ECOLOGIA BALKANICA

2023, Vol. 15, Issue 1

June 2023

pp. 49-76

Are Mytilus species suitable bioindicators for assessing aquatic pollution along the Black Sea Coast? A review

Vesela Yancheva¹, Iliana Velcheva¹, Elenka Georgieva², Stela Stoyanova², Borislava Todorova¹, László Antal^{3,4*}, Krisztián Nyeste^{3,4}

¹Department of Developmental Biology, Faculty of Biology, Plovdiv University, Plovdiv, BULGARIA

²Department of Ecology and Environmental Conservation, Faculty of Biology, Plovdiv University, Plovdiv, BULGARIA

³Department of Hydrobiology, Faculty of Science and Technology, Institute of Biology and Ecology, University of Debrecen, HUNGARY

⁴National Laboratory for Water Science and Water Safety, University of Debrecen, HUNGARY

*Corresponding author: antal.laszlo@science.unideb.hu

Abstract. This review aims to summarize the possibility of using mussels (*Mytilus* spp.) as bioindicators to assess aquatic pollution in the Black Sea in Bulgaria. In addition, the main responsive biomarkers that could be applied to study the negative effects of different toxicants on these species in terms of using the Marine Strategy Framework Directive and implementation of environmental quality standards (EQS) in marine biota are also discussed. A specific reference is made to plastic pollution, transplant mussel caging, and mussel watch programs - their application, challenges, and future perspectives in Bulgaria.

Key words: *Mytilus* spp., aquatic pollution, biomarkers, Black Sea, Bulgaria.

Introduction

Human activities produce various persistent organic and inorganic pollutants, such as polyromantic hydrocarbons (PAHs), polychlorinated bypehnils (PCBs), polybrominated diphenyl ethers (PBDEs), shortchain chlorinated paraffins (SCCPs), dioxins and trace metals, as well as plastic litter, which eventually end up in the marine environments and ocean (Guendouzi et al., 2020; Kouali et al., 2022). Hence, the marine environments, which are subjected to such diverse and toxic pollutants, are threatened – the integrity of habitats and their associated biota, from coastal to pelagic environments, and from benthonic to surface ecosystems (Anbuselvan et al., 2018; Bonanno & Orlando-Bonaca, 2018; Bonsignore et al., 2018; Urban-Malinga et al., 2018).

Environmental problems in Black Sea

The Black Sea is the world's largest landlocked inland sea between southeastern Europe and western Asia, and is surrounded by six countries - Romania, Bulgaria, Ukraine, Russia, Georgia, and Turkey (Fig. 1). It is 1210 km long from east to west and up to 560 km wide, has a maximum depth of 2212 m and a volume of 534000 km³, and covers an area of 432000 km² (Özsoy &

Ünlüata, 1997; Sari et al., 2018). The Black Sea communicates with the Mediterranean Sea to the south and the Azov Sea to the north (Topping & Mee, 1998; Boran & Altınok, 2010).

However, the Black Sea suffers from serious and much challenging environmental problems. The sea's shallow, mixed surface waters receive river discharges, heavily loaded with nitrogen and phosphorus nutrients, and polluted with industrial and mining wastes. In addition, coastal industries appear to discharge wastes directly into the sea with little or no treatment. Thus, the life-supporting surface laver's water quality has seriously deteriorated (Fabry et al., 1993). Moreover, The Black Sea coastal zone is densely populated. The total population in the catchment area of the Black Sea is about 160 million, almost half of which is from noncoastal countries in the catchment area of the Danube (Sari et al., 2018). It has a permanent population of about 6 million, with another 4 million tourists during summer. Another important pollutant source is the Mediterranean inflow that transports municipal and industrial pollutants from the mega metropolitan Istanbul city with a population of 16 million (Sari et al., 2018).



Fig. 1. Map of the Black Sea in Europe.

The Black Sea's primary pollution sources are the rivers flowing into it. The major rivers flowing into the Black Sea and their discharges are the Danube (203 km³ yr⁻¹), Dnieper (Dnepr) (54 km³ yr⁻¹), Dniester (Dnestr) (9.3 km³ yr⁻¹), Don (28 km³ yr⁻¹), and Kuban (13 km³ yr⁻¹) (Bakan & Özkoç, 2007). Furthermore, many smaller rivers along the Turkish and Bulgarian coasts contribute another 28 km³ yr⁻¹ to the water budget of the sea (Bakan & Özkoç, 2007). Moreover, the Danube is the major river delivering 58% of the total freshwater and sediment inputs to the Black Sea (Mee, 1992; Müftüoğlu, 2013). It is 2850 km long, with a drainage area of 817000 km² (Sari et al., 2018). The Danube and the other major rivers flow through the central and eastern European industrial towns and agricultural areas, and transport significant pollutants and natural inputs from mineralized and high-background rock-bearing areas. It drains into the Black Sea, passing through the lands of Germany, Austria, Slovakia, Hungary, Croatia, Serbia, Romania, Bulgaria, and Ukraine. 81 million people living in the drainage area influence its hydrological system (Sommerwerk et al., 2010). Hence, the Black Sea's sediments and water have been adversely affected by the anthropogenic transport of riverine pollutants over the last few centuries (Guieu et al., 1998; Secrieru & Secrieru, 2002; Yigiterhan & Murray, 2008; Mülayim & Balkıs, 2015) and sadly, this problem persists.

Plastic pollution is an environmental issue in many seas worldwide (Lebreton et al., 2017; Ryberg et al., 2019). The Black Sea is no exception, and it suffers from land-based pollution, including cities and sewage systems (Lechner et al., 2014; Berov & Klayn, 2020; D'Hont et al., 2021; González-Fernández et al., 2021; Pojar et al., 2021). The environmental problems are getting worse because the sea is semi-enclosed; thus, plastics tend to accumulate over time; the drainage area of the sea is approximately 2.5 million km² and is divided into 107 subbasins, which drain through more than 20 countries located in the European and Asian continents; the sea also receives plastics from the three large transboundary rivers: Danube, Don, and Dnieper (Strokal et al., 2022).

There are already several surveys of marine litter on beaches, floating litter in the

waters, and plastic pollution of the sediments that have been carried out in Turkey and Romania (Topçu et al., 2013; Suaria et al., 2015; Aytan et al., 2016; Kilinc, 2017; Oztekin & Bat, 2017; Săvucă et al., 2017; Terzi & Seyhan, 2017; Pojar & Stock, 2019; Aytan et al., 2020; Terzi et al., 2020), including a survey on the Western and Eastern Black Sea waters along the coasts of Ukraine, Russia, and Georgia (Slobodnik et al., 2018).

There are also some studies from Bulgaria (Moncheva et al., 2016; Simeonova et al., 2017; Simeonova & Chuturkova, 2019; Stanev & Ricker, 2019; Berov & Klayn, 2020; Miladinova et al., 2020); however, there is insufficient data on the negative effects of marine litter in the Bulgarian Black Sea, and how (micro)plastics could affect different biological indices in the aquatic organisms. The microplastic pollution in the Black Sea in Bulgaria as an environmental problem and its potential toxic threats for aquatic animals in Bulgaria has been recently reviewed by Todorova et al. (2023).

Marine Strategy Framework Directive

Within the Marine Strategy Framework Directive scope, the Member States of one marine region and neighboring countries, which share the same marine waters, must cooperate to protect the marine environment more effectively (Coatu et al., 2016). The Strategy Framework Directive Marine (MSFD, 2008/56/EC) requires all the EU achieve Member States to Good Environmental Status (GES) of the marine environment by 2020, considering 11 key descriptors environmental status of (Orlando-Bonaca et al., 2022). The presence of hazardous substances in biota represents relevant criteria and indicators for assessing the status of the Black Sea environment under Descriptor 8 ("Concentrations of contaminants are at levels not giving rise to pollution effects") and 9 ("Contaminants in fish and other seafood for human consumption do not exceed levels established by Community legislation or other relevant standards") (Coatu et al., (Properties 2016). Descriptor 10 and quantities of marine litter do not cause harm

to the coastal and marine environment) focuses on the emerging problem of marine litter and its effects on the marine environment and biota. Specifically, the secondary criterion D10C3 defines that "The amount of litter and micro-litter ingested by marine animals is at a level that does not adversely affect the health of the species concerned," "Litter and micro-litter classified in the categories 'artificial polymer materials' and 'other,' assessed in any species from the following groups: birds, mammals, reptiles, fish and invertebrates," and that "Member States shall establish that list of species to be assessed through regional or subregional cooperation." Indicators for this criterion should be developed to assess the GES in the current implementation cycle of MSFD. At the moment, this criterion has not been fully evaluated, as the threshold values for levels that may have lethal or sublethal effects on marine organisms have not yet been identified, while they should still be defined through regional or subregional cooperation. In addition, the secondary criterion D10C4 outlines "The number of individuals of each species, which are adversely affected due to litter, such as by entanglement, other types of injury or mortality, or health effects." Although this criterion appears ready to be used, monitoring campaigns for the census of marine organisms affected by litter are relatively occasional. A magnitude ranking of injuries due to litter (and explicitly plastic) ingestion has still to be developed. Eventually, the Commission Decision 2010/477/EU identified the indicator "Trends in amount and composition of litter ingested by marine animals (e.g., stomach analysis)" (10.2.1) (Galgani et al., 2013; Orlando-Bonaca et al., 2022).

According to the Barcelona Convention (2016), "Marine pollution knows no border, pollution in one country affects all other 21 countries." The awareness of the need for a regional approach has resulted in many formal and informal initiatives at global and regional levels (e.g., UNEP Regional Seas Program, 1974; OSPAR, 2009). Moreover, the "Common Indicator 18" of the Barcelona Convention proposed loggerheads as

indicators of marine debris levels ashore or at sea for monitoring, achieving, or maintaining the GES, as defined by D10 of the MSFD (Barcelona Convention, 2016). In addition, the Integrated Monitoring and Assessment Program (IMAP) adopted in 2016 includes Ecological Objective 10 on Marine Litter (IMAP, 2016). Within this framework, the proposal for marine litter monitoring also includes Candidate Indicator 24: "Trends in the amount of litter ingested by or entangling marine organisms focusing on selected mammals, marine birds, and marine turtles (EO10)". So far, the most suitable species for this indicator have not been identified vet (Orlando-Bonaca et al., 2022).

Mussels as an important food source in aquaculture

Mytilus species (Fig. 2) are common in temperate seas all around the globe. They are widely used both as seafood, not just as sentinel organisms monitoring in anthropogenic pollution trends in marine waters (Goldberg, 1975, 1980; Farrington et al., 2016). Blue mussels have been an important food for humans for thousands of years, and mussel farming dates back to the Ancient Romans (Beyer et al., 2017). Mussels are economically important food species, accounting for more than a third (roughly 470 thousand tons) of production by weight of the aquaculture industry in the European Union (Eurostat, 2016). Therefore, ingestion of mussel seafood contaminated with various pollutants is a key source of potential health risks, such as neurotoxic, carcinogenic, and cardiovascular diseases for a man (Ersoy & Çelik, 2009). This is of primary concern in the case of top marine predators, as some metals and persistent organic pollutants (dichlorodiphenyltrichloroethane, DDT) accumulation tends to magnify along the food web, resulting in a higher potential risk to human health when high trophic level predators are consumed (Barone et al., 2018).

In recent years marine aquaculture along the Bulgarian Black Sea coast has been mainly related to the construction of mussel farms, which number has exceeded 30 installations. The mussels from the family Mytilidae, are important and prospective species traditionally consumed in Bulgaria and Europe (Cammilleri et al., 2020).

Mediterranean (black) mussels (M. galloprovincialis) are a valuable protein source, and the favorable conditions of the Black Sea (temperature, salinity, and food availability) stimulated mussel farming in this region. In recent years, the growing market interest in this species is based on the proven high nutritional quality and M. galloprovincialis have a future as a promising source of high-quality protein, polyunsaturated fatty acids, and essential macro and microelements (Pevcheva et al., 2021a). Mussels are actually the only bivalves cultured on the Bulgarian coast of the Black Sea (Ministry of Agriculture and Food, 2016). They are mainly cultured on ropes suspended in the water column and attached to rafts. The aquacultured mussels are usually suspended 3-4 m above the sea bottom (Executive agency for fish and aquacultures - IARA, 2017). A few studies characterized this species as a beneficial food that could provide a well-balanced chemical composition and, through their consumption, could prevent various nonchronic diseases (Özden et al., 2010; Petrova-Pavlova et al., 2014; Merdzhanova et al., 2019; Peycheva et al., 2021b).

Blue mussels (M. edulis) are bivalve mollusks widely consumed as seafood, with a high content of proteins, omega-3 fatty acids, and vitamins (Akre et al., 2019; Gomez-Delgado et al., 2023). Moreover, Black Sea mussels are one of the most perspective novel food sources with a protein content equal to cattle meat. The mussels are also important natural biofilters - at 17°C temperature of the water, one mussel can filter almost 3 liters of seawater per hour. In recent years in Bulgaria, an increasing interest in this animal as profitable breeding culture has been increasing. However, today due to the industrial pollution of the Black Sea, there are serious obstacles in mussel farms (Ganchev et al., 2012). In this regard, the spatial extent of mussel beds in the Black Sea and Sea of Marmara was reported to have been declining drastically over the past years

(Zaitsev & Mamaev, 1997). The total biomass of mussels in the Black Sea was estimated at 25×106 tons in the 1960s, dropping to 7×106 tons in the 1980s, with juvenile specimens predominating (Zaitsev, 1992). Mass mortalities of *M. galloprovincialis* were observed in the region due to siltation and hypoxia (Gomoiu, 1992; Ozturk & Ozturk, 1996). Moreover, eutrophication, coastal constructions, sea dumping, and ship accidents (such as Rabunion-18, sinking with 20000 live sheep in the Bosphorus) were reported to be the leading cause of hypoxia in the region (Yurdun et al., 1995; Topçu et al., 2019).

Mussels as important sentinel organisms in aquatic toxicology

Using bioindicators as sentinels of the environment's health is a well-established biotechnology that provides qualitative and quantitative information on the impact of numerous pollutants and stressors (Siddig et al., 2016). The selection of appropriate bioindicator species follows general criteria that can also be applied to marine plastic pollution and should allow different marine habitats to be monitored. Given the ubiquitous nature of plastics in the oceans, cosmopolitan marine species should be considered the primary sentinels of environmental impacts. Larger ecological niches allow organisms to detect the same disturbances or stressors in different habitats (Bartell, 2006; Urban et al., 2012).

