

UNIVERSITI SAINS MALAYSIA
GERAN PENYELIDIKAN UNIVERSITI PENYELIDIKAN
LAPORAN AKHIR

ANTIPROLIFERATIVE MECHANISM OF CERVICAL CANCER
CELLS (HELA) TREATED WITH BIOLOGICAL-ACTIVE SUB-
FRACTION FROM QUERCUS INFECTORIA EXTRACT

PENYELIDIK

PROF. MADYA DR. HASMAH ABDULLAH

2017

ANTIPROLIFERATIVE MECHANISM OF CERVICAL
CANCER CELLS (HELA) TREATED WITH
BIOLOGICAL-ACTIVE SUBFRACTION FROM
QUERCUS INFECTORIA EXTRACT

FINAL REPORT FOR RESEARCH UNIVERSITI

GRANT1001/PPSK/813061



BY

HASMAH BINTI ABDULLAH

SCHOOL OF HEALTH SCIENCES
UNIVERSITI SAINS MALAYSIA

2017

i

Igrz, thy senau, saya tol
terima barang² saujana.
7& 7/6/17

TABLE OF CONTENTS

LIST OF TABLES	v
LIST OF FIGURES.....	vi
LIST OF ABBREVIATIONS	vii
LIST OF SYMBOLS.....	x
ABSTRAK	xi
ABSTRACT.....	xiii
CHAPTER 1 INTRODUCTION	1
1.1Background of the study.....	1
1.2Objectives of the study	4
CHAPTER 2 LITERATURE REVIEW.....	5
2.1 Cervical cancer.....	5
2.2 Chemotherapy of plant anticancer	9
2.3 <i>Quercus Infectoria</i> galls	11
2.3.1 Utilization of QI galls in traditional practices	12
2.3.2 Potential of QI galls as anticancer agent.....	13
2.4 Targeting apoptosis in treatment of cancer.....	14
2.4.1 Apoptosis	15
2.4.2 Apoptosis pathways	16
CHAPTER 3 METHODOLOGY	19
3.1 Experimental design.....	19

3.2 Materials	21
3.3 Methods	23
3.3.1 Plant's extracts preparation.....	23
3.3.2 Cell culture.....	24
3.3.3 Subculturing cells	24
3.3.4 MTT assay.....	25
3.4 Fractionation and purification of manjakani extract	27
3.4.1 Chromatographic techniques.....	27
3.4.2 Phytochemical test.....	29
3.4.3 Structure elucidation.....	30
3.5 Mechanism of cell death.....	33
3.5.1 Nuclear morphological assay	33
3.5.2 FITC Annexin V/propidium iodide double staining	34
3.5.3 Determination of apoptotic proteins expressions	35
3.6 Statistical analysis	36
CHAPTER 4 RESULT	37
4.1 Organoleptic properties of <i>Quercus infectoria</i> (QI) galls	37
4.2 Extraction yields of <i>Quercus infectoria</i> (QI) galls	37
4.3 Antiproliferative activity of <i>Quercus infectoria</i> (QI) galls extracts	38
4.4 Phytochemical screening	43
4.6 Structure elucidation of isolated compounnd	43
4.6.1 Isolation of Secondary Metabolites.....	43

4.7 Antiproliferative activity of QIEA extract subfraction	51
4.8 Mechanism of Hela cell death induced by MGA subfracton	51
4.8.1 Nuclear morphological changes.....	51
4.8.2 Detection of apoptotic rate.....	53
4.9 Regulation of apoptotic proteins in HeLa cell line	60
4.9.1 p53 expression.....	60
4.9.2 Bax expression	62
4.9.3 Bcl-2 expression.....	64
4.9.4 Caspase-3 expression.....	66
4.9.5 Cytochrome c secretion	68
CHAPTER 5 DISCUSSION	69
5.1 <i>Quercus infectoria</i> (QI) galls extraction	69
5.2 Antiproliferative activity of QI galls extracts.....	70
5.3 Isolation and purification of bioactive compound from QIEA.....	74
5.4 Induction of apoptosis by MGA subfracton	74
5.4.1 Nuclear morphological changes.....	75
5.4.2 Phosphatidylserine translocation.....	76
5.4.3 Regulation of apoptotic proteins in HeLa cells treated with MGA	78
CHAPTER 6 CONCLUSION.....	85
REFERENCES.....	87

