0.010	1.40	[1.09, 1.81]	0.010
Graduate or professional	Ref			Ref			Ref		
Area deprivation index									
Quartile 1 (least deprived)	Ref			Ref			Ref		
(Intercept)	0.05	[0.04, 0.06]	<0.001	–	–	–			
Quartile 2	1.23	[0.94, 1.60]	0.129	–	–	–	0.99	[0.75, 1.31]	0.929
Quartile 3	1.59	[1.23, 2.05]	<0.001	–	–	–	1.09	[0.82, 1.45]	0.549
Quartile 4 (most deprived)	1.89	[1.45, 2.46]	<0.001	–	–	–	1.18	[0.87, 1.62]	0.293

OR, odds ratio; CI, confidence interval; P, p-value.

had a 2.15 higher odds of caffeine intake 12h prior to sampling compared to high income households (Table 6; OR=2.15, 95% CI [1.62–2.85]; $p < 0.0125$). This same pattern was observed among poverty (OR=1.95, 95% CI [1.42–2.68]; $p < 0.05$), near poverty (OR=1.86, 95% CI [1.46–2.36]; $p < 0.0125$), and mid income households (OR=1.57, 95% CI [1.26–1.96]; $p < 0.0125$) (Table 6).

Lower levels of household education demonstrated a significantly higher odds of caffeine intake compared to households with Graduate/Professional educations (Table 6; $p < 0.0125$). Households with a less than HS education demonstrated a 2.88 higher odds of caffeine intake 12h prior to salivary sampling compared to Graduate/professional households (OR=2.88, 95% CI [2.07–4.01]; $p < 0.0125$). There was a pattern of decreasing odds of caffeine intake with higher education levels. Households with a HS graduate or Some College/Associate education demonstrated a respective 2.79, 2.33, 1.50 higher odds of caffeine intake compared to the reference group (Table 6).

ADI was only significantly associated with caffeine intake in univariate analyses (Table 6). Residing in highly deprived neighborhoods (e.g., quartile 3 and 4) was significantly associated with a 1.59–1.89 ($p < 0.0125$) higher odds of caffeine intake compared to participants residing in the least deprived neighborhoods (quartile 1).

3.6.1. Multivariate

In multivariate analyses adjusting for household marital status, education, and ADI, relationships between household poverty status and odds of caffeine intake, as well as ADI and caffeine intake became fully attenuated (Model 7 and Model 8). However, significant relationships between household education and caffeine intake were maintained (Model 7 and Model 8).

4. Discussion

The findings from this study demonstrate significant associations between several key salivary methodological variables (time since waking, collection time of day, physical activity, and caffeine intake) with key socioeconomic factors (poverty status, household education, neighborhood deprivation). In general, lower levels of household poverty and education were significantly associated with salivary collection methodological variables (e.g., longer times since waking, collections later in the day, higher odds of caffeine consumption, and lower odds of physical activity). Furthermore, household socioeconomic context and neighborhood socioeconomic context were differentially associated with these variables. This indicates multiple sources of socioeconomic factors can independently introduce methodological biases when not fully standardized across data collection sites and individual participants. Together, present findings ultimately suggest that analyte levels measured from these samples may be impacted by non-random systematic methodological biases, particularly among analytes sensitive to variability in pH levels (e.g., caffeine in sample), physical activity/exercise, or circadian patterns. Leveraging this large salivary data set will require additional care when leveraging salivary analytes in future examination of early life antecedents of health inequities. Finally, only a subset of key socioeconomic factors and salivary sampling methodological variables were assessed in the present analyses, therefore other factors that drive health inequities

may impact additional salivary methodological variables in addition to those examined in this current study.

Household poverty status was consistently significantly associated with salivary methodological variables in univariate analyses, often when comparing highly impoverished households with lesser impoverished households. These relationships were maintained in multivariate analyses when specifically predicting time since waking and collection time of day. Significant relationships between household poverty status and physical activity and caffeine intake were attenuated in multivariate analyses when adjusting for household marital status, household education, or ADI. To our knowledge, no study has examined direct relationships between household poverty status and salivary collection variables among pediatric populations. Our measure of poverty status (e.g., household income as a function of household size) may reflect more proximal measures of material or economic goods that, when scarce in impoverished households, facilitate longer durations between waking and arriving to the laboratory to provide a saliva sample, as well as sampling later in the day. With this, it may be that a reduction in economic goods associated with an impoverished household leads to unique barriers preventing an early arrival to the study site shortly after waking and earlier in the day, thereby performing salivary collections in the “tail” of diurnal rhythms when levels are low. Also, later sampling times among participants from impoverished households may have been partially or fully driven by site-specific differences in access (e.g., differences in travel time and distance). Alternatively, given the semi-flexible experimental design of the cohort study, it is possible households in poverty self-selected for a later study start time over an earlier start time in anticipation of additional barriers, such as prioritizing employment responsibilities, geographical or transportation barriers, or responsibilities of other children without funds for additional childcare. Differential preferences to come into the laboratory on a weekday versus a weekend may be another contributing source to this variability and not investigated in the present analysis. Additionally, attenuated relationships with household poverty status predicting physical activity and caffeine intake after accounting for additional socioeconomic factors, such as household education or ADI, suggest that differences in likelihood of physical activity or caffeine intake may be partially attributed to a complex interaction between several socioeconomic constructs. It is possible that individual measures of SES may be less apt to capture differences compared to composite forms of SES that include income, education, and neighborhood characteristics (53, 88). While these are only some explanations, these differences in salivary sampling methodological variables may partially, yet falsely, drive future SES-related health inequities, or null findings, in observed salivary analyte levels that are sensitive to variability in sampling methodological variables.

