Clinics in Oncology

9

The Role of Meningioma-1 (Mn1) Gene as Marker for Prognosis and Minimal Residual Disease Monitoring in Acute Myeloid Leukemia: A Concise Review

Eleonora Toffoletti, Mario Tiribelli*, Giorgia Maccari, Alexsia Chiarvesio, Anna Candoni, Renato Fanin and Daniela Damiani

Division of Hematology and Bone Marrow Transplantation, Azienda Sanitaria Universitaria Integrata di Udine, Italy

Introduction

Molecular markers are necessary for prognostic stratification and monitoring of Minimal Residual Disease (MRD) in Acute Myeloid Leukemia (AML) [1,2]. Cytogenetic aberrations have long been recognized as the most important prognostic variable in AML, and are still the major determinant for post-remission therapy [3]. Unfortunately, only 50-60% of AML patients present an abnormal karyotype at diagnosis, while the remaining cases display a Normal Karyotype (NK). NK AML patients are generally included in an "intermediate risk" prognostic group, that is however characterized by a heterogeneous clinical course. To stratify prognosis of NK AML patients, numerous studies have led, in the last decade, to the introduction of different molecular markers such as FLT3, NPM1, BAALC and CEBPA [4-7]. Still, their use to monitor disease, either defining remission status and detecting relapse as early as possible, is still somehow controversial, due to fluctuations during disease course, low incidence rates in AML and sensitivity of the technologies detecting the single marker [8-10]. These limitations have, to date, precluded a timely and precise quantification of disease in NK AML patients, thus preventing from a complete individualization of post-remission therapy and early treatment in case of impending relapse. In other words, in NK AML it has not been reached the precision achieved in BCR/ABL-positive chronic myeloid leukemia and PML/RAR alpha mutated acute promyelocytic leukemia.

OPEN ACCESS

*Correspondence:

Mario Tiribelli, Division of Hematology and Bone Marrow Transplantation, Azienda Sanitaria Universitaria Integrata di Udine, P.le S. M. Misericordia, 15 - 33100 Udine, Italy, Tel: +39-0432-559666; Fax: +39-0432-559661; E-mail: mario.tiribelli@uniud.it Received Date: 12 Jan 2017 Accepted Date: 07 Mar 2017 Published Date: 13 Mar 2017 Citation:

Toffoletti E, Tiribelli M, Maccari G, Chiarvesio A, Candoni A, Fanin R, et al. The Role of Meningioma-1 (Mn1) Gene as Marker for Prognosis and Minimal Residual Disease Monitoring in Acute Myeloid Leukemia: A Concise Review. Clin Oncol. 2017; 2: 1217.

Copyright © 2017 Mario Tiribelli. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. MRD is conventionally defined as the amount of residual AML cells still present in complete remission patients, undetectable at microscope level. The most widely used method for MRD detection is the quantitative Real-Time Polymerase Chain Reaction Assay (RT-PCR). Historically, MRD detection by RT-PCR was limited to those patients characterized by a genetic signature (e.g.: mutations, fusion genes or altered gene expressions) [11]. To overcome this lack, efforts have been made to identify so-called "pan-leukemic" markers validated for MRD detection in NK AML. The Wilms Tumor gene 1 (WT1) has been found to be overexpressed in a large proportion of AML (over 90% of cases, irrespective of karyotype) [12-14]. Although not associated with a specific leukemic clone, WT1 quantification provides information of disease persistence or relapse. However, being WT1 expressed also in normal cells, its determination has limited sensitivity; more, around 10% of AML patients does not express WT1. Therefore, is necessary to associate other possible markers in the characterization of AML.

The *Meningioma 1* (MN1) gene, located on chromosome 22q11, encodes a protein that participates in a gene transcription regulator complex. In cells derived from the bone marrow of healthy donors the MN1 levels are very low [15,16], supporting the hypothesis that high MN1 gene levels are specific for leukemic blasts and not a consequence of differentiation. However, the molecular mechanisms via which MN1 inhibits differentiation and stimulates proliferation of hematopoietic cells remain largely unknown.

