Check for updates

OPEN ACCESS

EDITED BY Yi Zhang, Zhejiang University, China

REVIEWED BY Zhiyong Zhang, Shanghai Jiao Tong University, China Haifeng Wang, Chinese Academy of Sciences, China

*CORRESPONDENCE Emmanuel L. Barbier Immanuel.barbier@univ-grenoble-alpes.fr

[†]These authors share last authorship

RECEIVED 23 February 2023 ACCEPTED 28 April 2023 PUBLISHED 02 June 2023

CITATION

de Souza DAR, Mathieu H, Deloulme J-C and Barbier EL (2023) Evaluation of kernel low-rank compressed sensing in preclinical diffusion magnetic resonance imaging. *Front. Neurosci.* 17:1172830. doi: 10.3389/fnins.2023.1172830

COPYRIGHT

© 2023 de Souza, Mathieu, Deloulme and Barbier. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Evaluation of kernel low-rank compressed sensing in preclinical diffusion magnetic resonance imaging

Diego Alves Rodrigues de Souza¹, Hervé Mathieu^{1,2}, Jean-Christophe Deloulme^{1†} and Emmanuel L. Barbier^{1,2*†}

¹Université Grenoble Alpes, INSERM, U1216, Grenoble Institut Neurosciences, Grenoble, France, ²Université Grenoble Alpes, INSERM, US17, CNRS, UAR 3552, CHU Grenoble Alpes, Grenoble, France

Compressed sensing (CS) is widely used to accelerate clinical diffusion MRI acquisitions, but it is not widely used in preclinical settings yet. In this study, we optimized and compared several CS reconstruction methods for diffusion imaging. Different undersampling patterns and two reconstruction approaches were evaluated: conventional CS, based on Berkeley Advanced Reconstruction Toolbox (BART-CS) toolbox, and a new kernel low-rank (KLR)-CS, based on kernel principal component analysis and low-resolution-phase (LRP) maps. 3D CS acquisitions were performed at 9.4T using a 4-element cryocoil on mice (wild type and a MAP6 knockout). Comparison metrics were error and structural similarity index measure (SSIM) on fractional anisotropy (FA) and mean diffusivity (MD), as well as reconstructions of the anterior commissure and fornix. Acceleration factors (AF) up to 6 were considered. In the case of retrospective undersampling, the proposed KLR-CS outperformed BART-CS up to AF = 6 for FA and MD maps and tractography. For instance, for AF = 4, the maximum errors were, respectively, 8.0% for BART-CS and 4.9% for KLR-CS, considering both FA and MD in the corpus callosum. Regarding undersampled acquisitions, these maximum errors became, respectively, 10.5% for BART-CS and 7.0% for KLR-CS. This difference between simulations and acquisitions arose mainly from repetition noise, but also from differences in resonance frequency drift, signal-tonoise ratio, and in reconstruction noise. Despite this increased error, fully sampled and AF = 2 yielded comparable results for FA, MD and tractography, and AF = 4showed minor faults. Altogether, KLR-CS based on LRP maps seems a robust approach to accelerate preclinical diffusion MRI and thereby limit the effect of the frequency drift.

KEYWORDS

diffusion MRI, compressed sensing (CS), kernel principal component analysis, mouse brain, tractography, composite real PCA, microtubule-associated protein, Berkeley Advanced Reconstruction Toolbox (BART)

1. Introduction

Diffusion MRI, based on diffusion-weighted imaging (DWI), is used in neuroscience research to characterize anatomical connectivity (Le Bihan and Johansen-Berg, 2012). At the preclinical level, anatomical connectivity mapping helps to characterize transgenic animal models used in various areas, from neurodevelopment (Oguz et al., 2012) to neurodegeneration (Gatto et al., 2019; Nishioka et al., 2019). To obtain reliable connectivity data, it is now accepted that at least 30 diffusion directions are required; the precise number depends on the analysis approach (Schilling et al., 2017). Advanced diffusion models require even richer datasets (more diffusion directions and/or use of several diffusion gradient values) and therefore take longer to acquire (Calabrese et al., 2015; Allan Johnson et al., 2019). Currently, these in vivo acquisitions can be used for humans owing to several well-known acquisition strategies: parallel imaging (Pruessmann et al., 1999; Griswold et al., 2002), compressed sensing (CS) (Lustig et al., 2007), and multiband acquisition (Setsompop et al., 2013; Lee et al., 2021).

In preclinical settings, however, these developments are not widely used yet, despite several proof-of-principle studies. Moreover, the small voxel sizes required to map tracts in mouse or rat brains (100 μ m or less) contribute to further increasing the DWI acquisition times, which are much longer than in clinical settings (Aggarwal et al., 2010; Calabrese et al., 2015). Therefore, most preclinical studies are performed *ex vivo* with acquisition durations that routinely exceed 8 h (Zhang et al., 2012; Gimenez et al., 2017; Wang et al., 2018; Allan Johnson et al., 2019). Such long acquisition times may decrease image quality because of hardware (e.g., resonance frequency) and/or sample drift (Vos et al., 2017) and can limit the number of samples used owing to cost and/or scan time availability.

One of the most attractive means to reduce acquisition duration is CS (Lustig et al., 2007; Michailovich et al., 2011; Bilgic et al., 2012; Mani et al., 2015), which has recently been evaluated in mouse brain connectomics (Allan Johnson et al., 2019; Zhang et al., 2020). The Johnson group reported connectivity data obtained with an acquisition pattern based on Monte-Carlo sampling, which was constant across diffusion directions (Wang et al., 2018; Allan Johnson et al., 2019). Acquired data were reconstructed using the SparseMRI CS approach developed by Lustig et al. (2007), and the impact of reducing the scan time on mean diffusivity (MD), fractional anisotropy (FA), and on the ability to detect whitematter tracts and fiber-crossings was evaluated (Wang et al., 2018). Their approach was also evaluated in a model of autism spectrum disorders (Allan Johnson et al., 2019). In their settings, an eightfold reduction in scan time (from 92.6 to 11.8 h) was acceptable, but the scan time remains long. More recently, Zhang et al. (2020) proposed a kernel low-rank (KLR) method to jointly reconstruct multiple diffusion-weighted images (Nakarmi et al., 2017), using undersampling patterns that vary with the diffusion direction (Mani et al., 2015). In their settings, the KLR-CS appears more efficient than the conventional CS proposed by Lustig et al. (2007) for a 5.56-fold reduction in scan time (from 8 to 1.4 h). However, the proposed KLR approach has been evaluated on magnitudefiltered data, i.e., data in which the Hermitian symmetry in k-space has been enhanced, and using retrospective undersampling (Zhang et al., 2020). Altogether, to our knowledge, there is no implementation of a KLR-CS approach able to efficiently handle complex undersampled DWI acquisitions.

In this study, we thus evaluated both the acquisition, using a four-channel receive coil, and a KLR-CS reconstruction protocol adapted to handle complex undersampled DWI data. Data were acquired on a preclinical mouse model that exhibits altered brain connectivity: the *MAP6* knockout model (Deloulme et al., 2015). The DWI undersampling parameters were optimized using simulations. A KLR-CS approach, based on the use of low-resolution-phase (LRP) maps, was introduced, and parametric maps and fiber tracts from undersampled acquisitions were analyzed, using the Berkeley Advanced Reconstruction Toolbox (BART)-CS as reference. Finally, differences between simulations and acquisitions were explored, considering the change in signal-to-noise ratio, frequency drift, and repetition noise.

