

Review **Influence of the Mediterranean Diet on Healthy Aging**

Maria Carmen Andreo-López 1,†, Victoria Contreras-Bolívar 1,2,†, Manuel Muñoz-Torres 1,2,3,4,[*](https://orcid.org/0000-0002-9645-3260) , Beatriz García-Fontana 2,3,5,[‡](https://orcid.org/0000-0002-4134-5561) and Cristina García-Fontana 1,2,3,[‡](https://orcid.org/0000-0002-9328-6022)

- ¹ Endocrinology and Nutrition Unit, University Hospital Clínico San Cecilio, 18016 Granada, Spain
- $\overline{2}$ Instituto de Investigación Biosanitaria de Granada (Ibs. Granada), 18014 Granada, Spain
- ³ CIBER on Frailty and Healthy Aging (CIBERFES), Instituto de Salud Carlos III, 18012 Granada, Spain
⁴ Department of Modisine, University of Canada, 18016 Canada, Spain
- ⁴ Department of Medicine, University of Granada, 18016 Granada, Spain
5 Department of Gill Biglaces University of Granada, 18016 Granada, Spain
- ⁵ Department of Cell Biology, University of Granada, 18016 Granada, Spain
- ***** Correspondence: mmt@mamuto.es; Tel.: +34-958246124
- † M.C.A.-L. and V.C.-B. contributed equally as first co-authors.
- ‡ These authors contributed equally to this work and are listed as co-last.

Abstract: The life expectancy of the global population has increased. Aging is a natural physiological process that poses major challenges in an increasingly long-lived and frail population. Several molecular mechanisms are involved in aging. Likewise, the gut microbiota, which is influenced by environmental factors such as diet, plays a crucial role in the modulation of these mechanisms. The Mediterranean diet, as well as the components present in it, offer some proof of this. Achieving healthy aging should be focused on the promotion of healthy lifestyle habits that reduce the development of pathologies that are associated with aging, in order to increase the quality of life of the aging population. In this review we analyze the influence of the Mediterranean diet on the molecular pathways and the microbiota associated with more favorable aging patterns, as well as its possible role as an anti-aging treatment.

Keywords: aging; Mediterranean diet; molecular pathways; microbiome

Citation: Andreo-López, M.C.; Contreras-Bolívar, V.; Muñoz-Torres, M.; García-Fontana, B.; García-Fontana, C. Influence of the Mediterranean Diet on Healthy Aging. *Int. J. Mol. Sci.* **2023**, *24*, 4491. [https://doi.org/10.3390/](https://doi.org/10.3390/ijms24054491) [ijms24054491](https://doi.org/10.3390/ijms24054491)

Academic Editor: Shian-Jang Yan

Received: 27 January 2023 Revised: 18 February 2023 Accepted: 21 February 2023 Published: 24 February 2023

Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license [\(https://](https://creativecommons.org/licenses/by/4.0/) [creativecommons.org/licenses/by/](https://creativecommons.org/licenses/by/4.0/) $4.0/$).

1. Introduction

Currently, the global population has a notably increased life expectancy compared to decades ago, exceeding 60 years of age in most cases. According to the World Health Organization (WHO), the percentage of people over 60 years of age will double globally by 2050 [\[1\]](#page-19-0). However, a longer life expectancy leads us to reconsider not only the health of older people but also what kind of implications aging has [\[2\]](#page-19-1).

Aging is a natural physiological process that leads to a progressive loss of cellular functionality, with consequences that predispose people to an increased risk of frailty, morbidity, and mortality [\[3\]](#page-19-2). The role of lifestyle and diet can promote "healthy aging", in which quality of life takes precedence. According to the WHO, this concept refers to the process of developing and maintaining a functional capacity that enables well-being in old age [\[1](#page-19-0)[,4\]](#page-19-3).

Several cellular and molecular hallmarks are involved in the aging process. In particular, there are nine hallmarks that are decisive in the aging process: genomic instability, telomere attrition, epigenetic alterations, loss of proteostasis, the dysregulation of nutrient sensing, mitochondrial dysfunction, cellular senescence, stem cell depletion, and altered intercellular communication [\[5\]](#page-19-4). These molecular mechanisms are involved in the development of age-related diseases such as cancer, obesity, diabetes, cardiovascular disease (CVD), and neurodegenerative diseases [\[3\]](#page-19-2). These age-related diseases have been associated with risk factors that can be modified mainly through nutrition, which constitutes one of the pillars of health [\[6\]](#page-19-5). In addition, the microbiota, which is modified by diet, has also been involved in aging [\[6\]](#page-19-5). It has been suggested that the age-related decline in immune system function (immunosenescence) and chronic low-grade inflammation could

lead to microbiota disturbances that are associated with several age-related pathologies. Thus, it has been argued that a balanced diet can modulate the proliferation of specific bacteria within the gut microbiota. This has been associated with improved health status in older people [\[7\]](#page-20-0).

In light of the above, the aim of our review was to analyze the available data regarding the potential effects of the Mediterranean diet (MedDiet) as a whole, or of individual elements of it, on the nine hallmarks of aging, to provide further evidence of its health benefits. Moreover, we have also reviewed the effect of MedDiet on the microbiota and its relationship to aging.

2. Aging and Frailty: Biological Links

From a biological point of view, aging can be defined as the physiological and progressive accumulation of senescent cells in organs and tissues, which occurs during the lifetime of an individual and leads to progressive functional slowing or the total loss of function [\[8](#page-20-1)[–10\]](#page-20-2).

Pleiotropic antagonist genes comprise a set of genes that regulate cellular senescence, performing an important role in preventing the degeneration of malignant cells in the cell cycle [\[11,](#page-20-3)[12\]](#page-20-4). These genes are also involved in protective mechanisms in physiological cellular senescence processes and in age-related diseases. However, aging cells produce proinflammatory and lytic extracellular matrix molecules in a process known as the senescence-associated complex secretory phenotype (SASP), resulting in degeneration and pathological senescence. Moreover, the aging process involves the immune system; in particular, the cell-mediated defense mechanism is slowed down. Senescent cells do not produce sufficient signals to activate immune cells. Likewise, senescence is induced by the accumulation of various factors at the cellular level that is responsible for macromolecular damage, such as secondary DNA alterations due to oxidative damage, telomere shortening, and endoplasmic reticulum (ER) degeneration [\[13\]](#page-20-5). Thus, aging is the result of multifactorial interactions between local and systemic environmental factors and involutional factors due to cellular senescence. Therefore, the number of senescent cells in a person's body increases with age as the aging immune system becomes less efficient and senescent cells accumulate. This makes individuals more vulnerable to further deterioration after exposure to environmental stressors [\[13\]](#page-20-5). The disease occurs when environmental stressors attack tissues that are already in the presence of senescent cells with very low resilience [\[14](#page-20-6)[,15\]](#page-20-7).

Frailty develops due to an increasing decline usually linked to age, severe deterioration, and the onset of pathological states. This leads to a condition of increased vulnerability and reduced adaptive capacity, and ultimately, negative health changes are triggered by even mild stressors. It is considered more appropriate to speak of "frailty syndrome": a chronic pathological condition resulting from the interaction between several factors, including aging-related physiological alterations, pluripathology, nutritional deficiencies up to severe malnutrition, and the negative impact of socio-environmental factors [\[16\]](#page-20-8). In fact, a high proportion of undernourished people are frail, and undernutrition leads to weight loss, which can contribute to frailty syndrome [\[17\]](#page-20-9). At the other extreme, obesity increases the risk of frailty [\[18\]](#page-20-10). In terms of body composition, frailty has been associated with a higher body fat mass and fat percentage and with a low muscle mass and is often without association with the body mass index [\[19–](#page-20-11)[21\]](#page-20-12).

All of this can lead to the frail elderly losing all self-sufficiency, increasing the risk of falls, and can result in a state of confusion with severe impairment of cognitive functions that ultimately increases the risk of the development of diseases [\[22\]](#page-20-13).

3. The Mediterranean Diet

The term MedDiet was first coined by Ancel Keys in the 1960s [\[23\]](#page-20-14). The MedDiet reflects the dietary patterns typical of civilizations based around the Mediterranean Sea, especially Greece, the island of Crete, and southern Italy in the early 1960s [\[24\]](#page-20-15). In fact, the MedDiet is closely linked to traditional olive growing areas in the Mediterranean region

and has been associated with low rates of chronic diseases (lower risk of CVD and metabolic diseases associated with excess weight) and, consequently, high life expectancy [\[24](#page-20-15)[,25\]](#page-20-16).

The MedDiet is characterized by a high consumption of olive oil (OO) as the main source of fat—especially virgin (VOO) and extra virgin (EVOO)—and the high consumption of plant foods (vegetables, fruits, legumes, potatoes, bread, and other cereals (minimally refined), nuts and seeds), as well as fresh seasonal, locally grown, and minimally processed foods. Dairy intake is moderate (mainly cheeses and yogurts), and fish (an excellent source of long-chain poly-unsaturated fatty acids (PUFAs), particularly omega-3) and poultry are consumed in low or moderate amounts. The MetDiet includes the low consumption of red meat and sweets and moderate consumption of wine at meals. No more than four eggs are consumed per week. In general terms, caloric intake in the form of fat does not exceed 30% of the intake, with less than 8–10% contributed by saturated fats. Some bioactive compounds in the MedDiet include vitamins, minerals, polyphenols, fiber, nitrates, PUFAs, and mono-unsaturated fatty acids (MUFAs) that, in combination or separately, are beneficial to health [\[26](#page-20-17)[–28\]](#page-20-18). Among the PUFAs, the essential omega-6 fatty acid is linoleic acid (LA). In addition, longer omega-3 PUFAs, such as eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA), are derivatives of alpha-linoleic acid. These are mainly present in fish oils [\[29\]](#page-20-19). For these reasons, MedDiet is unique and different from other healthy dietary patterns [\[29\]](#page-20-19).

4. Interplay between Mediterranean Diet, Aging and Frailty

4.1. Mediterranean Diet and Hallmarks of Ageing

The aging process has been linked to nine distinctive cellular and molecular features [\[5\]](#page-19-4) (Figure [1\)](#page-2-0). Each of these plays its role in the trajectory of natural aging; their experimental exacerbation accelerates the process, and their optimization slows it down, thus increasing lifespan [\[30\]](#page-20-20). External lifestyle factors such as diet can modulate the aging process [\[31](#page-20-21)[,32\]](#page-20-22).

Figure 1. Impact of MedDiet on the hallmarks of aging. **Figure 1.** Impact of MedDiet on the hallmarks of aging.
 Figure 1. Impact of MedDiet on the hallmarks of aging.

4.1.1. Genomic Instability

Aging increases susceptibility to DNA alterations resulting from a combination of oxidative stress, epigenetic alterations, damaged DNA, and telomere attrition [\[33\]](#page-20-23). Unrepaired DNA could increase the risk of mutations and favor the beginning or development of age-related diseases [\[34,](#page-20-24)[35\]](#page-20-25). The MedDiet could play a protective role against genomic alterations. Indeed, bioactive compounds that are contained in the MedDiet, such as melatonin, phytosterols, carotenoids, polyphenols (such as resveratrol and hydroxytyrosol (HT)), vitamins, and glucosinolates (in cruciferous vegetables), can promote DNA repair and attenuate telomere shortening [\[36,](#page-21-0)[37\]](#page-21-1). These positive effects have been explained by the anti-inflammatory effects of the MedDiet and the direct and indirect (epigenetic) modifications induced by the MedDiet on gene expression [\[38–](#page-21-2)[41\]](#page-21-3).

DNA damage due to oxidative stress is a result of the failure of oxidative damagerepair mechanisms as a result of excess reactive oxygen species (ROS) [\[42\]](#page-21-4). Guanine is an important target for DNA oxidation, generating oxidized metabolites. Among them, 8-oxo-2 0 -deoxyguanosine (8-OHdG) is considered a marker of oxidative stress with mutagenic potential [\[43–](#page-21-5)[45\]](#page-21-6). Urquiaga et al. related the MedDiet, including a moderate intake of red wine, with a reduction in 8-OHdG levels in the peripheral blood leukocyte DNA, which impacts positively on the control of oxidative stress [\[46\]](#page-21-7). Similarly, lower levels of another marker of oxidative stress, the deoxyguanosine adduct, were associated with greater adherence to the MedDiet in the Italian cohort of the EPIC (European Investigation into Cancer and Nutrition) study [\[47\]](#page-21-8).

Nuts and cooked tomato sauce using OO highlight the protective role against oxidative DNA damage [\[48](#page-21-9)[–50\]](#page-21-10). This defense phenomenon was found in intervention studies in the human population after the consumption of tomato [\[51,](#page-21-11)[52\]](#page-21-12), broccoli [\[53,](#page-21-13)[54\]](#page-21-14), spinach [\[55](#page-21-15)[,56\]](#page-21-16), and blueberries [\[57](#page-21-17)[,58\]](#page-21-18). In addition, a study in rats fed with a diet enriched with VOO found an association with less damage in the genetic material in peripheral blood cells vs. rats fed with sunflower oil [\[59\]](#page-21-19). In this context, MUFAs and PUFAs have been demonstrated to play a protective role against oxidative stress and DNA damage [\[60–](#page-22-0)[62\]](#page-22-1). In a sub-analysis of the PREDIMED study, the intervention group with the MedDiet with MUFA (EVOO) or PUFA (walnuts) compared to the control (low-fat diet) showed a significant improvement in cardiovascular outcomes and a lower proportion of oxidative markers in urine [\[63\]](#page-22-2). Likewise, another clinical trial demonstrated better control of atherosclerosis markers in individuals on the MedDiet consuming OO; however, only the group whose main fat source was VOO decreased urinary 8-OHdG levels [\[64\]](#page-22-3). In general, the MedDiet and OO intervention groups showed positive results in the regulation of DNA repair genes. In fact, a low expression of the polymerase k gene, which encodes a protein that is responsible for replicating damaged DNA, was identified in the MedDiet and OO intervention groups [\[64\]](#page-22-3). Thus, this dietary pattern showed a favorable effect on blood pressure, insulin sensitivity, and lipid levels exerted in people with a high cardiovascular risk [\[65–](#page-22-4)[69\]](#page-22-5).

Modifications in dietary habits towards the Mediterranean pattern have been proposed as positive in the reduction in oxidative DNA damage in cancer patients [\[70\]](#page-22-6). In addition, several studies indicate an inverse association between adherence to the MedDiet and very prevalent neoplasms such as breast cancer, colorectal cancer (CRC), bladder cancer, or prostate cancer [\[71](#page-22-7)[–73\]](#page-22-8). Specifically, individuals with CRC and high adherence to the MedDiet presented a lower histological grade and a lower frequency of synchronous adenomas compared to oncologic patients with low adherence to the MedDiet. In fact, elevated values of glutathione peroxidase (with antioxidant properties) and decreased 8-OHdG values were detected in patients with higher adherence to the MedDiet [\[74\]](#page-22-9).

Regarding neurodegenerative diseases, a decreased risk of Alzheimer's disease (AD) was associated with the direct effect of MedDiet components on the pathogenesis of AD [\[75,](#page-22-10)[76\]](#page-22-11). The beneficial effects of this neurodegenerative disorder were primarily due to EVOO consumption. Oleuropein (one of the main phenolic components of green olive pulp) reduced Poly ADP-ribose polymerase (PARP)-1 activation, protecting neuronal cells from oxidative damage. Furthermore, Zhang et al. observed an age-dependent increase in 8-OHdG in human brain tissues, particularly in those belonging to people with AD [\[77\]](#page-22-12). In people with mild cognitive impairment, an increase in 8-OHdG was detected in certain brain areas, and this could be interpreted as a biomarker predictive of AD pathogenesis [\[78\]](#page-22-13). Thus, when evaluating oxidative DNA damage according to 8-OHdG values in patients with mild cognitive impairment who consumed EVOO for one year, 8-OHdG levels were reduced [\[79\]](#page-22-14).

Overall, it seems that the Mediterranean pattern has potentially positive effects on genomic instability.

4.1.2. Telomere Attrition

Telomeres are nucleotide sequences at the ends of chromosomes, which progressively shorten with age. They are known as biomarkers of aging. Chronic oxidative stress is associated with telomere attrition [\[80\]](#page-22-15). In addition, inflammation stimulates telomere shortening by increasing the rate of hematopoietic stem cell replication to supply the leukocyte demand generated in the inflammatory process [\[81\]](#page-22-16). Although telomere length is inherited, it is influenced by external factors such as smoking, obesity, and a sedentary lifestyle. In turn, telomere shortening increases the risk of CVD, cancer, and mortality, especially at early ages [\[82](#page-22-17)[,83\]](#page-23-0).

Greater adherence to the MedDiet was associated with longer telomeres and greater telomerase activity [\[84](#page-23-1)[,85\]](#page-23-2), which could relate to lower levels of inflammation and oxidative stress [\[84,](#page-23-1)[86\]](#page-23-3). In their meta-analysis, Canudas et al. established a positive association between MedDiet adherence and telomere length in the blood cells except for samples taken from men [\[87\]](#page-23-4). However, it should be noted that the included studies were cross-sectional and, therefore, did not establish causality. Nevertheless, this result can be contrasted with the two prospective studies conducted to date [\[88,](#page-23-5)[89\]](#page-23-6). The work of Meinilä et al. prospectively studied 1046 Dutch subjects with a mean age of 61 years over a period of 10 years and did not establish an association between the male sex and adherence to the MedDiet. In the same study, women had faster telomere shortening [\[89\]](#page-23-6). On this subject, the only clinical trial found to date was by García-Calzón et al.; this was a sub-analysis of the PREDIMED-Navarra study, which evaluated 520 individuals, comprising 55% women aged 55–80 years. Three intervention groups were randomly assigned to a control or low-fat diet and there were two MedDiet groups, one supplemented with EVOO and the other with mixed nuts. In the cross-sectional analysis at the baseline, better adherence to the MedDiet was associated with longer telomeres only in women. However, assignment to the MedDiet-nuts group was associated with a higher risk of telomere shortening 5 years after the intervention, with no differences for the group supplemented with EVOO [\[88\]](#page-23-5). There was no consistent explanation for these findings, although cohort variables such as ethnicity, genetics, or sex could be related. In the work of Gu et al., a positive association was found between MedDiet adherence and telomere length in non-Hispanic white people [\[90\]](#page-23-7). This association was not found in African Americans or Hispanics [\[90\]](#page-23-7). However, in another subanalysis of the PREDIMED-Navarra study, it was shown that the Pro12Ala polymorphism in the peroxisome proliferator-activated receptor γ 2 (PPAR γ 2) gene interacted with MedDiet to prevent telomere shortening [\[91\]](#page-23-8).

Regarding PUFAs, omega-3 was better than omega-6 since attenuated telomere shortening was shown in a cohort of people over 65 years of age with mild cognitive impairment supplemented with this kind of fatty acid [\[92\]](#page-23-9). Similarly, the Breitas–Simoes clinical trial demonstrated attenuation in telomere attrition in cognitively unimpaired elderly people who supplemented their usual diet with walnuts (a source of omega-3 PUFAs) for 2 years [\[36\]](#page-21-0). However, LA supplementation in a population of 299 elderly people with recent myocardial infarction was associated with increased leukocyte telomere length, and no relationship was established with other PUFAs [\[93\]](#page-23-10). In general, it can be concluded that it is beneficial to follow a healthy diet in which PUFAs are present.

Telomere length has been linked to several types of cancer. The finding of short telomeres in CRC suggests that telomere shortening contributes to tumorigenesis and the genetic instability of premalignant cells. In fact, severely short telomeres have been shown to cause senescence in healthy cells or genomic instability in premalignant cells [\[83](#page-23-0)[,94](#page-23-11)[,95\]](#page-23-12). Indeed, altered telomere length homeostasis and unrepaired DNA damage were considered key in the onset, progression, and prognosis of CRC [\[95\]](#page-23-12). In this regard, the MedDiet could be useful primarily as a preventive therapy since several studies show the association between good adherence to the MedDiet and telomere preservation [\[84,](#page-23-1)[91\]](#page-23-8).

In addition, telomere shortening has been related to neurodegenerative diseases [\[96](#page-23-13)[,97\]](#page-23-14). In fact, Guo et al. suggested that telomere length has a causal effect on the risk of AD due to oxidative stress and inflammation [\[97\]](#page-23-14). Furthermore, telomere shortening has been associated with cognitive impairment, amyloid pathology, and tau protein hyperphosphorylation in AD [\[96\]](#page-23-13). In this regard, the oleuropein aglicone from OO inhibits protein aggregation in AD [\[98\]](#page-23-15). Further, the Mediterranean diet could prevent telomere shortening after oxidative damage thanks to antioxidant-rich vegetables such as nuts and seeds [\[99\]](#page-23-16). Resveratrol (an antioxidant present in grapes) generated neuroprotective effects in a study on a mouse model of AD [\[100\]](#page-23-17).

In summary, increased adherence to the MedDiet could attenuate telomere attrition. However, these beneficial effects could be limited to specific subgroups of the population. Further studies are needed to resolve the controversies raised.

4.1.3. Epigenetic Effects

Epigenetics encompass inherited genomic changes that occur in the absence of direct DNA damage. Young, healthy individuals maintain compact chromatin and the optimal epigenetic regulation of biological processes. However, aging favors the accumulation of chromatin damage, which compromises genome integrity and alters cellular function [\[101\]](#page-23-18). DNA methylation (mDNA) is considered one of the best-known epigenetic markers. mDNA is used as a clock for the calculation of biological age [\[102\]](#page-23-19). The long interspersed nuclear element (LINE-1) was used as a marker of global mDNA because it is the most common repetitive sequence in the human genome, and 1/3 of mDNA occurs in LINE-1 [\[103,](#page-23-20)[104\]](#page-23-21). Specifically, the hypomethylation of LINE-1 occurs during aging and is associated with multiple cancers and CVD [\[105\]](#page-23-22). In addition, oxidative stress plays a role in mDNA via the carbon cycle [\[42\]](#page-21-4), so the more ROS there is, the greater the DNA damage; finally, DNA undergoes hypomethylation to defend itself [\[106\]](#page-23-23). In this context, a diet rich in antioxidants, such as the Mediterranean diet, is proposed as an epigenetic diet.

To date, several studies have linked MedDiet adherence to LINE-1 hypomethylation. Specifically, there are two clinical trials that establish an inverse relationship between these two factors [\[107,](#page-23-24)[108\]](#page-23-25). In the work of Agodi et al., women of childbearing age with low adherence to the MedDiet, particularly those with lower fruit intake, were at higher risk of LINE-1 hypomethylation [\[109\]](#page-24-0). This finding may be related to the fact that fruit is a folate-rich food, and folate is an important donor of methyl groups. Other more specific components of the MedDiet, such as nuts and EVOO, are able to induce methylation changes in several peripheral white blood cell genes related to diabetes, inflammation, and signal transduction, which may have potential health benefits [\[110\]](#page-24-1). Similarly, the MedDiet could contribute to delaying the process of carcinogenesis that is related to DNA methylation processes since several studies have shown lower LINE-1 methylation levels in different types of tumors, such as CRC or breast cancer [\[111](#page-24-2)[,112\]](#page-24-3).

Regarding CVD, the review published by Muka et al. supported the suggestion that global mDNA, according to repeated LINE-1 hypomethylation, is inversely associated with CVD risk independently of established cardiovascular risk factors [\[105\]](#page-23-22). In general terms, LINE-1 hypomethylation is linked to an unfavorable cardiovascular risk profile due to its association with diabetes, obesity, lower HDL-cholesterol levels, elevated total cholesterol levels, and inflammation [\[113](#page-24-4)[,114\]](#page-24-5).

However, the expression of RNA (ribonucleic acid) contributes to the epigenetic modulation of gene expression that alters cellular functionality. In this context, the overexpression of miR-155-3p has been linked to carcinogenesis [\[115\]](#page-24-6). In fact, Ping Li et al. detected an

increased expression of miR-155-3p in relation to CRC tumor growth [\[116\]](#page-24-7). In addition, Let-7b, a regulator of histone H2B ubiquitination, showed a probable antitumor effect [\[117\]](#page-24-8). In respect of this line, Li et al. showed that let-7b-3p inhibited tumor growth and metastasis in lung cancer, correlating with the low expression of this molecule with poor prognosis in lung adenocarcinoma patients [\[118\]](#page-24-9). As a preventive therapeutic strategy, good adherence to the MedDiet could decrease the expression of miR-155-3p and increase the expression of let-7b-3p, improving the risk and evolution of cancer [\[119\]](#page-24-10).

In relation to neurodegenerative diseases associated with aging, epigenetic modifications play important roles in AD [\[120\]](#page-24-11). Specifically, mDNA is a highly controlled mechanism involving nicotinamide adenine dinucleotide (NAD)-dependent deacetylase Sirtuin 1 (SIRT1) that prevents altered methylation [\[121\]](#page-24-12). In this regard, Luccarini et al. demonstrated that quercetin and other EVOO polyphenols activate the SIRT1 pathway with suggestive therapeutic and preventive benefits [\[122](#page-24-13)[,123\]](#page-24-14).

Therefore, it is likely that there is an age-related disease—epigenetic interaction that may benefit from a healthy Mediterranean pattern diet.

4.1.4. Proteostasis

Proteostasis, or protein homeostasis, refers to the work of a complex network of pathways that are essential for cell function and viability, ensuring the appropriate concentration, folding, and interactions of proteins from synthesis to degradation [\[5\]](#page-19-4). Specifically, chaperones and two proteolytic systems (the ubiquitin-proteasome and the lysosome-autophagy system) are responsible for the maintenance of proteostasis [\[124\]](#page-24-15). The progressive loss of cellular protein homeotasis is detected during aging, and proteomes that are more stable or more resistant to alterations are found in the longest-lived species [\[125\]](#page-24-16).

The age-related deterioration of proteostasis affects chaperone functionality due to the cellular energy deficit that is inherent to senescence [\[126\]](#page-24-17). In addition, autophagy and the proteasome are altered with age, influencing proteotasis [\[127](#page-24-18)[,128\]](#page-24-19). In this regard, experimental interventions that enhanced autophagy-activating properties were associated with healthier aging [\[129\]](#page-24-20). Dietary habits could be beneficial for the optimization of proteostasis. Indeed, MedDiet polyphenols such as resveratrol can directly activate autophagy [\[129\]](#page-24-20). Likewise, oleuropein has been highlighted as an autophagy enhancer through a protein mammalian target of rapamycin (mTOR) and the adenosine monophosphate-activated protein kinase (AMPK)-dependent mechanism [\[130\]](#page-24-21). Furthermore, the antioxidant properties of these MedDiet components could attenuate the excess of oxidized proteins associated with senescence and age-related diseases [\[131\]](#page-24-22).

Age-related diseases such as neurodegenerative diseases (in particular, AD and Parkinson´s disease, PD) have been related to the deterioration of proteostasis [\[132\]](#page-24-23). In AD, hyperphosphorylated tau protein was aggregated abnormally and created insoluble neurofibrillary tangles, which were involved in neurodegeneration [\[133](#page-24-24)[,134\]](#page-25-0). In addition, amyloid-beta (Aβ) peptide accumulated and formed plaques that damaged neuronal cells [\[135\]](#page-25-1). In relation to PD, aggregates of insoluble α -synuclein protein fibrils were present in the neurons of people with PD and were neurotoxic [\[136\]](#page-25-2). Overall, this loss of proteostasis in neurodegenerative diseases is closely linked to inflammation and cellular senescence [\[13](#page-20-5)[,137\]](#page-25-3). Shannon et al. proposed the MedDiet as a mechanism by which to prevent neurodegeneration due to its modulating effect on protein homeostasis [\[30\]](#page-20-20). By enhancing autophagy, OO could mitigate the effects of toxic vascular agents, favoring the prevention of late-onset AD [\[29\]](#page-20-19). Specifically, oleocanthal could reduce tau protein polymerization [\[138\]](#page-25-4). The interaction between oleocanthal and tau proteins induces a tau rearrangement that may explain the antifibrillogenic effect of oleocanthal [\[139\]](#page-25-5). Thus, oleocanthal intervention in mice has been shown to increase the yield in the activity of blood–brain barrier transporter proteins that remove Aβ peptides (P-glycoprotein and low-density lipoprotein receptor-related protein 1). Thus, the percentage of degraded $Aβ$ peptides was higher in the treated group [\[140\]](#page-25-6). The beneficial effect of oleocanthal in mice was extensible to human cell lines since an improvement in $\Lambda\beta$ transport by $\Lambda\beta$ -secreting

cells was observed after administering oleocanthal for 72 h [\[141\]](#page-25-7). Further, resveratrol reduced β-secretase activity and Aβ-peptide aggregation in AD murine models and acted as a neuroprotectant in AD and PD [\[142\]](#page-25-8).