MN1 has been found to be over-expressed in AML with inv(16) [17], and high MN1 levels seems to have prognostic impact in NK AML patients (Table 1). In 2006 Heuser et al. [18] found that high MN1 (defined as more than the median expression) was associated with poor response to a double induction therapy of idarubicin + etoposide + cytarabine (71% *vs.* 87%, P = 0.02), and lower Relapse-Free Survival (RFS) (23% *vs.* 51% at 3 years, P = 0.002) and Overall Survivals (OS) (38% *vs.* 59% at 3 years, P = 0.03) in 142 adult patients with NK AML. In multivariate analysis, MN1 expression retained its prognostic significance. Few years later, Langer et al. [19] measured MN1 expression by real-time RT-PCR in 119 untreated NK AML patients younger than 60 years to confirm its

Reference	No. of patients	No. of controls	Cytogenetic	Molecular markers	Incidence of MN1 over expression (%)	Threshold of MN1 over expression	Prognostic impact			Multivariate analysis	
							CRR	RFS/DFS	os	Variables	Significant parameters
Heuser et al. [18]	142	0	NK	MN1, FLT3, MLL-PTD, NPM1	50	median value	0,02	RFS: 0.002 at 3y	0.03 at 3y	MN1 expression, ECOG performance status, Age above median	OS
Langer et al. [19]	119	0	NK	MN1, NPM1, BAALC, FLT3, WT1¤, MLL-PTD, ERG, CEBPA	25	highest quartile	0,006	DFS: <0.001 at 5y	0.001 at 5y	MN1 and ERG expression, FLT3-ITD, NPM1 and WT1¤, WBCs	CRR, DFS, OS
Aref et al. [20]	100	10	NK	MN1, NPM1, FLT3	48	median value	0,001	DFS: 0.04 at 1y	0.03 at 1y	MN1 expression, Age,WBCs, Blast count,FAB	DFS, OS
Xiang et al. [21]	158	20	not specified, but all FAB except M3	FLT3, NPM1, WT1 expression, miR-20a, miR-181b,	50	median value	fSWOG*: P=0.026 iSWOG§: P=0.04 aSWOG¥: P=0.024	n.s.	fSWOG : P=0.024 at 2yiSWOG: P=0.018 at 2y aSWOG: P=0.033 at 2y	MN1, miR- 20a, miR- 181b, WT1 expression, FLT3-ITD, NPM1, WBCs	CCR, RFS, OS
Zayed et al. [22]	50	10	NK	MN1, PTEN	50	median value	n.s.	n.s.	n.s.	n.a.	n.a.

Table 1: Expression and prognostic impact of MN1 in NK AML.

CCR: Complete Remission Rate; RFS: Relapse Free Survival; DFS: Disease Free Survival; OS: Overall Survival; NK: Normal Kariotype; ¤: Mutational Status of WT1; y: Years; WBCs: Count of White Blood Cells at Onset, *: Favorable SWOG Group; §: Intermediate SWOG Group; ¥: Adverse SWOG Group

prognostic role in the context of other predictive molecular markers. Their results indicate that higher MN1 expression was associated with NPM1 wild-type and high BAALC expression (P = 0.004); patients over-expressing MN1 had lower Complete Remission (CR) rate (P = 0.005 after adjustment for WBC count), shorter Disease-Free Survival (DFS) (P = 0.01 after adjustment for WT1and FLT3-ITD mutations); and shorter OS (P = 0.04 after adjustment for WT1, NPM1 and FLT3-ITD mutations, and for WBC).

In line with these first experiences, more recently Aref et al. [20] analyzed 100 NK AML patients, treated and followed up for at least 24 months, showing that MN1 overexpression (documented in 48 patients) is a predictor of poor response, as high gene levels at diagnosis were associated with poor CR after induction chemotherapy (8.4% vs. 62.5%, P = 0.001), higher risk of relapse (54.1 vs. 23%, P = 0.02) and shorter survival (mortality rate 75% *vs.* 46.1, P = 0.03). Multivariable analysis confirmed that MN1 over-expression was an independent risk factor for RFS and OS. Xiang et al. [21] studied MN1 gene and MN1-associated microRNAs expression level in 158 newly diagnosed Chinese AML patients and in 20 healthy donors, finding that MN1 was overexpressed in patients compared with normal controls and that high gene expression was associated with lower CR rates (P = 0.01) and shorter RFS (P = 0.02) and OS (P = 0.02).

If, overall, most of the published experiences identify MN1 overexpression as a marker of a more aggressive disease, a recent paper by Zayed et al. [22] on 50 NK AML patients and 10 controls did not confirm MN1 prognostic role nor suggested its use as a routine diagnostic tool. However, this latter study may suffer from the limited sample size and the consequently low statistical power.

