### THE UNIVERSITY OF RHODE ISLAND

#### University of Rhode Island DigitalCommons@URI

**Biological Sciences Faculty Publications** 

**Biological Sciences** 

2013

# Examination of *Ulva* bloom species richness and relative abundance reveals two cryptically cooccurring bloom species in Narragansett Bay, Rhode Island

Michele Guidone

Carol S. Thornber University of Rhode Island, thornber@uri.edu

Follow this and additional works at: https://digitalcommons.uri.edu/bio\_facpubs

Terms of Use All rights reserved under copyright.

#### Citation/Publisher Attribution

Michele Guidone, Carol S. Thornber, Examination of Ulva bloom species richness and relative abundance reveals two cryptically cooccurring bloom species in Narragansett Bay, Rhode Island. *Harmful Algae, 24*, April 2013, Pages 1-9. Available at: http://dx.doi.org/10.1016/j.hal.2012.12.007

This Article is brought to you for free and open access by the Biological Sciences at DigitalCommons@URI. It has been accepted for inclusion in Biological Sciences Faculty Publications by an authorized administrator of DigitalCommons@URI. For more information, please contact digitalcommons@etal.uri.edu.

1	
2	
3	Examination of Ulva bloom species richness and relative abundance reveals two cryptically co-
4	occurring bloom species in Narragansett Bay, Rhode Island
5	
6	Michele Guidone <sup>1, 2</sup> * and Carol S. Thornber <sup>1</sup>
7	
8	<sup>1</sup> University of Rhode Island, Department of Biological Sciences, 120 Flagg Road, Kingston, RI,
9	USA 02881
10	<sup>2</sup> Current address: Sacred Heart University, Biology Department, 5151 Park Avenue, Fairfield,
11	CT, USA 06492
12	* Corresponding author: guidonem@sacredheart.edu
13	Phone (203) 396-8492
14	Fax: (203) 365-4785
15	
16	Carol Thornber: thornber@uri.edu
17	

18 Abstract

19 Blooms caused by the green macroalga *Ulva* pose a serious threat to coastal ecosystems 20 around the world. Despite numerous studies of the causes and consequences of these blooms, we 21 still have a limited understanding of Ulva bloom species richness and abundance due to 22 difficulties in identifying *Ulva* species using morphological features. Along the northeastern U.S. 23 coastline, all blooms of distromatic Ulva blades were previously identified as U. lactuca. Recent molecular sequencing, however, discovered the presence of additional distromatic Ulva species. 24 25 Therefore, in order to determine the relative abundance of *Ulva* species within blooms, we 26 conducted monthly surveys at four Narragansett Bay, RI, sites representing a gradient of bloom 27 severity. We found that the biomass of *Ulva* within blooms was a mix of *U. compressa* and *U.* 28 *rigida*, not *U. lactuca* as previously reported. In contrast, sites not impacted by blooms that were 29 located near the mouth of Narragansett Bay were dominated by U. lactuca. We also observed 30 spatial and temporal differences in *Ulva* and total macroalgal diversity between bloom-impacted 31 sites, indicating that *Ulva* bloom composition can be radically different between similar sites 32 within close proximity. We discuss our results in the context of Ulva blooms worldwide, 33 highlighting the need to definitively determine bloom species composition in order to fully 34 understand bloom dynamics. 35 Key words: biomass, diversity, eutrophication, macroalgal bloom, survey, Ulva 36 37

38 1. Introduction

The formation of blooms of filamentous and/or thin foliose macroalgae are frequently a
consequence of coastal eutrophication (Fletcher, 1996; Valiela et al., 1997; Morand and

Merceron, 2005; Ye et al., 2011). Macroalgae with these morphologies have a high surface area
to volume ratio that enables them to rapidly uptake nutrients for greatly increased growth (Littler
and Littler, 1980; Hein et al., 1995; Pedersen and Borum, 1996), provided favorable bathymetric,
temperature, and light conditions exist (Rivers and Peckol, 1995; Taylor et al., 2001; Cohen and
Fong, 2004; Sousa et al., 2007; Liu et al., 2010).

46 Bloom macroalgae often form large floating mats in the water column, in which 47 individual thalli grow, fragment, and asexually reproduce via zoospores (Gao et al., 2010; Ye et 48 al., 2011). These floating mats of algae alter coastal light, nutrient, and water flow conditions, 49 causing decreases in perennial algae, seagrasses, and benthic invertebrates (Valiela et al., 1997; 50 Hauxwell et al., 1998, 2001; Thomsen and McGlathery, 2006; Worm and Lotze, 2006). Nightly 51 respiration and decomposition of bloom macroalgae contribute to hypoxic events (Valiela et al., 52 1997; Raffaelli et al., 1998) that can result in substantial mortality of invertebrates and fishes 53 (Deacutis et al., 2006; Berezina et al., 2007). In addition, several species produce toxins that 54 negatively impact co-occurring organisms (Nelson et al., 2003a; Eklund et al., 2005; Van 55 Alstyne et al., 2006). Moreover, blooms interfere with coastal commercial and recreational 56 activities (Lee and Olsen, 1985; Thomsen and McGlathery, 2006; Deacutis, 2008; Leliaert et al., 57 2009).

Bloom-forming macroalgal species can be found within the phyla Chlorophyta,
Heterokontophyta, and Rhodophyta, but most macroalgal blooms, including the largest ever
recorded, are caused by Chlorophyta species, such as those within the genus *Ulva* Linnaeus
(Fletcher, 1996; Valiela et al., 1997; Morand and Merceron, 2005). For example, the 2008 bloom
of *Ulva prolifera* offshore of Qingdao, China contained an estimated 20 million wet tons of algae
spanning approximately 13,000 km<sup>2</sup> in the Yellow Sea. This bloom required the removal of more

than 1 million tons of *U. prolifera* from the shoreline, at a cost of over \$100 million US dollars
(Leliaert et al., 2009; Gao et al., 2010).

*Ulva* species are notoriously difficult to identify due to a lack of distinguishing 66 67 morphological features among species and a tremendous degree of phenotypic plasticity within 68 species (Blomster et al., 1999; Blomster et al., 2002; Leskinen et al., 2004). Until recently, this 69 morphological uncertainty hindered our ability to accurately assess species richness within Ulva 70 blooms. In the last decade, however, numerous molecular studies from bloom and non-bloom 71 impacted habitats around the world have greatly increased our understanding of Ulva richness 72 (e.g. Hayden et al., 2003; Leliaert et al., 2009; Kraft et al., 2010; Liu et al., 2010). However, 73 detailed surveys of the relative abundance of different Ulva species, as well as physiological and 74 ecological studies utilizing molecularly confirmed *Ulva* species, remain lacking (but see Liu et 75 al., 2010; Yokoyama and Ishihi, 2010; Kim et al., 2011). These knowledge gaps pose a serious 76 barrier in our ability to understand *Ulva* bloom dynamics, and consequently hinder the 77 development of macroalgal bloom risk assessments and well-informed coastal management 78 practices.