Any good bioindicator, in particular, should have some basic characteristics, including natural occurrence, abundance, ease of identification and sampling, moderate tolerance to disturbances and stresses, and broad geographic distribution corresponding to a range of exposures to a certain pollutant or stressor (Carignan & Villard, 2002; Caro, 2010).

The high number of taxa impacted by different marine plastics underscores the magnitude of this threat to biodiversity. Some key taxa, such as sea turtles and marine mammals, and various pelagic fish species were particularly affected. Furthermore, to date, numerous species of different taxonomic groups have been used as bioindicators of various marine pollutants, such as mollusks (Naimo, 1995; Guerlet et al., 2007; Dirrigl et al., 2018; Yancheva et al., 2018; Yancheva et al., 2022), among them - mussels, in particular, the blue mussel (*Mytilus edulis* Linnaeus, 1758) and the Mediterranean mussel (*Mytilus galloprovincialis* Lamarck, 1819) (Cappello et al., 2013; Beyer et al., 2017), as well as various fish species (Yancheva et al., 2015).

Mytilus spp. as bioindicators for aquatic pollution

The genus *Mytilus* includes several closely related (congeneric) species (or subspecies) that can interbreed with each other and make fertile hybrids. It is often called the Mytilus edulis complex. Although the exact taxonomy within the *Mytilus* genus is not yet fully clarified, recent research indicates there are five species occurring in the Northern Hemisphere (M. edulis, M. galloprovincialis, M. trossulus, M. californianus, and M. coruscus) and two in the Southern Hemisphere (M. galloprovincialis and M. platensis). In contrast, the former M. chilensis, is currently considered a variant of M. platensis (Gaitan-Espitia et al., 2016). About 3.5 million years ago, M. trossulus and M. edulis diverged genetically, then around 2.5 million years ago, M. edulis and M. galloprovincialis diverged too (Fraïsse et al., 2014).

The blue mussel (M. edulis) and the Mediterranean mussel (M. galloprovincialis) are two well-known "early warning" bioindicators of marine pollution that are increasingly used to monitor the presence of microplastics (Avio et al., 2017; Phuong et al., 2017). Mussels are sessile suspension feeders that ingest microplastics to 500-fold higher concentrations (Van Cauwenberghe & Janssen, 2014; Van Cauwenberghe et al., 2015). Although short-term exposure to microplastics may not have significant biological effects (Browne et al., 2008), ingestion of microplastics by mussels has been shown to result in disruptive effects, such as a reduction in filtering activity (Wegner et al., 2012), tissue-dependent

changes at the transcriptome level (Détrée & Gallardo-Escárate, 2017), histological changes, and severe inflammatory responses (von Moos et al., 2012).

M. edulis is boreo-temperate in its distribution on both coasts of the Atlantic Ocean. It is found in abundance, intertidally and subtidally, in both sheltered and exposed sites, attached to hard substrates or forming biogenic reefs. As summarized by Lynch et al. (2014), in the western Atlantic, M. edulis is historically found from the Arctic Sea, Canada (Dall, 1889) to North Carolina, United States (Stimpson, 1860; McDougall, 1943) and in the eastern Atlantic occurs from Norway (Christiansen, 1965) to the border of France and Spain (Sanjuan et al., 1994). In favorable conditions, M. edulis can grow to a shell length of more than 10 cm and have a lifespan of more than 20 years (Powell & Cummins, 1985; Sukhotin et al., 2007), although specimens larger than 8 cm and older than 10 years are uncommon.

M. edulis is a keystone coastal species with essential roles in ecosystem functioning, including habitat formation for diverse benthic

communities (Joint Nature Conservation Committee, 2008) and nutrient recycling. They play an important role in benthic-pelagic coupling by removing large quantities of suspended organic matter from the water by filter-feeding, and through the production of faces and pseudofaces (Ward & Shumway, 2004) and process large volumes of water; for example, under optimal algal conditions, a 21.5 mm sized mussel will filter an average of 15 mL min⁻¹ (Riisgård et al., 2014). Coupled with their wide geographical range and low metabolic transformation rates, these traits make blue mussels useful in monitoring programs for many potential and dissolved chemical pollutants (Scott et al., 2019).

M. galloprovincialis is endemic to the Mediterranean, Black, and Adriatic Seas and has expanded its range to the British Isles (Gosling, 1992). As explained by Livingstone (1992) and Uluturhan et al. (2019), the sessile nature of Mediterranean mussels renders them ideal candidates for molecular to physiological and ecological studies, and biomonitoring purposes of the water column.



Fig. 2. *Mytilus edulis* - general anatomy and morphology of as presented by Eggermont et al. (2020).

Biomarkers in Mytilus species

The concept of biomarkers was set for the first time in 1987 by the US National Research Council (NRC) and defined as "xenobioticallyinduced variations in cellular or biochemical components or processes, structures or functions that are measurable in a biological system or sample". In addition, due to their quick response to stress and potentially high toxicological relevance, biomarkers are often considered early warning indicators in detecting molecular, biochemical, or ecological effects (Dellali et al., 2021).

To assess the impact of ingested debris on marine organisms, Fossi et al. (2020) proposed a threefold monitoring approach that can combine an accurate measurement of debris and microplastic levels in animals, the assessment of plastic additives and persistent organic pollutants levels in tissues, and associated toxicological effects. According to Fossi et al. (2018a) and Fossi et al. (2018b), such

a monitoring approach should be based on three types of collected data, and the three evaluations can be applied independently or simultaneously for different kinds of bioindicators commercial (e.g., species, protected species, stranded and/or hospitalized organisms, etc.): 1) the analysis of gastrointestinal contents to assess the debris (especially plastics) ingested by the animals, focusing on the occurrence (%), abundance (number), weight (g), color, size, and polymer topologies of the ingested macrodebris and microplastics; 2) the qualitative -quantitative analysis of plastic additives (various phthalates and polybrominated diphenyl ethers) and bioaccumulating, persistent, and toxic substances used as "potential" plastic tracers in the bioindicators tissues; 3) the analysis of the effects of debris ingestion through biomarker responses at different levels of biological organization (from variations in gene/protein expression to histological changes).



Fig. 3. Biomarkers in *Mytilus* species for assessing the effects of water pollution.

Here we present some of the most common and reliable biomarkers, which can be applied in *Mytilus* species when assessing aquatic pollution (Figure 3):

Neutral red retention time in lysosomes

Unlike fish, mussels do not have the same ability to metabolize organic compounds, but detoxification processes exist through other complex mechanisms that occur partly in the lysosomal compartment. Hence, the lysosomes play an important role as a natural immunological defense system in invertebrates (Lowe & Pipe, 1994; Lowe et al., 1995; Moore & Willows, 1998; Akcha et al., 2004; Mamaca et al., 2005). At the subcellular level, the lysosomal system has been identified as a particular target for the toxic of contaminants, and pathological effects alterations in lysosomes have been especially the identification useful in of adverse environmental impacts on marine organisms (Moore et al., 1996; Giamberini & Pihan, 1997; Moore et al., 2009).

When marine mollusks, such as mussels, are exposed to xenobiotics, one of the characteristic pathological alterations is decreased integrity of lysosomal membrane (Moore, 1988). the Lysosomal membrane integrity has also been reduced with increasing nonspecific stress (i.e., biotic and abiotic) (Moore, 1985). The mechanisms causing this alteration in membrane stability needs to be better understood. Still, it may involve the direct effects of chemicals on the membrane or the increased frequency of secondary lysosomes in toxicant-stressed cells (Mayer et al., 1992).

are important Lysosomes subcellular that contain many organelles hydrolytic enzymes, perform protein degradation and detoxify some foreign compounds. At the cellular level, lysosomal digestion pathways phago-cytosis, endocytosis, include and autophagy. The lysosomal membrane protects the cytosol and the rest of the cell from leakage of degradative enzymes. However, malfunctioning of lysosomes and their accumulation of toxic pollutants have been linked to lysosomal storage diseases and result in lysosomal injury and oxidative damage, in some cases leading to cell death (Moore et al., 2007). In this regard, the neutral red retention time (NRRT) assay takes advantage of this phenomenon by measuring the

decreased retention time of neutral red dye within phagocytic haemocytes of a range of aquatic organisms, including mussels (Regoli, 1992; Tedesco et al., 2008). In the popular sentinel species, *M. edulis*, the haemocytes (from the adductor muscles or digestive gland) are essential immune system components (Hu et al., 2015). Therefore, NRTT has been reported as a useful indicator of the organism's overall health status because animals exposed to pollutants often have compromised lysosomal stability (Borenfreund & Puerner, 1985; Moore et al., 2009) and the loss of red dye in the cytosol (reduction of NRRT) indicates destabilization of the lysosomal membrane.

Oxidative stress

Biomarker responses in bivalves include the induction of antioxidant enzymes. They have been widely used to assess organic xenobiotics' impact in marine environments (Doyotte et al., 1997; Lau & Wong, 2003). During detoxification, enzymes, such as carboxylesterases (CEs), convert toxic compounds into more hydrophilic and reactive molecules to facilitate their elimination. In the second step, glutathione xenobiotics transferases (GSTs) conjugate metabolites with glutathione to convert them into more hydrophilic and less reactive molecules (Falfushynska et al., 2019). Accompanying the detoxifying process, reactive oxygen species (ROS) can be produced.

In front of this, organisms have developed a complex antioxidant system to avoid oxidative stress damage (Livingstone, 2001; Rios-Fuster et al., 2022). The antioxidant system, which is composed of enzymes, such as catalase (CAT), superoxide dismutase glutathione (SOD), peroxidase (GPx), glutathione reductase (GRd), and glutathione-S-transferase (GST) (Capó et al., 2015; Capó et al., 2021) serves as a shield, which crucial role is to protect the cells against the ROS harmful effects and to reduce the possible damage due to their high reactivity (Vidal-Liñán et al., 2010; Kourdali et al., 2022).

However, if the ROS production is beyond the organisms' elimination capabilities, ROS can lead to the production of several biomolecules, such as malondialdehyde (MDA), which are lipids generating oxidative products and thus, can be used as biomarkers of oxidative damage (lipid peroxidation) (Bartoskova et al., 2013).

DNA damage

Many harmful substances sporadically present in the water can bioaccumulate in living organisms, induce DNA and/or cell damage, and even enter the trophic chain and affect distant ecosystems if some components of such chains are migratory. Increasing attention is being paid to using micronuclei (MN) as an index of cytogenetic damage in aquatic organisms.

Mussels are potentially suitable biological indicators of genotoxic pollution (Venier & Canova, 1996; Dixon et al., 2002). The induction of MN in different mussel species exposed to genotoxic compounds has been reported in several studies (Parolini et al., 2010; Dallas et al., 2013; Liu et al., 2014). Therefore, *Mytilus* spp. can be considered suitable for routine in situ surveys of genotoxic pollutants employing the micronucleus test as a biomarker, supporting Viarengo & Canesi (1991).

According to Bolognesi & Hayash (2011), the MN assay validation process in the genus Mytilus started more than 20 years ago (i.e., Heddle et al., 1983; Wrisberg & van der Gaag, 1992). Dose-related induction of MN by different pollutants or polluted water containing various mixtures of contaminants have been reported in gill cells and haemocytes - mitomycin C (Majone et al., 1987, 1990), ethyl methanesulfonate (EMS) (Wrisberg et al., 1992; Jha et al., 2005), dimethylbenz[a]anthracene (Bolognesi et al., 1996), benzo[a]pyrene (Venier et al., 1997), bisphenol A (Barsiene et al., 2006), phenanthrene (Koukouzika & Dimitriadis, 2008), and heavy metals (Bolognesi et al., 1999; Duroudier et al., 2021).

Histopathological and histochemical analyses

Histopathology is a sensitive tool for diagnosing direct and indirect toxic effects that affect the tissues (Kent et al., 2013); therefore, it is considered an excellent method for assessing environmental quality (Bignell et al., 2011). In addition, according to Hinton & Lauren (1990), histopathology is often the easiest method of assessing both short and long-term toxic effects for field assessments. On the other hand, Wester & Canton (1991) state that the histopathological methods are relatively labor-intensive and require good experience, but after all, they have the considerable advantage that pathological alterations in different tissues (e.g., gills, liver) can be observed individually, creating a direct link with physiological functions, such as growth, reproduction, respiration, and nutrition.

The histochemical techniques help to analyze the localization of lipids and glycogen the cellular level. Furthermore, at histochemistry's advantage main lies in analyzing biological phenomena in "particular cells". In this regard, the intensity of staining can be used for comparing the lipid and glycogen contents present in the (gills) cells of normal mussels compared to treated ones with different toxic compounds (Pathan et al., 2009).

Based on the studied literature on histochemical changes triggered by various toxicants, we can state that the histochemical methods, such as PAS (glycogen) and Sudan III (lipids) reactions mainly concern vertebrates, such as fish (and the liver), and to some smaller extent - invertebrates, such as mussels (and the gills and digestive gland) (Drastichová et al., 2005; Wolf & Wolfe, 2005, Figueiredo-Fernandes et al., 2006; El-Serafy, 2009; Singh, 2014).

Furthermore, glycogen synthesis and degradation mechanisms are studied primarily in mammal tissues, such as the liver (Smythe & Cohen, 1991; Bollen et al., 1998). These mechanisms seem similar in the gills, which are energy-consuming organs. Moreover, the mussel gills are attractive models for ecotoxicological studies because the gills are the first uptake site for many toxicants present in the aquatic environment and are often affected by exposure to pollutants (Gómez-Mendikute et al., 2005). In addition, the histochemical methods are relatively inexpensive compared to biochemical analyses; therefore, we encourage applying these tissue methods in monitoring programs and multi-biomarker approaches.

Biometric measurements

Biometric measurements are the easiest among all biomarkers to study. There are different approaches; some use wet weight, and

some use dry weight; however, all are associated with the length and weight of treated and controlled mussels. Here we present the formulas of some of the most common condition indexes, which can provide information about impacted physiological processes:

CI total – (soft tissue weight/total weight) x 100; CI 2 - soft tissue weight/shell weight;

CI 3 (state index) - soft tissue weight/shell length;

CI 4 (shell component index) - shell weight/shell weight + soft tissue weight;

CI 5 (condition factor) - soft tissue weight/shell lenght³

(Lucas & Beninger, 1985; Rios-Fuster et al., 2022).