2. Materials and methods

2.1. Animals

The study protocol was approved by the local animal welfare committee (Comité Local GIN, C2EA-04–APAFIS number 21234-2019031308592774) and complied with EU guidelines (Directive 2010/63/EU). Every precaution was taken to minimize the number of animals used and the stress on animals during experiments. The *MAP6*-deficient mouse line used in this study is on the C57BL/6 genetic background (Andrieux et al., 2002). Adult mice that were heterozygous for *MAP6* (*MAP6*^{+/-}) and their wild-type (WT) littermates were obtained by crossing *MAP6*^{+/-} mice.

As the complete *MAP6* knockout (*MAP6*^{-/-}) leads to major changes in brain connectivity (Deloulme et al., 2015), we used the partial (heterozygous *MAP6*^{+/-}) knockout to achieve subtle changes in tract morphology. Thus, we challenged the level of detail quantifiable by CS reconstructions.

2.2. Brain preparation for *ex vivo* MRI acquisitions

Brains were prepared according to a previously reported protocol (Gimenez et al., 2017). After transcardiac perfusion with a 4% paraformaldehyde solution containing Gd-chelate (6.25 mM of Gd-DOTA; Guerbet Laboratories, Roissy, France), mice were decapitated. After the removal of skin and muscles, the skull was immersed in the same fixing solution for 4 days and transferred into Fomblin oil (FenS chemicals, Goes, Netherlands), an oil that generates no MRI signal but has a magnetic susceptibility close to that of brain tissue. MRI was performed at least 7 days after

Abbreviations: AF, acceleration factor; b0, $b\sim 0 \text{ s/mm}^2$ images (baseline images acquired without diffusion weighting); KPCA, kernel principal component analysis, CS, compressed sensing; DWI, diffusion-weighted imaging; ESPIRIT, efficient L1 iterative self-consistent parallel imaging reconstruction; FA, fractional anisotropy; FS, fully sampled; KLR, kernel low-rank; LRP, low-resolution-phase; *MAP6*, microtubule-associated protein 6; MD, mean diffusivity; pSNR, peak signal-to-noise ratio; ROI, region of interest; SNR, signal-to-noise ratio; SSIM, structural similarity index measure; WT, wild type.

brain fixation. This protocol ensures a homogeneous distribution of the Gd-DOTA throughout the whole mouse brain (Gimenez et al., 2016). The contrast agent decreased brain T1 from 1000 ± 102 to 110 ± 13 ms and brain T2 from 27.3 ± 3.1 to 18.3 ± 2.6 ms (data not shown), allowing a reduction in acquisition times.

2.3. Optimization of the CS undersampling pattern for DWI

To select an undersampling approach, 20 different k-space undersampling patterns were explored, by retrospectively undersampling the FS DWI datasets acquired from the 3 WT brains and using the methods described below:

- Monte-Carlo vs. Poisson-disk sampling. Monte-Carlo sampling patterns were generated through a variable probability density function with a flat region in its center (Wang et al., 2018; Zhang et al., 2020). Poisson-disk sampling patterns were generated using the algorithm embedded in BART. For this pattern, we evaluated regular and elliptical sampling as well as uniform and variable sampling density. To ensure fair comparisons, the area of the FS center of k-space was kept constant across sampling patterns.
- Single- vs. multi-mask sampling (Senel et al., 2019). The single-mask approach stands for the use of the same undersampling pattern for all DWI volumes and the multi-mask approach for the use of different undersampling patterns for each DWI volume.
- Fully sampled (FS) vs. undersampled b0 images. To ensure a comparable global acceleration factors (AF) across the whole DWI datasets (b0 + diffusion direction images), the AF per diffusion direction (AF_{diff}) was increased in the case of FS b0 images. The behavior of AF and AF_{diff}, as a function of the sampling strategies for b0 images, is further described in **Supplementary Figure 1**.

Acceleration factors (AF) values of 2, 3, 4, and 6 were explored. Images were reconstructed and MD and FA were computed across the whole brain. The most robust undersampling strategy was determined (see section "2.7. Comparison metrics").

2.4. Image acquisitions

MRI acquisitions were carried out at 9.4 T (Biospec Avance III HD, Bruker, Ettlingen, Germany; IRMaGe facility) using a 4-channel head surface cryocoil for reception, a volume coil for transmission, and Paravision 7. The following sequences were performed:

- a 3D CS T1w gradient-echo MRI acquisition (cf Supplementary Table 1 for parameter details). These anatomical data were used to overlay an atlas for the DWI analysis (see section "2.6. MD and FA maps and tractography").
- A 3D CS, spin-echo, diffusion-weighted images (DWI). The gradient directions were generated using an optimal

distribution across the surface of the sphere (Caruyer et al., 2013). This sequence allows to assign a 3D undersampling pattern to each diffusion direction¹. Several acquisitions were performed with different AF, using $\delta = 4.0$ ms and $\Delta = 9.6$ ms, as summarized in **Supplementary Table 1**. To estimate and limit the impact of the frequency drift, the main resonance frequency was calibrated before and after each DWI acquisition.

2.5. Image reconstruction

Data were reconstructed using the Matlab environment (MATLAB2021b; The MathWorks, MA, USA). For the undersampled dataset, two reconstruction approaches were performed:

- Conventional CS reconstructions were performed using the BART (Uecker et al., 2015, 2021) toolbox v0.7.00², which calculates coil sensitivity maps using ESPIRiT (Efficient L1 iterative self-consistent parallel imaging reconstruction) calibration (Uecker et al., 2014). For the minimization process, we used the parameters optimized in Wang et al. (2018): $\lambda_1 = 0.005$, $\lambda_2 = 0.002$, max iteration = 200. This toolbox was chosen to have the same reference technique as in the original KLR-CS paper (Zhang et al., 2020).
- Kernel low-rank (KLR)-CS reconstructions of diffusionweighted images were performed using an in-house adaptation of the original KLR toolbox as published (Nakarmi et al., 2017; Zhang et al., 2020). However, the original reconstruction method was evaluated on data after magnitude filtering, a step that enhances the Hermitian symmetry of k-space (Zhang et al., 2020). Therefore, this method-hereafter called "magnitude-filtered KLR-CS"-is not designed for acquired undersampled datasets as the data-consistency step is no longer efficient. We therefore proposed and evaluated two adaptations of the original KLR toolbox: a composite kernel principal component analysis (KPCA) adapted from the composite real PCA of complex signals (Hellings et al., 2015) and LRP maps. For the KLR-CS based on composite KPCA, we replaced the original input to the KPCA module i.e., magnitude data - with composite data, which is a concatenation of the real and imaginary parts of the data. The output of composite KPCA is then reorganized to produce complex data before enforcing data consistency. For the KLR-CS based on LRP maps, the phase information related to the FS center of k-space (i.e., the low-resolution part of the dataset) is stored before the self-training step and added back to the output magnitude image, yielding a complex image for each channel that can then be used to enforce data consistency using the original undersampled k-space. This approach using the phase information of the entire undersampled k-space was also tested but led to lower quality results. For KLR training, the low-resolution, 4-channels, 4D images obtained from the FS region at the center of the k-space were used.

