

1 **Asian Honey Bee *Apis cerana* Foraging on Mushrooms**

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3 Jun-ichi Takahashi<sup>1\*</sup>, Kentaro Hosaka<sup>2</sup>, Stephen J. Martin<sup>3</sup>, Akira Kawabe<sup>1</sup>

4 <sup>1</sup> Faculty of Life sciences, Kyoto Sangyo University, Kyoto, Japan

5 <sup>2</sup> Department of Botany, National Museum of Nature and Science, Amakubo 4-1-1,

6 Tsukuba, Ibaraki, Japan

7 <sup>3</sup> School of Environment & Life Sciences University of Salford, Manchester UK

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10 **Corresponding author:**

11 Jun-ichi Takahashi, Faculty of Life sciences, Kyoto Sangyo University, Kyoto, Japan.

12 E-mail: [jit@cc.kyoto-su.ac.jp](mailto:jit@cc.kyoto-su.ac.jp)

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17 Introduction

18 Honey bees (*Apis* spp.) are well known for obtaining their nutrition from pollen and nectar  
19 collected from a wide range of flowers (Winston, 1991). They also collect honeydew  
20 secreted by aphids and scale insects as an additional source of carbohydrate. Many species  
21 of bees use plant resins, which when mixed with their saliva produces propolis, a  
22 sticky substance used to seal the hive and help combat pathogens and infections (Castro,  
23 2001). In some honey bees e.g. *Apis florea* propolis can be used as an ant deterrent  
24 (Duangphakdee, Koeniger, Koeniger, Wongsiri, & Deowanish, 2005). However, in Brazil  
25 the Africanized honey bee (*Apis mellifera*) workers gather mycelium and spores from  
26 *Cladosporium* sp. of fungi (Modro, Silva, & Luz, 2009) but are not known to visit the  
27 fruiting body of fungi. Here we report for the first-time honey bee (*A. mellifera*) workers  
28 apparently feeding on the fruiting body of fungi.

29

30 Material and Methods

31 The observational survey was conducted at the Kyoto Sangyo University campus in  
32 Kyoto City, Japan in autumn 2017. At the study site, 14 individual fruiting bodies of the  
33 same species were located within a 1 m<sup>2</sup> area. Continuous observations were made for  
34 at least two hours each day between September 26 and October 3, 2017. There were no

35 flowers to attract bees within a 50-m radius of the study site. Observations of fungi were  
36 made by four researchers and recorded using a digital camera (Sony HXR-MC1),  
37 resulting in total 48 hours of recordings. During the same period, control observations of  
38 honey bee visiting goldenrod (*Solidago canadensis*) flowers around 200 m away from the  
39 fungi using the same methodology resulted in a total of 21 hours of recordings. After that,  
40 we measured the duration and frequency of visits by the honey bee workers to the  
41 fruiting body used the video data.

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#### 44 Results and Discussion

45 Although more colonies of *A. mellifera* than *A. cerana* were in the surrounding area, only  
46 *A. cerana* workers were found visiting the fruiting body at a rate of 3.2 times per hour  
47 spending an average of 28.2 seconds during each visit (Figure 1). *A. cerana* visited the  
48 goldenrod flowers at a similar rate of 3.1 times per hour and stayed for an average of 31.6  
49 seconds, whereas *A. mellifera* visited the flowers 3.4 times per hour staying an average  
50 of 35.1 seconds (Figure 1).

51

52 We observed *A. cerana* workers gnawing with their mandibles and extending their

53 tongues into the tubes (Figure 2, Video 1) of the fruiting body. The *A. cerana* workers  
54 were also observed feeding on the decaying mushrooms later in the study. The fungi was  
55 initially identified as *Xerocomus* spp. in the order Boletales (Basidiomycota, Fungi). This  
56 identification was later confirmed via DNA sequence analysis, using the internal  
57 transcribed spacer (ITS) primers, ITS5 and ITS4, with reaction conditions as described in  
58 White, Bruns, Lee, & Taylor (1990). The resultant nucleotide sequence was registered  
59 with the DDBJ DNA data bank (accession number LC333562) and had a 96% homology  
60 with genus *Xerocomus* spp. as determined by BLAST of the resulting ITS sequence.  
61 Additional DNA sequences (accession numbers MG650105 and MG650106) were  
62 obtained from RPB1 and ATP6 genes, using the primer pairs bRPB2-6F/bRPB2-7.1R  
63 (Matheny, 2005) and ATP6-3/APT6-2 (Kretzer & Bruns, 1999), respectively. These  
64 sequences also confirmed *Xerocomus* spp.-based BLAST searches.