Concerning the potential usefulness of MN1 gene as MRD marker, a recent paper by Carturan et al. [15] showed not only that an increased MN1 expression can be of prognostic significance in predicting relapse of AML patient, but also that MN1 and WT1

gene expression had a good degree of concordance, as an increase above the upper normal limit was documentable four months before hematologic relapse. In our experience, we found an excellent concordance between MN1 and WT1 gene expression and with disease course (i.e. remission achievement and maintenance, relapse) [personal data, unpublished]. In our cohort, WT1 was over-expressed in about 90% of AML patients, while MN1 was high only in 40% of the cases, in line with most published experience [19]. Nonetheless, the concomitant evaluation of the two markers could give clinicians a better tool to early detect a leukemic relapse in AML lacking a specific molecular marker. In their work, Carturan and colleagues analyzed the different cytogenetic prognostic groups according to MN1 expression, finding a lower gene expression in cases with translocation t(8;21) compared to those with inv(16), that have been recognized as unique entities and are usually reported together as Core Binding Factor (CBF) AML [15]. This diversity is in line with an emerging branch of research investigating the clinical and biological heterogeneity of CBF AML [23]. Moreover, they found that MN1 levels in patients with Acute Promyelocytic Leukemia (APL) are very low, comparable to those of healthy controls. This could suggest that MN1 is involved in pathways not activated in APL, contrarily to WT1, whose levels are higher in APL than in other AML [24].

In summary, MN1 gene has peculiar patterns of expression in different AML subtypes, even among the same cytogenetic risk categories, and seems promising to gain further knowledge of leukemogenic mechanisms, particularly in CBF or in NK AML lacking other known mutations (such as FLT3 or NPM1). Even if the published experiences have given discordant results, most of the evidences point towards a prognostic role of MN1 in AML patients; however, further studies on large number of patients are warranted to define the correct weight of MN1 as a factor impacting on outcomes. Last, promising data are emerging on MN1 potential as a marker of MRD, both alone and in combination with other molecular abnormalities, if present, to allow for a precocious detection of AML relapse and to optimize post-remission therapy. In the last years, various paper focused on therapeutic strategies upon re-emergence of minimal residual disease in AML, mostly after allogeneic stem cell transplant [25,26], but an ultimate consensus on monitoring strategies and on a MDR-driven therapy is still lacking [27].

References

- 1. Döhner H, Estey E, Grimwade D, Amadori S, Appelbaum FR, Büchner T, et al. Diagnosis and management of AML in adults: 2017 ELN recommendations from an international expert panel. Blood. 2017;129: 424-447.
- Schlenk RF, Döhner K, Krauter J, Fröhling S, Corbacioglu A, Bullinger L, et al. Mutations and treatment outcome in cytogenetically normal acute myeloid leukemia. German-Austrian Acute Myeloid Leukemia Study Group. N Engl J Med. 2008; 358: 1909-1918.
- 3. Estey E. Acute myeloid leukemia: 2016 Update on risk-stratification and management. Am J Hematol. 2016; 91: 824-846.
- 4. Thiede C, Steudel C, Mohr B, Schaich M, Schäkel U, Platzbecker U, et al. Analysis of FLT3-activating mutations in 979 patients with acute myelogenous leukemia: association with FAB subtypes and identification of subgroups with poor prognosis. Blood. 2002; 99: 4326-4335.
- Schnittger S, Kern W, Tschulik C, Weiss T, Dicker F, Falini B, et al. Minimal residual disease levels assessed by NPM1 mutation-specific RQ-PCR provide important prognostic information in AML. Blood. 2009; 114: 2220-2231.
- 6. Schwind S, Marcucci G, Maharry K, Radmacher MD, Mrózek K, Holland KB, et al. BAALC and ERG expression levels are associated with outcome and distinct gene and micro RNA expression profiles in older patients with de novo cytogenetically normal acute myeloid leukemia: a Cancer and Leukemia Group B study. Blood. 2010; 116: 5660-5669.
- Li HY, Deng DH, Huang Y, Ye FH, Huang LL, Xiao Q, et al. Favorable prognosis of biallelic CEBPA gene mutations in acute myeloid leukemia patients: a meta-analysis. Eur J Haematol. 2015; 94: 439-448.
- Krönke J, Schlenk RF, Jensen KO, Tschürtz F, Corbacioglu A, Gaidzik VI, et al. Monitoring of minimal residual disease in NPM1-mutated acute myeloid leukemia: a study from the German-Austrian acute myeloid leukemia study group. J Clin Oncol. 2011; 29: 2709-2716.
- Damiani D, Tiribelli M, Raspadori D, Sirianni S, Meneghel A, Cavalllin M, et al. Clinical impact of CD200 expression in patients with acute myeloid leukemia and correlation with other molecular prognostic factors. Oncotarget. 2015; 6: 30212-30221.
- 10. Candoni A, De Marchi F, Zanini F, Zannier ME, Simeone E, Toffoletti E, et al. Predictive value of pretransplantation molecular minimal residual disease assessment by WT1 gene expression in FLT3-positive acute myeloid leukemia. Exp Hematol. 2017. [Epub ahead of print]
- 11. Del Principe MI, Buccisano F, Maurillo L, Sconocchia G, Cefalo M, Consalvo MI, et al. Minimal Residual Disease in Acute Myeloid Leukemia of Adults: Determination, Prognostic Impact and Clinical Applications. Mediterr J Hematol Infect Dis. 2016; 8: e2016052.
- 12. Inoue K, Ogawa H, Sonoda Y, Kimura T, Sakabe H, Oka Y, et al. Aberrant overexpression of the Wilmstumor gene (WT1) in human leukemia. Blood. 1997; 89: 1405–1412.
- Inoue K, Sugiyama H, Ogawa H, Nakagava M, Yamagami T, Miwa H, et al. WT1 as a new prognostic factor and a new marker for the detection of minimal residual disease in acute leukemia. Blood. 1994; 84: 3071–3079.

