

Accepted Manuscript

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PII: S0009-2509(18)30270-7
DOI: <https://doi.org/10.1016/j.ces.2018.04.056>
Reference: CES 14188

To appear in: *Chemical Engineering Science*

Received Date: 30 March 2017
Revised Date: 22 March 2018
Accepted Date: 25 April 2018

Please cite this article as: A.L.C.V. Lopes, A.F.G. Ribeiro, M.P.S. Reis, D.C.M. Silva, I. Portugal, C.M.S.G. Baptista, Distribution models for nitrophenols in a liquid-liquid system, *Chemical Engineering Science* (2018), doi: <https://doi.org/10.1016/j.ces.2018.04.056>

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Distribution models for nitrophenols in a liquid-liquid system

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Abstract

The formation of nitrophenols by-products is still of major concern for the economics and environmental impact of the industrial process of benzene (Bz) nitration to mononitrobenzene (MNB) with mixed acid (sulphuric and nitric acids). The knowledge of nitrophenol (NP) distribution ratios in the liquid-liquid mixture (D_j , $j = \{NP\}$) is desirable for process optimization and for understanding the reaction mechanisms behind nitrophenols formation.

In this study, a data-driven approach was implemented to provide prediction models for D_j of 2,4-dinitrophenol (DNP) and of 2,4,6-trinitrophenol (TNP) in a biphasic liquid system with a composition representative of the industrial processes. In the first step, screening tests were performed to identify the main variables influencing the experimental equilibrium weight fractions of nitrophenols in the aqueous phase ($w_{j,e}^A$). Subsequently two independent data sets were built for development and external validation of prediction multivariate linear regression (MLR) models, at 30 °C. The fitting results (R^2 and $R_{ad}^2 \geq 0.90$) and the prediction results ($R_{pred,DNP}^2 = 0.931$, $R_{pred,TNP}^2 = 0.908$) confirmed the quality of the $w_{j,e}^A$

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models. Statistical significant predictive MLR models were also developed for D_j (which is related with $w_{j,e}^A$), at 30 °C, with DNP evidencing a higher affinity for the organic phase (i.e. $D_{\text{DNP}} \approx 2D_{\text{TNP}}$).

Keywords: benzene nitration, distribution ratio, multivariate linear regression, nitrophenols, predictive models

2017 MSC: 00-01, 99-00.

1 1. Introduction

2 The nitration of benzene (Bz) to mononitrobenzene (MNB) in a mixture of
3 aqueous sulphuric and nitric acids (mixed acid), is an heterogeneous liquid-liquid
4 (L-L) reaction that is accompanied by the formation in a reduced concentration
5 (i.e. ppm) of undesirable and toxic nitrophenols by-products. In spite of the
6 great attention received, the production of the nitrophenols is not yet fully
7 understood (Burns and Ramshaw, 2002; Dummann et al., 2003; Hanson et al.,
8 1976; Quadros et al., 2005a). A mechanistic model for nitrophenols formation,
9 namely 2,4-dinitrophenol (DNP) and 2,4,6-trinitrophenol (TNP), has not been
10 established yet and data-driven approaches have been used to model the benzene
11 nitration (Nogueira et al., 2013; Portugal et al., 2009; Quadros et al., 2005b).

12 Another aspect is the presently unknown distribution of each NP between
13 the two liquid phases of the system, which is important for modelling the
14 system and better comprehension of reaction mechanism. Available studies for
15 the distribution of the mixture components include: the partition of Bz
16 between organic and aqueous phases characterized as a function of equilibrium
17 temperature and mole fractions in the mixed acid (Zaldivar et al., 1995); the
18 distributions of sulphuric and nitric acids aqueous phases in equilibrium with a
19 MNB organic phase, expressed in terms of weight ratio, at 298 K (Suresh
20 et al., 2009). Equilibrium data can also be found in the study of Schiefferle
21 et al. (1976) for all species in the reaction system except nitrophenols, and in
22 the works of Rahaman et al. (2007, 2010) for MNB in concentrated sulphuric
23 acid. On the other hand, experimental values for NP distributions between

24 L-L phases are available for octanol-water, cyclohexane-water, Bz-water and
 25 MNB-water systems (Abraham et al., 2000; Berthod and Carda-Broch, 2004;
 26 Azevedo, 2015). The effects of different temperatures, pH, and solute
 27 concentrations were studied on the distribution of each NP in MNB-water
 28 systems by Azevedo (2015). None of the above works studied the distribution
 29 of nitrophenols in an acid phase.

30 1.1. Fundamentals of liquid-liquid equilibrium

31 The equilibrium distribution of a solute in a closed L-L system at a given
 32 temperature (T) and pressure (P) is characterized by a certain concentration of
 33 the solute in both phases. When solutes are present at very low concentrations
 34 and the phases are mostly immiscible, it is usually considered that they do
 35 not interfere in the phases solubility (Leo et al., 1971; Sangster, 1989). In these
 36 conditions, the closed biphasic system reaches a minimum Gibbs energy in which
 37 the phase-specific contribution is given by,

$$G_i^f = \sum_i x_i^f \mu_i^f \quad (1)$$

38 with

$$\mu_i^f = \mu_i^o + RT \ln(\gamma_i^f x_i^f) \quad (2)$$

39 where x_i^f , μ_i^f and γ_i^f are the molar fraction, chemical potential and activity
 40 coefficient of the pure compound i in phase f , respectively; μ_i^o is the chemical
 41 potential of i in the standard state; and R is the universal constant (Elliott
 42 and Lira, 1999). Under equilibrium conditions, the chemical potentials in each
 43 phase, I and II , are equal, $\mu_i^I = \mu_i^{II}$, and according to Eq. (2):

$$\gamma_i^I x_i^I = \gamma_i^{II} x_i^{II} \quad (3)$$

44 In the case of dilute solutions, one can assume ideal behaviour ($\gamma_i^f \simeq 1$) and
 45 the molar fraction can be expressed as the product of the molar concentration
 46 (C_i^f) by the molar volume (V_m^f), leading to $x_i^f \simeq C_i^f V_m^f$ (Chiou et al., 1982).

47 The distribution of solutes in L-L systems at specific experimental conditions
 48 can be characterized by the thermodynamic distribution constant (K_x) and,
 49 in the case of dilute solutions, by the distribution constant (K_D) defined by
 50 (Berthod and Carda-Broch, 2004; Leo et al., 1971; Sangster, 1989):

$$K_x = \frac{x_i^I}{x_i^{II}} \quad (4)$$

$$K_D = \frac{C_i^I}{C_i^{II}} \quad (5)$$

51 For dilute solutions, when the molar volumes of phases *I* e *II* are identical
 52 K_x and K_D become similar ($K_x \simeq K_D$). The distribution constant (K_D)
 53 characterizes the partition of a solute in one chemical form (e.g. molecular,
 54 ionic, elemental) between the two phases at equilibrium. However, when the
 55 solute ionizes, precipitates, reacts or participates in complexation reactions
 56 with other species in the system, the distribution ratio (D) should be adopted
 57 instead (Berthod and Carda-Broch, 2004). This parameter is defined as the
 58 ratio between the sum of all concentrations of the chemical forms in which the
 59 solute may be present, in each phase (Berthod et al., 1999; Berthod and
 60 Carda-Broch, 2004; Berthod and Mekaoui, 2011; Ingram et al., 2011). Since
 61 the nitrophenols in this study are ionizable acids, the distribution ratio
 62 depends on the ionization extent in the aqueous phase ($AH \rightleftharpoons A^- + H^+$)
 63 and is given by,

$$D_{AH} = \frac{C_{AH}^O + C_{A^-}^O}{C_{AH}^A + C_{A^-}^A} \quad (6)$$

64 where C_{AH}^f and $C_{A^-}^f$ are the molar concentrations of solute in its molecular and

65 ionized forms in phase f , respectively. Rearranging Eq. (6) one obtain,

$$D_{\text{AH}} = \frac{K_{D,\text{AH}} + K_{D,\text{A}^-}(K_{\text{AH}}/C_{\text{H}^+}^{\text{A}})}{1 + K_{\text{AH}}/C_{\text{H}^+}^{\text{A}}} \quad (7)$$

66 where K_{AH} is the equilibrium ionization constant ($K_{\text{AH}} = (C_{\text{H}^+}^{\text{A}} C_{\text{A}^-}^{\text{A}})/C_{\text{AH}}^{\text{A}}$) and
 67 $K_{D,\text{AH}}$ and K_{D,A^-} are the distribution constants of the molecular and ionic
 68 forms of the solute (AH and A⁻, respectively).