In relation to CVD, alterations in protein homeostasis and stability in the proteome can influence healthy cardiac aging [\[143\]](#page-25-9). The increased accumulation of misfolded protein aggregates was detected in CVD by the downregulation of the HSP70 chaperone in vascular tissue during aging [\[144,](#page-25-10)[145\]](#page-25-11). In general terms, decreased proteasome activity was detected in the atherogenic plaques of aged rats and elderly patients [\[124](#page-24-15)[,146\]](#page-25-12). In this context, oleuropein raised the rates of proteosome-mediated degradation in human fibroblast cultures, their lifespan was increased, and senescence was delayed by 15% [\[147\]](#page-25-13). A diet rich in EVOO also increased messenger RNA levels of the autophagy marker LC3 in older rats compared to rats that were fed using other sources of dietary fat [\[148\]](#page-25-14).

Urra et al. identified altered proteostasis as a hallmark of cancer. The hypermetabolism of cancer cells and the overexpression of oncogenes were related to ER stress. In response to this stress, the unfolded protein response (UPR) was generated [\[149\]](#page-25-15). Therefore, the UPR functioned as an adaptive mechanism during cancer progression [\[149\]](#page-25-15). Nevertheless, the activation of UPR at different stages of cancer evolution experienced a complex progression [\[149\]](#page-25-15). The role of the UPR during the early phase of cancer development prevented oncogene-induced malignant progression [\[150\]](#page-25-16). Cells surviving oncogene-induced apoptosis elevated UPR activation levels. Thus, in later stages of cancer progression, the UPR modified part of its function and contributed to tumor growth, aggressiveness, microenvironment adaptation, and resistance to treatment [\[149\]](#page-25-15). In fact, in human biopsies of breast cancer, lymphoma, or multiple myeloma, x-box-binding protein-1 (XBP-1), a protein that signals the activation of the UPR complex, was highly expressed and correlated with poor prognosis [\[151–](#page-25-17)[154\]](#page-25-18). In addition, the IRE1 α -XBP1 UPR signaling pathway was linked to the promotion of triple human breast [\[155\]](#page-25-19), prostate [\[156\]](#page-25-20), and hepatocellular cancers [\[157\]](#page-25-21). In this sense, MedDiet could prevent the perpetuation of this process. The work of Yubero-Serrano et al. observed that the MedDiet decreased the expression of genes relating to endoplasmic erythrocyte stress, such as XBP1 [\[158\]](#page-25-22). In addition, OO MUFAs "colonized" cell lipid membranes, which was associated with reduced susceptibility to ER stress and apoptosis [\[159\]](#page-26-0). In addition, secoiridoids such as oleuropein from EVOO favored the turnover of misfolded proteins in the cell by promoting compensatory UPR activity. Nevertheless, these bioactive compounds were able to inhibit the growth of highly aggressive mammary malignant cells [\[160](#page-26-1)[,161\]](#page-26-2).

Overall, the prevention of neurodegenerative, CVD, and oncological diseases may be of great interest given the proteostasis-activating role played by some components of the MedDiet. Nevertheless, further studies are needed to clarify the role of specific MedDiet nutrients in the prevention of these pathologies.

4.1.5. Nutrient-Sensing Pathways

These pathways are signaling systems responsible for detecting the availability of cellular resources that are essential for maintaining functionality, growth, and reproduction and thus participate in the aging process [\[162\]](#page-26-3). In fact, it is hypothesized that the proper regulation of these signaling pathways may extend lifespan and decrease the risk of agerelated diseases [\[5\]](#page-19-4).

In particular, the dysregulation of some nutrient-sensing pathways such as insulin/insulinlike growth factor-1 (IIS), mTOR, AMPK, and sirtuins was linked to an increased risk of non-inheritable diseases associated with age [\[5](#page-19-4)[,163\]](#page-26-4). The IIS pathway is the most conserved aging control pathway in evolution, and it regulates glucose metabolism [\[5\]](#page-19-4). Multiple genetic polymorphisms or mutations that reduce the intensity of signaling in the IIS pathway were associated with an increased lifespan in mice [\[164\]](#page-26-5). Paradoxically, however, in physiological or pathological aging, growth hormone (GH) and insulin-like growth factor-1 (IGF-1) levels decrease [\[165\]](#page-26-6). This phenomenon is explained as a defensive response of the organism to minimize cell growth and metabolic response in a scenario of systemic damage [\[166\]](#page-26-7).

Nevertheless, low concentrations of IGF-1 at the peripheral level are associated with an increased risk of type 2 diabetes mellitus, CVD, sarcopenia, osteoporosis, and frailty in elderly humans [\[167](#page-26-8)[,168\]](#page-26-9). This has been explained by the decrease in insulin sensitivity that appears with age [\[168\]](#page-26-9). However, reduced IIS pathway signaling stimulates longevityrelated Forkhead Box O (FOXO) proteins, which improve mitochondrial function and promote glucose metabolism through lipid oxidation [\[169\]](#page-26-10). In this regard, Calnan et al. proposed to control FOXO activity by up-regulating the expression of genes involved in resistance to metabolic stress and apoptosis to promote healthy aging [\[170\]](#page-26-11).

Together with the IIS pathway, mTOR is the main accelerator of aging [\[5\]](#page-19-4). The mTOR pathway identifies high concentrations of amino acids, and its regulation has been associated with healthy aging. Specifically, mTOR is a kinase that is formed by two protein complexes: mTORC1 and mTORC2 [\[171\]](#page-26-12). In mice with low levels of mTORC1 or S6K1 (the ribosomal S6 protein kinase 1) activity (main substrate of mTORC1) and normal levels (the ribosomal S6 protein kinase 1) activity (main substrate of mTORC1) and normal levels of mTORC2, increased life expectancy was detected [\[172,](#page-26-13)[173\]](#page-26-14). The hyperstimulation of this pathway was frequently observed in diseases associated with aging, such as cancer [\[174](#page-26-15)[,175\]](#page-26-16), AD [\[176\]](#page-26-17), and diabetes [\[177\]](#page-26-18). However, the inhibition of mTOR activity also [174,175], AD [176], and diabetes [177]. However, the inhibition of mTOR activity also showed undesirable effects, including wound healing problems, insulin resistance, and showed undesirable effects, including wound healing problems, insulin resistance, and testicular degeneration in mice [\[178\]](#page-26-19). testicular degeneration in mice [178].

> In contrast to the IIS and mTOR pathways that sense nutrient abundance and favor In contrast to the IIS and mTOR pathways that sense nutrient abundance and favor anabolism, AMPKs, and sirtuins sense nutrient scarcity and promote energy catabolism. In anabolism, AMPKs, and sirtuins sense nutrient scarcity and promote energy catabolism. fact, the activation of AMPK promoted longevity by the inhibition of one mTOR complex, mTORC1 [\[179\]](#page-26-20), and one of the sirtuins, SIRT1, could have triggered peroxisome proliferator-activated receptor gamma 1-alpha (PGC-1α) coactivator upon deacetylation [\[180](#page-26-21)[,181\]](#page-26-22). This coactivator is involved in the transcription of antioxidant genes and is a key regulator of mitochondrial biogenesis [\[182\]](#page-26-23) (Figure 2).

Figure 2. Dysregulated nutrient sensing pathways. Overview of the somatotropic axis, involving **Figure 2.** Dysregulated nutrient sensing pathways. Overview of the somatotropic axis, involving GH and the IIS pathway. Molecules that promote aging are shown in yellow, while molecules with antianti-aging properties are shown in light green. Red lines in lines in $\frac{1}{2}$ in the slowed pathways and $\frac{1}{2}$ in $\frac{1}{2}$ aging properties are shown in light green. Red lines indicate inhibited or slowed pathways and blue
... lines indicate activated pathways that together promote healthy aging. GH, growth hormone; IGF-1, $\,$ insulin-like growth factor-1; mTOR, protein mammalian target of rapamycin; AMPK, adenosine monophosphate-activated protein kinase; FOXO, Forkhead Box; PGC-1α, peroxisome proliferatoractivated receptor gamma 1-alpha; SIRT-1, sirtuine-1.

In animal models, a dietary restriction has been shown to promote healthy aging me-diated by nutrient-sensing pathways [\[183\]](#page-26-24). However, in humans, less stringent and more realistic interventions are being proposed [\[163\]](#page-26-4). In this regard, MedDiet, characterized by low-moderate protein intake, low glycaemic index, and polyphenol-rich foods, may be an alternative [\[184\]](#page-26-25). Polyphenol-rich foods activate the AMPK and sirtuin pathways, while mTOR is inhibited and autophagy is stimulated [\[185](#page-26-26)[,186\]](#page-27-0). Interestingly, oleocanthal exhib-ited potent neuroprotective and antitumoral properties by inhibiting mTOR activity [\[187\]](#page-27-1). Antiproliferative effects were observed in certain breast cancer cell lines, although this effect was not fully clear in other neoplasms, such as CRC and cervical cancer, probably due to the lower expression of the mTOR pathway in these two malignant processes [\[141\]](#page-25-7). Moreover, oleuropein and HT showed an antidiabetic effect in addition to attenuating oxidative stress in rats with diabetes mellitus [\[188,](#page-27-2)[189\]](#page-27-3). Further, adherence to the MedDiet may be of interest to patients with AD. In these cases, neurons presented a reduced activity of glucotransporters, GLUT1 and GLUT3, leading to altered insulin signaling [\[190\]](#page-27-4), altering fasting blood glucose along with lipid metabolism dysfunction [\[191\]](#page-27-5). In addition, low IGF-1 values were detected with a low-glycemic-index diet compared with a highglycemic-index diet [\[192\]](#page-27-6), which could attenuate the signaling intensity of the IIS pathway favoring longevity.

However, in the study of Levine et al., people aged from 50 to 65 years who ingested more protein (above 20% of the daily caloric intake) presented higher mortality, and 25% of deaths were associated with cancer [\[193\]](#page-27-7). Those individuals with moderate intake recorded a lower IGF-1 concentration by down-modulating the activity of the IIS and mTOR pathways [\[192](#page-27-6)[,194\]](#page-27-8). In addition, high protein intake was linked to an increased risk of type 2 diabetes, obesity, and CVD [\[195\]](#page-27-9). Nevertheless, the protein quality is important. In this context, in murine models, methionine restriction showed a prolonged life expectancy and protection against multiple chronic diseases, particularly cancer [\[196\]](#page-27-10). Other essential branched-chain amino acids (in poultry, dairy, and eggs) such as leucine, isoleucine, and valine were identified as key in the regulation of insulin sensitivity via mTOR [\[197\]](#page-27-11). In fact, Fontana et al. reported that selective reduction in the dietary intake of branched-chain amino acids improved glucose tolerance, β-cell metabolic stress, and body composition [\[198\]](#page-27-12).

Accordingly, there is robust evidence that some components of the MedDiet have properties that favor healthy aging, especially EVOO, so it would be interesting to further explore this through its action on nutrient signaling pathways.

4.1.6. Mitochondrial Dysfunction

Mitochondria are cellular organelles that are responsible for producing much of the adenosine triphosphate (ATP) necessary for cell survival and modulating signaling toward apoptosis. Much of the total oxygen taken up by cells is metabolized in the electron transport chain located in the inner membrane of the mitochondria. The formation of ROS at this level represents potential intracellular damage to the organelle. In general, mitochondria are altered by oxidative damage with aging, and their deterioration is a consequence of the tissue's inability to repair or eradicate the damage [\[199\]](#page-27-13). In the elderly, an excess of dysfunctional mitochondria can result in less energy in the form of ATP and more ROS than in the younger populations [\[5\]](#page-19-4). Indeed, this mitochondrial imbalance has been associated with age-related neurological diseases such as PD and AD [\[200,](#page-27-14)[201\]](#page-27-15), cancer [\[202\]](#page-27-16), and metabolic syndrome [\[203\]](#page-27-17).

Mitochondrial dysfunction is also implicated in the pathophysiology of type 2 diabetes mellitus, obesity, dyslipidemia, and CVD [\[203\]](#page-27-17). These pathologies are usually described as metabolic syndrome. Impaired mitochondrial energy metabolism is considered the main cause of metabolic syndrome [\[203\]](#page-27-17). Specifically, in type 2 diabetes mellitus, high glucose levels increase ROS production with consequent damage to the mitochondria [\[204\]](#page-27-18) and inhibition of the IIF pathway, which favors lipid accumulation leading to metabolic disorders [\[205–](#page-27-19)[210\]](#page-27-20). Aging, the alteration of mitochondrial antioxidant effects, and genetic factors promoting insulin resistance are the main causes of many metabolic diseases [\[203\]](#page-27-17). In this sense, the MedDiet with MUFAs and antioxidants may have a beneficial effect. In fact, different experimental models have demonstrated that components of MedDiet, such as polyphenols, plant-derived compounds, and PUFAs, could correct mitochondrial dysfunction and improve mitochondrial metabolism [\[211\]](#page-27-21). Indeed, the PREDIMED study demonstrated the cardiometabolic benefits [\[212\]](#page-28-0).

Varela-López et al. demonstrated that OO has a favorable effect on the mitochondrial structure and function of aged rats [\[213\]](#page-28-1). Along the same lines, MUFA-rich OO played a key role in adapting the lipid profile of mitochondrial membranes to provide resistance against oxidative damage and dysfunction associated with aging [\[141\]](#page-25-7). In fact, the predominant dietary fat source impacted mitochondrial membrane biochemistry by modifying the fatty acid composition profile and electron transport systems [\[214\]](#page-28-2). In addition, PUFA sources favored the oxidation of mitochondrial membranes compared to saturated or MUFA sources [\[215\]](#page-28-3). In this sense, Ochoa et al. measured the mitochondrial fatty acid profile, catalase activity, and hydroperoxide levels in the liver, heart, and skeletal muscle of Wistar rats when supplemented with different fat sources (sunflower oil versus OO) [\[216\]](#page-28-4). Those with higher OO intake had more MUFAs in their mitochondrial membranes, and those fed mainly with sunflower oil had more omega-6 PUFAs. These variations were consistent with the levels of oxidative damage, i.e., those fed OO had fewer hydroperoxides in their body tissues compared to those who were fed sunflower oil. Therefore, a diet rich in OO generated membranes with fewer PUFAs, which attenuated an increase in agerelated lipid peroxidation in post-mitotic tissues such as the heart and skeletal muscle [\[217\]](#page-28-5). Furthermore, in the liver (the prototype of regenerative tissue) and heart, a greater increase in catalase activity—essential for antioxidant defense in relation to life expectancy—was observed in rats who were fed a diet rich in OO [\[218,](#page-28-6)[219\]](#page-28-7).

However, HT and oleuropein reduce oxidative stress and optimize mitochondrial function [\[220](#page-28-8)[,221\]](#page-28-9). HT improves neuronal inflammation and may delay the development of AD [\[222\]](#page-28-10). In fact, mitochondrial dysfunction is critical in the early stages of AD and PD, and the antioxidant power of wine polyphenols can protect organelles [\[200,](#page-27-14)[223,](#page-28-11)[224\]](#page-28-12). Quercetin and procyanidins (the main polyphenols in wine) can decrease ROS and improve the cell viability of neuronal and astrocytic cell lines [\[225,](#page-28-13)[226\]](#page-28-14). Specifically, quercetin decreases ROS production through the overexpression of the AMPK/SIRT1 signaling pathway [\[227\]](#page-28-15). Resveratrol improved the antioxidant status in PD rats and reduced dopamine loss [\[228\]](#page-28-16). However, the mechanism by which resveratrol protects mitochondrial function and homeostasis is not fully understood but has been investigated for its potential applications in the treatment of age-related diseases [\[229\]](#page-28-17). Overall, several polyphenols are present in wine and individually carry out promising mitochondrial protection. However, Kurin et al. demonstrated greater antioxidant potency when combining several polyphenols with respect to their individual activity [\[230\]](#page-28-18). Thus, light to moderate wine intake in humans can favor the expression of antioxidant enzymes in the blood [\[231\]](#page-28-19). Although most of the studies evaluating the impact of MedDiet components on mitochondrial dysfunction were conducted in animals, they are also consistent with those conducted in humans.

Fish oil, a high-omega-3 PUFA source, has a protective effect on age-related mitochondrial dysfunctions similar to that observed for OO. Afshordel et al. demonstrated that fish oil supplementation for 21 days restored the concentration of omega-3 PUFA derivatives, improving mitochondrial function and consequent ATP synthesis in the brains of older mice [\[232\]](#page-28-20). The benefits of omega-3 PUFAs in neurodegenerative diseases have been observed in preclinical studies, while most of the controlled clinical trials have not met expectations. In this regard, initiating clinical work prematurely in the course of the disease and increasing the durability of the study may be helpful in obtaining the expected outcomes [\[233\]](#page-28-21). In addition, in cerebral ischemia, DHA showed beneficial results, and the reduction in stroke events was related to less disruption of the blood–brain barrier, less brain edema, and less inflammatory cell swelling [\[234\]](#page-28-22).

Cell membranes with high concentrations of peroxidized PUFAs, which are typical of aging, led to apoptosis and growth inhibition [\[202\]](#page-27-16). In fact, the main products of lipid peroxidation are toxic and mutagenic aldehydes such as malondialdehyde (MDA) and 4-hydroxynonenal/4-hydroxy-2-nonenal (HNE). In addition, elevated MDA values were recorded in the plasma and blood of patients with breast, lung, and ovarian cancer [\[235](#page-28-23)[–239\]](#page-29-0). In the work of Li YP et al., HNE promoted breast cancer cell growth and angiogenesis [\[240\]](#page-29-1). In this sense, the antioxidant capacity of OO could attenuate peroxidation, preventing

or defending against the activation of carcinogenesis [\[217\]](#page-28-5). In fact, EVOO could have a beneficial effect on breast cancer risk [\[241\]](#page-29-2).

In conclusion, mitochondrial dysfunction and oxidative damage play a crucial role in the pathogenesis of aging and longevity-related diseases. Together, different components of the MedDiet, such as OO, PUFA, and red wine, contribute to the maintenance of mitochondrial function. However, more studies are needed to evaluate the synergistic effect of MedDiet components on this hallmark.

4.1.7. Cellular Senescence

Senescent cells often exhibit irreversible DNA damage, leading to cell cycle arrest. In addition, these cells produce a proinflammatory secretome or SASP that contributes to aging [\[242](#page-29-3)[,243\]](#page-29-4). Cellular senescence is associated with other features of aging, including mitochondrial dysfunction, autophagy disorders, altered nutrient signaling, and epigenetic effects [\[5,](#page-19-4)[244\]](#page-29-5). Overall, age increases the number of senescent cells, which increases the likelihood of age-related diseases [\[245\]](#page-29-6). This hallmark defends tissues from damaged and potentially oncogenic cells [\[5\]](#page-19-4). However, it requires progenitor cells with a regenerative capacity to compensate for the cellular deficit associated with aging [\[5\]](#page-19-4).

Similar to DNA damage, exaggerated mitogenic (senescence-inducing) signaling is the other stress that is strongly associated with senescence. There are important cellular mechanisms that defend an organism against oncogenic or mitogenic alterations, such as the p16 INK4a/Rb and p19 ARF/p53 pathways [\[246\]](#page-29-7). Indeed, p16 INK4a (and, to a lesser extent, p19 ARF) levels correlate with age in most of the tissues analyzed [\[247](#page-29-8)[,248\]](#page-29-9). In a meta-analysis performed by Jeck et al., the INK4a/ARF genomic locus was found to be the locus most closely linked to age-associated pathologies, including several types of CVD, diabetes, glaucoma, and AD [\[249\]](#page-29-10).

Senolytic therapies, including dietary intervention, may delay or prevent cellular aging [\[250](#page-29-11)[,251\]](#page-29-12). In fact, the MedDiet has demonstrated senolytic properties thanks to various food components. For example, nuts and certain vegetables seem to prevent the accumulation of senescent cells [\[53](#page-21-13)[,55](#page-21-15)[,252\]](#page-29-13). Further, the phenolic components of EVOO (oleocanthal or oleuropein), with antioxidant and anti-inflammatory effects, could play a relevant role in neurodegenerative diseases such as AD [\[253](#page-29-14)[–255\]](#page-29-15). Specifically, tauopathy was associated with astrocyte or microglia senescence [\[256,](#page-29-16)[257\]](#page-29-17) and oleocanthal-reduced tau protein polymerization [\[138\]](#page-25-4). In addition, the Aβ peptide was identified as a potent inducer of cellular senescence [\[258–](#page-29-18)[262\]](#page-30-0). The intervention with oleocanthal in mice showed increased performance in the activity of blood–brain barrier transporter proteins in charge of eliminating Aβ peptides (P-glycoprotein and low-density lipoprotein receptor-related protein 1), with the percentage of degraded Aβ peptides being higher in the treated group. The beneficial effect of olocanthal in mice may be extensible to human cell lines [\[141\]](#page-25-7).

Resveratrol can delay or prevent senescence, as proven in human cell models (mesenchymal stem cells) [\[263](#page-30-1)[,264\]](#page-30-2). Further, quercetin, when associated with Dasatinib (BCL family apoptotic inhibitors) in people with diabetic nephropathy, reduced the number of senescent cells in human adipose tissue [\[265\]](#page-30-3). However, there is clinical evidence of an association between age-related cardiac pathologies and the release of SASP components by senescent cells. The heart diseases that have been studied are heart failure, ischemia and myocardial infarction, and cardiotoxicity secondary to cancer chemotherapy [\[266\]](#page-30-4). However, the specific role of senescent cells in these conditions is unclear, and existing information is contradictory [\[266\]](#page-30-4). Presumably, the presence of maintained (and not transient) cellular senescence promotes deleterious effects in cardiac disease, such as the functional impairment of cardiac progenitor cells [\[267\]](#page-30-5). In addition, this hallmark can impair adult cardiomyocyte proliferation [\[267\]](#page-30-5). In this case, the MedDiet could reduce the production of proinflammatory substances since resveratrol inhibits the nuclear transcription of factor κB (NF-κB), which is essential for the genesis of SASP [\[268](#page-30-6)[,269\]](#page-30-7). In addition, quercetin (associated with dasatinib) can facilitate programmed senescent cell death by inhibiting the PI3K-AKT pathway [\[251\]](#page-29-12). Furthermore, the antioxidant properties of the

MedDiet can act on ROS by attenuating DNA damage and decreasing the excess number of senescent cells [\[42,](#page-21-4)[68\]](#page-22-18).

Regarding cancer, cellular senescence can protect tissues against tumorigenesis [\[270\]](#page-30-8). Indeed, anti-cancer therapies (chemotherapy or radiotherapy) induce senescence in cancer cells [\[271\]](#page-30-9). However, the persistence of therapy-induced senescent cells can be detrimental. Overall, a strategy that eliminates these persistent cells in the long term to minimize tumor progression and avoid adverse effects is of interest. Specifically, quercetin, in combination with dasatinib in aged mice, eliminated senescent cells and optimized cardiovascular function and survival [\[269,](#page-30-7)[272\]](#page-30-10). However, in the elderly, senescent cells represent a high percentage of the cellular reserve, and this could jeopardize tissue structural integrity or affect vascular endothelial cells, leading to liver damage and the fibrosis of perivascular tissue with important repercussions on health [\[273,](#page-30-11)[274\]](#page-30-12).

In summary, the MedDiet may be useful as a senolytic tool, although more molecular studies are needed to clarify the synergistic action of the various anti-aging foods on tissue senescent cells.

4.1.8. Stem Cell Exhaustion

Stem cells in humans have the capacity for self-renewal and differentiation in various tissues [\[275\]](#page-30-13). A decrease in the regenerative capacity of tissues is characteristic of aging [\[5\]](#page-19-4). This is a consequence of intrinsic and extrinsic causes that generate a vulnerable scenario for stem cell preservation in all human tissues [\[276\]](#page-30-14). This scenario includes reduced cell-cycle activity in longer-lived stem cells [\[277\]](#page-30-15), accumulated DNA damage [\[277\]](#page-30-15), the overexpression of p16 INK4a (cell cycle inhibitory) proteins [\[278\]](#page-30-16), and telomere shortening [\[279](#page-30-17)[,280\]](#page-30-18). In addition, an optimal balance between the activation of cell regeneration and the inactivation of the cellular process is essential for proper cell function. Therefore, circuits that safeguard progeroid stem cells' quiescence, such as INK4 induction and IGF-1 depletion, are essential [\[5\]](#page-19-4). Overall, the deterioration in stem cell regenerative capacity and its lack of control contribute to aging and increase the risk of age-related diseases. In the case of hematopoietic tissue, the regenerative potential diminishes with age, and immunosenescence occurs. This phenomenon can often favor subclinical inflammation, which contributes to the development of age-related diseases [\[281\]](#page-30-19).

Some components of the MedDiet, in combination or separately, have shown benefits in attenuating stem cell depletion. In the work of Cesari et al., adherence to the MedDiet in very old people was associated with increased numbers of endothelial progenitor cells [\[282\]](#page-30-20). Endothelial stem cells are essential for maintaining vascular homeostasis and renewing injured vascular cells [\[283,](#page-30-21)[284\]](#page-30-22). Thus, the MedDiet intervenes in the early stages of the atherosclerotic process, which could have important implications for the early prevention of CVD [\[30\]](#page-20-20). Particularly, oleuropein and oleacein play a protective role in the senescence of endothelial progenitor cells induced by angiotensin II (a key pathological factor in hypertension) [\[285\]](#page-30-23). In addition, oleopurein stimulates osteoblastogenesis and the mineralization of the cellular matrix and inhibits bone resorption. An increase in serum osteocalcin levels was detected in elderly patients on the MedDiet enriched with VOO [\[286,](#page-30-24)[287\]](#page-31-0). Overall, the risk of osteoporosis was reduced, including a bone protective effect in a two-year intervention study derived from PREDIMED [\[286\]](#page-30-24).

In relation to carcinogenesis, an increased incidence of hematological malignancies has been associated with immunosenescence [\[288\]](#page-31-1). In this sense, multiple polyphenols in OO preserve hematopoietic stem cells and their differentiation [\[141,](#page-25-7)[289\]](#page-31-2). Further, there is an increased risk of aggressive and invasive skin cancers (melanoma and basal cell carcinoma) associated with aging and exposure to ultraviolet radiation [\[290,](#page-31-3)[291\]](#page-31-4). Both circumstances induce the premature senescence of fibroblasts and the activation of fibroblastto-myofibroblast transitions [\[290](#page-31-3)[,292\]](#page-31-5). These findings could favor fibrosis along with the loss of skin elasticity and an increased risk of oncogenesis. As an anti-aging and preventive therapy against hyperplasia and skin cancers, resveratrol can induce anti-inflammatory and

antioxidant changes [\[293\]](#page-31-6). However, further studies are needed to conclude with certainty the benefits of resveratrol.

AD is included in the group of SASP or pathologies secondary to an altered secretome (growth factors, ROS, cytokines, and metalloproteinases). In fact, late astrocytes generate increased secretion of SASP factors that deregulate physiological functions. Dysfunctional astrocytes produce a chronic inflammatory response and pathologies of the central nervous system. In this context, altered synaptic plasticity, blood–brain barrier impairment, and glutamate excitoxicity with decreased neural stem cell proliferation have been observed [\[257](#page-29-17)[,294\]](#page-31-7). As a preventive option to avoid this pathological condition, studies with DHA plus EPA supplementation have shown beneficial effects in patients with mild AD [\[295](#page-31-8)[,296\]](#page-31-9). Therefore, MedDiet may be useful, as omega-3 PUFA-rich components are frequently ingested. Similarly, this consumption has shown a protective effect against the incidence of AD [\[296\]](#page-31-9). Flavonoids from cereals, vegetables, fruits, and OO had potentially beneficial effects, including free-radical scavenging, anti-inflammatory effects, and protection against $\text{A}β$ neurotoxicity [\[297](#page-31-10)[–299\]](#page-31-11). Further, the positive influence of polyphenols on cerebrovascular health is considered relevant, including lipoprotein oxidation, platelet aggregation, and endothelial cell reactivity [\[300\]](#page-31-12).

In conclusion, MedDiet can be useful in the prevention of prevalent age-related diseases thanks to foods such as OO, which can palliate or slow down the deterioration of stem cells. However, more studies are needed to contrast this approach, as there are controversies on some points.

4.1.9. Altered Intercellular Communication

Cellular coordination is essential for proper functionality. Different soluble molecules allow intercellular communication: cytokines, chemokines, growth factors, and neurotransmitters [\[301\]](#page-31-13). Multiple intercellular, endocrine, and neuronal pathways undergo alterations in the aging process. Specifically, neurohormonal signaling is altered in aging as inflammatory reactions increase and immune system reactivity to pathogens and precancerous cells decreases [\[5\]](#page-19-4). Aging is related to inflammation, one of the most relevant intercellular communication processes, and this association is multicausal. Among the causes are the accumulation of proinflammatory tissue damage by the increased secretion of cytokines and adipokines, excess ROS, immunosenescence, increased activation of the NK-κB pathway, changes in the gut microbiome and intestinal permeability, and altered autophagy response [\[302–](#page-31-14)[306\]](#page-31-15). Thus, the elderly presents a low-grade systemic inflammatory phenomenon that favors age-associated chronic diseases and increases the risk of mortality [\[307\]](#page-31-16).