65

66 It remains unclear why this foraging behavior of *A. cerana* at the fruiting bodies was  
67 occurring. The orchid *Cymbidium floribundum* is known to trick *A. cerana* bees into  
68 pollinating them for no reward of nectar or pollen by mimicking honey bee secretions  
69 (Sasaki, Ono, Asada, & Yoshida, 1991). It is possible that the mushroom may release  
70 secretions, which are acting as an attractant or psychostimulant. It has been shown that

71 honey bees preferred solutions with low concentrations of drugs such as nicotine and  
72 caffeine over a 20% sucrose control solution (Singaravelan, Nee'man, Inbar, & Izhaki,  
73 2005) and several insects are known to become addicted to drugs (Boppre, 1999).  
74 Furthermore, we found that some of the *Xerocomus* spp. fruiting bodies were parasitized  
75 by *Sepedonium* spp., an ascomycetous mold (Figure 2), which was identified as either a  
76 *Sepedonium* spp. (98% homology) or *Hypomyces chrysospermus* (97% homology) based  
77 on the ITS sequence obtained using the primers ITS5 and ITS4 (White, Bruns, Lee, &  
78 Taylor, 1990). These mold's are known to parasitize Boletaceae fruiting bodies, including  
79 the genus *Xerocomus* (Arora, 1986). *Hypomyces chrysospermus* produces sepedonin,  
80 which shows anti-fungal and -bacterial activities (Nagao, Yoshida, Iwai, Sakai, Tanaka,  
81 & Miyahara, 2006) that are highly prized for their culinary value (Arora, 1986). So, it is  
82 possible that *A. cerana* is selectively choosing parasitized *Xerocomus* as their food source.  
83 However, these ideas remain highly speculative, until more data is obtained. It is well  
84 established that some species of beetles and flies feed on the fruiting bodies of mushrooms  
85 (van Emden, 2013), but such behavior in bees is hitherto unknown (Winston 1991; Sasaki,  
86 2010).  
87  
88 In this study, we may have observed the mutualistic relationship between the honey bee and

89 fruiting body of a fungi. We hypothesize that *A. cerana* workers likely obtain some resources  
90 from the mushrooms. Honey bees are most likely to obtain proteins like pollen from  
91 fruiting body, but some extracts from the mycelium of amadou (*Fomes*) and reishi  
92 (*Ganoderma*) fungi have been suggested to have antiviral effect against pathogenic  
93 viruses (Stamets, Naeger, Evans, Han, Hopkins, Lopez, Moershel, Nally, Sumerlin,  
94 Taylor, Carris, & Sheppard, 2018). It has also been suggested that *A. mellifera* nurse bees  
95 prefers honey that has high antimicrobial activity (Gherman, Denner, Bobiș, Dezmirean,  
96 Mărghitas, Schlüns, Moritz, & Erler, 2014). Mushrooms often release secretions from the  
97 underside of their fruiting bodies, and the honey bees might be collecting that. In this  
98 observation, the type of the substance that the honey bees obtained from the mushrooms  
99 remains unknown.