69 The dependence of D_{AH} with pH usually exhibits a decreasing sigmoidal
 70 profile (Berthod et al., 1999) being higher at low pH, because the molecular
 71 form of the acid solute is mostly located in the organic phase. However, when
 72 the pH rises, D_{AH} decreases due to the enhanced solubility of the anionic form
 73 in the aqueous phase. Moreover, from Eq. (7), it can be seen that $D \simeq K_D$
 74 when $C_{\text{H}^+}^{\text{A}}$ is much higher than K_{AH} (Berthod et al., 1999). In the particular
 75 case of DNP, ionization can be minimized by operating at pH = 2.6 (Abraham
 76 et al., 2000), while for TNP this is achieved for pH < 0.5 (Azevedo, 2015).

77 Given the lack of detailed fundamental knowledge about reaction system,
 78 mechanistic models cannot be developed, and the characterization of the
 79 distribution of DNP and TNP in the L-L system representative of an
 80 industrial nitration process should be accomplished by developing prediction
 81 models based on experimental data.

82 1.2. Predictive Models Development

83 A systematic data-driven approach schematically represented in Fig. 1 was
 84 followed in order to obtain simple and reliable predictive models for the
 85 distribution ratio of each NP, D_j . The main steps of this approach include the
 86 identification of the main variables by conducting a set of experiments planned
 87 according to a statistical design of experiments (DOE) strategy, the
 88 development of the models following the MLR methodology, and evaluation of
 89 the fit and predictive capabilities of the MLR models.

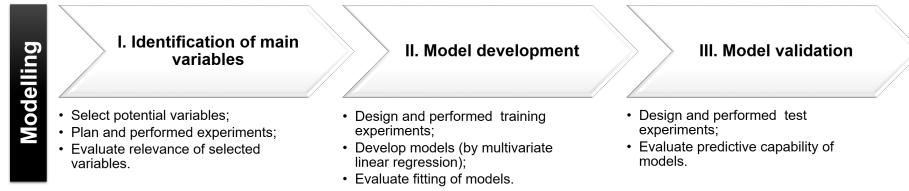


Fig. 1. Systematic approach for modelling the distribution of each NP in the L-L system.

90 The systematic approach for developing data-driven predictive models is
 91 strongly dependent of the availability or reliable and informative experimental
 92 data. In this context, DOE is a very efficient statistical methodology for
 93 planning the experiments for building and validation of predictive models
 94 (training and test sets, respectively) (Montgomery, 2001). The individual
 95 effects and the interactions between the independent variables (factors) can be
 96 estimated and evaluated with minimum effort (number of experiments) by
 97 application of a suitable experimental design.

98 Once experimental data becomes available, the conventional MLR
 99 homocedastic framework, based on the least squares method can be adopted,
 100 as long as the predictors are not highly correlated (a situation that can be
 101 avoided by selecting an appropriate DOE approach; for instance, the fractional
 102 factorial designs, used in this work, are orthogonal designs and therefore the
 103 factors do not present any mutual correlation) (Montgomery, 2012). The
 104 generic structure of the MLR models that is usually considered in DOE
 105 applications, is the following (quadratic model):

$$y_c = \hat{\beta}_0 + \sum_{g=1}^z \hat{\beta}_g x_{g,c} + \sum_{g=1}^z \hat{\beta}_{gg} x_{g,c}^2 + \sum_{g=1}^{z-1} \sum_{h=g+1}^z \hat{\beta}_{gh} x_{g,c} x_{h,c} + \epsilon \quad (8)$$

106 The error term, ϵ , is considered to have homocedastic variance, being estimate
 107 by the root mean squared error (RMSE) defined by the square root of the

108 variance, $\hat{\sigma}^2$,

$$\hat{\sigma}^2 = \frac{\sum_{c=1}^N (y_c - \hat{y}_c)^2}{N - p} \quad (9)$$

109 where y_c and \hat{y}_c are the experimental and predicted values of the response
 110 variable for observation c , respectively; $\hat{\beta}$ designates the estimates of the
 111 coefficients of the MLR terms ($\hat{\beta}_0$ is the intercept, $\hat{\beta}_g$ are the partial regression
 112 coefficients, $\hat{\beta}_{gg}$ and $\hat{\beta}_{gh}$ with $g \neq 0$ are the coefficients of the second order
 113 interaction and quadratic terms, respectively); $x_{g,c}$ and $x_{h,c}$ are the g^{th} and
 114 h^{th} regressors for observation c ; and z , p , and N are the number of regressors,
 115 of β parameters and of total observations, respectively.

116 When, upon inspection of the residuals, one verifies that the hypothesis
 117 of homocedasticity of the error term is not verified, the estimation machinery
 118 of the MLR model needs to be modified accordingly. In this case, instead of
 119 the usual least squares approach, it becomes necessary to apply a weighted
 120 least squares (WLS) scheme, that takes into consideration the heterocedastic
 121 (i.e., non-homogeneous) nature of the errors. In this method each response
 122 is weighted by a factor inversely proportional to the variance of its residual
 123 ($e_c = y_c - \hat{y}_c$), keeping unchanged the structure of the MLR model. Therefore,
 124 more importance is given to observations with lower variances than to those with
 125 higher variances (smaller weight). This consists in determining the estimates of
 126 the β parameters by minimizing the following WLS problem,

$$\sum_{c=1}^N W_c \left(y_c - \beta_0 - \sum_{g=1}^z \beta_g x_{g,c} + \sum_{g=1}^z \beta_{gg} x_{g,c}^2 + \sum_{g=1}^{z-1} \sum_{h=g+1}^z \beta_{gh} x_{g,c} x_{h,c} \right)^2 \quad (10)$$

127 with $W_c = 1/\sigma_{e_c}^2$ and where the variance was estimated from the residuals
 128 obtained in a preliminary least squares regression model fitted to the data.
 129 These residuals were grouped forming clusters and the variance of each cluster

130 was computed and used to calculate the weights of each observation, W_c .

131 The statistical significance of the conventional MLR models is usually
132 assessed following standard procedures based on hypothesis testing. Analysis
133 of variance (ANOVA) is used to evaluate the significance of the model as a
134 whole with the null hypothesis, $H_0: \beta_g = \beta_{gh} = 0$ with $g \neq 0$, and the
135 alternative hypothesis, H_1 : at least one of the β 's $\neq 0$ (with the exception of
136 β_0 , that is not considered in this test). The statistical significance is given by
137 the rejection criterion of H_0 , where the value of the test statistic F_0 is greater
138 than the *Fischer* or *F* distribution value for a certain significance level α
139 (normally $\alpha = 0,05$): $F_0 > F_{\alpha,z,N-p}$. The significance of each regression term
140 is evaluated by another type of statistical hypothesis test which checks if the
141 corresponding coefficient: is equal to zero, $\beta = 0(H_0)$, or not, $\beta \neq 0(H_1)$, for a
142 certain significance level, α (Montgomery, 2012). The rejection of H_0 in favor
143 of H_1 indicates that the regression term is significant and should be considered
144 in the model developed to explain the response. This is obtained when the
145 absolute value of the test statistic, t , is greater than the value of the t student
146 distribution: $|t| > t_{\alpha/2,N-p}$. The outcome of the statistical tests can also be
147 based on the p-value approach, with the rejection of H_0 occurring whenever
148 the p-value $< \alpha$.