In addition, reproduction can be inhibited by sublethal environmental stress because animals reallocate energy away from gamete production and toward defense, and repair mechanisms (Michalek-Wagner & Willis, 2001).

The gonadosomatic index (GSI) is calculated according to the formula (Roff, 1992): GSI = gonadal tissue weight / (gonadal + somatic tissue weight).

Stress-on-stress

According to Viarengo et al. (1995), contaminant exposure can decrease mussel tolerance to anoxia. The stress-on-stress (SOS) response represents the survival of mussels in the air (time to kill 50% of sample: LT_{50} ; Thomas et al., 1999; Hellou & Law, 2003). It appeared as a sensitive and straightforward indicator of mussels' health, and therefore has been since applied in monitoring programs and laboratory studies with mussels. Holwerda et al. (1985) reviewed the general survival of invertebrates in air and listed blue mussels as surviving more than 30 days, the second highest of 22 species tested (Hammen, 1976). SOS response can also significantly reveal mussel exposure to a mixture pollutants very of at low concentrations, resembling field conditions. Moreover, the methodology utilized to evaluate the stress on stress response is simple, rapid, and low in cost and does not require sophisticated equipment. The SOS response was therefore proposed as an index of general stress at the organismal level, and has been since

applied as a monitoring tool for assessing polluted aquatic ecosystems (Hellou & Law, 2003).

Scope for growth

Recent advances in environmental toxicology involving the close coupling of the sensitive stress response (scope for growth -SFG) and pollutant levels in the tissues of mussels have provided a powerful and costeffective method of assessing environmental pollution (Widdows & Donkin, 1992; Widdows, 1998; Widdows et al., 2002). SFG itself results from various vital functions (filtration, ingestion, absorption, and respiration), a technique involving calculating the energy available for growth (Albentosa et al., 2012). This approach complements the established chemical monitoring programs by assessing whether the recorded contaminant levels are causing deleterious effects and whether all are being relevant toxicants measured (Widdows et al., 2002). In addition, SFG is a biomarker at the individual/whole organism level of biological complexity with high ecological relevance. This is very applicable to biomonitoring programs.

Hence, the SFG concept and method are sensitive indicators of environmental pollution in European waters (Widdows et al., 2002; Halldórsson et al., 2005). As explained by Albentosa et al. (2012), in 2007, the Spanish Marine Pollution monitoring program (SMP) conducted by the Spanish Institute of Oceanography added SFG as an environmental assessment technique to be integrated with the chemical parameters. Determining growth in organisms is one of the most sensitive methods available for detecting, quantifying, and identifying changes over time and space to the water quality of marine ecosystems since growth results from a combination of different physiological processes involved in energy acquisition and consumption. In short, it consists of evaluating the energy acquired by an organism after absorbing the food it has ingested. The difference in the organism's energy available for production (growth and reproduction) is lost in the respiratory and excretory processes.

The calculations of SFG are based upon the following equation:

$SFG=(I-F)-R=(I \times AE)-R,$

where I is the consumption of the energy available in the diet, F is the energy lost in the feces, AE is the absorption efficiency, and R is the energy consumed by respiration (Bayne & Newell, 1983).

The clearance rate (CR, expressed in L ind⁻¹ h⁻¹) is calculated from the difference between inlet and outlet concentrations for the experimental system according to the equation (Riisgård, 2001):

CR=f x (Ci-Co)/Ci

where f is the flow of water expressed in L/h, Ci is the inlet concentration, and Co is the outlet concentration, both expressed in particulate volume units, mm³/L.

The organic ingestion rate (OIR, mg POM h⁻¹) was obtained by multiplying the clearance rate by the diet concentration (expressed in mg POM l⁻¹).

The absorption efficiency (AE) is calculated from the percentage of organic matter in the food and the feces according to Conover's ratio (1966);

 $AE=[(F-E)/((1-E) \times F)] \times 100,$

where F is the percentage of organic matter (ash-free dry weight) in the food and E is the percentage of organic matter in the feces.

The absorption rate (AR, mg POM h⁻¹) was obtained by multiplying the ingestion rate by the absorption efficiency (AR=OIR x AE).

Respiration intensity rate

The rate of respiration reflects the metabolic activities of animals and the responses due to changes in the surrounding environment are a good indicator of the adjustment capacity of the organism (Kumar et al., 2012). Bivalve mollusks reflect immediate responses to toxic substances present in the surrounding water through changes in their physiological responses (Basha et al., 1988) and histological arrangement (Kumar et al., 2012). It is known that without time for acclimation, mussels typically reduce their clearance rate (volume of water passing through gills per unit time), thus potentially lowering their intake of oxygen (Aldridge et al., 1987). However, most bivalve mollusks reflect immediate responses to toxic substances

present in the surrounding water by changes in physiological responses (Basha et al., 1988). In most cases, the respiration rate increases with the increase of the pollutant concentration and level of toxicity (Kumar et al., 2012). This is because the organism tries to deliver more oxygen to all tissues and organs triggered by the stress caused by toxic exposure.

Respiration intensity is calculated following Tsekov (1989);

$$[=Q_2/G,$$

where I – respiration rate index; G – weight of the mussels, in grams, Q_2 – oxygen consumed by the mussels between the two measurements (the difference between the oxygen levels before and after 1 h, $Q_2 = Q-Q_1$ hour).

Q is calculated following the formula:

$Q = V x q_{\prime}$

where: Q – total oxygen level; V – water volume, in liters; q –dissolved oxygen levels in 1 liter of water (mg L⁻¹).

Mussel caging and mussel watch programs

Environmental monitoring with mussels is often termed as Mussel Watch Program. Data from such monitoring is available from more than 50 nations, sometimes with data going back to the 1960s (Goldberg, 1986; Cantillo, 1998; Beliaeff et al., 1998). The relationship between the level of waterborne pollutants and bivalve tissue concentrations is well established within the Mussel Watch Program, which monitors over 150 organic and inorganic pollutants, including polycyclic aromatic hydrocarbons (PAHs), polychlorinated biphenyls (PCBs), the pesticide dichlorodiphenyltrichloroethane (DDT), etc. (National Oceanic and Atmospheric Administration, 2018). Since 1983, the Ontario Ministry of Environment (MOE) has used caged mussels (Elliptio complanata Lightfoot, 1786) to monitor contaminants in the Niagara River (Richman et al., 2011). Several studies have suggested that transplanted mussels can also be useful biomonitoring tools for evaluating environmental microplastic pollution (Brate et al., 2018). Mussel caging, transplants, and similar mussel watch programs have yet to be applied in Bulgaria. Gecheva et al. (2020) and Georgieva et al. (2022) are the first to report results from such field experiments in Bulgaria.

The current situation in Bulgaria – challenges and future perspectives

Data on mussels and their various biological disturbances due to pollution in the Black Sea in Bulgaria is extremely scarce. There is some research on the levels of heavy metals and various organic pollutants in the sea and mussels (Stoichev et al., 2007; Rizov & Georgieva, 2010; Georgieva et al., 2016a,b; Peteva et al., 2018a,b; Zhelyazkov et al., 2018; Manev et al., 2020; Bachvarova et al., 2022). At present, there are no studies regarding the spread and bioaccumulation of microplastics or their effects on the marine biota of the Bulgarian part of the Black Sea. Still, Alexandrova et al. (2022) recently provided the first preliminary data on the distribution and accumulation of microplastics in wedge clams (Donax trunculus Linnaeus, 1758) from selected localities along the Bulgarian Black Sea coast.

In sum, most of the results concern the qualities of mussels as a source of protein and therefore, the risk to humans (Peteva et al., 2020; Peycheva et al., 2022) or practices for growing and catching them or their importance for aquaculture (Petrova & Stoykov, 2009; Klisarova et al., 2020). Aquaculture is one of the pillars of the European Union's blue economy strategy. It is the subject of increasing interest on the Bulgarian Black Sea coast, but so far, mainly establishing farms to cultivate in М. galloprovincialis (Zahariev, 2021). Furthermore, during the last years, there has been an increased interest in cultivating Mediterranean mussels in Bulgaria, and several farms have been created in Sozopol, Kavarna, and Balchik (Nikolov et al., 2010). As mussels are considered a healthy food because of their high nutritional value, including high levels of polyunsaturated fatty acids, especially omega-3 fatty acids, recent research on the nutritional characteristics of shellfish from the Bulgarian coast showed high values of unsaturated fatty acids, high protein, and high fat-soluble vitamin content and how shellfish may provide health benefits for local populations (Merdzhanova et al., 2017; Stancheva et al., 2017; Merdzhanova et al., 2021; Panayotova et al., 2021).

Data on changes in different biological tools due to aquatic pollution is limited. Gorinstein et al. (2003) studied antioxidants in *M*. galloprovincialis as an indicator of Black Sea coastal pollution. Ganchev et al. (2012) tested in laboratory conditions how some spirohydantoins and their derivatives affected the mortality rate of M. galloprovincialis. Yakimov et al. (2018) estimated the pro/antioxidant status of M. galloprovincialis from different Bulgarian Black Sea coastal area sites and studied the oxidative stress levels to indicate stressful environmental conditions. In their recent study, Yakimov et al. (2020) further investigated the oxidative stress in different Bulgarian Black Sea bivalves -Chamelea gallina, D. trunculus, M. galloprovincialis, and their bioindicator potential. So did Nikolova et al. (2018) and Tsvetanova et al. (2022), but they investigated oxidative stress regarding seasonal changes. Nechev et al. (2006) and Nechev et al. (2007) followed the lipid and sterol changes due to the effect of cobalt ions on galloprovincialis. Peteva et al. (2018b) М. analyzed the marine toxin levels along the Black Sea food chain (phytoplankton and mussels). They discussed the metabolic changes they could undergo as they moved to higher trophic levels and assessed the potential human risk. In 2017 Vasileva et al. applied the Comet assay as a sensitive tool for genotoxicity assessment of environmental stress in M. galloprovincialis from the Bulgarian Black Sea coast.

Other species as potential bioindicators for aquatic pollution in the Black Sea in Bulgaria

Even though most of the results suggest that *Mytilus* spp. are probably the most suitable mollusk bioindicators, including for the assessment of microplastic pollution, not only in Bulgaria, but worldwide (Jamil et al., 1999; Beyer et al., 2017; Li et al., 2018, 2019; Monteiro et al., 2019; Gunaalan et al., 2020; Li et al., 2021; Abelouah et al., 2023; Xu et al., 2023) there are other species, which might need to be studied too.

Petrova-Pavlova (2014) describes that the sand mussel (*Mya arenaria* Linnaeus, 1758) is invasive species for Black Sea, which originates from the northern part of the Atlantic Ocean. In the Black Sea it was transported, probably in larval stage in 1960s. Along the Bulgarian Black Sea coast this species is distributed everywhere, but the largest aggregations are observed in the south coastal area in front of estuaries and bay aquatories. *Anadaraina equivalvis* (Bruguiere, 1789) is an invasive bivalve of Indo-Pacific fauna, which was first found in the Black Sea in 1968 and it has spread into the whole basin. The habitat of this clam is sandy-muddy bottoms between 3 to 15 m depth. *Chamelea gallina* (Linnaeus, 1758) is widely distributed in the Black Sea up to 25 meters, forming aggregations in the sublittoral zone of sandy ground. This species live buried in the sandy sediment at a depth of 15–20 cm and its life cycle is three years (Petrova-Pavlova, 2014).

The veined rapa whelk (*Rapana venosa* Valenciennes, 1846) is a predatory invasive species from Asia. It was first detected in the Black Sea in 1947 and fed mainly on mussels (*M. galloprovincialis*). The veined rapa whelks are suitable for human consumption and are gathered by divers. Data suggests this species could also be used for environmental pollution biomonitoring (Bat et al., 2000; Bat & Öztekin, 2015; Mülayim & Balkıs, 2015; Zhelyazkov et al., 2018).

The wedge clam (*D. trunculus*) inhabits fine sandy habitats of the upper littoral subzone and feeds by filtration on phytoplankton and suspended particulate matter. In the Bulgarian Black Sea coastal zone, D. trunculus usually dominates between 1.0 and 6.5 m depth, and is exposed to intense wave action and fluctuations of abiotic environmental factors (Gumus et al., Although people 2020). local do not consume wedge clams, traditionally D. trunculus are increasingly being collected with dredges for export due to their high price at foreign markets (Gumus et al., 2020). In this regard, Georgieva et al. (2021) presented results on PAH accumulation in *D. trunculus* from the Bulgarian Black Sea Coast. Georgieva et al. (2021) assessed the state of the marine environment along the Bulgarian Black Sea Coast by analyzing the acetylcholinesterase activity in wedge clam too. Olivieri et al. (2022) reported the first results on the uptake of microplastics in wedge clams from the Mediterranean Sea.

Conclusions

More scientists, among them, biologists and chemists from Bulgaria, are starting to study the negative consequences of Black Sea pollution on various marine species - not only mollusks, but also different economically important fish species. Therefore, we strongly recommend the implementation of a multi-biomarker approach that combines successfully most of the indicated in this review biomarkers for better results, the application of mussel caging, as well as an exchange of collective experience.

Acknowledgments

This study is financed by the European Union - NextGenerationEU, through the National Recovery and Resilience Plan of the Republic of Bulgaria, project № BG-RRP-2.004-0001-C01. The research was supported by ÚNKP-22-4 and ÚNKP-22-5 New National Excellence Program of the Ministry for Culture and Innovation from the source of the National Research, Development and Innovation Fund. Project no. **TKP2021-NKTA-32** was implemented with the support provided from the National Research, Development and Innovation Fund of Hungary, financed under the TKP2021-NKTA funding scheme. The Bolyai Fellowship of the Hungarian Academy of Sciences supported László Antal. The research presented in the article was carried out within the framework of the Széchenyi Plan Plus program with the support of the RRF 2.3.1-21-2022-00008 and GINOP_PLUSZ-2.1.1-21-2022-00245 projects.