4.7 Antiproliferative activity of isolated compound isolated from QIEA extract.....	51
4.8 Mechanism of Hela cell death induced by 4-O-methylgallic acid.....	51
4.8.1 Nuclear morphological changes.....	51
4.8.2 Detection of apoptotic rate.....	53
4.9 Regulation of apoptotic proteins in HeLa cell line	60
4.9.1 p53 expression.....	60
4.9.2 Bax expression	62
4.9.3 Bcl-2 expression.....	64
4.9.4 Caspase-3 expression.....	66
4.9.5 Cytochrome c secretion	68
CHAPTER 5 DISCUSSION.....	69
5.1 <i>Quercus infectoria</i> (QI) galls extraction	69
5.2 Antiproliferative activity of QI galls extracts.....	70
5.3 Isolation and purification of bioactive compound from QIEA.....	74
5.4 Induction of apoptosis by MGA	74
5.4.1 Nuclear morphological changes.....	75
5.4.2 Phosphatidylserine translocation.....	76
5.4.3 Regulation of apoptotic proteins in HeLa cells treated with MGA	78
CHAPTER 6 CONCLUSION.....	85
REFERENCES.....	87

LIST OF TABLES

Table 3.1	List of chemicals and reagents.....	21
Table 3.2	List of laboratory apparatus	22
Table 3.3	List of kit and antibodies.....	22
Table 3.4	List of laboratory instruments.....	23
Table 3.5	Concentration ranges of extracts and cisplatin used for cytotoxic activity screening	26
Table 4.1	Organoleptic properties of QI galls	37
Table 4.2	Extraction yields of QI galls.....	38
Table 4.3	The IC ₅₀ values for QI galls extracts and cisplatin againts cancerous and non-cancerous cell lines, P \leq 0.05 was taken as significantly different from positive control (cisplatin).....	42
Table 4.4	Phytochemical constituents in QIEA.....	43
Table 4.5	Compounds isolated from QIEA.....	44

LIST OF FIGURES

Figure 2.1	Cervical cancer estimated incidence and mortality worldwide in 2012 by estimated age-standardised rates (World) per 100,000 (GLOBOCAN, 2012).....	7
Figure 2.2	The physical morphology of QI galls.....	12
Figure 2.3	The sequence of morphological changes in apoptosis process (adapted from Gewies, 2003).....	16
Figure 2.4	Intrinsic (mitochondria) and extrinsic (death receptor) pathways of apoptosis (Dewson and Kluck, 2009).....	18
Figure 3.1	Flow diagram for experiment	20
Figure 4.1	Cytotoxic effects of QIH towards Hela, MCF-7, MDA-MB-231, HepG-2 and L929 cell lines. Each point is the percentage of viability as compared to negative control, DMSO (control=100%). Graph presents mean \pm SEM $\mu\text{g}/\text{ml}$ of three independent experiments.....	39
Figure 4.2	Cytotoxic effects of QIEA towards Hela, MCF-7, MDA-MB-231, HepG-2 and L929 cell lines. Each point is the percentage of viability as compared to negative control, DMSO (control=100%). Graph presents mean \pm SEM $\mu\text{g}/\text{ml}$ of three independent experiments.....	40
Figure 4.3	Cytotoxic effects of QIM towards Hela, MCF-7, MDA-MB-231, HepG-2 and L929 cell lines. Each point is the percentage of viability as compared to negative control, DMSO (control=100%). Graph presents mean \pm SEM $\mu\text{g}/\text{ml}$ of three independent experiments.....	40
Figure 4.4	Cytotoxic effects of cisplatin towards Hela, MCF-7, MDA-MB-231, HepG-2 and L929 cell lines. Each point is the percentage of viability as compared to negative control, DMSO (control=100%). Graph presents mean \pm SEM $\mu\text{g}/\text{ml}$ of three independent experiments.....	42
Figure 4.5	^1H NMR sepectrum of 4-O-Methylgallic acid in CD_3OD , 400 MHz.....	45
Figure 4.6	^{13}C NMR sepectrum of 4-O-Methylgallic acid in CD_3OD , 400 MHz.....	45

