



https://helda.helsinki.fi

Potential of Wood Hemicelluloses and Their Derivates as Food Ingredients

Abik, Felix

2023-02-15

```
Abik, F, Palasingh, C, Bhattarai, M, Leivers, S, Ström, A, Westereng, B, Mikkonen, K
S & Nypelö, T 2023, 'Potential of Wood Hemicelluloses and Their Derivates as Food
þÿlngredients ', Journal of Agricultural and Food Chemistry, vol. 71,
```

http://hdl.handle.net/10138/356832 https://doi.org/10.1021/acs.jafc.2c06449

cc_by publishedVersion

Downloaded from Helda, University of Helsinki institutional repository.

This is an electronic reprint of the original article.

This reprint may differ from the original in pagination and typographic detail.

Please cite the original version.

Potential of Wood Hemicelluloses and Their Derivates as Food Ingredients

Felix Abik, Chonnipa Palasingh, Mamata Bhattarai, Shaun Leivers, Anna Ström, Bjørge Westereng,* Kirsi S. Mikkonen,* and Tiina Nypelö*

Cite This: J. Agric. Food Chem. 2023, 71, 2667–2683		Read Online		
ACCESS	III Metrics & More		E Article Recommendations	

ABSTRACT: A holistic utilization of all lignocellulosic wood biomass, instead of the current approach of using only the cellulose fraction, is crucial for the efficient, ecological, and economical use of the forest resources. Use of wood constituents in the food and feed sector is a potential way of promoting the global economy. However, industrially established food products utilizing such components are still scarce, with the exception of cellulose derivatives. Hemicelluloses that include xylans and mannans are major constituents of wood. The wood hemicelluloses are structurally similar to hemicelluloses from crops, which are included in our diet, for example, as a part of dietary fibers. Hence, structurally similar wood hemicelluloses have the potential for similar uses. We review the current status and future potential of wood hemicelluloses as food ingredients. We include an inventory of the extraction routes of wood hemicelluloses, their physicochemical properties, and some of their gastrointestinal characteristics, and we also consider the regulatory route that research findings need to follow to be approved for food solutions, as well as the current status of the wood hemicellulose applications on that route.

KEYWORDS: xylans, mannans, emulsions, novel foods

1. INTRODUCTION

Motivated by the ever-increasing human population, new food ingredients are being discovered and developed at a remarkable rate. These new food ingredients must fulfill various quality criteria to be widely accepted for consumption, including safety, technological, nutritional, economical, and environmental aspects. There is also an increasing societal demand for cleanlabel food products that are derived naturally from biomass instead of relying on synthetic materials.¹ While there have been technological efforts to reduce food waste and make the food industry more circular, as well as the booming development of plant-based food, it is also possible to obtain edible material from seemingly inedible sources.² One such promising alternative source is lignocellulosic biomass, in particular, wood, which contains hemicelluloses that can be developed into functional food ingredients. From our perspective, wood hemicelluloses present a large opportunity to be developed into and marketed as naturally derived food ingredients, fulfilling the current industrial demands of clean label food and feed products.

Hemicelluloses compose a remarkable family of polysaccharides found in plant cell walls. In food systems, they are long known as important dietary fibers, which can be extracted from various sources. One promising source of such extractable fibers is wood biomass. Hemicelluloses can be liberated from wood biomass as polymeric, oligomeric, and monomeric fractions with coextraction of noncellulosic compounds. This wide range of possible extractable grades of material leads to diversity in molar mass and physicochemical properties as well as opening a window toward numerous applications, including those as potential food ingredients. Complete exploitation of wood hemicelluloses allows resource-efficient use of wood biomass and minimizes volumes of waste streams currently generated from biomass processing within the forest sector.

This Review focuses on presenting the status of wood hemicelluloses and their derivates as food ingredients. Although the structure and properties of wood hemicelluloses as well as chemical reactions for their derivatization are largely established, currently there are only few academically demonstrated commercial and industrial solutions, and it has proven difficult to progress the scientific demonstrations into emerging applications. Thus, a description of the route to regulatory and legislative approval, the essential step from demonstration to food applications, is also included. Many of the academic studies deal with hemicelluloses in emulsions, and therefore the scienceto-technology approach requires an understanding of the mechanisms behind hemicelluloses at interfaces; therefore, research within that area is included in this Review. This Review also covers wood and wood-like hemicelluloses in general, common derivatization methods that are potentially acceptable for food applications, and their key physicochemical properties.

Readers are also encouraged to refer to previous reviews on the structure and solubility of hemicelluloses,³ isolation,

Received:September 16, 2022Revised:January 9, 2023Accepted:January 12, 2023Published:February 1, 2023





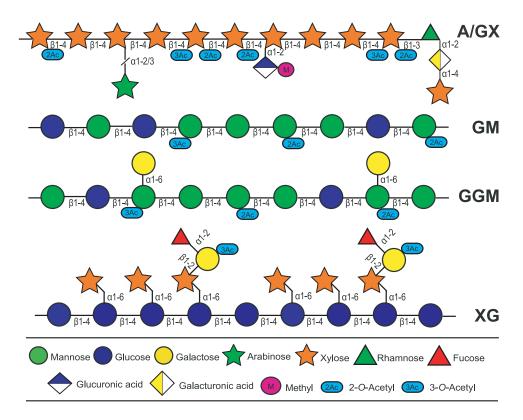


Figure 1. Structure of wood hemicelluloses, illustrating the backbones and side chains of the different types of hemicellulose. A/GX indicates that GX may exist in wood with or without arabinose residues. α 1-2, α 1-4, α 1-6, β 1-2, β 1-3, and β 1-4 indicate the type of glycosidic bonds; α 1-2/3 in A/GX indicates that the arabinose residue may attach to hydroxyl groups at position 2 or 3 of the xylose residue. 2-O-Acetyl and 3-O-acetyl indicate that acetylation may occur at the hydroxyl position 2 or 3 of the respective carbohydrate residue. The symbols are sketched according to Varki et al.¹⁷

structural characterization, and potential applications of bamboo hemicelluloses,⁴ recent developments and challenges of obtaining plant-based nanomaterials from plant residues,⁵ applications for functional foods,⁶ bioethanol,⁷ and oligosac-charide production,^{8,9} and polysaccharide degradation by microbial¹⁰ and technological¹¹ processing. Hemicelluloses in packaging materials have been reviewed recently¹² and are therefore excluded from this Review.

2. WOOD AND WOOD-LIKE HEMICELLULOSES

The hemicelluloses that can be liberated in industrially relevant yields from wood are xylans and glucomannans (GM), containing substituents such as arabinose (Ara), glucuronic acids (GlcA), and galacturonic acids (GalA) in xylans, and galactose (Gal) in mannans. Both types of polymers are also acetylated to various extents in the native state in the plants. Approximately 15-35 wt % of hardwood and 30-32 wt % of softwood macromolecular composition is made up of hemicelluloses.^{13–15} Softwoods such as Norway spruce (*Picea abies*) and white spruce (Picea glauca) are rich in galactoglucomannan (GGM) (16–17% of dry wood weight) with some arabinoglucuronoxylan (AGX) (8–10%), while hardwood, for example, paper birch (Betula papyrifera), is rich in glucuronoxylan (GX) (15–30%) with some GM (1-2%).^{14,15} Xyloglucans (XG) make up 20-25% of the cell wall in dicotyledonous angiosperms, 2-5% in grasses, and 10% in softwoods.¹⁶ However, due to their lesser abundance in wood, XGs are typically not considered as one of the main hemicellulose streams from wood, and as such we focus our discussions on

GGMs and GXs. Details regarding the structural features of both softwood and hardwood hemicelluloses have been extensively described by Sjöström¹⁴ and Ebringerova et al.,³ and an overview of the structure of these hemicelluloses is presented in Figure 1.

Because of the central and versatile role of hemicelluloses in the cell wall structure, wood is not the only source of xylans and mannans. In particular, xylans can be extracted as arabinoxylans (AX) from various types of nonwood biomass, such as sugar cane bagasse,¹⁸ straw,¹⁹ corn fibers,²⁰ cereal brans, and grass,² while mannans can be isolated as galactomannan (GalM) from seeds and beans²² or GM from konjac tubers.²³ Many of these hemicelluloses, such as GalM isolated from seeds, guar gum, and locust bean gum, entered the food system several decades ago.²² While there are structural differences between seed-based mannans and xylans as compared to those isolated from wood (such as the degree of acetylation for both, and glucuronic acid for the latter), they share a similar backbone and structural motifs. Because of the structural similarities, we therefore hypothesize that hemicelluloses from wood can also be considered as food ingredients. However, including a food ingredient from new sources, even if it is already in use from an existing source, requires approval and permission from the European Commission (EC).²⁴ The procedure is described later in this Review; acquiring the permission is currently a gatekeeper to the commercial use of wood hemicelluloses as food ingredients. Fortunately, wood as the source of biomass is not an intrinsic limitation. Cellulose derivatives obtained from wood are considered safe food and feed ingredients and are commonly

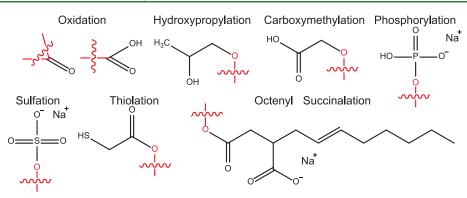


Figure 2. Structures of the common derivatizations performed on other polysaccharides potentially acceptable for food applications. Red wavy lines indicate the connection of the substituent groups to the polysaccharide chain.

used across industries (E460, E461, E462, E464, E465, E468, etc.). 25

2.1. Extraction of Hemicelluloses from Wood and Other Biomasses. There are multiple ways one can liberate hemicelluloses from wood biomass using hot water extraction of wood chips at neutral, acidic, and alkaline conditions, hot water, ^{26–28} neutral-, acidic-, and alkaline extraction of wood chips, delignified wood biomass, or wood pulp.^{29,30} New methods are continuously being developed to improve the efficiency, sustainability, and selectivity of the extraction process. In fact, improving the efficiency while reducing the cost of the extraction and purification processes may be the key to facilitating the exploitation of wood hemicelluloses when it comes to their extraction at an industrial scale.

The extraction conditions have been reported to have a large effect on the chemical composition of the hemicelluloses. Perez et al.³¹ reported that, for softwoods, near neutral or acidic extraction yields GGM with low residual lignin content, while alkaline extraction conditions lead to mixtures of GGM and AGX. Acidic treatment of hardwoods has been reported to produce mixtures of AGX and GGM, while the fractions obtained in alkaline conditions were comprised of AGX. In general, neutral and acidic conditions result in GGM fractions, while alkaline conditions favor xylan liberation.²⁶

Additionally, extraction conditions determine the purity of the isolated hemicellulose. Ebringerova and Heinze³² described a process for liberating xylan from the cell wall matrix. In a process targeting xylan, the hardwood species of choice is first delignified with acidic NaClO₂ or preferably with H₂O₂/NaOH. This leads to separation of biomass into a solubilized extract and a residue; the former requires purification if the other constituents from the plant material are not desired in the final product. The residue is subjected to alkaline extraction and results in a water-insoluble xylan extract. The purification step is considered demanding; nevertheless, fractionation of the hemicellulose and lignin components is most often desired. However, for certain applications, complete purification of the extract might not be necessary as was demonstrated for ligninrich spray-dried birch GX extract obtained by pressurized hotwater extraction (PHWE) that was found to be a more efficient emulsion stabilizer than its ethanol-precipitated counterpart containing less lignin.³³

Subjecting wood chips to hydrothermal treatments results in oligomeric fragments, either as part of a prehydrolysis step to generate dissolving pulp or exclusively to isolate the hemicelluloses as oligosaccharides or short polysaccharides. The most commonly explored method is the hydrothermal

hydrolysis of the wood biomass, either with the addition of a small amount of acid for prehydrolysis^{34,35} or without in the case of autohydrolysis, where the acetyl groups are released as acetic acid, subsequently acidifying the mixture.36,37 During the hydrolysis process, hemicellulose chains are cleaved into smaller fragments, which are then liberated into the hydrolysate. The hydrolysate may also contain several byproducts, including lignin, released as the wood structure is compromised. Furfurals from degraded pentoses and acetic acid from the cleaved acetyl groups of the hemicellulose may also be present. The production of these byproducts, in particular, furfurals and acetic acid, is increased as the extraction process becomes more severe.^{38,39} However, it is possible to eventually separate them from the hydrolysate to obtain the hemicelluloses and other components at a considerable purity. $^{40-42}$ Other hydrothermal extraction methods include steam explosion and microwave-assisted hydrothermal extraction, each with specific release and degradation kinetics as compared to conventional hydrothermal hydrolysis processes.^{43–46}

Ultimately, dealing with materials obtained from nature requires an understanding that the structure of the extracted materials will primarily be determined by the source. In the case of hemicelluloses, plants intrinsically have a source-specific dominant hemicellulose and carbohydrate composition, as well as various nonhemicellulose components that may be coextracted. In addition, it can be generally considered that hydrolysis leads to short chains, while hot water and alkaline extraction are more likely to generate polymeric hemicelluloses. With those points taken into consideration, a set of suitable extraction methods and conditions are chosen to achieve the desired product, as the extraction conditions also affect the physicochemical properties of the extracts. In essence, to obtain a specific type of hemicellulose with certain desired properties, one must choose carefully both the source of biomass and the suitable extraction method.