References

- Abelouah, M.R., Romdhani, I., Ben-Haddad, M., Hajji, S., De-la-Torre, G.E., Gaaied, S., Barra, I., Banni, M. & Alla, A.A. (2023). survey **Binational** using *Mytilus* galloprovincialis as a bioindicator of microplastic pollution: Insights into chemical analysis and potential risk on humans. Science of the Total 161894. Environment, 870, doi: 10.1016/j.scitotenv.2023.161894
- Akcha, F., Tanguy, A., Leday, G., Pelluhet, L., Budzinski, H. & Chiffoleau, J.F. (2004). Measurement of DNA single-strand breaks in gill and hemolymph cells of mussels, *Mytilus* sp., collected on the French Atlantic Coast. Marine Environmental Research, 58, 753–756. doi: 10.1016/j.marenvres.2004.03.090

- Albentosa, M., Viñas, L., Besada, V., Franco, A. & González-Quijano, A. (2012). First measurements of the scope for growth (SFG) in mussels from a large scale survey in the North-Atlantic Spanish coast. Science of the Total Environment, 435–436, 430–445. doi: 10.1016/j.scitotenv.2012.07.025
- Aldridge, D.W., Payne, B.S. & Miller, A.C. (1987). The effects of intermittent exposure to suspended solids and turbulence on three species of freshwater mussels. Environmental Pollution, 45, 17– 28. doi: 10.1016/0269-7491(87)90013-3
- Alexandrova, A.V., Ignatova-Ivanova, T.V., Bachvarova, D.G., Toschkova, S.G., Doichinov, A.H., Ibryamova, S.F. & Chipev, N.H. (2022). Pilot screening and assessment of microplastic bioaccumulation in wedge clams *Donax trunculus* Linnaeus, 1758 (Bivalvia) from the Bulgarian Black Sea Coast. Acta Zoologica Bulgarica, 74(4), 568-578.
- Anbuselvan, N., Senthil Nathan, D. & Sridharan, M. (2018). Heavy metal assessment in surface sediments off Coromandel Coast of India: implication on marine pollution. Marine Pollution Bulletin, 131, 712–726. doi: 10.1016/j.marpolbul.2018.04.074
- Avio, C.G., Cardelli, L.R., Gorbi, S., Pellegrini, D. & Regoli, F. (2017). Microplastics pollution after the removal of the Costa Concordia wreck: First evidences from a biomonitoring case study. Environmental Pollution, 227, 207-214.

doi: 10.1016/j.envpol.2017.04.066

- Aytan, U., Esensoy Sahin, F.B. & Karacan, F. (2020). Beach litter on Sarayköy Beach (SE Black Sea): density, composition, possible sources and associated organisms. Turkish Journal of Fisheries and Aquatic Sciences, 20, 137–145. doi: 10.4194/1303-2712-v20_2_06
- Aytan, U., Valente, A., Senturk, Y., Usta, R., Sahin, F.B.E., Mazlum, R.E. & Agirbas, E. (2016). First evaluation of neustonic microplastics in Black Sea waters. Marine Environmental Research, 119, 22–30. doi: 10.1016/j.marenvres.2016.05.009
- Bachvarova, D., Stancheva, M., Ignatova-Ivanova, T., Ibryamova, S., Chipev, N. & Alexandrova, A. (2022). Trace metal

accumulation in tissues of wedge clams from sandy habitats of the Bulgarian Black Sea coast. BioRisk, 17, 73–81. doi: 10.3897/biorisk.17.77290

- Bakan, G. & Özkoç, H.B. (2007). An ecological risk assessment of the impact of heavy metals in surface sediments on biota from the mid-Black Sea coast of Turkey. International Journal of Environmental Studies, 64(1), 45-57. doi: 10.1080/00207230601125069
- Barcelona Convention (2016). The 19th Meeting of the contracting parties to the convention for the protection of the marine environment and the coastal region of the Mediterranean. Decision G.22/10. Retrieved from https://wedocs.unep.org/bitstream/
- Barone, G., Dambrosio, A., Storelli, A., Garofalo, R., Busco, V.P. & Storelli, M.M. (2018). Estimated dietary intake of trace metals from swordfish consumption: a human health problem. Toxics, 6, 22. doi: 10.3390/toxics6020022
- Barsiene, J., Syvokiene, J. & Bjornstad, A. (2006). Induction of micronuclei and other nuclear abnormalities in mussels exposed to bisphenol A, diallyl phthalate and tetrabromodiphenyl ether-47. Aquatic Toxicology, 78(S1), 105–108. doi: 10.1016/j.aquatox.2006.02.023
- Bartell, S.M. (2006). Biomarkers, bioindicators, and ecological risk assessment—a brief review and evaluation. Environmental Bioindicators, 1, 39-52. doi: 10.1080/15555270591004920
- Bartoskova, M., Dobsikova, R., Stancova, V., Zivna, D., Blahova, J., Marsalek, P., Zelnickova, L., Bartos, M., Casuscelli Di Tocco, F. & Faggio, C. (2013). Evaluation of ibuprofen toxicity for zebrafish (*Danio rerio*) targeting on selected biomarkers of oxidative stress. Neuroendocrinology Letters, 34(S2), 102–108.
- Basha, S.M., Swami, K.S. & Puspanjali, A. (1988). Ciliary and cardiac activity of freshwater mussel *Lamellidens marginalis* (Lamarck) as an index of evaluating organophosphate toxicity. Journal of Environmental Biology, 9(3), 313–318.
- Bat, L., Gönlügür, G., Andaç, M., Öztürk, M. & Öztürk, M. (2000). Heavy metal concentrations in the sea snail *Rapana venosa*

(Valenciennes, 1846) from Sinop coasts of the Black Sea. Turkish Journal of Marine Sciences, 6, 227–240.

- Bat, L. & Öztekin, H.C. (2015). Heavy metals in *Mytilus galloprovincialis, Rapana venosa* and *Eriphia verrucosa* from the Black Sea coasts of Turkey as bioindicators of pollution. Walailak Journal of Science and Technology, 13, 715–728. Retrieved from https://wjst.wu.ac.th/
- Bayne B.L. & Newell R.C. (1983). 9 -Physiological energetics of marine mollusks. In A.S.M. Saleuddin, K.M. Wilbur (Eds.). The Mollusca. (pp. 407-515). Academic Press. doi: 10.1016/B978-0-12-751404-8.50017-7
- Beliaeff, B., O'Connor, T.P. & Claisse, D. (1998).
 Comparison of chemical concentrations in mussels and oysters from the United States and France. Environmental Monitoring and Assessment, 49, 87-95. doi: 10.1023/A:1005766321323
- Berov, D. & Klayn, S. (2020) Microplastics and floating litter pollution in Bulgarian Black Sea coastal waters. Marine Pollution Bulletin, 156, 111225. doi: 10.1016/j.marpolbul.2020.111225
- Beyer, J., Green, N.W., Brooks, S., Allan, I.J., Ruus, A., Gomes, T., Bråte, I.L.N. & Schøyen, M. (2017). Blue mussels (*Mytilus edulis* spp.) as sentinel organisms in coastal pollution monitoring: A review. Marine Environmental Research, 130, 338-365. doi: 10.1016/j.marenvres.2017.07.024
- Bignell, J.P, Stentiford, G.D., Taylor, N.G.H. & Lyons, B.P. (2011). Histopathology of mussels (sp.) from the Tamar estuary, UK. Marine Environmental Research, 72(1-2), 25-32. doi: 10.1016/j.marenvres.2011.05.004
- Bollen, M., Keppens, S. & Stalmans, W. (1998). Specific features of glycogen metabolism in the liver. Biochemical Journal, 336, 19– 31. doi: 10.1042/bj3360019
- Bolognesi, C., Rabboni, R. & Roggeri, P. (1996). Genotoxicity biomarkers in *M. galloprovincialis* as indicators of marine pollutants. Comparative Biochemistry and Physiology Part C, 113, 319–323. doi: 10.1016/0742-8413(95)02103-5
- Bolognesi, C., Landini, E., Roggieri, P., Fabbri, R. & Viarengo, A. (1999). Genotoxicity

biomarkers in the assessment of heavy metal effects in mussels: experimental studies. Environmental and Molecular Mutagenesis, 33, 287–292.

- Bolognesi, C. & Hayashi, M. (2011). Micronucleus assay in aquatic animals. Mutagenesis, 26(1), 205–213. doi: 10.1093/mutage/geq073
- Bonanno, G. & Orlando-Bonaca, M. (2018). Perspectives on using marine species as bioindicators of plastic pollution. Marine Pollution Bulletin, 137, 209–221. doi: 10.1016/j.marpolbul.2018.10.018
- Bonsignore, M., Manta, D.S., Al-Tayeb Sharif, E.A., D'Agostino, F., Traina, A., Quinci, E.M., Giaramita, L., Monastero, C., Benothman, M. & Sprovieri, M. (2018). Marine pollution in the Libyan coastal area: environmental and risk assessment. Marine Pollution Bulletin, 128, 340–352. doi: 10.1016/j.marpolbul.2018.01.043
- Boran, M. & Altınok, I. (2010). A review of heavy metals in water, sediment and living organisms in the Black Sea. Turkish Journal of Fisheries and Aquatic Sciences, 10, 565-572. doi: 10.4194/trjfas.2010.0418
- Borenfreund, E. & Puerner, J.A. (1985). Toxicity determined in vitro by morphological alterations and neutral red absorption. Toxicology Letters, 24, 119-124. doi: 10.1016/0378-4274(85)90046-3
- Brate, I.L.N., Hurley, R., Iversen, K., Beyer, J., Thomas, K.V., Steindal, C.C., Green, N.W., Olsen, M. & Lusher, A. (2018). *Mytilus* spp. as sentinels for monitoring microplastic pollution in Norwegian coastal waters: a qualitative and quantitative study. Environmental Pollution, 243, 383–393. doi: 10.1016/j.envpol.2018.08.077
- Browne, M.A., Dissanayake, A., Galloway, T.S., Lowe, D.M. & Thompson, R.C. (2008).
 Ingested microscopic plastic translocates to the circulatory system of the mussel, *Mytilus edulis* (L.). Environmental Science & Technology, 42, 5026-5031. doi: 10.1021/es800249a
- Cammilleri, G., Galluzzo, P., Pulvirenti, A., Giangrosso, I.E., Lo Dico, G.M., Montana, G., Lampiasi, N., Mobilia, M.A., Lastra, A., Vazzana, M., Vella, A., La Placa, P., Macaluso, A., & Ferrantelli, V. (2020). Toxic

mineral elements in *Mytilus galloprovincialis* from Sicilian coasts (Southern Italy). Natural Product Research, 34(1), 177–182. doi: 10.1080/14786419.2019.1610963

- Cantillo, A.Y. (1998). Comparison of results of mussel watch programs of the United States and France with worldwide mussel watch studies. Marine Pollution Bulletin, 36, 712-717. doi: 10.1016/S0025-326X(98)00049-6
- Capó, X., Tejada, S., Box, A., Deudero, S. & Sureda, A. (2015). Oxidative status assessment of the endemic bivalve *Pinna nobilis* affected by the oil spill from the sinking of the Don Pedro. Marine Environmental Research, 110, 19–24. doi: 10.1016/j. marenvres.2015.07.013
- Capó, X., Rubio, M., Solomando, A., Alomar, C., Compa, M. & Sureda, A. (2021). Microplastic intake and enzymatic responses in *Mytilus* galloprovincialis reared at the vicinities of an aquaculture station. Chemosphere, 280, 130575. doi: 10.1016/j.chemosphere.2021.130575
- Cappello, T., Maisano, M., D'Agata, A., Natalotto, A., Mauceri, A. & Fasulo, S. (2013). Effects of environmental pollution in caged mussels (*Mytilus galloprovincialis*). Marine Environmental Research, 91, 52-60. doi: 10.1016/j.marenvres.2012.12.010
- Carignan, V. & Villard, M.A. (2002). Selecting indicator species to monitor ecological integrity: review. Environmental Monitoring and Assessment, 78, 45-61. doi: 10.1023/A:1016136723584
- Caro, T. (2010). Conservation by proxy: indicator, umbrella, keystone, flagship, and other surrogate species. Washington, DC: Island Press.
- Christiansen, B.O. (1965). Notes of the littoral fauna of Bear Island. Astarte, 26, 1–15.
- Coatu, V., Oros, A., Țigănuş, D., Shtereva, G. & Bat, L. (2016). Assessment of the contaminants in biota from the Western Black Sea basin in respect with MSFD requirements in the frame of the MISIS project. Cercetări Marine, 46, 82-97. doi: 10.55268/CM.2016.46.82
- Conover, R.J. (1966). Assimilation of organic matter by zooplankton. Limnology and Oceanography, 11, 338–345.
- D'Hont, A., Gittenberger, A., Leuven, R.S. & Hendriks, A.J. (2021). Dropping the microbead: source and sink related microplastic

distribution in the Black Sea and Caspian Sea basins. Marine Pollution Bulletin, 173, 112982. doi: 10.1016/j.marpolbul.2021.112982

- Dall, W.H. (1889). Synopsis of the recent and tertiary Leptonacea of North America and the West Indies. Proceedings of the United States National Museum, 21(1177), 873– 897. doi: 10.5479/si.00963801.21-1177.873
- Dallas, L.J., Bean, T.P., Turner, A., Lyons, B.P. & Jha, A.N. (2013). Oxidative DNA damage may not mediate Ni-induced genotoxicity in marine mussels: Assessment of genotoxic biomarkers and transcriptional responses of key stress genes. Mutation Research/Genetic Toxicology and Environmental Mutagenesis, 754(1-2), 22-31. doi: 10.1016/j.mrgentox.2013.03.009
- Dellali, M., Hedfi, A., Ali, M.B., Noureldeen, A., Darwish, H., Beyrem, H., Gyedu-Ababio, T., Dervishi, A., Karachle, P.K. & Boufahja, F. (2021). Multi-biomarker approach in *Mytilus* galloprovincialis and *Ruditapes decussatus* as a predictor of pelago-benthic responses after exposure to benzo[a]pyrene. Comparative Biochemistry and Physiology Part C, 249, 109141. doi: 10.1016/j.cbpc.2021.109141
- Détrée, C. & Gallardo-Escárate, C. (2017). Polyethylene microbeads induce transcripttional responses with tissue-dependent patterns in the mussel *Mytilus galloprovincialis*. Journal of Molluscan Studies, 83(2), 220-225. doi: 10.1093/mollus/eyx005
- Dirrigl, F.J., Badaoui, Z., Tamez, C., Vitek, C.J. & Parsons, J.G. (2018). Use of the sea hare (*Aplysia fasciata*) in marine pollution biomonitoring of harbors and bays. Marine Pollution Bulletin, 129, 681–688. doi: 10.1016/j.marpolbul.2017.10.056
- Dixon, D.R., Pruski, A.M., Dixon, L.R.J. & Jha, A.N. (2002). Marine invertebrate ecogenotoxicology: a methodological overview. Mutagenesis, 17(6), 495–507. doi: 10.1093/mutage/17.6.495
- Doyotte, A., Cossu, C., Jacquin, M.C., Babut, M. Vasseur, P. (1997). Antioxidant & glutathione and enzymes, lipid peroxidation as relevant biomarkers of experimental or field exposure in the gills and the digestive gland of the freshwater bivalve Unio tumidus. Aquatic Toxicology, 39, doi: 10.1016/S0166-93-110. 445X(97)00024-6

- Drastichová, J., Švestková, E., Lusková, V. & Svobodová, Z. (2005). Cytochemical study of carp neutrophil granulocytes after acute exposure to cadmium. Journal of Applied Ichthyology, 21(3), 215–219. doi: 10.1111/j.1439-0426.2005.00618.x
- Duroudier, N., Katsumiti, A., Mikolaczyk, M., Schäfer, J., Bilbao, E. & Cajaraville, M.P. (2021). Cell and tissue level responses in mussels *Mytilus galloprovincialis* dietarily exposed to PVP/PEI coated Ag nanoparticles at two seasons. Science of the Total Environment, 750, 141303. doi: 10.1016/j.scitotenv.2020.141303.
- EC (2006). Commission Regulation (EC) No 1881/2006 of 19 December 2006 setting maximum levels for certain contaminants in foodstuffs (Text with EEA relevance). Official Journal of the European Union, L 364, 5-24.