149 In addition, it is important to validate the conventional MLR assumptions
150 namely, that observation errors are independent and identically distributed
151 with a normal distribution around the zero mean and with constant variance
152 σ^2 (homocedasticity). For this purpose the model residuals, e_c , which are
153 estimates of the observation errors, must be analysed (Chatterjee and Hadi,
154 2006; Montgomery, 2001). Normality can be assessed by a QQ-plot or a
155 Shapiro-Wilk test to the residual sample (SAS Institute, 2016). The
156 studentized residuals, r_c , defined in Eq. (11) for observation c , can also be

157 analysed for detection of outlier observations (Montgomery, 2012).

$$r_c = \frac{e_c}{\text{SE}(e_c)} \quad (11)$$

158 where $\text{SE}(e_c)$ is the standard error of the raw residual for observation c
 159 (SAS Institute, 2016). When this normalized residuals sample follows a
 160 Normal distribution then it is expected that about 95% of its values fall within
 161 ± 2 , and for r_c values higher than ± 3 the observations are considered potential
 162 outliers (Montgomery, 2001). Cook's distance measure ($d_c = r_c/p$) is used to
 163 assess the influence of an outlier observation in the model, which is indicated
 164 by a value of d_c higher than 1 (Montgomery, 2012). Homoscedasticity is
 165 observed by a random and an homogeneous dispersion (with no trend) of the
 166 e_c values plot against \hat{y}_c values (Montgomery, 2012).

167 The fitting quality of the conventional MLR models to the training set data
 168 is assessed by the coefficient of determination, R^2 , and the adjusted R^2 , R_{ad}^2 ,
 169 defined by:

$$R^2 = \frac{\sum_{c=1}^N (\hat{y}_c - \bar{y}_c)^2}{\sum_{c=1}^N (y_c - \bar{y}_c)^2} \quad (12)$$

$$R_{ad}^2 = \frac{\frac{\sum_{c=1}^N (y_c - \hat{y}_c)^2}{N - p}}{\frac{\sum_{c=1}^N (y_c - \bar{y}_c)^2}{N - 1}} \quad (13)$$

170 where \bar{y}_c is the mean of the N observations of the response variable. From
 171 Eqs. (12) and (13), it can be seen that $R^2 > R_{ad}^2$ with values close to the
 172 maximum value 1 indicating a good fit. The final validation of MLR models
 173 should be done using a test. This test set should be representative of the
 174 experimental conditions under analysis and independent from the training set
 175 (external validation). This validation approach is preferred to other

176 methodologies (Esbensen and Geladi, 2010) and should be followed whenever
 177 possible. In external validation, similar performance metrics to the fit quality
 178 are used to evaluate the predictive capability of the N_t experiments of the test
 179 set namely, the R^2 of prediction, R_{pred}^2 ,

$$R_{pred}^2 = \frac{\sum_{t=1}^{N_t} (\hat{y}_t - \bar{y}_t)^2}{\sum_{t=1}^{N_t} (y_t - \bar{y}_t)^2} \quad (14)$$

180 and the root mean squared error of prediction (RMSE_{pred}),

$$\text{RMSE}_{pred} = \sqrt{\frac{\sum_{t=1}^{N_t} (\hat{y}_t - y_t)^2}{N_t}} \quad (15)$$

181 also called root average squared error (RASE) (SAS Institute, 2016). Another
 182 error metric useful for model comparison is the mean absolute error of prediction
 183 (MAE_{pred}) given in Eq. (16), also designated as average absolute error (AAE)
 184 (SAS Institute, 2016).

$$\text{MAE}_{pred} = \frac{\sum_{t=1}^{N_t} |\hat{y}_t - y_t|}{N_t} \quad (16)$$

185 The aim of the present study is to build simple, robust and representative
 186 prediction models for the distribution ratios of DNP and TNP, D_j , under
 187 operating conditions as similar as possible to those prevailing in industrial
 188 nitration processes, but in the absence of reaction (i.e. with no nitric acid).
 189 The statistical software used in this work was JMP[®] Pro version 12.1.0 of
 190 SAS Institute Inc. .

191 2. Material and methods

192 During the experiments, the organic phase contained nitration grade
 193 benzene (99.994 wt%) from Gadiv Petrochemical Industries Ltd,

194 mononitrobenzene (99.995 wt%) from CUF - Químicos Industriais S.A.,
195 2,4-dinitrophenol (97 wt%) from Sigma-Aldrich, and 2,4,6-trinitrophenol
196 (99 wt%) from BDH. Other chemicals used in this work were: sulphuric acid
197 (95–97 wt%), sodium hydroxide solution (Titripur[®] 1 M) and potassium
198 dihydrogen phosphate (≥ 99.5 wt%) all from Merck, and acetonitrile (99.9 wt%)
199 acquired from VWR Chemicals. The water used in this study was obtained
200 from a Barnstead E-pure purification system.

201 Three different organic phases (Bz and MNB) were prepared, containing
202 10, 30 and 50 wt% of Bz. Precise amounts of DNP and TNP were weighted
203 (± 0.0001 g), added to the organic phase and placed in an ultrasound bath at
204 room temperature for about 10 min, in order to ensure complete dissolution of
205 nitrophenols. The aqueous phase consisted of five solutions of sulphuric acid,
206 in the range of 50–75 wt%, prepared by weighting and dilution with water.
207 The exact compositions were measured by titration with NaOH (1 M) in a 907
208 Titrand from Metrohm, equipped with a combined pH electrode iEcoTrobe
209 Plus[®] as described in the studies of Santos (2005); Nogueira (2015).

210 The experimental procedure for mixing and reaching phase equilibrium
211 followed the shake flask methodology (Berthod and Carda-Broch, 2004;
212 Sangster, 1989) using the manual agitation (≈ 2 min) and decantation times
213 (≈ 60 min) adapted and proposed by Azevedo (2015). Different volumes of
214 each phase were employed (initial weight ratio of phases in the range of 2 – 8)
215 and equilibrium temperature was reached by immersing the flasks in a F25-ED
216 thermostatic bath from Julabo (± 0.1 °C) for about 5 min. After mixing,
217 complete separation of phases was achieved by decantation followed by
218 centrifugation in a Rotanta 460 centrifuge from Hettich for 1 min at 1250 rpm.

219 The equilibrium weight fractions of nitrophenols in the aqueous phase, $w_{j,e}^A$
220 were measured by HPLC as described by Costa et al. (2013); Azevedo (2015).
221 Briefly, aliquots of the aqueous phase (ca. 0.5 g) were collected and diluted 10
222 times with water. The diluted samples (10 μ L) were injected in triplicate in
223 an Elite LaChrom HPLC from VWR Hitachi equipped with a LiChroCART[®]
224 125-4 column (5 μ m, 125 mm \times 4 mm) and a LiChroCART[®] 4-4 guard column

225 (5 μm ; 4 mm \times 4 mm), both from Merck. Calibration was done in acidic medium
 226 (1 wt% of sulphuric acid) for the NP weight fractions in the range $1\text{--}5 \times 10^{-6}$
 227 with $R_{\text{DNP}}^2 = 0.9998$ and a $R_{\text{TNP}}^2 = 0.9991$. The software EZChrom Elite
 228 version 3.1.7 from Agilent was used for peak area calculations. Capability and
 229 adequacy of the measurement system for nitrophenols in the aqueous phase was
 230 confirmed by a gauge reproducibility and repeatability (R&R) study. Weight
 231 fractions of each NP in the organic phase ($w_{j,e}^{\text{O}}$) were calculated by mass balances
 232 assuming an immiscible L-L system.