2.2. Common Derivatizations Potentially Acceptable for Food Applications. Although hemicelluloses have a range of naturally occurring derivatizations in the form of acetyl, methyl, and acidic groups as well as extensive branching,⁴⁷ further chemical modification and cross-linking may be required to achieve various specific effects in food.⁴⁸ We begin here by considering the available derivatizations for polysaccharides by inspecting examples of routinely chemically modified polysaccharides to illustrate the possible array of chemical synthesis tools, applicable also to wood hemicelluloses. Common derivatizations are shown in Figure 2. While reactions such as cross-linking and derivatization are accepted, particularly for

starches, highly defined inclusion levels must still be adhered to so as to meet strict purity criteria for use in food.^{49,50}

Starch, consisting of the polysaccharides amylose (α -(1 \rightarrow 4)glucoside) and amylopectin (α -(1 \rightarrow 4)-glucoside with α -(1 \rightarrow 6)-glucoside branching),⁵¹ can be readily modified, and numerous forms of derivatized starch have been approved for use as food additives.⁵⁰ Common, approved chemical derivatizations of starch include etherification ((carboxy)methylation,⁵² also used in food-grade cellulose; hydroxypropylation⁵³), esterification (acetylation, ⁵⁴ phosphorylation, ⁵⁵ and (octenyl)succinylation⁵⁶), and oxidation.⁵⁷ Cross-linking, enzymatic, and physical methodologies have also been routinely applied and extensively studied.⁵⁸ Such modifications can induce functionalities including lowering the viscosity, increasing the stability and/or clarity, improving the binding and moisture retention, gelation under controlled conditions, and increasing the crystallinity.⁵⁹ Pectins (E440), a class of galacturonans commonly used as food and beverage ingredients, are routinely de-esterified to alter the degree of methylester-ification.⁶⁰ They can also be amidated,⁶¹ and amidated pectin (E440b) is currently the only approved modified pectin used in the food industry.⁶² Nevertheless, numerous derivatized pectins have been assessed, particularly for their potential use in a medical setting (thiolated,⁶³ sulfated,⁶⁴ and oxidized⁶⁵ forms).

Many of the derivatizations and cross-linking strategies already approved for other polysaccharides have been fundamentally applied to both xylans and mannans. Beechwood xylan, a common target in xylan studies, has been shown to be successfully carboxymethylated⁶⁶ and homogeneously sulfated.⁶⁷ Other hemicellulose xylans, such as those derived from corn straw, have been phosphorylated and characterized with regards to functionality for further uses.⁶⁸ Spruce mannan was oxidized⁶⁹ and carboxymethylated⁷⁰ as a pretreatment technique to examine the effect on hydrolysis yields and emulsification, respectively. Other sources of mannan, such as that from yeast, have been studied extensively and chemically carboxymethylated, phosphorylated, and sulfated.⁷¹

3. KEY PHYSICOCHEMICAL PROPERTIES OF WOOD HEMICELLULOSES AND THEIR DERIVATIVES

3.1. Solubility of Extracted and Derivatized Hemicelluloses. Polysaccharide solubility governs their technological properties as emulsifiers and stabilizers, and potentially other functionalities resulting from colloidal interactions. Polysaccharide dissolution is a complex balance of chainchain interactions (inter/intramolecular) and chain-solvent interactions. Intrinsic polysaccharide characteristics such as molar mass, degree of side-chain substitution, type of substituents, and solvent conditions such as pH, ionic strength, and temperature alter these interactions.⁷² It is especially relevant in food items, where the system is composed of multiple ingredients that may influence or be influenced by the inclusion of wood hemicelluloses. We review here the key items relevant to wood hemicellulose solubility and include contributions not only relevant to the food field, to give a comprehensive picture of their solubility.

Inter- and intrachain bonding, as well as side-chain substitutions, provide both physical and chemical properties that can be controlled for dissolution. Branched polysaccharides are generally easier to dissolve in water, such as the case for water-soluble arabinogalactan;⁷³ however, other types of substitution also affect the solubility. Wood GGMs' solubility is affected by acetyl and Gal substituent units. Xu et al.⁷⁴ showed

that deacetylated GGMs formed macroscopic aggregates, while Parikka et al.⁷⁵ observed how enzymatic oxidation of Gal units in GGMs enhanced intramolecular interactions via hemiacetal bonds. Mannans are often partially acetylated and are highly soluble in water as compared to alkaline extracted xylans, where the labile acetyl groups undergo deacetylation, which reduces their water solubility.⁷⁶ To preserve the native acetyl groups, extraction can be performed with nonaqueous solvents, such as DMSO.⁷⁷ However, using DMSO as the extraction solvent may be problematic when considering food-related applications due to potential residual solvent.

The presence of lignin and lignin-derived compounds, which are usually coextracted with hemicelluloses to a varying degree during hot-water extraction, is reported to modify GGM solubility. Bhattarai et al.⁷⁸ showed that crude GGM extracts obtained via PHWE method were less soluble in water than the same crude extracts purified via ethanol-precipitation. It was shown that aqueous suspensions of crude GGM extracts contained soluble hemicelluloses of 10 kDa molar mass and insoluble submicrometer fractions. Using cryo-transmission electron microscope imaging and small-angle X-ray scattering analysis, it was shown that some crude GGM extracts were partly consolidated to nano- and submicrometer structured objects resembling lignin nanoparticles.

Studies by Kishani et al.⁷⁹ suggest that water is a poor dispersion medium for native grades of GGM. The research referred to GGMs recovered from different extraction approaches, namely high pressure heating (100 °C, 5 min), hot-water extraction (60 °C, 3 h) of wood chips, and effluent from a thermomechanical pulp (TMP) plant. Regardless of the extraction and postextraction process, such as the removal of extractives and lignin impurities, all extracts displayed poor to very poor solubility in water, rendering aggregates up to 1000 nm. The molar masses of the GGMs obtained using these extraction procedures are seemingly higher than those from the PHWE process (21-66 kDa vs 10 kDa), which can render solubility differences. However, differences in the chemical structure of hemicelluloses from different recovery procedures also play a crucial role in multifactorial aspects affecting wood hemicelluloses' solubility.

As some native hemicelluloses are sparingly soluble in water, solvents such as aqueous alkaline solutions, DMSO,80 and cosolvent systems^{77⁻} are used to dissolve or solubilize hemicelluloses. Additionally, chemical modification of hemicelluloses, for example, esterification, oxidation, and etherification, have been used to add functional groups, while modification of the backbone structure has also been employed to improve the hemicellulose–water interaction.^{29,81} The degree of substitution was observed to play a key role in the solubilization of carboxymethylated GGM as the hydrodynamic volume increased as a function of degree of substitution.⁷⁰ Carboxymethylation and dihydroxypropylation improved the water solubility of xylan, demonstrated by the reduced turbidity of the solutions as compared to unmodified xylans.⁸² The solubility of xylans is similarly affected by their side groups and acetylation.^{3,83} Kishani et al.⁷⁹ reported the hydrodynamic radii of AGX to be <10 nm at a 0.1 wt % concentration, increasing to >60 nm at a 1.5 wt % concentration. For etherified xylans, aggregation at low degrees of substitution has also been observed, while an increased degree of modification resulted in reduced aggregation.⁸⁴ For oxidized hemicelluloses, the aggregation has been reported to be correlated to concentration and inversely correlated to the degree of modification.^{29,81}

3.2. Degree of Polymerization (DP). Degradation during the liberation processes defines the molar mass of the extracted products. The wood hemicellulose extracts commonly possess a heterogeneous molar mass and exist as mixtures of polymers and oligomers. The length of a linear polymer chain often has a tremendous effect on potential end use applications. Short chains are useful when low viscosity, fast solubilization (or even depolymerization) during use is sought, or when aiming at high inclusion levels as, for example, in high fiber containing products. On the other hand, longer chains are a requisite for applications where, for example, high viscosity or emulsification properties are desired as well as engineering applications, where entangled networks form stronger materials.⁸⁵ Use in packaging materials requires film formation⁸⁶ and barrier-relevant properties⁸⁷ that small molecules often lack. Consequently, hemicelluloses incorporated in packaging films⁸⁸⁻⁹¹ utilize longchain polysaccharide fractions or include the hemicelluloses in a matrix of higher molar mass polymers.⁸⁴ Because of the aforementioned aspects, it is of key importance to have a precise DP determination.

Hemicellulose insolubility in water complicates molar mass analysis because, due to clustering, the molar mass distribution assigned to the single polysaccharides remains unelucidated. Therefore, gel permeation chromatography, also known as sizeexclusion chromatography, combined with universal calibration or multiangle light scattering techniques are commonly used for the molar mass determination. This size-based fractionation method uses porous columns to separate the analytes on the basis of hydrodynamic volume. Solubility is therefore crucial to characterize the individual polysaccharide chains and to avoid erroneous estimation of molar mass values. Hence, solubility is likely to be one of the prominent reasons for the discrepancies in molar mass values reported in the literature. Insolubility also reduces the column's lifetime as well as increases the risk of capillary blockage. The latter issue has been addressed to a large extent by an alternative method named asymmetric-flow fieldflow fractionation (AF4), which does not rely on a stationary column,⁷⁸ but where a disposable ultrafiltration membrane is used for separation, eliminating the risk of column blockage. AF4 has been applied in the analysis of molar mass and characterization of aggregates from spruce GGMs.⁷⁸ The residual lignin in hemicelluloses is also thought to interfere with molar mass analysis as the autofluorescence of lignin interferes with the scattered laser light, hampering the analysis and thus leading to an overestimation of molar mass values. The fluorescence interference can be partly reduced by applying narrow bandwidth filters that are transparent only for the applied wavelength for light scattering as discussed previously.⁹

When the molar mass of the hemicelluloses has been determined, it is possible to obtain the DP value by dividing the average molar mass value by the molar mass of the anhydrous monosaccharide unit. Typical denominator values are 162 and 132 for neutral hexoses and pentoses, respectively, although the value can be different when the hemicelluloses' backbone involves charged or oxidized monosaccharides or contains noncarbohydrate substituents. Using number-average molar mass will subsequently yield number-average DP, and, in the same way, weight-average molar mass will yield weight-average DP.⁹²

A range of molar masses from 5 to 85 kDa and 4 to 67 kDa are reported for polysaccharide fractions of mannans and xylans, respectively (Table 1). Deliberate extraction that leads to larger molecules has been achieved by incorporating microwave

Table 1. Molar Mass (Weight-Average) of Liberated Wood
Hemicellulose Polysaccharides Reported in the Literature

mannans	$M_{\rm w}~({\rm kDa})$	refs	
hot-water extraction of wood	5-85	27,28,97	
hot-water extraction from TM	48-53	27,28	
alkaline treatment		56	98
xylans	$M_{\rm w}~({\rm kDa})$	refs	
hot-water extraction	4-8	27,99	
alkaline extraction	8-67	26,29,30,100,101	

irradiation of raw wood material in aqueous alkaline solution, instead of traditional heating.²⁶ The atmospheric pressure and relatively mild conditions (temperature <100 °C and pH 5) led to materials generated with a molar mass up to 67 kDa from spruce and demonstrated less degradation than the hydrothermal treatment. The formation of oligosaccharides (3-10 residues)⁹⁴ is often a consequence of degradation in the process, for example, by oxidation, or due to autohydrolytic effects caused by the release of acetic acid.^{43,95} For alkaline pulping processes, the degradation mechanisms of wood carbohydrates have been characterized by hydrolysis, oxidative peeling, and endwise peeling (Figure 3). In alkaline media, the reducing xylose group is removed by a β -elimination, which leads to a reducing galacturonic acid end group.96 The reactivity of carbohydrates in the prevalent kraft pulping process is steered by the morphology, crystallinity, and degree of polymerization. Within carbohydrates in wood biomass, cellulose is of a higher degree of polymerization and is more resistant toward alkaline media, and hence degrades to a lesser extent than hemicelluloses. Furthermore, alkaline conditions at elevated temperature, such as in cooking of pulp, will induce alkaline deesterification resulting in deacetylated hemicelluloses. Degradation of hemicelluloses may also take place in post processing treatments such as chemical modification. Oxidation with sodium periodate and subsequent reduction has been reported to lead to degradation that is manifested in a lowered molar mass.⁸¹

3.3. Assembly at Solid-Liquid Interfaces. Understanding how polysaccharides and their clusters assemble at interfaces is central to unlocking potential uses in food and feed. A solidliquid interface is present, for example, when the polysaccharides of interest interact with other macromolecule components in food or the mucosal layer in the gastrointestinal tract. In these events, adsorption or lack of adsorption (depletion) may take place. At the solid-liquid interface, an adsorption occurs for two main reasons: the adsorbate has a strong affinity toward a surface and/or a poor interaction with the solvent. This means that the structure and chemistry of the adsorbate along with the surface morphology and adsorption conditions (e.g., adsorbate concentration, pH value, and salt concentration) govern the overall adsorption. Surface-sensitive techniques with nanoscale resolution such as quartz crystal microbalance with dissipation monitoring and surface plasmon resonance are typically employed for investigating solid-liquid adsorption, and both techniques have been successfully implemented in solid-liquid adsorption studies involving hemicelluloses. 102-104

Hemicelluloses and their derivatives have been adsorbed on biopolymer surfaces in studies that mainly use cellulose as the substrate biopolymer. The focus of this Review is on food ingredients and wood hemicelluloses, but examples of other uses and nonwood hemicelluloses are added here to enrich the description. Molar mass, conformation, substituting units, and

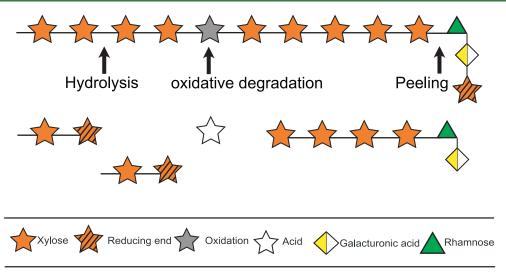


Figure 3. Mechanism of wood polysaccharide (shown for xylan) degradation during pulping. Adapted with permission from Sixta et al.¹⁵ Copyright 2006 Wiley.

degree of substitution of hemicelluloses are factors that determine the adsorption. The influence of the molar mass of XG,^{105,106} xylans,¹⁰⁷ and oxidized xylans²⁹ has been studied; it was found that a higher molar mass increased the adsorption of hemicelluloses on the cellulose surface, and this was partly assigned to their tendency to aggregate. However, substituting units attached to the hydroxyl moieties of the polysaccharide backbone may also influence the adsorption behavior, both for native and for chemically modified hemicelluloses. In the case of AX, a low amount of arabinose substitutions decreased the water solubility as unsubstituted parts tended to interact with other unsubstituted components and form aggregates, benefiting liquid-solid adsorption.^{102,108} However, a high degree of arabinose substitution could hinder adsorption because AX is better solvated and hence remains in the solution instead of adsorbing to the solid interface. Acetyl groups increased the steric hindrance on xylan and consequently decreased the selfassociation of xylans, hampering its adsorption on the cellulose surface.^{104,107,109} Fucose units on XG side chains were found to influence the interaction with cellulose positively; however, the molar mass and conformation of XG were found to be more important.^{110,111} On the contrary, in the case of GGM, the amount of Gal units had a greater impact on the interaction with cellulose rather than the molar mass.¹¹²

When charges are involved in the adsorption, surface charge, polymer charge, and ionic strength become decisive adsorption parameters,¹¹³ which is relevant in our discussion as several wood hemicelluloses contain GlcA residues. Lee et al.¹¹⁴ reported on the adsorption of cationic-modified xylan on model cellulose surfaces. They found that low charge-density xylan was adsorbed as a thicker and more viscoelastic layer as compared to its high charge-density counterpart. Additionally, the ionic strength of the solvent environment played a role in the adsorption behavior, where a high salt concentration increased the adsorbed mass of high charge-density xylan due to screening of the charges, resulting in a thicker adsorbed layer. Moreover, the charge of the solid substrate also participates in the adsorption process. Uncharged, oxidized xylans were reported to be adsorbed less on negatively charged cellulose as compared to an uncharged cellulose substrate, indicating an influence of electrostatics despite only the substrate being charged.²⁹

Surface morphological characteristics, such as surface area and porosity, can also influence adsorption. This aspect is especially relevant in food processing steps that involve membranes, as the adsorption of the hemicelluloses on the surface may lead to membrane fouling.¹¹⁵ In general, a large accessible surface area has a positive effect on adsorption; however, adsorption on internal surfaces, for example, in porous media, is also affected by the size of the pores, as the adsorbate may only adsorb at the internal surface if it can penetrate the pores.¹⁰⁷ Benselfelt et al.¹¹⁶ studied the effect of cellulose crystallinity on XG adsorption. XG did not only bind on the surface but penetrated the cellulose film. Therefore, the morphology of the substrate, for example, that of the food stuff or surfaces in the gastrointestinal tract, may affect the adsorption of wood-derived hemicelluloses.