Retrieved from https://eurlex.europa.eu

- EC (2008). Directive 2008/105/EC of the European Parliament and of the Council of 16 December 2008 on environmental quality standards in the field of water policy, amending and subsequently repealing Council Directives 82/176/EEC, 83/513/EEC, 84/156/EEC, 84/491/EEC, 86/280/EEC and amending Directive 2000/60/EC of the European Parliament and of the Council. Official Journal of the European Union, L 348, 84-96. Retrieved from https://eurlex.europa.eu
- EC. (2008). Marine Strategy Framework Directive 2008/56/EC of the European Parliament and of the Council of 17 June 2008 establishing a framework for community action in the field of marine environmental policy. Official Journal of the European Union. Retrieved from https://eurlex.europa.eu
- EC. (2010). Commission Decision of 1 September 2010 on criteria and methodological standards on good environmental status of marine waters (notified under document C(2010) 5956) (Text with EEA relevance) (2010/477/EU). Official Journal of the European Union, L 232, 14-24. Retrieved from https://eurlex.europa.eu
- Eggermont, M., Cornillie, P., Dierick, M., Adriaens, D., Nevejan, N., Bossier, P., Van

den Broeck, W., Sorgeloos, P., Defoirdt, T.& Declercq, A.M. (2020). The blue mussel inside: 3D visualization and description of the vascular-related anatomy of *Mytilus edulis* to unravel hemolymph extraction. Scientific Reports, 10, 6773. doi: 10.1038/s41598-020-62933-9

- El-Serafy, S.S., Abdel-Hameid, N.-A.H. & El-Daly, A.A. (2009). Histological and histochemical alterations induced by phenol exposure in *Oreochromis aureus* (Steindachner, 1864) juveniles. Egyptian Journal of Aquatic Biology and Fisheries, 13(2), 151–172. doi: 10.21608/EJABF.2009.2038
- Ersoy, B. & Çelik, M. (2009). Essential elements and contaminants in tissues of commercial pelagic fish from the Eastern Mediterranean Sea. Journal of the Science of Food and Agriculture, 89(9), 1615–1621. doi: 10.1002/jsfa.3646.
- Eurostat. (2016). Agriculture, Forestry and Fishery Statistics. Belgium.
- Fabry, V., Fröhlich, K. & Osvath, I. (1993). Environmental pollution of the Black Sea: A search for answer. IAEA Bulletin, 2, 20-24.
- Falfushynska, H., Sokolov, E.P., Haider, F., Oppermann, C., Kragl, U., Ruth, W., Stock, M., Glufke, S., Winkel, E.J. & Sokolova, I.M. (2019). Effects of a common pharmaceutical, atorvastatin, on energy metabolism and detoxification mechanisms of a marine bivalve *Mytilus edulis*. Aquatic Toxicology, 208, 47–61. doi: 10.1016/j.aquatox.2018.12.022.
- Farrington, J.W., Tripp, B.W., Tanabe, S., Subramanian, A., Sericano, J.L., Wade, T.L. & Knap, A.H. (2016). Edward D. Goldberg's proposal of "The Mussel Watch": reflections after 40 years. Marine Pollution Bulletin, 110, 501-510. doi: 10.1016/j.marpolbul.2016.05.074
- Figueiredo-Fernandes, A., Fontaínhas-Fernandes, A., Rocha, E. & Reis-Henriques, M. (2006). The effect of paraquat on hepatic EROD activity, liver and gonadal histology in males and females of Nile tilapia, Oreochromis niloticus, exposed at different tempeof Environmental ratures. Archives Contamination and Toxicology, 51(4), 626-632. doi: 10.1007/s00244-005-0208-3

- Fossi, M.C., Panti, C., Baini, M., Lavers, J.L. (2018a). A review of plastic-associated pressures: cetaceans of the Mediterranean Sea and Eastern Australian Shearwaters as case studies. Frontiers in Marine Science, 5, 173. doi: 10.3389/fmars.2018.00173
- Fossi, M.C., Pedà, C., Compa, M., Tsangaris, C., Alomar, C., Claro, F., Ioakeimidis, C., Galgani, F., Hema, T., Deudero, S., Romeo, T., Battaglia, P., Andaloro, F., Caliani, I., Casini, S., Panti, C. & Baini, M. (2018b). Bioindicators for monitoring marine litter ingestion and its impacts on Mediterranean biodiversity. Environmental Pollution, 237, 1023–1040. doi: 10.1016/j.envpol.2017.11.019
- Fossi, M.C., Baini, M. & Simmonds, M.P. (2020) Cetaceans as ocean health indicators of marine litter impact at global scale. Frontiers in Environmental Science, 8, 586627. doi: 10.3389/fenvs.2020.586627
- Fraïsse, C., Roux, C., Welch, J.J. & Bierne, N. (2014). Gene-flow in a mosaic hybrid zone: is local introgression adaptive? Genetics, 197(3), 939-951. doi: 10.1534/genetics.114.161380
- Gaitan-Espitia, J.D., Quintero-Galvis, J.F., Mesas,
 A. & D'Elia, G. (2016). Mitogenomics of southern hemisphere blue mussels (Bivalvia: Pteriomorphia): Insights into the evolutionary characteristics of the *Mytilus edulis* complex. Scientific Reports, 6, 26853. doi: 10.1038/srep26853
- Galgani, F., Hanke, G., Werner, S. & De Vrees, L. (2013). Marine litter within the European Marine Strategy Framework Directive. Journal of Marine Sciences, 70(6), 1055-1064. doi: 10.1093/icesjms/fst122
- Ganchev, D., Marinov, M., Marinova, P., Krustev, S., Zlateva, M., Atanasova, N., Nikolov, A. & Stoyanov, N. (2012). Ecotoxicological examination of some spirohydantoins and their derivatives towards Black Sea Mussel (*Mytilus galloprovincialis*). University of Ruse "Angel Kanchev" Proceedings, Biotechnologies and Food Technologies, 51(9.2), 14-17.
- Gecheva, G., Yancheva, V., Velcheva, I. Georgieva, E., Stoyanova, S., Arnaudova, D., Stefanova, V., Georgieva, D., Genina, V., Todorova, B. & Mollov, I. (2020). Integrated monitoring with moss-bag and mussel transplants in reservoirs. Water, 12, 1800. doi: 10.3390/w12061800

- Georgieva S., Stancheva M. & Makedonski L. (2016a). Investigation about the presence of organochlorine pollutants in mussels from the Black Sea, Bulgaria. Ovidius University Annals of Chemistry, 27(1), 8-12. doi: 10.1515/auoc-2016-0006
- Georgieva, S., Stancheva, M., Makedonski, L. & Peteva, Z. (2016b). Organochlorine compounds (PCBs and DDTs) in seafood from the Black Sea, Bulgaria. Journal of International Scientific Publications, Agriculture & Food, 4, 284-291.
- Georgieva, A.P., Alexandrova, A.V., Tsvetanova, E.R. & Chipev, N.H. (2021). State of the marine environment along the Bulgarian Black Sea coast as indicated by acetylcholinesterase activity of wedge clam (*Donax trunculus* Linnaeus, 1758). Ecologia Balkanica, Special Edition 4, 135-143.
- Georgieva, S.K., Stancheva, M.D., Peteva, Z.V., Ivanova, T.I. & Alexandrova, A.V. (2022). Assessment of PAHs accumulation in *Donax trunculus* (Linnaeus, 1758) (Bivalvia, Donacidae) from the Bulgarian Black Sea Coast. BioRisk, 17, 95–104. doi: 10.3897/biorisk.17.77343
- Georgieva, E., Antal, L., Stoyanova, S., Arnaudova, D., Velcheva, I., Iliev, I., Vasileva, T., Bivolarski, V., Mitkovska, V., Chassovnikarova, T., Todorova, B., Uzochukwu, I.E., Nyeste, K. & Yancheva, V. (2022). Biomarkers for pollution in caged mussels from three reservoirs in Bulgaria: a pilot study. Heliyion, 8(3), 09069. doi: 10.1016/j.heliyon.2022.e09069
- Giamberini, L. & Pihan, J.C. (1997). Lysosomal changes in the hemocytes of the freshwater mussel *Dreissena polymorpha* experimentally exposed to lead and zinc. Diseases of Aquatic Organisms, 28, 221–227.
- Goldberg, E. (1975). The mussel watch a first step in global marine monitoring. Marine Pollution Bulletin, 6, 111.
- Goldberg, E. (1980). The International MUSSEL WATCH - Report of a workshop sponsored by the Environmental Studies Board, Commission on Natural Resources and the National Research Council. Washington D.C: National Academy of Sciences.
- Goldberg, E.D. (1986). The mussel watch concept. Environmental Monitoring and

Assessment, 7, 91-103. DOI: 10.1007/BF00398031

- Gomez-Delgado, A.I., Tibon, J., Silva, M.S., Lundebye, A.-K., Agüera, A., Rasinger, J.D., Strohmeier, T. & Sele, V. (2023). Seasonal variations in mercury, cadmium, lead and arsenic species in Norwegian blue mussels (*Mytilus edulis* L.) – Assessing the influence of biological and environmental factors. Journal of Trace Elements in Medicine and Biology, 76, 127110. doi: 10.1016/j.jtemb.2022.127110
- Gómez-Mendikute, A., Elizondo, M., Venier, P. & Cajaraville, M.P. (2005). Characterization of mussel gill cells in vivo and in vitro. Cell Tissue Research, 21(1), 131–140. doi: 10.1007/s00441-005-1093-9
- Gomoiu, M.T. (1992). Marine eutrophication syndrome in the north-western part of the Black Sea. In R.A. Vollenweider, R. Marchetti, R. Viviani (Eds.). Marine coastal eutrophication. (pp. 683–692). Amsterdam: Elsevier.
- González-Fernández, D., Cózar, A., Hanke, G., Viejo, J., Morales-Caselles, C., Bakiu, R., Barceló, D., Bessa, F., Bruge, A., Cabrera, M., Castro Jiménez, J., Constant, M., Crosti, R., Galletti, Y., Kideys, A.E., Machitadze, N., Pereira De Brito, J., Pogojeva, M., Ratola, N., Rigueira, J., Rojo-Nieto, E., Savenko, O., Schöneich-Argent, R., Siedlewicz, G., Suaria, G. & Tourgeli, M. (2021). Floating macrolitter leaked from Europe into the ocean. Nature Sustainability, 4, 474-483. doi: 10.1038/s41893-021-00722-6
- Gorinstein, S., Moncheva, S., Katrich, E., Toledo, F., Arancibia, P., Goshev, I. & Trakhtenberg, S. (2003). Antioxidants in the black mussel (*Mytilus galloprovincialis*) as an indicator of Black Sea coastal pollution. Marine Pollution Bulletin, 46(10), 1317-1325. doi: 10.1016/S0025-326X(03)00239-X.
- Gosling, E.M. (1992). Systematics and geographic distribution of *Mytilus*. In E.M. Gosling (Ed.). The mussel *Mytilus*: ecology, physiology, genetics and culture. (pp. 1–20). Amsterdam: New York: Elsevier.
- Guendouzi, Y., Soualili, D.L., Fowler, S.W. & Boulahdid, M. (2020). Environmental and

human health risk assessment of trace metals in the mussel ecosystem from the southwestern Mediterranean. Marine Pollution Bulletin, 151, 110820. doi: 10.1016/j.marpolbul.2019.110820

- Guerlet, E., Ledy, K., Meyer, A. & Giamberini, L. (2007). Towards a validation of a cellular biomarker suite in native and transplanted zebra mussels: a 2-year integrative field study of seasonal and pollution induced variations. Aquatic Toxicology, 81, 377-388. doi: 10.1016/j.aquatox.2006
- Guieu, C., Martin, J.M., Tankere, S.P.C., Mousty, F., Trincherini, P., Bazot, M. & Dai, M.H. (1998). On trace metal geochemistry in the Danube River and western Black Sea. Estuarine, Coastal and Shelf Science, 47, 471-485. doi: 10.1006/ecss.1998.0377
- Gumus, M.R., Todorova, V.R. & Panayotova, M.D. (2020). Recent observations on the size structure of *Donax trunculus* (Linnaeus, 1758) and *Chamelea gallina* (Linnaeus, 1758) in the Bulgarian Black Sea as status indicators of commercially exploited shellfish under the Marine Strategy Framework Directive (MSFD). Ecologia Balkanica, Special Edition, 3, 63–71.
- Gunaalan, K., Fabbri, E. & Capolupo, M. (2020). The hidden threat of plastic leachates: A critical review on their impacts on aquatic organisms. Water Research, 184, 116170. doi: 10.1016/j.watres.2020.116170
- Halldórsson, H.P., Svavarsson, J. & Granmo, A. (2005). The effect of pollution on scope for growth of the mussel (*Mytilus edulis* L.) in Iceland. Marine Environmental Research, 59, 47–64. doi: 10.1016/j.marenvres.2004.02.001
- Hammen, C.S. (1976). Estuarine Processes, Vol 1: Uses, stresses and adaptations to estuaries. New York: Academic Press.
- Heddle, J.A., Hite, M., Kirkhart, B., Mavoumin, K., Mac Gregor, J.T., Newell, G.T. & Salamone, M.F. (1983). The induction of micronuclei as a measure of genotoxicity. A report of the U.S. Environmental Protection Agency Gene-Tox Program. Mutation Research, 123, 61–118. doi: 10.1016/0165-1110(83)90047-7
- Hellou, J. & Law, R.J. (2003). Stress on stress response of wild mussels, *Mytilus edulis* and *Mytilus trossulus*, as an indicator of

ecosystem health. Environmental Pollution, 126, 407–416. doi: 10.1016/s0269-7491(03)00231-8