We conducted extensive surveys at four Narragansett Bay, Rhode Island sites (Figure 1), to determine: 1) which *Ulva* species is (are) the main contributor(s) to *Ulva* blooms in Narragansett Bay; 2) if bloom-forming *Ulva* species are found throughout Narragansett Bay or only in bloom-impacted areas; and 3) how the species richness and relative abundance of all macroalgal species varies amongst bloom and non-bloom sites. We discuss our results in the context of previously studied *Ulva* bloom systems and highlight the importance of determining their species composition for understanding bloom dynamics.

#### 87 **2. Methods**

#### 88 2.1. Study locale and species

89 Narragansett Bay is a well-studied estuary in which annual blooms of distromatic Ulva 90 blades and, less often, monostromatic tubular Ulva spp. (formerly Enteromorpha, Hayden et al., 91 2003) and *Gracilaria* spp. occur in the anthropogenically impacted northern portions of the bay 92 (Granger et al., 2000; Calabretta and Oviatt, 2008; Deacutis, 2008; Oczkowski et al., 2008; 93 Thornber and Guidone, unpublished data). While tubular Ulva species in Rhode Island cannot be 94 identified to the species level based on morphological features alone, significant progress has 95 been made in distinguishing between *Ulva* blades in this region. Originally identified as 96 monospecific blooms of U. lactuca, molecular sequencing of Ulva blades within Narragansett 97 Bay and along the outer Rhode Island coast detected three species of Ulva blades: U. compressa 98 Linnaeus, U. lactuca Linnaeus, and U. rigida C. Agardh (Guidone et al., unpublished data). 99 Similar results were found in molecular assessments of *Ulva* in the Great Bay Estuarine System 100 in New Hampshire and Maine (Hofmann et al. 2010). These blade-forming species can be 101 reliably distinguished based on a suite of cellular features including cell size, shape, and 102 arrangement, chloroplast position, and pyrenoid number (Guidone et al., unpublished data; 103 Hofmann et al., 2010).

104

105 2.2. Survey methodology

We first assessed the biomass of *Ulva compressa*, *U. lactuca*, and *U. rigida*, and the percent cover of all algal species throughout Narragansett Bay by conducting monthly surveys from May-September 2009 at four field sites: Brushneck Cove and Chepiwanoxet, Warwick, RI and The Graduate School of Oceanography (GSO) and Pier 5, Narragansett, RI (Figure 1).

110	Brushneck Cove and Chepiwanoxet are located in Greenwich Bay, a subestuary of Narragansett
111	Bay that experiences annual Ulva blooms (Granger et al., 2000; Thornber and Guidone,
112	unpublished data). Both Brushneck Cove and Chepiwanoxet are intertidal mud flats bordered by
113	fringing salt marshes; however, Chepiwanoxet has a longer water residence time (1.5 days versus
114	0.3 days), a greater mean depth, and experiences lower temperatures and a greater incidence of
115	hypoxic events (Granger et al., 2000; Thornber and Guidone, unpublished data). These sites were
116	contrasted to GSO and Pier 5 (Figure 1), which are near the mouth of Narragansett Bay and are
117	not impacted by annual Ulva blooms. GSO is a narrow beach of mixed sand and cobblestone,
118	while Pier 5 is a boulder field.
119	For comparison across seasons, additional surveys were conducted in February and May
120	2010 at all four sites. Monthly surveys were subsequently continued at the two bloom-impacted
121	sites from June 2010 to November 2011.
122	We conducted all surveys during spring low tides. For each survey, at each site, we
123	placed two 10 m transects parallel to the shore; transects were at least 30 m apart and placed at
124	the same tidal height. Transect positioning corresponded to the tidal height where Ulva wrack
125	was most commonly observed at low tide; this was directly below the fringing marsh at
126	Brushneck Cove and Chepiwanoxet, and directly above the waterline at GSO and Pier 5. For our
127	initial May-September surveys, we used a $0.25 \text{ m}^2$ quadrat to calculate the percent cover of all
128	algal taxa at one meter intervals along each transect, for a total of twenty replicate samples per
129	site and sampling month. Subsequent surveys assessed percent cover at two-meter intervals, for a
130	total of ten replicate samples per site and sampling month. Due to the overlap of macroalgal
131	thalli within drift mats, the total percent cover of all species often totaled more than 100%. Algae
132	were identified to the lowest taxonomic unit possible in the field. The Ulva blades within each

quadrat were collected and returned to the laboratory. Additionally, beginning in February 2010
we collected *Ulva* tubes from the bloom-impacted sites for biomass comparison to *Ulva* blades.
In the laboratory, we identified each blade to species using distinguishing cellular features
determined from molecularly confirmed voucher specimens (Guidone et al., unpublished data).
Following identification, blades were spun to a constant weight using a salad spinner and then
weighed.

139

140 2.3 Statistical analyses

141 We analyzed Ulva bloom biomass data for our 2009-2010 survey and our bloom-142 impacted sties (2010-2011) using fully factorial nested ANOVAs with fixed factors for month-143 year, site, species, and transect nested within site (JMP, version 8, SAS Institute Inc., North 144 Carolina, USA). We were unable to normalize our data via transformation, however the analysis 145 of variance test is robust to departures from normality and homogeneity of variances when 146 datasets are large. In this instance, our datasets were sufficiently large to ensure that our results 147 were not impacted by violating these assumptions (Underwood, 1997). 148 Percent cover data for our 2009-2010 and 2010-2011 surveys were used to calculate the 149 average Shannon-diversity index (H') and Pielou's evenness (J') for each site. Additionally, algal 150 percent cover was assessed for differences in taxa among sites and sampling months using a two-151 way crossed analysis of similarity (ANOSIM). The contribution of each taxon to the average

similarity and dissimilarity among sites and months was determined using a similarity of

153 percentages analysis (SIMPER). Prior to ANOSIM and SIMPER analysis, data were fourth-root

transformed to increase the importance of rare species; all analyses were conducted on Bray-

155 Curtis similarities. Non-parametric analyses were conducted using Primer-E (version 6, Primer-E
156 Ltd., Plymouth UK).

