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Therapeutic Drug Monitoring and Dosage Adjustments of Immunosuppressive Drugs When Combined With Nirmatrelvir/Ritonavir in Patients With COVID-19

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Abstract: Nirmatrelvir/ritonavir (Paxlovid) consists of a peptidomimetic inhibitor (nirmatrelvir) of the SARS-CoV-2 main protease and a pharmacokinetic enhancer (ritonavir). It is approved for the treatment of mild-to-moderate COVID-19. This combination of nirmatrelvir and ritonavir can mediate significant and complex drug–drug interactions (DDIs), primarily due to the ritonavir component. Indeed, ritonavir inhibits the metabolism of nirmatrelvir through cytochrome P450 3A (CYP3A) leading to higher plasma concentrations and a longer half-life of nirmatrelvir. Coadministration of nirmatrelvir/ritonavir with immunosuppressive drugs (ISDs) is particularly challenging given the major involvement of CYP3A in the metabolism of most of these drugs and their narrow therapeutic ranges. Exposure of ISDs will be drastically increased through the potent ritonavir-mediated inhibition of CYP3A, resulting in an increased risk of adverse drug reactions. Although a decrease in the dosage of ISDs can prevent toxicity, an inappropriate dosage regimen may also result in insufficient exposure and a risk of rejection.

Here, we provide some general recommendations for therapeutic drug monitoring of ISDs and dosing recommendations when coadministered with nirmatrelvir/ritonavir. Particularly, tacrolimus should be discontinued, or patients should be given a microdose on day 1, whereas cyclosporine dosage should be reduced to 20% of the initial dosage during the antiviral treatment. Dosages of mammalian target of rapamycin inhibitors (m-TORis) should also be adjusted while dosages of mycophenolic acid and corticosteroids are expected to be less impacted.

Key Words: drug–drug interactions, transplantation, calcineurin inhibitors, mTOR inhibitors, tacrolimus

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INTRODUCTION

Paxlovid is a new solid oral formulation indicated for the treatment of mild-to-moderate COVID-19 in adults and pediatric patients (12 years of age and older weighing at least

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40 kg) who test positive for direct SARS-CoV-2 virus and who are at high risk for progression to severe COVID-19.¹⁻⁴ Paxlovid is supplied as 300-mg nirmatrelvir (2 150 mg tablets) with 100-mg ritonavir (one 100 mg tablet) with all the 3 tablets taken together orally with or without food twice daily for 5 days.¹ No dosage adjustment is needed in patients with mild renal impairment (eGFR ≥ 60 to < 90 L/min). In patients with moderate renal impairment (eGFR ≥ 30 to < 60 mL/min), the dosage of Paxlovid is modified to 150-mg nirmatrelvir and 100-mg ritonavir twice daily for 5 days. However, in patients with severe renal impairment or severe hepatic impairment, the use of the nirmatrelvir/ritonavir is not recommended due to limited availability of safety and efficacy data.^{1,2}

Pharmacodynamics

The chemical name of nirmatrelvir is (1*R*, 2*S*, 5*S*)-*N*-((1*S*)-1-Cyano-2-((3*S*)-2-oxopyrrolidin-3-yl) ethyl)-3-((2*S*)-3,3-dimethyl-2-(2,2,2-trifluoroacetamido) butanoyl)-6,6-dimethyl-3-azabicyclo [3.1.0] hexane-2-carboxamide (Fig. 1), a peptidomimetic inhibitor of the SARS-CoV-2 main protease, also referred to as 3C-like protease or nsp5 protease. Inhibition of SARS-CoV-2 main protease renders it incapable of processing polyprotein precursors, preventing viral replication.^{1,2} Nirmatrelvir exhibited antiviral activity against SARS-CoV-2 infection in a primary human lung alveolar epithelial cell line (EC50 value of 61.8 nM and EC90 value of 181 nM) after 3 days of drug exposure.⁵ It was shown to have efficacy against the Alpha (B.1.1.7), Gamma (P.1), Delta (B.1.617.2), Lambda (C.37), Mu (B.1.621), and Omicron (B.1.1.529) SARS-CoV-2 variants.⁵

Pharmacokinetics

After oral administration of a single dose of nirmatrelvir/ritonavir 300 mg/100 mg, the median time to peak concentration

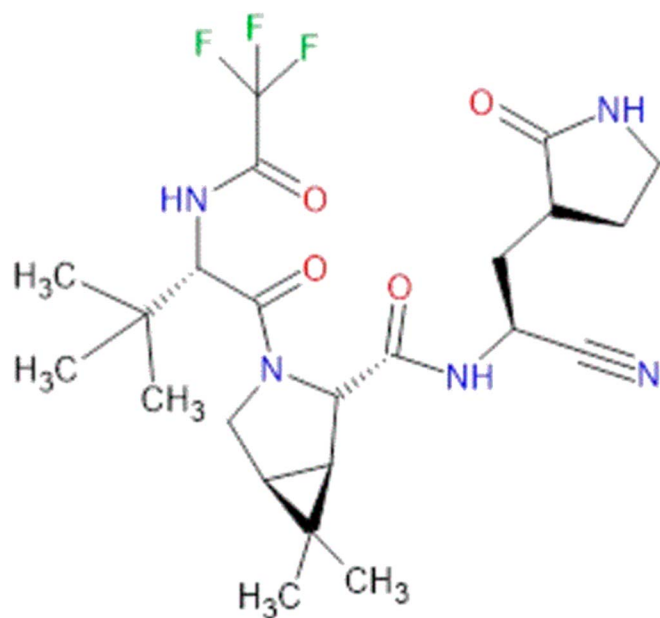


FIGURE 1. Chemical structure of nirmatrelvir.

of nirmatrelvir was 3 hours (range, 1–6 hours), indicating relatively rapid absorption. The observed geometric mean (CV%) concentration maximum (C_{max}) and the area under the plasma concentration versus time curve from zero to infinity (AUC_{0–inf}) for nirmatrelvir were 2.21 mg/L (33% CV) and 23.01 mcg**h*/mL (23% CV), respectively.^{1,5} Studies have shown that nirmatrelvir was moderately bound to plasma proteins (69%) and has a mean apparent volume of distribution (V_z/F) of 104.7 L when administered with ritonavir (300 mg nirmatrelvir/100 mg ritonavir).¹