Household education was not significantly associated with time since waking nor collection time of day but was significantly associated with physical activity and caffeine intake in univariate and multivariate analyses. Again, to our knowledge, no study has examined direct relationships between household education and on-site salivary collection methodological variables among adolescent populations. Even with this, Krieger et al. reported weak associations between education level and physical health status however only among those living below the poverty line (89). While this study was performed among adults and examined health status, this partially supports our non-significant findings between household education and time since

waking or collection time of day. In addition, relationships in our study between household education and physical activity were only significant when comparing households with Some College/Associate education to households with a Graduate/Professional education and adjusting for household poverty status. These findings are also in line with those of Krieger et al. where level of education operates on health differentially by poverty status (89). Nonetheless, this evidence may explain why household education was sparsely related to salivary collection variables. The inclusion of both household education and household poverty status in the same statistical models potentiates confounding, given evidence of strong positive correlations between one's education level and income (88). However, we checked variance inflation factor (VIF) values for these models, and all were below 2.09, indicating that these variables were not redundant in predicting the outcomes in this study sample.

When examining neighborhood socioeconomic contexts, significant relationships were observed with ADI when predicting multivariate odds of physical activity and univariate odds of caffeine intake, whereas ADI was not significantly associated with time since waking nor collection time of day. Cerin et al. demonstrated complex relationships between environmental factors and individual-level or household-level factors (e.g., household income and education) that impact participation in physical activity (90). Differences in performing moderate to vigorous physical activity due to area-level socioeconomic factors were significantly mediated by several individual-level factors (e.g., social support from friends and self-efficacy), but not significantly mediated by infrastructure nor area-level crime (90). While ADI is a well-validated measure of neighborhood-level socioeconomic context, there are other ways to assess this construct beyond the current version (91, 92) that may miss key characteristics that are important for understanding childhood origins of health inequities. The measure of ADI used in this study is a composite of multiple forms of area economic and resource deprivation. This indicates that relationships between area-level SES and physical activity may be partially explained by individual-level factors not recorded as part of this study. While limited in the ability to inform individual-level patterns (e.g., due to ecological fallacy), this ADI measure includes factors of basic resources (e.g., plumbing, telephone) that would not be captured by income and education alone.

4.1. Strengths and limitations

Despite evidence for potential non-random systematic bias in salivary sampling methodological variables in the present cohort study, several strengths of the study design were observed. First, the ABCD Study© achieved coordination among 21 sites for the successful self-collection of saliva among a large pediatric cohort repeated annually. This strength adds to both the salience of the observed findings in this nationally representative pediatric study sample and further highlights the utility of salivary bioscience research on large scales and with pediatric populations. Second, the cohort sample of children was successfully recruited from the general population, rather than a convenience sample among those presenting to a clinical site, thus adding to the heterogeneity of the cohort sample, and thereby increasing the external validity of the present findings for future large-scale salivary collections. Additionally, uncovering socioecological relationships using data obtained in a non-invasive way means that salivary biosciences are well-suited to understand

public health issues, particularly among children from families underrepresented in research (93). Salivary methodological variables examined in this study are often applicable to other forms of biological sample collection measuring acutely fluctuating levels (e.g., blood, urine) for analytes that vary across time of day yet correlate with salivary levels (94–98). Thus, our results may have increased generalizability beyond saliva in this study and may occur in other large biomedical research studies. Other biological methods measuring chronic levels would not be impacted by these methodological variations (e.g., hair, nails, teeth).

Nonetheless, there are several limitations to the current analyses. First, part of the exclusion criteria for the current analytical sample was a mismatch between parental report of “biological sex at birth” and the participant endorsed a binary “biological sex/gender” at the time of saliva collection from baseline (e.g., current analyses). Unfortunately, given that sex at birth determined the hormone panel for testing prior to Year 3, this protocol misrepresents associations between estradiol and variants of male sex or gender expression by not assaying saliva samples for the assumed “female” hormone. This experimental strategy potentially excludes important dynamics in gender identification throughout pubertal maturation (99) and may limit our ability to fully understand how hormones emerge across a diversity of gender identities in the current data set. In year 3, ABCD protocol solicited the participant's endorsement of any gender identity at saliva collection however this is not part of the Release 3.0 dataset used in these analyses. Additional gender identity specific assessments were added to the study at this Year 3 timepoint as well. After Year 3, biological males at birth endorsing a male gender identity were assessed for testosterone and DHEA only, and all other possible combinations of gender identity endorsement (including neither gender) were assessed for testosterone, DHEA, and estradiol. Future analyses using the ABCD dataset for year 3 and later should leverage the gender identity data that better capture the dynamics of gender identification with salivary hormones. Second, there are many ways to capture socioeconomic status (SES), including measures of employment or unemployment status, wealth, type or status of occupation, or numeric income level (100). The variables used in this study are mostly reflective of household economic resources and household education. Previous evidence indicates that education and poverty status represent just two of many overlapping yet distinct dimensions comprising SES, rather than being entirely reflective of SES (53). Given that SES is a dynamic, multi-dimensional construct, the exclusion of other aspects of SES may only provide a partial understanding of socioecological relationships on salivary collection methodological variables.

Another limitation is the relative difference in smaller sample size among the deep poverty and poverty groups compared to the high-income group, given that larger sample sizes are more statistically powered to detect small effect sizes. Thus, imbalances in sample sizes can bias the findings of smaller effect sizes between groups, especially where the comparison group (e.g., deep poverty or poverty) is a smaller sample size relative to the reference group (e.g., high income). The deep poverty and poverty groups are likely underpowered to detect small effects and are the most at risk for null findings. Null findings between deep poverty and poverty with salivary methodological variables in the present study should be interpreted with caution. However, the deep poverty and poverty sample size were $n = 798$ and $n = 616$, respectively, which is relatively robust for pediatric biomedical research. In addition, for many of the observed findings,

the effect sizes of the significant results in this analysis are relatively moderate to small. These results may not be observable within studies with smaller sample sizes, as sample sizes may be underpowered to detect small effect sizes. Without being contextualized to specific analytes of interest, the practical application of current findings is limited.