157

#### 158 **3. Results**

159 *3.1. Ulva species richness and relative abundance* 

Ulva lactuca was the dominant blade forming Ulva species at non-bloom sites, while U. *compressa* and U. *rigida* dominated at bloom sites. Overall, Ulva species biomass varied
significantly amongst sites and months (Table 1). In 2009-2010, U. lactuca was the only blade
species found at the non-bloom impacted Pier 5, with peak mean wet biomass of 75.48 g/m<sup>2</sup> in
May 2010 (Figure 2a). Ulva lactuca also dominated at GSO, where we only found small
fragments (< 0.4 g per piece) of U. compressa and U. rigida during three of the seven survey</li>
months (Figure 2b).

In contrast, over the entire course of this study (2009-2011), U. lactuca was rarely found 167 168 at either of our bloom-impacted sites, while U. compressa and U. rigida were consistently 169 present at both sites (Figure 3). The mean *Ulva* biomass was significantly greater at Brushneck 170 Cove than the other three sites during 2009-2010 (Tukey post-hoc test, p < 0.05); there was no 171 significant difference in biomass between Brushneck Cove and Chepiwanoxet in 2010-2011 (Table 1b). Additionally, while we observed no impact of transect placement during our 2009-172 173 2010 surveys, transect did have a significant impact on biomass at our bloom sites during 2010-174 2011 (Table 1).

At both bloom-impacted sites, biomass consistently peaked during June-July with
subsequent crashes in August (Figure 3). One exception to this pattern occurred during 2009 at
Brushneck Cove, which saw continued high biomass through September 2009 (Figure 3b). Total

mean wet biomass was greatest during June 2009 (882.80 g/m<sup>2</sup>; maximum observation 1,662.24  $g/m^2$ ) at Brushneck Cove and July 2010 (665.88 g/m<sup>2</sup>; maximum observation 1,804.60 g/m<sup>2</sup>) at Chepiwanoxet.

181 Throughout our surveys from 2009-2011, U. compressa was nearly always the largest 182 component of Ulva mats at Chepiwanoxet, except during April 2011, May of each year, and 183 Sept. 2009, when Ulva tubes dominated (Figures 3-5). By contrast, U. rigida biomass was 184 greater than U. compressa during 2009 and 2010 at Brushneck Cove, while summer peaks in 185 June and July 2011 were dominated by U. compressa. Low densities of tubular Ulva species 186 were only present at Brushneck Cove during May of each year and April 2011 (Figures 3-5). 187 Despite having reliable morphological descriptions for our three targeted *Ulva* species, 188 we were occasionally (< 12% of samples) unable to identify *Ulva* blades to species level. 189 Unidentifiable blades were more frequently encountered at Brushneck Cove and were more 190 abundant during the spring months (Figure 3). 191

192 *3.2. Total macroalgal species richness and abundance* 

193 *3.2.1. 2009-2010 survey* 

A total of 34 taxonomic groups were observed throughout our 2009-2010 surveys (Figure 4, Table 2). Species diversity (Shannon H') was highest at non-bloom Pier 5 and lowest at bloom-impacted Brushneck Cove (Table 3; one-way ANOVA  $F_{3, 456} = 91.40$ , p < 0.0001, Tukey post-hoc p < 0.05). However, no clear correlation between bloom-impacted and non-impacted sites was evident, as diversity was higher at bloom-impacted Chepiwanoxet than at GSO. In addition, evenness (Pielou's J') was highest at Chepiwanoxet. Of the 34 taxa recorded, 22 were found at only one or both of the lower-bay sites (Table 2). No individual taxa were unique to the 201 two bloom-impacted sites; however, Chepiwanoxet was the only site that contained mats of

202 intertwined Agardhiella subulata, Ceramium virgatum, Gracilaria spp., and Polysiphonia spp.

203 (hereafter ACGP mats). While these mats could reach greater than 50% cover, the thalli within

them were often small fragments, making separation of the component species impracticable.

205 Therefore, we considered these mats as a unique entity for this study.

206 Taxon assemblages were significantly different amongst all sites and sampling months 207 (ANOSIM, p = 0.001; Table 4). In concurrence with the Shannon diversity index, the largest 208 difference in algal composition was observed between bloom-impacted Brushneck Cove and 209 non-bloom Pier 5 (Table 4a), while the smallest difference was between bloom-impacted 210 Chepiwanoxet and non-bloom GSO. However, the high R-value and significance (p = 0.001) of 211 all pairwise tests between sites indicates strong separation of algal communities among all sites 212 (Table 4a). Differences in algal composition between the bloom and non-bloom impacted sites 213 were largely due to the greater percent cover of *Ulva* blades at the bloom impacted sites and the 214 presence of *Chondrus crispus* at the non-bloom sites (Figure 4, Table 5). In addition, while *Ulva* 215 tubes were found at all four sites, their occurrence and percent cover varied temporally (Figure 4, 216 Table 5).

Amongst months, the largest differences in flora were between February and August; the smallest differences were between July and August (Table 4b). Seasonal shifts in algal composition were apparent from June to July, August to September, and September to February (Figure 4, Table 4b).

223	As in the 2009-2010 survey, algal composition from June 2010-November 2011 differed
224	significantly between the two bloom-impacted sites (ANOSIM global $R = 0.272$ , $p = 0.001$ ),
225	with a greater diversity at Chepiwanoxet than at Brushneck Cove (Table 3). The dominant taxa at
226	both sites were also similar to 2009-2010, with Ulva blades dominating the algal community at
227	Brushneck Cove during all months except January-May 2011; Ulva blades, Ulva tubes, and
228	ACGP mats dominated at Chepiwanoxet (Figures 4, 5, Table 2). Taxa observed in 2010-2011
229	that were not previously observed during 2009-2010 included Ectocarpus spp., Porphyra spp.,
230	Cladophora spp., and Scytosiphon lomentaria (Table 2).
231	
232	4. Discussion
233	4.1. Ulva in Narragansett Bay
234	4.1.1. Species distribution
235	Our results demonstrate that U. compressa and U. rigida are responsible for the blooms
236	of Ulva blades in the northern portions of Narragansett Bay, RI. Neither of these bloom-forming
237	species was found at Pier 5, our field site closest to the open coast. In contrast, U. lactuca was
238	regularly observed at the two non-bloom sites near the mouth of Narragansett Bay and rarely
239	found at the bloom-impacted sites.
240	In a molecular assessment of Ulva species within the Great Bay Estuarine System of New
241	Hampshire and Maine, USA, Hofmann et al. (2010) found a similar distributional pattern for
242	these three species, with U. compressa and U. rigida typically occurring together at inner
243	estuarine sites and U. lactuca, along with U. pertusa, occurring at higher salinity sites closer to
244	the open coast. Their results, in conjunction with our own, suggest that U. compressa and U.
245	<i>rigida</i> are more physiologically and/or ecologically similar to each other than either is to U.