Based on in vitro studies, CYP3A4 is the major contributor (99%) to the oxidative metabolism of nirmatrelvir. However, nirmatrelvir undergoes minimal metabolism when coadministered with ritonavir, resulting in high and persisting plasma concentrations.^{1,5} When the protease inhibitor metabolism is inhibited by ritonavir, renal elimination becomes the major route of nirmatrelvir.^{1,5} After single or multiple oral doses of nirmatrelvir/ritonavir as oral suspension in healthy volunteers, the half-life ranged from 6.8 to 9.5 hours, thereby supporting a twice daily dosing regimen.^{1,5}

Clinical Trials

The first-in-human clinical trial of nirmatrelvir started on September 9, 2020, and was a phase-1B, placebo-controlled, single and multiple intravenous ascending dose study evaluating the safety, tolerability, and pharmacokinetics in patients with COVID-19.^{6,7} The efficacy of nirmatrelvir was evaluated in a preomicron variant era phase-2 to phase-3, double-blind, randomized, controlled trial in adult patients with a laboratory-confirmed diagnosis who were symptomatic, unvaccinated, nonhospitalized, and at high risk for progression to severe COVID-19. A total of 2246 adults were randomized to receive either 300 mg of nirmatrelvir along with 100 mg of ritonavir or placebo orally every 12 hours for 5 days.³ The treatment arm demonstrated a COVID-19–related hospitalization rate of 0.77% (3/389) at 28 days compared with the placebo arm at 7.01% (27/385). This led to an absolute risk reduction of 6.32% [95% confidence interval (CI), –9.04 to –3.59; *P* < 0.001] and a relative risk reduction of 87.8%. In addition, the COVID-19–related mortality through day 28 was 0% (0/389) in the treatment group compared with 1.2% (7/385) in the placebo group.³ Patients treated with nirmatrelvir and ritonavir displayed a decrease in SARS-CoV-2 viral load by a factor of 10 relative to the placebo at day 5.³ Due to the satisfactory demonstration of efficacy and safety, Paxlovid received the Emergency Use Authorisation from the US-FDA in December 2021 and conditional marketing authorization by the EMA in January 2022 as the first oral antiviral drug for treating COVID-19 in the outpatient setting.⁸ Of note, less than 1% of patients included in the trial were immunocompromised.

Warnings and Toxicity

The use of Paxlovid has been associated with potential complications. Dysgeusia, diarrhea, hypertension, and myalgia are the main adverse drug reactions reported in the phase-3 randomized clinical trial.^{2,4,5}

The combination of nirmatrelvir/ritonavir has significant and complex DDIs, primarily due to ritonavir. A careful review

of the patient's concomitant medications, including over-the-counter medications, herbal supplements, and recreational drugs should be performed to minimize the occurrence of any potential DDIs before prescribing nirmatrelvir/ritonavir.^{2,4,5} It is suggested that drug interaction potential be comprehensively evaluated. As an example, French recommendations on managing DDIs with nirmatrelvir/ritonavir have been published,⁹ and the Web site of the University of Liverpool also provides free access to drug interaction charts.¹⁰ The nature of the drug–drug interactions will be discussed in subsequent sections of this publication.

Paxlovid is contraindicated with drugs that are potent CYP3A inducers, where significantly reduced nirmatrelvir/ritonavir plasma concentrations may lead to potential loss of response to SARS-CoV-2 and possible development of resistance.

However, ritonavir is a strong inhibitor of cytochrome CYP3A4; as such, the use of Paxlovid is contraindicated with drugs that are highly dependent on CYP3A for clearance and for which elevated concentrations are associated with serious or life-threatening reactions.^{2,4,5} Moreover, nirmatrelvir/ritonavir could also exhibit potential life-threatening interactions in individuals on immunosuppressive drugs (ISDs) because these are mainly metabolized by CYP3A4.^{2,4,5}

Concomitant drug therapy of nirmatrelvir/ritonavir with ISDs requires specific consideration to avoid elevated concentrations of ISDs in the toxic range; in this scenario, therapeutic drug monitoring (TDM) can be beneficial to adjust dosing of ISDs. This will be the focus of the second part of this publication.

DDIS WITH RITONAVIR-BOOSTED NIRMATRELVIR

In vitro testing shows that nirmatrelvir does not significantly inhibit CYP1A2, CYP2B6, CYP2C8, CYP2C9, CYP2C19, or CYP2D6, and it does not induce any CYPs at clinically relevant concentrations. However, nirmatrelvir has the potential to irreversibly and time-dependently inhibit CYP3A4 and P-glycoprotein 1 (permeability glycoprotein, P-gp), also known as multidrug resistance protein 1 or ATP-binding cassette subfamily B member 1 or cluster of differentiation 243.¹ Although DDIs between nirmatrelvir and ritonavir is beneficial in the use of Paxlovid, the potent inhibition of 3A4 by ritonavir can lead to other significant drug interactions. The significant DDIs in the combination product are primarily due to ritonavir.

Ritonavir, a substrate of CYP3A and to a lesser extent, CYP2D6, is transported by P-gp.^{11,12} It is an inducer of CYP1A2, CYP2C9, CYP2C19, CYP2B6, and UGT1A1. Ritonavir is a potent inhibitor of CYP3A and, to a lesser extent of CYP2D6, CYP2C8. Ritonavir displays a paradoxical dose- and time-dependent inhibitory/induction effect on CYP3A and on the multidrug efflux transporter P-gp.¹¹ Some of the specific interactions are discussed below.

Of note, inflammation resulting from COVID-19 can also impact metabolism leading to drug–disease interaction. However, this phenomenon might be stronger in the later stage of infection (ie, the cytokine storm) than at the early stage, where nirmatrelvir/ritonavir is indicated.