Figure 4.7	Molecular structure of 4-O-Methylgallic acid (C ₈ H ₈ O ₅).....	46
Figure 4.8	FT-IR spectrum of 4-O-Methylgallic acid.....	47
Figure 4.9	¹ H NMR sepectrum of gallic acid in CD ₃ OD, 400 MHz.....	48
Figure 4.10	¹³ C NMR sepectrum of gallic acid in CD ₃ OD, 400 MHz.....	49
Figure 4.11	Molecular structure of gallic acid (C ₇ H ₆ O ₅).....	49
Figure 4.12	FT-IR spectrum of gallic acid.....	50
Figure 4.13	Nuclear morphological changes of HeLa cells stained with Hoechst 33258 stain.....	52
Figure 4.14	Scatter plots of FITC-AV/PI double staining in quadrant analysis..	54
Figure 4.15	The graph summarized the number of cells in each quadrant.....	59
Figure 4.16	p53 expressions in HeLa cell line.....	61
Figure 4.17	Bax expressions in HeLa cell line.....	63
Figure 4.18	Bcl-2 expressions in HeLa cell line.....	65
Figure 4.19	Caspase-3 expressions in HeLa cell line.....	67
Figure 4.20	Cytochrome c levels in HeLa cell line.....	68

LIST OF ABBREVIATIONS

BSC	Biosafety cabinet
Caov-3	Ovarian cancer cell line
CO ₂	Carbon dioxide
DMEM	Dulbecco's Modified Eagle Medium
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
EDTA	Ethylenediaminetetraacetic acid
ELISA	Enzyme-linked immunosorbent assay
FBS	Fetal bovine serum
FDA	Food and Drug Administration
HBV	Hepatitis B virus
HCV	Hepatitis C virus
HeLa	Cervical cancer cell line
HPV 16	Human papillomavirus strain 16
HPV 18	Human papillomavirus strain 18
HPV	Human papillomavirus
IC ₅₀	Inhibition concentration of 50% cell population
IDV	Integrated density value
MRI	Mean relative intensity
MTT	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide
p53	Tumour suppressor protein
PBS	Phosphate buffer saline
pRB	Retinoblastoma protein

QI	<i>Quercus infectoria</i>
QIA	<i>Quercus infectoria</i> aqueous
RNA	Ribonucleic acid
SEM	Standard error mean
GOG	Gynecologic Oncology Group
Cis	Cisplatin

LIST OF SYMBOLS

%	Percentage
μg	Microgram
$\mu\text{g}/\text{ml}$	Microgram per millilitre
μl	Microlitre
μm	Micrometre
$^{\circ}\text{C}$	Degree celcius
cells/ml	Cells per millilitre
cm	Centimetre
cm^2	Square centimetre
g	Gram
L	Litre
mg	Milligram
mg/ml	Milligram per millilitre
ml	Millilitre
mM	Millimolar
OD	Absorbance
rpm	Revolutions per minute
V	Voltage

ABSTRAK

Kanser servik merupakan kanser keempat paling kerap berlaku dalam kalangan wanita di seluruh dunia dengan anggaran kira-kira 530,000 kes pada tahun 2012. Kanser servik juga merupakan salah satu punca utama kematian yang disebabkan oleh kanser dalam kalangan wanita di seluruh dunia. Kaedah rawatan moden dan konvensional yang digunakan sekarang didapati mendatangkan pelbagai kesan sampingan terhadap pesakit. Oleh yang demikian, pelbagai kajian telah dijalankan untuk mencari rawatan alternative yang lebih selamat dan berkesan, seperti penggunaan produk hasilan semulajadi. Kajian terdahulu melaporkan aktiviti farmakologikal manjakani atau *Quercus infectoria* (QI) termasuk aktiviti antikanser. Walaubagaimanapun, mekanisma tindak balas aktiviti antikanser masih samar dan perlu kajian mendalam. Dalam kajian ini, QI diekstrak menggunakan beberapa jenis pelarut organik sebelum diuji untuk aktiviti sitotoksik dan mekanisma tindak balas kematian sel. Kaedah asai MTT digunakan untuk menentukan aktiviti sitotoksik bagi ekstrak n-hexane (QIH), etil asetat (QIEA) dan methanol (QIM) daripada QI terhadap sel kanser servik (Hela). Aktiviti sitotoksik ekstrak QI terhadap sel fibroblast normal juga ditentukan bagi melihat kesan toksik dan sifat keselektifan ekstrak. Di samping itu, sel yang dirawat dengan DMSO bertindak sebagai kawalan negatif dan sisplatin sebagai kawalan positif. Kemudian, graf dos-tindak balas diplotkan bagi menentukan nilai IC₅₀. Pemerhatian perubahan morfologi nukleus, peratusan sel apoptotik dan pengesanan paras pengekspresan protein apoptotik juga dilakukan. Pemeriksaan kandungan sebatian fitokimia dalam ekstrak yang paling poten juga dijalankan untuk menentukan hubung kait kumpulan bioaktif dalam ekstrak. Berdasarkan keputusan eksperimen, QIEA menunjukkan aktiviti sitotoksik