- 14. Cilloni D, Renneville A, Hermitte F, Hills RK, Daly S, Jovanovic JV, et al. Real-time quantitative polymerase chain reaction detection of minimal residual disease by standardized WT1 assay to enhance risk stratification in acute myeloid leukemia: a European Leukemia Net study. J ClinOncol. 2009; 27: 5195-5201.
- 15. Carturan S, Petiti J, Rosso V, Calabrese C, Signorino E, Bot-Sartor G, et al. Variable but consistent pattern of Meningioma 1 gene (MN1) expression in different genetic subsets of acute myelogenous leukaemia and its potential use as a marker for minimal residual disease detection. Oncotarget. 2016.
- 16. Schroeder T, Czibere A, Zohren F, Aivado M, Gattermann N, Germing U, et al. Meningioma 1 gene is differentially expressed in CD34 positive cells from bone marrow of patients with myelodysplastic syndromes with the highest expression in refractoryanemia with excess of blasts and secondary acute myeloid leukemia. Leukemia&Lymphoma. 2009; 50: 1043-1046.
- Carella C, Bonten J, Sirma S, Kranenburg TA, Terranova S, Klein-Geltink R, et al. MN1 overexpression is an important step in the development of inv(16) AML. Leukemia. 2007; 21: 1679-1690.
- Heuser M, Beutel G, Krauter J, Döhner K, von Neuhoff N, Schlegelberger B, et al. High meningioma 1 (MN1) expression as a predictor for poor outcome in acute myeloid leukemia with normal cytogenetics. Blood. 2006; 108: 3898-3905.
- 19. Langer C, Marcucci G, Holland KB, Radmacher MD, Maharry K, Paschka P, et al. Prognostic importance of MN1 transcript levels, and biologic insights from MN1-associated gene and micro RNA expression signatures in cytogenetically normal acute myeloid leukemia: a cancer and leukemia group B study. J Clin Oncol. 2009; 27: 3198-3204.
- Aref S, Ibrahim L, Morkes H, Azmy E, Ebrahim M. Meningioma 1 (MN1) expression: Refined risk stratification in acute myeloid leukemia with normal cytogenetics (CN-AML). Hematology. 2013; 18: 277-283.
- 21. Xiang L, Li M, Liu Y, Cen J, Chen Z, Zhen X, et al. The clinical characteristics and prognostic significance of MN1 gene and MN1- associated microRNA expression in adult patients with de novo acute myeloid leukaemia. Ann Hematol. 2013; 92: 1063-1069.
- 22. Zayed RA, Eltaweel MA, BotrosSK, Zaki MA. MN1 and PTEN gene expression in acute myeloid leukemia. Cancer Biomark. 2016.
- Duployez N, Marceau-Renaut A, Boissel N, Petit A, Bucci M, Geffroy S, et al. Comprehensive mutational profiling of core binding factor acute myeloid leukemia. Blood. 2016; 127: 2451-2459.
- 24. Gaur GC, Ramadan SM, Cicconi L, Noguera NI, Luna I, Such E, et al. Analysis of mutational status, SNP rs16754, and expression levels of Wilms tumor 1 (WT1) gene in acute promyelocytic leukemia. AnnHematol. 2012; 91: 1855-1860.
- 25. Platzbecker U, Wermke M, Radke J, Oelschlaegel U, Seltmann F, Kiani A, et al. Azacitidine for treatment of imminent relapse in MDS or AML patients after allogeneic HSCT: results of the RELAZA trial. Leukemia. 2012; 26: 381-389.
- 26. Ommen HB, Hokland P, Haferlach T, Abildgaard L, Alpermann T, Haferlach C, et al. Relapse kinetics in acute myeloid leukaemias with MLL translocations or partial tandem duplications within the MLL gene. Br J Haematol. 2014;165: 618-628.
- 27. Ossenkoppele G, Schuurhuis GJ. MRD in AML: does it already guide therapy decision-making? Hematology Am Soc Hematol Educ Program. 2016; 2016: 356-365.