In addition, an area-level measure such as ADI is subject to an ecological fallacy because aggregate-level patterns may not actually reflect individual-level socioeconomic measures (53, 101). Although we leverage multi-level models accounting for participant clustering by study site, we observed different relationships to salivary collection methodological variables between household income/education and ADI. One potential explanation as to why ADI was not related to time-dependent salivary collection variables is that ADI may not be as proximal to household level factors, and thus would not reflect direct relationships to time since waking or collection time of day.

Another limitation of this analyses is the focus on salivary bioscience methodological variables only. The observed relationships discovered in the present analyses were not examined further in relation to specific salivary analytes that have been assayed in the samples (e.g., DHEA, testosterone, estradiol). Associations of socioeconomic-based differences in salivary collection methodological variables with salivary analyte levels were not directly tested in the present analysis. Further, our examination of baseline relationships may also limit interpretability over time, especially with longitudinal changes in SES for a participant, changes in salivary methodological variables (e.g., sampling at different time of the day or different physical activity/caffeine intake habits as participants age), and even longitudinal changes in analyte levels. Given the breadth of research questions and corresponding analytical approaches with this dataset, associations between methodological biases and analyte levels could vary across independent and longitudinal investigations. There are important considerations for whether these relationships are stable over time. We rely on existing literature that points to interference of accurate analyte measurement due to collection methodological variables (11, 29, 30, 35–37, 39, 40). Rather, this analysis encourages researchers examining health inequities to conduct a thorough examination of salivary collection methods prior to leveraging analyte levels.

While we observed significant relationships between socioeconomic and salivary sampling methodological variables, we cannot make conclusions about magnitude and directionality of relationships to specific analytes. Based on previous literature of neuroendocrine circadian patterns (30–34), we predict that these differences in salivary sampling methodological variables will become more problematic as participants continue to mature, as circadian patterns become more pronounced with maturation, and differences in exercise and caffeine intake may grow with age as a function of key socioeconomic factors. However, not all salivary analytes demonstrate a circadian rhythm or are sensitive to changes in pH of the sample, or physical activity. Thus, some specific analytes may be relatively unaffected by the observations discovered in these analyses. Researchers should evaluate whether their salivary analytes of interest reflect the observed patterns in their own analyses, and if so, intentionally address them in analyses and interpretation of salivary analyte results.

Examination of socioeconomic factors with other salivary sampling methodological variables that were collected in ABCD were out of scope for the current analyses, including cotinine levels from

first and/or second-hand tobacco exposure in the children and medications that may alter salivary flow rates. However, future studies of bio-banked salivary samples could measure cotinine directly from the sample to statistically control for these confounders. There are additional salivary sampling methodological considerations that were not fully collected in this large data set, such as participant reported factors of the oral environment (e.g., blood from sores, lost teeth, injury). Research assistants used a 5-point scale to document visible alterations in the saliva sample, including presence of discoloration from food dye or blood, and food particles. A visual inspection of the salivary sample was also conducted by professional laboratory staff during the time of assaying (e.g., Salimetrics, Carlsbad, CA, United States) to note any abnormalities in the sample. Future studies would benefit from a thorough oral health questionnaire at the time of sample collection to account for salivary sample contamination. To achieve the most rigorous use of salivary analytes, all of these methodological factors should be controlled for either through upfront experimental design in future studies, expansion of questionnaires on oral health, or with careful and intentional statistical analyses to fully understand how socioeconomic factors may drive experimental noise and interfere with results. This includes maintaining strict protocols for saliva sampling regarding time since waking, time of day, sample collection duration, abstaining from caffeine, smoking, and rigorous physical activity 12h prior to sampling.

Lastly, the examination of race/ethnicity differences was outside of the scope of this analysis; however, we encourage investigators to consider intentionally integrating upstream measures when investigating research questions pertaining to racial and ethnic minoritized groups (102–104). For example, structural racism has been identified as an important factor of adverse health among racial and ethnic minoritized groups including adolescents (105–107). Future salivary bioscience research studies must acknowledge root causes of racial/ethnic differences in health, and should be integrated in salivary bioscience research when examining race/ethnicity particularly through collaboration with experts in structural racism.

4.2. Conclusion

Significant associations were observed between socioeconomic factors and salivary collection methodological variables. Specifically, lower levels of household poverty and education were significantly associated with more sources of potential bias in salivary collection methodological variables (e.g., longer times since waking, collections later in the day, higher odds of caffeine consumption, and lower odds of physical activity). These novel findings serve as a thorough cautionary tale for future analyses leveraging analyte levels from these salivary samples to examine early antecedents of health inequities, as results may reflect variations in methodological variables of salivary collections (e.g., time since waking to sampling, time of day of sampling, physical activity, and caffeine intake) and not actual biological mechanisms. Entangled contributions to biological functioning from socioeconomic factors remain a potential source of non-random systematic biases. Conclusions made about biological functioning using saliva while only accounting for salivary collection methodological variables, without the consideration of socioeconomic factors, may erroneously attribute group differences to differences in biological functioning rather than the broader upstream socioeconomic environment.