*lactuca*. Based on the distributional pattern of these species in Narragansett Bay and the Great
Bay Estuarine System, we hypothesize that *U. lactuca* is rarely found in northern Narragansett
Bay bloom-impacted sites due to a lower tolerance of high water temperatures, salinity
fluctuations, and/or hypoxia or other factors. Likewise, if *U. compressa* and *U. rigida* are
adapted to the abiotic conditions found in shallow, low-flow eutrophic estuaries, they might be
absent from open coastal areas due to nutrient limitations or intolerance to higher salinities or
wave exposure.

253 One alternate explanation exists for the distributional pattern of U. compressa. Tan et al. 254 (1999) observed that distromatic blades of U. compressa were concentrated in low salinity areas 255 of an estuary in Aberdeenshire, Scotland, while U. compressa with a tubular morphology was 256 found at higher salinity sites near the North Sea. Taking this into consideration, it is possible that 257 U. compressa in Narragansett Bay persists in lower salinity eutrophic areas as a distromatic 258 blade and is present at lower bay and outer coast sites as a monostromatic tube. Although we did 259 not identify tubular Ulva to species in this survey, prior molecular analysis of outer coast 260 samples supports this hypothesis (Guidone et al., unpublished data).

261 In addition to abiotic factors, *Ulva* species distribution may be restricted by differences in 262 *Ulva* palatability and/or herbivore communities amongst the study sites. Nelson et al. (2008) 263 found that Ulva and Ulvaria (both in the family Ulvaceae) differed in abiotic tolerances and 264 palatability, causing the more palatable but stress tolerant *Ulva* to dominate intertidally while the 265 unpalatable *Ulvaria* thrived in the herbivore populated subtidal. Similarly, blooms in the Baltic 266 Sea were dominated by the unpalatable Pilayella littoralis when herbivores were abundant and 267 the palatable *Ulva intestinalis* when herbivores were absent or nutrient levels were enriched 268 (Lotze et al., 2000; Lotze and Worm, 2000). Although not directly quantified in this study, based

- 269 on previous studies and our own field observations, herbivore communities between our bloom
- and non-bloom sites can be substantially different (Guidone et al., unpublished data).
- 271
- 272 *4.1.2. Bloom species relative abundance*

273 While similar densities of *Ulva* were found at both bloom-impacted sites during the 274 summers of 2010 and 2011, the relative abundance of each *Ulva* blade species, as well as the 275 proportion of tubular *Ulva* species present, differed significantly. These spatial and temporal 276 fluctuations indicate that even between eutrophic sites within close proximity (only 3.5 km 277 apart), small abiotic or biotic differences, or stochasticity, may lead to markedly different Ulva 278 bloom compositions. Nelson et al. (2003b) observed similar patterns amongst Ulva blades, Ulva 279 tubes, and *Ulvaria*, on a slightly larger scale in the Pacific Northwest. Our observation that *Ulva* 280 biomass differed amongst transects during 2010-2011 indicates that temporal changes in water 281 flow and/or wind patterns may play an important role in *Ulva* bloom deposition patterns in the 282 intertidal.

283

#### *4.2. Total macroalgal diversity at bloom and non-bloom sites*

As we had expected, Pier 5 had the highest diversity of the four sites sampled in 2009-286 2010. This site is closest to the open coast and likely receives drift from a large area of the lower 287 Narragansett Bay and open ocean sites. Pier 5 also has an abundance of hard substrata available 288 for algal attachment, unlike the other three field sites.

289 Contrary to our expectations, we did not find a strict pattern of high diversity (H') at non-290 bloom sites vs. low diversity at bloom-impacted sites, as Chepiwanoxet had the second highest 291 diversity of the four sites. This is particularly perplexing in light of environmental measurements

292 (dissolved oxygen, water residence time; Granger et al., 2000) that indicate Chepiwanoxet is the 293 most eutrophic of the four sites. Since the algae sampled at all sites was largely drift, it is unclear 294 whether the diversity observed at Chepiwanoxet is representative of the site itself, or if 295 circulation patterns deposit a wide diversity of species from adjacent areas. However, all areas 296 within close proximity to Chepiwanoxet are also bloom-impacted, suggesting that the diversity 297 observed at this site is truly representative of the bloom-impacted community. Furthermore, 298 Chepiwanoxet had the highest evenness of the four sites sampled from 2009-2010, which differs 299 from general patterns that indicate eutrophication has a larger negative impact on evenness than 300 species richness (Hillebrand et al., 2007). Similar results were found in subestuaries of Waquoit 301 Bay, Massachusetts, USA, where macroalgal bloom biomass (Cladophora vagabunda and 302 *Gracilaria tikvahiae*) was linked to nutrient enrichment, while species richness was not (Fox et 303 al., 2008).

304

#### 305 *4.3. Narragansett Bay blooms compared to Ulva blooms around the world*

306 To our knowledge, this is the first report of *Ulva* bloom biomass that has extensively 307 examined the relative contribution of cryptically co-occurring distromatic blade species 308 following molecular confirmation of the *Ulva* species present within an area (Guidone et al., 309 unpublished data). We are unaware of any previous estimates of *U. compressa* bloom biomass, 310 but reports of *Ulva* bloom biomass based solely on morphology have identified *U. rigida* as the 311 causative species of blooms in Europe (Sfriso et al., 1992; Coat et al., 1998; Balducci et al., 312 2001; Merceron and Morand, 2004 as U. armoricana) and the Philippines (Largo et al., 2004 as 313 U. armoricana). Ulva lactuca has been reported to bloom in North America (Lyons et al., 2009), 314 New Zealand (Park, 1992), and South Africa (Anderson et al., 1996). The density of

Narragansett Bay blooms fall within the range of densities reported for most blooms of *U. rigida* and *U. lactuca* (150-3,000 g/m<sup>2</sup> wet mass). One notable exception to this range is the bloom of *U. rigida* in the Venice Lagoon, Italy, reported to have a biomass range of 5-20 kg/m<sup>2</sup> wet mass (Schramm, 1999).