233 3. Results and Discussion

234 This work consists of three steps (Fig. 1) based on experimental data. The
 235 relevant process variables were identified in the first step. The second and third
 236 steps take advantage of this information to build and validate prediction MLR
 237 models.

238 3.1. Screening of relevant variables

239 This first step of the systematic data-driven approach (Fig. 1) was divided in
 240 four stages, in order to identify the most influential variables on the equilibrium
 241 weight fractions of nitrophenols in aqueous phase ($w_{j,e}^{\text{A}}$). The variables assessed
 242 in each stage were:

- 243 • Stage I – equilibrium temperature (T);
- 244 • Stage II – initial weight fraction of each NP in the organic phase ($w_{j,0}^{\text{O}}$);
- 245 • Stage III – initial composition of organic and aqueous phases (expressed
 246 in terms of weight fraction of Bz ($w_{\text{Bz},0}^{\text{O}}$) and sulphuric acid ($w_{\text{AS},0}^{\text{A}}$));
- 247 • Stage IV – initial weight ratio of the aqueous and organic phases
 248 ($W_0^{\text{A}}/W_0^{\text{O}}$).

249 3.1.1. Stage I – equilibrium temperature

250 The influence of temperature in the distribution of nitrophenols was
 251 assessed at six temperatures in the range 20–60 $^{\circ}\text{C}$, with $w_{\text{AS},0}^{\text{A}} = 0.65$,
 252 $w_{\text{Bz},0}^{\text{O}} = 0.50$, $W_0^{\text{A}}/W_0^{\text{O}} = 2$ and $w_{j,0}^{\text{A}} = 0.001$ for each NP. The experiments

253 were carried out in triplicate. The equilibrium weight fractions of DNP and
 254 TNP in the aqueous phase, $w_{j,e}^A$, are plotted as a function of temperature in
 255 Fig. 2. The experimental results reveal a slight reduction in $w_{\text{TNP},e}^A$ with
 256 increasing equilibrium temperature while, on the other hand, $w_{\text{DNP},e}^A$ did not
 257 vary significantly. Therefore for TNP the temperature increase could lead to a
 258 smooth increased of its affinity for organic phase. It may also be observed that
 259 $w_{\text{TNP},e}^A$ is about twice $w_{\text{DNP},e}^A$. This difference may be explained by the
 260 stronger acidity of TNP in comparison with DNP ($\text{p}K_{\text{DNP}} = 4.0$ and
 261 $\text{p}K_{\text{TNP}} = 0.4$ (Carey, 2006)). In fact, the presence of strong
 262 electron-withdrawing nitro groups bonded to the aromatic ring of NP increases
 263 the stability of the phenoxide ion by resonance.

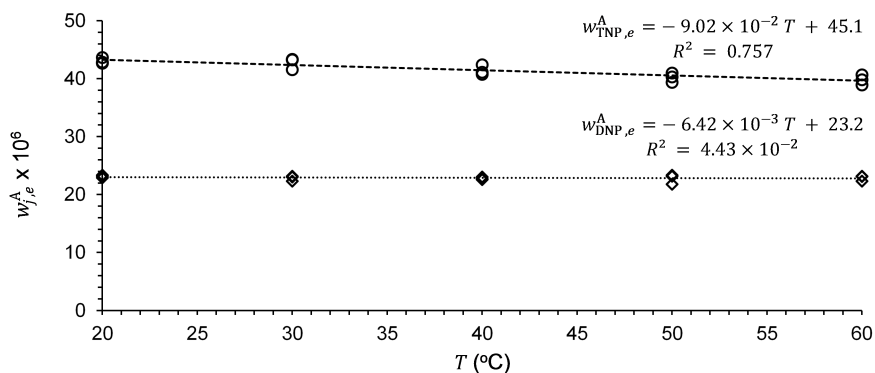


Fig. 2. Influence of temperature in the equilibrium composition of the aqueous phase (\diamond , DNP; and \circ , TNP).

264 Accordingly to these results, the effect found for temperature was weak and
 265 the experiments in the next stages were all performed at 30 °C.

266 3.1.2. Stage II – Initial weight fraction of each NP in the organic phase

267 The effect of the initial composition of each NP in the organic phase on their
 268 equilibrium distributions between phases was investigated based on a three-
 269 level complete factorial DOE, with $w_{j,0}^A$ factors in the range $0.25\text{--}2 \times 10^{-3}$. The
 270 experiments were performed in triplicate at the following conditions: $w_{\text{AS},0}^A =$
 271 0.65 , $w_{\text{Bz},0}^O = 0.5$ and $W_0^A/W_0^O = 2$. In this stage a three-level factorial was

272 implemented for better modelling of the expected curvature, as reported by
 273 Azevedo (2015). However, in this work, non-linear dependencies were not found
 274 as can be seen in Fig. 3, and therefore, in section 3.2 ahead a two level factorial
 275 will be adopted.

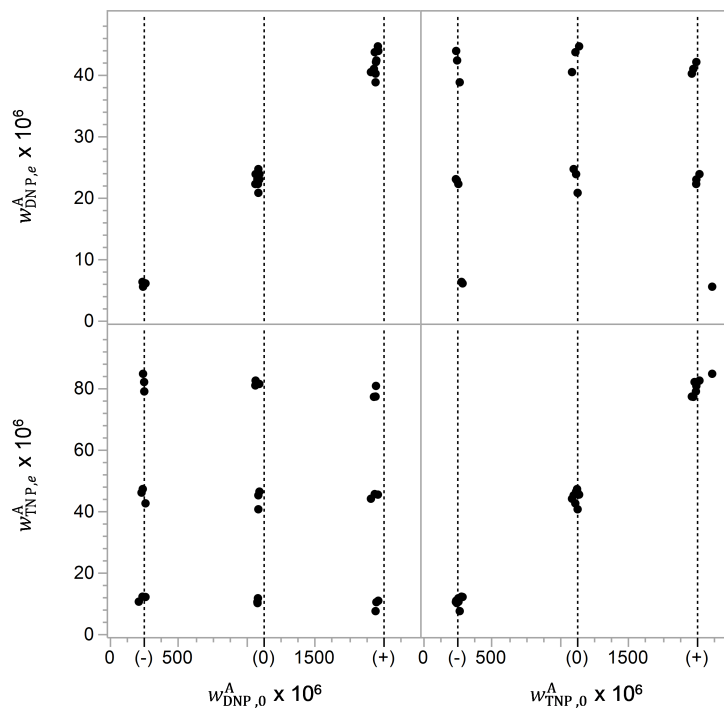


Fig. 3. Scatter plot of the equilibrium fractions in the aqueous phase in function of initial fractions in the organic phase data for DNP and TNP in the low (-), intermediate (0) and high (+) levels of the DOE.

276 Fig. 3 shows the linear associations between $w_{j,e}^A$ and $w_{j,0}^A$ with no effect of
 277 one NP in the equilibrium weight fraction of the other. Analysis of the
 278 regressors' coefficients of the MLR models (Table S.1 in Supplementary
 279 Material) revealed that these associations were positive. The quality of fits
 280 was also found to be good (R^2 and $R_{ad}^2 > 0.98$).

281 These data were also used to relate the equilibrium weight fractions of each
 282 NP in aqueous and organic phases. Fig. 4 shows that DNP has an higher
 283 affinity for the organic phase than TNP as $w_{DNP,e}^O/w_{DNP,e}^A \approx 2w_{TNP,e}^O/w_{TNP,e}^A$.