3.4. Assembly at Liquid–Liquid Interfaces. Emulsions constitute an integral part of the food industry in the form of beverages and oil-containing foods. Aside from their adsorption at the solid-liquid interface, wood hemicelluloses are known to adsorb at the liquid-liquid interface, leading to the physical stabilization of liquid and semisolid emulsions. The emulsion stabilizing capability has been investigated for hemicelluloses obtained from different extraction methods and puri-ties, ^{27,117,118} as well as chemically modified derivatives. ^{70,119–121} As wood hemicelluloses are known to have low viscosity both in solution and as emulsions,^{74,122} the emulsion stabilizing capability has been attributed to interfacial activity (i.e., their ability to lower the interfacial tension and to assemble at the oilwater interface).^{122,123} Understanding this assembly and adsorption at the oil-water interface would allow for the generation of emulsions with tailor-made stabilities for various applications, utilizing the many different types of wood hemicelluloses.

There are three possible models of hemicellulose adsorption at the oil–water interface, an overview of which is illustrated in Figure 4. The first model follows the classical surfactant model, where the hemicellulose molecule is divided into a hydrophilic head and a hydrophobic tail. The hydrophobic tail anchors the entire molecule to the oil surface, while the hydrophilic head faces the aqueous phase.¹²⁴ The hydrophobic moiety can be composed of residual lipids, proteins, or other hydrophobic materials.¹²⁵ For example, it is well established that the protein-

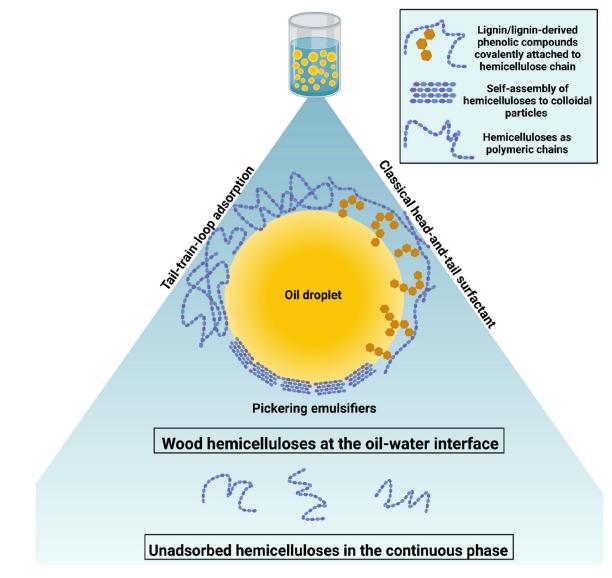


Figure 4. Adsorption modes of wood hemicelluloses at the liquid—liquid interface. Yellow spheres indicate oil droplets in an emulsion, while the blue background indicates the aqueous phase. Blue hexagons represent carbohydrate units, while brown hexagons represent lignin residues. The illustration also indicates that there may be a fraction of the hemicelluloses that are unadsorbed and are simply dispersed in the continuous phase. Created with BioRender.com.

rich fractions in gum arabic are responsible for its interfacial activity,^{126,127} which consist of proteoglycans and lipids associated with the protein.¹²⁸ For wood hemicelluloses, this model is particularly prominent in describing the interfacial activity of hemicelluloses that contain lignin-carbohydrate complexes (LCC). The covalent bond between lignin and hemicelluloses indicates that the lignin moiety could act as a hydrophobic anchor to the oil phase, while the aqueous phasefacing hemicellulose moiety could impart steric stability.^{129,130} Evidence of surfactant-like anchoring was provided by partitioning of adsorbed and unadsorbed hemicelluloses from model emulsions, followed by semiquantitative 2D nuclear magnetic resonance spectroscopy studies of the fractions.¹³¹ LCC species were found to be present in both fractions, with benzyl ether lignin-carbohydrate bonds (present in unpurified and nonpolar fractions of both PHWE GGM and GX) found exclusively bound to the oil-water interface. This finding explains the increased stability of emulsions stabilized by

unpurified PWHE hemicelluloses as compared to the partially purified extract.¹³⁰

The second model of adsorption details how a polymer chain adsorbs following the tail-train-loop conformation at the oil– water interface, such as in methylcellulose (MC). MC is a cellulose derivative in which some of the hydroxyl moieties are methyl-etherified, altering its inter- and intramolecular interactions as well as imparting interfacial activity. Following this model, the hemicelluloses would be adsorbed as random coils with loops that associate preferentially between the two phases.¹³² There have been previous studies that modified wood hemicelluloses into amphiphilic polymers by grafting hydrophobic moieties, both through esterification^{120,133,134} and through etherification^{119,135} of the hydroxyl groups, with the aim of altering the interfacial activity of the hemicelluloses in a similar manner by which cellulose is modified into MC. The results of those studies exhibited an increased surface activity of the hemicelluloses and evidenced consequent self-assembly. Naturally, wood hemicelluloses are acetylated, which contributed to reducing the extent of hydrogen bonding, subsequently decreasing their tendency to aggregate, and thus increasing water solubility;¹³⁶ as such, it is plausible to assume that they might adsorb at the oil—water interface in a way similar to MC. However, there has been limited investigation into the correlation between the degree of acetylation of a given hemicellulose and the interfacial activity.

The third mode of adsorption is commonly known as Pickering stabilization, which requires solid particles or clusters of particles termed Pickering emulsifiers. Pickering emulsifiers adsorb at the oil-water interface, not as dissolved molecules, but as solid particles. The particles are considered to be adsorbed irreversibly at the oil-water interface due to the high free energy barrier associated with their desorption.¹³⁷ There have been many explorations into the use of polysaccharide-based solid particles as Pickering emulsifiers.^{138,139} It is possible to prepare nanocrystals with xylan extracted from birchwood kraft pulp¹⁴⁰ as well as nanoparticles from xylan and xylan derivates.¹⁴¹⁻¹⁴³ Xylan nanocrystals from sugar cane bagasse have been successfully utilized as Pickering emulsifiers,¹⁴⁴ which might indicate that wood-derived xylan nanocrystals have the potential to be used similarly. Additionally, insoluble clusters of alkaliextracted xylans from birch and beech also demonstrated efficient stabilization properties.47 Nevertheless, it is worth noting that, in a hemicellulose-stabilized emulsion, there is a possibility that nonpolysaccharide solid particles are also responsible for the stabilization. As mentioned previously, spruce GGM extracts containing lignin particles¹⁴⁵ contributed to emulsion stabilization. 138,146

In addition to their mode of adsorption, another interesting question to address regarding wood hemicelluloses' adsorption at the liquid-liquid interface is the fate of the aggregates (in which they typically exist in aqueous systems), as the hemicelluloses are adsorbed at the oil-water interface. Despite documented evidence that wood hemicelluloses form aggregates in aqueous solutions/dispersions,^{78,147} there has only been limited information on whether the same form of assembly exists at the interface, especially those that are not adsorbed as Pickering emulsifiers. For example, previous studies of PHWE hemicellulose-stabilized emulsions indicated that both the hemicellulose and the phenolic fractions are adsorbed at the oil-water interface and contributed to both physical and chemical stability of the emulsion, 130,145 and that the benzyl ether-type LCC is preferentially adsorbed at the interface as compared to phenyl glycosides and gamma esters.¹³¹ While those are valuable insights, the available data did not offer any indications regarding their assembly at the interface, as the adsorbed species had to be extracted prior to analysis and their conformation may potentially have been compromised. Considering their association with lignin, inspiration could be drawn from the investigation into the adsorption of gum arabic, which is known to be associated with protein species and form assemblies in its aqueous solution.^{148,149} Neutron scattering investigations of gum arabic-stabilized emulsions indicated that aggregates, present in its aqueous solution, dissociate at the oilwater interface to form an interfacial network that contributed to emulsion stability.^{150,151} Hence, it may be intriguing in the future to apply direct characterization methods for emulsion interfaces, such as neutron scattering¹⁵² and atomic force microscopy.¹⁵³

4. WOOD HEMICELLULOSES AS FOOD INGREDIENTS

4.1. Gastrointestinal Behavior of Hemicelluloses. Complex dietary glycans can affect the gastrointestinal tract in a number of ways, including influencing the microbiota composition. The gut microbiome is well-known to affect host well-being, with the preponderance of several bacterial species either positively or negatively correlated to anorexia,¹⁵⁴ obesity and diabetes,¹⁵⁵ and colorectal cancer,¹⁵⁶ among others. The structural compositions of wood-derived xylans and mannans resemble other complex dietary fibers found in a variety of foods that stimulate beneficial bacteria and are therefore likely to be good candidates as alternatives to dietary fibers used today.

One prominent role of xylans and mannans as dietary fibers lies in their fermentability by the gut microbiota. More detailed knowledge about individual microbes' potential to degrade specific complex carbohydrates present in food stuffs has recently emerged. Several detailed studies on complex degradation machineries in bacterial representatives from Bifidobacterium, Firmicutes, and Bacteroidetes¹⁰ have been reported for xylans^{157,158} and mannans.^{159–162} This knowledge further enables rationally modifying the fibers to be preferentially digested by the microbe of interest, also known as microbiota-directed fiber (MDF). The use of MDF in food and feed has been investigated in several in vivo studies with positive results; specifically, xylans and mannans have been shown to stimulate growth of probiotic targets like Bifidobacterium, 163,164 Roseburia intestinalis,¹⁵⁷ and Faecalibacterium prausnitzii.^{156,164,165} However, understanding the mechanisms of how different bacteria interact and affect their surroundings is still limited. For example, the observed effects might have arisen either directly due to the bacteria targeted by MDF or indirectly, from the proliferation of other microbial species that benefit from the MDF-targeted bacteria. This secondary "butterfly effect" may, in some cases, be more important than the direct effect of the MDF itself.¹⁶² As such, there are still challenges in demonstrating the beneficial effect toward the host in using xylans and mannans as MDFs. Fortunately, no adverse effects have yet been observed in the in vivo studies, and, as mentioned above, both wood-based hemicelluloses promoted the growth of probiotic target bacteria.^{160,162,164}

4.2. Food Emulsions. Emulsions are indispensable in modern food product formulations, and a rising consumer demand for food products with all-natural, minimally processed ingredients has increased the global demand of functional polysaccharides from natural sources to act as emulsion stabilizers. Because of their interfacial activity, wood hemicelluloses have attracted interest as possible food emulsifiers and stabilizers. Spruce GGMs have exhibited both abilities in oil-in-water emulsion systems in many studies.^{117,122} Mikkonen et al.¹²³ showed that GGMs promoted the long-term stability of emulsions more effectively than both gum arabic and corn fiber gum, which are commonly used hydrocolloids in commercial food formulations. It was also noted that the emulsion stabilization mechanism of cereal and seed-based hemicelluloses differs from that of wood-based hemicelluloses. Aqueous GGM and GX solutions exhibit a much lower viscosity and do not significantly modify the rheological properties of emulsions' continuous phases at concentrations below 8 wt %;¹²² however, as elaborated on in the previous section, recent studies have proven the central role of residual lignin in their emulsifying capability. In bamboo xylans, coextracted proteins together with lignin contributed to the surface-active property and formed

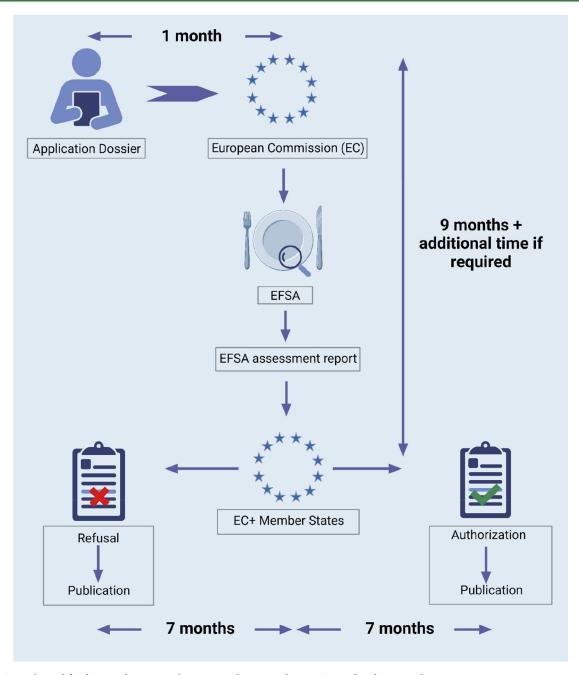


Figure 5. General novel food premarket approval summarized route in the EU. Created with BioRender.com.

stable soy oil-in-water emulsions.¹⁶⁶ Hemicellulose–protein complexes have also been formed to improve their emulsification abilities; for example, soyhull hemicelluloses conjugated with soy protein via the Maillard reaction have exhibited improved emulsifying properties.¹⁶⁷

However, the stability of an emulsion is not only limited to physical stability against phase separation, but also the chemical stability of the oil-soluble compounds loaded in the oil phase. By the virtue of the lignin and lignin-derived phenolic groups, spruce GGM extracts acquire antioxidant capabilities and protect the dispersed oil phase against lipid oxidation.^{130,168} GGM was used to formulate a functional, drinkable yogurt with cod liver oil¹⁶⁹ within this concept. Hemicelluloses have been used as stand-alone stabilizers or in conjunction with other polysaccharides such as gum arabic,¹⁷⁰ providing a method to protect fragile bioactive components in various food matrices. The presence of lignin and other impurities, however, might become a hindrance with respect to consumer acceptance of hemicelluloses. Pure hemicelluloses have a neutral flavor profile (similar to gum arabic); however, sensory analysis of yogurt supplemented by emulsions made with wood hemicelluloses indicated that a higher lignin content correlates to a more pronounced wood flavor.¹⁷¹ While no consumer acceptance test has been conducted thus far, the woody flavor might not necessarily be unwanted; it is known that flavors derived from wood are desirable in food items aged in wooden barrels such as alcoholic beverages¹⁷² and wine vinegar.¹⁷³ Therefore, instead of a hindrance, it might open new avenues for woody flavors in foods.

