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Molecular Mechanisms of FSH Muscular Dystrophy Pathogenesis

Peter L. Jones

University of Massachusetts Medical School Worcester

Et al.

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Molecular mechanisms of FSH muscular dystrophy pathogenesis

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Departments of Cell and Developmental Biology & Neurology

Facioscapulohumeral Muscular Dystrophy (FSHD)

Most prevalent muscular dystrophy afflicting children and adults (~1:7,000-15,000)

Autosomal dominant

Facio: refers to face

Scapulo: refers to shoulders

Humeral: refers to humerus (upper arm bone)

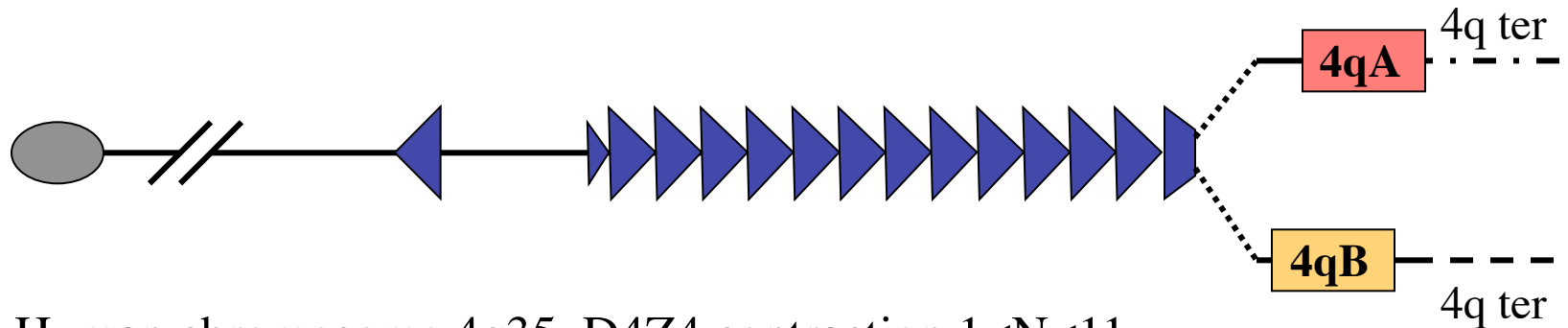


Winging on both sides in a patient with FSHD due to weakness of all the scapula stabilizing muscles

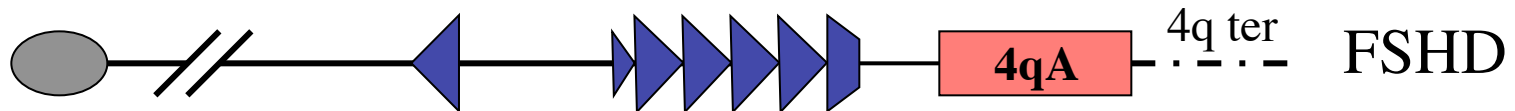
**Great genetic and clinical heterogeneity
Each patient may differ in severity
Most patients exhibit symptoms by age 20
>50% of patients retain ability to walk**