These results advance salivary bioscience research by applying a health equity perspective in considering socioeconomic factors on standardizing salivary methodology. These findings highlight the importance of developing an experimental design that standardizes salivary collections, to prevent potential unintentional non-random systematic biases in saliva sampling methodology. Specifically, our results suggest that future studies ensure participants self-collect at the same time of day, for the same collection duration, and in the absence of rigorous physical activity or caffeine consumption 12h prior to providing a sample. If stringent sample collection protocols are not feasible, we recommend that future studies collect information on potentially important salivary methodological variables (e.g., time since waking, collection time of day, physical activity, caffeine intake, oral health, medications), utilize post-hoc statistical techniques (e.g., adjustment) to cautiously disentangle effects, and target analytes that are robust to variability in salivary methodological variables. Nonetheless, salivary samples were collected effectively in participants across 21 sites, demonstrating feasibility of guided self-sampling as a non-invasive biological specimen in a large-scale pediatric study. These samples have strong potential to be leveraged in investigations of biological mechanisms across the entire sample, yet more cautiously when leveraging factors in analyses that drive health inequities.

Data availability statement

Publicly available datasets were analyzed in this study. This data can be found in the NIMH Data Archive (NDA), Adolescent Brain Cognitive Development (ABCD) Study (<http://dx.doi.org/10.15154/1519007>).

Ethics statement

Ethical review and approval was not required for the study on human participants in accordance with the local legislation and institutional requirements. Written informed consent to participate in this study was provided by the participants' legal guardian/next of kin.

Author contributions

HM and KU contributed to the design of the study. HM led analyses and writing. KU contributed to analyses and writing. All authors contributed to the article and approved the submitted version.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

- Chen E, Martin AD, Matthews KA. Understanding health disparities: the role of race and socioeconomic status in children's health. *Am J Public Health.* (2006) 96:702–8. doi: 10.2105/AJPH.2004.048124
- Devenish B, Hooley M, Mellor D. The pathways between socioeconomic status and adolescent outcomes: a systematic review. *Am J Community Psychol.* (2017) 59:219–38. doi: 10.1002/ajcp.12115
- Hanson MD, Chen E. Socioeconomic status and health behaviors in adolescence: a review of the literature. *J Behav Med.* (2007) 30:263–85. doi: 10.1007/s10865-007-9098-3
- Condon EM. Chronic stress in children and adolescents: a review of biomarkers for use in pediatric research. *Biol Res Nurs.* (2018) 20:473–96. doi: 10.1177/1099800418779214