CYP3A

Ritonavir is metabolized by CYP3A4 with a Km value of around 20 μM . However, the KI value for CYP3A4-mediated drug metabolism is 0.015 μM .¹³ Ritonavir's potent inhibitory potential is based on the efficient blocking of CYP3A4 and CYP3A5 (kinact/KI), and it was reported to be as large as 3200 and 667 $\text{min}^{-1}\cdot\text{mM}^{-1}$, respectively.¹⁴ Studies have shown that ritonavir is a type II ligand that perfectly fits into the CYP3A4 active site and irreversibly binds to the heme iron by means of the thiazole nitrogen. This binding results in decreased redox potential of the CYP protein.¹⁵ This is the basis of the interaction between ritonavir and immunosuppressant drugs, such as tacrolimus, cyclosporine, everolimus, and sirolimus, that are metabolized by CYP3A. Enzymatic activity can only be restored through de novo protein synthesis; therefore, these mechanism-based inhibitions are prolonged in vivo and are observed even after stopping the inhibitors.

Ritonavir is well acknowledged as a strong CYP3A inhibitor with an inhibitory effect starting from the first intake and lasting at least 3 days after the end of ritonavir administration. Therefore, ritonavir has been combined with other drugs as a pharmacokinetic booster such as the prescription for antihuman immune deficiency virus drugs and direct-acting antiviral drug against hepatitis C virus (HCV). Studies have demonstrated drug interactions with the direct-acting antiviral combination of ombitasvir, paritaprevir, and ritonavir (Viekirax) and 11 medications in healthy volunteers.¹⁶ Although only formulated with ritonavir 100 mg once daily, the exposure of ketoconazole, digoxin, pravastatin, and rosuvastatin with Viekirax were increased by up to 105%, 58%, 76%, and 161%, respectively, whereas omeprazole exposure decreased by approximately 50%. However, during this phase-2 study on DDIs, no clinically meaningful changes in ombitasvir, paritaprevir, and ritonavir exposures were observed in the presence of the 11 medications.

P-Glycoprotein

Ritonavir is unidirectionally transported by P-gp.^{17,18} Several clinical studies have shown that ritonavir pretreatment in healthy subjects slightly decreased renal and nonrenal clearance of digoxin administered orally or intravenously.^{19,20} The simultaneous administration of ritonavir increased time-to-peak concentration and AUC_{0-24} and lowered digoxin renal clearance, implying that ritonavir mainly affects digoxin's tubular secretion through P-gp.²¹ Taken together, the administration of ritonavir mainly affects the transport activity of renal P-gp and may only minimally decrease intestinal and liver P-gp activity following saturation of the CYP3A enzyme. This implies minimal impact of ritonavir on ISD interaction mediated through P-gp.

Additional Information

Ritonavir is a strong activator of the pregnane X receptor, which regulates the expression of various metabolic enzymes, including CYP1A2, CYP2B6, CYP2C9, CYP2C19, and uridine diphospho-glucuronosyltransferases (UGTs).¹² The activities of CYP2C8, CYP2D6, organic anion transport protein, and breast cancer resistance protein decreased in the presence of ritonavir.^{12,17}

NIRMATRELVIR/RITONAVIR WITH CALCINEURIN INHIBITORS

The calcineurin inhibitors (CNIs), tacrolimus, and cyclosporine are both narrow therapeutic index drugs.²² For organ transplant recipients, target concentrations of CNIs have been defined, depending on immunological risk, time after transplantation, cotreatment with other ISDs, and other factors.²³ DDIs are frequent because cyclosporine and tacrolimus are metabolized by CYP3A enzymes (in the gut wall and liver) and they are also substrates of P-gp,²⁴ which is expressed at high levels in the liver, gastrointestinal tract, and kidneys.²⁵

A formal study of DDIs in healthy volunteers showed that in the presence of steady-state concentrations of ritonavir (100 mg once daily), the dose-normalized cyclosporine concentration at 24 hours (C₂₄), and the AUC_{0–inf} were 15.8-fold and 5.8-fold higher, respectively.²⁶ The effect on tacrolimus pharmacokinetics was even more drastic with a 17-fold and 57-fold increase in dose-normalized tacrolimus C₂₄ and AUC_{0–inf}, accordingly. The half-lives of cyclosporine and tacrolimus increased from 7 to 25 h and 32–232 hours, correspondingly. For both CNIs, the effect on C_{max} was lower, indicating that the reduced clearance by the liver was more likely the primary mechanism of the interaction than increased oral bioavailability. Thus, these results suggest that CYP3A inhibition, rather than P-gp, is the primary mechanism of this interaction. Keeping the same dose of tacrolimus after the initial ritonavir treatment will lead to an extremely high tacrolimus exposure within 24 hours. There is a need for a significant dose reduction, to avoid potentially toxic concentrations. For anti-HIV therapies in the context of liver transplantation, it has been suggested that a dose reduction of tacrolimus as low as 0.5 or 1.0 mg once a week when coadministered with ritonavir boosted lopinavir therapy is necessary.²⁷

Drug interaction with both CNIs is significant and may lead to toxic concentrations of CNIs for a prolonged period; however, monitoring of the magnitude of the interaction through drug concentration measurements can provide a way to manage this potentially dangerous interaction in those patients who may simultaneously need both drugs.

RECOMMENDATIONS

Tacrolimus

For most patients, it seems best to discontinue tacrolimus 12 h before nirmatrelvir/ritonavir therapy is started. The predose concentration measured at the time of starting nirmatrelvir/ritonavir is expected to be maintained at approximately 75% of the initial concentration at the end of the 5 days of the antiviral treatment, whereas the magnitude of change in tacrolimus total exposure (AUC_{0–120 h}) has been more difficult to estimate. Conversely, in the context of active COVID-19, a small reduction in the exposure of ISDs may contribute to faster viral clearance. If possible, we recommend a “keep it simple” approach and discontinue tacrolimus during the entire 5 days of antiviral treatment with nirmatrelvir/ritonavir.