4.3. Animal Feed. As established in the previous section, hemicelluloses are indigestible polysaccharides fermented by the gut microbiota. Spruce GGMs and birch GXs have exhibited this

potential during in vitro^{164,174} and in vivo experiments in animal models, such as sheep,¹⁷⁵ pigs,¹⁶² and mice.^{176,177} It is therefore implied that wood hemicelluloses could be applicable for use in animal feeds as well. The earliest known commercial product from wood hemicelluloses used in animal feed was Masonex, a hardwood hemicellulose extract produced by mild hydrolysis of wooden hardboard. It was included as a binding agent for pellet feeds. Masonex was found to be indigestible by broiler chickens¹⁷⁸ and protected protein feed from degradation in the rumen.¹⁷⁹ Over the years, more details on the effects of adding wood hemicelluloses to animal feed have been discovered. For example, mice fed softwood hemicellulose supplementation experienced a lower level of cholesterol, free fatty acid, and bile acids in their serum lipid composition as compared to the control group fed without supplementation, implying a possible cardiac protective property.¹⁷⁷ In addition, spruce GGM may impart beneficial effects toward chronic prostate inflammation due to observed changes in the gut microbiota composition.¹⁸⁰ Ruminants may also benefit from the addition of hemicelluloses in their feed, where it has been documented to enhance cellulose digestibility,^{181,182} improve lactation in dairy cows,¹⁸¹ and modify methanogenesis, which can potentially reduce methane emissions.¹⁸³ While there are still knowledge gaps to be filled, such potential should be further explored.

4.4. Regulatory Items. Before entering commercial use as food ingredients, wood hemicelluloses must go through the premarket approval from food regulatory bodies specific to geographical locations where they will be marketed and distributed. The process of gaining premarket approval varies from one country or region to another. For example, in the United States (U.S.), food manufacturers may follow a process that relies on assessment independent of the federal agency, Food and Drug Administration (FDA), while the European Union (EU) follows a process regulated by the EC, facilitated by the European Food Safety Agency (EFSA), that provides independent scientific opinions on the safety of food. In this section, we will cover existing regulatory frameworks in the EU and the U.S.

Any food that was not consumed to a significant degree prior to May 1997 within the EU falls under the category "novel foods" pursuant to Regulation (EU) 2015/2283.²⁴ It also includes food from new sources, new substances used in food, and new ways and technologies for producing food. Application for the approval of such a substance or food requires the submission of a dossier on the substance, which must include its identity and nutritional information. Required information in the dossier includes compositional data including stability; production process; toxicological data including genotoxicity, subchronic, chronic, reproductive, and developmental toxicities; absorption, distribution, metabolism, and excretion (ADME); proposed uses; usage levels; and anticipated intake including precautions and restrictions.¹⁸⁴ After a complete submission of the information, a detailed risk assessment is performed by the EFSA as per EC mandate, which will take around 9 months provided no additional data are required from the applicant. When the risk assessment is adopted, the application reaches the postadoption phase where EFSA publishes the scientific output and EC drafts an implementation act. The complete process of authorization of a novel food substance, which requires a risk assessment, can take up to two years¹⁸⁵ from the submission of the application (Figure 5).

While the molecular structure and the technological functions of wood hemicelluloses are known, few studies have focused on nutritional aspects and toxic contaminants, or side products formed during processing. Moreover, chemical and microbiological characterization of the intended substance pertaining to shelf life should be provided or addressed, and, when the novel food is expected to be used as an ingredient added in food products, assessment of its stability in the intended food matrices must be performed.¹⁸⁶ Specific safety considerations with regards to wood hemicelluloses involve, for example, the following items: furans, such as furfural, 5-hydroxymethylfurfural, and furfural diethylacetal, can be formed from pentose or hexose sugars during thermal treatment and oligophenolic compounds from pentose self-condensation.^{11,49,187} The formation of acids, for example, cinnamic acid, abietic acid, and levulinic acid, has also been reported.¹¹ While it is possible to reduce the amount of these small molecule contaminants down to trace amounts, it is nevertheless important to consider their safety limits. Additionally, the presence of other minor contaminants and microbial contamination over time are also relevant issues to assess.

The anticipated daily intake of the wood hemicelluloses would vary depending on the intended technical and nutritional function in specific foods. As emulsion stabilizers, the daily intake was estimated to be 32 mg/kg bodyweight/day, which amounts to 2.2 g per person per day, assuming a 70 kg bodyweight. This value is based on the average food consumption of the Finnish population, assuming GGM as the sole emulsifier in dressings, beverages, and margarines;⁴⁹ however, the value may be different from country to country, with regards to the consumption rate of emulsion-based food products. It is less than the intake value of modified and unmodified cellulose (which can have wood origin), which EFSA considers safe up to an intake of 660-900 mg/kg bodyweight/day.²⁵ Alternatively, wood hemicelluloses can be further explored as a source of dietary fibers, where a higher level of intake applies. Under EFSA NDA 2010,¹⁸⁸ dietary fiber is defined as "polysaccharides and lignin that are not digested in the human small intestine". Dietary fibers are resistant to hydrolysis and absorption in the small intestine and enter the colon substantially unmodified, where they are subjected to fermentation by the colonic microbiota, and, as such, if deemed to demonstrate a positive effect, may be used as a prebiotic; encouragingly, wood hemicelluloses have recently demonstrated such properties, as elaborated on in section 4.1. The present intake of dietary fibers is below the recommended levels at a general population scale (on average 18-24 g/day for adult males and 16-20 g/day for adult females in Europe),¹⁸⁹ which is lower than EFSA's recommendation of 25 g/day of dietary fibers to be adequate for normal laxation in adults.¹¹

In the U.S., the FDA does not, as yet, recognize novel foods in a separate category. On the basis of the technological aspects described in the previous sections, wood hemicelluloses would be classified as food additives and regulated through the food additive approval process. However, any new food or ingredient can obtain a "generally recognized as safe" (GRAS) designation if a panel of recognized experts affirm that pivotal studies related to the risk assessment of the food under the conditions of its intended use are published in peer-reviewed scientific journals or if the food/ingredient in question was commonly used before 1958. As such, an ingredient is referred to as "self-affirmed GRAS" and voluntarily notified to the FDA.^{190,191} An alternative approach is to petition the FDA to classify the ingredient as a food additive.¹⁹² In both cases, FDA requires a dossier of risk assessment data similar to that in EU. The key difference between the GRAS and food additive petition is that, in the latter case, data and information on the use of the substance are held privately and sent to the FDA, who establishes the safety of the substance under the intended conditions, in contrast to the GRAS process, where the safety determination is made by qualified experts outside the federal government. Therefore, most companies opt to list new food ingredients as GRAS, when possible, for a rapid premarket approval process.

5. OUTLOOK

The growing focus on a circular economy and an increasing need for new food solutions have increased interest in the more efficient usage of wood biomass and application of wood processing side-streams in food instead of disposing them. While forests can be harvested less frequently than crops, they can be harvested all year around and are thus not dependent on seasonal harvesting. Forest biomass is currently used for the production of cellulose and lignin, which leads to low yield and wastage of the prominent hemicellulose fraction. Implementing a wider usage of wood hemicelluloses would provide incentives for the development of new harvesting strategies that could both optimize the use of biomass and preserve the role of forests in the ecosystem. This is highlighted by the fact that there is a vast amount of available wood-based feedstocks that are inadequate for timber/construction material that is still rich in hemicelluloses. Additionally, this volume may rise further due to the spread of wood degrading insects due to the changing climate.¹⁹³ Thus, exploiting wood hemicellulose as food could improve the utilization of wood biomass and contribute to feeding a population that has just passed 8 billion.

We have exhibited in this Review that the current state-of-theart technological aspects that wood hemicelluloses possess are useful for food applications. Using various extraction methods and postextraction treatments, it is possible to produce hemicelluloses from wood with tailored functionalities for a vast array of food items, ranging from emulsion stabilizers, sources of dietary fiber and probiotics, and even food packaging material. There have also been successful efforts to scale up the extraction process, 194,195 proving the industrial feasibility of producing wood hemicelluloses. Nevertheless, we acknowledge that there are still several gaps in the knowledge that need to be addressed, such as their behavior in the human digestion process, toxicology, and consumer acceptance. However, vigorous research activities are being conducted to fill those gaps as well as discovering possible new functionalities. Hence, we believe it will not be long before wood hemicelluloses find a way onto kitchen tables around the world.

AUTHOR INFORMATION

Corresponding Authors

Bjørge Westereng – Faculty of Chemistry, Biotechnology and Food Science, Norwegian University of Life Sciences, Ås 1430, Norway; orcid.org/0000-0002-5141-7231; Email: bjorge.westereng@nmbu.no

- Kirsi S. Mikkonen Department of Food and Nutrition and Helsinki Institute of Sustainability Science (HELSUS), University of Helsinki, Helsinki 00014, Finland; Octid.org/ 0000-0003-4490-5912; Email: kirsi.s.mikkonen@helsinki.fi
- **Tiina Nypelö** Department of Chemistry and Chemical Engineering, Chalmers University of Technology, Gothenburg 41296, Sweden; Wallenberg Wood Science Center, Chalmers

University of Technology, Gothenburg 41296, Sweden; Department of Bioproducts and Biosystems, Aalto University, Espoo 00760, Finland; orcid.org/0000-0003-0158-467X; Email: tiina.nypelo@chalmers.se

Authors

- Felix Abik Department of Food and Nutrition, University of Helsinki, Helsinki 00014, Finland
- **Chonnipa Palasingh** Department of Chemistry and Chemical Engineering, Chalmers University of Technology, Gothenburg 41296, Sweden
- Mamata Bhattarai Department of Food and Nutrition, University of Helsinki, Helsinki 00014, Finland; Department of Bioproducts and Biosystems, Aalto University, Espoo 00076, Finland
- Shaun Leivers Faculty of Chemistry, Biotechnology and Food Science, Norwegian University of Life Sciences, Ås 1430, Norway
- Anna Ström Department of Chemistry and Chemical Engineering, Chalmers University of Technology, Gothenburg 41296, Sweden; orcid.org/0000-0002-9743-1514

Complete contact information is available at: https://pubs.acs.org/10.1021/acs.jafc.2c06449

Funding

We thank the Nordic Council of Ministers for funding a joint Nordic project (SNS Hemisurf 127) in which framework this Review was drafted. T.N. and C.P. thank the Swedish Research Council grant (registration number 2017-05138), the Wallenberg Wood Science Center, and the Materials Science Area of Advance at Chalmers for funding. B.W. and S.L. appreciate funding from Norwegian research council grants 295501, 296083, and 244259. F.A. and K.S.M. acknowledge funding from the Academy of Finland 322514.

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

We thank Troy Faithfull for his help in proofreading the manuscript.

ABBREVIATIONS USED

Ara, arabinose; AX, arabinoxylan; AGX, arabinoglucuronoxylan; AF4, asymmetric-flow field flow fractionation; DMSO, dimethyl sulfoxide; EC, European Commission; EFSA, European Food Safety Authority; FDA, Food and Drug Administration; Gal, galactose; GalA, galacturonic acid; GalM, galactomannan; GGM, galactoglucomannan; GRAS, generally recognized as safe; GM, glucomannan; GlcA, glucuronic acid; GX, glucuronoxylan; LCC, lignin–carbohydrate complex; MC, methyl cellulose; MDF, microbiota-directed fiber; NDA, panel on nutrition, novel foods, and food allergens; PHWE, pressurized hot-water extraction; TMP, thermomechanical pulp; XG, xyloglucan

REFERENCES

(1) Asioli, D.; Aschemann-Witzel, J.; Caputo, V.; Vecchio, R.; Annunziata, A.; Næs, T.; Varela, P. Making sense of the "clean label" trends: A review of consumer food choice behavior and discussion of industry implications. *Food Research International* **2017**, *99*, 58–71.

(2) Valoppi, F.; Agustin, M.; Abik, F.; Morais de Carvalho, D.; Sithole, J.; Bhattarai, M.; Varis, J. J.; Arzami, A. N. A. B.; Pulkkinen, E.; Mikkonen, K. S. Insight on Current Advances in Food Science and

Technology for Feeding the World Population. *Frontiers in Sustainable Food Systems* **2021**, *5*, 1.

(3) Ébringerová, A.; Hromádková, Z.; Heinze, T. Hemicellulose. In *Polysaccharides I: Structure, Characterization and Use*, 1st ed.; Heinze, T., Ed.; Springer-Verlag: Berlin, Heidelberg, 2005; pp 1–67.

(4) Peng, P.; She, D. Isolation, structural characterization, and potential applications of hemicelluloses from bamboo: A review. *Carbohydr. Polym.* **2014**, *112*, 701–720.