¹ https://github.com/nifm-gin/compressedSensing

² https://mrirecon.github.io/bart/

In addition, all the FS DWI datasets (n = 6 brains; 3 WT and 3 $MAP6^{+/-}$) were retrospectively undersampled, using the same undersampling pattern and AF values as the one used for the acquisitions, and reconstructed with the two methods described above. Thereby, metrics obtained from simulated (i.e., retrospective) undersampling could be compared to that obtained from undersampled acquisitions.

2.6. MD and FA maps and tractography

Diffusion-weighted imaging (DWI) data was processed using MRtrix (v3.0_RC3-137).³ After image denoising, unringing, and brain masking, tensor-derived FA and MD maps were generated. FA and MD were measured in the whole brain and two regions of interest (ROI), corpus callosum and caudate putamen.

The fiber orientation distributions were computed with constrained spherical deconvolution (CSD) (Tournier et al., 2004, 2007) and default parameter values [for instance, maximum harmonic order lmax = 4 (Tournier et al., 2013)]. Whole brain tractography was computed with a probabilistic streamline tracking (Parker and Alexander, 2003; Behrens et al., 2007) (iFOD2 algorithm) and three seeds per voxel. Empirically optimized tracking parameters on our mouse brain data were applied after normalization of the fiber orientation distributions: step size = 0.01 mm, radius of curvature = 0.07 mm (Moldrich et al., 2010), min length = 0.1 mm, max length = 50 mm (Aydogan et al., 2018), and cutoff = 0.2. In addition, several major tracts were isolated as follows. For each tract, several ROIs were defined, according to a mouse brain atlas (Paxinos and Franklin, 2019). An ROI was a 2D surface in one orientation (coronal, horizontal, sagittal). Each ROI-tract association then received either the label "AND" or "NOT." The fibers of one particular tract were selected as all the fibers that cross the ROIs labeled "AND" minus the fibers that cross the ROIs labeled "NOT." Supplementary Table 2 shows the list of ROIs, with their labels and the orientation in which they were considered, for each tract evaluated in this study.

2.7. Comparison metrics

To compare parametric maps, two metrics were used: the absolute error (%) (called "error" in the rest of the manuscript) and the structural similarity index measure [SSIM, ranging from 0 (worst) up to 1 (best) (MathWorks, 2022)]. The details regarding the computation of both metrics are provided as **Supplementary** material.

Both metrics were calculated for each reconstruction, considering the whole 3D brain. Error and SSIM maps were obtained using the FS dataset as the reference, unless mentioned otherwise. In addition, to allow tractogram comparisons, the mean fiber length was computed. To remove the image shift caused by the frequency drift that occurs during hours-long acquisitions, raw DWI images were registered to the FS images

Statistical analyses were performed in GraphPad Prism version 9.4.1 for Windows (GraphPad Software, La Jolla, CA, USA).⁴ The p-values mentioned along this article correspond to paired t-tests, unless stated otherwise.

3. Results

3.1. Optimization of the CS for DWI using simulations

Regarding the choice of undersampling patterns, examples based on multi-mask and Monte-Carlo or Poisson-disk sampling are shown in Figure 1A, for different AF values. Overall, while the choices of Monte-Carlo multi-mask sampling clearly yielded better results (Figures 1B–E), the full sampling of b0 images did not provide a major advantage. To ease comparison with the previous study (Zhang et al., 2020), we choose to keep that full sampling of b0 images. Altogether, the undersampling pattern based on Monte-Carlo with FS b0 images and multi-mask was used for all subsequent evaluations (simulations and acquisitions).

Using this sampling pattern, we then compared the effects of composite KPCA and LRP using complex data and AF between 2 and 6. For sake of comparison, we evaluated the original KLR-CS method using k-spaces from complex images instead of k-spaces from magnitude-only images. Figure 2 shows FA and MD maps obtained from the two proposed KLR-CS, BART-CS, and the magnitude-filtered KLR-CS reconstructions. The corresponding error and SSIM maps are shown in Figures 3A, B, E, F, respectively. As expected, BART-CS, LRP KLR-CS, and KPCA KLR-CS, which all handle complex data, yielded better results than the magnitudefiltered KLR-CS. For example, for AF = 2, the median error on FA with LRP KLR-CS was 4.79 \pm 0.12% while it was 9.69 \pm 0.12% with magnitude-filtered KLR-CS (Figures 3C, D, G, H). This confirms that the original, magnitude-filtered, KLR CS approach is not adapted to complex data. In the rest of the manuscript, we thus focus on LRP and KPCA KLR-CS. Regarding FA (Figures 3C, D), the LRP KLR-CS outperformed BART-CS for AF = 2 and AF = 3 (p < 0.0001) and KPCA KLR-CS for all AFs (p < 0.0001). For AF = 4 and 6, LRP KLR-CS had similar results to BART-CS. Regarding MD (Figures 3G, H), LRP KLR-CS outperformed BART-CS and KPCA KLR-CS for all AFs (p < 0.01). Therefore, LRP KLR-CS, hereafter called "KLR-CS" for simplicity, was used to reconstruct all the data reported in the subsequent studies.

Figures 4A, B shows examples of FA and MD maps obtained from a WT mouse as a function of AF using BART-CS and KLR-CS reconstructions, optimized as described. There were no obvious faults observed in the brain slices. The focus on the hippocampus shows decreased contrast as AF increases, in particular for BART-CS reconstructions. For quantitative analysis, the corresponding

using an in-house rigid registration tool prior to computing error and SSIM metrics. Across each brain, the median error and the median SSIM were retained, to limit the effect of locally high errors such as the ones observed on the edges or in moving structures (ventricles, vessels).

³ https://www.mrtrix.org/

⁴ www.graphpad.com



FIGURE 1

Optimization of the undersampling strategy. (A) Examples of undersampling patterns for AF = 2 to 6. Corresponding (B,D) error and (C,E) SSIM values, using MD maps and AF = 6. (B,C) Correspond to BART-CS and (D,E) to KLR-CS reconstruction. Results are expressed as mean \pm standard deviation across animals (n = 3). Columns show different Poisson-disk and Monte-Carlo undersampling patterns and their resulting masks for the first b0 image and the first diffusion direction. Rows correspond to AF (between 2 and 6) and sampling mode of b0 images (FS: fully sampled; US: undersampled). Note that only phase encoding directions are undersampled. "N.A." stands for "not available" (a mask cannot be generated in these specific conditions). A total of 20 possible combinations of undersampling patterns per AF was evaluated (Poisson-disk; fully sampled or undersampled b0/uniform or variable density/single or multi-mask mode/common or elliptical scanning; Monte-Carlo: fully sampled or undersampled b0/single or multi-mask mode). The naming of the bar graph: MC: Monte-Carlo, PD: Poisson-disk, Ud: uniform density, Vd: variable density, Rs: regular scanning, Es: elliptical scanning. With the KLR-CS reconstruction, the multi-mask undersampling outperformed the single-mask undersampling (D,E). For instance, for a Monte-Carlo with FS b0s undersampling scheme ("MC_FS-b0" bars) and AF = 6, the error reduced from 5.92 \pm 0.21 to 4.06 \pm 0.16% (p = 0.0054), and the SSIM increased from 0.781 \pm 0.063 to 0.917 \pm 0.020 (p = 0.0479) when replacing the single-mask undersampling with a multi-mask. With the BART-CS reconstruction, single and multi-mask undersampling has no effect on error or SSIM (B,C).