The MedDiet demonstrated anti-inflammatory effects in relation to different markers, such as interleukin 6 (IL-6) or tumor necrosis factor-alpha (TNFα) [\[38](#page-21-2)[,308](#page-31-17)[,309\]](#page-31-18). In this context, multiple in vitro studies have positioned OO polyphenols as sources of healthpromoting properties [\[141\]](#page-25-7). Zhang et al. highlighted the anti-inflammatory properties of HT mediated by the suppression of cyclooxygenase-2 (COX-2) and the expression of inducible nitric oxide synthase [\[310\]](#page-31-19). HT also reduces superoxide ions and inhibits the excess of important inflammatory mediators in humans, prostaglandin E2, probably due to the reduced expression of COX-2 $[311]$. Thus, it is intuited that the ability of polyphenols to regulate the production of proinflammatory molecules may have a salutary impact on older people. In addition to HT, other polyphenols such as oleocanthal and oleouropein inhibit TNFα-induced matrix metalloproteinase 9 through the anti-inflammatory pathway shared with ibuprofen (a non-steroidal anti-inflammatory drug that inhibits COX-2) [\[253,](#page-29-14)[312\]](#page-31-21). Further, the flavonoid apigenin can act as an immunomodulator and principal regulator of TNF α in lipopolysaccharide-induced inflammation [\[313\]](#page-32-0).

Atherosclerosis is an inflammatory disease and is closely linked to endothelial dysfunction. Dell'Agli et al. showed that OO polyphenols slow down the expression of proatherogenic molecules through the inactivation of NF-kB in endothelial cells [\[314\]](#page-32-1). These polyphenols were also potent against ROS, prevented the oxidative damage of genetic materials, and enhanced the antioxidant power of endothelial cells [\[315\]](#page-32-2). In this sense, the study of Meza-Miranda et al. determined that a breakfast based on VOO favorably regulated the pathophysiological mechanisms of premature atherosclerosis in endothelial cells [\[316\]](#page-32-3). Camargo et al. demonstrated that a breakfast rich in OO could curb the expression of proinflammatory genes and favor a less harmful inflammatory profile [\[317\]](#page-32-4). In addition, EVOO enhances the anti-inflammatory effect of high-density lipoprotein cholesterol and increases age-related antiatherogenic activity [\[318\]](#page-32-5).

Moreover, the inflammation of neuronal cells is a fundamental mechanism in neurodegeneration. Proinflammatory immune-mediated mechanisms are essential in the pathogenesis and progression of AD and PD [\[319\]](#page-32-6). MedDiet has been associated with a lower risk of developing AD and PD [\[320\]](#page-32-7). In addition, the positive effects of omega-3 PUFAs on AD are due to their antioxidant and anti-inflammatory power. However, these PUFAs tend to oxidize. Therefore, the antioxidant capacity of polyphenols may be of interest as adjuvants [\[29\]](#page-20-19). This is more supportive of adherence to the Mediterranean diet than the individual intake of a certain food that is commonly present in the MedDiet. Further, fruits and vegetables (including in the MedDiet), with great antioxidant and anti-inflammatory properties, reduce the risk of PD [\[321,](#page-32-8)[322\]](#page-32-9).

Regarding carcinogenesis, senescent tumor cells produce SAPS that control the senescence of neighboring cells [\[323\]](#page-32-10). Initially, secretome factors could prevent tumor progression or eradicate malignant cells except in situations where there is acute aggression to SASP [\[323\]](#page-32-10). At this point, some polyphenols, such as quercetin and phytosine, exhibit senolytic properties through the inhibition of the PI3K-AKT pathway [\[324,](#page-32-11)[325\]](#page-32-12). Regarding the most prevalent cancers today, MedDiet, when applied to CRC, has shown a protective effect thanks to different compounds [\[326\]](#page-32-13). Among them, oleuropein is promising as a protective agent against colitis-associated CRC, and in mice with induced CRC and inflammatory cytokines such as TNF-α, IL-6, and COX-2 decreased [\[327](#page-32-14)[,328\]](#page-32-15). Further, oleocanthal could reduce the risk of inflammatory bowel diseases and CRC [\[329,](#page-32-16)[330\]](#page-32-17). Therefore, the inflammatory response is regulated by OO polyphenols as they inhibit NF-κB, and this implies the lower expression of different interleukins and COX-2. This microenvironment hinders tumor proliferation [\[331](#page-32-18)[,332\]](#page-32-19).

Therefore, the evidence supports the suggestion that the MedDiet has anti-inflammatory properties and, consequently, may positively influence the aging phenotype.

5. Aging, Mediterranean Diet and Microbiome

5.1. Microbiome

The gastrointestinal tract is colonized by an array of microorganisms, including bacteria, viruses, fungi, and protozoa. These coexist symbiotically with enterocytes without being identified by the immune system as pathogens [\[333\]](#page-32-20). These microorganisms make up the microbiota, which consists of a total of 52 different phyla and up to 35,000 species of bacteria, mainly Actinobacteria, Bacteroidetes, Firmicutes, and Proteobacteria [\[334\]](#page-32-21).

The intestinal microbiota has its origin in the placenta, with low levels of nonpathogenic bacteria, mostly Bacteroidetes, and Firmicutes. After birth, the intestine of the newborn and infant is rapidly colonized. Factors such as the type of delivery (vaginal or caesarean) or the type of feeding (breastfeeding or formula feeding) are determinants of the microbiota [\[333\]](#page-32-20). During the first three years of life, the microbiota has low diversity. After the third year, the microbiota is similar to that of the adult stage. With aging, changes occur in the morphology and function of the microbiota. Thus, after 65 years of age, the microbiota experiences a decrease in Firmicutes and Bifidobacterium, with an increase in diversity for Clostridium [\[28\]](#page-20-18).

The changes produced throughout life could alter the diversity of the microbiota, giving rise to metabolic and inflammatory alterations and causing the appearance of conditions such as inflammatory bowel disease or irritable bowel syndrome, among others [\[335\]](#page-32-22). Moreover, the microbiota has not only been linked to diseases of the gastrointestinal tract but also to other diseases such as obesity, diabetes, CVD, or cancer [\[336\]](#page-32-23). In fact, since

the concept of the "gut–brain axis" was created, gut microbiota has also been linked to neurodegenerative diseases [\[337\]](#page-32-24).

5.2. Interplay between Aging and the Microbiome

The mechanisms by which the microbiota change with age are not fully understood. It is known that, in aging, physiological changes occur, such as alterations in dentition or decreased digestion and absorption, and modifications of lifestyle conditions, such as hospitalization or nursing homes. These modifications could be responsible, in part, for changes in diet and thus for the nutritional status of the elderly [\[338\]](#page-33-0). Moreover, in aging, especially in respect of frailty, there is usually a reduction in the amount and variety of food, which leads to the appearance of malnutrition [\[339\]](#page-33-1). Diet seems to be one of the pillars of changes in the microbiota. The microbiota may modulate changes in aging-related to innate immunity, cognitive function, and sarcopenia, which are components of frailty syndrome [\[340\]](#page-33-2). In fact, recent studies have suggested that loss of the gut microbiota is more related to age-associated frailty than to chronological age [\[341\]](#page-33-3).

During the transition from adult to elderly, the main changes in the intestinal microbiota occur. Microbial diversity decreases compared to young adults [\[342\]](#page-33-4). In elderly centenarians, the microbiota consists mostly of Bacteroidetes and Firmicutes. However, in comparison with young adults, there are changes in subgroups such as Firmicutes, with an increase in Bacilli and a decrease in Clostridium. In addition, there is also an increase in Protebacteria [\[343\]](#page-33-5). Intestinal dysbiosis mainly involves changes in the abundance of commensal bacteria, also including some that function as opportunistic pathogens. The importance of the dysbiosis phenomenon is that it stimulates the excretion of endotoxins, i.e., amyloid and microbial lipopolysaccharides, to promote intestinal wall permeability and increase the peripheral circulation of proinflammatory cytokines [\[344\]](#page-33-6).

In the ELDERMET study, the microbiota was studied in elderly people living in a community or living in long-stay homes in Ireland [\[338\]](#page-33-0). In the first group, microbiota configurations were more affected by antibiotic use than the microbiota of individuals in long-stay residences. However, this first group presented greater recovery after antibiotic use. The second group showed a loss of microbial components associated with ill health and a gain in altered microbiota associated with aging [\[338\]](#page-33-0). These findings on the relationship between microbiota, diet, and health status are supported by Claesson et al. [\[342\]](#page-33-4). They demonstrated, through an analysis of the composition of the fecal microbiota separated in 178 elderly subjects, that a change in diet associated with a transfer to a nursing home caused a change in the composition of intestinal bacteria, which correlated with nutritional status, inflammatory markers, comorbidity, and frailty [\[342\]](#page-33-4). Thus, the aging process and other environmental factors may alter the composition of the microbiota and contribute to the development of chronic low-grade inflammation [\[7\]](#page-20-0).

Therefore, maintaining diversity in the microbiota appears key to maintaining health status and preventing frailty.

5.3. Mediterranean Diet and Microbiome: Health Status and Disease

Diet has a major impact on the biology of the gut microbiota [\[345\]](#page-33-7). Some nutrients have effects on the structure, function, and secretion of metabolites of the gut microbiota that can modulate immune functions and multiple metabolic and inflammatory pathways [\[346](#page-33-8)[,347\]](#page-33-9). Emerging evidence is showing that adherence to the MedDiet promotes beneficial effects on the microbiota, favoring microbial diversity mainly in the colon, and it is associated with a reduction in Clostridium and an increase in Bacteroidetes and Firmicutes [\[28\]](#page-20-18) (Figure [3\)](#page-16-0).

Figure 3. Influence of diet in gut microbiome related to aging. **Figure 3.** Influence of diet in gut microbiome related to aging.

5.3.1. Mediterranean diet, Microbiome and CVD, Obesity and Diabetes 5.3.1. Mediterranean Diet, Microbiome and CVD, Obesity and Diabetes

Variations in the microbiota have been linked to the development of diseases. The Variations in the microbiota have been linked to the development of diseases. The microbiota can be modified through diet. In fact, polyphenols from the MedDiet play a microbiota can be modified through diet. In fact, polyphenols from the MedDiet play a key role in the microbiota. These compounds can reach the gut microbiota and modify the bacterial population and its metabolism. In this respect, it has been reported that administration of polyphenols in rats, specifically resveratrol and curcumin, was the administration of polyphenols in rats, specifically resveratrol and curcumin, was associated with alterations in the Bacteroidetes and Clostridium groups of bacteria, thus associated with alterations in the Bacteroidetes and Clostridium groups of bacteria, thus providing metabolic benefits in glycemic control [\[348\]](#page-33-10). Additionally, the high content and bioavailability of fiber in the MedDiet (two times higher than in a Western diet) have a material of the Me have beneficial effects on the cardiovascular system of older adults. These benefits could
. be due, in part, to changes in the microbiota. Fiber appears to have a positive impact on the composition of the gut microbiota, increasing the number of beneficial bacteria, inhibiting the growth of pathogens, and reducing atherogenic serum cholesterol in the microbiome. It also prevents glucose intolerance by reducing postprandial hyperglycemia
through the contract of the co through the formation of a viscous layer around the small intestine, which slows down the change of a viscous layer around the small intestine, which slows down the chyme transition [\[7\]](#page-20-0). This, in turn, increases the thickness of the aqueous layer through
which solutes word were to weak the extensity weakened at a direction developed in which solutes must pass to reach the enterocyte membrane, leading to a decrease in glucose in glucose in glucose in glucose in glucose in glucose in the enteror of the enteror solution in glucose in the enteror solution of the enterocyte blood and resulting in a decreased absorption of glucose, lipids, and amino and amino acids [\[7\]](#page-20-0). High fiber intake has been found to promote modifications of the acids [7]. High fiber intake has been found to promote modifications of the gut microbiota gut microbiota with an increase in Bacteroidetes (in particular, *Bacteroides acidifaciens*), gat increased with an increase in Bacteroidetes (in particular, *Bacteroides acidifaciens*), which produce high levels of short-chain fatty acids, including acetate, butyrate, and high levels of short-chain fatty acids, including acetate, butyrate, and propionate [349]. propionate [\[349\]](#page-33-11). Some of the beneficial effects of these metabolites are thought to be mediated by binding to specific G-protein-coupled receptors expressed on enteroendocrine and immune cells $[349]$. Conversely, poor adherence to the MedDiet was associated with an increase in l-Ruminococcus and Streptococcus bacteria and an increased concentration of trimethylamine N-oxide (TMAO) in urine. Compared to the Western diet, the MedDiet has significantly lower contents of choline and L-carnitine (present in egg, cheese, and red meat), and the production of TMAO by the microbiota has been shown to be lower [\[350\]](#page-33-12). This could reduce the risk of CVD, independent of the presence of cardiovascular factors. Zhu et al. concluded that an elevated level of TMAO could also be involved in the pathogenesis of obesity and type 2 diabetes mellitus, as it induces vascular inflammation and a prothrombotic effect by increasing platelet hypersensitivity to multiple agonists [\[351\]](#page-33-13). Indeed, the review by Cornejo-Pareja et al. concludes that the increase in fat mass in obese patients is not only due to more efficient energy uptake but that the microbiota glucose in the enterocyte blood and resulting in a decreased absorption of glucose, lipids,

is involved in changes in endotoxemia, intestinal permeability, insulin resistance, the hormonal environment, the expression of lipogenesis regulatory genes, interaction with bile acids and changes in the proportion of brown adipose tissue [\[352\]](#page-33-14).

5.3.2. Mediterranean Diet, Microbiome and Cancer

Numerous epidemiological studies have supported the importance of lifestyle factors and exposure to known or suspected carcinogens in the development of cancer. In fact, it is estimated that 30–35% of cancer risk factors are associated with diet, physical activity, and/or energy imbalance [\[353\]](#page-33-15), and 15–20% of cancers are caused by infectious agents [\[354\]](#page-33-16). The microbiota that inhabits our body can be considered an environmental factor to which we are continuously exposed throughout life. However, the underlying mechanisms by which the MedDiet decreases the risk of cancer are not entirely clear [\[355\]](#page-33-17). In the diet–microbiota interaction, it has been observed that many dietary and digestive components are metabolized by bacteria in the gastrointestinal tract, leading to tumor suppressor metabolites and putative oncometabolites [\[356,](#page-33-18)[357\]](#page-33-19). As an example, the excessive consumption of red meat, present in the Western diet, is a risk factor for CRC and other cancers by several mechanisms, including some that are dependent on intestinal bacteria. Elevated levels of protein intake can lead to an increase in certain types of bacteria, including Bacteroides and Firmicutes. These ferment amino acids into N-nitroso compounds, which induce DNA alkylation and mutations in the host [\[357\]](#page-33-19). Proteobacterias that encode nitroreductases and nitrate reductases are also related to this process, which is strongly associated with inflammation [\[358\]](#page-33-20).

In addition, in the process of the digestion of saturated fat associated with red meat consumption, approximately 5% of the primary bile acids escape from the enterohepatic circulation and reach the colon, where they are converted by bacteria into secondary bile acids. Primary cholic acid is converted to secondary deoxycholic acid by certain bacteria, including *Clostridium scindens*. Secondary deoxycholic acid functions as a tumor promoter by disrupting cell membranes to release arachidonic acid, which is converted by cyclooxygenase-2 and lipooxygenase into prostaglandins and ROS that trigger inflammation and DNA damage [\[359\]](#page-33-21). By contrast, the dietary fiber present in the MedDiet is fermented by certain types of colonic bacteria, such as Clostridium groups IV and XIVa, into short-chain fatty acids. Butyrate, which is one of the most abundant short-chain fatty acids, is the main source of energy for colonocytes and is involved in the prevention of CRC. It has been observed that butyrate probably exerts its tumor-suppressive properties through multiple mechanisms. Butyrate epigenetically regulates the expression of genes that are involved in apoptosis and cell proliferation apoptosis [\[116\]](#page-24-7). It also acts as a ligand for certain G-protein-coupled receptors due to its involvement in tumor suppression [\[360\]](#page-33-22). Both mechanisms are believed to be important for butyrate's ability to induce regulatory T cells. In addition, butyrate helps maintain the epithelial barrier function, which is important for preventing inflammation. Other components of the MedDiet that are related to cancer prevention are polyphenols. Ellagitannins are polyphenols found in nuts and berries. When they reach the intestine, they are modified by the microbiota and transformed into different compounds. Urolithin is one of the most studied products, and it has been shown that it can be absorbed by the enterohepatic circulation and transported by the blood and thus distributed to different tissues. It has anticarcinogenic effects through the inhibition of the Wnt signaling pathway, which could have a protective effect against CRC [\[361\]](#page-33-23).

Thus, our diet dictates whether the microbiota produces metabolites that exacerbate or enhance tumor progression [\[362\]](#page-33-24).

5.3.3. Mediterranean Diet, Microbiome and Neurological Diseases

Regarding neurodegenerative diseases, better cognitive functions and a lower risk of dementia have been associated with higher adherence to the MedDiet. The PREDIMED study demonstrated a modest beneficial effect of adherence to the MedDiet for 4–6 years on cognitive functions in cognitively healthy adults at high risk of CVD, especially in the domains of global cognition, memory, and executive function [\[363](#page-33-25)[,364\]](#page-33-26). By contrast, no benefit on cognitive function was reported after 1 year of the MedDiet in older adults in the NU-AGE trial. However, participants with higher adherence to the MedDiet demonstrated better global cognition and episodic memory compared with those that have low adherence [\[365\]](#page-34-0). These benefits appear to be related to certain components of MedDiet (omega-3 fatty acids, antioxidants, and polyphenols) as they may inhibit neuroinflammation associated with AD and other degenerative diseases [\[366\]](#page-34-1).

Changes in the microbiota could also be involved in the pathogenesis of these diseases by initiating and perpetuating neuroinflammatory processes. In this respect, a study demonstrated the existence of the brain microbiota in cerebral blood vessels through micrographs of the human brain [\[367\]](#page-34-2). These bacteria and gut-derived toxins appear to compromise the integrity of the blood–brain barrier and could contribute to early neuroinflammatory changes by stimulating microglia and hindering amyloid clearance [\[368](#page-34-3)[,369\]](#page-34-4). In addition, microbial amyloid and circulating liposaccharides activate innate resistance receptors, such as the Toll-like receptor and the receptor for advanced glycation end products, to increase proinflammatory signaling and to promote chronic neuroinflammation and progressive neurodegeneration, especially in sensitive brain regions such as the hippocampus [\[368](#page-34-3)[,370\]](#page-34-5). Moreover, the microbiota has also been linked to other disorders, such as epilepsy. In fact, one study in epileptic patients found that antibiotic treatment reduced seizure frequency by 10% [\[371\]](#page-34-6).

Nearly 60% of the variation in gut microbiota is attributable to diet [\[372\]](#page-34-7); therefore, modulation of the gut microbiota through diet could be an effective approach for reducing the inflammation associated with neurological diseases. Preliminary data have shown positive associations between the MedDiet and increased numbers of beneficial species of the microbiota, e.g., Bacteroidetes, and their short-chain fatty acid metabolites, which have anti-inflammatory effects [\[373](#page-34-8)[,374\]](#page-34-9). However, only a few studies have evaluated dietary patterns and gut microbiota, most of them being observational, which prevents establishing causality [\[375](#page-34-10)[,376\]](#page-34-11). Further research is therefore needed to understand the complex relationships between the gut microbiota and cognitive health and whether dietinduced effects are mediated by alterations in gut microbiota.

All of this is important because increasing evidence suggests that the reprogramming of gut microbial functions through long-term adherence to healthier diets can influence physiological responses to nutrients and other features of host biology that are critical to promoting health and longevity [\[377\]](#page-34-12). Thus, the modification of the microbiota through MedDiet could benefit the evolution and prognosis of these diseases.

6. Methods

Interventional studies involving animals or humans, and other studies that require ethical approval, must list the authority that provided approval and the corresponding ethical approval code. A comprehensive search of the literature published in PubMed from November 2022 was conducted to identify articles relating to MedDiet, microbiota, aging, and frailty. Search strategies were based on the following search terms: MedDiet, polyphenols, omega-3 PUFAs, healthy aging, hallmarks of aging, telomere length, microbiota, oxidative stress, mitochondrial function, inflammation, cellular senescence, anti-senescence compounds, frailty, and sarcopenia. A selection of articles published in English providing original human research, observational prospective and retrospective studies, randomized controlled trials, reviews, and meta-analyses were included.

In addition, we considered case series, single-case reports, editorials, research or original articles, letters to the editor, comments (on an article or from the editor), responses (to a comment, letter, or article), corrections, short reports, short communications, perspectives, opinions, and discussions. Priority was given to the largest studies and to the strongest available evidence and most recent studies.

7. Conclusions

There is an increasingly aging global population. However, the way to achieve healthy aging has not yet been fully elucidated. The loss of function and frailty syndrome associated with aging increases the vulnerability of the elderly and their propensity to disease. There are different molecular pathways or hallmarks involved in aging that bring us closer to understanding the deterioration associated with the senescence process, such as genomic instability, telomere attrition, epigenetic effects, proteostasis, nutrient-sensing pathways, mitochondrial dysfunction, cellular senescence, stem cell depletion, and altered intercellular communication. Likewise, microbiota disturbances seem to play a relevant role in frailty in the elderly.

It has been shown that MedDiet promotes healthy aging, increasing the life expectancy of the population. This review has shown that MedDiet positively influences the molecular pathways that determine age. Consequently, MedDiet has been associated with a lower risk of age-related diseases, mainly CVD, neurodegenerative, and oncological diseases. Therefore, further evidence of the beneficial effects of this dietary pattern on human health and longevity has been provided. However, most studies do not evaluate the impact of the Mediterranean diet pattern as a whole on the hallmarks of aging but rather its individual components, especially certain bioactive components. Certainly, there are some clinical trials exploring the role of the Mediterranean diet (mostly PREDIMED substudies), but they focus on specific dietary supplementation with nuts or EVOO. Therefore, it would be useful to evaluate the pattern as a whole without special emphasis on these more studied components. In addition, more quality studies on the MedDiet and the prevention of frailty and disease in aging are needed, as many studies are observational, and causality cannot be determined.

Overall, more research is needed to provide a better understanding of the mechanism of action of MedDiet on aging. However, at present, MedDiet could be recommended as a baseline anti-aging therapy to prevent frailty and maintain functionality until the later stages of life, as the benefits of MedDiet on human health present robust evidence.

Author Contributions: Study design: M.C.A.-L. and V.C.-B., study conduct: M.C.A.-L. and V.C.-B., data collection: M.C.A.-L. and V.C.-B., data interpretation: M.C.A.-L., V.C.-B., B.G.-F., C.G.-F. and M.M.-T., drafting of the manuscript: M.C.A.-L. and V.C.-B., reviewing the manuscript and approving final version of manuscript: All authors have read and agreed to the published version of the manuscript.