(5) Barhoum, A.; Jeevanandam, J.; Rastogi, A.; Samyn, P.; Boluk, Y.; Dufresne, A.; Danquah, M. K.; Bechelany, M. Plant celluloses, hemicelluloses, lignins, and volatile oils for the synthesis of nanoparticles and nanostructured materials. *Nanoscale* **2020**, *12* (45), 22845–22890.

(6) Garrote, G.; Cruz, J. M.; Moure, A.; Domínguez, H.; Parajó, J. C. Antioxidant activity of byproducts from the hydrolytic processing of selected lignocellulosic materials. *Trends in Food Science & Technology* **2004**, *15* (3), 191–200.

(7) Herrera, S. Wood-based ethanol advances on international front...Cellulosic fuel from trees gets a closer look. *Industrial Biotechnology* **2006**, 2 (2), 101–107.

(8) Mano, M. C. R.; Neri-Numa, I. A.; da Silva, J. B.; Paulino, B. N.; Pessoa, M. G.; Pastore, G. M. Oligosaccharide biotechnology: an approach of prebiotic revolution on the industry. *Appl. Microbiol. Biotechnol.* **2018**, *102* (1), 17–37.

(9) Santibáñez, L.; Henríquez, C.; Corro-Tejeda, R.; Bernal, S.; Armijo, B.; Salazar, O. Xylooligosaccharides from lignocellulosic biomass: A comprehensive review. *Carbohydr. Polym.* **2021**, *251*, 117118.

(10) McKee, L. S.; La Rosa, S. L.; Westereng, B.; Eijsink, V. G.; Pope, P. B.; Larsbrink, J. Polysaccharide degradation by the Bacteroidetes: mechanisms and nomenclature. *Environmental Microbiology Reports* **2021**, *13* (5), 559–581.

(11) Rasmussen, H.; Sørensen, H. R.; Meyer, A. S. Formation of degradation compounds from lignocellulosic biomass in the biorefinery: sugar reaction mechanisms. *Carbohydr. Res.* **2014**, 385, 45–57.

(12) Asgher, M.; Qamar, S. A.; Bilal, M.; Iqbal, H. M. N. Bio-based active food packaging materials: Sustainable alternative to conventional petrochemical-based packaging materials. *Food Research International* **2020**, *137*, 109625.

(13) Bowyer, J. L.; Haygreen, J. G.; Shmulsky, R. Forest Products and Wood Science: An Introduction, 5th ed.; Blackwell Pub: Ames, IA, 2007.
(14) Sjöström, E. Wood Chemistry: Fundamentals and Applications, 2nd ed.; Academic Press: San Diego, CA, 1993.

(15) Sixta, H. *Handbook of Pulp*; WILEY-VCH Verlag GmbH: New York, 2006; Vol. 2.

(16) Ebringerová, A. Structural Diversity and Application Potential of Hemicelluloses. *Macromol. Symp.* **2005**, 232 (1), 1–12.

(17) Varki, A.; Cummings, R. D.; Aebi, M.; Packer, N. H.; Seeberger, P. H.; Esko, J. D.; Stanley, P.; Hart, G.; Darvill, A.; Kinoshita, T.; Prestegard, J. J.; Schnaar, R. L.; Freeze, H. H.; Marth, J. D.; Bertozzi, C. R.; Etzler, M. E.; Frank, M.; Vliegenthart, J. F.; Lütteke, T.; Perez, S.; Bolton, E.; Rudd, P.; Paulson, J.; Kanehisa, M.; Toukach, P.; Aoki-Kinoshita, K. F.; Dell, A.; Narimatsu, H.; York, W.; Taniguchi, N.; Kornfeld, S. Symbol Nomenclature for Graphical Representations of Glycans. *Glycobiology* **2015**, *25* (12), 1323–1324.

(18) Alokika; Anu; Kumar, A.; Kumar, V.; Singh, B. Cellulosic and hemicellulosic fractions of sugarcane bagasse: Potential, challenges and future perspective. *Int. J. Biol. Macromol.* **2021**, *169*, 564–582.

(19) Huang, L.-Z.; Ma, M.-G.; Ji, X.-X.; Choi, S.-E.; Si, C. Recent Developments and Applications of Hemicellulose From Wheat Straw: A Review. *Frontiers in Bioengineering and Biotechnology* **2021**, *9*, 1.

(20) Doner, L. W.; Hicks, K. B. Isolation of Hemicellulose from Corn Fiber by Alkaline Hydrogen Peroxide Extraction. *Cereal Chem.* **1997**, *74* (2), 176–181.

(21) Stoklosa, R. J.; Hodge, D. B. Extraction, Recovery, and Characterization of Hardwood and Grass Hemicelluloses for Integration into Biorefining Processes. *Ind. Eng. Chem. Res.* **2012**, *51* (34), 11045–11053.

(22) Prajapati, V. D.; Jani, G. K.; Moradiya, N. G.; Randeria, N. P.; Nagar, B. J.; Naikwadi, N. N.; Variya, B. C. Galactomannan: A versatile biodegradable seed polysaccharide. *Int. J. Biol. Macromol.* **2013**, *60*, 83– 92.

(23) Yang, D.; Yuan, Y.; Wang, L.; Wang, X.; Mu, R.; Pang, J.; Xiao, J.; Zheng, Y. A Review on Konjac Glucomannan Gels: Microstructure and Application. *International Journal of Molecular Sciences* **2017**, *18* (11), 2250.

(24) Regulation (EU) 2015/2283 of the European parliament and of the Council of 25 November 2015 on novel foods, amending Regulation (EU) no. 1169/2011 of the European Parliament and of the Council and repealing Regulation (EC) no. 258/97 of the European Parliament and of the Council and Commision Regulation (EC) Nu 1852/2001). Ed., 2015.

(25) Younes, M.; Aggett, P.; Aguilar, F.; Crebelli, R.; Di Domenico, A.; Dusemund, B.; Filipič, M.; Jose Frutos, M.; Galtier, P.; Gott, D.; Gundert-Remy, U.; Georg Kuhnle, G.; Lambré, C.; Leblanc, J.-C.; Lillegaard, I. T.; Moldeus, P.; Mortensen, A.; Oskarsson, A.; Stankovic, I.; Tobback, P.; Waalkens-Berendsen, I.; Wright, M.; Tard, A.; Tasiopoulou, S.; Woutersen, R. A. Re-evaluation of celluloses E 460(i), E 460(ii), E 461, E 462, E 463, E 464, E 465, E 466, E 468 and E 469 as food additives. *EFSA J.* **2018**, *16*, e05047.

(26) Chadni, M.; Bals, O.; Ziegler-Devin, I.; Brosse, N.; Grimi, N. Microwave-assisted extraction of high-molecular-weight hemicelluloses from spruce wood. *Comptes Rendus Chimie* **2019**, *22* (8), 574–584.

(27) Mikkonen, K. S.; Kirjoranta, S.; Xu, C.; Hemming, J.; Pranovich, A.; Bhattarai, M.; Peltonen, L.; Kilpeläinen, P.; Maina, N.; Tenkanen, M. Environmentally-compatible alkyd paints stabilized by wood hemicelluloses. *Ind. Crops Prod.* **2019**, *133*, 212–220.

(28) Kishani, S.; Vilaplana, F.; Xu, W.; Xu, C.; Wågberg, L. Solubility of Softwood Hemicelluloses. *Biomacromolecules* **2018**, *19* (4), 1245–1255.

(29) Palasingh, C.; Ström, A.; Amer, H.; Nypelö, T. Oxidized xylan additive for nanocellulose films – A swelling modifier. *Int. J. Biol. Macromol.* **2021**, *180*, 753–759.

(30) Bai, L.; Hu, H.; Xu, J. Influences of configuration and molecular weight of hemicelluloses on their paper-strengthening effects. *Carbohydr. Polym.* **2012**, *88* (4), 1258–1263.

(31) Perez, D. D. S.; Huber, P.; Janodet, A.; Soranzo, A.; Guillemain, A.; Schelcher, M. Impact of the extraction conditions and wood species on the chemical composition of hemicelluloses. *ATIP Assoc. Technol. L'Industrie Papet.* **2011**, *65*, 14–20.

(32) Ebringerová, A.; Heinze, T. Xylan and xylan derivatives – biopolymers with valuable properties, 1. Naturally occurring xylans structures, isolation procedures and properties. *Macromol. Rapid Commun.* **2000**, *21* (9), 542–556.

(33) Lahtinen, M. H.; Valoppi, F.; Juntti, V.; Heikkinen, S.; Kilpeläinen, P. O.; Maina, N. H.; Mikkonen, K. S. Lignin-Rich PHWE Hemicellulose Extracts Responsible for Extended Emulsion Stabilization. *Frontiers in Chemistry* **2019**, *7*, 1.

(34) Wu, C. The Potential of Pre-Hydrolysis Liquor from the Dissolving Pulp Process as Recovery Source of Xylooligosaccharide - A Mini-Review. *BioResources* **2016**, *11* (3), 11.

(35) Reyes, P.; Teixeira Mendonça, R.; Rodríguez, J.; Fardim, P.; Vega, B. Characterization of the Hemicellulosic Fraction Obtained After Pre-Hydrolysis of *Pinus radiata* Wood Chips With Hot-Water at Different Initial pH. *Journal of the Chilean Chemical Society* **2013**, *58*, 1614–1618.

(36) Garrote, G.; Domínguez, H.; Parajó, J. C. Mild autohydrolysis: an environmentally friendly technology for xylooligosaccharide production from wood. *J. Chem. Technol. Biotechnol.* **1999**, 74 (11), 1101–1109.

(37) Kabel, M. A.; Carvalheiro, F.; Garrote, G.; Avgerinos, E.; Koukios, E.; Parajó, J. C.; Gírio, F. M.; Schols, H. A.; Voragen, A. G. J. Hydrothermally treated xylan rich by-products yield different classes of xylo-oligosaccharides. *Carbohydr. Polym.* **2002**, *50* (1), 47–56.

(38) Garrote, G.; Kabel, M. A.; Schols, H. A.; Falqué, E.; Domínguez, H.; Parajó, J. c. Effects of Eucalyptus globulus Wood Autohydrolysis Conditions on the Reaction Products. J. Agric. Food Chem. 2007, 55 (22), 9006–9013.

(39) Nabarlatz, D.; Farriol, X.; Montané, D. Kinetic Modeling of the Autohydrolysis of Lignocellulosic Biomass for the Production of Hemicellulose-Derived Oligosaccharides. *Ind. Eng. Chem. Res.* **2004**, *43* (15), 4124–4131.

(40) Francillon, J.; Chirat, C.; Boisset, C.; Buon, L. Multi-step purification method of water-soluble oligosaccharides produced from hardwood and softwood. *Holzforschung* **2020**, *74* (6), 615–623.

(41) Xu, F.; Chen, J.; Yang, G.; Ji, X.; Wang, Q.; Liu, S.; Ni, Y. Combined Treatments Consisting of Calcium Hydroxide and Activate Carbon for Purification of Xylo-Oligosaccharides of Pre-Hydrolysis Liquor. *Polymers* **2019**, *11* (10), 1558.

(42) Cara, C.; Ruiz, E.; Carvalheiro, F.; Moura, P.; Ballesteros, I.; Castro, E.; Gírio, F. Production, purification and characterisation of oligosaccharides from olive tree pruning autohydrolysis. *Industrial Crops and Products* **2012**, *40*, 225–231.

(43) Michalak, L.; Knutsen, S. H.; Aarum, I.; Westereng, B. Effects of pH on steam explosion extraction of acetylated galactoglucomannan from Norway spruce. *Biotechnology for Biofuels* **2018**, *11* (1), 311.

(44) Remón, J.; Li, T.; Chuck, C. J.; Matharu, A. S.; Clark, J. H. Toward Renewable-Based, Food-Applicable Prebiotics from Biomass: A One-Step, Additive-Free, Microwave-Assisted Hydrothermal Process for the Production of High Purity Xylo-oligosaccharides from Beech Wood Hemicellulose. ACS Sustainable Chem. Eng. 2019, 7 (19), 16160–16172.

(45) Rigual, V.; Santos, T. M.; Domínguez, J. C.; Alonso, M. V.; Oliet, M.; Rodriguez, F. Evaluation of hardwood and softwood fractionation using autohydrolysis and ionic liquid microwave pretreatment. *Biomass and Bioenergy* **2018**, *117*, 190–197.

(46) Sarker, T. R.; Pattnaik, F.; Nanda, S.; Dalai, A. K.; Meda, V.; Naik, S. Hydrothermal pretreatment technologies for lignocellulosic biomass: A review of steam explosion and subcritical water hydrolysis. *Chemosphere* **2021**, *284*, 131372.

(47) Mikkonen, K. S. Strategies for structuring diverse emulsion systems by using wood lignocellulose-derived stabilizers. *Green Chem.* **2020**, 22 (4), 1019–1037.

(48) Qaseem, M. F.; Shaheen, H.; Wu, A.-M. Cell wall hemicellulose for sustainable industrial utilization. *Renewable and Sustainable Energy Reviews* **2021**, *144*, 110996.

(49) Pitkänen, L.; Heinonen, M.; Mikkonen, K. S. Safety considerations of plant polysaccharides for food use: a case study on phenolic-rich softwood galactoglucomannan extract. *Food & Function* **2018**, *9* (4), 1931–1943.

(50) Mortensen, A.; Aguilar, F.; Crebelli, R.; Di Domenico, A.; Dusemund, B.; Frutos, M. J.; Galtier, P.; Gott, D.; Gundert-Remy, U.; Lambré, C.; Leblanc, J.-C.; Lindtner, O.; Moldeus, P.; Mosesso, P.; Parent-Massin, D.; Oskarsson, A.; Stankovic, I.; Waalkens-Berendsen, I.; Wright, M.; Younes, M.; Tobback, P.; Horvath, Z.; Tasiopoulou, S.; Woutersen, R. A. Re-evaluation of oxidised starch (E 1404), monostarch phosphate (E 1410), distarch phosphate (E 1412), phosphated distarch phosphate (E 1413), acetylated distarch phosphate (E 1414), acetylated starch (E 1420), acetylated distarch adipate (E 1422), hydroxypropyl starch (E 1440), hydroxypropyl distarch phosphate (E 1442), starch sodium octenyl succinate (E 1450), acetylated oxidised starch (E 1451) and starch aluminium octenyl succinate (E 1452) as food additives. *EFSA Journal* 2017, *15*, e04911.