FIGURE 2

FA and MD maps from fully sampled data and after simulated undersampling, reconstructed using BART-CS, KLR-CS based on composite KPCA, KLR-CS with low-resolution-phase (LRP) maps, and magnitude-filtered KLR-CS. **(A)** FA and **(B)** MD maps from a zoom on the hippocampus, obtained from simulated reconstructions by BART (blue) and three different implementations of KLR-CS (green), for different AFs. A reconstruction from a fully sampled dataset (red) is shown as the reference.



low-resolution-phase (LRP) maps, and magnitude-filtered KLR-CS. Error and SSIM of (**A**,**B**) FA and (**E**,**F**) MD maps using BART-CS and three different implementations of KLR-CS, for different AFs. Full 3D brain values of (**C**,**G**) median error and (**D**,**H**) median SSIM, using the fully sampled data as the reference. Results expressed as mean \pm standard deviation across the animals (*n* = 3). The three KLR-CS implementations are KLR-CS based on composite KPCA (Comp), KLR-CS using low-resolution-phase (LRP) maps, and magnitude-filtered KLR-CS (Magn).

error and SSIM maps are shown in Figures 5A, D, and the median error and median SSIM values across the six brains are shown in Figures 5B, C for FA and Figures 5E, F for MD. Since the median error and median SSIM of FA and MD maps were not significantly different between the WT and $MAP6^{+/-}$ groups (unpaired *t*-test, p > 0.05), data were pooled to improve statistical power in the methodological study. As AF increased from 2 to 6, there was an increase in median error together with a decrease in median SSIM. Overall, KLR-CS reconstructions led to smaller median errors and higher median SSIM than BART-CS, regardless of AF. For instance, with AF = 2 and for FA maps, BART-CS and KLR-CS reconstructions had a median error of 5.78 \pm 0.11 and 4.79 \pm 0.12% (p < 0.0001) and a median SSIM of 0.976 \pm 0.001 and 0.982 \pm 0.001 (p < 0.0001), respectively. For MD maps under AF = 2, BART-CS and KLR-CS had a median error of 1.41 \pm 0.04 and 0.71 \pm 0.02% (p < 0.0001) and a median SSIM of 0.988 \pm 0.001 and 0.997 \pm 0.001 (p < 0.0001), respectively. Altogether, the KLR-CS reconstructions were of very good quality up to AF = 4, with an error below 2% and a SSIM above 0.970.

Tractographies of the anterior commissure of WT and $MAP6^{+/-}$ mice — a model for which we expect tract differences —

are shown in **Figure 6** for different AF values. First, as expected, the WT and $MAP6^{+/-}$ mice exhibited different mean fiber lengths, as observed on the FS data (unpaired *t*-test, *p* = 0.0281) (**Figure 6C**). For AF = 2, 3, and 4 and with the parameters used for the tractography, the two reconstruction methods had similar performances (*p* > 0.05 for AF = 2, 3 and 4 vs. AF = 1) and maintained the ability to observe the reduction in mean fiber length between WT and $MAP6^{+/-}$ mice. For AF = 6, KLR-CS remained robust while BART-CS sometimes missed the anterior part of the anterior commissure in $MAP6^{+/-}$ mice (**Figure 6B**). Overall, KLR-CS and BART-CS yielded comparable results for AF = 2, 3, and 4, but KLR-CS remained reliable at AF = 6 while BART-CS did not.

3.2. Evaluation of CS DWI acquisitions

After evaluating BART-CS and KLR-CS methods using simulated undersamplings, we reconstructed undersampled acquisitions carried out with our in-house DWI spin-echo CS sequence. AF = 2 and 4 were used as they corresponded to the most promising simulation results. For the acquisitions, we evaluated



FA and MD maps from fully sampled data and after simulated undersampling, using BART-CS and KLR-CS reconstructions. Examples of **(A)** FA and **(B)** MD maps from a magnification on the hippocampus, obtained from fully sampled (red; AF = 1), BART-CS (blue), and KLR-CS (green) reconstructions, using different AFs.

the FA and MD maps and the ability to reconstruct tracts. No difference may be seen on the whole maps (not shown), however, magnifying the complex region of the hippocampus revealed reduced image contrasts and some spatial smoothing as AF increased (**Figure** 7). Visually, this smoothing was less pronounced for KLR-CS than for BART-CS, in line with the simulation results. Corresponding error and SSIM maps (**Figures 8A–D**) and values (**Figures 8E–H**) confirmed the better performance of KLR-CS over BART-CS (e.g., for AF = 2, MD error: 2.59 ± 0.12 and $2.13 \pm 0.12\%$, p < 0.0001; MD SSIM: $0.964 \pm 0.002\%$ and 0.977 ± 0.002 , p < 0.0001).

We surprisingly observed that the error increased and SSIM decreased in acquisitions compared with simulations, for all AF values and for both FA and MD. For AF = 2 and KLR-CS, the FA error increased from 4.79 \pm 0.12% in simulations to 10.28 \pm 0.44% in acquisitions (p < 0.0001), and the FA SSIM decreased from 0.981 \pm 0.001 to 0.922 \pm 0.004% (p < 0.0001). Similarly, the MD error increased from 0.71 \pm 0.02% in simulations to 2.13 \pm 0.12% in acquisitions (p < 0.0001), and the MD SSIM decreased from 0.997 \pm 0.001 to 0.977 \pm 0.002% (p < 0.0001). As the errors in acquisitions are surprisingly much larger than that observed in simulations, we explored the different contributions to

these errors to ensure that the proposed acquisition method was working as intended.

First, the repetition noise was evaluated. The FS and the AF = 2 DWI datasets were acquired three times on the same fixed brain. The comparison of the three FS datasets between themselves had a median error of $7.97 \pm 0.57\%$ and a median SSIM of 0.965 ± 0.004 for FA and $1.50 \pm 0.19\%$ and 0.987 ± 0.003 for MD (Supplementary Figure 2). These repetition errors are shown in Figure 8 as a red dashed line, and they contribute to about two thirds of the observed median error.