Funding: Victoria Contreras-Bolívar was funded by a postdoctoral fellowship from the Junta de Andalucia (RH-0141-2020) and C.G.-F. was funded by a postdoctoral fellowship from the Instituto de Salud Carlos III (CD20/00022).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. World Health Organization. *Envejecimiento y La Salud*; World Health Organization: Geneva, Switzerland, 2022.
- 2. Beard, J.R.; Officer, A.; De Carvalho, I.A.; Sadana, R.; Pot, A.M.; Michel, J.P.; Lloyd-Sherlock, P.; Epping-Jordan, J.E.; Peeters, G.M.E.E.; Mahanani, W.R.; et al. The World Report on Ageing and Health: A Policy Framework for Healthy Ageing. *Lancet* **2016**, *387*, 2145–2154. [\[CrossRef\]](http://doi.org/10.1016/S0140-6736(15)00516-4) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26520231)
- 3. Niccoli, T.; Partridge, L. Ageing as a Risk Factor for Disease. *Curr. Biol.* **2012**, *22*, R741–R752. [\[CrossRef\]](http://doi.org/10.1016/j.cub.2012.07.024) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22975005)
- 4. Rudnicka, E.; Napierała, P.; Podfigurna, A.; Męczekalski, B.; Smolarczyk, R.; Grymowicz, M. The World Health Organization (WHO) Approach to Healthy Ageing. *Maturitas* **2020**, *139*, 6–11. [\[CrossRef\]](http://doi.org/10.1016/j.maturitas.2020.05.018)
- 5. López-Otín, C.; Blasco, M.A.; Partridge, L.; Serrano, M.; Kroemer, G. The Hallmarks of Aging. *Cell* **2013**, *153*, 1194. [\[CrossRef\]](http://doi.org/10.1016/j.cell.2013.05.039) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23746838)
- 6. Vaiserman, A.M.; Koliada, A.K.; Marotta, F. Gut Microbiota: A Player in Aging and a Target for Anti-Aging Intervention. *Ageing Res. Rev.* **2017**, *35*, 36–45. [\[CrossRef\]](http://doi.org/10.1016/j.arr.2017.01.001) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28109835)
- 7. Sanchez-Morate, E.; Gimeno-Mallench, L.; Stromsnes, K.; Sanz-Ros, J.; Román-Domínguez, A.; Parejo-Pedrajas, S.; Inglés, M.; Olaso, G.; Gambini, J.; Mas-Bargues, C. Relationship between Diet, Microbiota, and Healthy Aging. *Biomedicines* **2020**, *8*, 287. [\[CrossRef\]](http://doi.org/10.3390/biomedicines8080287)
- 8. Brown, M.K.; Naidoo, N. The Endoplasmic Reticulum Stress Response in Aging and Age-Related Diseases. *Front. Physiol.* **2012**, *3*, 263. [\[CrossRef\]](http://doi.org/10.3389/fphys.2012.00263)
- 9. Morley, J.E.; Haren, M.T.; Rolland, Y.; Kim, M.J. Frailty. *Med. Clin. N. Am.* **2006**, *90*, 837–847. [\[CrossRef\]](http://doi.org/10.1016/j.mcna.2006.05.019)
- 10. Titus, S.; Li, F.; Stobezki, R.; Akula, K.; Unsal, E.; Jeong, K.; Dickler, M.; Robson, M.; Moy, F.; Goswami, S.; et al. Impairment of BRCA1-Related DNA Double-Strand Break Repair Leads to Ovarian Aging in Mice and Humans. *Sci. Transl. Med.* **2013**, *5*, 837–847. [\[CrossRef\]](http://doi.org/10.1126/scitranslmed.3004925)
- 11. Rose, M.R.; Flatt, T.; Graves, J.L.; Greer, L.F.; Martinez, D.E.; Matos, M.; Mueller, L.D.; Shmookler Reis, R.J.; Shahrestani, P. What Is Aging? *Front. Genet.* **2012**, *3*, 134. [\[CrossRef\]](http://doi.org/10.3389/fgene.2012.00134)
- 12. Giaimo, S.; D'Adda di Fagagna, F. Is Cellular Senescence an Example of Antagonistic Pleiotropy? *Aging Cell* **2012**, *11*, 378–383. [\[CrossRef\]](http://doi.org/10.1111/j.1474-9726.2012.00807.x) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22329645)
- 13. Childs, B.G.; Durik, M.; Baker, D.J.; Van Deursen, J.M. Cellular Senescence in Aging and Age-Related Disease: From Mechanisms to Therapy. *Nat. Med.* **2015**, *21*, 1424–1435. [\[CrossRef\]](http://doi.org/10.1038/nm.4000) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26646499)
- 14. Shimizu, I.; Yoshida, Y.; Katsuno, T.; Tateno, K.; Okada, S.; Moriya, J.; Yokoyama, M.; Nojima, A.; Ito, T.; Zechner, R.; et al. P53-Induced Adipose Tissue Inflammation Is Critically Involved in the Development of Insulin Resistance in Heart Failure. *Cell Metab.* **2012**, *15*, 51–64. [\[CrossRef\]](http://doi.org/10.1016/j.cmet.2011.12.006) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22225876)
- 15. Ryan, A.S. Insulin Resistance with Aging. *Sport. Med.* **2012**, *30*, 327–346. [\[CrossRef\]](http://doi.org/10.2165/00007256-200030050-00002) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/11103847)
- 16. Capurso, C.; Bellanti, F.; Buglio, A.L.; Vendemiale, G. The Mediterranean Diet Slows down the Progression of Aging and Helps to Prevent the Onset of Frailty: A Narrative Review. *Nutrients* **2020**, *12*, 35. [\[CrossRef\]](http://doi.org/10.3390/nu12010035) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31877702)
- 17. Chang, S.F. Frailty Is a Major Related Factor for at Risk of Malnutrition in Community-Dwelling Older Adults. *J. Nurs. Scholarsh.* **2017**, *49*, 63–72. [\[CrossRef\]](http://doi.org/10.1111/jnu.12258)
- 18. Crow, R.S.; Lohman, M.C.; Titus, A.J.; Cook, S.B.; Bruce, M.L.; Mackenzie, T.A.; Bartels, S.J.; Batsis, J.A. Association of Obesity and Frailty in Older Adults: NHANES 1999–2004. *J. Nutr. Health Aging* **2019**, *23*, 138–144. [\[CrossRef\]](http://doi.org/10.1007/s12603-018-1138-x)
- 19. Falsarella, G.R.; Gasparotto, L.P.R.; Barcelos, C.C.; Coimbra, I.B.; Moretto, M.C.; Pascoa, M.A.; Ferreira, T.C.B.R.; Coimbra, A.M.V. Body Composition as a Frailty Marker for the Elderly Community. *Clin. Interv. Aging* **2015**, *10*, 1661–1667. [\[CrossRef\]](http://doi.org/10.2147/CIA.S84632)
- 20. Ferriolli, E.; Pessanha, F.P.A.d.S.; Moreira, V.G.; Dias, R.C.; Neri, A.L.; Lourenço, R.A. Body Composition and Frailty Profiles in Brazilian Older People: Frailty in Brazilian Older People Study-FIBRA-BR. *Arch. Gerontol. Geriatr.* **2017**, *71*, 99–104. [\[CrossRef\]](http://doi.org/10.1016/j.archger.2017.03.008)
- 21. Xu, L.; Zhang, J.; Shen, S.; Hong, X.; Zeng, X.; Yang, Y.; Liu, Z.; Chen, L.; Chen, X. Association between Body Composition and Frailty in Elder Inpatients. *Clin. Interv. Aging* **2020**, *15*, 313–320. [\[CrossRef\]](http://doi.org/10.2147/CIA.S243211)
- 22. Clegg, A.; Young, J.; Iliffe, S.; Rikkert, M.O.; Rockwood, K. Frailty in Elderly People. *Lancet* **2013**, *381*, 752–762.
- 23. Keys, A.; Menotti, A.; Aravanis, C.; Blackburn, H.; Djordevič, B.S.; Buzina, R.; Dontas, A.S.; Fidanza, F.; Karvonen, M.J.; Kimura, N.; et al. The Seven Countries Study: 2,289 Deaths in 15 Years. *Prev. Med.* **1984**, *13*, 141–154. [\[CrossRef\]](http://doi.org/10.1016/0091-7435(84)90047-1)
- 24. Keys, A. Mediterranean Diet and Public Health: Personal Reflections. *Am. J. Clin. Nutr.* **1995**, *61*, 1321S–1323S. [\[CrossRef\]](http://doi.org/10.1093/ajcn/61.6.1321S) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/7754982)
- 25. Dominguez, L.J.; Di Bella, G.; Veronese, N.; Barbagallo, M. Impact of Mediterranean Diet on Chronic Non-Communicable Diseases and Longevity. *Nutrients* **2021**, *13*, 2028. [\[CrossRef\]](http://doi.org/10.3390/nu13062028)
- 26. Hernández, J.M.; Rentero, M.P.Z. Bioactive Compounds Contained in Mediterranean Diet and Their Effects on Neurodegenerative Diseases. In *Current Topics on Superfoods*; InTech: London, UK, 2018.
- 27. Shannon, O.M.; Stephan, B.C.M.; Minihane, A.M.; Mathers, J.C.; Siervo, M. Nitric Oxide Boosting Effects of the Mediterranean Diet: A Potential Mechanism of Action. *J. Gerontol. Ser. A Biol. Sci. Med. Sci.* **2018**, *73*, 902–904. [\[CrossRef\]](http://doi.org/10.1093/gerona/gly087) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/29684102)
- 28. Tosti, V.; Bertozzi, B.; Fontana, L. Health Benefits of the Mediterranean Diet: Metabolic and Molecular Mechanisms. *J. Gerontol. Ser. A Biol. Sci. Med. Sci.* **2018**, *73*, 318–326. [\[CrossRef\]](http://doi.org/10.1093/gerona/glx227) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/29244059)
- 29. Román, G.C.; Jackson, R.E.; Gadhia, R.; Román, A.N.; Reis, J. Mediterranean Diet: The Role of Long-Chain ω-3 Fatty Acids in Fish; Polyphenols in Fruits, Vegetables, Cereals, Coffee, Tea, Cacao and Wine; Probiotics and Vitamins in Prevention of Stroke, Age-Related Cognitive Decline, and Alzheimer Disease. *Rev. Neurol.* **2019**, *175*, 724–741. [\[CrossRef\]](http://doi.org/10.1016/j.neurol.2019.08.005) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31521398)
- 30. Shannon, O.M.; Ashor, A.W.; Scialo, F.; Saretzki, G.; Martin-Ruiz, C.; Lara, J.; Matu, J.; Griffiths, A.; Robinson, N.; Lillà, L.; et al. Mediterranean Diet and the Hallmarks of Ageing. *Eur. J. Clin. Nutr.* **2021**, *75*, 1176–1192. [\[CrossRef\]](http://doi.org/10.1038/s41430-020-00841-x)
- 31. Stevenson, E.J.; Shannon, O.M.; Minihane, A.M.; Adamson, A.; Burns, A.; Hill, T.; Sniehotta, F.; Muniz-Terrera, G.; Ritchie, C.W. NuBrain: UK Consortium for Optimal Nutrition for Healthy Brain Ageing. *Nutr. Bull.* **2020**, *45*, 223–229. [\[CrossRef\]](http://doi.org/10.1111/nbu.12429)
- 32. Mathers, J.C. Impact of Nutrition on the Ageing Process. *Br. J. Nutr.* **2015**, *113*, S18–S22. [\[CrossRef\]](http://doi.org/10.1017/S0007114514003237)
- 33. Aguilera, A.; Gómez-González, B. Genome Instability: A Mechanistic View of Its Causes and Consequences. *Nat. Rev. Genet.* **2008**, *9*, 204–217. [\[CrossRef\]](http://doi.org/10.1038/nrg2268)
- 34. Mahat, R.K.; Singh, N.; Gupta, A.; Rathore, V. Oxidative DNA Damage and Carotid Intima Media Thickness as Predictors of Cardiovascular Disease in Prediabetic Subjects. *J. Cardiovasc. Dev. Dis.* **2018**, *5*, 15. [\[CrossRef\]](http://doi.org/10.3390/jcdd5010015)
- 35. Basu, A.K. DNA Damage, Mutagenesis and Cancer. *Int. J. Mol. Sci.* **2018**, *19*, 970. [\[CrossRef\]](http://doi.org/10.3390/ijms19040970)
- 36. Freitas-Simoes, T.M.; Cofán, M.; Blasco, M.A.; Soberón, N.; Foronda, M.; Serra-Mir, M.; Roth, I.; Valls-Pedret, C.; Doménech, M.; Ponferrada-Ariza, E.; et al. Walnut Consumption for Two Years and Leukocyte Telomere Attrition in Mediterranean Elders: Results of a Randomized Controlled Trial. *Nutrients* **2018**, *10*, 1907. [\[CrossRef\]](http://doi.org/10.3390/nu10121907)
- 37. Kiokias, S.; Proestos, C.; Oreopoulou, V. Effect of Natural Food Antioxidants against LDL and DNA Oxidative Changes. *Antioxidants* **2018**, *7*, 133. [\[CrossRef\]](http://doi.org/10.3390/antiox7100133) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30282925)
- 38. Arpón, A.; Riezu-Boj, J.I.; Milagro, F.I.; Marti, A.; Razquin, C.; Martínez-González, M.A.; Corella, D.; Estruch, R.; Casas, R.; Fitó, M.; et al. Adherence to Mediterranean Diet Is Associated with Methylation Changes in Inflammation-Related Genes in Peripheral Blood Cells. *J. Physiol. Biochem.* **2016**, *73*, 445–455. [\[CrossRef\]](http://doi.org/10.1007/s13105-017-0552-6) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28181167)
- 39. Corella, D.; Ordovas, J.; Sorli, J.; Asensio, E.; Ortega, C.; Carrasco, P.; Portoles, O.; Coltell, O. Effect of the Mediterranean Diet on DNA Methylation of Selected Genes in the PREDIMED-Valencia Intervention Trial. *FASEB J.* **2015**, *29*, LB242. [\[CrossRef\]](http://doi.org/10.1096/fasebj.29.1_supplement.lb242)
- 40. Herrera-Marcos, L.V.; Lou-Bonafonte, J.M.; Arnal, C.; Navarro, M.A.; Osada, J. Transcriptomics and the Mediterranean Diet: A Systematic Review. *Nutrients* **2017**, *9*, 472. [\[CrossRef\]](http://doi.org/10.3390/nu9050472) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28486416)
- 41. Ungvari, Z.; Bagi, Z.; Feher, A.; Recchia, F.A.; Sonntag, W.E.; Pearson, K.; de Cabo, R.; Csiszar, A. Resveratrol Confers Endothelial Protection via Activation of the Antioxidant Transcription Factor Nrf2. *Am.J. Physiol Heart Circ. Physiol* **2010**, *299*, 18–24. [\[CrossRef\]](http://doi.org/10.1152/ajpheart.00260.2010)
- 42. Menezo, Y.J.R.; Silvestris, E.; Dale, B.; Elder, K. Oxidative Stress and Alterations in DNA Methylation: Two Sides of the Same Coin in Reproduction. *Reprod. BioMedicine Online* **2016**, *33*, 668–683. [\[CrossRef\]](http://doi.org/10.1016/j.rbmo.2016.09.006)
- 43. Valko, M.; Izakovic, M.; Mazur, M.; Rhodes, C.J.; Telser, J. *Role of Oxygen Radicals in DNA Damage and Cancer Incidence*; Kluwer Academic Publishers: Norwell, MA, USA, 2004; Volume 266, pp. 37–56.
- 44. Zhang, Y.; Yuan, F.; Wu, X.; Wang, M.; Rechkoblit, O.; Taylor, J.-S.; Geacintov, N.E.; Wang, Z. Error-Free and Error-Prone Lesion Bypass by Human DNA Polymerase κ in Vitro. *Nucleic Acids Res.* **2000**, *28*, 4138–4146. [\[CrossRef\]](http://doi.org/10.1093/nar/28.21.4138)
- 45. Kaneko, K.; Akuta, T.; Sawa, T.; Kim, H.W.; Fujii, S.; Okamoto, T.; Nakayama, H.; Ohigashi, H.; Murakami, A.; Akaike, T. Mutagenicity of 8-Nitroguanosine, a Product of Nitrative Nucleoside Modification by Reactive Nitrogen Oxides, in Mammalian Cells. *Cancer Lett.* **2008**, *262*, 239–247. [\[CrossRef\]](http://doi.org/10.1016/j.canlet.2007.12.007)
- 46. Urquiaga, I.; Strobel, P.; Perez, D.; Martinez, C.; Cuevas, A.; Castillo, O.; Marshall, G.; Rozowski, J.; Leighton, F. Mediterranean Diet and Red Wine Protect against Oxidative Damage in Young Volunteers. *Atherosclerosis* **2010**, *211*, 694–699. [\[CrossRef\]](http://doi.org/10.1016/j.atherosclerosis.2010.04.020) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/20451910)
- 47. Saieva, C.; Peluso, M.; Palli, D.; Cellai, F.; Ceroti, M.; Selvi, V.; Bendinelli, B.; Assedi, M.; Munnia, A.; Masala, G. Dietary and Lifestyle Determinants of Malondialdehyde DNA Adducts in a Representative Sample of the Florence City Population. *Mutagenesis* **2016**, *31*, 475–480. [\[CrossRef\]](http://doi.org/10.1093/mutage/gew012) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26961145)
- 48. Calcabrini, C.; De Bellis, R.; Mancini, U.; Cucchiarini, L.; Stocchi, V.; Potenza, L. Protective Effect of Juglans Regia L. Walnut Extract Against Oxidative DNA Damage. *Plant Foods Hum. Nutr.* **2017**, *72*, 192–197. [\[CrossRef\]](http://doi.org/10.1007/s11130-017-0609-2)
- 49. Erol, Ö.; Arda, N.; Erdem, G. Phenols of Virgin Olive Oil Protects Nuclear DNA against Oxidative Damage in HeLa Cells. *Food Chem. Toxicol.* **2012**, *50*, 3475–3479. [\[CrossRef\]](http://doi.org/10.1016/j.fct.2012.07.048) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22877972)
- 50. Vilahur, G.; Cubedo, J.; Padró, T.; Casaní, L.; Mendieta, G.; González, A.; Badimon, L. Intake of Cooked Tomato Sauce Preserves Coronary Endothelial Function and Improves Apolipoprotein A-I and Apolipoprotein J Protein Profile in High-Density Lipoproteins. *Transl. Res.* **2015**, *166*, 44–56. [\[CrossRef\]](http://doi.org/10.1016/j.trsl.2014.11.004) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25514506)
- 51. Chen, L.; Stacewicz-Sapuntzakis, M.; Duncan, C.; Sharifi, R.; Ghosh, L.; Van Breemen, R.; Ashton, D.; Bowen, P.E. Oxidative DNA Damage in Prostate Cancer Patients Consuming Tomato Sauce-Based Entrees as a Whole-Food Intervention. *JNCI J. Natl. Cancer Inst.* **2001**, *93*, 1872–1879. [\[CrossRef\]](http://doi.org/10.1093/jnci/93.24.1872)
- 52. Riso, P.; Pinder, A.; Santangelo, A.; Porrini, M. Does Tomato Consumption Effectively Increase the Resistance of Lymphocyte DNA to Oxidative Damage? *Am. J. Clin. Nutr.* **1999**, *69*, 712–718. [\[CrossRef\]](http://doi.org/10.1093/ajcn/69.4.712) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/10197573)
- 53. Riso, P.; Martini, D.; Visioli, F.; Martinetti, A.; Porrini, M. Effect of Broccoli Intake on Markers Related to Oxidative Stress and Cancer Risk in Healthy Smokers and Nonsmokers. *Nutr. Cancer* **2009**, *61*, 232–237. [\[CrossRef\]](http://doi.org/10.1080/01635580802425688)
- 54. Riso, P.; Martini, D.; Møller, P.; Loft, S.; Bonacina, G.; Moro, M.; Porrini, M. DNA Damage and Repair Activity after Broccoli Intake in Young Healthy Smokers. *Mutagenesis* **2010**, *25*, 595–602. [\[CrossRef\]](http://doi.org/10.1093/mutage/geq045)
- 55. Moser, B.; Szekeres, T.; Bieglmayer, C.; Wagner, K.H.; Mišík, M.; Kundi, M.; Zakerska, O.; Nersesyan, A.; Kager, N.; Zahrl, J.; et al. Impact of Spinach Consumption on DNA Stability in Peripheral Lymphocytes and on Biochemical Blood Parameters: Results of a Human Intervention Trial. *Eur. J. Nutr.* **2011**, *50*, 587–594. [\[CrossRef\]](http://doi.org/10.1007/s00394-011-0167-6)
- 56. Porrini, M.; Riso, P.; Oriani, G. Spinach and Tomato Consumption Increases Lymphocyte DNA Resistance to Oxidative Stress but This Is Not Related to Cell Carotenoid Concentrations. *Eur. J. Nutr.* **2014**, *41*, 95–100. [\[CrossRef\]](http://doi.org/10.1007/s003940200014) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/12111045)
- 57. Del Bo', C.; Riso, P.; Campolo, J.; Møller, P.; Loft, S.; Klimis-Zacas, D.; Brambilla, A.; Rizzolo, A.; Porrini, M. A Single Portion of Blueberry (*Vaccinium corymbosum* L.) Improves Protection against DNA Damage but Not Vascular Function in Healthy Male Volunteers. *Nutr. Res.* **2013**, *33*, 220–227. [\[CrossRef\]](http://doi.org/10.1016/j.nutres.2012.12.009) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23507228)
- 58. Riso, P.; Klimis-Zacas, D.; Del Bo', C.; Martini, D.; Campolo, J.; Vendrame, S.; Møller, P.; Loft, S.; De Maria, R.; Porrini, M. Effect of a Wild Blueberry (Vaccinium Angustifolium) Drink Intervention on Markers of Oxidative Stress, Inflammation and Endothelial Function in Humans with Cardiovascular Risk Factors. *Eur. J. Nutr.* **2013**, *52*, 949–961. [\[CrossRef\]](http://doi.org/10.1007/s00394-012-0402-9) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22733001)
- 59. Quiles, J.L.; Ochoa, J.J.; Ramirez-Tortosa, C.; Battino, M.; Huertas, J.R.; Martín, Y.; Mataix, J. Dietary Fat Type (Virgin Olive vs. Sunflower Oils) Affects Age-Related Changes in DNA Double-Strand-Breaks, Antioxidant Capacity and Blood Lipids in Rats. *Exp. Gerontol.* **2004**, *39*, 1189–1198. [\[CrossRef\]](http://doi.org/10.1016/j.exger.2004.05.002)
- 60. Dizdaroglu, M. Oxidatively Induced DNA Damage and Its Repair in Cancer. *Mutat. Res./Rev. Mutat. Res.* **2015**, *763*, 212–245. [\[CrossRef\]](http://doi.org/10.1016/j.mrrev.2014.11.002)
- 61. Kikugawa, K.; Yasuhara, Y.; Ando, K.; Koyama, K.; Hiramoto, K.; Suzuki, M. Protective Effect of Supplementation of Fish Oil with High N−3 Polyunsaturated Fatty Acids against Oxidative Stress-Induced DNA Damage of Rat Liver in Vivo. *J. Agric. Food Chem.* **2003**, *51*, 6073–6079. [\[CrossRef\]](http://doi.org/10.1021/jf030141v)
- 62. Müllner, E.; Brath, H.; Pleifer, S.; Schiermayr, C.; Baierl, A.; Wallner, M.; Fastian, T.; Millner, Y.; Paller, K.; Henriksen, T.; et al. Vegetables and PUFA-Rich Plant Oil Reduce DNA Strand Breaks in Individuals with Type 2 Diabetes. *Mol. Nutr. Food Res.* **2013**, *57*, 328–338. [\[CrossRef\]](http://doi.org/10.1002/mnfr.201200343)
- 63. Mitjavila, M.T.; Fandos, M.; Salas-Salvadó, J.; Covas, M.I.; Borrego, S.; Estruch, R.; Lamuela-Raventós, R.; Corella, D.; Martínez-Gonzalez, M.; Sánchez, J.M.; et al. The Mediterranean Diet Improves the Systemic Lipid and DNA Oxidative Damage in Metabolic Syndrome Individuals. A Randomized, Controlled, Trial. *Clin. Nutr.* **2013**, *32*, 172–178. [\[CrossRef\]](http://doi.org/10.1016/j.clnu.2012.08.002)
- 64. Konstantinidou, V.; Covas, M.-I.; Muñz-Aguayo, D.; Khymenets, O.; de la Torre, R.; Saez, G.; del Carmen Tormos, M.; Toledo, E.; Marti, A.; Ruiz-Gutiérrez, V.; et al. In Vivo Nutrigenomic Effects of Virgin Olive Oil Polyphenols within the Frame of the Mediterranean Diet: A Randomized Controlled Trial. *FASEB J.* **2010**, *24*, 2546–2557. [\[CrossRef\]](http://doi.org/10.1096/fj.09-148452)
- 65. Garcia-Arellano, A.; Ramallal, R.; Ruiz-Canela, M.; Salas-Salvadó, J.; Corella, D.; Shivappa, N.; Schröder, H.; Hébert, J.R.; Ros, E.; Gómez-Garcia, E.; et al. Dietary Inflammatory Index and Incidence of Cardiovascular Disease in the PREDIMED Study. *Nutrients* **2015**, *7*, 4124–4138. [\[CrossRef\]](http://doi.org/10.3390/nu7064124) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26035241)
- 66. Guasch-Ferré, M.; Salas-Salvadó, J.; Ros, E.; Estruch, R.; Corella, D.; Fitó, M.; Martínez-González, M.A.; Arós, F.; Gómez-Gracia, E.; Fiol, M.; et al. The PREDIMED Trial, Mediterranean Diet and Health Outcomes: How Strong Is the Evidence? *Nutr. Metab. Cardiovasc. Dis.* **2017**, *27*, 624–632. [\[CrossRef\]](http://doi.org/10.1016/j.numecd.2017.05.004)
- 67. Martínez-González, M.A.; Salas-Salvadó, J.; Estruch, R.; Corella, D.; Fitó, M.; Ros, E. Benefits of the Mediterranean Diet: Insights From the PREDIMED Study. *Prog. Cardiovasc. Dis.* **2015**, *58*, 50–60. [\[CrossRef\]](http://doi.org/10.1016/j.pcad.2015.04.003) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25940230)
- 68. Koloverou, E.; Panagiotakos, D.B.; Pitsavos, C.; Chrysohoou, C.; Georgousopoulou, E.N.; Grekas, A.; Christou, A.; Chatzigeorgiou, M.; Skoumas, I.; Tousoulis, D.; et al. Adherence to Mediterranean Diet and 10-Year Incidence (2002– 2012) of Diabetes: Correlations with Inflammatory and Oxidative Stress Biomarkers in the ATTICA Cohort Study. *Diabetes/Metab. Res. Rev.* **2016**, *32*, 73–81. [\[CrossRef\]](http://doi.org/10.1002/dmrr.2672)
- 69. Buil-Cosiales, P.; Toledo, E.; Salas-Salvadó, J.; Zazpe, I.; Farràs, M.; Basterra-Gortari, F.J.; Diez-Espino, J.; Estruch, R.; Corella, D.; Ros, E.; et al. Association between Dietary Fibre Intake and Fruit, Vegetable or Whole-Grain Consumption and the Risk of CVD: Results from the PREvención Con DIeta MEDiterránea (PREDIMED) Trial. *Br. J. Nutr.* **2016**, *116*, 534–546. [\[CrossRef\]](http://doi.org/10.1017/S0007114516002099) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/27264785)
- 70. Del Bo', C.; Marino, M.; Martini, D.; Tucci, M.; Ciappellano, S.; Riso, P.; Porrini, M. Overview of Human Intervention Studies Evaluating the Impact of the Mediterranean Diet on Markers of DNA Damage. *Nutrients* **2019**, *11*, 391. [\[CrossRef\]](http://doi.org/10.3390/nu11020391)
- 71. Giacosa, A.; Barale, R.; Bavaresco, L.; Gatenby, P.; Gerbi, V.; Janssens, J.; Johnston, B.; Kas, K.; La Vecchia, C.; Mainguet, P.; et al. Cancer Prevention in Europe: The Mediterranean Diet as a Protective Choice. *Eur. J. Cancer Prev.* **2013**, *22*, 90–95. [\[CrossRef\]](http://doi.org/10.1097/CEJ.0b013e328354d2d7)
- 72. Grosso, G.; Buscemi, S.; Galvano, F.; Mistretta, A.; Marventano, S.; Vela, V.L.; Drago, F.; Gangi, S.; Basile, F.; Biondi, A. Mediterranean Diet and Cancer: Epidemiological Evidence and Mechanism of Selected Aspects. *BMC Surg.* **2013**, *13*, S14. [\[CrossRef\]](http://doi.org/10.1186/1471-2482-13-S2-S14)
- 73. Schwingshackl, L.; Hoffmann, G. Adherence to Mediterranean Diet and Risk of Cancer: A Systematic Review and Meta-Analysis of Observational Studies. *Int. J. Cancer* **2014**, *135*, 1884–1897. [\[CrossRef\]](http://doi.org/10.1002/ijc.28824)
- 74. Acevedo-León, D.; Gómez-Abril, S.Á.; Monzó-Beltrán, L.; Estañ-Capell, N.; Arroyo-Montañés, R.; Bañuls, C.; Salas-Salvadó, J.; Sáez, G. Adherence to the Mediterranean Diet Has a Protective Role against Metabolic and DNA Damage Markers in Colorectal Cancer Patients. *Antioxidants* **2022**, *11*, 499. [\[CrossRef\]](http://doi.org/10.3390/antiox11030499)