A less radical alternative and intended only for high-risk patients (patients in the early posttransplant period or

those with a high risk of rejection) would be to administer a small tacrolimus dose (one-eighth of patient standard daily dose) on day 1 of nirmatrelvir/ritonavir therapy (with no further dosing for at least 5 days) to maintain a certain level of tacrolimus exposure during the next consecutive 5 days of nirmatrelvir/ritonavir therapy. This approach also has the advantage of maintaining exposure (considering AUC_{0–120 h}) after antiviral treatment initiation closer to the exposure provided by the patient’s initial dosage before nirmatrelvir/ritonavir initiation. Such a strategy should be supported by a close collaboration between pharmacologists and clinicians (Table 1).

Upon discontinuation of nirmatrelvir/ritonavir, CYP3A activity will recover with time. The half-life of ritonavir is short (3–5 hours), but published data suggest that it may take 3 days for CYP3A activity to completely restore, achieving approximately 75% of metabolic activity after 48 hours.²⁸ Therefore, a pragmatic approach would be to extend the temporary discontinuation of tacrolimus treatment for a few more days with cautious dosing under close surveillance after tacrolimus exposure. After 7–8 days, tacrolimus can be reintroduced, either in the dose given before nirmatrelvir/ritonavir treatment or based on measured tacrolimus concentrations at the time of restarting tacrolimus treatment (as suggested by Lange et al).²⁹

Cyclosporine

The impact of cyclosporine on ritonavir is somewhat less than for tacrolimus. Nevertheless, cyclosporine overexposure accompanied by signs of severe nephrotoxicity has been reported in patients on ritonavir.³⁰ After nirmatrelvir/ritonavir introduction, the total daily dose of cyclosporine should be reduced to 20% of the initial dose and administered once daily to maintain similar blood cyclosporine concentrations. This degree of dose reduction is comparable to what has been described with direct antiviral agents.^{26,31} Regular cyclosporine concentration monitoring is advised to guide dosing, especially after discontinuation of nirmatrelvir/ritonavir (Table 1). TDM of tacrolimus and cyclosporine is advised to guide safe dosing (see dedicated section).

NIRMATRELVIR/RITONAVIR WITH MTORIS: EVEROLIMUS AND SIROLIMUS

To date, there are no reports on the optimal dosing strategy for mTORis, such as everolimus and sirolimus, when combined with nirmatrelvir/ritonavir or other ritonavir formulations. In the clinical trials that were performed, patients who used medications that are highly dependent on CYP3A4 for clearance, such as sirolimus and everolimus, were excluded. To provide tentative dose adjustments for solid organ transplant recipients on mTORis when starting on nirmatrelvir/ritonavir, we therefore need to rely on the information from drug interactions that have similar properties with respect to CYP3A-mediated metabolism and P-gp-mediated transport inhibition. For everolimus and sirolimus, there are few retrospective reports that could provide guidance to an appropriate dosing strategy. A study of sirolimus dosing requirements in patients on HIV therapy with ritonavir

TABLE 1. Recommendations for Dose Adjustment of Immunosuppressive Drugs When Combined With Nirmatrelvir/Ritonavir, Restart Strategies, and Therapeutic Drug Monitoring

Immunosuppressive Agent	Recommended Immunosuppressive Drug Dose Adjustment	Alternative Immunosuppressive Drug Dose Adjustment	Restart of Immunosuppressive Drug	Proposition for TDM (if Feasible)
Tacrolimus	Discontinue tacrolimus 12 h before initiation of nirmatrelvir/ritonavir	Administer one-eighth of the usual daily dose on day 1 and then discontinue	Tacrolimus can be reintroduced on day 7 or 8	Two sample time points to capture drug elimination and TDM at treatment restoration or shortly thereafter (if possible, an AUC monitoring should be conducted)
Cyclosporine	Reduce to 20% of the initial dose from day-1	—	Progressively reintroduced from day-6	Trough concentrations (be aware that pharmacokinetic profile can change during antiviral treatment) and TDM at treatment restoration or shortly thereafter
m-TOR inhibitors	Discontinue m-TOR inhibitor 12 h before the initiation of nirmatrelvir/ritonavir	Administer one-eighth of the usual daily dose on day 1, 3, and 5	m-TOR can be reintroduced on day 7	Trough concentrations (on day 3 in case of applying the alternative drug adjustment) and then TDM at treatment restoration or shortly thereafter
MPA	No need to adjust drug dosage given the short antiviral treatment duration	MPA can be discontinued in some cases	—	TDM according to local practice
Corticosteroids	No need to adjust drug dosage	—	—	No need for TDM

AUC, area under the curve of drug concentrations; m-TOR, mammalian target of rapamycin inhibitor.

combination, 1/10 to 1/20 of the typical dose of sirolimus has been recommended.³²

In a crossover study of 12 healthy subjects, a single oral everolimus dose of 2 mg was given on day 1 under fasting conditions. Starting on day 10, 200 mg of ketoconazole was administered orally every 12 hours, and 4 days later (day 13), everolimus was given again, at a dose of 1 mg. When the results were normalized to a 2-mg dose for comparison, the C_{max} increased on average 3.9-fold (90% CI, 3.4–4.6); however, the AUC increases were clustered largely in the range from 11.2-fold to 17.5-fold (n = 11) with ketoconazole. The average increase in AUC across all subjects was 15.0-fold (90% CI, 13.6–16.6). The half-life of everolimus increased from an average of 30 hours (SD = 4) to 56 hours (SD = 5).³³ Although the results were obtained with the combination of ketoconazole and not with ritonavir, they indicate the potential influence of CYP3A4 and P-gp inhibition on changing in everolimus exposure. It is expected that the effect of ritonavir would be much greater.