284 Therefore, for the same $w_{j,0}^O$, more TNP had been transferred once equilibrium
 285 was reached.

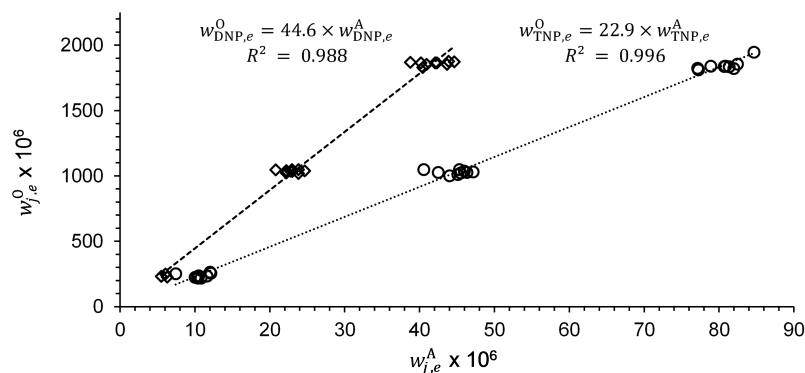


Fig. 4. Equilibrium distribution of each NP between the two phases (\diamond , DNP; and \circ , TNP).

286 3.1.3. Stage III – Initial composition of organic and aqueous phases

287 The dependence of the $w_{j,e}^A$ on the initial composition of the two phases was
 288 examined for $0.1 \leq w_{Bz,0}^O \leq 0.5$ and $0.5 \leq w_{AS,0}^A \leq 0.75$, which are within the
 289 range of industrial adiabatic nitrators operating conditions. The experiments
 290 were carried out in duplicate and designed according to a three-level complete
 291 factorial with $w_{Bz,0}^O$ and $w_{AS,0}^A$ as factors, keeping constant the initial weight
 292 fractions of nitrophenols, $w_{j,0}^O = 1 \times 10^{-3}$, and the initial weight ratio of phases,
 293 $W_0^A/W_0^O = 2$. A positive association of the $w_{j,e}^A$ with the phases composition
 294 was observed, in particular for $w_{AS,0}^A$ (Fig. S.1 in Supplementary Material). The
 295 statistical significance of the MLR model terms for the variables $w_{Bz,0}^O$ and $w_{AS,0}^A$
 296 was confirmed and the model also presented good fitting ability, as can be seen
 297 from the high values of R^2 and R_{ad}^2 (> 0.895) - see Table S.2 in Supplementary
 298 Material. The magnitude of the test statistic t for the coefficients of $w_{AS,0}^A$ and
 299 $w_{Bz,0}^O$ proved that the former is the most influential among the two.

300 The positive coefficient of $w_{AS,0}^A$ indicates that the affinity of the nitrophenols
 301 with the aqueous phase increased when this phase was richer in sulphuric acid.
 302 One possible explanation is that higher values of $w_{AS,0}^A$ lead to an increase of the
 303 electronic charge which, on the other hand, may favour the electrostatic forces

304 between the nitrophenols and the ionic species. Another interpretation is based
 305 on the solubility increment of Bz and MNB for higher sulphuric acid content
 306 (Schiefferle et al., 1976; Suresh et al., 2009). Since the nitrophenols are mostly
 307 dissolved in the organic phase in acidic conditions, they might be transferred
 308 to the aqueous phase in a higher extent due to the higher solubilities of Bz and
 309 MNB.

310 On the other hand, the positive coefficient of $w_{\text{Bz},0}^{\text{O}}$ demonstrates that the
 311 affinity of nitrophenols for organic phase was reduced by the higher content in
 312 Bz leading to an increase of $w_{j,e}^{\text{A}}$. This result may be explained by the fact that
 313 nitrophenols have a higher affinity with MNB rather than with Bz.

314 The MLR model for $w_{\text{DNP},e}^{\text{A}}$ exhibited an additional and significant quadratic
 315 term with the variable $w_{\text{AS},0}^{\text{A}}$ (see Eq. (8)) indicating that the relation between
 316 these two variables is non linear (Table S.2). The importance of this effect will
 317 be re-evaluated again in section 3.1.4.

318 3.1.4. Stage IV – Initial weight ratio of the aqueous and organic phases

319 This last stage aimed at inferring about the effect of aqueous and organic
 320 weight ratio ($W_0^{\text{A}}/W_0^{\text{O}}$) on the equilibrium compositions, $w_{j,e}^{\text{A}}$. The experiments
 321 were based on a two-level complete factorial DOE for $w_{\text{AS},0}^{\text{A}}$, $w_{\text{Bz},0}^{\text{O}}$, $w_{j,0}^{\text{O}}$ and
 322 $W_0^{\text{A}}/W_0^{\text{O}}$ factors, with $0.5 \leq w_{\text{AS},0}^{\text{A}} \leq 0.75$, $0.1 \leq w_{\text{Bz},0}^{\text{O}} \leq 0.5$, $1 \leq w_{j,0}^{\text{O}} \times 10^3 \leq 2$,
 323 and $2 \leq W_0^{\text{A}}/W_0^{\text{O}} \leq 8$.

324 The experimental data were also well fitted by MLR models (R^2 and
 325 $R_{\text{ad}}^2 > 0.98$) and the coefficients of the model terms (Table S.3 in
 326 Supplementary Material) confirmed the associations previously mentioned in
 327 stages II and III. The slightly negative relation between $w_{j,e}^{\text{A}}$ and the ratio
 328 $W_0^{\text{A}}/W_0^{\text{O}}$ (illustrated in Fig. S.2. in Supplementary Material) is consistent
 329 with the higher affinity of nitrophenols with the organic phase under acidic
 330 conditions.

331 The order of relevance (from high to low importance) of the main variables on
 332 $w_{j,e}^{\text{A}}$, ranked by the magnitude of the respective test statistic (t), was: $w_{\text{AS},0}^{\text{A}} >$
 333 $w_{j,0}^{\text{O}} > W_0^{\text{A}}/W_0^{\text{O}} > w_{\text{Bz},0}^{\text{O}}$ (Table S.3). The statistical importance of an aliased

334 quadratic term was verified for both nitrophenols. In stage III this was valid
 335 only for DNP and identified in the $w_{AS,0}^A$ term. On the other hand, for TNP
 336 the quadratic term with higher t value, although not significant, was the $w_{AS,0}^A$
 337 term. Moreover, these MLR models showed that interaction terms (Eq. (8))
 338 between $w_{AS,0}^A$ and $w_{Bz,0}^O$, and between $w_{AS,0}^A$, $w_{Bz,0}^O$ and $w_{j,0}^O$ have also significant
 339 contributions (Table S.3).

340 3.2. Equilibrium prediction models for DNP and TNP

341 Prediction models for the distribution ratio of DNP and TNP in L-L systems
 342 at equilibrium conditions were developed and validated (steps II and III of the
 343 systematic data-driven approach depicted in Fig. 1) with two independent data
 344 sets, namely training and test experiments. The experiments were carried out
 345 in the same range of initial conditions at 30 °C: $0.5 \leq w_{AS,0}^A \leq 0.75$, $0.1 \leq$
 346 $w_{Bz,0}^O \leq 0.5$, $250 \leq w_{j,0}^O \times 10^6 \leq 2000$, $2 \leq W_0^A/W_0^O \leq 8$. The training set was
 347 designed with a two-level complete factorial for $w_{AS,0}^A$, $w_{Bz,0}^O$, $w_{j,0}^O$ and W_0^A/W_0^O
 348 factors with one central point leading to 17 experimental conditions that were
 349 run in duplicate. The test set comprised 9 experiments planned by a two-level
 350 fractional factorial for the same factors, with one central point.