319 Given the difficulty in identifying *Ulva* species using morphology alone, and based on 320 our observation that U. rigida and U. compressa often bloom simultaneously, it is likely that 321 some prior reports of Ulva blooms have either misidentified the Ulva species involved or 322 underestimated the number of species present within the bloom. For example, a recent molecular 323 survey of *Ulva* in New Zealand found *U. lactuca* to be present at only 3 out of 195 sampled sites 324 (Heesch et al., 2009), indicating that New Zealand blooms are likely formed by another, more 325 abundant *Ulva* species. Additionally, based on a small sampling of *Ulva* blades within blooms in 326 Brittany, France, Merceron and Morand (2004) tentatively identified three co-occurring ulvoid 327 species (U. rigida as U. armoricana, U. rotundata, and Umbraulva olivascens as U. olivascens). 328 Unfortunately, even when armed with molecularly verified species descriptions, if morphological 329 features cannot be found to separate co-occurring species, detailed surveys of Ulva bloom 330 diversity such as we conducted will be challenging. 331

#### 332 5. Acknowledgements

333 We would like to thank C. Blewett, J. Bonamusa, K. Eldredge, L. Falconer, M. Gennazio,

A. Heinze, K. Hyman, B. Konkle, M. O'Brien, C. Newton, B. Pestone, S. Rinehart, N. Rohr, T.

335 Schollmeier, E. Vincent, M. Wands, and A. Ziegler for help with field collections and biomass

- 336 processing. We would also like to thank C. Newton and two anonymous reviewers for their
- 337 evaluation of earlier versions of this manuscript. Funding for this research was provided by Bay

338	Window (NOAA), the Rhode Island Natural History Survey, Rhode Island Sea Grant, the
339	Sounds Conservancy Quebec-Labrador Foundation, and the University of Rhode Island. This
340	material is based, in part, upon work supported in part by the National Science Foundation
341	EPSCoR Cooperative Agreement #EPS-1004057 and the State of Rhode Island.
342	
343	6. References
344	Anderson, R.J., Monteiro, P.M.S., Levitt, G.J., 1996. The effect of localised
345	eutrophication on competition between Ulva lactuca (Ulvaceae, Chlorophyta) and a
346	commercial resource of Gracilaria verrucosa (Gracilariaceae, Rhodophyta).
347	Hydrobiologia 326/327, 291-296.
348	Balducci, C., Sfriso, A., Pavoni, B., 2001. Macrofauna impact on Ulva rigida C. Ag.
349	production and the relationship with environmental variables in the lagoon of Venice.
350	Mar. Environ. Res. 52, 27-49.
351	Berezina, N.A., Tsiplenkina, I.G., Pankova, E.S, Gubelit, J.I., 2007. Dynamics of
352	invertebrate communities on the stony littoral of the Neva Estuary (Baltic Sea) under
353	macroalgal blooms and bioinvasions. Transitional Waters Bulletin 1, 65-76.
354	Blomster, J., Bäck, S., Fewer, D.P., Kiirikki, M., Lehvo, A., Maggs, C.A., Stanhope,
355	M.J., 2002. Novel morphology in Enteromorpha (Ulvophyceae) forming green tides. J.
356	Bot. 89, 1756-1763.
357	Blomster, J., Maggs, C.A., Stanhope, M.J., 1999. Extensive intraspecific
358	morphological variation in Enteromorpha muscoides (Chlorophyta) revealed by
359	molecular analysis. J. Phycol. 35, 575-586.
200	Calabratta C.I. Oriett C.A. 2008 The response of hearthis means from to

360 Calabretta, C.J., Oviatt, C.A., 2008. The response of benthic macrofauna to

361	anthropogenic stress in Narragansett Bay, Rhode Island: a review of human stressors and
362	assessment of community conditions. Mar. Pollut. Bull. 56, 1680-1695.
363	Coat, G., Dion, P., Noailles, MC., de Reviers, B., Fontaine, JM., Berger-Perrot, Y.,
364	Loiseaux-de Goér, S., 1998. Ulva armoricana (Ulvales, Chlorophyta) from the coasts of
365	Brittany (France). II. Nuclear rDNA ITS sequence analysis. Eur. J. Phycol. 33, 81-86.
366	Cohen, R.A., Fong, P., 2004. Physiological responses of a bloom-forming green
367	macroalga to short-term change in salinity, nutrients, and light help explain its ecological
368	success. Estuaries 27, 209-216.
369	Deacutis, C., 2008. Evidence of ecological impacts from excess nutrients in upper
370	Narragansett Bay, In: Desbonnet, A., Costa-Pierce, B.A. (Eds.), Science for Ecosystem-
371	Based Management. Springer, New York, pp. 349-381.
372	Deacutis, C., Murray, D., Prell, W., Saarman, E., Korhun, L., 2006. Hypoxia in the
373	upper half of Narragansett Bay, RI, during August 2001 and 2002. Northeast. Nat. 13,
374	173-198.
375	Eklund, B., Svensson, A.P., Jonsson, C., Malm, T., 2005. Toxic effects of
376	decomposing red algae on littoral organisms. Estuar. Coast. Shelf S. 62, 621-626.
377	Fletcher, R.L., 1996. The occurrence of green tides a review, In: Schramm, W.,
378	Niehuis, P.K. (Eds.), Marine benthic vegetation: recent changes and the effects of
379	eutrophication. Springer, Berlin, pp. 7-43.
380	Fox, S.E., Stieve, E., Valiela, I., Hauxwell, J., McClelland, J., 2008. Macrophyte
381	abundance in Waquoit Bay: effects of land-derived nitrogen loads on seasonal and multi-
382	year biomass patterns. Estuar. Coast. 31, 532-541.
383	Gao, S., Chen, X., Yi, Q., Wang, G., Pan, G., Lin, A., Peng, G., 2010. A strategy for

385 of sporangia by fragmentation. PLoS ONE 5, e8571. 386 Granger, S.L., Mark, J., Buckley, B.A., Schwartz, M., 2000. An assessment of 387 eutrophication in Greenwich Bay. Rhode Island Sea Grant, Narragansett. 388 Hauxwell, J., Cebrián, J., Furlong, C., Valiela, I., 2001. Macroalgal canopies 389 contribute to eelgrass (*Zostera marina*) decline in temperate estuarine ecosystems. 390 Ecology 82, 1007-1022. 391 Hauxwell, J., McClelland, J., Behr, P.J., Valiela, I., 1998. Relative importance of 392 grazing and nutrient controls of macroalgal biomass in three temperate shallow estuaries. 393 Estuaries 21, 347-360. 394 Hayden, H.S., Blomster, J., Maggs, C.A., Silva, P.C., Stanhope, M.J., Waaland, J.R., 395 2003. Linnaeus was right all along: *Ulva* and *Enteromorpha* are not distinct genera. Eur. 396 J. Phycol. 38, 277-294. 397 Heesch, S., Broom, J.E.S., Neill, K.F., Farr, T.J., Dalen, J.L., Nelson, W.A., 2009. 398 Ulva, Umbraulva, and Gemina: genetic survey of New Zealand taxa reveals diversity and 399 introduced species. Eur. J. Phycol. 44, 143-154. 400 Hein, M., Pedersen, M.F., Sand-Jensen, K., 1995. Size dependent nitrogen uptake in 401 micro- and macroalgae. Mar. Ecol. Prog. Ser. 118, 247-253. 402 Hillebrand, H., Gruner, D.S., Borer, E.T., Bracken, M.E.S., Cleland, E.E., Elser, J.J., 403 Harpole, W.S., Ngai, J.T., Seabloom, E.W., Shurin, J.B., Smith, J.E., 2007. Consumer 404 versus resource control of producer diversity depends on ecosystem type and producer

the proliferation of *Ulva prolifera*, main causative species of green tides, with formation

405 community structure. P. Natl. Acad. Sci. 104, 10904-10909.