Second, we evaluated the difference in the resonance frequency drift during an FS acquisition (13 h 37 min) and during an AF = 2 acquisition (6 h 49 min). We observed drifts of 33.9 ± 13.6 and 10.9 ± 5.1 Hz, respectively. A part of this frequency drift induces a spatial shift, which is corrected during post-processing by image registration (0.6 pixels and 0.3 pixels for our two experimental conditions). The rest of the frequency drift may not be corrected for and alters the image. Because of the acquisition durations, we expected the FS data to be more affected than the CS data. To evaluate this contribution, we simulated a spatial drift and computed the median error and the SSIM between the FA and MD maps obtained with and without simulated drift. We observed an

FIGURE 5

Error and SSIM, derived from FA and MD maps obtained after simulated undersampling and reconstruction by BART-CS and KLR-CS. Error and SSIM maps for (A) FA and (D) MD, using BART-CS and KLR-CS reconstructions, for different AF values. Corresponding (B,E) median error and (C,F) median SSIM obtained for the full 3D brain, using fully sampled data as the reference. Results are expressed as mean \pm standard deviation across the animals (n = 6).

error that represents about 10% of the error between the FS and CS acquisitions (Figure 8).

Third, we evaluated the change in peak signal-to-noise ratio (pSNR) between the FS and the CS acquisitions. For AF = 2, the average pSNRs across diffusion directions of FS, CS simulations, and CS acquisitions were 318.0 \pm 26.9, 179.3 \pm 15.1, and 155.9 \pm 14.1, respectively. The 13% reduction in pSNR between CS simulations and CS acquisitions (p < 0.0001) contributes to increasing the error on acquired FA and MD maps. In addition, we observed strong correlations between pSNR and the median error as a function of the diffusion direction: -0.7144 (p < 0.0001) and -0.6785 (p < 0.0001) for BART-CS and KLR-CS acquisitions, respectively. The noise level was also independent of the direction, suggesting that it remains stable when the undersampling pattern changes, as expected.

To evaluate the utility of the proposed CS approach, we considered two additional metrics. First, as all the reported metrics so far were obtained at the pixel level for the whole brain, we considered instead the error and the SSIM within two ROIs: the corpus callosum, a white matter ROI, and the caudate putamen, a gray matter ROI. This approach corresponds to what is usually performed to report diffusion-based estimates. Regarding the change in absolute value, the ROI-level error was lower than the pixel-level error reported in Figure 8 for MD of gray matter and FA of white matter regions, and it was higher for MD of white matter and in some cases for FA of gray matter regions. In addition, we observed that KLR-CS almost always outperforms BART-CS in simulations and acquisitions (see Supplementary Figure 3 for details).

Second, we considered white-matter tracts. The main shape of the anterior commissure is preserved in both simulations and acquisitions (Figures 9A–D). Both BART-CS and KLR-CS methods did not show significant differences in the mean fiber length in simulations and acquisitions to their corresponding WT and $MAP6^{+/-}$ references for AF = 2 (p > 0.05 for all) (Figures 9E, F). However, for AF = 4, CS acquisitions showed a reduction in mean

(Simulation) and maps in the right column (**B,D**) after CS acquisitions (Acquisition), using different AFs. Fully sampled (red), BART-CS (blue), and KLR-CS (green). The fully sampled reconstruction (AF = 1), which does not differ between simulations and acquisitions, has been repeated to facilitate figure reading.

fiber length for both WT (-11.9%, unpaired *t*-test, p = 0.0059) and $MAP6^{+/-}$ (-5.9%, unpaired *t*-test p = 0.0424) mice when compared with CS simulations. With our small dataset, the difference in fiber length between WT and $MAP6^{+/-}$ mice, visible for the FS and AF = 2 data, is no longer significant for AF = 4. A similar analysis was made for the fornix, a tract with increased complexity, given its wide distribution in the 3D space and higher levels of defasciculation than the anterior commissure. As previously found, the main shape of the fornix and the mean fiber length were conserved by both CS methods (see Supplementary Figure 4 for details). Moreover, KLR-CS showed a smaller variance on the mean fiber lengths than BART-CS (p < 0.05, considering data from the anterior commissure and fornix). These results suggest that AF = 2 seems to have no impact on fiber tract analysis, and AF = 4 has a small but acceptable impact. In addition, KLR-CS appears as the most robust method.

4. Discussion

In this study, we implemented an original 3D spin-echo CS acquisition method for preclinical DWI imaging, with the possibility of using the same or different sampling masks per diffusion direction. Two reconstruction algorithms were evaluated: BART-CS and the more recently proposed KLR-CS approach. After an optimization step based on retrospective undersampling, FA and MD maps, as well as tractographies, were produced based on CS data acquired at 9.4T using fixed mouse brains. The two reconstruction protocols yielded robust results for a reduction in scan time up to 4, with a clear advantage for KLR-CS. When FA and MD were estimated in a white-matter ROI, the difference between FS and KLR-CS reconstructions remained below 3.9 and 7.0%, for AF = 2 and 4, respectively. Regarding tracts, a reduction in scan time up to 4 produced only minor faults, even in a complex structure such as the fornix.

Moreover, we used a definition of AF that differs from the one used in previous studies (Zhang et al., 2020): AF was considered here as the effective reduction in scan time. Therefore, as we chose to fully sample the b0 images, the AF used per diffusion direction (AF_{diff}) was higher. For instance, to reach an effective AF of 4, we used an AF_{diff} of about 5.71. To facilitate the comparison with previous CS DWI studies, **Supplementary Table 1** provides a summary of simulation and acquisition parameters in this and previous studies, including both AF and AF_{diff}. de Souza et al.

FIGURE 8

Error and SSIM, derived from FA and MD maps obtained using BART-CS and KLR-CS, for simulated undersamplings and for CS acquisitions. Error and SSIM for (**A**,**B**) FA and (**C**,**D**) MD maps and corresponding full 3D brain, (**E**,**G**) median error and (**F**,**H**) median SSIM obtained using BART-CS (blue) and KLR-CS (green) reconstructions, for the full 3D brain, and using fully sampled data as the reference. The left column (**A**,**C**,**E**,**G**) corresponds to retrospective undersampling (Simulation), and the right one (**B**,**D**,**F**,**H**) to CS acquisitions (Acquisition). The repeatability error and SSIM, calculated from three repetitions of a fully sampled acquisition on the same sample, are represented as red dashed lines for comparison purposes. Results expressed as mean \pm standard deviation across animals (n = 6).

Tractography of the anterior commissure (ac) from fully sampled data, after simulated undersampling, and for CS acquisitions, using BART-CS and KLR-CS reconstructions. Anterior commissure (ac) of a (A,B) WT and a (C,D) MAP6+/- mouse from fully sampled (red), BART-CS (blue), and KLR-CS (green) reconstructions, using different AFs. Mean fiber length of the ac of (E) WT and (F) MAP6+/- mice. The left column (A,C,E) shows results obtained with simulations and the right column (B,D,F) with acquisitions. The anterior commissure from fully sampled data is repeated between simulation and acquisition columns, to facilitate figure reading. Results are expressed as mean ± standard deviation across animals (n = 3 per group). acp, ac posterior part; aca, ac anterior part.