A 35-year-old patient with COVID-19 developed a very high everolimus concentration during cotreatment with lopinavir/ritonavir (400/100 mg BID). The everolimus dose was initially reduced by one third, but predose concentrations reached a maximum of 31.1 ng/mL. The authors recommended immediate withdrawal of mTORi therapy and close monitoring of blood concentrations, clinical status, and signs of drug toxicity.³⁴

A recent study included adult transplant recipients treated with nirmatrelvir/ritonavir for 5 days; among these patients, 3 were on everolimus and 1 on sirolimus.³⁵

According to the protocol,²⁹ the mTORi was withheld at the start of nirmatrelvir/ritonavir. The most recent mTORi predose concentration had been measured 77 days (IQR, 58–140) before the start of nirmatrelvir/ritonavir, and these concentrations averaged 4.8 ng/mL (IQR, 3–4.9). After completion of the nirmatrelvir/ritonavir, 2 patients had undetectable concentrations on day 7 and day 9, whereas the third patient had 1.4 ng/mL on day 8. The patient on sirolimus had a trough concentration of 5 ng/mL 13 days before starting nirmatrelvir/ritonavir and 9.5 ng/mL on day 14. Another case report advised sirolimus dosage reductions to 1.5 mg per week and 1 mg per 14 days.³⁶

Recommendations

Based on these very limited observations, the following is recommended for the dosing of mTORis during and following nirmatrelvir/ritonavir treatment:

- Approximately 12 hours before nirmatrelvir/ritonavir initiation, the mTORi should be put on hold. From the day after the cessation of nirmatrelvir/ritonavir, the mTORi may be reintroduced in one-fifth of the original dose, and the dose may be increased by 20% each day or according to the measured mTORi trough concentrations.³⁷ The timing and selection of dose for reintroduction of the mTORi may also depend on which combination of other immunosuppressants are used and the importance of the contribution of mTORis in the immunosuppressive regimen.

Alternatively, if it is considered important to avoid subtherapeutic mTORi concentrations, and given the low risk

of immediate adverse drug reactions with these drugs, another strategy that can be considered is as follows:

- A microdose of one-eighth of the patient's initial daily dose should be given on days 1, 3, and 5 after starting nirmatrelvir/ritonavir, and the initial treatment dosage could be restarted on day 7 (Table 1).

It is recommended to frequently monitor immunosuppressant drug levels, at least right before dose administration and shortly after the end of nirmatrelvir/ritonavir treatment (see dedicated section).

NIRMATRELVIR/RITONAVIR WITH MYCOPHENOLIC ACID

Mycophenolic acid (MPA), administered as a prodrug (mycophenolate mofetil, the morpholinoethyl ester of MPA) or an enteric-coated salt (mycophenolate sodium), is one of the most important drugs for the immunosuppressive regimen in organ transplant recipients. During viral infections, MPA is usually the first ISD with a dose reduction, if not discontinued.³⁸ The drug has also been shown to strongly decrease SARS-CoV-2 vaccine efficacy.³⁹ Therefore, therapeutic management of transplant recipients infected with SARS-CoV-2 requires careful review, and a special consideration should be given to potential interactions with approved COVID-19 treatments, such as nirmatrelvir/ritonavir.

The metabolism of MPA is extensive and mostly occurs in the liver, intestine, and kidney through the uridine 5'-diphospho-glucuronosyltransferase (UGT) system. The main UGT isoforms involved in MPA glucuronidation are UGT1A9 and UGT2B7.^{40,41} The glucuronidation pathway produces a major metabolite, MPA 7-O-glucuronide, which is inactive. A minor phase-I metabolite from human cytochrome P450 isoforms, CYP3A4 and CYP3A5, the 6-O-desmethyl-MPA (DM-MPA) and its related glucuronide were also identified in blood and urine from transplant patients.⁴²

Expected DDIs with nirmatrelvir/ritonavir may result from ritonavir being an inhibitor and inducer of UGT. Different studies have underscored the distinct effects of ritonavir on UGT isoforms. Ritonavir has been shown *in vitro* to consistently inhibit UGT1A1, UGT1A3, and UGT1A4 and weakly inhibit UGT1A6, UGT1A9, and UGT2B7 ($IC_{50} > 100 \mu\text{M}$).⁴³ Conversely, ritonavir has been shown to induce UGTs, in clinical studies because it significantly decreased systemic exposure to some drugs eliminated from the body by the glucuronidation pathway like ethinyl oestradiol⁴⁴ and lamotrigine.⁴⁵

Ritonavir seems to only weakly interact with UGT isoforms (1A9 and 2B7) involved in MPA metabolism as evidenced by the absence of significant modification in AUC in a HCV-infected patient treated with the combination of ombitasvir, paritaprevir/ritonavir, dasabuvir, and MPA for vasculitis.⁴⁶ Moreover, a successful recovery from COVID-19 has been reported in a patient with systemic lupus erythematosus treated concomitantly with lopinavir/ritonavir and the SLE drugs including MPA.⁴⁷ Overall, a limited impact of nirmatrelvir/ritonavir is expected on MPA metabolism and exposure.

Recommendations

Given the short duration (5 days) of nirmatrelvir/ritonavir treatment, and because of the low potential of pharmacokinetic interactions, there is no need to adjust the dosage of MPA. However, in solid organ transplant patients with COVID-19, the first drug to be temporarily discontinued to allow clearance of the virus is MPA (Table 1).

NIRMATRELVIR/RITONAVIR WITH CORTICOSTEROIDS

Corticosteroids are an integral part of induction and maintenance in immunosuppressive regimens in solid organ transplantation. Corticosteroids are also a drug category that is used in acute therapy but at the severe, advanced stage of COVID-19 infection. Prednisone, prednisolone, and methylprednisolone are the most commonly used synthetic corticosteroids in transplant recipients. Prednisone is a prodrug converted through first-pass metabolism by 11- β -hydroxy dehydrogenase to its active form, prednisolone.⁴⁸

Both prednisolone and prednisone undergo 6 β -hydroxylation by means of the CYP3A4 metabolic pathway and are inducers of multidrug-associated resistance protein 2, as well as substrates, inhibitors, and inducers of P-gp.^{49,50} Corticosteroid clearance has been reported to be significantly reduced in patients on ritonavir-boosted protease inhibitors resulting in increased concentration.⁵¹ In these patients, the higher exposure has been associated with bone toxicities (eg, osteonecrosis) and Cushing syndrome.⁵² In the French Pharmacovigilance Database, antiretrovirus-boosting agents in combination with corticosteroids were incriminated in several cases of iatrogenic Cushing syndrome, but this has only been reported in chronic treatment and is unlikely to happen during a 5-day treatment course with ritonavir.⁵³

Recommendations

Nirmatrelvir/ritonavir combination with corticosteroids in transplant patients could presumably lead to a temporary increase in prednisolone exposure due to CYP3A4 and P-glycoprotein inhibition. We recommend maintaining the same dosage of corticosteroid during the 5-day nirmatrelvir/ritonavir course (Table 1).