351 3.2.1. Multivariate linear regression models for DNP

352 The predictive MLR model for the equilibrium weight fraction $w_{DNP,e}^A$,
 353 expressed in Eq. (17), was developed with the training data set using the main
 354 variables identified in section 3.1.

$$\begin{aligned} \hat{w}_{DNP,e}^A = & -8.44 \times 10^{-5} + 1.36 \times 10^{-4} w_{AS,0}^A + 1.13 \times 10^{-5} w_{Bz,0}^O \\ & + 1.96 \times 10^{-2} w_{DNP,0}^O - 5.03 \times 10^{-7} W_0^A/W_0^O \\ & + 9.25 \times 10^{-2} (w_{AS,0}^A - 0.6239)(w_{DNP,0}^O - 1.51 \times 10^{-3}) \end{aligned} \quad (17)$$

355 The positive associations between $w_{DNP,e}^A$ and the initial compositions of
 356 both phases and of DNP ($w_{AS,0}^A$, $w_{Bz,0}^O$, $w_{DNP,0}^O$) were previously mentioned in

section 3.1.4, as well as the slightly negative relation with W_0^A/W_0^O and an interaction term between the main variables, see Eq. (17).

The distribution ratio of DNP, D_{DNP} , in the L-L system was modelled taking into consideration Eq. (6), for initial conditions expressed in molar concentrations (C), $6.946 \leq C_{\text{AS},0}^A \leq 12.42$, $1.471 \leq C_{\text{Bz},0}^O \leq 6.442$, $0.529 \leq C_{\text{DNP},0}^O \times 10^2 \leq 1.32$, $0.430 \leq C_{\text{TNP},0}^O \times 10^2 \leq 1.05$, and the ratio between phases expressed as molar ratio (n), $4.0403 \leq n_0^A/n_0^O \leq 30.255$. The prediction MLR model for \hat{D}_{DNP} given by Eq. (18) and illustrated in Fig. 5, shows that an aqueous phase with a higher sulphuric acid concentration, or an organic phase richer in Bz, lead to lower values of D_{DNP} , being $C_{\text{AS},0}^A$ the more influential factor. The MLR model for \hat{D}_{DNP} also exhibited a centred interaction term for $C_{\text{AS},0}^A$ and $C_{\text{Bz},0}^O$.

$$\begin{aligned} \hat{D}_{\text{DNP}} = & 175 - 11.7C_{\text{AS},0}^A - 3.45C_{\text{Bz},0}^O \\ & + 0.852(C_{\text{AS},0}^A - 9.682)(C_{\text{Bz},0}^O - 3.968) \end{aligned} \quad (18)$$

The statistical significance of the two prediction MLR models (ANOVA in Table S.4 of Supplementary Material) and of their terms (coefficients analysis in Table S.5 of Supplementary Material) was further confirmed by analyses of their quality of fits. The significance of the main variables followed the order $w_{\text{AS},0}^A > w_{\text{DNP},0}^O > w_{\text{Bz},0}^O > W_0^A/W_0^O$ and $C_{\text{AS},0}^A > C_{\text{Bz},0}^O$ depending on the prediction model (Table S.5). These results demonstrate that the content of sulphuric acid in the aqueous phase is the most influential variable for both $w_{\text{DNP},e}^A$ and D_{DNP} .

The prediction capability of the MLR models for DNP, in Eqs. (17) and (18), was evaluated using the test data set. The results for RMSE, quality of fit metrics R^2 and R_{ad}^2 (Eqs. (12) and (13)), and prediction capability metrics R_{pred}^2 , RMSE_{pred} and MAE_{pred} (Eqs. (15) and (16)) are presented in the Table 1. The high values of the coefficients R^2 , R_{ad}^2 and R_{pred}^2 (near or higher than 0.90) prove the high fitting and prediction capabilities of these models. In residual

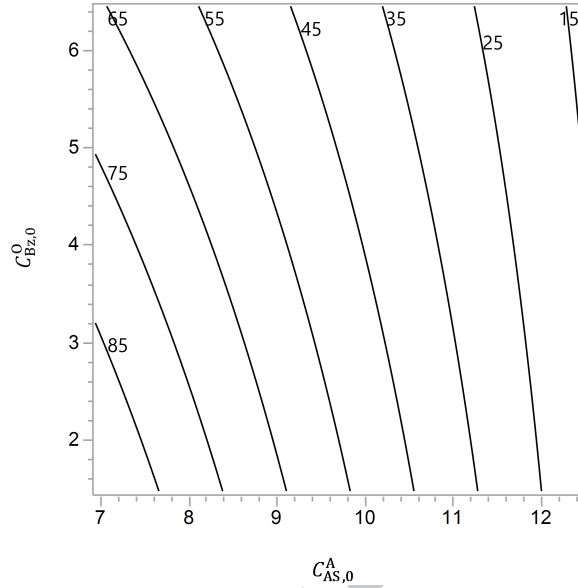


Fig. 5. Counter plot of the predicted D_{DNP} MLR model in Eq. (18).

383 analysis, the plots of r_c as a function of predicted $w_{\text{TNP},e}^A$ and D_{TNP} in Fig. 6 do
 384 not show outliers (values of r_c are within ± 3). As well influential observations
 385 were not identified (d_e values in Figs. S.3 and S.4 of Supplementary Material).

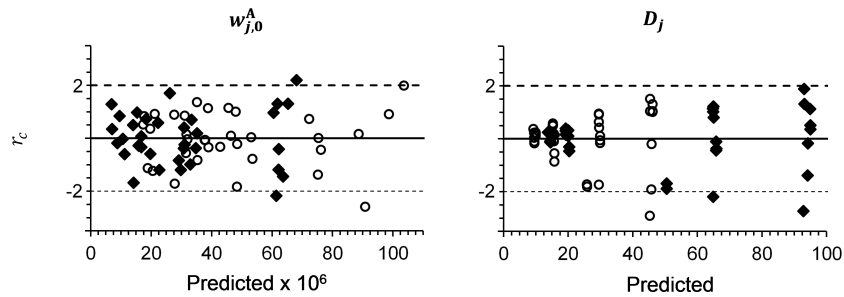


Fig. 6. Studentized residuals in function of the model values of $w_{j,e}^A$ and D_j for training set (\blacklozenge , DNP; \circ , TNP).

386 The residuals e_c for predicted values of $\hat{w}_{\text{DNP},e}^A$ exhibit an aleatory
 387 distribution and were homoscedastic (Fig. S.5 of Supplementary Material),
 388 which behaviour was reflected on r_c sample as observed in Fig. 6. The

389 normality of the e_c distribution was confirmed by a QQ-plot, with the
 390 majority of the points falling along the bisection line, and by the Shapiro-Wilk
 391 test, where the p-value was greater than α , validating the MLR assumptions
 392 (Fig. S.3). On the other hand, the e_c for \hat{D}_{DNP} presented an heteroscedastic
 393 behaviour (funnel pattern in Fig. S.5) and, as consequence, did not followed a
 394 normal distribution (Fig. S.4). Therefore, for the later model, the conventional
 395 MLR assumptions were not fully validated. This was attributed to the
 396 calculation procedure of D_{DNP} , where a small variation on the equilibrium
 397 concentration of NP in the aqueous phase ($C_{j,e}^{\text{A}}$) on the lower range, introduces
 398 a great difference in D_{DNP} values. This effect is attenuated as $C_{j,e}^{\text{A}}$ increases.