- Hinton, D.E. & Lauren, D.J. (1990). Liver structural alterations accompanying chronic toxicity in fishes: potential biomarkers of exposure. In J.F. McCarthy, L.R. Shugart (Eds.). Biomakers of Environmental Contamination. (pp. 17-57). Boca Raton, FL: Lewis Publishers.
- Holwerda, D.A., De Zwaan, A., Kluytmans, J.H. & Zandee, D.I. (1985). Metabolic adaptations to environmental anoxia in the intertidal bivalve mollusc *Mytilus edulis* L. Netherlands Journal of Zoology, 36(3), 322-343. doi: 10.1163/002829686X00117
- Hu, W., Culloty, S., Darmody, G., Lynch, S., Davenport, J., Ramirez-Garcia, S., Dawson, K., Lynch, I., Doyle, H. & Sheehan, D. (2015). Neutral red retention time assay in determination of toxicity of nanoparticles. Marine Environmental Research, 111, 158-161. doi: 10.1016/j.marenvres.2015.05.007
- IARA (Excecutive agency for fish and aquacultures). (2017). Register of fish farms from 3.07.2017. (In Bulgarian).
- IMAP (Integrated Monitoring and Assessment Program). (2016). Review of the current state of knowledge regarding marine litter in wastes dumped at sea under the London Convention and Protocol. Final Report. pp. 35.
- Jamil, A., Lajtha, K., Radan, S., Ruzsa, G., Cristofor, S. & Postolache, C. (1999). Mussels as bioindicators of trace metal pollution in the Danube Delta of Romania. Hydrobiologia, 392, 143–158. doi: 10.1023/A:1003555130831
- Jha, A.N., Dogra, Y., Turner, A. & Millward, G.E. (2005). Impact of low doses of tritium on the marine mussel, *Mytilus edulis*: genotoxic effects and tissue-specific bioconcentration. Mutation Research, 586, 47–57. doi: 10.1016/j.mrgentox.2005.05.008
- Joint Nature Conservation Committee. (2008). UK Biodiversity Action Plan. Priority Habitat Descriptions. Retrieved from http://jncc.defra.gov.uk/page-5705
- Kent, M.L., Benda, S., St-Hilaire, S. & Schreck, C.B. (2013). Sensitivity and specificity of histology for diagnoses of four common

pathogens and detection of nontarget pathogens in adult Chinook salmon (*Oncorhynchus tshawytscha*) in fresh water. Journal of Veterinary Diagnostic Investigation, 25(3), 341–351. doi: 10.1177/1040638713482124

- Kilinc, S.O. (2017). Microplastic contamination in Black Sea beach samples from the Turkish Coast – a low cost approach. doi: 10.13140/RG.2.2.33019.41762
- Klisarova, D., Gerdzhikov, D., Kostadinova, G., Petkov, G., Cao, X., Song, C. & Zhou, Y. (2020) Bulgarian marine aquaculture: Development and prospects – A review. Bulgarian Journal of Agricultural Science, 26(1), 163-174.
- Kouali, H., Chaouti, A., Achtak, H., Elkalay, K. & Dahbi, A. (2022). Trace metal contents in the mussel *Mytilus galloprovincialis* from Atlantic coastal areas in northwestern Morocco: Levels of contamination and assessment of potential risks to human health. Marine Pollution Bulletin, 179, 113680. doi: 10.1016/j.marpolbul.2022.113680
- Koukouzika, N. & Dimitriadis, V.K. (2008). Aspects of the usefulness of five marine pollution biomarkers, with emphasis on MN and lipid content. Marine Pollution Bulletin, 56, 941– 949. doi: 10.1016/j.marpolbul.2008.01.043
- Kourdali, S., Boudjema, K., Meknachi, A., Bounakous, N., Jaouadi, B., Mechri, S. & Badis, A. (2022). An ecotoxicological approach for assessing marine pollution: Comparative study of multi-responses of marine mussels, *Mytilus galloprovincialis* and *Perna perna*, exposed to pollutant heavy metals (copper and lead). Regional Studies in Marine Science, 52, 102334. doi: 10.1016/j.rsma.2022.102334
- Kumar, S., Pandey, R.K., Das, S. & Das, V.K. (2012). Dimehoate alters respiratory rate and gill histopathology in freshwater mussel *Lamellidens marginatus* (Lamarck). Journal of Applied Biosciences, 38(2), 154–158.
- Lau, P.S. & Wong, H.L. (2003). Effect of size, tissue parts and location on six biochemical markers in the green-lipped mussel, *Perna viridis*. Marine Pollution Bulletin, 46, 1563– 1572. doi: 10.1016/S0025-326X(03)00321-7
- Lebreton, L.C., Van der Zwet, J., Damsteeg, J.-W., Slat, B., Andrady, A. & Reisser, J.

(2017). River plastic emissions to the world's oceans. Nature Communications, 8, 15611. doi: 10.1038/ncomms15611

- Lechner, A., Keckeis, H., Lumesberger-Loisl, F., Zens, B., Krusch, R., Tritthart, M., Glas, M. & Schludermann, E. (2014). The Danube so colourful: a potpourri of plastic litter outnumbers fish larvae in Europe's second largest river. Environmental Pollution, 188, 177–181. doi: 10.1016/j.envpol.2014.02.006
- Li, J.N., Green, C., Reynolds, A., Shi, H.H. & Rotchell, J.M. (2018). Microplastics in mussels sampled from coastal waters and supermarkets in the United Kingdom. Environmental Pollution, 241, 35–44. doi: 10.1016/j.envpol.2018.05.038
- Li, J., Lusher, A.L., Rotchell, J.M., Deudero, S., Turra, A., Bråte, I.L.N., Sun, C., Hossain, M.S., Li, Q., Kolandhasamy, P. & Shi, H. (2019). Using mussel as a global bioindicator of coastal microplastic pollution. Environmental Pollution, 244, 522-533. doi: 10.1016/j.envpol.2018.10.032
- Li, J., Wang, Zh., Rotchell, J.M., Shen, X., Li, Q. & Zhu, J. (2021). Where are we? Towards an understanding of the selective accumulation of microplastics in mussels. Environmental Pollution, 286, 117543. doi: 10.1016/j.envpol.2021.117543
- Liu, C., Chang, V.W.C., Gin, K.Y.H.& Nguyen, V.T. (2014). Genotoxicity of perfluorinated chemicals (PFCs) to the green mussel (*Perna viridis*). Science of the Total Environment, 487, 117-122. doi: 10.1016/j.scitotenv.2014.04.017.
- Livingstone, D.R. (1992). Mussels and environmental contaminants: molecular and cellular aspects. In E. Gosling (Ed.). The Mussel *Mytilus*: ecology, physiology, genetics and culture. (pp. 589). Amsterdam; New York: Elsevier.
- Livingstone, D.R. (2001). Contaminant-stimulated reactive oxygen species production and oxidative damage in aquatic organisms. Marine Pollution Bulletin, 42(8), 656-66. doi: 10.1016/s0025-326x(01)00060-1
- Lowe, D.M. & Pipe, R.K. (1994). Contaminant induced lysosomal membrane damage in marine mussel digestive cells—an in vitro study. Aquatic Toxicology, 30, 357–365. doi: 10.1016/0166-445X(94)00045-X

- Lowe, D.M., Fossato, V.U., Depledge, M.H. (1995). Contaminant induced lysosomal membrane damage in blood cells of mussels *M. galloprovincialis* from the Venice Lagoon: an in vitro study. Marine Ecology-Progress Series, 129, 189-196.
- Lucas, A. & Beninger, P.G. (1985). The use of physiological condition indices in marine bivalve aquaculture. Aquaculture, 44, 187– 200. doi: 10.1016/0044-8486(85)90243-1
- Lynch, S.A., Morgan, E., Carlsson, J., Mackenzie, C., Wooton, E.C., Rowley, A.F., Malham, S., Culloty, S.C. (2014). The health status of mussels, *Mytilus* spp., in Ireland and Wales with the molecular identification of a previously undescribed haplosporidian. Journal of Invertebrate Pathology, 118, 59– 65. doi: 10.1016/j.jip.2014.02.012
- Majone, F., Brunetti, R., Gola, I. & Levis, A.G. (1987). Persistence of micronuclei in the marine mussel, *Mytilus galloprovincialis*, after treatment with mitomycin C. Environmental Pollution, 191, 157–161. doi: 10.1016/0165-7992(87)90147-3
- Majone, F., Brunetti, R., Fumagalli, O., Gabriele, M. & Levis, A.G. (1990). Induction of micronuclei by mitomycin C and colchicine in the marine mussel *Mytilus* galloprovincialis. Mutation Research, 244, 147–151. doi: 10.1016/0165-7992(90)90064-q
- Mamaca, E., Bechmann, R.K., Torgrimsen, S., Aas, E., Bjørnstad, A., Baussant, T. & Le Floch, S. (2005). The neutral red lysosomal retention assay and Comet assay on haemolymph cells from mussels (*Mytilus edulis*) and fish (*Symphodus melops*) exposed to styrene. Aquatic Toxicology, 75, 191– 201. doi: 10.1016/j.aquatox.2005.08.001
- Manev, I., Kirov, V. & Neshovska, H. (2020). Heavy metals accumulation in Black Sea ecosystems: seawater, sediment, algae, benthic organisms. Tradition and modernity in veterinary medicine, 5, 2(9), 88–99.
- Mayer, F.L., Versteeg, D.J., McKee, M.J., Folmar, L.C., Graney, R.L., McCume, D.C. & Rattner, B.A. (1992). Physiological and nonspecific biomarkers. In R.J. Huggett, R.A. Kimerle, P.M. Mehrle Jr., H.L. Bergman (Eds.). Biomarkers: Biochemical, physiological, and histological markers of anthropogenic stress. (pp. 125–153). USA, FL, Boca Raton: Lewis Publishers.

- McDougall, K.D. (1943). Sessile marine invertebrates of Beaufort, North Carolina: a study of settlement, growth, and seasonal fluctuations among pile-dwelling organisms. Ecological Monographs, 13, 321-374. doi: 10.2307/1943225
- Mee, L.D. (1992). The Black Sea in crisis a need for concerted international action. Ambio (Sweden), 21, 278-286.
- Merdzhanova, A., Ivanov, I., Dobreva, D., & Makedonski, L. (2017). Fish lipids as a valuable source of polyunsaturated fatty acids. Acta Scientifica Naturalis, 4(1), 70-75. doi: 10.1515/asn-2017-0011
- Merdzhanova, A., Panayotova, V., Dobreva, D.
 A., & Stancheva, R. (2019). Effect of thermal stress on the biologically active lipids of *Mytilus galloprovincialis*. Bulgarian Chemical Communications, 51, 256–261.
- Merdzhanova, A., Panayotova, V., Dobreva, D.A. & Peycheva, K. (2021). Can fish and shellfish species from the Black Sea supply health beneficial amounts of bioactive fatty acids? Biomolecules, 11, 1661. doi: 10.3390/biom11111661
- Michalek-Wagner, K. & Willis, B.L. (2001). Impacts of bleaching on the soft coral *Lobophytum compactum*. I. Fecundity, fertilization and offspring viability. Coral Reefs, 19, 231–239. doi: 10.1007/s003380170003
- Miladinova, S., Macias, D., Stips, A. & Garcia-Gorriz, E. (2020). Identifying distribution and accumulation patterns of floating marine debris in the Black Sea. Marine Pollution Bulletin, 153, 110964. doi: 10.1016/j.marpolbul.2020.110964
- Ministry of Agriculture and Food. (2016). Annual report on the condition and development of agriculture Bulgaria. (In Bulgarian)
- Moncheva, S., Stefanova, K., Krastev, A., Apostolov, A., Bat, L., Sezgin, M., Sahin, F. & Timofte, F. (2016). Marine litter quantification in the Black Sea: a pilot assessment. Turkish Journal of Fisheries and Aquatic Sciences, 16, 213–218. doi: 10.4194/1303-2712-v16_1_22.
- Monteiro, R., Costa, S., Coppola, F., Freitas, R., Vale, C. & Pereira, E. (2019). Toxicity beyond accumulation of titanium after exposure of *Mytilus galloprovincialis* to spiked seawater. Environmental Pollution, 244, 845-854. doi: 10.1016/j.envpol.2018.10.035

- Moore, M.N. (1985). Cellular response to pollutants. Marine Pollution Bulletin, 16, 134–139. doi: 10.1016/0025-326X(85)90003-7
- Moore, M.N. (1988). Cytochemical responses of the lysosomal system and NADPHferrihemoprotein reductase in molluscan digestive cells of environmental and experimental exposure to xenobiotics. Marine Ecology Progress Series, 46, 81–89.
- Moore, M.N., Wedderburn, R.J., Lowe, D.M., Depledge, M.H. (1996). Lysosomal reaction to xenobiotics in mussel hemocytes using BODIPY-FL-verapamil. Marine Environmental Research, 42, 99–105. doi: 10.1016/0141-1136(96)00082-7
- Moore, M.N. & Willows, R.I. (1998). A model for cellular uptake and intracellular behaviour of particulate-bound micropollutants. Marine Environmental Research, 46, 509– 514. doi: 10.1016/S0141-1136(97)00078-0
- Moore, M.N., Viarengo, A., Donkin, P. & Hawkins, A.J.S. (2007). Autophagic and lysosomal reactions to stress in the hepato- pancreas of blue mussels. Aquatic Toxicology, 84, 80-91. doi: 10.1016/j.aquatox.2007.06.007
- Moore, M.N., Readman, J.A.J., Readman, J.W., Lowe, D.M., Frickers, P.E. & Beesley, A. (2009). Lysosomal cytotoxicity of carbon nanoparticles in cells of the molluscan immune system: an in vivo study. Nanotoxicology, 3, 40-45. doi: 10.1080/17435390802593057
- Müftüoğlu, A.E. (2013). Transportation of pollutants of the Danube river into the(western) Black Sea. Asian Journal of Chemistry, 25, 5013-5018. doi: 10.14233/ajchem.2013.14279
- Mülayim, A. & Balkıs, H. (2015). Toxic metal (Pb, Cd, Cr, and Hg) levels in *Rapana venosa* (Valenciennes, 1846), *Eriphia verrucosa* (Forskal, 1775), and sediment samples from the Black Sea littoral (Thrace, Turkey). Marine Pollution Bulletin, 95, 215–222. doi: 10.1016/j.marpolbul.2015.04.016
- Naimo, T.J. (1995). A review of the effects of heavy metals on freshwater mussels. Ecotoxicology, 4, 341-362. doi: 10.1007/BF00118870
- Nechev, J., Stefanov, K. & Popov, S. (2006). Effect of cobalt ions on lipid and sterol metabolism in the marine invertebrates *Mytilus galloprovincialis* and *Actinia equine*. Comparative Biochemistry and Physiology Part A, 144, 112–118. doi: 10.1016/j.cbpa.2006.02.022