- 75. Knopman, D.S. Mediterranean Diet and Late-Life Cognitive Impairment: A Taste of Benefit. *JAMA J. Am. Med. Assoc.* **2009**, *302*, 686. [\[CrossRef\]](http://doi.org/10.1001/jama.2009.1149) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/19671912)
- 76. Tsolaki, M.N.; Koutsouraki, E.S.; Katsipis, G.K.; Myserlis, P.G.; Chatzithoma, M.A.; Pantazaki, A.A. Alternative Anti-Infective/Anti-Inflammatory Therapeutic Options for Fighting Alzheimer's Disease. *Front. Anti-Infect. Drug Discov.* **2017**, *6*, 3–161. [\[CrossRef\]](http://doi.org/10.2174/9781681084794117060003)
- 77. Zhang, C.; Nestorova, G.; Rissman, R.A.; Feng, J. Detection and Quantification of 8-Hydroxy-2'-Deoxyguanosine in Alzheimer's Transgenic Mouse Urine Using Capillary Electrophoresis. *Electrophoresis* **2013**, *34*, 2268–2274. [\[CrossRef\]](http://doi.org/10.1002/elps.201300036) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23712533)
- 78. Wang, J.; Markesbery, W.R.; Lovell, M.A. Increased Oxidative Damage in Nuclear and Mitochondrial DNA in Mild Cognitive Impairment. *J. Neurochem.* **2006**, *96*, 825–832. [\[CrossRef\]](http://doi.org/10.1111/j.1471-4159.2005.03615.x) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/16405502)
- 79. Tzekaki, E.E.; Tsolaki, M.; Geromichalos, G.D.; Pantazaki, A.A. Extra Virgin Olive Oil Consumption from Mild Cognitive Impairment Patients Attenuates Oxidative and Nitrative Stress Reflecting on the Reduction of the PARP Levels and DNA Damage. *Exp. Gerontol.* **2021**, *156*, 111621. [\[CrossRef\]](http://doi.org/10.1016/j.exger.2021.111621) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/34748951)
- 80. Coluzzi, E.; Colamartino, M.; Cozzi, R.; Leone, S.; Meneghini, C.; O'Callaghan, N.; Sgura, A. Oxidative Stress Induces Persistent Telomeric DNA Damage Responsible for Nuclear Morphology Change in Mammalian Cells. *PLoS ONE* **2014**, *9*, e110963. [\[CrossRef\]](http://doi.org/10.1371/journal.pone.0110963) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25354277)
- 81. Aviv, A. Leukocyte Telomere Length: The Telomere Tale Continues. *Am. J. Clin. Nutr.* **2009**, *89*, 1721–1722. [\[CrossRef\]](http://doi.org/10.3945/ajcn.2009.27807)
- 82. Boonekamp, J.J.; Simons, M.J.P.; Hemerik, L.; Verhulst, S. Telomere Length Behaves as Biomarker of Somatic Redundancy Rather than Biological Age. *Aging Cell* **2013**, *12*, 330–332. [\[CrossRef\]](http://doi.org/10.1111/acel.12050)
- 83. Calado, R.T.; Young, N.S. Telomere Diseases. *N. Engl. J. Med.* **2009**, *361*, 2353–2365. [\[CrossRef\]](http://doi.org/10.1056/NEJMra0903373)
- 84. Boccardi, V.; Esposito, A.; Rizzo, M.R.; Marfella, R.; Barbieri, M.; Paolisso, G. Mediterranean Diet, Telomere Maintenance and Health Status among Elderly. *PLoS ONE* **2013**, *8*, e62781. [\[CrossRef\]](http://doi.org/10.1371/journal.pone.0062781)
- 85. Crous-Bou, M.; Fung, T.T.; Prescott, J.; Julin, B.; Du, M.; Sun, Q.; Rexrode, K.M.; Hu, F.B.; De Vivo, I. Mediterranean Diet and Telomere Length in Nurses' Health Study: Population Based Cohort Study. *BMJ* **2014**, *349*, g6674. [\[CrossRef\]](http://doi.org/10.1136/bmj.g6674)
- 86. García-Calzón, S.; Zalba, G.; Ruiz-Canela, M.; Shivappa, N.; Hébert, J.R.; Martínez, J.A.; Fitó, M.; Gómez-Gracia, E.; Martínez-González, M.A.; Marti, A. Dietary Inflammatory Index and Telomere Length in Subjects with a High Cardiovascular Disease Risk from the PREDIMED-NAVARRA Study: Cross-Sectional and Longitudinal Analyses over 5 y. *Am. J. Clin. Nutr.* **2015**, *102*, 897–904. [\[CrossRef\]](http://doi.org/10.3945/ajcn.115.116863)
- 87. Canudas, S.; Becerra-Tomas, N.; Hernandez-Alonso, P.; Galie, S.; Leung, C.; Crous-Bou, M.; De Vivo, I.; Gao, Y.; Gu, Y.; Meinila, J.; et al. Mediterranean Diet and Telomere Length: A Systematic Review and Meta-Analysis. *Adv. Nutr.* **2020**, *11*, 1544–1554. [\[CrossRef\]](http://doi.org/10.1093/advances/nmaa079)
- 88. García-Calzón, S.; Martínez-González, M.A.; Razquin, C.; Arós, F.; Lapetra, J.; Martínez, J.A.; Zalba, G.; Marti, A. Mediterranean Diet and Telomere Length in High Cardiovascular Risk Subjects from the PREDIMED-NAVARRA Study. *Clin. Nutr.* **2016**, *35*, 1399–1405. [\[CrossRef\]](http://doi.org/10.1016/j.clnu.2016.03.013)
- 89. Meinilä, J.; Perälä, M.M.; Kautiainen, H.; Männistö, S.; Kanerva, N.; Shivappa, N.; Hébert, J.R.; Iozzo, P.; Guzzardi, M.A.; Eriksson, J.G. Healthy Diets and Telomere Length and Attrition during a 10-Year Follow-Up. *Eur. J. Clin. Nutr.* **2019**, *73*, 1352–1360. [\[CrossRef\]](http://doi.org/10.1038/s41430-018-0387-4)
- 90. Gu, Y.; Honig, L.S.; Schupf, N.; Lee, J.H.; Luchsinger, J.A.; Stern, Y.; Scarmeas, N. Mediterranean Diet and Leukocyte Telomere Length in a Multi-Ethnic Elderly Population. *Age* **2015**, *37*, 24. [\[CrossRef\]](http://doi.org/10.1007/s11357-015-9758-0) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25750063)
- 91. García-Calzón, S.; Martínez-González, M.A.; Razquin, C.; Corella, D.; Salas-Salvadó, J.; Alfredo Martínez, J.; Zalba, G.; Marti Del Moral, A. Pro12Ala Polymorphism of the PPARγ2 Gene Interacts with a Mediterranean Diet to Prevent Telomere Shortening in the PREDIMED-NAVARRA Randomized Trial. *Circ. Cardiovasc. Genet.* **2015**, *8*, 91–99. [\[CrossRef\]](http://doi.org/10.1161/CIRCGENETICS.114.000635) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25406242)
- 92. O'Callaghan, N.; Parletta, N.; Milte, C.M.; Benassi-Evans, B.; Fenech, M.; Howe, P.R.C. Telomere Shortening in Elderly Individuals with Mild Cognitive Impairment May Be Attenuated with ω-3 Fatty Acid Supplementation: A Randomized Controlled Pilot Study. *Nutrition* **2014**, *30*, 489–491. [\[CrossRef\]](http://doi.org/10.1016/j.nut.2013.09.013) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24342530)
- 93. Kalstad, A.A.; Tveit, S.; Myhre, P.L.; Laake, K.; Opstad, T.B.; Tveit, A.; Schmidt, E.B.; Solheim, S.; Arnesen, H.; Seljeflot, I. Leukocyte Telomere Length and Serum Polyunsaturated Fatty Acids, Dietary Habits, Cardiovascular Risk Factors and Features of Myocardial Infarction in Elderly Patients. *BMC Geriatr.* **2019**, *19*, 1–9. [\[CrossRef\]](http://doi.org/10.1186/s12877-019-1383-9) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31881852)
- 94. Barnes, R.P.; Fouquerel, E.; Opresko, P.L. The Impact of Oxidative DNA Damage and Stress on Telomere Homeostasis. *Mech. Ageing Dev.* **2019**, *177*, 37–45. [\[CrossRef\]](http://doi.org/10.1016/j.mad.2018.03.013)
- 95. Tomasova, K.; Kroupa, M.; Forsti, A.; Vodicka, P.; Vodickova, L. Telomere Maintenance in Interplay with DNA Repair in Pathogenesis and Treatment of Colorectal Cancer. *Mutagenesis* **2020**, *35*, 261–271. [\[CrossRef\]](http://doi.org/10.1093/mutage/geaa005)
- 96. Cai, Z.; Yan, L.J.; Ratka, A. Telomere Shortening and Alzheimer's Disease. *Neuromolecular Med.* **2013**, *15*, 25–48. [\[CrossRef\]](http://doi.org/10.1007/s12017-012-8207-9)
- 97. Guo, Y.; Yu, H. Leukocyte Telomere Length Shortening and Alzheimer's Disease Etiology. *J. Alzheimer's Dis. JAD* **2019**, *69*, 881–885. [\[CrossRef\]](http://doi.org/10.3233/JAD-190134) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31156167)
- 98. Daccache, A.; Lion, C.; Sibille, N.; Gerard, M.; Slomianny, C.; Lippens, G.; Cotelle, P. Oleuropein and Derivatives from Olives as Tau Aggregation Inhibitors. *Neurochem. Int.* **2011**, *58*, 700–707. [\[CrossRef\]](http://doi.org/10.1016/j.neuint.2011.02.010) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21333710)
- 99. Crous-Bou, M.; Molinuevo, J.L.; Sala-Vila, A. Plant-Rich Dietary Patterns, Plant Foods and Nutrients, and Telomere Length. *Adv. Nutr.* **2019**, *10*, S296. [\[CrossRef\]](http://doi.org/10.1093/advances/nmz026) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31728493)
- 100. Porquet, D.; Casadesús, G.; Bayod, S.; Vicente, A.; Canudas, A.M.; Vilaplana, J.; Pelegrí, C.; Sanfeliu, C.; Camins, A.; Pallàs, M.; et al. Dietary Resveratrol Prevents Alzheimer's Markers and Increases Life Span in SAMP8. *Age* **2013**, *35*, 1851–1865. [\[CrossRef\]](http://doi.org/10.1007/s11357-012-9489-4)
- 101. Pal, S.; Tyler, J.K. Epigenetics and Aging. *Sci. Adv.* **2016**, *2*, e1600584. [\[CrossRef\]](http://doi.org/10.1126/sciadv.1600584)
- 102. Horvath, H.; Horvath, S. DNA Methylation Age of Human Tissues and Cell Types. *Genome Biol.* **2013**, *14*, 115. [\[CrossRef\]](http://doi.org/10.1186/gb-2013-14-10-r115)
- 103. Lander, E.S.; Linton, L.M.; Birren, B.; Nusbaum, C.; Zody, M.C.; Baldwin, J.; Devon, K.; Dewar, K.; Doyle, M.; Fitzhugh, W.; et al. Initial Sequencing and Analysis of the Human Genome. *Nature* **2001**, *409*, 860–921. [\[CrossRef\]](http://doi.org/10.1038/35057062)
- 104. Yang, A.S.; Estécio, M.R.H.; Doshi, K.; Kondo, Y.; Tajara, E.H.; Issa, J.P.J. A Simple Method for Estimating Global DNA Methylation Using Bisulfite PCR of Repetitive DNA Elements. *Nucleic Acids Res.* **2004**, *32*, e38. [\[CrossRef\]](http://doi.org/10.1093/nar/gnh032)
- 105. Muka, T.; Koromani, F.; Portilla, E.; O'Connor, A.; Bramer, W.M.; Troup, J.; Chowdhury, R.; Dehghan, A.; Franco, O.H. The Role of Epigenetic Modifications in Cardiovascular Disease: A Systematic Review. *Int. J. Cardiol.* **2016**, *212*, 174–183. [\[CrossRef\]](http://doi.org/10.1016/j.ijcard.2016.03.062) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/27038728)
- 106. Caradonna, F.; Consiglio, O.; Luparello, C.; Gentile, C. Science and Healthy Meals in the World: Nutritional Epigenomics and Nutrigenetics of the Mediterranean Diet. *Nutrients* **2020**, *12*, 1748. [\[CrossRef\]](http://doi.org/10.3390/nu12061748) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32545252)
- 107. Barchitta, M.; Maugeri, A.; Quattrocchi, A.; Barone, G.; Mazzoleni, P.; Catalfo, A.; De Guidi, G.; Iemmolo, M.G.; Crimi, N.; Agodi, A. Mediterranean Diet and Particulate Matter Exposure Are Associated With LINE-1 Methylation: Results From a Cross-Sectional Study in Women. *Front. Genet.* **2018**, *9*, 514. [\[CrossRef\]](http://doi.org/10.3389/fgene.2018.00514) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30425730)
- 108. Maugeri, A.; Barchitta, M.; Magnano San Lio, R.; Favara, G.; La Rosa, M.C.; La Mastra, C.; Basile, G.; Agodi, A. Adherence to the Mediterranean Diet Partially Mediates Socioeconomic Differences in Leukocyte LINE-1 Methylation: Evidence from a Cross-Sectional Study in Italian Women. *Sci. Rep.* **2020**, *10*, 14360. [\[CrossRef\]](http://doi.org/10.1038/s41598-020-71352-9) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32873815)
- 109. Agodi, A.; Barchitta, M.; Quattrocchi, A.; Maugeri, A.; Canto, C.; Marchese, A.E.; Vinciguerra, M. Low Fruit Consumption and Folate Deficiency Are Associated with LINE-1 Hypomethylation in Women of a Cancer-Free Population. *Genes Nutr.* **2015**, *10*, 480. [\[CrossRef\]](http://doi.org/10.1007/s12263-015-0480-4)
- 110. Arpón, A.; Milagro, F.I.; Razquin, C.; Corella, D.; Estruch, R.; Fitó, M.; Marti, A.; Martínez-González, M.A.; Ros, E.; Salas-Salvadó, J.; et al. Impact of Consuming Extra-Virgin Olive Oil or Nuts within a Mediterranean Diet on DNA Methylation in Peripheral White Blood Cells within the PREDIMED-Navarra Randomized Controlled Trial: A Role for Dietary Lipids. *Nutrients* **2018**, *10*, 15. [\[CrossRef\]](http://doi.org/10.3390/nu10010015)
- 111. Barchitta, M.; Quattrocchi, A.; Maugeri, A.; Vinciguerra, M.; Agodi, A. LINE-1 Hypomethylation in Blood and Tissue Samples as an Epigenetic Marker for Cancer Risk: A Systematic Review and Meta-Analysis. *PLoS ONE* **2014**, *9*, e109478. [\[CrossRef\]](http://doi.org/10.1371/journal.pone.0109478)
- 112. Park, S.Y.; Seo, A.N.; Jung, H.Y.; Gwak, J.M.; Jung, N.; Cho, N.Y.; Kang, G.H. Alu and LINE-1 Hypomethylation Is Associated with HER2 Enriched Subtype of Breast Cancer. *PLoS ONE* **2014**, *9*, e100429. [\[CrossRef\]](http://doi.org/10.1371/journal.pone.0100429)
- 113. Martín-Núñez, G.M.; Rubio-Martín, E.; Cabrera-Mulero, R.; Rojo-Martínez, G.; Olveira, G.; Valdés, S.; Soriguer, F.; Castaño, L.; Morcillo, S. Type 2 Diabetes Mellitus in Relation to Global LINE-1 DNA Methylation in Peripheral Blood: A Cohort Study. *Epigenetics* **2014**, *9*, 1322. [\[CrossRef\]](http://doi.org/10.4161/15592294.2014.969617)
- 114. Turcot, V.; Tchernof, A.; Deshaies, Y.; Pérusse, L.; Bélisle, A.; Marceau, S.; Biron, S.; Lescelleur, O.; Biertho, L.; Vohl, M.-C. LINE-1 Methylation in Visceral Adipose Tissue of Severely Obese Individuals Is Associated with Metabolic Syndrome Status and Related Phenotypes. *Clin. Epigenetics* **2012**, *4*, 10. [\[CrossRef\]](http://doi.org/10.1186/1868-7083-4-10)
- 115. Tang, B.; Lei, B.; Qi, G.; Liang, X.; Tang, F.; Yuan, S.; Wang, Z.; Yu, S.; He, S. MicroRNA-155-3p Promotes Hepatocellular Carcinoma Formation by Suppressing FBXW7 Expression. *J. Exp. Clin. Cancer Res.* **2016**, *35*, 93. [\[CrossRef\]](http://doi.org/10.1186/s13046-016-0371-6) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/27306418)
- 116. Li, P.; Weng, Z.; Li, P.; Hu, F.; Zhang, Y.; Guo, Z.; Shen, W.; Zhao, C.; Dai, S. BATF3 Promotes Malignant Phenotype of Colorectal Cancer through the S1PR1/p-STAT3/MiR-155-3p/WDR82 Axis. *Cancer Gene Ther.* **2020**, *28*, 400–412. [\[CrossRef\]](http://doi.org/10.1038/s41417-020-00223-2) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33057139)
- 117. Spolverini, A.; Fuchs, G.; Bublik, D.R.; Oren, M. Let-7b and Let-7c MicroRNAs Promote Histone H2B Ubiquitylation and Inhibit Cell Migration by Targeting Multiple Components of the H2B Deubiquitylation Machinery. *Oncogene* **2017**, *36*, 5819–5828. [\[CrossRef\]](http://doi.org/10.1038/onc.2017.187)
- 118. Li, Y.; Dong, R.; Lu, M.; Cheng, C.; Feng, Z.; Zhao, R.; Liang, J.; Han, J.; Jiang, J.; Xu-Welliver, M.; et al. Let-7b-3p Inhibits Tumor Growth and Metastasis by Targeting the BRF2-Mediated MAPK/ERK Pathway in Human Lung Adenocarcinoma. *Transl. Lung Cancer Res.* **2021**, *10*, 1841–1856. [\[CrossRef\]](http://doi.org/10.21037/tlcr-21-299)
- 119. Marques-Rocha, J.L.; Milagro, F.I.; Mansego, M.L.; Zulet, M.A.; Bressan, J.; Martínez, J.A. Expression of Inflammation-Related MiRNAs in White Blood Cells from Subjects with Metabolic Syndrome after 8 Wk of Following a Mediterranean Diet-Based Weight Loss Program. *Nutrition* **2016**, *32*, 48–55. [\[CrossRef\]](http://doi.org/10.1016/j.nut.2015.06.008) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26421388)
- 120. Gao, X.; Chen, Q.; Yao, H.; Tan, J.; Liu, Z.; Zhou, Y.; Zou, Z. Epigenetics in Alzheimer's Disease. *Front. Aging Neurosci.* **2022**, *14*, 195. [\[CrossRef\]](http://doi.org/10.3389/fnagi.2022.911635) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/35813941)
- 121. Heo, J.; Lim, J.; Lee, S.; Jeong, J.; Kang, H.; Kim, Y.H.; Kang, J.W.; Yu, H.Y.; Jeong, E.M.; Kim, K.; et al. Sirt1 Regulates DNA Methylation and Differentiation Potential of Embryonic Stem Cells by Antagonizing Dnmt3l. *Cell Rep.* **2017**, *18*, 1930–1945. [\[CrossRef\]](http://doi.org/10.1016/j.celrep.2017.01.074)
- 122. Heger, V.; Tyni, J.; Hunyadi, A.; Horáková, L.; Lahtela-Kakkonen, M.; Rahnasto-Rilla, M. Quercetin Based Derivatives as Sirtuin Inhibitors. *Biomed. Pharmacother.* **2019**, *111*, 1326–1333. [\[CrossRef\]](http://doi.org/10.1016/j.biopha.2019.01.035)
- 123. Luccarini, I.; Grossi, C.; Rigacci, S.; Coppi, E.; Pugliese, A.M.; Pantano, D.; la Marca, G.; Ed Dami, T.; Berti, A.; Stefani, M.; et al. Oleuropein Aglycone Protects against Pyroglutamylated-3 Amyloid-ß Toxicity: Biochemical, Epigenetic and Functional Correlates. *Neurobiol. Aging* **2015**, *36*, 648–663. [\[CrossRef\]](http://doi.org/10.1016/j.neurobiolaging.2014.08.029)
- 124. Kaushik, S.; Cuervo, A.M. Proteostasis and Aging. *Nat. Med.* **2015**, *21*, 1406–1415. [\[CrossRef\]](http://doi.org/10.1038/nm.4001)
- 125. Treaster, S.B.; Ridgway, I.D.; Richardson, C.A.; Gaspar, M.B.; Chaudhuri, A.R.; Austad, S.N. Superior Proteome Stability in the Longest Lived Animal. *Age* **2014**, *36*, 1009–1017. [\[CrossRef\]](http://doi.org/10.1007/s11357-013-9597-9) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24254744)
- 126. Ma, Y.; Li, J. Metabolic Shifts during Aging and Pathology. *Compr. Physiol.* **2015**, *5*, 667. [\[CrossRef\]](http://doi.org/10.1002/CPHY.C140041)
- 127. Morimoto, R.I.; Cuervo, A.M. Proteostasis and the Aging Proteome in Health and Disease. *J. Gerontol. Ser. A Biol. Sci. Med. Sci.* **2014**, *69* (Suppl. S1), S33–S38. [\[CrossRef\]](http://doi.org/10.1093/gerona/glu049) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24833584)
- 128. Rubinsztein, D.C.; Mariño, G.; Kroemer, G. Autophagy and Aging. *Cell* **2011**, *146*, 682–695. [\[CrossRef\]](http://doi.org/10.1016/j.cell.2011.07.030) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21884931)
- 129. Madeo, F.; Zimmermann, A.; Maiuri, M.C.; Kroemer, G. Essential Role for Autophagy in Life Span Extension. *J. Clin. Investig.* **2015**, *125*, 85–93. [\[CrossRef\]](http://doi.org/10.1172/JCI73946)
- 130. Rigacci, S.; Miceli, C.; Nediani, C.; Berti, A.; Cascella, R.; Pantano, D.; Nardiello, P.; Luccarini, I.; Casamenti, F.; Stefani, M. Oleuropein Aglycone Induces Autophagy via the AMPK/MTOR Signalling Pathway: A Mechanistic Insight. *Oncotarget* **2015**, *6*, 35344–35357. [\[CrossRef\]](http://doi.org/10.18632/oncotarget.6119)
- 131. Schmitt, C.A.; Fridman, J.S.; Yang, M.; Lee, S.; Baranov, E.; Hoffman, R.M.; Lowe, S.W. A Senescence Program Controlled by P53 and P16INK4a Contributes to the Outcome of Cancer Therapy. *Cell* **2002**, *109*, 335–346. [\[CrossRef\]](http://doi.org/10.1016/S0092-8674(02)00734-1)
- 132. Yerbury, J.J.; Ooi, L.; Dillin, A.; Saunders, D.N.; Hatters, D.M.; Beart, P.M.; Cashman, N.R.; Wilson, M.R.; Ecroyd, H. Walking the Tightrope: Proteostasis and Neurodegenerative Disease. *J. Neurochem.* **2016**, *137*, 489–505. [\[CrossRef\]](http://doi.org/10.1111/jnc.13575)
- 133. Arendt, T.; Rödel, L.; Gärtner, U.; Holzer, M. Expression of the Cyclin-Dependent Kinase Inhibitor P16 in Alzheimer's Disease. *Neuroreport* **1996**, *7*, 3047–3049. [\[CrossRef\]](http://doi.org/10.1097/00001756-199611250-00050)
- 134. Mcshea, A.; Harris, P.L.R.; Webster, K.R.; Wahl, A.F.; From, M.A.S.; Squibb, B.-M.; Research Institute, P.; Washingtoni, P. Abnormal Expression of the Cell Cycle Regulators P16 and CDK4 in Alzheimer's Disease. *Am. J. Pathol.* **1997**, *150*, 1933.
- 135. Duyckaerts, C.; Delatour, B.; Potier, M.C. Classification and Basic Pathology of Alzheimer Disease. *Acta Neuropathol.* **2009**, *118*, 5–36. [\[CrossRef\]](http://doi.org/10.1007/s00401-009-0532-1)
- 136. Melki, R. Role of Different Alpha-Synuclein Strains in Synucleinopathies, Similarities with Other Neurodegenerative Diseases. *J. Park. Dis.* **2015**, *5*, 217–227. [\[CrossRef\]](http://doi.org/10.3233/JPD-150543) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25757830)
- 137. Hou, Y.; Dan, X.; Babbar, M.; Wei, Y.; Hasselbalch, S.G.; Croteau, D.L.; Bohr, V.A. Ageing as a Risk Factor for Neurodegenerative Disease. *Nat. Rev. Neurol.* **2019**, *15*, 565–581. [\[CrossRef\]](http://doi.org/10.1038/s41582-019-0244-7)
- 138. Monti, M.C.; Margarucci, L.; Tosco, A.; Riccio, R.; Casapullo, A. New Insights on the Interaction Mechanism between Tau Protein and Oleocanthal, an Extra-Virgin Olive-Oil Bioactive Component. *Food Funct.* **2011**, *2*, 423–428. [\[CrossRef\]](http://doi.org/10.1039/c1fo10064e) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21894330)
- 139. Monti, M.C.; Margarucci, L.; Riccio, R.; Casapullo, A. Modulation of Tau Protein Fibrillization by Oleocanthal. *J. Nat. Prod.* **2012**, *75*, 1584–1588. [\[CrossRef\]](http://doi.org/10.1021/np300384h)
- 140. Abuznait, A.H.; Qosa, H.; Busnena, B.A.; El Sayed, K.A.; Kaddoumi, A. Olive-Oil-Derived Oleocanthal Enhances β-Amyloid Clearance as a Potential Neuroprotective Mechanism against Alzheimer's Disease: In Vitro and in Vivo Studies. *ACS Chem. Neurosci.* **2013**, *4*, 973. [\[CrossRef\]](http://doi.org/10.1021/cn400024q)
- 141. Del Río, L.F.; Gutiérrez-Casado, E.; Varela-López, A.; Villalba, J.M. Olive Oil and the Hallmarks of Aging. *Molecules* **2016**, *21*, 163. [\[CrossRef\]](http://doi.org/10.3390/molecules21020163)
- 142. Albani, D.; Polito, L.; Signorini, A.; Forloni, G. Neuroprotective Properties of Resveratrol in Different Neurodegenerative Disorders. *BioFactors* **2010**, *36*, 370–376. [\[CrossRef\]](http://doi.org/10.1002/biof.118)
- 143. Dai, D.F.; Karunadharma, P.P.; Chiao, Y.A.; Basisty, N.; Crispin, D.; Hsieh, E.J.; Chen, T.; Gu, H.; Djukovic, D.; Raftery, D.; et al. Altered Proteome Turnover and Remodeling by Short-Term Caloric Restriction or Rapamycin Rejuvenate the Aging Heart. *Aging Cell* **2014**, *13*, 529–539. [\[CrossRef\]](http://doi.org/10.1111/acel.12203) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24612461)
- 144. Gouveia, M.; Xia, K.; Colón, W.; Vieira, S.I.; Ribeiro, F. Protein Aggregation, Cardiovascular Diseases, and Exercise Training: Where Do We Stand? *Ageing Res. Rev.* **2017**, *40*, 1–10. [\[CrossRef\]](http://doi.org/10.1016/j.arr.2017.07.005) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28757291)
- 145. Chin, J.H.; Okazaki, M.; Hu, Z.W.; Miller, J.W.; Hoffman, B.B. Activation of Heat Shock Protein (Hsp)70 and Proto-Oncogene Expression by Alpha1 Adrenergic Agonist in Rat Aorta with Age. *J. Clin. Investig.* **1996**, *97*, 2316. [\[CrossRef\]](http://doi.org/10.1172/JCI118674) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/8636412)
- 146. Marfella, R.; Di Filippo, C.; Laieta, M.T.; Vestini, R.; Barbieri, M.; Sangiulo, P.; Crescenzi, B.; Ferraraccio, F.; Rossi, F.; D'Amico, M.; et al. Effects of Ubiquitin-Proteasome System Deregulation on the Vascular Senescence and Atherosclerosis Process in Elderly Patients. *J. Gerontol. Ser. A* **2008**, *63*, 200–203. [\[CrossRef\]](http://doi.org/10.1093/gerona/63.2.200) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/18314458)
- 147. Katsiki, M.; Chondrogianni, N.; Chinou, I.; Rivett, A.J.; Gonos, E.S. The Olive Constituent Oleuropein Exhibits Proteasome Stimulatory Properties in Vitro and Confers Life Span Extension of Human Embryonic Fibroblasts. *Rejuvenation Res.* **2007**, *10*, 157–172. [\[CrossRef\]](http://doi.org/10.1089/rej.2006.0513) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/17518699)
- 148. Bullon, P.; Battino, M.; Varela-Lopez, A.; Perez-Lopez, P.; Granados-Principal, S.; Ramirez-Tortosa, M.C.; Ochoa, J.J.; Cordero, M.D.; Gonzalez-Alonso, A.; Ramirez-Tortosa, C.L.; et al. Diets Based on Virgin Olive Oil or Fish Oil but Not on Sunflower Oil Prevent Age-Related Alveolar Bone Resorption by Mitochondrial-Related Mechanisms. *PLoS ONE* **2013**, *8*, 74234. [\[CrossRef\]](http://doi.org/10.1371/journal.pone.0074234) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24066124)