(51) Gani, A.; Ashwar, B. A. Food Biopolymers: Structural, Functional and Nutraceutical Properties; Springer: New York, 2021.

(52) Spychaj, T.; Wilpiszewska, K.; Zdanowicz, M. Medium and high substituted carboxymethyl starch: Synthesis, characterization and application. *Starch - Stärke* **2013**, *65* (1–2), 22–33.

(53) Woggum, T.; Sirivongpaisal, P.; Wittaya, T. Characteristics and properties of hydroxypropylated rice starch based biodegradable films. *Food Hydrocolloids* **2015**, *50*, 54–64.

(54) Simsek, S.; Ovando-Martínez, M.; Whitney, K.; Bello-Pérez, L. A. Effect of acetylation, oxidation and annealing on physicochemical properties of bean starch. *Food Chem.* **2012**, *134* (4), *1796*–1803.

(55) Muhammad, K.; Hussin, F.; Man, Y. C.; Ghazali, H. M.; Kennedy, J. F. Effect of pH on phosphorylation of sago starch. *Carbohydr. Polym.* **2000**, *42* (1), 85–90.

(56) Bai, Y.; Shi, Y.-C. Structure and preparation of octenyl succinic esters of granular starch, microporous starch and soluble maltodextrin. *Carbohydr. Polym.* **2011**, 83 (2), 520–527.

(57) Sandhu, K. S.; Kaur, M.; Singh, N.; Lim, S.-T. A comparison of native and oxidized normal and waxy corn starches: Physicochemical, thermal, morphological and pasting properties. *LWT - Food Science and Technology* **2008**, *41* (6), 1000–1010.

(58) Amaraweera, S. M.; Gunathilake, C.; Gunawardene, O. H. P.; Fernando, N. M. L.; Wanninayaka, D. B.; Dassanayake, R. S.; Rajapaksha, S. M.; Manamperi, A.; Fernando, C. A. N.; Kulatunga, A. K.; Manipura, A. Development of Starch-Based Materials Using Current Modification Techniques and Their Applications: A Review. *Molecules* **2021**, *26* (22), 6880.

(59) Guarás, M. P.; Ludueña, L. N.; Alvarez, V. A. Chapter 4 -Development of Biodegradable Products from Modified Starches. In *Starch-Based Materials in Food Packaging*; Villar, M. A., Barbosa, S. E., García, M. A., Castillo, L. A., López, O. V., Eds.; Academic Press: New York, 2017; pp 77–124.

(60) Thakur, B. R.; Singh, R. K.; Handa, A. K.; Rao, M. A. Chemistry and uses of pectin — A review. *Critical Reviews in Food Science and Nutrition* **1997**, 37 (1), 47–73.

(61) Sinitsya, A.; Čopíková, J.; Prutyanov, V.; Skoblya, S.; Machovič, V. Amidation of highly methoxylated citrus pectin with primary amines. *Carbohydr. Polym.* **2000**, *42* (4), 359–368.

(62) Mortensen, A.; Aguilar, F.; Crebelli, R.; Di Domenico, A.; Dusemund, B.; Frutos, M. J.; Galtier, P.; Gott, D.; Gundert-Remy, U.; Lambré, C.; Leblanc, J.-C.; Lindtner, O.; Moldeus, P.; Mosesso, P.; Oskarsson, A.; Parent-Massin, D.; Stankovic, I.; Waalkens-Berendsen, I.; Wright, M.; Younes, M.; Tobback, P.; Ioannidou, S.; Tasiopoulou, S.; Woutersen, R. A. Re-evaluation of pectin (E 440i) and amidated pectin (E 440ii) as food additives. *EFSA Journal* **2017**, *15*, e04866.

(63) Sharma, R.; Ahuja, M. Thiolated pectin: Synthesis, characterization and evaluation as a mucoadhesive polymer. *Carbohydr. Polym.* **2011**, *85* (3), 658–663.

(64) Hu, Y.; Ye, X.; Yin, X.; Chen, S. Sulfation of citrus pectin by pyridine-sulfurtrioxide complex and its anticoagulant activity. *LWT* - *Food Science and Technology* **2015**, *60* (2, Part 2), 1162–1167.

(65) Tummalapalli, M.; Berthet, M.; Verrier, B.; Deopura, B. L.; Alam, M. S.; Gupta, B. Drug loaded composite oxidized pectin and gelatin networks for accelerated wound healing. *Int. J. Pharm.* **2016**, 505 (1), 234–245.

(66) Konduri, M. K. R.; Fatehi, P. Synthesis and characterization of carboxymethylated xylan and its application as a dispersant. *Carbohydr. Polym.* **2016**, *146*, 26–35.

(67) Daus, S.; Petzold-Welcke, K.; Kötteritzsch, M.; Baumgaertel, A.; Schubert, U. S.; Heinze, T. Homogeneous Sulfation of Xylan from Different Sources. *Macromol. Mater. Eng.* **2011**, *296* (6), 551–561.

(68) Qian, S.; Yu, Y.; Ma, J.; Diao, E.; Ye, S.; Gao, J.; Liu, Y.; Hu, W. Evaluation of a novel phosphorylated corn straw xylan for enhancement of thermal stability, crystallinity and functional activity. *International Journal of Food Science & Technology* **2022**, *57* (1), 481–488.

(69) Kallioinen, A.; Hakola, M.; Řiekkola, T.; Repo, T.; Leskelä, M.; von Weymarn, N.; Siika-aho, M. A novel alkaline oxidation pretreatment for spruce, birch and sugar cane bagasse. *Bioresour. Technol.* **2013**, *140*, 414–420.

(70) Xu, C.; Eckerman, C.; Smeds, A.; Reunanen, M.; Eklund, P. C.; Sjöholm, R.; Willför, S. Carboxymethylated spruce galactoglucomannans: preparation, characterisation, dispersion stability, water-inoil emulsion stability, and sorption on cellulose surface. *Nordic Pulp & Paper Research Journal* **2011**, *26* (2), 1–12.

(71) Liu, Y.; Huang, G. The derivatization and antioxidant activities of yeast mannan. *Int. J. Biol. Macromol.* **2018**, *107*, 755–761.

(72) BeMiller, J. N. Polysaccharides: Occurrence, Structures, and Chemistry. In *Carbohydrate Chemistry for Food Scientists*, 3rd ed.; BeMiller, J. N., Ed.; AACC International Press: UK,2019; pp 75–101.

pubs.acs.org/JAFC

(73) Willför, S.; Sjöholm, R.; Laine, C.; Holmbom, B. Structural features of water-soluble arabinogalactans from Norway spruce and Scots pine heartwood. *Wood Science and Technology* **2002**, *36* (2), 101–110.

(74) Xu, C.; Willför, S.; Sundberg, K.; Pettersson, C.; Holmbom, B. Physico-chemical characterization of spruce galactoglucomannan solutions: stability, surface activity and rheology. *Cellulose Chem. Technol.* **2007**, *41*, 51.

(75) Parikka, K.; Leppänen, A.-S.; Pitkänen, L.; Reunanen, M.; Willför, S.; Tenkanen, M. Oxidation of Polysaccharides by Galactose Oxidase. *J. Agric. Food Chem.* **2010**, 58 (1), 262–271.

(76) Kong, F.; Engler, C. R.; Soltes, E. J. Effects of cell-wall acetate, xylan backbone, and lignin on enzymatic hydrolysis of aspen wood. *Appl. Biochem. Biotechnol.* **1992**, *34* (1), 23–35.

(77) Sixta, H.; Iakovlev, M.; Testova, L.; Roselli, A.; Hummel, M.; Borrega, M.; van Heiningen, A.; Froschauer, C.; Schottenberger, H. Novel concepts of dissolving pulp production. *Cellulose* **2013**, *20* (4), 1547–1561.

(78) Bhattarai, M.; Sulaeva, I.; Pitkänen, L.; Kontro, I.; Tenkanen, M.; Potthast, A.; Mikkonen, K. S. Colloidal features of softwood galactoglucomannans-rich extract. *Carbohydr. Polym.* **2020**, *241*, 116368.

(79) Kishani, S.; Escalante, A.; Toriz, G.; Vilaplana, F.; Gatenholm, P.; Hansson, P.; Wagberg, L. Experimental and Theoretical Evaluation of the Solubility/Insolubility of Spruce Xylan (Arabino Glucuronoxylan). *Biomacromolecules* **2019**, *20* (3), 1263–1270.

(80) Mozdyniewicz, D. J.; Nieminen, K.; Sixta, H. Alkaline steeping of dissolving pulp. Part I: cellulose degradation kinetics. *Cellulose* **2013**, 20 (3), 1437–1451.

(81) Palasingh, C.; Nakayama, K.; Abik, F.; Mikkonen, K. S.; Evenäs, L.; Ström, A.; Nypelö, T. Modification of xylan via oxidation-reduction reaction. *Carbohydr. Polym.* **2022**, *292*, 119660.

(82) Hettrich, K.; Drechsler, U.; Loth, F.; Volkert, B. Preparation and Characterization of Water-Soluble Xylan Ethers. *Polymers* **2017**, *9* (4), 129.

(83) Guo, M. Q.; Hu, X.; Wang, C.; Ai, L. Polysaccharides: structure and solubility. *Solubility of Polysaccharides*; IntechOpen: 2017; Vol. 2, pp 8–21.

(84) Nypelö, T.; Laine, C.; Colson, J.; Henniges, U.; Tammelin, T. Submicron hierarchy of cellulose nanofibril films with etherified hemicelluloses. *Carbohydr. Polym.* **2017**, *177*, 126–134.

(85) Hartman, J.; Albertsson, A.-C.; Lindblad, M. S.; Sjöberg, J. Oxygen barrier materials from renewable sources: Material properties of softwood hemicellulose-based films. *J. Appl. Polym. Sci.* **2006**, *100* (4), 2985–2991.

(86) Felton, L. A. Mechanisms of polymeric film formation. Int. J. Pharm. 2013, 457 (2), 423–7.

(87) Hedenqvist, M. S. Barrier packaging materials. *Handbook of Environmental Degradation of Materials*; Elsevier: New York, 2012; pp 833–862.

(88) Escalante, A.; Goncalves, A.; Bodin, A.; Stepan, A.; Sandstrom, C.; Toriz, G.; Gatenholm, P. Flexible oxygen barrier films from spruce xylan. *Carbohydr. Polym.* **2012**, *87* (4), 2381–2387.

(89) Laine, C.; Harlin, A.; Hartman, J.; Hyvärinen, S.; Kammiovirta, K.; Krogerus, B.; Pajari, H.; Rautkoski, H.; Setälä, H.; Sievänen, J. Hydroxyalkylated xylans–Their synthesis and application in coatings for packaging and paper. *Industrial crops and products* **2013**, *44*, 692–704.

(90) Mikkonen, K. S.; Laine, C.; Kontro, I.; Talja, R. A.; Serimaa, R.; Tenkanen, M. Combination of internal and external plasticization of hydroxypropylated birch xylan tailors the properties of sustainable barrier films. *Eur. Polym. J.* **2015**, *66*, 307–318.

(91) Mikkonen, K. S.; Tenkanen, M. Sustainable food-packaging materials based on future biorefinery products: Xylans and mannans. *Trends Food Sci. Tech* **2012**, *28* (2), 90–102.

(92) Chen, M.-H.; Bergman, C. J. Method for determining the amylose content, molecular weights, and weight- and molar-based distributions of degree of polymerization of amylose and fine-structure of amylopectin. *Carbohydr. Polym.* **2007**, *69* (3), 562–578.

(93) Zinovyev, G.; Sulaeva, I.; Podzimek, S.; Rössner, D.; Kilpeläinen, I.; Sumerskii, I.; Rosenau, T.; Potthast, A. Getting Closer to Absolute Molar Masses of Technical Lignins. *ChemSusChem* **2018**, *11* (18), 3259–3268.

(94) Borrega, M.; Nieminen, K.; Sixta, H. Degradation kinetics of the main carbohydrates in birch wood during hot water extraction in a batch reactor at elevated temperatures. *Bioresour. Technol.* **2011**, *102* (22), 10724–10732.

(95) Pedersen, M.; Meyer, A. S. Lignocellulose pretreatment severity – relating pH to biomatrix opening. *New Biotechnology* **2010**, *27* (6), 739–750.

(96) Johansson, M. H.; Samuelson, O. Reducing end groups in brich xylan and their alkaline degradation. *Wood science and technology* **1977**, *11* (4), 251–263.

(97) Song, T.; Pranovich, A.; Sumerskiy, I.; Holmbom, B. Extraction of galactoglucomannan from spruce wood with pressurised hot water. *HF* **2008**, *62*, 659–666.

(98) Chadni, M.; Bals, O.; Ziegler-Devin, I.; Brosse, N.; Grimi, N. Microwave-assisted extraction of high-molecular-weight hemicelluloses from spruce wood. *C. R. Chim.* **2019**, *22* (8), 574–584.

(99) Kilpeläinen, P.; Leppänen, K.; Spetz, P.; Kitunen, V.; Ilvesniemi, H.; Pranovich, A.; Willför, S. BIOREFINERY. Pressurised hot water extraction of acetylated xylan from birch sawdust. *Nordic Pulp & Paper Research Journal* **2012**, *27* (4), 680–688.

(100) Koshijima, T.; Timell, T. E.; Zinbo, M. The number-average molecular weight of native hardwood xylans. *Journal of Polymer Science Part C: Polymer Symposia* **1965**, *11* (1), 265–279.

(101) Laine, C.; Kemppainen, K.; Kuutti, L.; Varhimo, A.; Asikainen, S.; Grönroos, A.; Määttänen, M.; Buchert, J.; Harlin, A. Extraction of xylan from wood pulp and brewer's spent grain. *Industrial Crops and Products* **2015**, *70*, 231–237.

(102) Bosmans, T. J.; Stépán, A. M.; Toriz, G.; Renneckar, S.; Karabulut, E.; Wågberg, L.; Gatenholm, P. Assembly of Debranched Xylan from Solution and on Nanocellulosic Surfaces. *Biomacromolecules* **2014**, *15* (3), 924–930.

(103) Schaubeder, J. B.; Ravn, J. L.; Orzan, E. J. Q.; Manfrão-Netto, J. H. C.; Geijer, C.; Nypelö, T.; Spirk, S. Xylan-cellulose thin film platform for assessing xylanase activity. *Carbohydr. Polym.* **2022**, 294, 119737.