5. Granger DA, Fortunato CK, Beltzer EK, Virag M, Bright MA, Out D. Focus on methodology: salivary bioscience and research on adolescence: an integrated perspective. *J Adolesc.* (2012) 35:1081–95. doi: 10.1016/j.adolescence.2012.01.005
6. Granger DA, Kivlighan KT, El-Sheikh M, Gordis EB, Stroud LR. Salivary α -amylase in biobehavioral research. *Annu Rev Public Health.* (2007) 1098:122–44. doi: 10.1196/annals.1384.008
7. Kokka I, Chrousos GP, Darviri C, Bacopoulou F. Measuring adolescent chronic stress: a review of established biomarkers and psychometric instruments. *Horm Res Paediatr.* (2023) 96:74–82. doi: 10.1159/000522387
8. Hoyt MA, Granger DA. Salivary bioscience and the future of behavioral medicine. *Int J Behav Med.* (2020) 27:257–61. doi: 10.1007/s12529-020-09900-w
9. Granger DA, Kivlighan KT, Fortunato C, Harmon AG, Hibel LC, Schwartz EB, et al. Integration of salivary biomarkers into developmental and behaviorally-oriented research: problems and solutions for collecting specimens. *Physiol Behav.* (2007) 92:583–90. doi: 10.1016/j.physbeh.2007.05.004
10. Granger DA, Taylor MK. Foundations of interdisciplinary salivary bioscience: An Introduction In: *Salivary Bioscience: Foundations of Interdisciplinary Saliva Research and Applications* (2020) (Springer), 3–9.
11. Smyth N, Hucklebridge F, Thorn L, Evans P, Clow A. Salivary cortisol as a biomarker in social science research. *Soc Pers Psychol Compass.* (2013) 7:605–25. doi: 10.1111/spc3.12057
12. Granger DA, Johnson SB, Szanton SL, Out D, Schumann LL. Incorporating salivary biomarkers into nursing research: an overview and review of best practices. *Biol Res Nurs.* (2012b) 14:347–56. doi: 10.1177/1099800412443892
13. Ramirez J, Elmofy M, Castillo E, DeRouen M, Shariff-Marco S, Allen L, et al. Evaluation of cortisol and telomere length measurements in ethnically diverse women with breast cancer using culturally sensitive methods. *J Community Genet.* (2017) 8:75–86. doi: 10.1007/s12687-016-0288-y
14. Wilde C, Out D, Johnson S, Granger DA. Sample collection, including participant preparation and sample handling In: *The Immunoassay Handbook* (2013). ed. D. Wild (Elsevier), 427–440.
15. Johnson SB, Voegtline KM. Salivary bioscience and pediatrics In: *Salivary Bioscience: Foundations of Interdisciplinary Saliva Research and Applications* (2020) (Springer), 643–646.
16. Parkin GM, McCarthy MJ, Thein SH, Piccerillo HL, Warikoo N, Granger DA, et al. Saliva testing as a means to monitor therapeutic lithium levels in patients with psychiatric disorders: identification of clinical and environmental covariates, and their incorporation into a prediction model. *Bipolar Disord.* (2021) 23:679–88. doi: 10.1111/bdi.13128
17. Granger D. A., Hamilton K. (2020). Saliva as a biospecimen: then and now in the era of COVID-19. Association for Psychological Science (APS) Observer 33. Available at: <https://www.psychologicalscience.org/observer/saliva-as-a-biospecimen-then-and-now-in-the-era-of-covid-19> (Accessed April 19, 2023).
18. Fernandes A, Skinner ML, Woelfel T, Carpenter T, Haggerty KP. Implementing self-collection of biological specimens with a diverse sample. *Field Methods.* (2013) 25:58–73. doi: 10.1177/1525822X12453526
19. Arbour L, Cook D. DNA on loan: issues to consider when carrying Out genetic research with aboriginal families and communities. *Public Health Genom.* (2006) 9:153–60. doi: 10.1159/000092651
20. Rajakumar K, Thomas SB, Musa D, Almario D, Garza MA. Racial differences in parents' distrust of medicine and research. *Arch Pediatr Adolesc Med.* (2009) 163:108–14. doi: 10.1001/archpediatrics.2008.521
21. Scharff DP, Mathews KJ, Jackson P, Hoffsuemmer J, Martin E, Edwards D. More than Tuskegee: understanding mistrust about research participation. *J Health Care Poor Underserved.* (2010) 21:879–97. doi: 10.1353/hpu.0.0323
22. George S, Duran N, Norris K. A systematic review of barriers and facilitators to minority research participation among African Americans, Latinos, Asian Americans, and Pacific islanders. *Am J Public Health.* (2014) 104:e16–31. doi: 10.2105/AJPH.2013.301706
23. Condon EM, Londono Tobon A, Mayes LC, Sadler LS. Acceptability and feasibility of hair and salivary biomarker collection among multiethnic school-age children. *Matern Child Health J.* (2020) 24:865–74. doi: 10.1007/s10995-020-02926-2
24. Ford JL, Boch SJ, McCarthy DO. Feasibility of Hair collection for cortisol measurement in population research on adolescent health. *Nurs Res.* (2016) 65:249–55. doi: 10.1097/NNR.0000000000000154
25. Padilla GA, Calvi JL, Taylor MK, Granger DA. Saliva collection, handling, transport, and storage: special considerations and best practices for interdisciplinary salivary bioscience research In: *Salivary Bioscience: Foundations of Interdisciplinary Saliva Research and Applications* (Springer) (2020). ed. D. Wild (Elsevier), 23–24.
26. Henson BS, Wong DT. Collection, storage, and processing of saliva samples for downstream molecular applications In: editors. *Oral biology: methods in molecular biology*. Eds. GJ Seymour, M Cullinan and N Heng. Totowa, NJ: Humana Press (2010). 21–30.
27. Bellagambi FG, Lomonaco T, Salvo P, Vivaldi F, Hangouët M, Ghimenti S, et al. Saliva sampling: methods and devices. An overview. *TrAC Trends Anal Chem.* (2020) 124:115781. doi: 10.1016/j.trac.2019.115781