- 149. Urra, H.; Dufey, E.; Avril, T.; Chevet, E.; Hetz, C. Endoplasmic Reticulum Stress and the Hallmarks of Cancer. *Trends Cancer* **2016**, *2*, 252–262. [\[CrossRef\]](http://doi.org/10.1016/j.trecan.2016.03.007)
- 150. Huber, A.L.; Lebeau, J.; Guillaumot, P.; Pétrilli, V.; Malek, M.; Chilloux, J.; Fauvet, F.; Payen, L.; Kfoury, A.; Renno, T.; et al. P58IPK-Mediated Attenuation of the Proapoptotic PERK-CHOP Pathway Allows Malignant Progression upon Low Glucose. *Mol. Cell* **2013**, *49*, 1049–1059. [\[CrossRef\]](http://doi.org/10.1016/j.molcel.2013.01.009)
- 151. Chen, X.; Iliopoulos, D.; Zhang, Q.; Tang, Q.; Greenblatt, M.B.; Hatziapostolou, M.; Lim, E.; Tam, W.L.; Ni, M.; Chen, Y.; et al. XBP1 Promotes Triple-Negative Breast Cancer by Controlling the HIF1α Pathway. *Nature* **2014**, *508*, 103–107. [\[CrossRef\]](http://doi.org/10.1038/nature13119)
- 152. Lhomond, S.; Avril, T.; Dejeans, N.; Voutetakis, K.; Doultsinos, D.; McMahon, M.; Pineau, R.; Obacz, J.; Papadodima, O.; Jouan, F.; et al. Dual IRE1 RNase Functions Dictate Glioblastoma Development. *EMBO Mol. Med.* **2018**, *10*, e7929. [\[CrossRef\]](http://doi.org/10.15252/emmm.201707929)
- 153. Lee, A.H.; Iwakoshi, N.N.; Anderson, K.C.; Glimcher, L.H. Proteasome Inhibitors Disrupt the Unfolded Protein Response in Myeloma Cells. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 9946–9951. [\[CrossRef\]](http://doi.org/10.1073/pnas.1334037100)
- 154. Tang, C.H.A.; Ranatunga, S.; Kriss, C.L.; Cubitt, C.L.; Tao, J.; Pinilla-Ibarz, J.A.; Del Valle, J.R.; Hu, C.C.A. Inhibition of ER Stress-Associated IRE-1/XBP-1 Pathway Reduces Leukemic Cell Survival. *J. Clin. Investig.* **2014**, *124*, 2585–2598. [\[CrossRef\]](http://doi.org/10.1172/JCI73448)
- 155. Song, B.; Scheuner, D.; Ron, D.; Pennathur, S.; Kaufman, R.J. Chop Deletion Reduces Oxidative Stress, Improves Beta Cell Function, and Promotes Cell Survival in Multiple Mouse Models of Diabetes. *J. Clin. Investig.* **2008**, *118*, 3378–3389. [\[CrossRef\]](http://doi.org/10.1172/JCI34587)
- 156. Sheng, X.; Nenseth, H.Z.; Qu, S.; Kuzu, O.F.; Frahnow, T.; Simon, L.; Greene, S.; Zeng, Q.; Fazli, L.; Rennie, P.S.; et al. IRE1α-XBP1s Pathway Promotes Prostate Cancer by Activating c-MYC Signaling. *Nat. Commun.* **2019**, *10*, 323. [\[CrossRef\]](http://doi.org/10.1038/s41467-018-08152-3) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30679434)
- 157. Wu, Y.; Shan, B.; Dai, J.; Xia, Z.; Cai, J.; Chen, T.; Lv, S.; Feng, Y.; Zheng, L.; Wang, Y.; et al. Dual Role for Inositol-Requiring Enzyme 1a in Promoting the Development of Hepatocellular Carcinoma During Diet-Induced Obesity in Mice A HE STUDY OF LIVER D I S E ASES T MERICAN ASSOCIATION FOR. *Hepatology* **2018**, *68*, 533–546. [\[CrossRef\]](http://doi.org/10.1002/hep.29871) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/29506314)
- 158. Yubero-Serrano, E.M.; Gonzalez-Guardia, L.; Rangel-Zuñiga, O.; Delgado-Lista, J.; Gutierrez-Mariscal, F.M.; Perez-Martinez, P.; Delgado-Casado, N.; Cruz-Teno, C.; Tinahones, F.J.; Villalba, J.M.; et al. Mediterranean Diet Supplemented with Coenzyme Q 10 Modifies the Expression of Proinflammatory and Endoplasmic Reticulum Stress-Related Genes in Elderly Men and Women. *J. Gerontol.-Ser. A Biol. Sci. Med. Sci.* **2012**, *67A*, 3–10. [\[CrossRef\]](http://doi.org/10.1093/gerona/glr167)
- 159. Quiles, J.L.; Huertas, J.R.; Mañas, M.; Battino, M.; Mataix, J. Physical Exercise Affects the Lipid Profile of Mitochondrial Membranes in Rats Fed with Virgin Olive Oil or Sunflower Oil. *Br. J. Nutr.* **1999**, *81*, 21–24. [\[CrossRef\]](http://doi.org/10.1017/S0007114599000094)
- 160. Lozano-Sánchez, J.; Segura-Carretero, A.; Menendez, J.A.; Oliveras-Ferraros, C.; Cerretani, L.; Fernández-Gutiérrez, A. Prediction of Extra Virgin Olive Oil Varieties through Their Phenolic Profile. Potential Cytotoxic Activity against Human Breast Cancer Cells. *J. Agric. Food Chem.* **2010**, *58*, 9942–9955. [\[CrossRef\]](http://doi.org/10.1021/JF101502Q)
- 161. Oliveras-Ferraros, C.; Fernández-Arroyo, S.; Vazquez-Martin, A.; Lozano-Sánchez, J.; Cufí, S.; Joven, J.; Micol, V.; Fernández-Gutiérrez, A.; Segura-Carretero, A.; Menendez, J.A. Crude Phenolic Extracts from Extra Virgin Olive Oil Circumvent de Novo Breast Cancer Resistance to HER1/HER2-Targeting Drugs by Inducing GADD45-Sensed Cellular Stress, G2/M Arrest and Hyperacetylation of Histone H3. *Int. J. Oncol.* **2011**, *38*, 1533–1547. [\[CrossRef\]](http://doi.org/10.3892/IJO.2011.993) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21455577)
- 162. Templeman, N.M.; Murphy, C.T. Regulation of Reproduction and Longevity by Nutrient-Sensing Pathways. *J. Cell Biol.* **2018**, *217*, 93–106. [\[CrossRef\]](http://doi.org/10.1083/jcb.201707168)
- 163. de Lucia, C.; Murphy, T.; Steves, C.J.; Dobson, R.J.B.; Proitsi, P.; Thuret, S. Lifestyle Mediates the Role of Nutrient-Sensing Pathways in Cognitive Aging: Cellular and Epidemiological Evidence. *Commun. Biol.* **2020**, *3*, 157. [\[CrossRef\]](http://doi.org/10.1038/s42003-020-0844-1) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32242137)
- 164. Fontana, L.; Partridge, L.; Longo, V.D. Extending Healthy Life Span–from Yeast to Humans. *Science* **2010**, *328*, 321–326. [\[CrossRef\]](http://doi.org/10.1126/science.1172539) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/20395504)
- 165. Schumacher, B.; Van Der Pluijm, I.; Moorhouse, M.J.; Kosteas, T.; Robinson, A.R.; Suh, Y.; Breit, T.M.; Van Steeg, H.; Niedernhofer, L.J.; Van Ijcken, W.; et al. Delayed and Accelerated Aging Share Common Longevity Assurance Mechanisms. *PLoS Genet.* **2008**, *4*, e1000161. [\[CrossRef\]](http://doi.org/10.1371/journal.pgen.1000161)
- 166. Garinis, G.A.; van der Horst, G.T.J.; Vijg, J.; Hoeijmakers, J.H.J. DNA Damage and Ageing: New-Age Ideas for an Age-Old Problem. *Nat. Cell Biol.* **2008**, *10*, 1241–1247. [\[CrossRef\]](http://doi.org/10.1038/ncb1108-1241)
- 167. Barzilai, N.; Huffman, D.M.; Muzumdar, R.H.; Bartke, A. The Critical Role of Metabolic Pathways in Aging. *Diabetes* **2012**, *61*, 1315. [\[CrossRef\]](http://doi.org/10.2337/db11-1300)
- 168. Rincon, M.; Rudin, E.; Barzilai, N. The Insulin/IGF-1 Signaling in Mammals and Its Relevance to Human Longevity. *Exp. Gerontol.* **2005**, *40*, 873–877. [\[CrossRef\]](http://doi.org/10.1016/j.exger.2005.06.014)
- 169. Van Heemst, D. Insulin, IGF-1 and Longevity. *Aging Dis.* **2010**, *1*, 147–157.
- 170. Calnan, D.R.; Brunet, A. The FoxO Code. *Oncogene* **2008**, *27*, 2276–2288. [\[CrossRef\]](http://doi.org/10.1038/onc.2008.21)
- 171. Johnson, S.C.; Rabinovitch, P.S.; Kaeberlein, M. MTOR Is a Key Modulator of Ageing and Age-Related Disease. *Nature* **2013**, *493*, 338–345. [\[CrossRef\]](http://doi.org/10.1038/nature11861)
- 172. Lamming, D.W.; Ye, L.; Katajisto, P.; Goncalves, M.D.; Saitoh, M.; Stevens, D.M.; Davis, J.G.; Salmon, A.B.; Richardson, A.; Ahima, R.S.; et al. Rapamycin-Induced Insulin Resistance Is Mediated by MTORC2 Loss and Uncoupled from Longevity. *Science* **2012**, *335*, 1638–1643. [\[CrossRef\]](http://doi.org/10.1126/science.1215135)
- 173. Selman, C.; Tullet, J.M.A.; Wieser, D.; Irvine, E.; Lingard, S.J.; Choudhury, A.I.; Claret, M.; Al-Qassab, H.; Carmignac, D.; Ramadani, F.; et al. Ribosomal Protein S6 Kinase 1 Signaling Regulates Mammalian Life Span. *Science* **2009**, *326*, 140–144. [\[CrossRef\]](http://doi.org/10.1126/science.1177221)
- 174. Meric-Bernstam, F.; Gonzalez-Angulo, A.M. Targeting the MTOR Signaling Network for Cancer Therapy. *J. Clin. Oncol. Off. J. Am. Soc. Clin. Oncol.* **2009**, *27*, 2278–2287. [\[CrossRef\]](http://doi.org/10.1200/JCO.2008.20.0766)
- 175. Engelman, J.A. Targeting PI3K Signalling in Cancer: Opportunities, Challenges and Limitations. Nature reviews. *Cancer* **2009**, *9*, 550–562. [\[CrossRef\]](http://doi.org/10.1038/NRC2664)
- 176. Caccamo, A.; Majumder, S.; Richardson, A.; Strong, R.; Oddo, S. Molecular Interplay between Mammalian Target of Rapamycin (MTOR), Amyloid-Beta, and Tau: Effects on Cognitive Impairments. *J. Biol. Chem.* **2010**, *285*, 13107–13120. [\[CrossRef\]](http://doi.org/10.1074/jbc.M110.100420)
- 177. Zoncu, R.; Efeyan, A.; Sabatini, D.M. MTOR: From Growth Signal Integration to Cancer, Diabetes and Ageing. Nature reviews. *Mol. Cell Biol.* **2011**, *12*, 21–35. [\[CrossRef\]](http://doi.org/10.1038/NRM3025)
- 178. Wilkinson, J.E.; Burmeister, L.; Brooks, S.V.; Chan, C.C.; Friedline, S.; Harrison, D.E.; Hejtmancik, J.F.; Nadon, N.; Strong, R.; Wood, L.K.; et al. Rapamycin Slows Aging in Mice. *Aging Cell* **2012**, *11*, 675. [\[CrossRef\]](http://doi.org/10.1111/j.1474-9726.2012.00832.x)
- 179. El-Khamisy, S.F.; Saifi, G.M.; Weinfeld, M.; Johansson, F.; Helleday, T.; Lupski, J.R.; Caldecott, K.W. Defective DNA Single-Strand Break Repair in Spinocerebellar Ataxia with Axonal Neuropathy-1. *Nature* **2005**, *434*, 108–113. [\[CrossRef\]](http://doi.org/10.1038/nature03314)
- 180. Alers, S.; Löffler, A.S.; Wesselborg, S.; Stork, B. Role of AMPK-MTOR-Ulk1/2 in the Regulation of Autophagy: Cross Talk, Shortcuts, and Feedbacks. *Mol. Cell. Biol.* **2012**, *32*, 2–11. [\[CrossRef\]](http://doi.org/10.1128/MCB.06159-11)
- 181. Rodgers, J.T.; Lerin, C.; Haas, W.; Gygi, S.P.; Spiegelman, B.M.; Puigserver, P. Nutrient Control of Glucose Homeostasis through a Complex of PGC-1alpha and SIRT1. *Nature* **2005**, *434*, 113–118. [\[CrossRef\]](http://doi.org/10.1038/nature03354)
- 182. Clark, J.; Reddy, S.; Zheng, K.; Betensky, R.A.; Simon, D.K. Association of PGC-1alpha Polymorphisms with Age of Onset and Risk of Parkinson's Disease. *BMC Med. Genet.* **2011**, *12*, 69. [\[CrossRef\]](http://doi.org/10.1186/1471-2350-12-69)
- 183. Colman, R.J.; Anderson, R.M.; Johnson, S.C.; Kastman, E.K.; Kosmatka, K.J.; Beasley, T.M.; Allison, D.B.; Cruzen, C.; Simmons, H.A.; Kemnitz, J.W.; et al. Caloric Restriction Delays Disease Onset and Mortality in Rhesus Monkeys. *Science* **2009**, *325*, 201–204. [\[CrossRef\]](http://doi.org/10.1126/science.1173635)
- 184. Vasto, S.; Buscemi, S.; Barera, A.; Di Carlo, M.; Accardi, G.; Caruso, C. Mediterranean Diet and Healthy Ageing: A Sicilian Perspective. *Gerontology* **2014**, *60*, 508–518. [\[CrossRef\]](http://doi.org/10.1159/000363060)
- 185. Lee, I.H.; Cao, L.; Mostoslavsky, R.; Lombard, D.B.; Liu, J.; Bruns, N.E.; Tsokos, M.; Alt, F.W.; Finkel, T. A Role for the NAD-Dependent Deacetylase Sirt1 in the Regulation of Autophagy. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 3374–3379. [\[CrossRef\]](http://doi.org/10.1073/pnas.0712145105)
- 186. Jung, C.H.; Ro, S.H.; Cao, J.; Otto, N.M.; Kim, D.H. MTOR Regulation of Autophagy. *FEBS Lett.* **2010**, *584*, 1287–1295. [\[CrossRef\]](http://doi.org/10.1016/j.febslet.2010.01.017)
- 187. Khanfar, M.A.; Bardaweel, S.K.; Akl, M.R.; El Sayed, K.A. Olive Oil-Derived Oleocanthal as Potent Inhibitor of Mammalian Target of Rapamycin: Biological Evaluation and Molecular Modeling Studies. *Phytother. Res. PTR* **2015**, *29*, 1776–1782. [\[CrossRef\]](http://doi.org/10.1002/ptr.5434)
- 188. Jemai, H.; Feki, A.E.L.; Sayadi, S. Antidiabetic and Antioxidant Effects of Hydroxytyrosol and Oleuropein from Olive Leaves in Alloxan-Diabetic Rats. *J. Agric. Food Chem.* **2009**, *57*, 8798–8804. [\[CrossRef\]](http://doi.org/10.1021/jf901280r)
- 189. Al-Azzawie, H.F.; Alhamdani, M.S.S. Hypoglycemic and Antioxidant Effect of Oleuropein in Alloxan-Diabetic Rabbits. *Life Sci.* **2006**, *78*, 1371–1377. [\[CrossRef\]](http://doi.org/10.1016/j.lfs.2005.07.029)
- 190. Camandola, S.; Mattson, M.P. Brain Metabolism in Health, Aging, and Neurodegeneration. *EMBO J.* **2017**, *36*, 1474–1492. [\[CrossRef\]](http://doi.org/10.15252/embj.201695810)
- 191. Cai, H.; Cong, W.; Ji, S.; Rothman, S.; Maudsley, S.; Martin, B. Metabolic Dysfunction in Alzheimer's Disease and Related Neurodegenerative Disorders. *Curr. Alzheimer Res.* **2012**, *9*, 5–17. [\[CrossRef\]](http://doi.org/10.2174/156720512799015064)
- 192. Runchey, S.S.; Pollak, M.N.; Valsta, L.M.; Coronado, G.D.; Schwarz, Y.; Breymeyer, K.L.; Wang, C.; Wang, C.Y.; Lampe, J.W.; Neuhouser, M.L. Glycemic Load Effect on Fasting and Post-Prandial Serum Glucose, Insulin, IGF-1 and IGFBP-3 in a Randomized, Controlled Feeding Study. *Eur. J. Clin. Nutr.* **2012**, *66*, 1146–1152. [\[CrossRef\]](http://doi.org/10.1038/ejcn.2012.107)
- 193. Levine, M.E.; Suarez, J.A.; Brandhorst, S.; Balasubramanian, P.; Cheng, C.W.; Madia, F.; Fontana, L.; Mirisola, M.G.; Guevara-Aguirre, J.; Wan, J.; et al. Low Protein Intake Is Associated with a Major Reduction in IGF-1, Cancer, and Overall Mortality in the 65 and Younger but Not Older Population. *Cell Metab.* **2014**, *19*, 407–417. [\[CrossRef\]](http://doi.org/10.1016/j.cmet.2014.02.006)
- 194. Fontana, L.; Adelaiye, R.M.; Rastelli, A.L.; Miles, K.M.; Ciamporcero, E.; Longo, V.D.; Nguyen, H.; Vessella, R.; Pili, R. Dietary Protein Restriction Inhibits Tumor Growth in Human Xenograft Models of Prostate and Breast Cancer. *Oncotarget* **2013**, *4*, 2451. [\[CrossRef\]](http://doi.org/10.18632/oncotarget.1586)
- 195. Menendez, J.A.; Joven, J.; Aragonès, G.; Barrajón-Catalán, E.; Beltrán-Debón, R.; Borrás-Linares, I.; Camps, J.; Corominas-Faja, B.; Cufí, S.; Fernández-Arroyo, S.; et al. Xenohormetic and Anti-Aging Activity of Secoiridoid Polyphenols Present in Extra Virgin Olive Oil: A New Family of Gerosuppressant Agents. *Cell Cycle* **2013**, *12*, 555–578. [\[CrossRef\]](http://doi.org/10.4161/cc.23756)
- 196. Brown-Borg, H.M.; Buffenstein, R. Cutting Back on the Essentials: Can Manipulating Intake of Specific Amino Acids Modulate Health and Lifespan? *Ageing Res. Rev.* **2017**, *39*, 87–95. [\[CrossRef\]](http://doi.org/10.1016/j.arr.2016.08.007)
- 197. Lynch, C.J.; Adams, S.H. Branched-Chain Amino Acids in Metabolic Signalling and Insulin Resistance. Nature reviews. *Endocrinology* **2014**, *10*, 723–736. [\[CrossRef\]](http://doi.org/10.1038/NRENDO.2014.171)
- 198. Fontana, L.; Cummings, N.E.; Arriola Apelo, S.I.; Neuman, J.C.; Kasza, I.; Schmidt, B.A.; Cava, E.; Spelta, F.; Tosti, V.; Syed, F.A.; et al. Decreased Consumption of Branched-Chain Amino Acids Improves Metabolic Health. *Cell Rep.* **2016**, *16*, 520–530. [\[CrossRef\]](http://doi.org/10.1016/j.celrep.2016.05.092)
- 199. Remmen, H.V.; Richardson, A. Oxidative Damage to Mitochondria and Aging. *Exp. Gerontol.* **2001**, *36*, 957–968. [\[CrossRef\]](http://doi.org/10.1016/S0531-5565(01)00093-6)
- 200. Birnbaum, J.H.; Wanner, D.; Gietl, A.F.; Saake, A.; Kündig, T.M.; Hock, C.; Nitsch, R.M.; Tackenberg, C. Oxidative Stress and Altered Mitochondrial Protein Expression in the Absence of Amyloid-β and Tau Pathology in IPSC-Derived Neurons from Sporadic Alzheimer's Disease Patients. *Stem Cell Res.* **2018**, *27*, 121–130. [\[CrossRef\]](http://doi.org/10.1016/j.scr.2018.01.019)
- 201. Park, J.S.; Davis, R.L.; Sue, C.M. Mitochondrial Dysfunction in Parkinson's Disease: New Mechanistic Insights and Therapeutic Perspectives. *Curr. Neurol. Neurosci. Rep.* **2018**, *18*, 21. [\[CrossRef\]](http://doi.org/10.1007/s11910-018-0829-3)
- 202. Kudryavtseva, A.V.; Krasnov, G.S.; Dmitriev, A.A.; Alekseev, B.Y.; Kardymon, O.L.; Sadritdinova, A.F.; Fedorova, M.S.; Pokrovsky, A.V.; Melnikova, N.V.; Kaprin, A.D.; et al. Mitochondrial Dysfunction and Oxidative Stress in Aging and Cancer. *Oncotarget* **2016**, *7*, 44879. [\[CrossRef\]](http://doi.org/10.18632/oncotarget.9821)
- 203. Bhatti, J.S.; Bhatti, G.K.; Reddy, P.H. Mitochondrial Dysfunction and Oxidative Stress in Metabolic Disorders—A Step towards Mitochondria Based Therapeutic Strategies. *Biochim. Biophys. Acta Mol. Basis Dis.* **2017**, *1863*, 1066–1077. [\[CrossRef\]](http://doi.org/10.1016/j.bbadis.2016.11.010)
- 204. Kowaltowski, A.J.; de Souza-Pinto, N.C.; Castilho, R.F.; Vercesi, A.E. Mitochondria and Reactive Oxygen Species. *Free Radic. Biol. Med.* **2009**, *47*, 333–343. [\[CrossRef\]](http://doi.org/10.1016/j.freeradbiomed.2009.05.004)
- 205. Choksi, K.B.; Boylston, W.H.; Rabek, J.P.; Widger, W.R.; Papaconstantinou, J. Oxidatively Damaged Proteins of Heart Mitochondrial Electron Transport Complexes. *Biochim. Biophys. Acta Mol. Basis Dis.* **2004**, *1688*, 95–101. [\[CrossRef\]](http://doi.org/10.1016/j.bbadis.2003.11.007)
- 206. Sowers, J.R. Insulin Resistance and Hypertension. *Am. J. Physiol. Heart Circ. Physiol.* **2004**, *286*, H1597-602. [\[CrossRef\]](http://doi.org/10.1152/ajpheart.00026.2004)
- 207. Smith, S.C. Multiple Risk Factors for Cardiovascular Disease and Diabetes Mellitus. *Am. J. Med.* **2007**, *120*, S3–S11. [\[CrossRef\]](http://doi.org/10.1016/j.amjmed.2007.01.002)
- 208. Morino, K.; Petersen, K.F.; Dufour, S.; Befroy, D.; Frattini, J.; Shatzkes, N.; Neschen, S.; White, M.F.; Bilz, S.; Sono, S.; et al. Reduced Mitochondrial Density and Increased IRS-1 Serine Phosphorylation in Muscle of Insulin-Resistant Offspring of Type 2 Diabetic Parents. *J. Clin. Investig.* **2005**, *115*, 3587–3593. [\[CrossRef\]](http://doi.org/10.1172/JCI25151)
- 209. Maassen, J.A.; 'T Hart, L.M.; Van Essen, E.; Heine, R.J.; Nijpels, G.; Jahangir Tafrechi, R.S.; Raap, A.K.; Janssen, G.M.C.; Lemkes, H.H.P.J. Mitochondrial Diabetes: Molecular Mechanisms and Clinical Presentation. *Diabetes* **2004**, *53* (Suppl. S1), S103–S109. [\[CrossRef\]](http://doi.org/10.2337/diabetes.53.2007.S103)
- 210. Cooper, S.A.; Whaley-Connell, A.; Habibi, J.; Wei, Y.; Lastra, G.; Manrique, C.; Stas, S.; Sowers, J.R. Renin-Angiotensin-Aldosterone System and Oxidative Stress in Cardiovascular Insulin Resistance. *Am. J. Physiol. Heart Circ. Physiol.* **2007**, *293*, H2009–H2023. [\[CrossRef\]](http://doi.org/10.1152/ajpheart.00522.2007)
- 211. Khalil, M.; Shanmugam, H.; Abdallah, H.; John Britto, J.S.; Galerati, I.; Gómez-Ambrosi, J.; Frühbeck, G.; Portincasa, P. The Potential of the Mediterranean Diet to Improve Mitochondrial Function in Experimental Models of Obesity and Metabolic Syndrome. *Nutrients* **2022**, *14*, 3112. [\[CrossRef\]](http://doi.org/10.3390/nu14153112)
- 212. Estruch, R.; Ros, E.; Salas-Salvadó, J.; Covas, M.-I.; Corella, D.; Arós, F.; Gómez-Gracia, E.; Ruiz-Gutiérrez, V.; Fiol, M.; Lapetra, J.; et al. Primary Prevention of Cardiovascular Disease with a Mediterranean Diet Supplemented with Extra-Virgin Olive Oil or Nuts. *N. Engl. J. Med.* **2018**, *378*, e34. [\[CrossRef\]](http://doi.org/10.1056/NEJMoa1800389)
- 213. Varela-Lopez, A.; Pérez-López, M.P.; Ramirez-Tortosa, C.L.; Battino, M.; Granados-Principal, S.; Ramirez-Tortosa, M.d.C.; Ochoa, J.J.; Vera-Ramirez, L.; Giampieri, F.; Quiles, J.L. Gene Pathways Associated with Mitochondrial Function, Oxidative Stress and Telomere Length Are Differentially Expressed in the Liver of Rats Fed Lifelong on Virgin Olive, Sunflower or Fish Oils. *J. Nutr. Biochem.* **2018**, *52*, 36–44. [\[CrossRef\]](http://doi.org/10.1016/j.jnutbio.2017.09.007)
- 214. Quiles, J.L.; Huertas, J.R.; Mañas, M.; Ochoa, J.J.; Battino, M.; Mataix, J. Oxidative Stress Induced by Exercise and Dietary Fat Modulates the Coenzyme Q and Vitamin A Balance between Plasma and Mitochondria. *Int. J. Vitam. Nutr. Research. Int. Z. Fur Vitam.-Und Ernahrungsforschung. J. Int. Vitaminol. Nutr.* **1999**, *69*, 243–249. [\[CrossRef\]](http://doi.org/10.1024/0300-9831.69.4.243)
- 215. Ochoa, J.J.; Quiles, J.L.; Ibáñez, S.; Martínez, E.; López-Frías, M.; Huertas, J.R.; Mataix, J. Aging-Related Oxidative Stress Depends on Dietary Lipid Source in Rat Postmitotic Tissues. *J. Bioenerg. Biomembr.* **2003**, *35*, 267–275. [\[CrossRef\]](http://doi.org/10.1023/A:1024615816839)
- 216. Mataix, J.; Ochoa, J.J.; Quiles, J.L. Olive Oil and Mitochondrial Oxidative Stress. International journal for vitamin and nutrition research. *Int. Z. Fur Vitam.-Und Ernahrungsforschung. J. Int. Vitaminol. Nutr.* **2006**, *76*, 178–183. [\[CrossRef\]](http://doi.org/10.1024/0300-9831.76.4.178)
- 217. Barja, G. Rate of Generation of Oxidative Stress-Related Damage and Animal Longevity. *Free Radic. Biol. Med.* **2002**, *33*, 1167–1172. [\[CrossRef\]](http://doi.org/10.1016/S0891-5849(02)00910-3)
- 218. Brown-Borg, H.M.; Rakoczy, S.G. Catalase Expression in Delayed and Premature Aging Mouse Models. *Exp. Gerontol.* **2000**, *35*, 199–212. [\[CrossRef\]](http://doi.org/10.1016/S0531-5565(00)00079-6)
- 219. Beckman, K.B.; Ames, B.N. The Free Radical Theory of Aging Matures. *Physiol. Rev.* **1998**, *78*, 547–581. [\[CrossRef\]](http://doi.org/10.1152/physrev.1998.78.2.547) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/9562038)
- 220. Sun, W.; Wang, X.; Hou, C.; Yang, L.; Li, H.; Guo, J.; Huo, C.; Wang, M.; Miao, Y.; Liu, J.; et al. Oleuropein Improves Mitochondrial Function to Attenuate Oxidative Stress by Activating the Nrf2 Pathway in the Hypothalamic Paraventricular Nucleus of Spontaneously Hypertensive Rats. *Neuropharmacology* **2017**, *113*, 556–566. [\[CrossRef\]](http://doi.org/10.1016/j.neuropharm.2016.11.010)
- 221. Schaffer, S.; Podstawa, M.; Visioli, F.; Bogani, P.; Müller, W.E.; Eckert, G.P. Hydroxytyrosol-Rich Olive Mill Wastewater Extract Protects Brain Cells in Vitro and Ex Vivo. *J. Agric. Food Chem.* **2007**, *55*, 5043–5049. [\[CrossRef\]](http://doi.org/10.1021/jf0703710) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/17530860)
- 222. Peng, Y.; Hou, C.; Yang, Z.; Li, C.; Jia, L.; Liu, J.; Tang, Y.; Shi, L.; Li, Y.; Long, J.; et al. Hydroxytyrosol Mildly Improve Cognitive Function Independent of APP Processing in APP/PS1 Mice. *Mol. Nutr. Food Res.* **2016**, *60*, 2331–2342. [\[CrossRef\]](http://doi.org/10.1002/mnfr.201600332) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/27287957)
- 223. Camilleri, A.; Vassallo, N. The Centrality of Mitochondria in the Pathogenesis and Treatment of Parkinson's Disease. *CNS Neurosci. Ther.* **2014**, *20*, 591–602. [\[CrossRef\]](http://doi.org/10.1111/cns.12264)
- 224. Lin, M.T.; Beal, M.F. Mitochondrial Dysfunction and Oxidative Stress in Neurodegenerative Diseases. *Nature* **2006**, *443*, 787–795. [\[CrossRef\]](http://doi.org/10.1038/nature05292) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/17051205)
- 225. Martín, S.; González-Burgos, E.; Carretero, M.E.; Gómez-Serranillos, M.P. Neuroprotective Properties of Spanish Red Wine and Its Isolated Polyphenols on Astrocytes. *Food Chem.* **2011**, *128*, 40–48. [\[CrossRef\]](http://doi.org/10.1016/j.foodchem.2011.02.074) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25214327)
- 226. Martín, S.; González-Burgos, E.; Carretero, M.E.; Gómez-Serranillos, M.P. Protective Effects of Merlot Red Wine Extract and Its Major Polyphenols in PC12 Cells under Oxidative Stress Conditions. *J. Food Sci.* **2013**, *78*, H112–H118. [\[CrossRef\]](http://doi.org/10.1111/1750-3841.12000) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23278327)