TDM OF IMMUNOSUPPRESSIVE DRUGS WITH NIRMATRELVIR/RITONAVIR

TDM Recommendations for CNIs to Modulate DDIs With Nirmatrelvir/Ritonavir-Based Treatment

The primary objective of TDM is to minimize the toxicity of ISDs while preventing any under- and over-immunosuppression possibly leading to graft rejection due to improper discontinuation or of immunosuppression or toxicity, respectively. Here, we provide some general guidance for monitoring patients simultaneously on nirmatrelvir/ritonavir and ISDs. They are also summarized in Table 1.

When tacrolimus is discontinued before the antiviral treatment, as proposed in these recommendations, there is a

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limited risk of toxicity. Given the fact that nirmatrelvir/ritonavir treatment has been designed for outpatients, it is possible to avoid TDM during the 5-day course unless there is a safe way to apply TDM (ideally autosampling at home).

The same conclusion can be formulated for cyclosporine. It is important to note that blood concentration versus time curve of CNIs combined with ritonavir is flat, with a low C_{max} (related to the low dose) and an almost horizontal slope (given the long half-life). As a result, the relationship between the predose concentration and the AUC is different from the situation where CNI is taken twice daily in a much higher dose. The mean AUC in ritonavir-treated patients was 40% lower compared with the AUC seen in those not cotreated with ritonavir, when similar trough levels were targeted.⁵⁴

In case of using a small tacrolimus boost (one-eighth of the standard dose corresponding to a 40-fold dose reduction with respect to the 5 previous days of tacrolimus treatment) on day 1 of nirmatrelvir/ritonavir therapy, for hospitalized patients, patients at high-risk of rejection or at the initial postoperative period, a more comprehensive TDM approach can be proposed. This may include 2 early sample points (eg, just before starting nirmatrelvir/ritonavir, at the end of day 1 and day 2) to capture the elimination rate constant of the drug, inform patient's exposure by approximating the AUC, and help defining the time to restart drug dosing. This approach will prevent toxic tacrolimus exposure and minimize levels falling below the target concentrations. For TDM of tacrolimus and cyclosporine, the trough concentrations are more feasible to obtain, but the AUC may better reflect the impact of DDIs in the whole PK profile.²³ These patients may benefit from limited sampling strategy, with Bayesian estimation of the total AUC, and dried blood spot analysis should be considered, particularly for tacrolimus.^{55,56}

Given that 70%–90% of CYP3A enzyme activity can be expected to recover after day 3 of ritonavir, discontinuation of nirmatrelvir/ritonavir TDM can also be proposed at the time of CNI treatment restoration or shortly thereafter. If the treatment with nirmatrelvir/ritonavir would be continued for a much longer time than just 5 days, it would make sense to aim for higher predose target concentrations.⁵⁷

TDM Recommendations for mTORis to Modulate DDIs With Nirmatrelvir/Ritonavir-Based Treatment

For sirolimus and everolimus, a similar interaction with boosted protease inhibitors, to what is reported with tacrolimus, is expected, although less well-defined.³² If the mTOR is withdrawn, ideally, mTOR concentrations should be monitored before starting the antiviral drug. Further dose adjustments will be necessary to reach target concentrations. In case of using a microdose of one-eighth of the patient's initial daily dose given on day 1, 3, and 5, trough concentration may be measured on day 3 of antiviral treatment to potentially adjust the subsequent dose. In any case, when the mTORi is reintroduced, trough concentrations should be monitored every second day until stable on a fixed dose, and the mTORi should be restarted according to the TDM results. If out of therapeutic range, measurements should be repeated

until the concentration is within range, and then, the mTORi dose restarted or adjusted.⁵⁸ Again, the implementation of TDM in an outpatient setting can be facilitated using a microsampling approach.

TDM Recommendations for Mycophenolic Acid to Modulate DDIs With Nirmatrelvir/Ritonavir-Based Treatment

If MPA treatment is maintained during antiviral therapy, TDM can be performed, if needed, according to local practices. This would help generate more data on this topic, including careful descriptions of case reports. Until such data are available, TDM might be a useful tool to maintain adequate exposure of MPA, for example, using a limited sampling strategy or a Bayesian approach.⁵⁸

CONCLUSIONS

Combining nirmatrelvir/ritonavir with narrow therapeutic index drugs metabolized by CYP3A4 and P-gp is challenging. This is particularly true for tacrolimus and, to a lesser extent, for cyclosporine, everolimus, and sirolimus. Data from studies on DDIs between ISDs and potent CYP3A4 inhibitors obtained in the field of HIV and HCV enable the generation of recommendations for dosage adjustments of these drugs and to secure their prescription during antiviral treatment. Although not easy to implement in an outpatient setting, TDM may be a useful tool to confirm that drug adjustments are appropriate in a specific case and to help reintroducing ISDs at the end of nirmatrelvir/ritonavir treatment. A TDM approach during the treatment also appears appropriate when a strict drug exposure conservation must be ensured.

