

"Carry-Over" Effects After Magnetic Stimulation – A Mechanistic Study

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Results

Introduction

Neuromodulation is a field of medical science focused on modulating the activity of the nervous system for therapeutic purposes. Neuromodulation with magnetic fields is widely used (TMS, rTMS, dTMS, MST, and MRgFUS) and neurons are expected to return to their pre-stimulation state and function after the treatment.

However, recent research has shown that the effects of magnetic stimulation can persist even after the stimulation has ended, a phenomenon we call "carry-over" effects. These carry-over effects can have profound implications on the outcome of the stimulation. Understanding the cellular and molecular mechanisms underlying these effects is crucial for predicting and optimizing neuromodulatory outcomes.

In this context, our research aims to review the current understanding of the cellular and molecular basis underlying carry-over effects of magnetic field stimulation and highlight the need for further research to unravel the unpredictability associated with magnetic field neuromodulation.



Discussion

- \rightarrow The state of neurons may be crucial for carry-over effects occurrence after magnetic field stimulation.
- \rightarrow Understanding the mechanisms underlying these carryover effects is crucial for achieving optimal control of neural inhibition with magnetic stimulation.
- \rightarrow Further development of magnetic stimulation technology would benefit from a deeper understanding of the mechanisms responsible for carry-over effects.
- \rightarrow Strategies that can modulate the state of neurons may be explored to mitigate carry-over effects and improve the precision and reliability of magnetic stimulation.

Methods

- Micromagnetic stimulation (μ MS) stimulation on Aplysia c. mollusk neurons of the buccal ganglion
- Trans-sheath µMS on *Aplysia c*. mollusk neurons of the buccal ganglion and recording from the axon
- Multi-compartment NEURON computational model

Buccal ganglia (BC)

Cerebral ganglia (CG)

Pleural ganglia (PlG)

Pedal ganglia (PeG)

Abdominal ganglion (AG)



Figure 1. Aplysia californica and its central nervous system.



Figure 4. Carry over (post-stimulation inhibition) effects after high frequency magnetic stimulation in spontaneously firing neurons.

A. Before magnetic stimulation

soma[50].ik(0.5)

0.3

B. After magnetic stimulation

soma[50].ik(0.5)



soma[50].ik(0.5)

50Hz 100Hz 200Hz 400Hz

oma[50].v(0.5)

soma[50].ina(0.5)

soma[50].ik(0.5)

 \rightarrow Investigating the effects of different magnetic field parameters on the occurrence of carry-over effects may provide valuable insights for optimizing the application of magnetic field stimulation in neuromodulation.

References

- Oliviero A, Strens LH, Di Lazzaro V, Tonali PA, Brown P (2003), Persistent effects of high frequency repetitive TMS on the coupling between motor areas in the human. Exp Brain Res 149:107-113. Pashut T, Wolfus S, Friedman A, Lavidor M, Bar-Gad I, Yeshurun Y, Korngreen A (2011), Mechanisms of magnetic stimulation of central nervous system neurons. PLoS Comput Biol 7:e1002022. Pasley BN, Allen EA, Freeman RD (2009), State-dependent variability of neuronal responses to transcranial magnetic stimulation of the visual cortex. Neuron 62:291-303.
- Ye H, Steiger A (2015), Neuron matters: electric activation of neuronal tissue is dependent on the interaction between the neuron and the electric field. J Neuroeng Rehabil 12:65



Figure 2. Position of the miniature coil for magnetic stimulation of the neurons in the buccal ganglion from Aplysia californica.



Figure 3. NEURON simulation of magnetic stimulation with a magnetic coil. Distribution of induced electric field around the coil and the modeled neuron.



Figure 5. Carry-over effects mediated by the altered sodium and potassium channel conductance after magnetic stimulation. The sodium and potassium currents were measured and compared before and after the magnetic stimulation in a voltage clamp experiment (NEURON) simulation).



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