yang lebih baik daripada QIM dan QIH dengan perencatan pertumbuhan paling poten terhadap sel Hela ($6.33 \pm 0.33 \mu\text{g/ml}$) dan bersifat sitoselektif. Hasil kajian pengesanannya sebatian fitokimia pula menunjukkan kehadiran tanin, alkaloid, glikosida, saponin, terpenoids, flavonoid dan sebatian fenolik dalam ekstrak QIEA. Ekstrak QIEA kemudian ditulenkkan dan menghasilkan dua komponen aktif iaitu *4-O-methylgallic acid (MGA)* dan *gallic acid (GA)*. Subfraksi MGA adalah lebih poten dengan $\text{IC}_{50} 11 \pm 0.58 \mu\text{g/ml}$. Hasil kajian juga menunjukkan sel Hela yang dirawat dengan subfraksi MGA telah mengalami apoptosis, yang mana dibuktikan melalui perubahan morfologi nukleus dengan kehadiran jasad apoptotik serta peningkatan kadar peratusan apoptosis. Tambahan pula, hasil kajian menunjukkan subfraksi MGA mencetuskan apoptosis melalui tapak jalan p53. Peningkatan aras protein p53 didapati telah menurunkan paras protein Bcl-2 dan mengaktifkan kaspase-3. Kesimpulannya, dua subfraksi mengandungi terbitan asid galik MGA dan GA menunjukkan ciri aktif secara biologi terhadap sel HeLa. Subfraksi MGA merencatkan pertumbuhan sel kanser Hela secara melalui mekanisma kematian sel secara apoptosis.

ABSTRACT

Cervical cancer is placed at the fourth most frequent cancer among women worldwide, which was estimated approximately 530, 000 new cases in 2012. This type of cancer has become one of the leading causes of cancer death among women worldwide. Conventional and modern cancer treatment nowadays comes with negative and adverse side effects to the patients. Therefore, lot of studies have been conducted to search for new alternative treatment which is more safe and effective such as utilisation of natural product based medications. Previous studied revealed various pharmacological activities of *Quercus infectoria* (QI) galls or manjakani including anticancer activity. However, the mechanism of action lay behind was not well explained. Therefore, in this present study, QI gall was selected for the evaluation of cytotoxic activity and cell death mode of action. In this study, QI was extracted using different types of organic solvents before being tested for cytotoxicity activity and mode of cell death. MTT assay was used to determine cytotoxic activity for n-hexane (QIH), ethyl acetate (QIEA) and methanol (QIM) of QI galls extracts against cervical cancer (Hela). Cytotoxic activity of QI extracts also tested against normal fibroblast (L929) cell line to determine cytotoxic effects and cytoselective properties. Meanwhile, DMSO-treated cells served as negative control while cisplatin-treated cells served as positive control. After that, dose-response graph was plotted to determine IC_{50} values. Observation on nuclear morphology, apoptotic percentage and detection of apoptotic protein expressions were also conducted. Moreover, phytochemical screening analysis had been carried out to determine bioactive groups that present in the most potent extract. From the findings,

QIEA extract exhibited better cytotoxic activity with best growth inhibition against Hela cells (IC_{50} value = $6.33 \pm 0.33 \mu\text{g/ml}$) compared to other extracts and showed cytoselective property as no cytotoxicity observed in the treated normal fibroblast (L929) cells. As for the phytochemical analysis of QIEA extract, it was revealed the presence of tannin, alkaloids, glycosides, saponins, terpenoids, flavonoids and phenolic compounds. The QIEA extract was then purified and resulted two compounds 4-O methylgallic acid (MGA) and gallic acid (GA). The most potent antiproliferative activity was exhibited by MGA subfraction with IC_{50} $11 \pm 0.58 \mu\text{g/ml}$. The current study also revealed that Hela cells treated with MGA subfraction has undergone apoptosis as exerted by the alteration of nuclear morphology and the presence of apoptotic bodies as well as the increment of apoptosis rate in the treated cells. Furthermore, MGA subfraction triggered apoptosis through upregulation of tumor suppression protein p53, with down regulation of Bcl-2 expression and facilitated the apoptosis execution through caspase-3 activation. In conclusion, the purification of QIEA has resulted two biologically active subfraction namely MGA and GA. It was cleared that MGA subfraction reduced Hela cell growth through induction of apoptosis.