- 227. Qiu, L.; Luo, Y.; Chen, X. Quercetin Attenuates Mitochondrial Dysfunction and Biogenesis via Upregulated AMPK/SIRT1 Signaling Pathway in OA Rats. *Biomed. Pharmacother.* **2018**, *103*, 1585–1591. [\[CrossRef\]](http://doi.org/10.1016/j.biopha.2018.05.003) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/29864946)
- 228. Khan, M.M.; Ahmad, A.; Ishrat, T.; Khan, M.B.; Hoda, M.N.; Khuwaja, G.; Raza, S.S.; Khan, A.; Javed, H.; Vaibhav, K.; et al. Resveratrol Attenuates 6-Hydroxydopamine-Induced Oxidative Damage and Dopamine Depletion in Rat Model of Parkinson's Disease. *Brain Res.* **2010**, *1328*, 139–151. [\[CrossRef\]](http://doi.org/10.1016/j.brainres.2010.02.031)
- 229. Jardim, F.R.; de Rossi, F.T.; Nascimento, M.X.; da Silva Barros, R.G.; Borges, P.A.; Prescilio, I.C.; de Oliveira, M.R. Resveratrol and Brain Mitochondria: A Review. *Mol. Neurobiol.* **2018**, *55*, 2085–2101. [\[CrossRef\]](http://doi.org/10.1007/s12035-017-0448-z)
- 230. Kurin, E.; Muĉaji, P.; Nagy, M. In Vitro Antioxidant Activities of Three Red Wine Polyphenols and Their Mixtures: An Interaction Study. *Molecules* **2012**, *17*, 14336–14348. [\[CrossRef\]](http://doi.org/10.3390/molecules171214336) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23208468)
- 231. Fernández-Pachón, M.S.; Berná, G.; Otaolaurruchi, E.; Troncoso, A.M.; Martín, F.; García-Parrilla, M.C. Changes in Antioxidant Endogenous Enzymes (Activity and Gene Expression Levels) after Repeated Red Wine Intake. *J. Agric. Food Chem.* **2009**, *57*, 6578–6583. [\[CrossRef\]](http://doi.org/10.1021/jf901863w)
- 232. Afshordel, S.; Hagl, S.; Werner, D.; Röhner, N.; Kögel, D.; Bazan, N.G.; Eckert, G.P. Omega-3 Polyunsaturated Fatty Acids Improve Mitochondrial Dysfunction in Brain Aging - Impact of Bcl-2 and NPD-1 like Metabolites. *Prostaglandins Leukot. Essent. Fat. Acids* **2015**, *92*, 23–31. [\[CrossRef\]](http://doi.org/10.1016/j.plefa.2014.05.008)
- 233. Eckert, G.P.; Lipka, U.; Muller, W.E. Omega-3 Fatty Acids in Neurodegenerative Diseases: Focus on Mitochondria. *Prostaglandins Leukot. Essent. Fat. Acids* **2013**, *88*, 105–114. [\[CrossRef\]](http://doi.org/10.1016/j.plefa.2012.05.006)
- 234. Chauveau, F.; Cho, T.-H.; Perez, M.; Guichardant, M.; Riou, A.; Aguettaz, P.; Picq, M.; Lagarde, M.; Berthezene, Y.; Nighoghossian, N.; et al. Brain-Targeting Form of Docosahexaenoic Acid for Experimental Stroke Treatment: MRI Evaluation and Anti-Oxidant Impact. *Curr. Neurovascular Res.* **2011**, *8*, 95–102. [\[CrossRef\]](http://doi.org/10.2174/156720211795495349)
- 235. Karki, K.; Pande, D.; Negi, R.; Khanna, S.; Khanna, R.S.; Khanna, H.D. Correlation of Serum Toll like Receptor 9 and Trace Elements with Lipid Peroxidation in the Patients of Breast Diseases. *J. Trace Elem. Med. Biol. Organ Soc. Miner. Trace Elem. (GMS)* **2015**, *30*, 11–16. [\[CrossRef\]](http://doi.org/10.1016/j.jtemb.2014.12.003)
- 236. Do Val Carneiro, J.L.; Nixdorf, S.L.; Mantovani, M.S.; Da Silva Do Amaral Herrera, A.C.; Aoki, M.N.; Amarante, M.K.; Fabris, B.A.; Pelegrinelli Fungaro, M.H.; Ehara Watanabe, M.A. Plasma Malondialdehyde Levels and CXCR4 Expression in Peripheral Blood Cells of Breast Cancer Patients. *J. Cancer Res. Clin. Oncol.* **2009**, *135*, 997–1004. [\[CrossRef\]](http://doi.org/10.1007/s00432-008-0535-7)
- 237. Maia, F.M.M.; Santos, E.B.; Reis, G.E. lias Oxidative Stress and Plasma Lipoproteins in Cancer Patients. *Einstein* **2014**, *12*, 480–484. [\[CrossRef\]](http://doi.org/10.1590/S1679-45082014RC3110)
- 238. Didžiapetriene, J.; Bublevič, J.; Smailyte, G.; Kazbariene, B.; Stukas, R. Significance of Blood Serum Catalase Activity and Malondialdehyde Level for Survival Prognosis of Ovarian Cancer Patients. *Medicina* **2014**, *50*, 204–208. [\[CrossRef\]](http://doi.org/10.1016/j.medici.2014.09.001)
- 239. Chole, R.H.; Patil, R.N.; Basak, A.; Palandurkar, K.; Bhowate, R. Estimation of Serum Malondialdehyde in Oral Cancer and Precancer and Its Association with Healthy Individuals, Gender, Alcohol, and Tobacco Abuse. *J. Cancer Res. Ther.* **2010**, *6*, 487–491. [\[CrossRef\]](http://doi.org/10.4103/0973-1482.77106)
- 240. Li, Y.P.; Tian, F.G.; Shi, P.C.; Guo, L.Y.; Wu, H.M.; Chen, R.Q.; Xue, J.M. 4-Hydroxynonenal Promotes Growth and Angiogenesis of Breast Cancer Cells through HIF-1α Stabilization. *Asian Pac. J. Cancer Prev. APJCP* **2014**, *15*, 10151–10156. [\[CrossRef\]](http://doi.org/10.7314/APJCP.2014.15.23.10151)
- 241. Moral, R.; Escrich, E. Influence of Olive Oil and Its Components on Breast Cancer: Molecular Mechanisms. *Molecules* **2022**, *27*, 477. [\[CrossRef\]](http://doi.org/10.3390/molecules27020477)
- 242. Rodier, F.; Campisi, J. Four Faces of Cellular Senescence. *J. Cell Biol.* **2011**, *192*, 547. [\[CrossRef\]](http://doi.org/10.1083/jcb.201009094)
- 243. Kuilman, T.; Michaloglou, C.; Mooi, W.J.; Peeper, D.S. The Essence of Senescence. *Genes Dev.* **2010**, *24*, 2463–2479. [\[CrossRef\]](http://doi.org/10.1101/gad.1971610)
- 244. Von Zglinicki, T.; Wan, T.; Miwa, S. Senescence in Post-Mitotic Cells: A Driver of Aging? *Antioxid. Redox Signal.* **2021**, *34*, 308–323. [\[CrossRef\]](http://doi.org/10.1089/ars.2020.8048)
- 245. Tchkonia, T.; Zhu, Y.; Van Deursen, J.; Campisi, J.; Kirkland, J.L. Cellular Senescence and the Senescent Secretory Phenotype: Therapeutic Opportunities. *J. Clin. Investig.* **2013**, *123*, 966–972. [\[CrossRef\]](http://doi.org/10.1172/JCI64098)
- 246. Serrano, M.; Lin, A.W.; McCurrach, M.E.; Beach, D.; Lowe, S.W. Oncogenic Ras Provokes Premature Cell Senescence Associated with Accumulation of P53 and P16INK4a. *Cell* **1997**, *88*, 593–602. [\[CrossRef\]](http://doi.org/10.1016/S0092-8674(00)81902-9)
- 247. Krishnamurthy, J.; Torrice, C.; Ramsey, M.R.; Kovalev, G.I.; Al-Regaiey, K.; Su, L.; Sharpless, N.E. Ink4a/Arf Expression Is a Biomarker of Aging. *J. Clin. Investig.* **2004**, *114*, 1299–1307. [\[CrossRef\]](http://doi.org/10.1172/JCI22475) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/15520862)
- 248. Ressler, S.; Bartkova, J.; Niederegger, H.; Bartek, J.; Scharffetter-Kochanek, K.; Jansen-Dürr, P.; Wlaschek, M. P16INK4A Is a Robust in Vivo Biomarker of Cellular Aging in Human Skin. *Aging Cell* **2006**, *5*, 379–389. [\[CrossRef\]](http://doi.org/10.1111/j.1474-9726.2006.00231.x)
- 249. Jeck, W.R.; Siebold, A.P.; Sharpless, N.E. Review: A Meta-Analysis of GWAS and Age-Associated Diseases. *Aging Cell* **2012**, *11*, 727–731. [\[CrossRef\]](http://doi.org/10.1111/j.1474-9726.2012.00871.x) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22888763)
- 250. Baker, D.J.; Childs, B.G.; Durik, M.; Wijers, M.E.; Sieben, C.J.; Zhong, J.A.; Saltness, R.; Jeganathan, K.B.; Verzosa, G.C.; Pezeshki, A.; et al. Naturally Occurring P16 Ink4a-Positive Cells Shorten Healthy Lifespan. *Nature* **2016**, *530*, 184–189. [\[CrossRef\]](http://doi.org/10.1038/nature16932) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26840489)
- 251. Xu, M.; Pirtskhalava, T.; Farr, J.N.; Weigand, B.M.; Palmer, A.K.; Weivoda, M.M.; Inman, C.L.; Ogrodnik, M.B.; Hachfeld, C.M.; Fraser, D.G.; et al. Senolytics Improve Physical Function and Increase Lifespan in Old Age. *Nat. Med.* **2018**, *24*, 1246–1256. [\[CrossRef\]](http://doi.org/10.1038/s41591-018-0092-9)
- 252. López-Uriarte Patricia, P.; Nogués, R.; Saez, G.; Bulló, M.; Romeu, M.; Masana, L.; Tormos, C.; Casas-Agustench, P.; Salas-Salvadó, J. Effect of Nut Consumption on Oxidative Stress and the Endothelial Function in Metabolic Syndrome. *Clin. Nutr.* **2010**, *29*, 373–380. [\[CrossRef\]](http://doi.org/10.1016/j.clnu.2009.12.008) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/20064680)
- 253. Beauchamp, G.K.; Keast, R.S.J.; Morel, D.; Lin, J.; Pika, J.; Han, Q.; Lee, C.H.; Smith, A.B.; Breslin, P.A.S. Ibuprofen-like Activity in Extra-Virgin Olive Oil. *Nature* **2005**, *437*, 45–46. [\[CrossRef\]](http://doi.org/10.1038/437045a) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/16136122)
- 254. Rodríguez-Morató, J.; Xicota, L.; Fitó, M.; Farré, M.; Dierssen, M.; De La Torre, R. Potential Role of Olive Oil Phenolic Compounds in the Prevention of Neurodegenerative Diseases. *Molecules* **2015**, *20*, 4655–4680. [\[CrossRef\]](http://doi.org/10.3390/molecules20034655)
- 255. Umeno, A.; Takashima, M.; Murotomi, K.; Nakajima, Y.; Koike, T.; Matsuo, T.; Yoshida, Y. Radical-Scavenging Activity and Antioxidative Effects of Olive Leaf Components Oleuropein and Hydroxytyrosol in Comparison with Homovanillic Alcohol. *J. Oleo Sci.* **2015**, *64*, 793–800. [\[CrossRef\]](http://doi.org/10.5650/jos.ess15042)
- 256. Bussian, T.J.; Aziz, A.; Meyer, C.F.; Swenson, B.L.; van Deursen, J.M.; Baker, D.J. Clearance of Senescent Glial Cells Prevents Tau-Dependent Pathology and Cognitive Decline. *Nature* **2018**, *562*, 578–582. [\[CrossRef\]](http://doi.org/10.1038/s41586-018-0543-y) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30232451)
- 257. Gaikwad, S.; Puangmalai, N.; Bittar, A.; Montalbano, M.; Garcia, S.; McAllen, S.; Bhatt, N.; Sonawane, M.; Sengupta, U.; Kayed, R. Tau Oligomer Induced HMGB1 Release Contributes to Cellular Senescence and Neuropathology Linked to Alzheimer's Disease and Frontotemporal Dementia. *Cell Rep.* **2021**, *36*, 109419. [\[CrossRef\]](http://doi.org/10.1016/j.celrep.2021.109419) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/34289368)
- 258. Hou, Y.; Wei, Y.; Lautrup, S.; Yang, B.; Wang, Y.; Cordonnier, S.; Mattson, M.P.; Croteau, D.L.; Bohr, V.A. NAD+ Supplementation Reduces Neuroinflammation and Cell Senescence in a Transgenic Mouse Model of Alzheimer's Disease via CGAS-STING. *Proc. Natl. Acad. Sci. USA* **2021**, *118*, e2011226118. [\[CrossRef\]](http://doi.org/10.1073/pnas.2011226118)
- 259. Li, Y.; Lu, J.; Cao, X.; Zhao, H.; Gao, L.; Xia, P.; Pei, G. A Newly Synthesized Rhamnoside Derivative Alleviates Alzheimer's Amyloid- β-Induced Oxidative Stress, Mitochondrial Dysfunction, and Cell Senescence through Upregulating SIRT3. *Oxidative Med. Cell. Longev.* **2020**, *2020*, 7698560. [\[CrossRef\]](http://doi.org/10.1155/2020/7698560)
- 260. Shang, D.; Hong, Y.; Xie, W.; Tu, Z.; Xu, J. Interleukin-1β Drives Cellular Senescence of Rat Astrocytes Induced by Oligomerized Amyloid β Peptide and Oxidative Stress. *Front. Neurol.* **2020**, *11*, 929. [\[CrossRef\]](http://doi.org/10.3389/fneur.2020.00929)
- 261. Xie, Y.Y.; Pan, T.T.; Xu, D.E.; Huang, X.; Tang, Y.; Huang, W.; Chen, R.; Lu, L.; Chi, H.; Ma, Q.H. Clemastine Ameliorates Myelin Deficits via Preventing Senescence of Oligodendrocytes Precursor Cells in Alzheimer's Disease Model Mouse. *Front. Cell Dev. Biol.* **2021**, *9*, 733945. [\[CrossRef\]](http://doi.org/10.3389/fcell.2021.733945)
- 262. Hu, Y.; Fryatt, G.L.; Ghorbani, M.; Obst, J.; Menassa, D.A.; Martin-Estebane, M.; Muntslag, T.A.O.; Olmos-Alonso, A.; Guerrero-Carrasco, M.; Thomas, D.; et al. Replicative Senescence Dictates the Emergence of Disease-Associated Microglia and Contributes to Aβ Pathology. *Cell Rep.* **2021**, *35*, 109228. [\[CrossRef\]](http://doi.org/10.1016/j.celrep.2021.109228) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/34107254)
- 263. Mikuła-Pietrasik, J.; Sosińska, P.; Murias, M.; Michalak, M.; Wierzchowski, M.; Piechota, M.; Sikora, E.; Ksiézek, K. Resveratrol Derivative, 3,3',4,4'-Tetrahydroxy-Trans-Stilbene, Retards Senescence of Mesothelial Cells via Hormetic-Like Prooxidative Mechanism. *J. Gerontol.-Ser. A Biol. Sci. Med. Sci.* **2014**, *70*, 1169–1180. [\[CrossRef\]](http://doi.org/10.1093/gerona/glu172) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25238775)
- 264. Peltz, L.; Gomez, J.; Marquez, M.; Alencastro, F.; Atashpanjeh, N.; Quang, T.; Bach, T.; Zhao, Y. Resveratrol Exerts Dosage and Duration Dependent Effect on Human Mesenchymal Stem Cell Development. *PLoS ONE* **2012**, *7*, 37162. [\[CrossRef\]](http://doi.org/10.1371/journal.pone.0037162)
- 265. Hickson, L.T.J.; Langhi Prata, L.G.P.; Bobart, S.A.; Evans, T.K.; Giorgadze, N.; Hashmi, S.K.; Herrmann, S.M.; Jensen, M.D.; Jia, Q.; Jordan, K.L.; et al. Senolytics Decrease Senescent Cells in Humans: Preliminary Report from a Clinical Trial of Dasatinib plus Quercetin in Individuals with Diabetic Kidney Disease. *EBioMedicine* **2019**, *47*, 446–456. [\[CrossRef\]](http://doi.org/10.1016/j.ebiom.2019.08.069) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31542391)
- 266. Mehdizadeh, M.; Aguilar, M.; Thorin, E.; Ferbeyre, G.; Nattel, S. The Role of Cellular Senescence in Cardiac Disease: Basic Biology and Clinical Relevance. *Nat. Rev. Cardiol.* **2022**, *19*, 250–264. [\[CrossRef\]](http://doi.org/10.1038/s41569-021-00624-2) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/34667279)
- 267. Liberale, L.; Montecucco, F.; Tardif, J.C.; Libby, P.; Camici, G.G. Inflamm-Ageing: The Role of Inflammation in Age-Dependent Cardiovascular Disease. *Eur. Heart J.* **2020**, *41*, 2974–2982. [\[CrossRef\]](http://doi.org/10.1093/eurheartj/ehz961) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32006431)
- 268. Di Micco, R.; Krizhanovsky, V.; Baker, D.; d'Adda di Fagagna, F. Cellular Senescence in Ageing: From Mechanisms to Therapeutic Opportunities. *Nat. Rev. Mol. Cell Biol.* **2021**, *22*, 75–95. [\[CrossRef\]](http://doi.org/10.1038/s41580-020-00314-w)
- 269. Paez-Ribes, M.; González-Gualda, E.; Doherty, G.J.; Muñoz-Espín, D. Targeting Senescent Cells in Translational Medicine. *EMBO Mol. Med.* **2019**, *11*, e10234. [\[CrossRef\]](http://doi.org/10.15252/emmm.201810234)
- 270. Collado, M.; Serrano, M. Senescence in Tumours: Evidence from Mice and Humans. *Nat. Rev. Cancer* **2010**, *10*, 51–57. [\[CrossRef\]](http://doi.org/10.1038/nrc2772)
- 271. Ewald, J.A.; Desotelle, J.A.; Wilding, G.; Jarrard, D.F. Therapy-Induced Senescence in Cancer. *J. Natl. Cancer Inst.* **2010**, *102*, 1536–1546. [\[CrossRef\]](http://doi.org/10.1093/jnci/djq364)
- 272. Yousefzadeh, M.J.; Zhu, Y.; McGowan, S.J.; Angelini, L.; Fuhrmann-Stroissnigg, H.; Xu, M.; Ling, Y.Y.; Melos, K.I.; Pirtskhalava, T.; Inman, C.L.; et al. Fisetin Is a Senotherapeutic That Extends Health and Lifespan. *EBioMedicine* **2018**, *36*, 18–28. [\[CrossRef\]](http://doi.org/10.1016/j.ebiom.2018.09.015)
- 273. Grosse, L.; Wagner, N.; Emelyanov, A.; Molina, C.; Lacas-Gervais, S.; Wagner, K.D.; Bulavin, D.V. Defined P16High Senescent Cell Types Are Indispensable for Mouse Healthspan. *Cell Metab.* **2020**, *32*, 87–99.e6. [\[CrossRef\]](http://doi.org/10.1016/j.cmet.2020.05.002)
- 274. Omori, S.; Wang, T.W.; Johmura, Y.; Kanai, T.; Nakano, Y.; Kido, T.; Susaki, E.A.; Nakajima, T.; Shichino, S.; Ueha, S.; et al. Generation of a P16 Reporter Mouse and Its Use to Characterize and Target P16high Cells In Vivo. *Cell Metab.* **2020**, *32*, 814–828.e6. [\[CrossRef\]](http://doi.org/10.1016/j.cmet.2020.09.006)
- 275. Mihaylova, M.M.; Sabatini, D.M.; Yilmaz, Ö.H. Dietary and Metabolic Control of Stem Cell Function in Physiology and Cancer. *Cell Stem Cell* **2014**, *14*, 292–305. [\[CrossRef\]](http://doi.org/10.1016/j.stem.2014.02.008) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24607404)
- 276. Gruver, A.L.; Hudson, L.L.; Sempowski, G.D. Immunosenescence of Ageing. *J. Pathol.* **2007**, *211*, 144–156. [\[CrossRef\]](http://doi.org/10.1002/path.2104) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/17200946)
- 277. Rossi, D.J.; Bryder, D.; Seita, J.; Nussenzweig, A.; Hoeijmakers, J.; Weissman, I.L. Deficiencies in DNA Damage Repair Limit the Function of Haematopoietic Stem Cells with Age. *Nature* **2007**, *447*, 725–729. [\[CrossRef\]](http://doi.org/10.1038/nature05862) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/17554309)
- 278. Janzen, V.; Forkert, R.; Fleming, H.E.; Saito, Y.; Waring, M.T.; Dombkowski, D.M.; Cheng, T.; DePinho, R.A.; Sharpless, N.E.; Scadden, D.T. Stem-Cell Ageing Modified by the Cyclin-Dependent Kinase Inhibitor P16INK4a. *Nature* **2006**, *443*, 421–426. [\[CrossRef\]](http://doi.org/10.1038/nature05159) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/16957735)
- 279. Flores, I.; Blasco, M.A. The Role of Telomeres and Telomerase in Stem Cell Aging. *FEBS Lett.* **2010**, *584*, 3826–3830. [\[CrossRef\]](http://doi.org/10.1016/j.febslet.2010.07.042)
- 280. Sharpless, N.E.; DePinho, R.A. How Stem Cells Age and Why This Makes Us Grow Old. Nature reviews. *Mol. Cell Biol.* **2007**, *8*, 703–713. [\[CrossRef\]](http://doi.org/10.1038/NRM2241)
- 281. Fulop, T.; Larbi, A.; Dupuis, G.; Page, A.L.; Frost, E.H.; Cohen, A.A.; Witkowski, J.M.; Franceschi, C. Immunosenescence and Inflamm-Aging As Two Sides of the Same Coin: Friends or Foes? *Front. Immunol.* **2018**, *8*, 1960. [\[CrossRef\]](http://doi.org/10.3389/fimmu.2017.01960)
- 282. Cesari, F.; Sofi, F.; Molino Lova, R.; Vannetti, F.; Pasquini, G.; Cecchi, F.; Marcucci, R.; Gori, A.M.; Macchi, C.; Boni, R.; et al. Aging Process, Adherence to Mediterranean Diet and Nutritional Status in a Large Cohort of Nonagenarians: Effects on Endothelial Progenitor Cells. *Nutr. Metab. Cardiovasc. Dis.* **2018**, *28*, 84–90. [\[CrossRef\]](http://doi.org/10.1016/j.numecd.2017.09.003)
- 283. Gulati, R.; Jevremovic, D.; Peterson, T.E.; Witt, T.A.; Kleppe, L.S.; Mueske, C.S.; Lerman, A.; Vile, R.G.; Simari, R.D. Autologous Culture-Modified Mononuclear Cells Confer Vascular Protection after Arterial Injury. *Circulation* **2003**, *108*, 1520–1526. [\[CrossRef\]](http://doi.org/10.1161/01.CIR.0000089084.48655.49)
- 284. Markoski, M.M.; Garavaglia, J.; Oliveira, A.; Olivaes, J.; Marcadenti, A. Molecular Properties of Red Wine Compounds and Cardiometabolic Benefits. *Nutr. Metab. Insights* **2016**, *9*, 51–57. [\[CrossRef\]](http://doi.org/10.4137/NMI.S32909)
- 285. Parzonko, A.; Czerwińska, M.E.; Kiss, A.K.; Naruszewicz, M. Oleuropein and Oleacein May Restore Biological Functions of Endothelial Progenitor Cells Impaired by Angiotensin II via Activation of Nrf2/Heme Oxygenase-1 Pathway. *Phytomedicine* **2013**, *20*, 1088–1094. [\[CrossRef\]](http://doi.org/10.1016/j.phymed.2013.05.002) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23809250)
- 286. Fernández-Real, J.M.; Bulló, M.; Moreno-Navarrete, J.M.; Ricart, W.; Ros, E.; Estruch, R.; Salas-Salvadó, J. A Mediterranean Diet Enriched with Olive Oil Is Associated with Higher Serum Total Osteocalcin Levels in Elderly Men at High Cardiovascular Risk. *J. Clin. Endocrinol. Metab.* **2012**, *97*, 3792–3798. [\[CrossRef\]](http://doi.org/10.1210/jc.2012-2221) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22855341)
- 287. Liu, H.; Huang, H.; Li, B.; Wu, D.; Wang, F.; Zheng, X.H.; Chen, Q.; Wu, B.; Fan, X. Olive Oil in the Prevention and Treatment of Osteoporosis after Artificial Menopause. *Clin. Interv. Aging* **2014**, *9*, 2087–2095. [\[CrossRef\]](http://doi.org/10.2147/CIA.S72006) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25506212)
- 288. Shaw, A.C.; Joshi, S.; Greenwood, H.; Panda, A.; Lord, J.M. Aging of the Innate Immune System. *Curr. Opin. Immunol.* **2010**, *22*, 507–513. [\[CrossRef\]](http://doi.org/10.1016/j.coi.2010.05.003)
- 289. Samet, I. Effect of Olive Leaf Components on the Proliferation and Viability of Hematopoietic Stem Cells. *Asian J. Biomed. Pharm.* **2014**, *4*, 1–7. [\[CrossRef\]](http://doi.org/10.15272/ajbps.v4i39.612)
- 290. Chen, W.; Kang, J.; Xia, J.; Li, Y.; Yang, B.; Chen, B.; Sun, W.; Song, X.; Xiang, W.; Wang, X.; et al. P53-Related Apoptosis Resistance and Tumor Suppression Activity in UVB-Induced Premature Senescent Human Skin Fibroblasts. *Int. J. Mol. Med.* **2008**, *21*, 645–653. [\[CrossRef\]](http://doi.org/10.3892/ijmm.21.5.645)
- 291. Marsh, D.; Dickinson, S.; Neill, G.W.; Marshall, J.F.; Hart, I.R.; Thomas, G.J. Alpha Vbeta 6 Integrin Promotes the Invasion of Morphoeic Basal Cell Carcinoma through Stromal Modulation. *Cancer Res.* **2008**, *68*, 3295–3303. [\[CrossRef\]](http://doi.org/10.1158/0008-5472.CAN-08-0174)
- 292. Kang, J.; Chen, W.; Xia, J.; Li, Y.; Yang, B.; Chen, B.; Sun, W.; Song, X.; Xiang, W.; Wang, X.; et al. Extracellular Matrix Secreted by Senescent Fibroblasts Induced by UVB Promotes Cell Proliferation in HaCaT Cells through PI3K/AKT and ERK Signaling Pathways. *Int. J. Mol. Med.* **2008**, *21*, 777–784. [\[CrossRef\]](http://doi.org/10.3892/ijmm.21.6.777)
- 293. Baxter, R.A. Anti-Aging Properties of Resveratrol: Review and Report of a Potent New Antioxidant Skin Care Formulation. *J. Cosmet. Dermatol.* **2008**, *7*, 2–7. [\[CrossRef\]](http://doi.org/10.1111/j.1473-2165.2008.00354.x)
- 294. Han, X.; Zhang, T.; Liu, H.; Mi, Y.; Gou, X. Astrocyte Senescence and Alzheimer's Disease: A Review. *Front. Aging Neurosci.* **2020**, *12*, 148. [\[CrossRef\]](http://doi.org/10.3389/fnagi.2020.00148)
- 295. Yurko-Mauro, K. Cognitive and Cardiovascular Benefits of Docosahexaenoic Acid in Aging and Cognitive Decline. *Curr. Alzheimer Res.* **2010**, *7*, 190–196. [\[CrossRef\]](http://doi.org/10.2174/156720510791050911)
- 296. Ajith, T.A. A Recent Update on the Effects of Omega-3 Fatty Acids in Alzheimer's Disease. *Curr. Clin. Pharmacol.* **2018**, *13*, 252–260. [\[CrossRef\]](http://doi.org/10.2174/1574884713666180807145648)
- 297. Colizzi, C. The Protective Effects of Polyphenols on Alzheimer's Disease: A Systematic Review. *Alzheimer's Dement. Transl. Res. Clin. Interv.* **2019**, *5*, 184. [\[CrossRef\]](http://doi.org/10.1016/j.trci.2018.09.002) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31194101)
- 298. Gaudreault, R.; Mousseau, N. Mitigating Alzheimer's Disease with Natural Polyphenols: A Review. *Curr. Alzheimer Res.* **2019**, *16*, 529–543. [\[CrossRef\]](http://doi.org/10.2174/1567205016666190315093520) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30873922)
- 299. Zheng, Q.; Kebede, M.T.; Kemeh, M.M.; Islam, S.; Lee, B.; Bleck, S.D.; Wurfl, L.A.; Lazo, N.D. Inhibition of the Self-Assembly of Aβ and of Tau by Polyphenols: Mechanistic Studies. *Molecules* **2019**, *24*, 2316. [\[CrossRef\]](http://doi.org/10.3390/molecules24122316)
- 300. Ayaz, M.; Sadiq, A.; Junaid, M.; Ullah, F.; Ovais, M.; Ullah, I.; Ahmed, J.; Shahid, M. Flavonoids as Prospective Neuroprotectants and Their Therapeutic Propensity in Aging Associated Neurological Disorders. *Front. Aging Neurosci.* **2019**, *11*, 155. [\[CrossRef\]](http://doi.org/10.3389/fnagi.2019.00155)
- 301. Mittelbrunn, M.; Sánchez-Madrid, F. Intercellular Communication: Diverse Structures for Exchange of Genetic Information. *Nat. Rev. Mol. Cell Biol.* **2012**, *13*, 328–335. [\[CrossRef\]](http://doi.org/10.1038/nrm3335)
- 302. Fransen, F.; van Beek, A.A.; Borghuis, T.; El Aidy, S.; Hugenholtz, F.; van der Gaast - de Jongh, C.; Savelkoul, H.F.J.; de Jonge, M.I.; Boekschoten, M.V.; Smidt, H.; et al. Aged Gut Microbiota Contributes to Systemical Inflammaging after Transfer to Germ-Free Mice. *Front. Immunol.* **2017**, *8*, 1385. [\[CrossRef\]](http://doi.org/10.3389/fimmu.2017.01385) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/29163474)