(104) Jaafar, Z.; Mazeau, K.; Boissière, A.; Le Gall, S.; Villares, A.; Vigouroux, J.; Beury, N.; Moreau, C.; Lahaye, M.; Cathala, B. Meaning of xylan acetylation on xylan-cellulose interactions: A quartz crystal microbalance with dissipation (QCM-D) and molecular dynamic study. *Carbohydr. Polym.* **2019**, *226*, 115315.

(105) Kishani, S.; Vilaplana, F.; Ruda, M.; Hansson, P.; Wågberg, L. Influence of Solubility on the Adsorption of Different Xyloglucan Fractions at Cellulose–Water Interfaces. *Biomacromolecules* **2020**, *21* (2), 772–782.

(106) Lopez, M.; Bizot, H.; Chambat, G.; Marais, M.-F.; Zykwinska, A.; Ralet, M.-C.; Driguez, H.; Buléon, A. Enthalpic Studies of Xyloglucan–Cellulose Interactions. *Biomacromolecules* **2010**, *11* (6), 1417–1428.

(107) Kabel, M. A.; van den Borne, H.; Vincken, J.-P.; Voragen, A. G. J.; Schols, H. A. Structural differences of xylans affect their interaction with cellulose. *Carbohydr. Polym.* **2007**, *69* (1), 94–105.

(108) Andrewartha, K. A.; Phillips, D. R.; Stone, B. A. Solution properties of wheat-flour arabinoxylans and enzymically modified arabinoxylans. *Carbohydr. Res.* **1979**, *77* (1), 191–204.

(109) Linder, Å.; Bergman, R.; Bodin, A.; Gatenholm, P. Mechanism of Assembly of Xylan onto Cellulose Surfaces. *Langmuir* **2003**, *19* (12), 5072–5077.

(110) Chambat, G.; Karmous, M.; Costes, M.; Picard, M.; Joseleau, J.-P. Variation of xyloglucan substitution pattern affects the sorption on celluloses with different degrees of crystallinity. *Cellulose* **2005**, *12* (2), 117–125.

(111) Lima, D. U.; Loh, W.; Buckeridge, M. S. Xyloglucan-cellulose interaction depends on the sidechains and molecular weight of xyloglucan. *Plant Physiology and Biochemistry* **2004**, *42* (5), 389–394.

(112) Hannuksela, T.; Tenkanen, M.; Holmbom, B. Sorption of dissolved galactoglucomannans and galactomannans to bleached kraft pulp. *Cellulose* **2002**, *9* (3), 251–261.

(113) Fleer, G.; Stuart, M. C.; Scheutjens, J. M.; Cosgrove, T.; Vincent, B. *Polymers at Interfaces*; Springer Science & Business Media: New York, 1993.

(114) Lee, S. H.; Lee, H. L.; Youn, H. J. Adsorption and viscoelastic properties of cationic xylan on cellulose film using QCM-D. *Cellulose* **2014**, *21* (3), 1251–1260.

(115) Meng, X.; Luosang, D.; Meng, S.; Wang, R.; Fan, W.; Liang, D.; Li, X.; Zhao, Q.; Yang, L. The structural and functional properties of polysaccharide foulants in membrane fouling. *Chemosphere* **2021**, *268*, 129364.

(116) Benselfelt, T.; Cranston, E. D.; Ondaral, S.; Johansson, E.; Brumer, H.; Rutland, M. W.; Wågberg, L. Adsorption of Xyloglucan onto Cellulose Surfaces of Different Morphologies: An Entropy-Driven Process. *Biomacromolecules* **2016**, *17* (9), 2801–2811.

(117) Bhattarai, M.; Pitkänen, L.; Kitunen, V.; Korpinen, R.; Ilvesniemi, H.; Kilpeläinen, P. O.; Lehtonen, M.; Mikkonen, K. S. Functionality of spruce galactoglucomannans in oil-in-water emulsions. *Food Hydrocolloids* **2019**, *86*, 154–161.

(118) Hromádková, Z.; Ebringerová, A.; Malovíková, A. The Structural, Molecular and Functional Properties of Lignin-Containing Beechwood Glucuronoxylan. *Macromol. Symp.* **2005**, *232* (1), 19–26. (119) Ebringerova, A.; Srokova, I.; Talaba, P.; Kacurakova, M.;

Hromadkova, Z. Amphiphilic beechwood glucuronoxylan derivatives. J. Appl. Polym. Sci. **1998**, 67 (9), 1523–1530.

(120) Skalková, P.; Sroková, I.; Sasinková, V.; Ebringerová, A. Polymeric Surfactants from Beechwood Glucuronoxylan. *Tenside Surfactants Detergents* **2006**, *43* (3), 137–141.

(121) Hu, Z.; Xiang, Z.; Lu, F. Synthesis and emulsifying properties of long-chain succinic acid esters of glucuronoxylans. *Cellulose* **2019**, *26* (6), 3713–3724.

(122) Mikkonen, K. S.; Merger, D.; Kilpeläinen, P.; Murtomäki, L.; Schmidt, U. S.; Wilhelm, M. Determination of physical emulsion stabilization mechanisms of wood hemicelluloses via rheological and interfacial characterization. *Soft Matter* **2016**, *12* (42), 8690–8700.

(123) Mikkonen, K. S.; Xu, C.; Berton-Carabin, C.; Schroën, K. Spruce galactoglucomannans in rapeseed oil-in-water emulsions: Efficient stabilization performance and structural partitioning. *Food Hydrocolloids* **2016**, *52*, 615–624.

(124) McClements, D. J. Food Emulsions: Principles, Practices, and Techniques, 3rd ed.; CRC Press: New York, 2015.

(125) Dickinson, E. Hydrocolloids at interfaces and the influence on the properties of dispersed systems. *Food Hydrocolloids* **2003**, *17* (1), 25–39.

(126) Randall, R. C.; Phillips, G. O.; Williams, P. A. The role of the proteinaceous component on the emulsifying properties of gum arabic. *Food Hydrocolloids* **1988**, *2* (2), 131–140.

(127) Atgié, M.; Masbernat, O.; Roger, K. Emulsions Stabilized by Gum Arabic: Composition and Packing within Interfacial Films. *Langmuir* **2019**, *35* (4), 962–972.

(128) Yadav, M. P.; Manuel Igartuburu, J.; Yan, Y.; Nothnagel, E. A. Chemical investigation of the structural basis of the emulsifying activity of gum arabic. *Food Hydrocolloids* **2007**, *21* (2), 297–308.

(129) Carvalho, D. M. d.; Lahtinen, M. H.; Lawoko, M.; Mikkonen, K. S. Enrichment and Identification of Lignin–Carbohydrate Complexes in Softwood Extract. *ACS Sustainable Chem. Eng.* **2020**, *8* (31), 11795–11804.

(130) Lehtonen, M.; Merinen, M.; Kilpeläinen, P. O.; Xu, C.; Willför, S. M.; Mikkonen, K. S. Phenolic residues in spruce galactoglucomannans improve stabilization of oil-in-water emulsions. *J. Colloid Interface Sci.* **2018**, *512*, 536–547.

(131) Carvalho, D. M. d.; Lahtinen, M. H.; Bhattarai, M.; Lawoko, M.; Mikkonen, K. S. Active role of lignin in anchoring wood-based stabilizers to the emulsion interface. *Green Chem.* **2021**, *23* (22), 9084– 9098. (132) Sarkar, N. Structural interpretation of the interfacial properties of aqueous solutions of methylcellulose and hydroxypropyl methylcellulose. *Polymer* **1984**, 25 (4), 481–486.

(133) Dax, D.; Eklund, P.; Hemming, J.; Sarfraz, J.; Backman, P.; Xu, C.; Willför, S. Amphiphilic Spruce Galactoglucomannan Derivatives Based on Naturally-Occurring Fatty Acids. *BioResources* **2013**, *8*, 3771.

(134) Shen, F.; Ling, H.; Ge, W.; Yang, Y.; Wang, X.; Ren, J.; Wang, X. Self-assembly behavior and conformation of amphiphilic hemicellulosegraft-fatty acid micelles. *Carbohydr. Polym.* **2021**, *261*, 117886.

(135) Nypelö, T.; Laine, C.; Aoki, M.; Tammelin, T.; Henniges, U. Etherification of Wood-Based Hemicelluloses for Interfacial Activity. *Biomacromolecules* **2016**, *17* (5), 1894–1901.

(136) Pawar, P.; Koutaniemi, S.; Tenkanen, M.; Mellerowicz, E. Acetylation of woody lignocellulose: significance and regulation. *Frontiers in Plant Science* **2013**, *4*, 118.

(137) Chevalier, Y.; Bolzinger, M.-A. Emulsions stabilized with solid nanoparticles: Pickering emulsions. *Colloids Surf.*, A **2013**, 439, 23–34.

(138) Calabrese, V.; Courtenay, J. C.; Edler, K. J.; Scott, J. L. Pickering emulsions stabilized by naturally derived or biodegradable particles. *Current Opinion in Green and Sustainable Chemistry* **2018**, *12*, 83–90.

(139) Murray, B. S. Pickering emulsions for food and drinks. *Current Opinion in Food Science* 2019, 27, 57–63.

(140) Meng, Z.; Sawada, D.; Laine, C.; Ogawa, Y.; Virtanen, T.; Nishiyama, Y.; Tammelin, T.; Kontturi, E. Bottom-up Construction of Xylan Nanocrystals in Dimethyl Sulfoxide. *Biomacromolecules* **2021**, 22 (2), 898–906.

(141) Heinze, T.; Hornig, S. Versatile concept for the structure design of polysaccharide-based nanoparticles. *Polysaccharide Materials: Performance by Design*; American Chemical Society: Washington, DC, 2009; pp 169–183.

(142) Heinze, T.; Petzold, K.; Hornig, S. Novel nanoparticles based on xylan. *Cellulose Chem. Technol.* **2007**, *41*, 13–18.

(143) Hornig, S.; Heinze, T.; Becer, C. R.; Schubert, U. S. Synthetic polymeric nanoparticles by nanoprecipitation. *J. Mater. Chem.* **2009**, *19* (23), 3838–3840.

(144) Wang, S.; Xiang, Z. Highly Stable Pickering Emulsions with Xylan Hydrate Nanocrystals. *Nanomaterials* **2021**, *11* (10), 2558.

(145) Valoppi, F.; Lahtinen, M. H.; Bhattarai, M.; Kirjoranta, S. J.; Juntti, V. K.; Peltonen, L. J.; Kilpeläinen, P. O.; Mikkonen, K. S. Centrifugal fractionation of softwood extracts improves the biorefinery workflow and yields functional emulsifiers. *Green Chem.* **2019**, *21* (17), 4691–4705.

(146) Dai, L.; Li, Y.; Kong, F.; Liu, K.; Si, C.; Ni, Y. Lignin-Based Nanoparticles Stabilized Pickering Emulsion for Stability Improvement and Thermal-Controlled Release of trans-Resveratrol. *ACS Sustainable Chem. Eng.* **2019**, *7* (15), 13497–13504.

(147) Westbye, P.; Köhnke, T.; Glasser, W.; Gatenholm, P. The influence of lignin on the self-assembly behaviour of xylan rich fractions from birch (Betula pendula). *Cellulose* **2007**, *14* (6), 603–613.

(148) Renard, D.; Garnier, C.; Lapp, A.; Schmitt, C.; Sanchez, C. Structure of arabinogalactan-protein from Acacia gum: From porous ellipsoids to supramolecular architectures. *Carbohydr. Polym.* **2012**, *90* (1), 322–332.

(149) Atgié, M.; Garrigues, J. C.; Chennevière, A.; Masbernat, O.; Roger, K. Gum Arabic in solution: Composition and multi-scale structures. *Food Hydrocolloids* **2019**, *91*, 319–330.

(150) Isobe, N.; Sagawa, N.; Ono, Y.; Fujisawa, S.; Kimura, S.; Kinoshita, K.; Miuchi, T.; Iwata, T.; Isogai, A.; Nishino, M.; Deguchi, S. Primary structure of gum arabic and its dynamics at oil/water interface. *Carbohydr. Polym.* **2020**, *249*, 116843.

(151) Atgié, M.; Chennevière, A.; Masbernat, O.; Roger, K. Emulsions Stabilized by Gum Arabic: How Diversity and Interfacial Networking Lead to Metastability. *Langmuir* **2019**, 35 (45), 14553–14565.

(152) Penfold, J.; Thomas, R. K. Neutron reflectivity and small angle neutron scattering: An introduction and perspective on recent progress. *Curr. Opin. Colloid Interface Sci.* **2014**, *19* (3), 198–206.

(153) Ho, T. M.; Abik, F.; Mikkonen, K. S. An overview of nanoemulsion characterization via atomic force microscopy. *Crit. Rev. Food Sci. Nutr.* **2021**, 1–21.

pubs.acs.org/JAFC

(154) Liu, L.; Poveda, C.; Jenkins, P. E.; Walton, G. E. An In Vitro Approach to Studying the Microbial Community and Impact of Pre and Probiotics under Anorexia Nervosa Related Dietary Restrictions. *Nutrients* **2021**, *13* (12), 4447.

(155) Patterson, E.; Ryan, P. M.; Cryan, J. F.; Dinan, T. G.; Ross, R. P.; Fitzgerald, G. F.; Stanton, C. Gut microbiota, obesity and diabetes. *Postgraduate Medical Journal* **2016**, *92* (1087), 286.

(156) Lopez-Siles, M.; Martinez-Medina, M.; Surís-Valls, R.; Aldeguer, X.; Sabat-Mir, M.; Duncan, S. H.; Flint, H. J.; Garcia-Gil, L. J. Changes in the Abundance of Faecalibacterium prausnitzii Phylogroups I and II in the Intestinal Mucosa of Inflammatory Bowel Disease and Patients with Colorectal Cancer. *Inflammatory Bowel Diseases* **2016**, *22* (1), 28–41.

(157) Leth, M. L.; Ejby, M.; Workman, C.; Ewald, D. A.; Pedersen, S. S.; Sternberg, C.; Bahl, M. I.; Licht, T. R.; Aachmann, F. L.; Westereng, B.; Abou Hachem, M. Differential bacterial capture and transport preferences facilitate co-growth on dietary xylan in the human gut. *Nature Microbiology* **2018**, *3* (5), 570–580.