REFERENCES

1. The U.S. Food and Drug Administration (FDA). Fact sheet for healthcare providers: emergency use authorization for Paxlovid. Available at: <https://www.fda.gov/media/155050/download>. Accessed March 18, 2022.
2. National Institutes of Health (NIH). COVID-19 Treatment Guidelines Panel. Coronavirus disease 2019 (COVID-19) treatment guidelines. Available at: https://files.covid19treatmentguidelines.nih.gov/guidelines/section/section_171.pdf. Accessed February 24, 2022.
3. Hammond J, Leister-Tebbe H, Gardner A, et al. Oral nirmatrelvir for high-risk, nonhospitalized adults with covid-19. *N Engl J Med*. 2022; 386:1397–1408.
4. Cohen P. COVID-19: outpatient evaluation and management of acute illness in adults. *UpToDate*. 2022. Available at: <https://www.uptodate.com/contents/covid-19-outpatient-evaluation-and-management-of-acute-illness-in-adults>. Accessed March 20, 2022.
5. European Medicines Agency (EMA). Paxlovid European public assessment report (EPAR). Available at: https://www.ema.europa.eu/en/documents/product-information/paxlovid-epar-product-information_en.pdf. Accessed January 28, 2022.
6. ClinicalTrials.gov. Available at: <https://clinicaltrials.gov/ct2/show/record/NCT04535167>. Accessed March 21, 2022.
7. ClinicalTrials.gov. Available at: <https://clinicaltrials.gov/ct2/show/NCT04756531>. Accessed March 21, 2022.
8. The U.S. Food and Drug Administration (FDA). Emergency Use Authorization 105 Paxlovid (nirmatrelvir tablets and ritonavir tablets, co-packaged for oral use). Available at: <https://www.fda.gov/media/155071/download>. Accessed December 22, 2021.
9. Lemaitre F, Grégoire M, Monchaud C, et al. Management of drug-drug interactions with nirmatrelvir/ritonavir in patients treated for Covid-19: guidelines from the French Society of Pharmacology and Therapeutics (SFPT). *Therapies*. 2022;77(5):509–521.

10. Liverpool Drug Interactions Group. COVID-19 drug interactions. Available at: <https://www.covid19-druginteractions.org/>. Accessed April 25, 2022.
11. Foisy MM, Yakiwchuk EM, Hughes CA. Induction effects of ritonavir: implications for drug interactions. *Ann Pharmacother*. 2008;42:1048–1059.
12. Cattaneo D, Cossu MV, Rizzardini G. Pharmacokinetic drug evaluation of ritonavir (versus cobicistat) as adjunctive therapy in the treatment of HIV. *Expert Opin Drug Metab Toxicol*. 2019;15:927–935.
13. Zhou SF, Xue CC, Yu XQ, et al. Clinically important drug interactions potentially involving mechanism-based inhibition of cytochrome P450 3A4 and the role of therapeutic drug monitoring. *Ther Drug Monit*. 2007; 29:687–710.
14. Ernest CS, II, Hall SD, Jones DR. Mechanism-based inactivation of CYP3A by HIV protease inhibitors. *J Pharmacol Exp Ther*. 2005;312: 583–591.
15. Sevrioukova IF, Poulos TL. Structure and mechanism of the complex between cytochrome P4503A4 and ritonavir. *Proc Natl Acad Sci U S A*. 2010;107:18422–18427.
16. Badri PS, Dutta S, Wang H, et al. Drug interactions with the direct-acting antiviral combination of ombitasvir and paritaprevir-ritonavir. *Antimicrob Agents Chemother*. 2016;60:105–114.
17. Pal D, Mitra AK. MDR- and CYP3A4-mediated drug-drug interactions. *J Neuroimmune Pharmacol official J Soc NeuroImmune Pharmacol*. 2006; 1:323–339.
18. Huisman MT, Smit JW, Wiltshire HR, et al. P-glycoprotein limits oral availability, brain, and fetal penetration of saquinavir even with high doses of ritonavir. *Mol Pharmacol*. 2001;59:806–813.
19. Penzak SR, Shen JM, Alfaro RM, et al. Ritonavir decreases the nonrenal clearance of digoxin in healthy volunteers with known MDR1 genotypes. *Ther Drug Monit*. 2004;26:322–330.
20. Ding R, Tayrouz Y, Riedel KD, et al. Substantial pharmacokinetic interaction between digoxin and ritonavir in healthy volunteers. *Clin Pharmacol Ther*. 2004;76:73–84.
21. Kirby BJ, Collier AC, Kharasch ED, et al. Complex drug interactions of the HIV protease inhibitors 3: effect of simultaneous or staggered dosing of digoxin and ritonavir, nelfinavir, rifampin, or bupropion. *Drug Metab Dispos*. 2012;40:610–616.
22. Neuberger JM, Bechstein WO, Kuypers DRJ, et al. Practical recommendations for long-term management of modifiable risks in kidney and liver transplant recipients: a guidance report and clinical checklist by the consensus on managing modifiable risk in transplantation (COMMIT) group. *Transplantation*. 2017;101:S1–S56.
23. Brunet M, van Gelder T, Asberg A, et al. Therapeutic drug monitoring of tacrolimus-personalized therapy: second consensus report. *Ther Drug Monit*. 2019;41:261–307.
24. van Gelder T. Drug interactions with tacrolimus. *Drug Saf*. 2002;25:707–712.
25. Thiebaut F, Tsuruo T, Hamada H, et al. Cellular localization of the multidrug-resistance gene product P-glycoprotein in normal human tissues. *Proc Natl Acad Sci U S A*. 1987;84:7735–7738.
26. Badri P, Dutta S, Coakley E, et al. Pharmacokinetics and dose recommendations for cyclosporine and tacrolimus when coadministered with ABT-450, ombitasvir, and dasabuvir. *Am J Transpl*. 2015;15:1313–1322.
27. Jain AB, Venkataramanan R, Egtesad B, et al. Effect of coadministered lopinavir and ritonavir (Kaletra) on tacrolimus blood concentration in liver transplantation patients. *Liver Transpl*. 2003;9:954–960.
28. Katzenmaier S, Markert C, Riedel KD, et al. Determining the time course of CYP3A inhibition by potent reversible and irreversible CYP3A inhibitors using a limited sampling strategy. *Clin Pharmacol Ther*. 2011;90: 666–673.
29. Lange NW, Salerno DM, Jennings DL, et al. Nirmatrelvir/ritonavir use: managing clinically significant drug-drug interactions with transplant immunosuppressants. *Am J Transpl*. 2022;22:1925–1926.