399 To deal with the heteroscedasticity of the residuals of \hat{D}_{DNP} the WLS
 400 methodology was applied. The corresponding WLS model for DNP is
 401 presented in Eq. (19):

$$\begin{aligned} \hat{D}_{\text{DNP,WLS}} = & 174 - 11.5C_{\text{AS},0}^{\text{A}} - 3.40C_{\text{Bz},0}^{\text{O}} \\ & + 0.840(C_{\text{AS},0}^{\text{A}} - 9.682)(C_{\text{Bz},0}^{\text{O}} - 3.968) \end{aligned} \quad (19)$$

402 All parameters of the $\hat{D}_{\text{DNP,WLS}}$ model fall within the confidence interval
 403 at 95% of the \hat{D}_{DNP} MLR model and the prediction results for training and
 404 test sets values (R_{pred}^2) were identical, with the WLS model being only slightly
 405 worse. Hence, the use of the conventional MLR model - Eq. (18) was validated
 406 for prediction of D_{DNP} in L-L systems similar to those industrially implemented
 407 for Bz nitration.

408 3.2.2. Multivariate linear regression models for TNP

409 The same procedure described in section 3.2.1 was adopted for the
 410 development, analysis and validation of the prediction MLR models for $w_{\text{TNP},e}^{\text{A}}$
 411 and D_{TNP} in the L-L system.

412 The MLR model for $w_{\text{TNP},e}^{\text{A}}$ - Eq. (20), exhibits similar associations between
 413 the main variables and the response variables as those previously observed in

414 the prediction models of DNP (Eqs. (17) and (18)). The coefficients in Eq. (20)
 415 confirm that higher values of $w_{AS,0}^A$ or of $w_{Bz,0}^O$ lead to higher values of $w_{TNP,e}^A$,
 416 and enable the identification of a larger number of interactions.

$$\begin{aligned}
 \hat{w}_{TNP,e}^A = & -8.98 \times 10^{-5} + 1.42 \times 10^{-4} w_{AS,0}^A + 3.03 \times 10^{-5} w_{Bz,0}^O \\
 & + 2.89 \times 10^{-2} w_{TNP,0}^O - 1.02 \times 10^{-6} W_0^A / W_0^O \\
 & + 9.91 \times 10^{-5} (w_{AS,0}^A - 0.6239)(w_{Bz,0}^O - 0.2992) \\
 & + 0.105 (w_{AS,0}^A - 0.6239)(w_{TNP,0}^O - 1.52 \times 10^{-3}) \\
 & + 2.15 \times 10^{-2} (w_{Bz,0}^O - 0.2992)(w_{TNP,0}^O - 1.52 \times 10^{-3}) \\
 & - 3.25 \times 10^{-6} (w_{Bz,0}^O - 0.2992)(W_0^A / W_0^O - 4.9060)
 \end{aligned} \tag{20}$$

417 In the MLR model for prediction of D_{TNP} expressed by Eq. (21), it is also
 418 evident that higher $C_{AS,0}^A$ or $C_{Bz,0}^O$ (i.e. $w_{AS,0}^A$ or $w_{Bz,0}^O$) lead to lower values of
 419 \hat{D}_{TNP} as shown in Fig. 7. The interaction term between $C_{AS,0}^A$ and $C_{Bz,0}^O$ has
 420 also an important contribution.

$$\begin{aligned}
 \hat{D}_{TNP} = & 79.7 - 4.71 C_{AS,0}^A - 2.23 C_{Bz,0}^O \\
 & + 0.370 (C_{AS,0}^A - 9.682)(C_{Bz,0}^O - 3.968)
 \end{aligned} \tag{21}$$

421 The statistical significance of both $w_{TNP,e}^A$ and D_{TNP} prediction models and
 422 of their terms was confirmed (results in Tables S.4 and S.5). The significance of
 423 the variables follows the same order as previously reported for the DNP models,
 424 i.e. $w_{AS,0}^A > w_{TNP,0}^O > w_{Bz,0}^O > W_0^A / W_0^O$ for $\hat{w}_{TNP,e}^A$ model and $C_{AS,0}^A > C_{Bz,0}^O$
 425 for \hat{D}_{TNP} model.

426 In residuals analysis, no outliers and influential observations were identified
 427 (see Fig. 6 and Figs. S.3 and S.4, respectively). The e_c of $\hat{w}_{TNP,e}^A$ were
 428 homocedastic and exhibit a Normal distribution (see Fig. S.5). In comparison
 429 to \hat{D}_{DNP} , the e_c of \hat{D}_{TNP} presented the same heteroscedastic pattern following
 430 a nonnormal distribution (Fig. S.5).

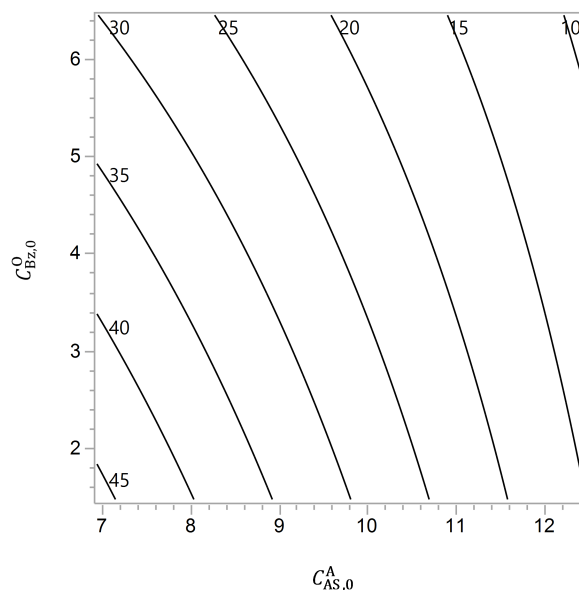


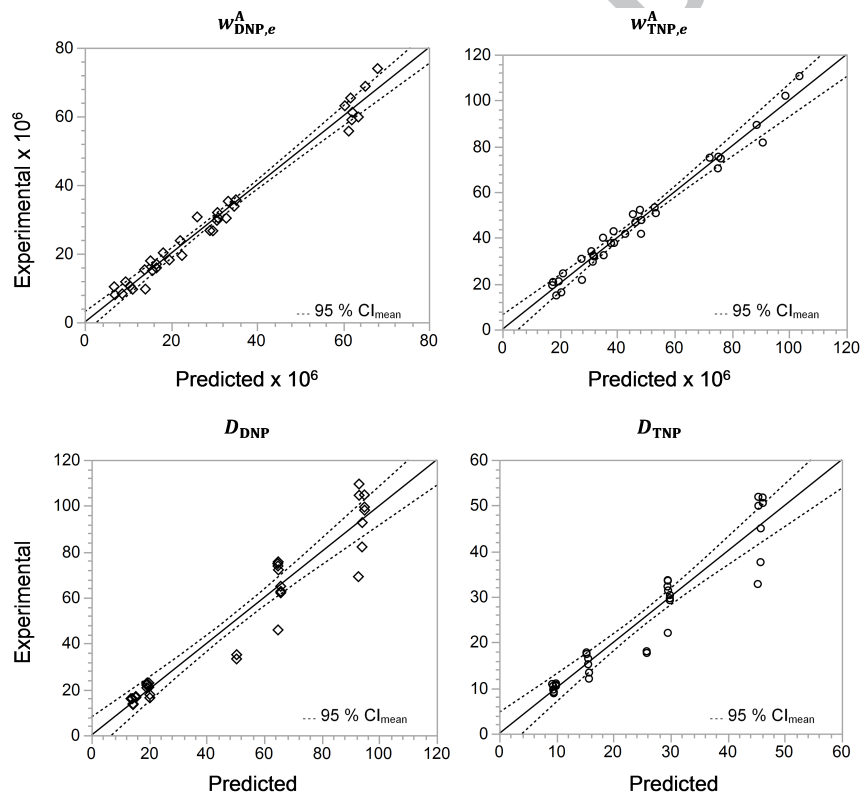
Fig. 7. Counter plot of the predicted D_{TNP} MLR model in Eq. (21).