- Nechev, J., Stefanov, K., Nedelcheva, D. & Popov, S. (2007). Effect of cobalt ions on the metabolism of some volatile and polar compounds in the marine invertebrates *Mytilus galloprovincialis* and *Actinia equine*. Comparative Biochemistry and Physiology Part B, 146, 568–575. doi: 10.1016/j.cbpb.2006.12.017
- Nikolov, G.Y., Atanasov, A.P., Georgie, D.M. & Raichev, E.G. (2010). Analysis of the plankton in the area around the cape Maslen Nos, Bulgaria: possibilities for cultivation of Mediterranean mussels (*Mytilus galloprovincialis*). Ecologia Balkanica, 2, 15-18.
- Nikolova G., Karamalakova Y., Zheleva A., Stratev D., Vashin I., Zhelyazkov G. & Gadjeva V. (2018). Comparative analysis of real-time oxidative stress biomarkers measured in mussels (*Mytilus galloprovincialis*) and veined rapa whelks (*Rapana venosa*) in relation to two seasons - An electron paramagnetic resonance study. Bulgarian Chemical Communications, 50C, 58–63.
- NOAA (National Oceanic and Atmospheric Administration). (2018). Mussel Watch Programme: National Status and Trends. Retrieved from https://www.fisheries.noaa.gov.
- NRC (National Research Council). (1987). Biological markers in environmental health research. Environmental Health Perspectives, 74, 3–9.
- Olivieri, Z., Cesarini, G., Orsini, M., De Santis, S. & Scalici, M. (2022). Uptake of microplastics in the wedge clam *Donax trunculus*: first evidence from the Mediterranean Sea. Water, 14, 4095. doi: 10.3390/w14244095
- Orlando-Bonaca, M., Avio, C.G. & Bonanno, G. (2022). 6 - Marine organisms as bioindicators of plastic pollution. Plastic Pollution and Marine Conservation (Approaches to Protect Biodiversity and Marine Life), 187-248. doi: 10.1016/B978-0-12-822471-7.00003-1
- OSPAR Commission. (2009). Marine litter in the North-East Atlantic Region: assessment and priorities for response. London, United Kingdom, pp. 127.
- Özden, Ö., Ulusoy, S., & Erkan, N. (2010). Study on the behavior of the trace metal and macro minerals in *Mytilus galloprovincialis* as a bioindicator species; the case of

Marmara Sea, Turkey. Journal Für Verbraucherschutz Und Lebensmittelsicherheit, 5, 407–412. doi: 10.1007/s0000 3-009-0544-8

- Özsoy, E. & Ünlüata, U. (1997). Oceanography of the Black Sea: a review of some recent results. Earth-Science Reviews, 42, 231-272. doi: 10.1016/S0012-8252(97)81859-4
- Oztekin, A. & Bat, L. (2017). Microlitter pollution in sea water: a preliminary study from Sinop Sarikum coast of the southern Black Sea. Turkish Journal of Fisheries and Aquatic Sciences, 17, 1431–1440. doi: 10.4194/1303-2712-v17_6_37
- Ozturk, B, & Ozturk, A.A. (1996). On the biology of the Turkish Strait System. Bulletin de l'Institut océanographique de Monaco, 17, 205–221.
- Panayotova, V., Merdzhanova, A., Stancheva, R., Dobreva, D., Peycheva, K. & Makedonski, L. (2021). Farmed mussels (*Mytilus galloprovincialis*) from the Black Sea reveal seasonal differences in their neutral and polar lipid fatty acids profile. Regional Studies in Marine Science, 44, 101782. doi: 10.1016/j.rsma.2021.101782
- Parolini, M., Binelli, A., Cogni, D. & Provini, A. (2010). Multi-biomarker approach for the evaluation of the cyto-genotoxicity of paracetamol on the zebra mussel (*Dreissena polymorpha*). Chemosphere, 79(5), 489-498. doi: 10.1016/j.chemosphere.2010.02.053
- Pathan, T.S., Thete, P.B., Shinde, S.E., Sonawane, D.L. & Khillare, Y.K. (2009). Histochemical changes in the liver of freshwater fish, *Rasbora daniconius*, exposed to paper mill effluent. Emirates Journal of Food and Agriculture, 21(2), 71–78. doi: 10.9755/ejfa.v21i2.5166
- Peteva, Z., Georgieva, S., Krock, B. & Stancheva, M. (2018a). Selected contaminants in fish and mussels from the Bulgarian Black Sea. CBU International Conference on Innovations in Science and Education, March 21-23, 2018, Prague, Czech Republic. doi: 10.12955/cbup.v6.1307
- Peteva, Z., Stancheva, M., Georgieva, S., Gerasimova, A., Makedonski, L. (2018b). Phycotoxin profile of plankton net and shellfish samples from Bulgarian Black Sea South coast: a case study. In Gastescu, P., Bretcan, P. Water resources and wetlands (Ed. 2018, pp. 224-231). 4th International

Conference Water resources and wetlands, 5-9 September 2018, Tulcea (Romania).

- Peteva, Z., Georgieva, S., Krock, B., Gerasimova, A., Stancheva, M. & Merdzhanova, A. (2020). Lipophilic marine biotoxins in mussels from Bulgarian Coast and dietary intake of different population groups. Proceedings of the Nutrition Society, 79 (OCE2), E325. doi: 10.1017/S0029665120002736
- Petrova, E. & Stoykov, S. (2011). Investigations of some bivalve mollusks in Bourgas Bay (Bulgarian Black Sea coast). Macedonian Journal of Animal Science, 1(1), 223–226.
- Petrova-Pavlova, E. (2014). Non-marine resources and their exploitation along the Bulgarian Black Sea coast. Agricultural Science and Technology, 6(2), 215-218.
- Peycheva, K., Panayotova, V., Stancheva, R., Makedonski, L., Merdzhanova, A., Cicero, N., Camilleri, G., & Fazio, F. (2021a). Trace elements and omega-3 fatty acids of Black Sea (Bulgaria) bivalve species *Mytilus* galloprovincialis, Chamelea gallina and Donax trunculus: Human health risk. Natural Product Research, 7, 1–8. doi: 10.1080/14786 419.2021.1921770
- Peycheva, K., Panayotova, V., Stancheva, R., Makedonski, L., Merdzhanova, A., Cicero, N., Parrino, V., & Fazio, F. (2021b). Trace elements and omega-3 fatty acids of wild and farmed mussels (*Mytilus galloprovincialis*) consumed in Bulgaria: Human health risks. International Journal of Environmental Research and Public Health, 18, 10023. doi: 10.3390/ijerp h1819 10023
- Peycheva, K., Panayotova, V., Stancheva, R., Makedonski, L., Merdzhanova, A., Cammilleri, G., Ferrantelli, V., Calabrese, V., Cicero, N. & Fazio, F. (2022). Effect of steaming on chemical composition of Mediterranean mussel (*Mytilus galloprovincialis*): Evaluation of potential risk associated with human consumption. Food Science & Nutrition, 10(9), 3052–3061. doi: 10.1002/fsn3.2903
- Phuong, N.N., Poirier, L., Pham, Q.T., Lagarde, F. & Zalouk-Vergnoux, A. (2017). Factors influencing the microplastic contamination of bivalves from the French Atlantic coast: location, season and/or mode of life? Marine Pollution Bulletin, 129(2), 664-674. doi: 10.1016/j.marpolbul.2017.10.054

- Pojar, I. & Stock, F. (2019). Microplastics in surface waters from the northwestern Black Sea: an abundance and composition approach. In Geophysical Research Abstracts. (pp. EGU2019-8357). Presented at the EGU General Assembly 2019.
- Pojar, I., Kochleus, C., Dierkes, G., Ehlers, S.M., Reifferscheid, G. & Stock, F. (2021). Quantitative and qualitative evaluation of plastic particles in surface waters of the Western Black Sea. Environmental Pollution, 268, 115724. doi: 10.1016/j.envpol.2020.115724
- Powell, E.N. & Cummins, H. (1985). Are molluscan maximum life spans determined by longterm cycles in benthic communities? Oecologia, 67, 177-182. doi: 10.1007/BF00384281
- Regoli, F. (1992). Lysosomal responses as a sensitive stress index in biomonitoring heavy-metal pollution. Marine Ecology Progress Series, 84, 63-69.
- Richman, L.A., Hobson, G., Williams, D.J. & Reiner,
 E. (2011.) The Niagara River mussel biomonitoring program (*Elliptio complanata*): 1983–2009. Journal of Great Lakes Research, 37(2), 213–225. doi: 10.1016/j.jglr.2011.03.012
- Riisgård, H.U. (2001). On measurement of filtration rates in bivalves-the stony road to reliable data: review and interpretation. Marine Ecology Progress Series, 215, 307–310.
- Riisgård, H.U., Larsen, P.S. & Pleissner, D. (2014). Allometric equations for maximum filtration rate in blue mussels *Mytilus edulis* and importance of condition index. Helgoland Marine Research, 68, 193–198. doi: 10.1007/s10152-013-0377-9
- Rios-Fuster B., Alomar C., Capó X., González G.P., Garcinuño Martínez R.M., Rojas D.L.S., Silva M., Hernando P.F., Solé M., Freitas R. & Deudero S. (2022). Assess-ment of the impact of aquaculture facilities on transplanted mussels (*Mytilus galloprovincialis*): Integrating plasticizers and physiological analyses as a biomonitoring strategy. Journal of Hazardous Materials, 424, 127264. doi: 10.1016/j.jhazmat.2021.127264
- Rizov, T. & Georgieva, St. (2010). Polychlorinated biphenyls and organochlorine pesticides in black mussel and goby from Black Sea, Bulgaria. Scripta Scientifica Medica, 42(4), 241-244.
- Roff, D.A. (1992). Evolution of life histories: theory and analysis. New York: Chapman and Hall.

- Ryberg, M.W., Hauschild, M.Z., Wang, F., Averous-Monnery, S. & Laurent, A. (2019). Global environmental losses of plastics across their value chains. Resources, Conservation and Recycling, 151, 104459. doi: 10.1016/j.resconrec.2019.104459
- Sanjuan, A., Zapata, C. & Alvarez, G. (1994). *Mytilus galloprovincialis* and *M. edulis* on the coasts of the Iberian Peninsula. Marine Ecology Progress Series, 113, 131–146.
- Sari, E., Çağatay, M.N., Acar, D., Belivermiş, M., Kılıç, Ö., Arslan, T.N., Tutay, A., Kurt, M.A. & Sezer, N. (2018). Geochronology and sources of heavy metal pollution in sediments of Istanbul Strait (Bosporus) outlet area, SW Black Sea, Turkey. Chemosphere, 205, 387-395. doi: 10.1016/j.chemosphere.2018.04.096
- Săvucă, A., Strungaru, S.-A., Nicoară, M. & Plăvan, G. (2017). A study regarding the pollution of sediments from Romanian Black Sea coast with microplastic fibers. Analele Științifice ale Universității "Alexandru Ioan Cuza" din Iași, s. Biologie animal, 63, 85–90.
- Scott, N., Porter, A., Santillo, D., Simpsona, H., Lloyd-Williams, S. & Lewis, C. (2019).
 Particle characteristics of microplastics contaminating the mussel *Mytilus edulis* and their surrounding environments. Marine Pollution Bulletin, 146, 125–133. doi: 10.1016/j.marpolbul.2019.05.041
- Secrieru, D. & Secrieru, A. (2002). Heavy metal enrichment of man-made origin of superficial sediment on the continental shelf of the North-western Black Sea. Estuarine, Coastal and Shelf Science, 54, 513-526. doi: 10.1006/ecss.2000.0671
- Siddig, A.A.H., Ellison, A.M., Ochs, A., Villar-Leeman, C. & Lau, M.K. (2016). How do ecologists select and use indicator species to monitor ecological change? Insights from 14 years of publication in Ecological Indicators. Ecological Indicators, 60, 223– 230. doi: 10.1016/j.ecolind.2015.06.036
- Simeonova, A. & Chuturkova, R. (2019). Marine litter accumulation along the Bulgarian Black Sea coast: categories and predominance. Waste Management, 84, 182– 193. doi: 10.1016/j.wasman.2018.11.001.
- Simeonova, A., Chuturkova, R. & Yaneva, V. (2017). Seasonal dynamics of marine litter along the Bulgarian Black Sea coast.

Marine Pollution Bulletin, 119, 110–118. doi: 10.1016/j.marpolbul.2017.03.035.