28. Riis JL, Bryce CI, Matin MJ, Stebbins JL, Kornienko O, van Huisstede L, et al. The validity, stability, and utility of measuring uric acid in saliva. *Biomark. Med.* (2018) 12:583–96. doi: 10.2217/bmm-2017-0336
29. Navazesh M. Methods for collecting saliva. *Ann N Y Acad Sci.* (1993) 694:72–7. doi: 10.1111/j.1749-6632.1993.tb18343.x
30. Blume C, Garbaza C, Spitschan M. Effects of light on human circadian rhythms, sleep and mood. *Somnologie.* (2019) 23:147–56. doi: 10.1007/s11818-019-00215-x
31. Granger DA, Schwartz EB, Booth A, Curran M, Zakaria D. Assessing dehydroepiandrosterone in saliva: a simple radioimmunoassay for use in studies of children, adolescents and adults. *Psychoneuroendocrinology.* (1999) 24:567–79. doi: 10.1016/S0306-4530(99)00013-X
32. Hucklebridge F, Hussain T, Evans P, Clow A. The diurnal patterns of the adrenal steroids cortisol and dehydroepiandrosterone (DHEA) in relation to awakening. *Psychoneuroendocrinology.* (2005) 30:51–7. doi: 10.1016/j.psyneuen.2004.04.007
33. Matchock RL, Dorn LD, Susman EJ. Diurnal and seasonal cortisol, testosterone, and DHEA rhythms in boys and girls during puberty. *Chronobiol Int.* (2007) 24:969–90. doi: 10.1080/07420520701649471
34. Whetzel CA, Klein LC. Measuring DHEA-S in saliva: time of day differences and positive correlations between two different types of collection methods. *BMC Res Notes.* (2010) 3:1–5. doi: 10.1186/1756-0500-3-204
35. di Luigi L, Guidetti L, Baldari C, Gallotta MC, Sgrò P, Perroni F, et al. Cortisol, dehydroepiandrosterone sulphate and dehydroepiandrosterone sulphate/cortisol ratio responses to physical stress in males are influenced by pubertal development. *J Endocrinol Investig.* (2006) 29:796–804. doi: 10.1007/BF03347373
36. Lennartsson AK, Kushnir MM, Bergquist J, Jonsdottir IH. DHEA and DHEA-S response to acute psychosocial stress in healthy men and women. *Biol Psychol.* (2012) 90:143–9. doi: 10.1016/j.biopsycho.2012.03.003
37. Mastorakos G, Pavlatou M, Diamanti-Kandarakis E, Chrousos GP. Exercise and the stress system. *Hormones.* (2005) 4:73–89.
38. Herting MM, Uban KA, Gonzalez MR, Baker FC, Kan EC, Thompson WK, et al. Correspondence between perceived pubertal development and hormone levels in 9-10 year-olds from the adolescent brain cognitive development study. *Front Endocrinol.* (2021) 11:549928. doi: 10.3389/fendo.2020.549928
39. Schwartz EB, Granger DA, Susman EJ, Gunnar MR, Laird B. Assessing salivary cortisol in studies of child development. *Child Dev.* (1998) 69:1503–13. doi: 10.1111/j.1467-8624.1998.tb06173.x
40. Klein LC, Bennett JM, Whetzel CA, Granger DA, Ritter FE. Caffeine and stress alter salivary α -amylase activity in young men. *Hum Psychopharmacol.* (2010) 25:359–67. doi: 10.1002/hup.1126
41. de Jong WHA, Eisenhofer G, Post WJ, Muskiet FAJ, de Vries EGE, Kema IP. Dietary influences on plasma and urinary metanephrines: implications for diagnosis of catecholamine-producing tumors. *J Clin Endocr Metab.* (2009) 94:2841–9. doi: 10.1210/jc.2009-0303
42. Deuschbein T, Unger N, Jaeger A, Broecker-Preuss M, Mann K, Petersenn S. Influence of various confounding variables and storage conditions on metanephrine and normetanephrine levels in plasma. *Clin Endocrinol.* (2010) 73:153–60. doi: 10.1111/j.1365-2265.2009.03761.x
43. Robertson D, Frölich JC, Carr RK, Watson JT, Hollifield JW, Shand DG, et al. Effects of caffeine on plasma renin activity, catecholamines and blood pressure. *N Engl J Med.* (1978) 298:181–6. doi: 10.1056/NEJM197801262980403
44. Garavan H, Bartsch H, Conway K, Decastro A, Goldstein RZ, Heeringa S, et al. Recruiting the ABCD sample: design considerations and procedures. *Dev Cogn Neurosci.* (2018) 32:16–22. doi: 10.1016/j.dcn.2018.04.004
45. Adler N, Ostrove J. Socioeconomic status and health: what we know and what we don't. *Ann N Y Acad Sci.* (1999) 896:3–15. doi: 10.1111/j.1749-6632.1999.tb08101.x
46. Brito NH, Noble KG. Socioeconomic status and structural brain development. *Front Neurosci.* (2014) 8:276. doi: 10.3389/fnins.2014.00276
47. Chaudry A, Wimer C. Poverty is not just an Indicator: the relationship between income, poverty, and child well-being. *Acad Pediatr.* (2016) 16:S23–9. doi: 10.1016/j.acap.2015.12.010
48. Hair NL, Hanson JL, Wolfe BL, Pollak SD. Association of child poverty, brain development, and academic achievement. *JAMA Pediatr.* (2015) 169:822–9. doi: 10.1001/jamapediatrics.2015.1475
49. Hughes M, Tucker W. Poverty as an adverse childhood experience. *N C Med J.* (2018) 79:124–6. doi: 10.18043/ncm.79.2.124
50. Roos LL, Wall-Wieler E, Lee JB. Poverty and early childhood outcomes. *Pediatrics.* (2019) 143:e20183426. doi: 10.1542/peds.2018-3426
51. Chen E, Cohen S, Miller GE. How low socioeconomic status affects 2-year hormonal trajectories in children. *Psychol Sci.* (2010) 21:31–7. doi: 10.1177/0956797609355566
52. Lupien SJ, King S, Meaney MJ, McEwen BS. Can poverty get under your skin? Basal cortisol levels and cognitive function in children from low and high socioeconomic status. *Dev Psychopathol.* (2001) 13:653–76. doi: 10.1017/S0954579401003133
53. Krieger N, Williams DR, Moss NE. Measuring social class in us public health research: concepts, methodologies, and guidelines. *Annu Rev Public Health.* (1997) 18:341–78. doi: 10.1146/annurev.publhealth.18.1.341
54. Ford ES, Merritt RK, Heath GW, Powell KE, Washburn RA, Kriska A, et al. Physical activity behaviors in lower and higher socioeconomic status populations. *Am J Epidemiol.* (1991) 133:1246–56. doi: 10.1093/oxfordjournals.aje.a115836