As the published KLR-CS requires a fully sampled dataset for regularization purposes (Nakarmi et al., 2017; Zhang et al., 2020), we proposed two approaches to deal with acquired complex data and chose, based on simulations, the KLR-CS based on LRP maps, which, with AF \leq 6, outperformed BART-CS. Further improvements may be proposed. First, coil sensitivity maps

calculated by robust algorithms such as ESPIRiT (Uecker et al., 2014), important for the quality of BART-CS reconstructions, are not used in the current implementation of KLR-CS. Indeed, KLR-CS uses simple sensitivity maps calculated by dividing the images of each channel by the sum of squares image. Second, the size of the central part of the k-space was set based on Zhang et al.'s (2020) study and therefore on their magnitudefiltered approach. This size, important for the learning step of the KLR-CS, could be optimized for the proposed LRP approach. To further improve the quality of the reconstruction, the LRP KLR-CS approach could also be challenged by different theoretical frameworks, such as manifold-modeled signal recovery algorithms (Poddar and Jacob, 2016) or model-based methods with spatial and parametric constraints (Zhu et al., 2017). Further work is required to evaluate the combination of CS with parallel imaging (Griswold et al., 2002). Note that additional regularization along the diffusion direction could be performed with the BART toolbox (Uecker et al., 2021). This would require further tuning of the CS parameters but could improve the BART results.

With our data and choice of parameters, both BART-CS and KLR-CS reconstructions yielded lower errors than that reported by Zhang et al.'s (2020) study, which was obtained on simulated CS data, using equivalent ROIs but with magnitude-filtered KLR-CS (Supplementary Figure 3 in our study; Figure 3 in Zhang et al., 2020). The difference could be related to the difference in acquisition conditions (Supplementary Table 1), but lower errors remain nevertheless encouraging. Regarding major white-matter tracts, the main shape is always conserved in both simple (anterior commissure) and more complex (fornix) tracts when using CS acquisitions, for all AFs studied. The appearance of some falsepositive fibers, especially in the fornix, can partially be controlled by the addition of exclusion ROIs. For AF = 4 and both CS approaches, further adjustment of tractography parameters, use of deep-learning-based versions of CS (Cao et al., 2020; Dar et al., 2020; Baul et al., 2021; Xie and Li, 2022), and possibly introducing priors (Güngör et al., 2022; Korkmaz et al., 2022) could help improve the reconstruction, but they have not yet been applied to preclinical data. To allow comparisons with other reconstruction methods, the data acquired in this study have been made available.

The switch from simulations to acquisitions increased the observed error and, consequently, the observed SSIM was decreased. Additional fluctuations with diffusion directions were also observed. We, therefore, analyzed the different contributions to this increase in error and fluctuations.

First, we characterized the repeatability (Supplementary Figure 2). Indeed, when using simulations, the values in the undersampled dataset are exactly the same as in the FS datasets. In the case of acquisitions, the values in the undersampled dataset differ from that in the FS dataset. We observed that this repetition error ($7.97 \pm 0.57\%$ for FA, $1.50 \pm 0.19\%$ for MD) was about twice the error of the KLR-CS reconstruction in simulations (AF = 2; $4.79 \pm 0.12\%$ for FA, $0.71 \pm 0.02\%$ for MD, Figure 8). The repetition noise appears as the main contribution to the difference between acquisitions and simulations.

Second, there are differences in acquisition conditions between simulated and acquired data. These differences include the effect of the frequency drift (about 10% of the error) and the small reduction in pSNR (about 13%), which also contributes to increasing the error on the derived parameters (FA, MD, and tracts). To limit the impact of the frequency drift, a navigator could be introduced in the MRI sequence (Vos et al., 2017). Note that, concerning the frequency drift, the FS dataset is more blurred than the undersampled ones. The "error" is therefore not always in the CS datasets.

Third, the spatial correction, applied to limit the effect of the frequency drift during acquisition, could be improved. Here, we used an algorithm to modify the phase in k-space and thereby shift the image, to linearly register DWI images to the reference. However, the effect of the difference in acquisition times between FS and CS acquisitions on image distortion remains to be evaluated and, if necessary, corrected for. In addition, the MRtrix3 probabilistic reconstruction can add some noise to the fiber tract reconstructions. Fifty repetitions of the MRtrix3 reconstruction on the same dataset led to 1% repetition variability on mean fiber length — a small but not negligible contribution to the error.

To summarize, estimates made on acquired data are in good agreement with simulations, in which only CS reconstruction noise is present. Altogether, our results suggest that a reduction in scan time by a factor of 4 is acceptable in our experimental setting, in line with previous preclinical (Wang et al., 2018; Zhang et al., 2020) and clinical results (Wu et al., 2014; Zhu et al., 2017). Further analysis includes increasing the number of directions and the spatial resolution (Wang et al., 2018; Allan Johnson et al., 2019) to achieve higher AFs without degrading image quality (Lazarus et al., 2019) and limiting the increase in scan time as the blurring for KLR-CS is in both the spatial and diffusion directions. Indeed, this reconstruction approach should benefit from a larger learning set in both spatial and diffusion dimensions. A concurrent optimization of the spatial resolution and the number of diffusion directions could thus be performed (Wang et al., 2018).

There were some limitations to the study. We used only three animals per group, given that our objective was not biological but methodological; this small number limits the power of the statistical analysis. Further studies are required to fully characterize this mouse model. Similarly, we focused on a few ROI and a few tracts, in parallel to 3D FA and MD maps. More tracts could be analyzed based on the data that have been shared (see section "Data availability statement"). Moreover, acquisitions were performed using only one MRI scanner. A multicentric study would be more appropriate to estimate acquisition and reconstruction errors as the choice of hardware may affect the outcomes (Deruelle et al., 2020). Application to Human would also be of interest.

5. Conclusion

In conclusion, LRP KLR-CS reconstruction of undersampled preclinical DWI data seems a robust approach to reducing scan time, thereby limiting the effect of the frequency drift, while maintaining accurate estimates of FA, MD, and tracts.

Data availability statement

The datasets presented in this study can be found in online repositories. Raw data and code are available at https://github.com/nifm-gin/compressedSensing.

Ethics statement

This animal study was reviewed and approved by the Comité Local GIN, C2EA-04–APAFIS number 21234-2019031308592774.

Author contributions

DS, J-CD, and HM acquired the data. DS processed the data and performed the statistical analysis. DS and EB wrote the first draft of the manuscript. All authors contributed to conception and design of the study, manuscript revision, read, and approved the submitted version.

Funding

This work was supported by the French National Research Agency in the framework of the "Investissements d'Avenir" program (ANR-15-IDEX-02). The MRI facility IRMaGe is partly funded by the French National Research Agency, grant Infrastructure d'Avenir en Biologie Santé (ANR-11-INBS-0006).