384

406 Hofmann, L.C., Nettleton, J.C., Neefus, C.D., Mathieson, A.C., 2010. Cryptic

407	diversity of Ulva (Ulvales, Chlorophyta) in the Great Bay Estuarine System (Atlantic
408	USA): introduced and indigenous distromatic species. Eur. J. Phycol. 45, 230-239.
409	Kim, JH., Kang, E.J., Park, M.G., Lee, BG., Kim, K.Y., 2011. Effects of
410	temperature and irradiance on photosynthesis and growth of a green-tide-forming species
411	(Ulva linza) in the Yellow Sea. J. App. Phycol. 23, 421-432.
412	Kraft, L.G.K., Kraft, G.T., Waller, R.F., 2010. Investigations in southern Australian
413	Ulva (Ulvophyceae, Chlorophyta) taxonomy and molecular phylogeny indicate both
414	cosmopolitanism and endemic cryptic species. J. Phycol. 46, 1257-1277.
415	Largo, D.B., Sembrano, J., Hiraoka, M., Ohno, M., 2004. Taxonomic and ecological
416	profile of 'green tide' species of Ulva (Ulvales, Chlorophyta) in central Philippines.
417	Hydrobiologia 512, 247-253.
418	Lee, V., Olsen, S., 1985. Eutrophication and management initiatives for the control of
419	nutrient inputs to Rhode Island coastal lagoons. Estuaries 8, 191-202.
420	Leliaert, F., Zhang, X., Ye, N., Malta, E., Engelen, A., Mineur, F., Verbruggen, H., De
421	Clerck, O., 2009. Research note: Identity of the Qingdao algal bloom. Phycol. Res. 57,
422	147-151.
423	Leskinen, E., Alström-Rapaport, C., Pamilo, P., 2004. Phylogeographical structure,
424	distribution and genetic variation of the green algae Ulva intestinalis and U. compressa
425	(Chlorophyta) in the Baltic Sea area. Mol. Ecol. 13, 2257-2265.
426	Littler, M.M., Littler, D.S., 1980. The evolution of thallus form and survival strategies
427	in benthic marine macroalgae: field and laboratory tests of a functional form model. Am.
428	Nat. 116, 25-44.
429	Liu, D., Keesing, J.K., Dong, Z., Zhen, Y., Di, B., Shi, Y., Fearns, P., Shi, P., 2010.

430	Recurrence of the world's largest green-tide in 2009 in Yellow Sea, China: Porphyra
431	yezoensis aquaculture rafts confirmed as nursery for macroalgal blooms. Mar. Pollut.
432	Bull. 60, 1423-1432.
433	Lotze, H., Worm, B., Sommer, U., 2000. Propagule banks, herbivory and nutrient
434	supply control population development and dominance patterns in macroalgal blooms.
435	Oikos 89, 46-58.
436	Lotze, H.K., Worm, B., 2000. Variable and complementary effects of herbivores on
437	different life stages of bloom-forming macroalgae. Mar. Ecol. Prog. Ser. 200, 167-175.
438	Lyons, P., Thornber, C., Portnoy, J., Gwilliam, E., 2009. Dynamics of macroalgal
439	blooms along the Cape Cod National Seashore. Northeast. Nat. 16, 53-66.
440	Merceron, M., Morand, P., 2004. Existence of a deep subtidal stock of drifting Ulva in
441	relation to intertidal algal mat developments. J. Sea Res. 52, 269-280.
442	Morand, P., Merceron, M., 2005. Macroalgal population and sustainability. J. Coast.
443	Res. 21, 1009-1020.
444	Nelson, T.A., Haberlin, K., Nelson, A.V., Ribarich, H., Hotchkiss, R., Van Alstyne,
445	K.L., Buckingham, L., Simunds, D.J., Fredrickson, K., 2008. Ecological and
446	physiological controls of species composition in green macroalgal blooms. Ecology 89,
447	1287-1298.
448	Nelson, T.A., Lee, D.J., Smith, B.C., 2003a. Are "green tides" harmful algal blooms?
449	Toxic properties of water-soluble extracts from two bloom-forming macroalgae, Ulva
450	fenestrata and Ulvaria obscura (Ulvophyceae). J. Phycol. 39, 874-879.
451	Nelson, T.A., Nelson, A.V., Tjoelker, M., 2003b. Seasonal and spatial patterns of "green tides"

452	(ulvoid algal blooms) and related water quality parameters in the coastal waters of
453	Washington State, USA. Bot. Mar. 46, 263-275.
454	Oczkowski, A., Nixon, S., Henry, K., DiMilla, P., Pilson, M., Granger, S., Buckley, B.,
455	Thornber, C., McKinney R., Chaves, J., 2008. Distribution and trophic importance of
456	anthropogenic nitrogen in Narragansett Bay: an assessment using stable isotopes. Estuar.
457	Coast. 31, 53-69.
458	Park, S., 1992. Ulva lactuca monitoring programme. Bay of Plenty Regional Council,
459	Tech. Rep. 32.
460	Pedersen, M.F., Borum, J., 1996. Nutrient control of algal growth in estuarine waters.
461	Nutrient limitation and the importance of nitrogen requirements and nitrogen storage
462	among phytoplankton and species of macroalgae. Mar. Ecol. Prog. Ser. 142, 261-272.
463	Raffaelli, D., Raven, J.A., Poole, L.A., 1998. Ecological impact of green macroalgal
464	blooms. Oceanogr. Mar. Biol. 36, 97-125.
465	Rivers, J.S., Peckol, P., 1995. Summer decline of Ulva lactuca (Chlorophyta) in a
466	eutrophic embayment: interactive effects of temperature and nitrogen availability? J.
467	Phycol 31, 223-228.
468	Schramm, W., 1999. Factors influencing seaweed responses to eutrophication: some
469	results from EU-project EUMAC. J. App. Phycol. 11, 69-78.
470	Sfriso, A., Pavoni, B., Marcomini, A., Orio, A.A., 1992. Macroalgae, nutrient cycles,
471	and pollutants in the Lagoon of Venice. Estuar. Coast. 15(4), 517-528.
472	Sousa, A.I., Martins, I., Lillebø, A.I., Flindt, M.R., Pardal, M.A., 2007. Influence of
473	salinity, nutrients and light on the germination and growth of Enteromorpha sp. spores. J.
474	Exp. Mar. Biol. Ecol. 341, 142-150.