55. Stalsberg R, Pedersen AV. Effects of socioeconomic status on the physical activity in adolescents: a systematic review of the evidence. *Scand J Med Sci Sports*. (2010) 20:368–83. doi: 10.1111/j.1600-0838.2009.01047.x
56. Branum AM, Rossen LM, Schoendorf KC. Trends in caffeine intake among US children and adolescents. *Pediatrics*. (2014) 133:386–93. doi: 10.1542/peds.2013-2877
57. Eicher-Miller HA, Boushey CJ, Bailey RL, Yang YJ. Frequently consumed foods and energy contributions among food secure and insecure U.S. children and adolescents. *Nutrients*. (2020) 12:304. doi: 10.3390/nu12020304
58. Uban KA, Horton MK, Jacobus J, Heyser C, Thompson WK, Tapert SF, et al. Biospecimens and the ABCD study: rationale, methods of collection, measurement and early data. *Dev Cogn Neurosci*. (2018) 32:97–106. doi: 10.1016/j.dcn.2018.03.005
59. Salimetrics LLC. *Salimetrics supports adolescent brain cognitive development (ABCD) study with salivary bioscience* Salimetrics (2016) Available at: https://www.prweb.com/releases/abcd_study/salivary_biomarkers/prweb13634957.htm.
60. Spilsbury JC, Storfer-Isser A, Drotar D, Rosen CL, Kirchner LH, Benham H, et al. Sleep behavior in an urban US sample of school-aged children. *Arch Pediatr Adolesc Med*. (2004) 158:988–94. doi: 10.1001/archpedi.158.10.988
61. Thorleifsdottir B, Björnsson JK, Benediktsdottir B, Gislason T, Kristbjarnarson H. Sleep and sleep habits from childhood to young adulthood over a 10-year period. *J Psychosom Res*. (2002) 53:529–37. doi: 10.1016/S0022-3999(02)00444-0
62. Centers for Disease Control and Prevention (CDC) (2003). Physical activity levels among children aged 9–13 years—United States, 2002. *MMWR* 52:785–88.
63. Craggs C, Corder K, van Sluijs EMF, Griffin SJ. Determinants of change in physical activity in children and adolescents: a systematic review. *Am J Prev Med*. (2011) 40:645–58. doi: 10.1016/j.amepre.2011.02.025
64. Nader PR, Bradley RH, Houts RM, McRitchie SL, O'Brien M. Moderate-to-vigorous physical activity from ages 9 to 15 years. *JAMA*. (2008) 300:295–305. doi: 10.1001/jama.300.3.295
65. Ellison RC, Singer MR, Moore LL, Nguyen U-SDT, Garrahe EJ, Marmor JK. Current caffeine intake of young children: amount and sources. *J Am Diet Assoc*. (1995) 95:802–4. doi: 10.1016/S0002-8223(95)00222-7
66. Morgan KJ, Stults VJ, Zabik ME. Amount and dietary sources of caffeine and saccharin intake by individuals ages 5 to 18 years. *Regul Toxicol Pharmacol*. (1982) 2:296–307. doi: 10.1016/0273-2300(82)90003-4
67. Shafer H. L., Edin K. (2012). Extreme poverty in the United States, 1996 to 2011. Available at: https://npc.umich.edu/publications/policy_briefs/brief28/policybrief28.pdf.
68. Davis RP, Williams WR. Bringing psychologists to the fight against deep poverty. *Am Psychol*. (2020) 75:655–67. doi: 10.1037/amp0000650
69. Ekono M, Jiang Y, Smith S. Young children in deep poverty. New York, NY: National Center for Children in Poverty (NCCP) (2016) Available at: <https://www.nccp.org/publication/young-children-in-deep-poverty/>
70. Nguyen US, Smith S, Granja MR, Street B. Young children in deep poverty: racial/ethnic disparities and child well-being compared to other income groups. New York, NY: National Center for Children in Poverty (NCCP) (2020) Available at: <https://www.nccp.org/publication/young-children-in-deep-poverty-racial-ethnic-disparities-and-child-well-being-compared-to-other-income-groups/>
71. Goodman E. The role of socioeconomic status gradients in explaining differences in US adolescents' health. *Am J Public Health*. (1999) 89:1522–8. doi: 10.2105/AJPH.89.10.1522
72. Braveman P, Cubbin C, Marchi K, Egerter S, Chavez G. Measuring socioeconomic status/position of in studies racial/ethnic disparities: maternal and infant health. *Public Health Rep*. (2001) 116:449–63. doi: 10.1093/phr/116.5.449
73. Brooks H. (2021). Exploring a nonbinary approach to health. NICHQ - National Institute for Children's Health Quality. Available at: <https://www.nichq.org/insight/exploring-nonbinary-approach-health> (Accessed April 13, 2023).
74. Kind AJH, Jencks S, Brock J, Yu M, Bartels C, Ehlenbach W, et al. Neighborhood socioeconomic disadvantage and 30-day rehospitalization: a retrospective cohort study. *Ann Intern Med*. (2014) 161:765–74. doi: 10.7326/M13-2946
75. Singh GK. Area deprivation and widening inequalities in US mortality, 1969–1998. *Am J Public Health*. (2003) 93:1137–43. doi: 10.2105/ajph.93.7.1137
76. Hooper A, Hustedt JT, Slicker G, Hallam RA, Gaviria-Loaiza J, Vu JA, et al. Area deprivation index as a predictor of economic risk and social and neighborhood perceptions among families enrolled in early head start. *Child Youth Serv Rev*. (2022) 137:106468. doi: 10.1016/j.chilcyouth.2022.106468
77. Oates G, Rutland S, Juarez L, Friedman A, Schechter MS. The association of area deprivation and state child health with respiratory outcomes of pediatric patients with cystic fibrosis in the United States. *Pediatr Pulmonol*. (2021) 56:883–90. doi: 10.1002/ppul.25192
78. Wall LB, Wright M, Samora J, Bae DS, Steinman S, Goldfarb CA. Social deprivation and congenital upper extremity differences—an assessment using PROMIS. *J Hand Surg Am*. (2021) 46:114–8. doi: 10.1016/j.jhsa.2020.08.017
79. Adise S, Marshall AT, Kan E, Gonzalez MR, Sowell ER. Relating neighborhood deprivation to childhood obesity in the ABCD study: evidence for theories of neuroinflammation and neuronal stress. *Health Psychol Adv*. (2022). Advance online publication. doi: 10.1037/hea0001250