431 The fitting and prediction capabilities of the MLR models of $w_{\text{TNP},e}^{\text{A}}$ and
 432 D_{TNP} are very satisfactory as indicated by the high values of the coefficients
 433 R^2 , R_{ad}^2 and R_{pred}^2 (>0.880) presented in Table 1. Furthermore, the linear
 434 relationship between the experimental and predicted values illustrated in Fig. 8
 435 not only for TNP but also for DNP models corroborates the high quality of the
 436 fit results.

437 Fig. 8 also shows that the experimental values of $w_{\text{TNP},e}^{\text{A}}$ are higher than
 438 the values of $w_{\text{DNP},e}^{\text{A}}$ and, consequently, the values of D_{DNP} are almost twice
 439 the values of D_{TNP} . Recalling that $D_j \approx K_{D,j}$, when nitrophenols ionization is
 440 minimized (i.e. under acidic conditions - see section 1.1) the D_j values from this
 441 work were compared with the K_D values calculated for Bz with the correlation
 442 of Zaldivar et al. (1995). The experimental D_j values differed significantly from
 443 the values calculated for $K_{D,\text{Bz}}$ for the same equilibrium conditions, with relative
 444 deviations between 77–99% for DNP and 89–100% for TNP.

Table 1Fitting and prediction capabilities of the prediction models of $w_{j,e}^A$ and D_j in the L-L system.

Model	RMSE	Quality of fit		Prediction capability		
		R^2	R_{ad}^2	R_{pred}^2	$RMSE_{pred}$	MAE_{pred}
$\hat{w}_{DNP,e}^A$	2.95×10^{-6}	0.982	0.979	0.924	6.16×10^{-6}	4.70×10^{-6}
$\hat{w}_{TNP,e}^A$	4.22×10^{-6}	0.978	0.971	0.884	9.92×10^{-6}	6.22×10^{-6}
\hat{D}_{DNP}	9.27	0.931	0.924	0.936	7.47	5.24
\hat{D}_{TNP}	4.59	0.908	0.899	0.917	4.06	3.08

**Fig. 8.** Experimental versus model values for training set (\diamond , DNP; \circ , TNP).

445 4. Conclusions

446 The distribution of DNP and TNP in a L-L system similar to the industrial
 447 production of MNB was characterized through relationships between the
 448 distribution ratio parameter, D_j , and the most influential operating variables
 449 in a range of interest for the full scale process. The values for D_{DNP} (between
 450 10 and 120) were about two times larger than the values for D_{TNP} (between 5
 451 and 55). Moreover, these values of D_j differed in one to two orders of
 452 magnitude when compared to the distribution of Bz ($K_{D,\text{Bz}}$) calculated with
 453 the Zaldivar et al. (1995) correlation for identical experimental conditions.

454 The systematic data-driven approach adopted in this work proved to be
 455 very efficient in getting simple, representative and reliable prediction models.
 456 Predictive models were developed for the parameter D_j and for the equilibrium
 457 weight fraction of each NP in the aqueous phase ($w_{j,e}^{\text{A}}$) with high quality of
 458 fit (R^2 and $R_{ad}^2 > 0.90$) and prediction capability ($0.88 < R_{pred}^2 < 0.93$ for
 459 the $w_{j,e}^{\text{A}}$ and $0.91 < R_{pred}^2 < 0.94$ for the D_j MLR models). The $w_{j,e}^{\text{A}}$ MLR
 460 model was significantly affected by the composition of both phases (expressed
 461 by $w_{\text{AS},0}^{\text{A}}$ and $w_{\text{Bz},0}^{\text{O}}$), the weight fraction of the NP in the organic phase ($w_{j,0}^{\text{O}}$),
 462 and the weight ratio of phases ($W_0^{\text{A}}/W_0^{\text{O}}$) in the following order of importance:
 463 $w_{\text{AS},0}^{\text{A}} > w_{j,0}^{\text{O}} > w_{\text{Bz},0}^{\text{O}} > W_0^{\text{A}}/W_0^{\text{O}}$. The effect of temperature was found to
 464 be negligible and the hypotheses of interaction between the two nitrophenols
 465 was discarded, because during the development of the models, in the screening
 466 step, the respective terms were not statistically significant. In the D_j MLR
 467 model, the variables found to be statistically significant were the aqueous phase
 468 composition ($C_{\text{AS},0}^{\text{A}}$) followed by the organic phase composition ($C_{\text{Bz},0}^{\text{O}}$).

469 The predictive models developed showed that an aqueous phase richer in
 470 sulphuric acid and higher concentrations of Bz in the organic phase (in depleting
 471 of MNB) provide higher $w_{j,e}^{\text{A}}$ and, consequently, lower values of D_j .

472 This study also indicates that an industrial process at similar L-L system
 473 composition, with each NP tested in a concentration lower than 5000 ppm, will
 474 have at least 95% of the NP in the organic phase.

475 **Nomenclature**476 *Abbreviations*

AAE	Average absolute error
ANOVA	Analysis of variance
Bz	Benzene
CI	Confidence interval
DNP	2,4-Dinitrophenol
DOE	Design of experiments
L-L	Liquid-liquid
MLR	Multivariate linear regression
MNB	Mononitrobenzene
MAE _{pred}	Mean absolute error
MSE	Mean squared error
NP	Nitrophenol
RASE	Root average squared error
RMSE	Root mean squared error
R&R	Reproducibility and repeatability
SA	Sulphuric acid
SE	Standard error
TNP	2,4,6-Trinitrophenol
WLS	Weighted least squares

477 *Symbols*

C	Molar concentration, mol dm ⁻³
D	Distribution ratio
d_c	Cook's distance measure
e	Residual
$F_{0, t}$	Test statistics
F_α	Fisher distribution

G	Gibbs energy, J mol^{-1}
K_{AH}	Equilibrium ionization constant
$K_{D,i}$	Distribution constant for species i
K_x	Thermodynamic distribution constant
N	Number of observations
n	Number of moles, mol
p	Number of parameters β
P	Pressure, Pa
R	Universal constant, $\text{J mol}^{-1} \text{K}^{-1}$
r	Studentized residual
R^2	Coefficient of determination
t	Observation index of test set
t_α	t Student distribution
T	Temperature, $^\circ\text{C}$
V_m	Molar volume, $\text{dm}^3 \text{mol}^{-1}$
w	Weight fraction
$W^{\text{A}}/W^{\text{O}}$	Initial weight ratio of aqueous phase for organic phase
x	Molar fraction
x_g, x_h	Regressors
y	Response
\bar{y}	Mean response
z	Number of regressor terms
\wedge	Estimate

⁴⁷⁸ *Superscript*

A	Aqueous
f	Phase
o	Standard state
O	Organic

479 *Subscript*

A^-	Acid in ionized form
ad	Adjusted
AH	Acid in molecular form
c	Observation index
e	Equilibrium condition
i	Generic species
j	DNP or TNP
$pred$	Prediction
t	Test set
0	Initial condition

480 *Greek symbols*

α	Significance level
β	Coefficient regressor
γ	Activity coefficient
μ	Chemical potential
σ^2	Variance

481 **Acknowledgements**

482 This work was supported by *Fundação para a Ciência e Tecnologia* (FCT)
483 and *CUF - Químicos Industriais S.A.* [Ph.D. Grant SFRH/BDE/52071/2013].

484 **Appendix A. Supplementary Material**

485 Supplementary data associated with this article can be found, in the online
486 version, at URL.

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Highlights

- Experimental data for liquid-liquid equilibrium distribution of nitrophenols (DNP and TNP);
 - Statistical predictive models for distribution ratio based on operating conditions;
 - Distribution ratio of each nitrophenol between the phases differed ($D_{DNP} \approx 2D_{TNP}$).
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