475	Tan, I.H., Blomster, J., Hansen, G., Leskinen, E., Maggs, C.A., Mann, D.G., Sluiman,
476	H.J., Stanhope, M.J., 1999. Molecular phylogenetic evidence for a reversible
477	morphogenetic switch controlling the gross morphology of two common genera of green
478	seaweeds, Ulva and Enteromorpha. Mol. Biol. Evol. 16, 1011-1018.
479	Taylor, R., Fletcher, R.L., Raven, J.A., 2001. Preliminary studies on the growth of
480	selected 'green tide' algae in laboratory culture: effects of irradiance, temperature, salinity
481	and nutrients on growth rate. Bot. Mar. 44, 327-336.
482	Thomsen, M., McGlathery, K., 2006. Effects of accumulations of sediments and drift
483	algae on recruitment of sessile organisms associated with oyster reefs. J. Exp. Mar. Biol.
484	Ecol. 328, 22-34.
485	Underwood, A.J., 1997. Experiments in ecology. Cambridge University Press, Cambridge.
486	Valiela, I., McClelland, J., Hauxwell, J., Behr, P.J., Hersh, D., Foreman, K., 1997.
487	Macroalgal blooms in shallow estuaries: controls and ecophysiological and ecosystem
488	consequences. Limnol. Oceanogr. 42, 1105-1118.
489	Van Alstyne, K.L., Nelson, A.V., Vyvyan, J.R., Cancilla, D.A., 2006. Dopamine
490	functions as an antiherbivore defense in the temperate green alga Ulvaria obscura.
491	Oecologia 148, 304-311.
492	Worm, B., Lotze, H.K., 2006. Effects of eutrophication, grazing, and algal blooms on
493	rocky shores. Limnol. Oceanogr. 51, 569-579.
494	Ye, NH., Zhang, XW., Mao, YZ., Liang, CW., Xu, D., Zou, J., Zhuang, ZM.,
495	Wang, QY., 2011. 'Green tides' are overwhelming the coastline of our blue planet:
496	taking the world's largest example. Ecol. Res. 26, 477-485.
497	Yokoyama, H., Ishihi, Y., 2010. Bioindicator and biofilter function of Ulva spp.

- 498 (Chlorophyta) for dissolved inorganic nitrogen discharged from a coastal fish farm —
- 499 potential role in integrated multi-trophic aquaculture. Aquaculture 310, 74-83.

503 Figure 1 Map of Rhode Island, USA, showing the location of our four study sites.

504

Figure 2 *Ulva* biomass during the May-September 2009 and February and May 2010 surveys at
A) Pier 5 and B) GSO. Error bars are ± 1 SE.

507

Figure 3 *Ulva* biomass at the two bloom-impacted sites A) Chepiwanoxet and B) Brushneck
Cove. Error bars are ± 1 SE.

510

511 Figure 4 Algal percent cover at all sites during 2009-2010. Species comprising less than 10%

512 cover in all months are not shown. Site abbreviations follow Table 2. ACGP refers to mixed mats

of *Agardhiella subulata*, *Ceramium virgatum*, *Gracilaria* spp., and *Polysiphonia* spp. \* indicates
no sampling due to ice cover.

515

516 **Figure 5** Algal percent cover at bloom-impacted sites during 2010-2011. Species comprising

517 less than 4% cover in all months are not shown. Site abbreviations follow Table 2. ACGP refers

518 to mixed mats of Agardhiella subulata, Ceramium virgatum, Gracilaria spp., and Polysiphonia

519 spp. \* indicates no sampling due to storm surge.

- **Table 1.** Results of a nested ANOVA on *Ulva* biomass among month-year, site, transect nested
- 522 within site, and species for a) all study sites from 2009-2010, and b) Brushneck Cove and

523 Chepiwanoxet from 2010-2011.

(a)				
Source	df	MS	F	Р
Month-Year	6	6,267.02	19.19	< 0.0001
Site	3	24,238.75	74.21	< 0.0001
Transect[Site]	4	109.38	0.33	0.85
Species	3	8,006.74	24.51	< 0.0001
Month-Year*Site	18	4,348.83	13.31	< 0.0001
Month-Year*Transect[Site]	24	2,202.50	6.74	< 0.0001
Month-Year*Species	18	3,645.81	11.16	< 0.0001
Site*Species	9	5,645.93	17.28	< 0.0001
Transect[Site]*Species	12	650.59	1.99	0.0217
Month-Year*Site*Species	54	3,210.02	9.83	< 0.0001
Month-Year*Transect[Site]*Species	72	1,107.17	3.39	< 0.0001
Error	1,624	326.65		

$(\mathbf{h})$		
(b)		

Source	df	MS	F	р
Month-Year	15	12,946.93	30.22	< 0.0001
Site	1	6.79	0.016	0.90
Transect[Site]	2	6,889.20	16.08	< 0.0001
Species	3	44,216.10	103.20	< 0.0001
Month-Year*Site	15	721.90	1.68	0.048
Month-Year*Transect[Site]	30	5,333.94	12.45	< 0.0001
Month-Year*Species	45	8,984.53	20.97	< 0.0001
Site*Species	3	8,087.41	18.88	< 0.0001
Transect[Site]*Species	6	4,578.26	10.69	0.0013
Month-Year*Site*Species	45	2,230.41	5.21	< 0.0001
Month-Year*Transect[Site]*Species	90	4,134.88	9.65	< 0.0001
Error	1227	428.47		

526 <b>Ta</b>	Table 2. Taxa observed during our	2009-2010 field surveys. Site abbrev	viations are: P Pier 5, G GSO, G	C Chepiwanoxet, and B
---------------	-----------------------------------	--------------------------------------	----------------------------------	-----------------------