Acknowledgments

We are particularly grateful to Dr. Leslie Ying, Ukash Nakarmi, and Chaoyi Zhang at the University of New York at Buffalo, for their toolbox for KLR compressed sensing reconstruction and

References

Aggarwal, M., Mori, S., Shimogori, T., Blackshaw, S., and Zhang, J. (2010). Three-dimensional diffusion tensor microimaging for anatomical characterization of the mouse brain. *Magn. Reson. Med.* 64, 249–261. doi: 10.1002/mrm.2 2426

Allan Johnson, G., Wang, N., Anderson, R., Chen, M., Cofer, G., Gee, J., et al. (2019). Whole mouse brain connectomics. J. Comp. Neurol. 527, 2146–2157. doi: 10.1002/cne.24560

Andrieux, A., Salin, P., Vernet, M., Kujala, P., Baratier, J., Gory-Fauré, S., et al. (2002). The suppression of brain cold-stable microtubules in mice induces synaptic defects associated with neuroleptic-sensitive behavioral disorders. *Genes Dev.* 16, 2350–2364. doi: 10.1101/gad.223302

Aydogan, D., Jacobs, R., Dulawa, S., Thompson, S., Francois, M., Toga, A., et al. (2018). When tractography meets tracer injections: A systematic study of trends and variation sources of diffusion-based connectivity. *Brain Struct. Funct.* 223, 2841–2858. doi: 10.1007/s00429-018-1663-8

Baul, A., Wang, N., Zhang, C., Ying, L., Chang, Y., and Nakarmi, U. (2021). Selflearned kernel low rank approach to accelerated high resolution 3D diffusion MRI. *Arvix* [Preprint]. doi: 10.48550/ARXIV.2110.08622

Behrens, T., Berg, H., Jbabdi, S., Rushworth, M., and Woolrich, M. (2007). Probabilistic diffusion tractography with multiple fibre orientations: What can we gain? *Neuroimage* 34, 144–155. doi: 10.1016/j.neuroimage.2006.09.018

Bilgic, B., Setsompop, K., Cohen-Adad, J., Wedeen, V., Wald, L., and Adalsteinsson, E. (2012). Accelerated diffusion spectrum imaging with compressed sensing using adaptive dictionaries. *Med. Image Comput. Comput. Assist. Interv.* 15, 1–9. doi: 10. 1007/978-3-642-33454-2_1

Calabrese, E., Badea, A., Cofer, G., Qi, Y., and Johnson, G. A. A. (2015). Diffusion MRI tractography connectome of the mouse brain and comparison with neuronal tracer data. *Cereb. Cortex* 25, 4628–4637. doi: 10.1093/cercor/bhv121

Cao, Y., Wang, L., Huang, J., Cheng, X., Zhang, J., Yang, F., et al. (2020). CS-GAN for high-quality diffusion tensor imaging. *Review* [Preprint]. doi: 10.21203/rs.3.rs-655 72/v1

the support to handle it and Sylvie Gory-Fauré for providing the WT and $MAP6^{+/-}$ mice. We thank Dr. Sandrine Humbert, from INSERM/Université Grenoble Alpes, for her support to J-CD and DS.

Conflict of interest

EB was a consultant at Bruker BioSpin.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fnins.2023. 1172830/full#supplementary-material

Caruyer, E., Lenglet, C., Sapiro, G., and Deriche, R. (2013). Design of multishell sampling schemes with uniform coverage in diffusion MRI. *Magn. Reson. Med.* 69, 1534–1540. doi: 10.1002/mrm.24736

Dar, S., Özbey, M., Çatlı, A., and Çukur, T. A. (2020). Transfer-learning approach for accelerated MRI using deep neural networks. *Magn. Reson. Med.* 84, 663–685. doi: 10.1002/mrm.28148

Deloulme, J., Gory-Fauré, S., Mauconduit, F., Chauvet, S., Jonckheere, J., Boulan, B., et al. (2015). Microtubule-associated protein 6 mediates neuronal connectivity through Semaphorin 3E-dependent signalling for axonal growth. *Nat. Commun.* 6:7246. doi: 10.1038/ncomms8246

Deruelle, T., Kober, F., Perles-Barbacaru, A., Delzescaux, T., Noblet, V., Barbier, E., et al. (2020). A multicenter preclinical MRI study: Definition of rat brain relaxometry reference maps. *Front. Neuroinform.* 14:22. doi: 10.3389/fninf.2020.00022

Gatto, R., Amin, M., Finkielsztein, A., Weissmann, C., Barrett, T., Lamoutte, C., et al. (2019). Unveiling early cortical and subcortical neuronal degeneration in ALS mice by ultra-high field diffusion MRI. *Amyotroph. Lateral Scler. Frontotemporal. Degener.* 20, 549–561. doi: 10.1080/21678421.2019.1620285

Gimenez, U., Boulan, B., Mauconduit, F., Taurel, F., Leclercq, M., Denarier, E., et al. (2017). 3D imaging of the brain morphology and connectivity defects in a model of psychiatric disorders: MAP6-KO mice. *Sci. Rep.* 7:10308. doi: 10.1038/s41598-017-10544-2

Gimenez, U., Mauconduit, F., Boulan, B., Denarier, E., Brocard, J., Gory-Fauré, S., et al. (2016). "Microscopic DTI for quantitative tractography of MAP6-KO mice: Validation by fluorescent microscopy on cleared brains," in *ISMRM 24th Annual Meeting & Exhibition* (Toronto, ON: ISMRM).

Griswold, M., Jakob, P., Heidemann, R., Nittka, M., Jellus, V., Wang, J., et al. (2002). Generalized autocalibrating partially parallel acquisitions (GRAPPA). *Magn. Reson. Med.* 47, 1202–1210. doi: 10.1002/mrm.10171

Güngör, A., Dar, S., Öztürk, Ş., Korkmaz, Y., Elmas, G., Özbey, M., et al. (2022). Adaptive Diffusion priors for accelerated MRI reconstruction. *Arvix* [Preprint]. doi: 10.48550/ARXIV.2207.05876 Hellings, C., Gogler, P., and Utschick, W. (2015). "Composite real principal component analysis of complex signals," in *2015 23rd European Signal Processing Conference (EUSIPCO)* (Piscataway, NJ: IEEE), 2216–2220. doi: 10.1109/EUSIPCO. 2015.7362778

Korkmaz, Y., Dar, S., Yurt, M., Ozbey, M., and Cukur, T. (2022). Unsupervised MRI Reconstruction via Zero-Shot Learned Adversarial Transformers. *IEEE Trans. Med. Imaging* 41, 1747–1763. doi: 10.1109/TMI.2022.3147426

Lazarus, C., Weiss, P., Chauffert, N., Mauconduit, F., El Gueddari, L., Destrieux, C., et al. (2019). SPARKLING: Variable-density k-space filling curves for accelerated T2* -weighted MRI. *Magn. Reson. Med.* 81, 3643–3661. doi: 10.1002/mrm.27678

Le Bihan, D., and Johansen-Berg, H. (2012). Diffusion MRI at 25: Exploring brain tissue structure and function. *Neuroimage* 61, 324–341. doi: 10.1016/j.neuroimage. 2011.11.006

Lee, H., Zhou, X., Li, Z., and Chuang, K. (2021). Optimizing diffusion MRI acquisition efficiency of rodent brain using simultaneous multislice EPI. *NMR Biomed.* 34:e4398. doi: 10.1002/nbm.4398

Lustig, M., Donoho, D., and Pauly, J. (2007). Sparse MRI: The application of compressed sensing for rapid MR imaging. *Magn. Reson. Med.* 58, 1182–1195. doi: 10.1002/mrm.21391

Mani, M., Jacob, M., Guidon, A., Magnotta, V., and Zhong, J. (2015). Acceleration of high angular and spatial resolution diffusion imaging using compressed sensing with multichannel spiral data. *Magn. Reson. Med.* 73, 126–138. doi: 10.1002/mrm.2 5119

MathWorks (2022). *Structural similarity (SSIM) index for measuring image quality.* Avaliable online at: https://mathworks.com/help/images/ref/ssim.html (accessed on Aug 18, 2022).