- Singh, R.N. (2014). Effects of dimethoate (EC 30%) on gill morphology, oxygen consumption and serum electrolyte levels of Common carp, *Cyprinus carpio* (Linn). International Journal of Scientific Research in Environmental Sciences, 2(6), 192–198. doi: 10.12983/ijsres-2014-p0192-0198
- Slobodnik, J., Alexandrov, B., Komorin, V., Mikaelvan, A.S., Guchmanidze, A., Arabidze, A. & Korshenko, A. (2018). VII. Descriptor 10: Part I - Marine litter. In National Pilot Monitoring Studies and Joint Open Sea Surveys in Georgia, Russian Federation and Ukraine, 2017. (pp. 573). Final Report on EU/UNDP Project "Improving Environmental Monitoring in the Black Sea-Phase II (EMBLAS-II)".
- Smythe, C. & Cohen, P. (1991). The discovery of glycogenin and the priming mechanism for glycogen biogenesis. European Journal of Biochemistry, 200, 625–631. doi: 10.1111/j.1432-1033.1991.tb16225.x
- Sommerwerk, N., Bloesch, J., Paunovic, M., Baumgartner, C., Venohr, M., Schneider-Jacoby, M., Hein, T. & Tockner, K. (2010). Managing the world's most international river: the Danube River Basin. Marine and Freshwater Research, 61, 736-748.
- Stancheva, M., Merdzhanova, A., & Dobreva, D. (2017). Fat soluble vitamins, cholesterol and fatty acid composition of wild and farmed Black mussel (*Mytilus galloprovincialis*) consumed in Bulgaria. Journal of Aquatic Food Product Technology, 26(2), 181-191. doi: 10.1080/10498850.2015.1108378
- Stanev, E.V. & Ricker, M. (2019). The fate of marine litter in semi-enclosed seas: a case study of the Black Sea. Frontiers in Marine Science, 6, 660. doi: 10.3389/fmars.2019.00660
- Stimpson, W. (1860). A trip to Beaufort. N. Carolina. American Journal of Science, 29, 442–445.
- Stoichev, T., Makedonski, L., Trifonova, T., Stancheva, M. & Ribarova, F. (2007). DDT in fish from the Bulgarian region of the Black Sea. Chemistry and Ecology, 23(3), 191-200. doi: 10.1080/02757540701339851
- Strokal, V., Kuiper, E.J., Bak, M.P., Vriend, P., Wang, M., van Wijnen, J. & Strokal, M. (2022). Future microplastics in the Black Sea:

River exports and reduction options for zero pollution. Marine Pollution Bulletin, 178, 113633. doi: 10.1016/j.marpolbul.2022.113633

- Suaria, G., Melinte-Dobrinescu, M.C., Ion, G. & Aliani, S. (2015). First observations on the abundance and composition of floating debris in the north-western Black Sea. Marine Environmental Research, 107, 45– 49. doi: 10.1016/j.marenvres.2015.03.011.
- Sukhotin, A.A., Strelkov, P.P., Maximovich, N.V. & Hummel, H. (2007). Growth and longevity of *Mytilus edulis* (L.) from northeast Europe. Marine Biology Research, 3, 155-167. doi: 17451000701364869
- Tedesco, S., Doyle, H., Redmond, G. & Sheehan, D. (2008). Gold nanoparticles and oxidative stress in *Mytilus edulis*. Marine Environmental Research, 66, 131-133. doi: 10.1016/j.marenvres.2008.02.044
- Terzi, Y. & Seyhan, K. (2017). Seasonal and spatial variations of marine litter on the southeastern Black Sea coast. Marine Pollution Bulletin, 120, 154–158. doi: 10.1016/j.marpolbul.2017.04.041.
- Terzi, Y., Erüz, C. & Özşeker, K. (2020). Marine litter composition and sources on coasts of south-eastern Black Sea: a long-term case study. Waste Management, 105, 139–147. doi: 10.1016/j.wasman.2020.01.032
- Thomas, R.E., Harris, P.M. & Rice, S.D. (1999). Survival in air of *Mytilus trossulus* following long-term exposure to spilled Exxon Valdez crude oil in Prince Williams sound. Comparative Biochemistry and Physiology Part C, 122, 147–152. doi: 10.1016/s0742-8413(98)10098-1
- Todorova, B., Todorova-Bambaldokova, D., Stoyanova, S., Georgieva, E., Velcheva, I. & Yancheva, V. (2023). Microplastic pollution – are there potential toxic threats for aquatic animals in Bulgaria? Zoonotes, 212, 1-4.
- Topçu, E.N., Tonay, A.M., Dede, A., Öztürk, A.A. & Öztürk, B. (2013). Origin and abundance of marine litter along sandy beaches of the Turkish Western Black Sea coast. Marine Environmental Research, 85, 21–28. doi: 10.1016/j.marenvres.2012.12.006
- Topçu, N.E., Turgay, E., Yardımcı, R.E., Topaloğlu, B., Yuksek, A., Steinum, T.M., Karataş, S. & Ozturk, B. (2019). Impact of excessive sedimentation caused by anthro-

pogenic activities on benthic suspension feeders in the Sea of Marmara. Journal of the Marine Biological Association UK, 99, 1075– 1086. doi:10.1017/S0025315418001066

- Topping, G. & Mee, L.D. (1998). Overview. In L.D. Mee, G. Topping (Eds.). Black Sea pollution assessment, Black Sea environmental series. (pp. 1-29). New York: United Nation Publications.
- Tsekov, A. (1989). Studies on transferrin polymerphism in carp and its resistance to oxygen deficiency. Genetics and Selection, 22(6), 517– 522. (In Bulgarian)
- Tsvetanova, E., Georgieva, A., Chipev, N. & Alexandrova, A. (2022). Seasonal changes in the pro/antioxidant status of mussels *Mytilus galloprovincialis* (Lamarck, 1819) from Bulgarian Black Sea coastal habitats. BioRisk, 17, 241–251. doi: 10.3897/biorisk.17.77279
- Uluturhan, E., Darilmaz, E., Kontas, A., Bilgin, M., Alyuruk, H., Alta, O. & Sevgi, S. (2019). Seasonal variations of multi-biomarker responses to metals and pesticides pollution in *M. galloprovincialis* and *T. decussatus* from Homa Lagoon, Eastern Aegean Sea. Marine Pollution Bulletin, 141, 176–186. doi: 10.1016/j.marpolbul.2019.02.035
- UNEP Regional Seas Program. (1974). Retrieved from https://www.unep.org/explore-topics/
- Urban, N.A., Swihart, R.K., Malloy, M.C. & Dunning Jr., J.B. (2012). Improving selection of indicator species when detection is imperfect. Ecological Indicators, 15, 188-197. doi: 10.1016/j.ecolind.2011.09.031
- Urban-Malinga, B., Wodzinowski, T., Witalis, B., Zalewski, M., Radtke, K. & Grygiel, W. (2018). Marine litter on the seafloor of the southern Baltic. Marine Pollution Bulletin, 127, 612–617. doi: 10.1016/j.marpolbul.2017.12.052
- Van Cauwenberghe, L. & Janssen, C.R. (2014). Microplastics in bivalves cultured for human consumption. Environmental Pollution, 193, 65-70. doi: 10.1016/j.envpol.2014.06.010
- Van Cauwenberghe, L., Claessens, M., Vandegehuchte, M.B. & Janssen, C.R. (2015). Microplastics are taken up by mussels (*Mytilus edulis*) and lugworms (*Arenicola marina*) living in natural habitats. Environmental Pollution, 199, 10-17. doi: 10.1016/j.envpol.2015.01.008
- Vasileva, B., Yakimov, L., Kukurina, B., Georgieva, M., Miloshev, G. & Chipev, N. (2017). Comet assay - a sensitive tool for genotoxicity

assessment of environmental stress in *Mytilus galloprovincialis* from the Bulgarian Black Sea coast. BioDiscovery, 20, 19265. doi: 10.3897/biodiscovery.20.e19265

- Venier, P. & Canova, S. (1996). Formation of DNA adducts in the gill tissue of *Mytilus galloprovincialis* treated with benzo[a]pyrene. Aquatic Toxicology, 34(2), 119-133. doi: 10.1016/0166-445X(95)00035-3
- Venier, P., Maron, S. & Canova, S. (1997). Detection of micronuclei in gill cells and haemocytes of mussel exposed to bezon(a)pyrene. Mutation Research, 390, 33–44. doi: 10.1016/s0165-1218(96)00162-0
- Viarengo, A. & Canesi, L. (1991). Mussels as biological indicators of pollution. Aquaculture, 94(2–3), 225-243. doi: 10.1016/0044-8486(91)90120-V
- Viarengo, A., Canesi, L., Pertica, M., Mancinelli, G., Accomando, R., Smaal, A.C. & Orunesu, M. (1995). Stress on Stress response: A simple monitoring tool in the assessment of a general stress syndrome in mussels. Marine Environmental Research, 39, 245-248. doi: 10.1016/0141-1136(94)00075-Z
- Vidal-Liñán, L., Bellas, J., Campillo, J.A. & Beiras, R. (2010). Integrated use of antioxidant enzymes in mussels, *Mytilus galloprovincialis*, for monitoring pollution in highly productive coastal areas of Galicia (NW Spain). Chemosphere, 78, 265–272. doi: 10.1016/j.chemosphere.2009.10.060
- von Moos, N., Burkhardt-Holm, P. & Khöler, A. (2012). Uptake and effects of microplastics on cells and tissue of the blue mussel *Mytilus edulis* L. after an experimental exposure. Environmental Science & Technology, 46, 11327-11335. doi: 10.1021/es302332w
- Ward, E.J. & Shumway, S.E. (2004). Separating the grain from the chaff: particle selection in suspension- and deposit-feeding bivalves. Journal of Experimental Marine Biology and Ecology, 300, 83–130. doi: 10.1016/j.jembe.2004.03.002
- Wegner, A., Besseling, E., Foekema, E., Kamermans, P. & Koelmans, A. (2012). Effects of nanopolystyrene on the feeding behavior of the blue mussel (*Mytilus edulis* L). Environmental Toxicology and Chemistry, 31, 2490-2497. doi: 10.1002/etc.1984
- Wester, P.W. & Canton, J.H. (1991). The usefulness of histopathology in aquatic toxicity

studies. Comparative Biochemistry and Physiology, 100(1-2), 115-117. doi: 10.1016/0742-8413(91)90135-g

- Widdows, J., & Donkin, P. (1992). Chapter 8: mussels and environmental contaminants: bioaccumulation and physiological aspects. In E. Gosling (Ed.). The mussel *Mytilus* (pp. 383–424). Amsterdam: Elsevier Press.
- Widdows, J. (1998). Chapter 9: marine and estuarine invertebrate toxicity tests. In P. Calow (Ed.). Handbook of ecotoxicology (pp. 145–166). Oxford: Blackwell Science
- Widdows, J., Donkin, P., Staff, F.J., Matthiessen, P., Law, R.J., Allen, Y.T., Thain, J.E., Allchin, C.R. & Jones, B.R. (2002). Measurement of stress effects (scope for growth) and contaminant levels in mussels (*Mytilus edulis*) collected from the Irish Sea. Marine Environmental Research, 53, 327–356. doi: 10.1016/s0141-1136(01)00120-9
- Wolf, J. & Wolfe, M. (2005). A brief overview of non-neoplastic hepatic toxicity in fish. Toxicologic Pathology, 33(1), 75–85. doi: 10.1080/01926230590890187
- Wrisberg, M.N. & van der Gaag, M.A. (1992). In vivo detection of genotoxicity in waste water from a wheat and rye straw paper pulp factory. Science of the Total Environment, 121, 95-108. doi: 10.1016/0048-9697(92)90309-g.
- Wrisberg, M.N., Bilbo, C.M. & Spliid, H. (1992). Induction of micronuclei in hemocytes of *Mytilus edulis* and statistical analysis. Ecotoxicology and Environmental Safety, 23, 191–205. doi: 10.1016/0147-6513(92)90058-b
- Xu, M., Zhang, Y., Cao, S., Li, Y., Wang, J., Dong, H. & Wang, Y. (2023). A simulated toxic assessment of cesium on the blue mussel *Mytilus edulis* provides evidence for the potential impacts of nuclear wastewater discharge on marine ecosystems. Environmental Pollution, 316, 120458. doi: 10.1016/j.envpol.2022.120458
- Yakimov L., Tsvetanova E., Georgieva A., Petrov L., Alexandrova A. (2018). Assessment of the oxidative status of black sea mussels (*Mytilus galloprovincialis* Lamarck, 1819) from Bulgarian coastal areas with introduction of specific oxidative stress index. Journal of Environmental Protection and Ecology, 19(4), 1614–1622.
- Yakimov, L.P., Tsvetanova, E.R., Georgieva, A.P., Nenkova, G.T., Chipev, N.H. & Alexan-

drova, A.V. (2020). Comparative analysis of the oxidative stress in Bulgarian Black-Sea bivalves and their bioindicator potential. Acta Zoologica Bulgarica, Suppl. 15, 147-153.

- Yancheva, V., Velcheva, I., Stoyanova, S. & Georgieva, E. (2015). Fish in ecotoxicological studies. Ecologia Balkanica, 7(1), 149-169.
- Yancheva, V., Stoyanova, S., Georgieva, E. & Velcheva, I. (2018). Mussels in ecotoxicological studies – are they better than fish? Ecologia Balkanica, 10(1), 57-84.
- Yancheva, V., Stoyanova, S., Todorova, B., Georgieva, E. & Velcheva, I. (2022). Zebra mussel (*Dreissena polymorpha* Pallas, 1771): the invasive bioindicator for freshwater quality? Zoonotes, 197, 1-4.
- Yigiterhan, O. & Murray, J.W. (2008). Trace metal composition of particulate matter of the Danube River and Turkish rivers draining into the Black Sea. Marine Chemistry, 111, 63-76. doi: 10.1016/j.marchem.2007.06.019
- Yurdun, T., Kaleağasıoğlu, F., Guven, K.C. & Ozturk, B. (1995). Investigations on the pollution of İstanbul Strait (Bosphorus) by M/S Rab Union-18 sunk with live sheep. Turkish Journal of Marine Sciences, 1, 55–61.
- Zahariev, D. (2021). Editorial Note: Black Sea a pearl in the crown of Bulgaria. Part III. Future. Acta Scientifica Naturalis, 8(3), 1-4. doi: 10.2478/asn-2021-0022
- Zaitsev, Y.P. (1992). Recent changes in the trophic structure of the Black Sea. Fisheries Oceanography, 1(2), 180–189. doi: 10.1111/j.1365-2419.1992.tb00036.x
- Zaitsev, Y.P. & Mamaev, V. (1997). Marine biological diversity in the Black Sea: a study of change and decline. GEF Black Sea Environmental Programme. New York: United Nations Publications.

Zhelyazkov, G., Yankovska-Stefanova, T., Mineva, E., Stratev, D., Vashin, I., Dospatliev, L., Valkova, E. & Popova, T. (2018). Risk assessment of some heavy metals in mussels (*Mytilus galloprovincialis*) and veined rapa whelks (*Rapana venosa*) for human health. Marine Pollution Bulletin, 128, 197–201. doi: 10.1016/j.marpolbul.2018.01.024

> Received: 13.03.2023 Accepted: 24.03.2023