- 303. Szarc Vel Szic, K.; Declerck, K.; Vidaković, M.; Vanden Berghe, W. From Inflammaging to Healthy Aging by Dietary Lifestyle Choices: Is Epigenetics the Key to Personalized Nutrition? *Clin. Epigenetics* **2015**, *7*, 33. [\[CrossRef\]](http://doi.org/10.1186/s13148-015-0068-2)
- 304. Xia, S.; Zhang, X.; Zheng, S.; Khanabdali, R.; Kalionis, B.; Wu, J.; Wan, W.; Tai, X. An Update on Inflamm-Aging: Mechanisms, Prevention, and Treatment. *J. Immunol. Res.* **2016**, *2016*, 8426874. [\[CrossRef\]](http://doi.org/10.1155/2016/8426874)
- 305. Samieri, C.; Sun, Q.; Townsend, M.K.; Chiuve, S.E.; Okereke, O.I.; Willett, C.; Stampfer, M.; Grodstein, F. The Association between Dietary Patterns at Midlife and Health in Aging an Observational Study. *Ann. Intern. Med.* **2013**, *159*, 584–591. [\[CrossRef\]](http://doi.org/10.7326/0003-4819-159-9-201311050-00004)
- 306. Frasca, D.; Blomberg, B.B. Inflammaging Decreases Adaptive and Innate Immune Responses in Mice and Humans. *Biogerontology* **2016**, *17*, 7–19. [\[CrossRef\]](http://doi.org/10.1007/s10522-015-9578-8) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25921609)
- 307. Michaud, M.; Balardy, L.; Moulis, G.; Gaudin, C.; Peyrot, C.; Vellas, B.; Cesari, M.; Nourhashemi, F. Proinflammatory Cytokines, Aging, and Age-Related Diseases. *J. Am. Med. Dir. Assoc.* **2013**, *14*, 877–882. [\[CrossRef\]](http://doi.org/10.1016/j.jamda.2013.05.009)
- 308. Estruch, R.; Ngel Martínez-Gonzá Lez, M.Á.; Corella, D.; Salas-Salvadó, J.; Ruiz-Gutié Rrez, V.; Covas, M.I.; Fiol, M.; GóMez-Gracia, E.; López-Sabater, M.C. Effects of a Mediterranean-Style Diet on Cardiovascular Risk Factors A Randomized Trial. *Ann. Intern. Med.* **2006**, *145*, 1–11. [\[CrossRef\]](http://doi.org/10.7326/0003-4819-145-1-200607040-00004) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/16818923)
- 309. Salas-Salvadó, J.; Garcia-Arellano, A.; Estruch, R.; Marquez-Sandoval, F.; Corella, D.; Fiol, M.; Gómez-Gracia, E.; Viñoles, E.; Arós, F.; Herrera, C.; et al. Components of the Mediterranean-Type Food Pattern and Serum Inflammatory Markers among Patients at High Risk for Cardiovascular Disease. *Eur. J. Clin. Nutr.* **2008**, *62*, 651–659. [\[CrossRef\]](http://doi.org/10.1038/sj.ejcn.1602762)
- 310. Zhang, X.; Cao, J.; Zhong, L. Hydroxytyrosol Inhibits Pro-Inflammatory Cytokines, INOS, and COX-2 Expression in Human Monocytic Cells. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **2009**, *379*, 581–586. [\[CrossRef\]](http://doi.org/10.1007/s00210-009-0399-7) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/19198806)
- 311. Rosignoli, P.; Fuccelli, R.; Fabiani, R.; Servili, M.; Morozzi, G. Effect of Olive Oil Phenols on the Production of Inflammatory Mediators in Freshly Isolated Human Monocytes. *J. Nutr. Biochem.* **2013**, *24*, 1513–1519. [\[CrossRef\]](http://doi.org/10.1016/j.jnutbio.2012.12.011)
- 312. Dell'Agli, M.; Fagnani, R.; Galli, G.V.; Maschi, O.; Gilardi, F.; Bellosta, S.; Crestani, M.; Bosisio, E.; De Fabiani, E.; Caruso, D. Olive Oil Phenols Modulate the Expression of Metalloproteinase 9 in THP-1 Cells by Acting on Nuclear Factor-KappaB Signaling. *J. Agric. Food Chem.* **2010**, *58*, 2246–2252. [\[CrossRef\]](http://doi.org/10.1021/jf9042503)
- 313. Arango, D.; Diosa-Toro, M.; Rojas-Hernandez, L.S.; Cooperstone, J.L.; Schwartz, S.J.; Mo, X.; Jiang, J.; Schmittgen, T.D.; Doseff, A.I. Dietary Apigenin Reduces LPS-Induced Expression of MiR-155 Restoring Immune Balance during Inflammation. *Mol. Nutr. Food Res.* **2015**, *59*, 763–772. [\[CrossRef\]](http://doi.org/10.1002/mnfr.201400705)
- 314. Dell'Agli, M.; Fagnani, R.; Mitro, N.; Scurati, S.; Masciadri, M.; Mussoni, L.; Galli, G.V.; Bosisio, E.; Crestani, M.; De Fabiani, E.; et al. Minor Components of Olive Oil Modulate Proatherogenic Adhesion Molecules Involved in Endothelial Activation. *J. Agric. Food Chem.* **2006**, *54*, 3259–3264. [\[CrossRef\]](http://doi.org/10.1021/jf0529161)
- 315. Zambonin, L.; Caliceti, C.; Vieceli Dalla Sega, F.; Fiorentini, D.; Hrelia, S.; Landi, L.; Prata, C. Dietary Phenolic Acids Act as Effective Antioxidants in Membrane Models and in Cultured Cells, Exhibiting Proapoptotic Effects in Leukaemia Cells. *Oxidative Med. Cell. Longev.* **2012**, *2012*, 839298. [\[CrossRef\]](http://doi.org/10.1155/2012/839298) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22792417)
- 316. Meza-Miranda, E.R.; Rangel-Zúñiga, O.A.; Marín, C.; Pérez-Martínez, P.; Delgado-Lista, J.; Haro, C.; Peña-Orihuela, P.; Jiménez-Morales, A.I.; Malagón, M.M.; Tinahones, F.J.; et al. Virgin Olive Oil Rich in Phenolic Compounds Modulates the Expression of Atherosclerosis-Related Genes in Vascular Endothelium. *Eur. J. Nutr.* **2016**, *55*, 519–527. [\[CrossRef\]](http://doi.org/10.1007/s00394-015-0868-3) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25733165)
- 317. Camargo, A.; Ruano, J.; Fernandez, J.M.; Parnell, L.D.; Jimenez, A.; Santos-Gonzalez, M.; Marin, C.; Perez-Martinez, P.; Uceda, M.; Lopez-Miranda, J.; et al. Gene Expression Changes in Mononuclear Cells in Patients with Metabolic Syndrome after Acute Intake of Phenol-Rich Virgin Olive Oil. *BMC Genom.* **2010**, *11*, 253. [\[CrossRef\]](http://doi.org/10.1186/1471-2164-11-253) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/20406432)
- 318. Loued, S.; Berrougui, H.; Componova, P.; Ikhlef, S.; Helal, O.; Khalil, A. Extra-Virgin Olive Oil Consumption Reduces the Age-Related Decrease in HDL and Paraoxonase 1 Anti-Inflammatory Activities. *Br. J. Nutr.* **2013**, *110*, 1272–1284. [\[CrossRef\]](http://doi.org/10.1017/S0007114513000482) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23510814)
- 319. Parhizkar, S.; Holtzman, D.M. APOE Mediated Neuroinflammation and Neurodegeneration in Alzheimer's Disease. *Semin. Immunol.* **2022**, *59*, 101594. [\[CrossRef\]](http://doi.org/10.1016/j.smim.2022.101594) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/35232622)
- 320. Solch, R.J.; Aigbogun, J.O.; Voyiadjis, A.G.; Talkington, G.M.; Darensbourg, R.M.; O'Connell, S.; Pickett, K.M.; Perez, S.R.; Maraganore, D.M. Mediterranean Diet Adherence, Gut Microbiota, and Alzheimer's or Parkinson's Disease Risk: A Systematic Review. *J. Neurol. Sci.* **2022**, *434*, 120166. [\[CrossRef\]](http://doi.org/10.1016/j.jns.2022.120166)
- 321. Ciulla, M.; Marinelli, L.; Cacciatore, I.; Di Stefano, A. Role of Dietary Supplements in the Management of Parkinson's Disease. *Biomolecules* **2019**, *9*, 271. [\[CrossRef\]](http://doi.org/10.3390/biom9070271)
- 322. Abdul-Latif, R.; Stupans, I.; Allahham, A.; Adhikari, B.; Thrimawithana, T. Natural Antioxidants in the Management of Parkinson's Disease: Review of Evidence from Cell Line and Animal Models. *J. Integr. Med.* **2021**, *19*, 300–310. [\[CrossRef\]](http://doi.org/10.1016/j.joim.2021.03.007)
- 323. Guo, J.; Huang, X.; Dou, L.; Yan, M.; Shen, T.; Tang, W.; Li, J. Aging and Aging-Related Diseases: From Molecular Mechanisms to Interventions and Treatments. *Signal Transduct. Target. Ther.* **2022**, *7*, 391. [\[CrossRef\]](http://doi.org/10.1038/s41392-022-01251-0)
- 324. Jia, S.; Xu, X.; Zhou, S.; Chen, Y.; Ding, G.; Cao, L. Fisetin Induces Autophagy in Pancreatic Cancer Cells via Endoplasmic Reticulum Stress- and Mitochondrial Stress-Dependent Pathways. *Cell Death Dis.* **2019**, *10*, 142. [\[CrossRef\]](http://doi.org/10.1038/s41419-019-1366-y) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30760707)
- 325. Tang, S.M.; Deng, X.T.; Zhou, J.; Li, Q.P.; Ge, X.X.; Miao, L. Pharmacological Basis and New Insights of Quercetin Action in Respect to Its Anti-Cancer Effects. *Biomed. Pharmacother.* **2020**, *121*, 109604. [\[CrossRef\]](http://doi.org/10.1016/j.biopha.2019.109604) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31733570)
- 326. Borzì, A.M.; Biondi, A.; Basile, F.; Luca, S.; Vicari, E.S.D.; Vacante, M. Olive Oil Effects on Colorectal Cancer. *Nutrients* **2019**, *11*, 32. [\[CrossRef\]](http://doi.org/10.3390/nu11010032)
- 327. Giner, E.; Andújar, I.; Recio, M.C.; Ríos, J.L.; Cerdá-Nicolás, J.M.; Giner, R.M. Oleuropein Ameliorates Acute Colitis in Mice. *J. Agric. Food Chem.* **2011**, *59*, 12882–12892. [\[CrossRef\]](http://doi.org/10.1021/jf203715m) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22114936)
- 328. Giner, E.; Recio, M.C.; Ríos, J.L.; Cerdá-Nicolás, J.M.; Giner, R.M. Chemopreventive Effect of Oleuropein in Colitis-Associated Colorectal Cancer in C57bl/6 Mice. *Mol. Nutr. Food Res.* **2016**, *60*, 242–255. [\[CrossRef\]](http://doi.org/10.1002/mnfr.201500605) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26502315)
- 329. Andrewes, P.; Busch, J.L.H.C.; De Joode, T.; Groenewegen, A.; Alexandre, H. Sensory Properties of Virgin Olive Oil Polyphenols: Identification of Deacetoxy-Ligstroside Aglycon as a Key Contributor to Pungency. *J. Agric. Food Chem.* **2003**, *51*, 1415–1420. [\[CrossRef\]](http://doi.org/10.1021/jf026042j) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/12590491)
- 330. García Rodríguez, L.A.; Huerta-Alvarez, C. Reduced Risk of Colorectal Cancer among Long-Term Users of Aspirin and Nonaspirin Nonsteroidal Antiinflammatory Drugs. *Epidemiology* **2001**, *12*, 88–93. [\[CrossRef\]](http://doi.org/10.1097/00001648-200101000-00015)
- 331. Godos, J.; Biondi, A.; Galvano, F.; Basile, F.; Sciacca, S.; Giovannucci, E.L.; Grosso, G. Markers of Systemic Inflammation and Colorectal Adenoma Risk: Meta-Analysis of Observational Studies. *World J. Gastroenterol.* **2017**, *23*, 1909–1919. [\[CrossRef\]](http://doi.org/10.3748/wjg.v23.i10.1909)
- 332. Santangelo, C.; Varì, R.; Scazzocchio, B.; Filesi, C. Polyphenols, Intracellular Signalling and Inflammation. *Ann.-Ist. Super. Sanita* **2007**, *43*, 394.
- 333. Jandhyala, S.M.; Talukdar, R.; Subramanyam, C.; Vuyyuru, H.; Sasikala, M.; Reddy, D.N. Role of the Normal Gut Microbiota. *World J. Gastroenterol.* **2015**, *21*, 8836–8847. [\[CrossRef\]](http://doi.org/10.3748/wjg.v21.i29.8787)
- 334. Frank, D.N.; St Amand, A.L.; Feldman, R.A.; Boedeker, E.C.; Harpaz, N.; Pace, N.R. Molecular-Phylogenetic Characterization of Microbial Community Imbalances in Human Inflammatory Bowel Diseases. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 13780–13785. [\[CrossRef\]](http://doi.org/10.1073/pnas.0706625104)
- 335. Power, S.E.; O'Toole, P.W.; Stanton, C.; Ross, R.P.; Fitzgerald, G.F. Intestinal Microbiota, Diet and Health. *Br. J. Nutr.* **2014**, *111*, 387–402. [\[CrossRef\]](http://doi.org/10.1017/S0007114513002560)
- 336. Arias, A.; Mach, N. Efecto de Los Probióticos En El Control de La Obesidad En Humanos: Hipótesis No Demostradas. *Rev. Esp. Nutr. Hum. Y Diet.* **2012**, *16*, 100–107. [\[CrossRef\]](http://doi.org/10.14306/renhyd.16.3.75)
- 337. Castillo-Álvarez, F.; Marzo-Sola, M.E. Papel de La Microbiota Intestinal En El Desarrollo de Diferentes Enfermedades Neurológicas. *Neurología* **2022**, *37*, 492–498. [\[CrossRef\]](http://doi.org/10.1016/j.nrl.2019.03.017) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/35779869)
- 338. Jeffery, I.B.; Lynch, D.B.; O'Toole, P.W. Composition and Temporal Stability of the Gut Microbiota in Older Persons. *ISME J.* **2016**, *10*, 170–182. [\[CrossRef\]](http://doi.org/10.1038/ismej.2015.88) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26090993)
- 339. Alang, N.; Kelly, C.R. Weight Gain after Fecal Microbiota Transplantation. *Open Forum Infect. Dis.* **2015**, *2*, ofv004. [\[CrossRef\]](http://doi.org/10.1093/ofid/ofv004) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26034755)
- 340. Mills, S.; Stanton, C.; Lane, J.A.; Smith, G.J.; Ross, R.P. Precision Nutrition and the Microbiome, Part I: Current State of the Science. *Nutrients* **2019**, *11*, 923. [\[CrossRef\]](http://doi.org/10.3390/nu11040923)
- 341. Rondanelli, M.; Giacosa, A.; Faliva, M.A.; Perna, S.; Allieri, F.; Castellazzi, A.M. Review on Microbiota and Effectiveness of Probiotics Use in Older. *World J. Clin. Cases* **2015**, *3*, 156–162. [\[CrossRef\]](http://doi.org/10.12998/wjcc.v3.i2.156)
- 342. Claesson, M.J.; Cusack, S.; O'Sullivan, O.; Greene-Diniz, R.; De Weerd, H.; Flannery, E.; Marchesi, J.R.; Falush, D.; Dinan, T.; Fitzgerald, G.; et al. Composition, Variability, and Temporal Stability of the Intestinal Microbiota of the Elderly. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 4586–4591. [\[CrossRef\]](http://doi.org/10.1073/pnas.1000097107)
- 343. Nagpal, R.; Mainali, R.; Ahmadi, S.; Wang, S.; Singh, R.; Kavanagh, K.; Kitzman, D.W.; Kushugulova, A.; Marotta, F.; Yadav, H. Gut Microbiome and Aging: Physiological and Mechanistic Insights. *Nutr. Healthy Aging* **2018**, *4*, 267–285. [\[CrossRef\]](http://doi.org/10.3233/NHA-170030)
- 344. Kaliannan, K.; Wang, B.; Li, X.Y.; Kim, K.J.; Kang, J.X. A Host-Microbiome Interaction Mediates the Opposing Effects of Omega-6 and Omega-3 Fatty Acids on Metabolic Endotoxemia. *Sci. Rep.* **2015**, *5*. [\[CrossRef\]](http://doi.org/10.1038/srep11276)
- 345. David, L.A.; Maurice, C.F.; Carmody, R.N.; Gootenberg, D.B.; Button, J.E.; Wolfe, B.E.; Ling, A.V.; Devlin, A.S.; Varma, Y.; Fischbach, M.A.; et al. Diet Rapidly and Reproducibly Alters the Human Gut Microbiome. *Nature* **2014**, *505*, 559–563. [\[CrossRef\]](http://doi.org/10.1038/nature12820)
- 346. Clemente, J.C.; Ursell, L.K.; Parfrey, L.W.; Knight, R. The Impact of the Gut Microbiota on Human Health: An Integrative View. *Cell* **2012**, *148*, 1258–1270. [\[CrossRef\]](http://doi.org/10.1016/j.cell.2012.01.035) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22424233)
- 347. Richards, J.L.; Yap, Y.A.; McLeod, K.H.; MacKay, C.R.; Marinõ, E. Dietary Metabolites and the Gut Microbiota: An Alternative Approach to Control Inflammatory and Autoimmune Diseases. *Clin. Transl. Immunol.* **2016**, *5*, e82. [\[CrossRef\]](http://doi.org/10.1038/cti.2016.29) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/27350881)
- 348. Sreng, N.; Champion, S.; Martin, J.C.; Khelaifia, S.; Christensen, J.E.; Padmanabhan, R.; Azalbert, V.; Blasco-Baque, V.; Loubieres, P.; Pechere, L.; et al. Resveratrol-Mediated Glycemic Regulation Is Blunted by Curcumin and Is Associated to Modulation of Gut Microbiota. *J. Nutr. Biochem.* **2019**, *72*, 108218. [\[CrossRef\]](http://doi.org/10.1016/j.jnutbio.2019.108218)
- 349. Thorburn, A.N.; Macia, L.; Mackay, C.R. Diet, Metabolites, and "Western-Lifestyle" Inflammatory Diseases. *Immunity* **2014**, *40*, 833–842. [\[CrossRef\]](http://doi.org/10.1016/j.immuni.2014.05.014)
- 350. Tang, W.H.W.; Wang, Z.; Levison, B.S.; Koeth, R.A.; Britt, E.B.; Fu, X.; Wu, Y.; Hazen, S.L. Intestinal Microbial Metabolism of Phosphatidylcholine and Cardiovascular Risk. *N. Engl. J. Med.* **2013**, *368*, 1575–1584. [\[CrossRef\]](http://doi.org/10.1056/NEJMoa1109400)
- 351. Zhu, W.; Gregory, J.C.; Org, E.; Buffa, J.A.; Gupta, N.; Wang, Z.; Li, L.; Fu, X.; Wu, Y.; Mehrabian, M.; et al. Gut Microbial Metabolite TMAO Enhances Platelet Hyperreactivity and Thrombosis Risk. *Cell* **2016**, *165*, 111–124. [\[CrossRef\]](http://doi.org/10.1016/j.cell.2016.02.011)
- 352. Cornejo-Pareja, I.; Muñoz-Garach, A.; Clemente-Postigo, M.; Tinahones, F.J. Importance of Gut Microbiota in Obesity. *Eur. J. Clin. Nutr.* **2019**, *72*, 26–37. [\[CrossRef\]](http://doi.org/10.1038/s41430-018-0306-8)
- 353. Anand, P.; Kunnumakara, A.B.; Sundaram, C.; Harikumar, K.B.; Tharakan, S.T.; Lai, O.S.; Sung, B.; Aggarwal, B.B. Cancer Is a Preventable Disease That Requires Major Lifestyle Changes. *Pharm. Res.* **2008**, *25*, 2097–2116. [\[CrossRef\]](http://doi.org/10.1007/s11095-008-9661-9) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/18626751)
- 354. IARC Working Group on the Evaluation of Carcinogenic Risks to Humans Biological Agents. Volume 100 B. A Review of Human Carcinogens. *Iarc Monogr. Eval. Carcinog. Risks Hum.* **2012**, *100*, 1.
- 355. Ostan, R.; Lanzarini, C.; Pini, E.; Scurti, M.; Vianello, D.; Bertarelli, C.; Fabbri, C.; Izzi, M.; Palmas, G.; Biondi, F.; et al. Inflammaging and Cancer: A Challenge for the Mediterranean Diet. *Nutrients* **2015**, *7*, 2589–2621. [\[CrossRef\]](http://doi.org/10.3390/nu7042589)
- 356. Louis, P.; Hold, G.L.; Flint, H.J. The Gut Microbiota, Bacterial Metabolites and Colorectal Cancer. *Nat. Rev. Microbiol.* **2014**, *12*, 661–672. [\[CrossRef\]](http://doi.org/10.1038/nrmicro3344)
- 357. Gill, C.I.R.; Rowland, I.R. Diet and Cancer: Assessing the Risk. *Br. J. Nutr.* **2002**, *88*, s73–s87. [\[CrossRef\]](http://doi.org/10.1079/BJN2002632) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/12215186)
- 358. Shin, N.R.; Whon, T.W.; Bae, J.W. Proteobacteria: Microbial Signature of Dysbiosis in Gut Microbiota. *Trends Biotechnol.* **2015**, *33*, 496–503. [\[CrossRef\]](http://doi.org/10.1016/j.tibtech.2015.06.011)
- 359. Ridlon, J.M.; Wolf, P.G.; Gaskins, H.R. Taurocholic Acid Metabolism by Gut Microbes and Colon Cancer. *Gut Microbes* **2016**, *7*, 201–215. [\[CrossRef\]](http://doi.org/10.1080/19490976.2016.1150414)
- 360. Singh, N.; Gurav, A.; Sivaprakasam, S.; Brady, E.; Padia, R.; Shi, H.; Thangaraju, M.; Prasad, P.D.; Manicassamy, S.; Munn, D.H.; et al. Activation of Gpr109a, Receptor for Niacin and the Commensal Metabolite Butyrate, Suppresses Colonic Inflammation and Carcinogenesis. *Immunity* **2014**, *40*, 128–139. [\[CrossRef\]](http://doi.org/10.1016/j.immuni.2013.12.007)
- 361. Larrosa, M.; González-Sarrías, A.; Yáñez-Gascón, M.J.; Selma, M.V.; Azorín-Ortuño, M.; Toti, S.; Tomás-Barberán, F.; Dolara, P.; Espín, J.C. Anti-Inflammatory Properties of a Pomegranate Extract and Its Metabolite Urolithin-A in a Colitis Rat Model and the Effect of Colon Inflammation on Phenolic Metabolism. *J. Nutr. Biochem.* **2010**, *21*, 717–725. [\[CrossRef\]](http://doi.org/10.1016/j.jnutbio.2009.04.012) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/19616930)
- 362. Bhatt, A.P.; Redinbo, M.R.; Bultman, S.J. The Role of the Microbiome in Cancer Development and Therapy. *CA A Cancer J. Clin.* **2017**, *67*, 326–344. [\[CrossRef\]](http://doi.org/10.3322/caac.21398) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28481406)
- 363. Martinez-Lapiscina, E.H.; Clavero, P.; Toledo, E.; Estruch, R.; Salas-Salvadó, J.; San Julián, B.; Sanchez-Tainta, A.; Ros, E.; Valls-Pedret, C.; Martinez-Gonzalez, M.A. Mediterranean Diet Improves Cognition: The PREDIMED-NAVARRA Randomised Trial. *J. Neurol Neurosurg. Psychiatry* **2013**, *84*, 1318–1325. [\[CrossRef\]](http://doi.org/10.1136/jnnp-2012-304792) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23670794)
- 364. Valls-Pedret, C.; Sala-Vila, A.; Serra-Mir, M.; Corella, D.; De La Torre, R.; Martínez-González, M.Á.; Martínez-Lapiscina, E.H.; Fitó, M.; Pérez-Heras, A.; Salas-Salvadó, J.; et al. Mediterranean Diet and Age-Related Cognitive Decline: A Randomized Clinical Trial. *JAMA Intern. Med.* **2015**, *175*, 1094–1103. [\[CrossRef\]](http://doi.org/10.1001/jamainternmed.2015.1668)
- 365. Marseglia, A.; Xu, W.; Fratiglioni, L.; Fabbri, C.; Berendsen, A.A.M.; Bialecka-Debek, A.; Jennings, A.; Gillings, R.; Meunier, N.; Caumon, E.; et al. Effect of the NU-AGE Diet on Cognitive Functioning in Older Adults: A Randomized Controlled Trial. *Front. Physiol.* **2018**, *9*, 349. [\[CrossRef\]](http://doi.org/10.3389/fphys.2018.00349)
- 366. Loughrey, D.G.; Lavecchia, S.; Brennan, S.; Lawlor, B.A.; Kelly, M.E. The Impact of the Mediterranean Diet on the Cognitive Functioning of Healthy Older Adults: A Systematic Review and Meta-Analysis. *Adv. Nutr.* **2017**, *8*, 571–586. [\[CrossRef\]](http://doi.org/10.3945/an.117.015495) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28710144)
- 367. Servick, K. Do Gut Bacteria Make a Second Home in Our Brains. *Science* 2018. [\[CrossRef\]](http://doi.org/10.1126/science.aaw0147)
- 368. Pistollato, F.; Iglesias, R.C.; Ruiz, R.; Aparicio, S.; Crespo, J.; Lopez, L.D.; Manna, P.P.; Giampieri, F.; Battino, M. Nutritional Patterns Associated with the Maintenance of Neurocognitive Functions and the Risk of Dementia and Alzheimer's Disease: A Focus on Human Studies. *Pharmacol. Res.* **2018**, *131*, 32–43. [\[CrossRef\]](http://doi.org/10.1016/j.phrs.2018.03.012) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/29555333)
- 369. Kowalski, K.; Mulak, A. Brain-Gut-Microbiota Axis in Alzheimer's Disease. *J. Neurogastroenterol. Motil.* **2019**, *25*, 48–60. [\[CrossRef\]](http://doi.org/10.5056/jnm18087) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30646475)
- 370. Sochocka, M.; Donskow-Łysoniewska, K.; Diniz, B.S.; Kurpas, D.; Brzozowska, E.; Leszek, J. The Gut Microbiome Alterations and Inflammation-Driven Pathogenesis of Alzheimer's Disease—A Critical Review. *Mol. Neurobiol.* **2019**, *56*, 1841–1851. [\[CrossRef\]](http://doi.org/10.1007/s12035-018-1188-4)
- 371. Braakman, H.M.H.; van Ingen, J. Can epilepsy be treated by antibiotics? *J. Neurol.* **2018**, *265*, 1934–1936. [\[CrossRef\]](http://doi.org/10.1007/s00415-018-8943-3) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/29931545)
- 372. Jin, M.; Qian, Z.; Yin, J.; Xu, W.; Zhou, X. The Role of Intestinal Microbiota in Cardiovascular Disease. *J. Cell. Mol. Med.* **2019**, *23*, 2343–2350. [\[CrossRef\]](http://doi.org/10.1111/jcmm.14195)
- 373. Minihane, A.M.; Vinoy, S.; Russell, W.R.; Baka, A.; Roche, H.M.; Tuohy, K.M.; Teeling, J.L.; Blaak, E.E.; Fenech, M.; Vauzour, D.; et al. Low-Grade Inflammation, Diet Composition and Health: Current Research Evidence and Its Translation. *Br. J. Nutr.* **2015**, *114*, 999–1012. [\[CrossRef\]](http://doi.org/10.1017/S0007114515002093)
- 374. Garcia-Mantrana, I.; Selma-Royo, M.; Alcantara, C.; Collado, M.C. Shifts on Gut Microbiota Associated to Mediterranean Diet Adherence and Specific Dietary Intakes on General Adult Population. *Front. Microbiol.* **2018**, *9*, 890. [\[CrossRef\]](http://doi.org/10.3389/fmicb.2018.00890)
- 375. Lange, K.W.; Guo, J.; Kanaya, S.; Lange, K.M.; Nakamura, Y.; Li, S. Medical Foods in Alzheimer's Disease. *Food Sci. Hum. Wellness* **2019**, *8*, 1–7. [\[CrossRef\]](http://doi.org/10.1016/j.fshw.2019.02.002)
- 376. Shannon, O.M.; Stephan, B.C.M.; Granic, A.; Lentjes, M.; Hayat, S.; Mulligan, A.; Brayne, C.; Khaw, K.T.; Bundy, R.; Aldred, S.; et al. Mediterranean Diet Adherence and Cognitive Function in Older UK Adults: The European Prospective Investigation into Cancer and Nutrition-Norfolk (EPIC-Norfolk) Study. *Am. J. Clin. Nutr.* **2019**, *110*, 938–948. [\[CrossRef\]](http://doi.org/10.1093/ajcn/nqz114) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31204785)
- 377. Dey, N.; Wagner, V.E.; Blanton, L.V.; Cheng, J.; Fontana, L.; Haque, R.; Ahmed, T.; Gordon, J.I. Regulators of Gut Motility Revealed by a Gnotobiotic Model of Diet-Microbiome Interactions Related to Travel. *Cell* **2015**, *163*, 95–107. [\[CrossRef\]](http://doi.org/10.1016/j.cell.2015.08.059) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26406373)

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.