Michailovich, O., Rathi, Y., and Dolui, S. (2011). Spatially regularized compressed sensing for high angular resolution diffusion imaging. *IEEE Trans. Med. Imaging* 30, 1100–1115. doi: 10.1109/TMI.2011.2142189

Moldrich, R., Pannek, K., Hoch, R., Rubenstein, J., Kurniawan, N., and Richards, L. (2010). Comparative mouse brain tractography of diffusion magnetic resonance imaging. *Neuroimage* 51, 1027–1036. doi: 10.1016/j.neuroimage.2010. 03.035

Nakarmi, U., Wang, Y., Lyu, J., Liang, D., and Ying, L. A. (2017). Kernel-based low-rank (KLR) Model for low-dimensional manifold recovery in highly accelerated dynamic MRI. *IEEE Trans. Med. Imaging* 36, 2297–2307. doi: 10.1109/TMI.2017. 2723871

Nishioka, C., Liang, H., Barsamian, B., and Sun, S. (2019). Amyloid-beta induced retrograde axonal degeneration in a mouse tauopathy model. *Neuroimage* 189, 180–191. doi: 10.1016/j.neuroimage.2019.01.007

Oguz, I., McMurray, M., Styner, M., and Johns, J. (2012). The translational role of diffusion tensor image analysis in animal models of developmental pathologies. *Dev. Neurosci.* 34, 5–19. doi: 10.1159/000336825

Parker, G., and Alexander, D. (2003). Probabilistic Monte Carlo based mapping of cerebral connections utilising whole-brain crossing fibre information. *Inf. Process. Med. Imaging* 18, 684–695. doi: 10.1007/978-3-540-45 087-0_57

Paxinos, G., and Franklin, K. (2019). Paxinos and Franklin's the mouse brain in stereotaxic coordinates, 5th Edn. Cambridge, MA: Academic Press.

Poddar, S., and Jacob, M. (2016). Dynamic MRI using smoothness regularization on manifolds (SToRM). *IEEE Trans. Med. Imaging* 35, 1106–1115. doi: 10.1109/TMI. 2015.2509245

Pruessmann, K., Weiger, M., Scheidegger, M., and Boesiger, P. (1999). Sense: Sensitivity encoding for fast MRI. *Magn. Reson. Med.* 42, 952–962.

Schilling, K., Nath, V., Blaber, J., Harrigan, R., Ding, Z., Anderson, A., et al. (2017). Effects of b-value and number of gradient directions on diffusion mri measures obtained with Q-ball Imaging. *Proc. SPIE Int. Soc. Opt. Eng.* 10133:101330N. doi: 10.1117/12.2254545

Senel, L., Kilic, T., Gungor, A., Kopanoglu, E., Guven, H., Saritas, E., et al. (2019). Statistically segregated k-space sampling for accelerating multiple-acquisition MRI. *IEEE Trans. Med. Imaging* 38, 1701–1714. doi: 10.1109/TMI.2019.2892378

Setsompop, K., Kimmlingen, R., Eberlein, E., Witzel, T., Cohen-Adad, J., McNab, J., et al. (2013). Pushing the limits of in vivo diffusion MRI for the human connectome project. *Neuroimage* 80, 220–233. doi: 10.1016/j.neuroimage.2013.05. 078

Tournier, J., Calamante, F., and Connelly, A. (2007). Robust determination of the fibre orientation distribution in diffusion MRI: Non-negativity constrained super-resolved spherical deconvolution. *Neuroimage*35, 1459–1472. doi: 10.1016/j. neuroimage.2007.02.016

Tournier, J., Calamante, F., and Connelly, A. (2013). Determination of the appropriate b value and number of gradient directions for high-angular-resolution diffusion-weighted imaging: Appropriate *b* value and number of gradient directions for hardi. *NMR Biomed.* 26, 1775–1786. doi: 10.1002/nbm.3017

Tournier, J., Calamante, F., Gadian, D., and Connelly, A. (2004). Direct estimation of the fiber orientation density function from diffusion-weighted MRI data using spherical deconvolution. *Neuroimage* 23, 1176–1185. doi: 10.1016/j.neuroimage.2004. 07.037

Uecker, M., Holme, C., Blumenthal, M., Wang, X., Tan, Z., Scholand, N., et al. (2021). *mrirecon/bart: Version 0.7.00 (v0.7.00)*. Genéve: Zenodo. Available online at: https://doi.org/10.5281/zenodo.4570601

Uecker, M., Lai, P., Murphy, M., Virtue, P., Elad, M., Pauly, J., et al. (2014). ESPIRiT–an eigenvalue approach to autocalibrating parallel MRI: Where SENSE meets GRAPPA. *Magn. Reson. Med.* 71, 990–1001. doi: 10.1002/mrm.24751

Uecker, M., Ong, F., Tamir, J., Bahri, D., Virtue, P., Cheng, J., et al. (2015). Berkeley Advanced Reconstruction Toolbox. ISMRM 2015. *Proc. Soc. Mag. Reson. Med.* 23:2486.

Vos, S., Tax, C., Luijten, P., Ourselin, S., Leemans, A., and Froeling, M. (2017). The importance of correcting for signal drift in diffusion MRI. *Magn. Reson. Med.* 77, 285–299. doi: 10.1002/mrm.26124

Wang, N., Anderson, R., Badea, A., Cofer, G., Dibb, R., Qi, Y., et al. (2018). Whole mouse brain structural connectomics using magnetic resonance histology. *Brain Struct. Funct.* 223, 4323–4335. doi: 10.1007/s00429-018-1750-x

Wu, Y., Zhu, Y., Tang, Q., Zou, C., Liu, W., Dai, R., et al. (2014). Accelerated MR diffusion tensor imaging using distributed compressed sensing. *Magn. Reson. Med.* 71, 763–772. doi: 10.1002/mrm.24721

Xie, Y., and Li, Q. A. (2022). Review of deep learning methods for compressed sensing image reconstruction and its medical applications. *Electronics* 11:586. doi: 10.3390/electronics11040586

Zhang, C., Arefin, T., Nakarmi, U., Lee, C., Li, H., Liang, D., et al. (2020). Acceleration of three-dimensional diffusion magnetic resonance imaging using a kernel low-rank compressed sensing method. *Neuroimage* 210:116584. doi: 10.1016/ j.neuroimage.2020.116584

Zhang, J., Jones, M., McMahon, M., Mori, S., and Calabresi, P. (2012). In vivo and ex vivo diffusion tensor imaging of cuprizone-induced demyelination in the mouse corpus callosum. *Magn. Reson. Med.* 67, 750–759. doi: 10.1002/mrm. 23032

Zhu, Y., Peng, X., Wu, Y., Wu, E., Ying, L., Liu, X., et al. (2017). Direct diffusion tensor estimation using a model-based method with spatial and parametric constraints. *Med. Phys.* 44, 570–580. doi: 10.1002/mp.12054

16