527 Brushneck Cove. Genera marked with an asterisk require microscopic examination for species determinations.

			2009			20	010
	May PGCB	June PGCB	July PGCB	August PGCB	September P G C B	February P G C B	May PGCB
Ulva blades	ХХХХ	X X X	ХХХХ	ХХХХ	ХХ	ХХ	XXXX
Ulva tubes Agardhiella subulata	XXXX	XXX	X X X X	X X X X	X X	X X X	X X X X
Ahnfeltia plicata						Х	
Ascophyllum nodosum	Х	Х	Х	Х	Х	Х	Х
Bonnemaisonia hamifera		Х	ХХ				Х
Bryopsis plumosa			Х	Х			
Ceramium virgatum	ХХХХ	X X	ХХ	X X	ХХ	Х	ХХХХ
<i>Chaetomorpha</i> spp.*			ХХ	Х			
Champia parvula			ХХ				
Chondrus crispus	X X	Х	ХХ	ХХ	Х	ХХ	X X
<i>Codium fragile</i> ssp. <i>fragile</i>	Х		X X	X X	Х	X X	Х
Corallina officinalis						Х	

Cystoclonium	X X	Х					Х
purpureum Desmarestia	Х	Х	Х				Х
viridis Dumontia contorta							ХХ
Ectocarpus	Х			Х			
spp. Fucus distichus		Х	Х	Х	Х	Х	Х
Fucus vesiculosus		Х					
Gracilaria tikvahiae & G. vermiculophylla	ХХХ	X X	X X	Х	X X	Х	X X
Grateloupia turuturu	X X		Х	ХХ		Х	
Heterosiphonia japonica							Х
Hypnea musciformis	Х						
Leathesia marina	Х						
Mastocarpus stellatus	Х	Х		Х			Х
Palmaria palmata	Х	Х	Х	Х	Х	Х	Х
Petalonia & Punctaria spp.*	ХХХ	Х				ХХ	Х
Phyllophora membranifolia				Х		Х	
Polyides rotundus	Х	Х				Х	Х

Polysiphonia & Neosiphonia	Х	Х	Х	X X	Х	X X	ХХ	X X
spp.* <i>Porphyra</i> spp.*	x		Х					ХХ
	X X		X	Х	Х	Х		X
Scytosiphon lomentaria	Х							
Ulothrix flacca							Х	
Vertebrata lanosa	Х		Х					Х

Table 3. Average species richness (S), Pielou's evenness (J'), and Shannon diversity index (H')

Survey	Site	S	J'	H'	
2009-2010	Brushneck Cove	1.56	0.34	0.12	
	Chepiwanoxet	2.92	0.76	0.76	
	GSÔ	1.45	0.72	0.33	
	Pier 5	4.45	0.68	0.92	
2010-2011	Brushneck Cove	1.81	0.67	0.33	
	Chepiwanoxet	2.38	0.60	0.52	

for our 2009-2010 and 2010-2011 field surveys.

**Table 4.** Results from a two-way crossed ANOSIM for differences amongst sites and months in536our 2009-2010 survey. R-values close to 1.00 indicate complete separation between groups while537R-values close to 0 indicate little separation between groups. All pairwise tests were significant538(p = 0.001). (a) Tests for differences between site groups across all month groups. (b) Tests for539differences between month groups across all site groups.

541	54	1	
-----	----	---	--

(a)							
Global R: 0.760	Brush	Brushneck Cove		Chepiwanoxet		Pier 5	
Brushneck Cove							
Chepiwanoxet	0.736						
GSO	0.787		0.70	)3			
Pier 5	0.908		0.83	30	0.766		
(b)							
Global R: 0.470	May	June	July	August	September	February	May
	2009					2010	
May 2009							
June	0.179						
July	0.481	0.518					
August	0.413	0.515	0.162				
September	0.468	0.444	0.689	0.681			
February	0.460	0.690	0.683	0.755	0.702		
May 2010	0.215	0.379	0.560	0.565	0.558	0.256	
*							

543 **Table 5.** Results from a two-way crossed SIMPER analysis for average similarity and

dissimilarity amongst sites across all months in our 2009-2010 survey. ACGP refers to mats of

545 intertwined Agardhiella subulata, Ceramium virgatum, Gracilaria spp., and Polysiphonia spp.

- 546 Site abbreviations follow Table 2.
- 547

	Percent contribution	Cumulative percent
Within site similarity		
Brushneck Cove– Average similarity: 77.69		
<i>Ulva</i> blades	95.74	95.74
Chepiwanoxet – Average similarity: 65.76		
<i>Ulva</i> tubes	34.73	34.73
<i>Ulva</i> blades	33.30	68.03
ACGP	20.71	88.74
<i>Gracilaria</i> spp.	5.47	94.21
GSO – Average similarity: 45.89		
<i>Ulva</i> tubes	58.22	58.22
Polysiphonia spp.	13.72	71.95
Ceramium virgatum	11.53	83.48
Chondrus crispus	7.68	91.16
Pier 5 – Average similarity: 54.19		
Chondrus crispus	43.58	43.58
Saccharina latissima	17.87	61.45
<i>Ulva</i> blades	9.77	71.22
Palmaria palmata	9.33	80.55
Ulva tubes	6.21	86.76
Desmarestia viridis	3.45	90.21

Between site dissimilarity	Perce	Percentage contribution to average dissimilarity between sites						
Taxon	B-C	B-G	B-P	C-G	C-P	G-P		
Ulva blades	21.87	60.42	24.08	29.66	12.58	8.43		
Ulva tubes	32.11	15.82	9.57	22.95	14.15	8.62		
ACGP	24.52	-	-	14.94	10.24	-		
Ascophyllum nodosum	-	-	-	-	-	2.48		
Ceramium virgatum	6.10	5.39	-	8.15	2.80	3.63		
Chondrus crispus	-	-	20.65	4.33	17.73	30.73		
Codium fragile ssp. fragile	-	-	-	-	1.86	-		
Desmarestia viridis	-	-	2.86	-	3.17	4.20		
<i>Fucus</i> spp.	-	-	5.21	-	4.30	5.91		
Gracilaria spp.	9.72	8.01	4.31	7.86	4.89	-		
Grateloupia turuturu	-	-	-	-	-	1.50		
Palmaria palmata	-	-	8.31	-	5.58	7.63		

Polysiphonia spp.	-	5.14	4.53	4.62	3.65	2.49
Porphyra spp.	-	-	-	-	-	2.20
Saccharina latissima	-	-	10.81	-	7.84	15.75
Ulothrix flacca	-	-	-	-	2.68	-
Total contribution of species	94.32	94.77	90.33	92.51	91.47	91.35
Average between site dissimilarity	57.16	92.68	85.55	82.62	83.83	87.21