(158) Rogowski, A.; Briggs, J. A.; Mortimer, J. C.; Tryfona, T.; Terrapon, N.; Lowe, E. C.; Baslé, A.; Morland, C.; Day, A. M.; Zheng, H.; Rogers, T. E.; Thompson, P.; Hawkins, A. R.; Yadav, M. P.; Henrissat, B.; Martens, E. C.; Dupree, P.; Gilbert, H. J.; Bolam, D. N. Glycan complexity dictates microbial resource allocation in the large intestine. *Nat. Commun.* **2015**, *6* (1), 7481.

(159) La Rosa, S. L.; Leth, M. L.; Michalak, L.; Hansen, M. E.; Pudlo, N. A.; Glowacki, R.; Pereira, G.; Workman, C. T.; Arntzen, M. Ø.; Pope, P. B.; Martens, E. C.; Hachem, M. A.; Westereng, B. The human gut Firmicute Roseburia intestinalis is a primary degrader of dietary β -mannans. *Nat. Commun.* **2019**, *10* (1), 905.

(160) Lindstad, L. J.; Lo, G.; Leivers, S.; Lu, Z.; Michalak, L.; Pereira, G. V.; Røhr, Å. K.; Martens, E. C.; McKee, L. S.; Louis, P.; Duncan, S. H.; Westereng, B.; Pope, P. B.; Rosa, S. L. L.; Bello, M. G. D. Human Gut Faecalibacterium prausnitzii Deploys a Highly Efficient Conserved System To Cross-Feed on β -Mannan-Derived Oligosaccharides. *mBio* **2021**, *12* (3), e03628–20.

(161) Bågenholm, V.; Reddy, S. K.; Bouraoui, H.; Morrill, J.; Kulcinskaja, E.; Bahr, C. M.; Aurelius, O.; Rogers, T.; Xiao, Y.; Logan, D. T. Galactomannan catabolism conferred by a polysaccharide utilization locus of Bacteroides ovatus: enzyme synergy and crystal structure of a β -mannanase. J. Biol. Chem. **2017**, 292 (1), 229–243.

(162) Michalak, L.; Gaby, J. C.; Lagos, L.; La Rosa, S. L.; Hvidsten, T. R.; Tétard-Jones, C.; Willats, W. G. T.; Terrapon, N.; Lombard, V.; Henrissat, B.; Dröge, J.; Arntzen, M. Ø.; Hagen, L. H.; Øverland, M.; Pope, P. B.; Westereng, B. Microbiota-directed fibre activates both targeted and secondary metabolic shifts in the distal gut. *Nat. Commun.* **2020**, *11* (1), 5773.

(163) Ejby, M.; Fredslund, F.; Vujicic-Zagar, A.; Svensson, B.; Slotboom, D. J.; Abou Hachem, M. Structural basis for arabinoxylooligosaccharide capture by the probiotic Bifidobacterium animalis subsp. lactis Bl-04. *Mol. Microbiol.* **2013**, *90* (5), 1100–1112.

(164) Rosa, S. L. L.; Kachrimanidou, V.; Buffetto, F.; Pope, P. B.; Pudlo, N. A.; Martens, E. C.; Rastall, R. A.; Gibson, G. R.; Westereng, B.; Marco, M. L. Wood-Derived Dietary Fibers Promote Beneficial Human Gut Microbiota. *mSphere* **2019**, *4*, e00554.

(165) Miquel, S.; Martín, R.; Rossi, O.; Bermúdez-Humarán, L. G.; Chatel, J. M.; Sokol, H.; Thomas, M.; Wells, J. M.; Langella, P. Faecalibacterium prausnitzii and human intestinal health. *Curr. Opin. Microbiol.* **2013**, *16* (3), 255–261.

(166) Li, Y.-F.; Yue, P.-P.; Hao, X.; Bian, J.; Ren, J.-L.; Peng, F.; Sun, R.-C. Comparison of emulsifying capacity of two hemicelluloses from moso bamboo in soy oil-in-water emulsions. *RSC Adv.* **2020**, *10* (8), 4657–4663.

(167) Wang, L.; Wu, M.; Liu, H.-M. Emulsifying and physicochemical properties of soy hull hemicelluloses-soy protein isolate conjugates. *Carbohydr. Polym.* **201**7, *163*, 181–190.

(168) Lehtonen, M.; Teräslahti, S.; Xu, C.; Yadav, M. P.; Lampi, A.-M.; Mikkonen, K. S. Spruce galactoglucomannans inhibit lipid oxidation in rapeseed oil-in-water emulsions. *Food Hydrocolloids* **2016**, 58, 255–266. (169) Valoppi, F.; Maina, N.; Allén, M.; Miglioli, R.; Kilpeläinen, P. O.; Mikkonen, K. S. Spruce galactoglucomannan-stabilized emulsions as essential fatty acid delivery systems for functionalized drinkable yogurt and oat-based beverage. *European Food Research and Technology* **2019**, *245* (7), 1387–1398.

(170) Tatar, F.; Tunç, M. T.; Dervisoglu, M.; Cekmecelioglu, D.; Kahyaoglu, T. Evaluation of hemicellulose as a coating material with gum arabic for food microencapsulation. *Food Research International* **2014**, *57*, 168–175.

(171) Kirjoranta, S.; Knaapila, A.; Kilpeläinen, P.; Mikkonen, K. S. Sensory profile of hemicellulose-rich wood extracts in yogurt models. *Cellulose* **2020**, *27* (13), 7607–7620.

(172) Maga, J. A. The contribution of wood to the flavor of alcoholic beverages. *Food Reviews International* **1989**, *5* (1), 39–99.

(173) Cerezo, A. B.; Tesfaye, W.; Soria-Díaz, M. E.; Torija, M. J.; Mateo, E.; Garcia-Parrilla, M. C.; Troncoso, A. M. Effect of wood on the phenolic profile and sensory properties of wine vinegars during ageing. *Journal of Food Composition and Analysis* **2010**, 23 (2), 175–184.

(174) Polari, L.; Ojansivu, P.; Mäkelä, S.; Eckerman, C.; Holmbom, B.; Salminen, S. Galactoglucomannan Extracted from Spruce (Picea abies) as a Carbohydrate Source for Probiotic Bacteria. *J. Agric. Food Chem.* **2012**, *60* (44), 11037–11043.

(175) Rinne, M.; Kautto, O.; Kuoppala, K.; Ahvenjärvi, S.; Kitunen, V.; Ilvesniemi, H.; Wilför, S.; Sormunen-Cristian, R. Digestion of woodbased hemicellulose extracts as screened by in vitro gas production method and verified in vivo using sheep. *Agricultural and Food Science* **2016**, 25 (1), 13–21.

(176) Kynkäänniemi, E.; Lahtinen, M. H.; Jian, C.; Salonen, A.; Hatanpää, T.; Mikkonen, K. S.; Pajari, A.-M. Gut microbiota can utilize prebiotic birch glucuronoxylan in production of short-chain fatty acids in rats. *Food & Function* **2022**, *13* (6), 3746–3759.

(177) Deloule, V.; Boisset, C.; Hannani, D.; Suau, A.; Le Gouellec, A.; Chroboczek, J.; Botté, C.; Yamaryo-Botté, Y.; Chirat, C.; Toussaint, B. Prebiotic role of softwood hemicellulose in healthy mice model. *Journal* of Functional Foods **2020**, *64*, 103688.

(178) Damron, B. L.; Harms, R. H. The Value of Masonex for Use in Broiler Diets. *Poultry Science* **1968**, 47 (4), 1330–1333.

(179) Hartnell, G. F.; Satter, L. D. Effect of Masonex (Hemicellulose Extract) on Protein Degradation by Rumen Microorganisms in Vitro and in Vivo. *Journal of Animal Science* **1978**, 47 (4), 935–943.

(180) Konkol, Y.; Keskitalo, A.; Vuorikoski, H.; Pietilä, S.; Elo, L. L.; Munukka, E.; Bernoulli, J.; Tuomela, J. Chronic nonbacterial prostate inflammation in a rat model is associated with changes of gut microbiota that can be modified with a galactoglucomannan-rich hemicellulose extract in the diet. *BJU International* **2019**, *123* (5), 899– 908.

(181) Herrick, K. J.; Hippen, A. R.; Kalscheur, K. F.; Anderson, J. L.; Ranathunga, S. D.; Patton, R. S.; Abdullah, M. Lactation performance and digestibility of forages and diets in dairy cows fed a hemicellulose extract. *Journal of Dairy Science* **2012**, *95* (6), 3342–3353.

(182) Crawford, D. F.; Anthony, W. B.; Harris, R. R. Evaluation of Concentrated Hemicellulose Extract as Cattle Feed. *Journal of Animal Science* **1978**, *46* (1), 32–40.

(183) Drehmel, O. R.; Brown-Brandl, T. M.; Judy, J. V.; Fernando, S. C.; Miller, P. S.; Hales, K. E.; Kononoff, P. J. The influence of fat and hemicellulose on methane production and energy utilization in lactating Jersey cattle. *Journal of Dairy Science* **2018**, *101* (9), 7892–7906.

(184) Turck, D.; Bresson, J.-L.; Burlingame, B.; Dean, T.; Fairweather-Tait, S.; Heinonen, M.; Hirsch-Ernst, K. I.; Mangelsdorf, I.; McArdle, H. J.; Naska, A.; Neuhäuser-Berthold, M.; Nowicka, G.; Pentieva, K.; Sanz, Y.; Siani, A.; Sjödin, A.; Stern, M.; Tomé, D.; Vinceti, M.; Willatts, P.; Engel, K.-H.; Marchelli, R.; Pöting, A.; Poulsen, M.; Salminen, S.; Schlatter, J.; Arcella, D.; Gelbmann, W.; de Sesmaisons-Lecarré, A.; Verhagen, H.; van Loveren, H. Guidance on the preparation and submission of an application for authorisation of a novel food in the context of Regulation (EU) 2015/22831 (Revision 1)2. *EFSA Journal* **2021**, *19*, e06555. (185) de Boer, A.; Bast, A. Demanding safe foods – Safety testing under the novel food regulation (2015/2283). *Trends in Food Science & Technology* **2018**, *72*, 125–133.

(186) Ververis, E.; Ackerl, R.; Azzollini, D.; Colombo, P. A.; de Sesmaisons, A.; Dumas, C.; Fernandez-Dumont, A.; Ferreira da Costa, L.; Germini, A.; Goumperis, T.; Kouloura, E.; Matijevic, L.; Precup, G.; Roldan-Torres, R.; Rossi, A.; Svejstil, R.; Turla, E.; Gelbmann, W. Novel foods in the European Union: Scientific requirements and challenges of the risk assessment process by the European Food Safety Authority. *Food Research International* **2020**, *137*, 109515.

(187) Lange, L.; Meyer, A. S. Potentials and possible safety issues of using biorefinery products in food value chains. *Trends in Food Science* & *Technology* **2019**, *84*, 7–11.

(188) EFSA Panel on Dietetic Products, N.. Allergies, Scientific Opinion on Dietary Reference Values for carbohydrates and dietary fibre. *EFSA J.* **2010**, *8*, 1462.

(189) Stephen, A. M.; Champ, M. M. J.; Cloran, S. J.; Fleith, M.; van Lieshout, L.; Mejborn, H.; Burley, V. J. Dietary fibre in Europe: current state of knowledge on definitions, sources, recommendations, intakes and relationships to health. *Nutrition Research Reviews* **2017**, *30* (2), 149–190.

(190) Frequently Asked Questions About GRAS for Substances Intended for Use in Human or Animal Food: Guidance for Industry. Food and Drug Administration, 2016.

(191) Food and Drug Administration, About the GRAS Notification Program; https://www.fda.gov/food/generally-recognized-safe-gras/ about-gras-notification-program (accessed November 9, 2022).

(192) Guidance for Industry: Questions and Answers About the Food Additive or Color Additive Petition Process; https://www.fda.gov/ regulatory-information/search-fda-guidance-documents/guidanceindustry-questions-and-answers-about-food-additive-or-color-additivepetition-process#answerA (accessed November 9, 2022).

(193) Ayres, M. P.; Lombardero, M. a. J. Assessing the consequences of global change for forest disturbance from herbivores and pathogens. *Science of The Total Environment* **2000**, *262* (3), *263*–286.

(194) Kilpeläinen, P. O.; Hautala, S. S.; Byman, O. O.; Tanner, L. J.; Korpinen, R. I.; Lillandt, M. K. J.; Pranovich, A. V.; Kitunen, V. H.; Willför, S. M.; Ilvesniemi, H. S. Pressurized hot water flow-through extraction system scale up from the laboratory to the pilot scale. *Green Chem.* **2014**, *16* (6), 3186–3194.

(195) Gallina, G.; Alfageme, E. R.; Biasi, P.; García-Serna, J. Hydrothermal extraction of hemicellulose: from lab to pilot scale. *Bioresour. Technol.* **2018**, 247, 980–991.

Recommended by ACS

Feeding Habits, Nutritional Condition, Feeal Intestinal Metabolites, and Risk of Developing Diabetes Mellitus II in the Mexican Population: A Cross-Sectional Study

Diana Laura García-Martínez, Víctor Manuel Zamora-Gasga, et al. MARCH 23, 2023 ACS FOOD SCIENCE & TECHNOLOGY

READ 🗹

Mass Spectrometry-Based Untargeted Metabolomics Reveals the Importance of Glycosylated Flavones in Patterned Lentil Seed Coats

Fatma M. Elessawy, Randy W. Purves, et al.	
FEBRUARY 08, 2023	
JOURNAL OF AGRICULTURAL AND FOOD CHEMISTRY	READ 🗹

Multielement Profiling of Diverse Food Samples

Jacqueline M. Chaparro, Jessica E. Prenni, *et al.* FEBRUARY 24, 2023

In Vitro Digestibility and Physicochemical Properties of Potato (*Solanum tuberosum*) Fermented by Traditional and Alternative Processes under Water Currents

Karla Escandón, Marco A. Lazo-Vélez, et al. FEBRUARY 27, 2023 ACS FOOD SCIENCE & TECHNOLOGY

READ 🗹

Get More Suggestions >