100

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107 References

- 108 Arora, D. (1986). *Mushrooms Demystified. A Comprehensive Guide to the Fleshy Fungi.*  
109 Second Edition. Ten Speed Press, Berkeley.
- 110 Boppre, M. (1999). Drug-addicted insects in Africa. *Metamorphosis* 10,3–15.
- 111 Castro de, S.L. (2001). Propolis: biological and pharmacological activities. Therapeutic  
112 uses of this bee-product. *Annual Review of Biomedical Sciences*, 3, 49-83.
- 113 Duangphakdee, O., Koeniger, N., Koeniger, G., Wongsiri, S., & Deowanish, S. (2005).  
114 Reinforcing a barrier – a specific social defense of the dwarf honeybee (*Apis florea*)  
115 released by the weaver ant (*Oecophylla smaragdina*). *Apidologie*, 36(4), 505-511.
- 116 Gherman, B.I., Denner, A., Bobiș, O., Dezmirean, D.S., Mărghitas, L.A., Schlüns, H.,  
117 Moritz, R.F.A., & Erler, S. (2014). Pathogen-associated self-medication behavior in  
118 the honeybee *Apis mellifera*. *Behavioral Ecology and Sociobiology*, 68, 1777-1784.
- 119 Kretzer, A.M., & Bruns, T.D. (1999). Use of *atp6* in fungal phylogenetics: an example  
120 from the Boletales. *Molecular Phylogenetics and Evolution*, 13, 483-492.
- 121 Matheny, P.B. (2005). Improving phylogenetic inference of mushrooms with RPB1 and  
122 RPB2 nucleotide sequences (*Inocybe*; Agaricales). *Molecular Phylogenetics and*  
123 *Evolution*, 35, 1-20.
- 124 Modro, A.F.H., Silva, I.C., & Luz, C.F.P. (2009). Saprophytic fungus collection by

125 africanized bees in Brazil. *Neotropical entomology*, 38(3), 434-436.

126 Nagao, K., Yoshida, N., Iwai, K., Sakai, T., Tanaka, M., & Miyahara, T. (2006).  
127 Production of sepedonin by *Sepedonium chrysospermum* NT-1 in submerged culture.  
128 *Environmental Sciences*, 13(5), 251-256.

129 Sasaki, M., Ono, M., Asada, S., & Yoshida, T. (1991). Oriental orchid (*Cymbidium*  
130 *pumilum*) attracts drones of the Japanese honeybee (*Apis cerana japonica*) as  
131 pollinators. *Experientia*, 47, 1229-1231.

132 Sasaki, M. (2010). Bee's eye View of Flowering Plants: Nectar- and Pollen-source Plants  
133 and Related Honeybee Products. Kaiyusya, Tokyo. [In Japanese].

134 Singaravelan, N., Nee'man, G., Inbar, M., & Izhaki, I. (2005). Feeding responses of Free-  
135 flying Honeybees to Secondary Compounds mimicking Floral Nectars. *Journal of*  
136 *Chemical Ecology*, 31, 2791-2804.

137 Stamets, P.R., Naeger, N.L., Evans, J.D., Han, J.O., Hopkins, B.K., Lopez, D., Moershel,  
138 H.M., Nally, R., Sumerlin, D., Taylor, A.W., Carris, L.M., & Sheppard, W.S. (2018).  
139 Extracts of Polypore Mushroom Mycelia Reduce Viruses in Honey Bees. *Scientific*  
140 *Reports*, 8, 13936.

141 Van Emden, H.F. (2013). *Handbook of Agricultural Entomology*. Wiley-Blackwell, New  
142 Jersey.



143 White, T.J., Bruns, T., Lee, S., & Taylor, J. (1990). Amplification and direct sequencing  
144 of fungal ribosomal RNA genes for phylogenetics, in: Innis, M.A., Gelfand, D.H.,  
145 Sninsky, J.J., & White, T.J. (Eds.), PCR protocols, a guide to methods and  
146 applications, Academic Press, pp.315-322. Academic Press, Massachusetts.

147 Winston, M.L. (1991). The Biology of the Honey Bee. Harvard University Press,  
148 Massachusetts.

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151 **Figures captions**

152

153 **Figure 1**

154 Comparison of (a) duration and (b) visits (mean  $\pm$  s.e.) in the honey bees *Apis cerana* and  
155 *Apis mellifera* on the fungi *Xerocomus* cf. *chrysenteron* and goldenrod flowers *Solidago*  
156 *canadensis*. N = The total number of honey bees observed.. Observations of fungi and  
157 flower were made for 42 and 21 hours respectively.

158

159 **Figure 2**

160 *Apis cerana* worker foraging on the tubes of the mushroom *Xerocomus* cf. *chrysenteron*  
161 (left) and biting the fruit body of the mushroom *Xerocomus* cf. *chrysenteron* parasitized  
162 by *Sepedonium* sp. (white mycelia on cap surface) (right). Please click the image to play

163 **Video 1.**