30. Gregoor PJ, van Gelder T, van der Ende ME, et al. Cyclosporine and triple-drug treatment with human immunodeficiency virus protease inhibitors. *Transplantation*. 1999;68:1210.
31. Mori T, Aisa Y, Kato J, et al. Drug interaction between voriconazole and calcineurin inhibitors in allogeneic hematopoietic stem cell transplant recipients. *Bone Marrow Transpl*. 2009;44:371–374.
32. Krown SE, Roy D, Lee JY, et al. Rapamycin with antiretroviral therapy in AIDS-associated Kaposi sarcoma: an AIDS Malignancy Consortium study. *J Acquir Immune Defic Syndr*. 2012;59:447–454.
33. Kovarik JM, Beyer D, Bizot MN, et al. Blood concentrations of everolimus are markedly increased by ketoconazole. *J Clin Pharmacol*. 2005; 45:514–518.
34. Meziyerh S, Zwart TC, van Etten RW, et al. Severe COVID-19 in a renal transplant recipient: a focus on pharmacokinetics. *Am J Transpl*. 2020; 20:1896–1901.
35. Salerno DM, Jennings DL, Lange NW, et al. Early clinical experience with nirmatrelvir/ritonavir for the treatment of COVID-19 in solid organ transplant recipients. *Am J Transpl*. 2022;22:2083–2088.
36. Dolman GE, Selby P, Gelson WT. Ombitasvir/paritaprevir/ritonavir plus dasabuvir regimen may be used safely in combination with sirolimus for the treatment of chronic hepatitis C. *BMJ Case Rep*. 2018;2018: bcr2018224664.
37. Zijp TR, Toren-Wielema ML, Nannan Panday PV, et al. Important interactions of immunosuppressants with experimental therapies for novel coronavirus disease (COVID-19): how to act. *Ther Drug Monit*. 2020; 42:652–653.
38. Lai Q, Spoletini G, Bianco G, et al. SARS-CoV2 and immunosuppression: a double-edged sword. *Transpl Infect Dis*. 2020;22:e13404.
39. Rabinowich L, Grupper A, Baruch R, et al. Low immunogenicity to SARS-CoV-2 vaccination among liver transplant recipients. *J Hepatol*. 2021;75:435–438.
40. Picard N, Ratanasavanh D, Premaud A, et al. Identification of the UDP-glucuronosyltransferase isoforms involved in mycophenolic acid phase II metabolism. *Drug Metab Dispos*. 2005;33:139–146.
41. Mackenzie PI. Identification of uridine diphosphate glucuronosyltransferases involved in the metabolism and clearance of mycophenolic acid. *Ther Drug Monit*. 2000;22:10–13.
42. Picard N, Cresteil T, Premaud A, Marquet P. Characterization of a phase I metabolite of mycophenolic acid produced by CYP3A4/5. *Ther Drug Monit*. 2004;26:600–608.
43. Zhang D, Chando TJ, Everett DW, et al. In vitro inhibition of UDP glucuronosyltransferases by atazanavir and other HIV protease inhibitors and the relationship of this property to in vivo bilirubin glucuronidation. *Drug Metab Dispos*. 2005;33:1729–1739.
44. Ouellet D, Hsu A, Qian J, et al. Effect of ritonavir on the pharmacokinetics of ethinyl oestradiol in healthy female volunteers. *Br J Clin Pharmacol*. 1998;46:111–116.
45. van der Lee MJ, Dawood L, ter Hofstede HJM, et al. Lopinavir/ritonavir reduces lamotrigine plasma concentrations in healthy subjects. *Clin Pharmacol Ther*. 2006;80:159–168.
46. Lemaitre F, Ben Ali Z, Tron C, et al. Managing drug-drug interaction between ombitasvir, paritaprevir/ritonavir, dasabuvir, and mycophenolate mofetil. *Ther Drug Monit*. 2017;39:305–307.
47. He F, Luo Q, Lei M, et al. Successful recovery of recurrence of positive SARS-CoV-2 RNA in COVID-19 patient with systemic lupus erythematosus: a case report and review. *Clin Rheumatol*. 2020;39:2803–2810.
48. Lindenfeld J, Miller GG, Shakar SF, et al. Drug therapy in the heart transplant recipient: part II: immunosuppressive drugs. *Circulation*. 2004;110:3858–3865.
49. Christians U, Strom T, Zhang YL, et al. Active drug transport of immunosuppressants: new insights for pharmacokinetics and pharmacodynamics. *Ther Drug Monit*. 2006;28:39–44.
50. Bergmann TK, Barraclough KA, Lee KJ, Staatz CE. Clinical pharmacokinetics and pharmacodynamics of prednisolone and prednisone in solid organ transplantation. *Clin Pharmacokinet*. 2012;51:711–741.
51. Lauterio A, Valsecchi M, Santambrogio S, et al. Successful recovery from severe COVID-19 pneumonia after kidney transplantation: the interplay between immunosuppression and novel therapy including tocilizumab. *Transpl Infect Dis*. 2020;22:e13334.
52. Penzak SR, Formentini E, Alfaro RM, et al. Prednisolone pharmacokinetics in the presence and absence of ritonavir after oral prednisone administration to healthy volunteers. *J Acquir Immune Defic Syndr*. 2005;40:573–580.
53. Peyro-Saint-Paul L, Besnier P, Demessine L, et al. Cushing's syndrome due to interaction between ritonavir or cobicistat and corticosteroids: a case-control study in the French Pharmacovigilance Database. *J Antimicrob Chemother*. 2019;74:3291–3294.

54. van Maarseveen EM, Crommelin HA, Mudrikova T, et al. Pretransplantation pharmacokinetic curves of tacrolimus in HIV-infected patients on ritonavir-containing cART: a pilot study. *Transplantation*. 2013;95:397–402.
55. Benkali K, Rostaing L, Premaud A, et al. Population pharmacokinetics and Bayesian estimation of tacrolimus exposure in renal transplant recipients on a new once-daily formulation. *Clin Pharmacokinet*. 2010;49:683–692.
56. Velghe S, De Troyer R, Stove C. Dried blood spots in therapeutic drug monitoring and toxicology. *Expert Opin Drug Metab Toxicol*. 2018;14:1–3.
57. van Maarseveen EM, van Zuilen AD, Mudrikova T. Outcomes of kidney transplantation in HIV-infected recipients. *N Engl J Med*. 2011;364:683. author reply 684.
58. Bergan S, Brunet M, Hesselink DA, et al. Personalized therapy for mycophenolate: consensus report by the international association of therapeutic drug monitoring and clinical toxicology. *Ther Drug Monit*. 2021;43:150–200.

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