80. Dennis E, Manza P, Volkow ND. Socioeconomic status, BMI, and brain development in children. *Transl Psychiatry*. (2022) 12:33–10. doi: 10.1038/s41398-022-01779-3
81. Mullins TS, Campbell EM, Hogeveen J. Neighborhood deprivation shapes motivational-neurocircuit recruitment in children. *Psychol Sci*. (2020) 31:881–9. doi: 10.1177/0956797620929299
82. Barch DM, Albaugh MD, Avenevoli S, Chang L, Clark DB, Glantz MD, et al. Demographic, physical and mental health assessments in the adolescent brain and cognitive development study: rationale and description. *Dev Cogn Neurosci*. (2018) 32:55–66. doi: 10.1016/j.dcn.2017.10.010
83. Pinheiro J, Bates D., Deb Roy S., Sarkar D.R Core Team (2022). nlme: linear and nonlinear mixed effects models. R package version 3.1-162. Available at: <https://CRAN.R-project.org/package=nlme>.
84. Fox J, Weisberg S. *An R companion to applied regression*. 3rd ed. Thousand Oaks, CA: SAGE Publications Inc. (2019).
85. Lefcheck JS. Piecewise SEM: piecewise structural equation modeling in R for ecology, evolution, and systematics. *Methods Ecol Evol*. (2016) 7:573–9. doi: 10.1111/2041-210X.12512
86. Grolemond G, Wickham H. Dates and times made easy with lubridate. *J Stat Softw*. (2011) 40:1–25. doi: 10.18637/jss.v040.i03
87. Harrell F. E. Jr. (2020). *Hmisc: harrell miscellaneous*. Available at: <https://CRAN.R-project.org/package=Hmisc>.
88. Liberatos P, Link BG, Kelsey JL. The measurement of social class in epidemiology. *Epidemiol Rev*. (1988) 10:87–121. doi: 10.1093/oxfordjournals.epirev.a036030
89. Krieger N, Fee E. Social class: the missing link in U.S. health data. *Int J Health Serv*. (1994) 24:25–44. doi: 10.2190/2JG7-YMD5-WCP2-XXNT
90. Cerin E, Leslie E. How socio-economic status contributes to participation in leisure-time physical activity. *Soc Sci Med*. (2008) 66:2596–609. doi: 10.1016/j.socscimed.2008.02.012
91. Fu M, Exeter DJ, Anderson A. So, is that your 'relative' or mine? A political-ecological critique of census-based area deprivation indices. *Soc Sci Med*. (2015) 142:27–36. doi: 10.1016/j.socscimed.2015.07.036
92. Trinidad S, Brokamp C, Mor Huertas A, Beck AF, Riley CL, Rasnick E, et al. Use of area-based socioeconomic deprivation indices: a scoping review and qualitative analysis. *Health Aff*. (2022) 41:1804–11. doi: 10.1377/hlthaff.2022.00482
93. Pellicchia M, Nuske HJ, Straiton D, McGhee Hassrick E, Gulsrud A, Iadarola S, et al. Strategies to engage underrepresented parents in child intervention services: a review of effectiveness and co-occurring use. *J Child Fam Stud*. (2018) 27:3141–54. doi: 10.1007/s10826-018-1144-Y/TABLES/6
94. Choe JK, Khan-Dawood FS, Dawood MY. Progesterone and estradiol in the saliva and plasma during the menstrual cycle. *Am J Obstet Gynecol*. (1983) 147:557–62. doi: 10.1016/0002-9378(83)90016-9
95. Ellison PT, Lipson SF. Salivary estradiol—a viable alternative? *Fertil Steril*. (1999) 72:951–2. doi: 10.1016/s0015-0282(99)00344-1
96. Rilling JK, Worthman CM, Campbell BC, Stallings JF, Mbizva M. Ratios of plasma and salivary testosterone throughout puberty: production versus bioavailability. *Steroids*. (1996) 61:374–8. doi: 10.1016/0039-128x(96)00043-8
97. Shirtcliff EA, Granger DA, Schwartz EB, Curran MJ, Booth A, Overman WH. Assessing estradiol in biobehavioral studies using saliva and blood spots: simple radioimmunoassay protocols, reliability, and comparative validity. *Horm Behav*. (2000) 38:137–47. doi: 10.1006/hbeh.2000.1614
98. Vining RF, McGinley RA, Symons RG. Hormones in saliva: mode of entry and consequent implications for clinical interpretation. *Clin Chem*. (1983) 29:1752–6. doi: 10.1093/clinchem/29.10.1752
99. Diamond LM. Gender fluidity and nonbinary gender identities among children and adolescents. *Child Dev Perspect*. (2020) 14:110–5. doi: 10.1111/cdep.12366
100. Shavers VL. Measurement of socioeconomic status in health disparities research. *J Natl Med Assoc*. (2007) 99:1013–23. doi: 10.13016/avw3-9cvx
101. Schwartz S. The fallacy of the ecological fallacy: the potential misuse of a concept and the consequences. *Am J Public Health*. (1994) 84:819–24. doi: 10.2105/AJPH.84.5.819
102. Braveman P, Dominguez TP. Abandon “race.” Focus on racism. *Front Public Health*. (2021) 9:689462. doi: 10.3389/fpubh.2021.689462
103. White K, Lawrence JA, Tchangalova N, Huang SJ, Cummings JL. Socially-assigned race and health: a scoping review with global implications for population health equity. *Int J Equity Health*. (2020) 19:25. doi: 10.1186/s12939-020-1137-5
104. Williams DR, Lawrence JA, Davis BA. Racism and health: evidence and needed research. *Annu Rev Public Health*. (2019) 40:105–25. doi: 10.1146/annurev-publhealth-040218-043750
105. Gee GC, Ford CL. Structural racism and health inequities: old issues, New Directions. *Du Bois Rev*. (2011) 8:115–32. doi: 10.1017/S1742058X11000130
106. Heard-Garris N, Boyd R, Kan K, Perez-Cardona L, Heard NJ, Johnson TJ. Structuring poverty: how racism shapes child poverty and child and adolescent health. *Acad Pediatr*. (2021) 21:S108–16. doi: 10.1016/j.acap.2021.05.026
107. Trent M, Dooley DG, Doug J Section On Adolescent Health, Council on Community Pediatrics, & Committee on Adolescence. The impact of racism on child and adolescent health. *Pediatrics*. (2019) 144:e20191765. doi: 10.1542/peds.2019-1765