The FSHD1 genetic lesion is a deletion in a tandem repeat array at 4q35

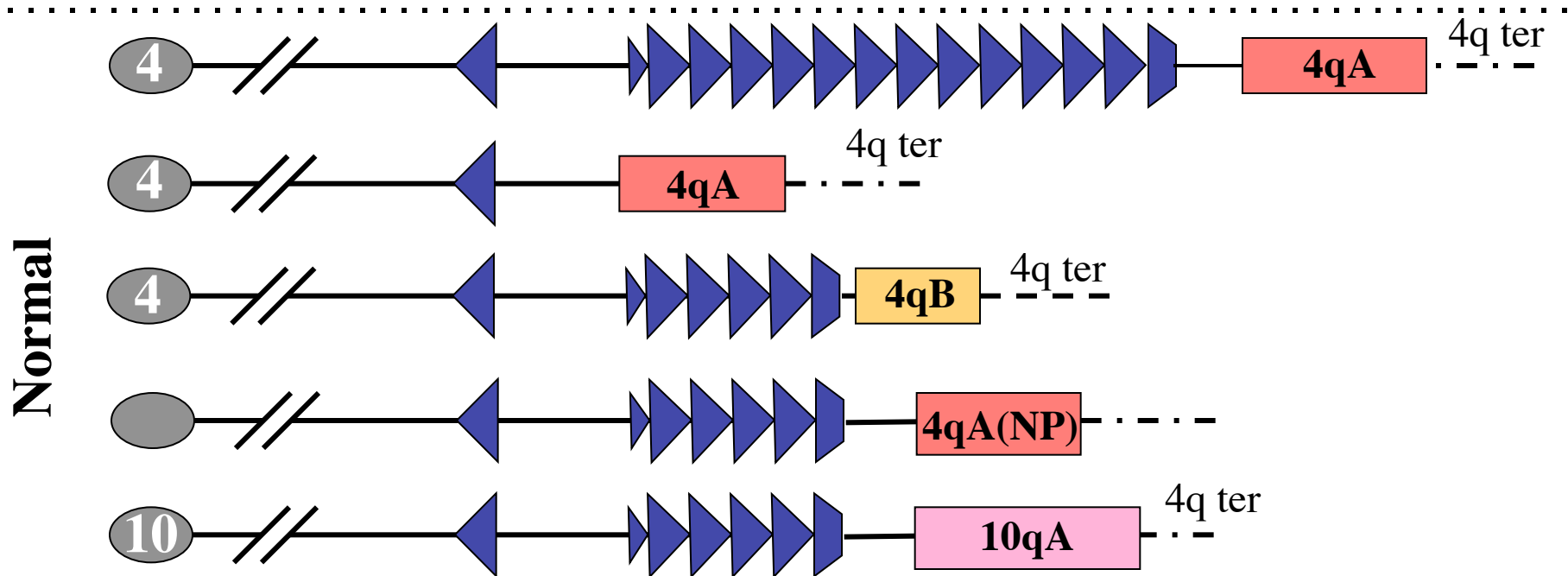
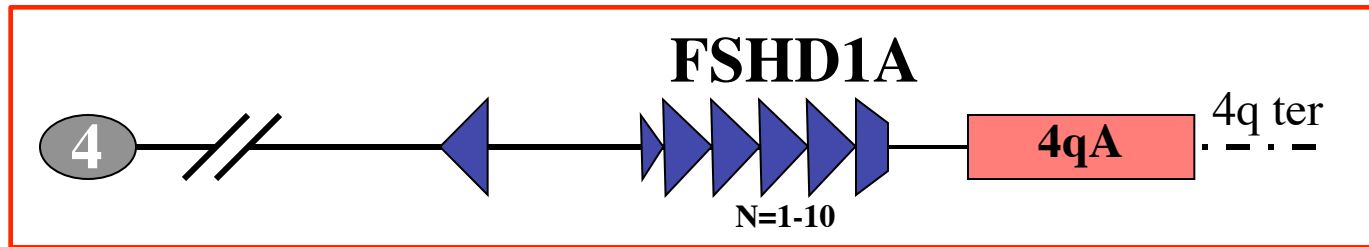
Human chromosome 4q35, Normal D4Z4 n=11-150



Human chromosome 4q35, D4Z4 contraction $1 < N < 11$



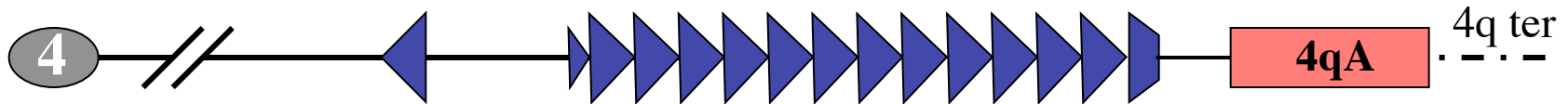
A pathogenic FSHD1 deletion is complex



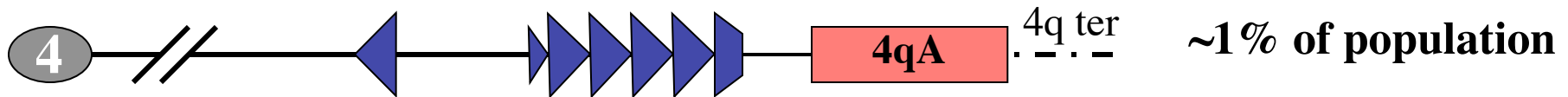
- The deletion itself is not pathogenic
- The 4qA sub-telomere is permissive, not pathogenic
- The 4qB sub-telomere is not permissive
- Requires at least 1 D4Z4 repeat unit for FSHD
- Chromosome 10 arrays (devoid of chr. 4 D4Z4) are not linked to FSHD

A putatively pathogenic FSHD1 deletion shows very low penetrance

Normal

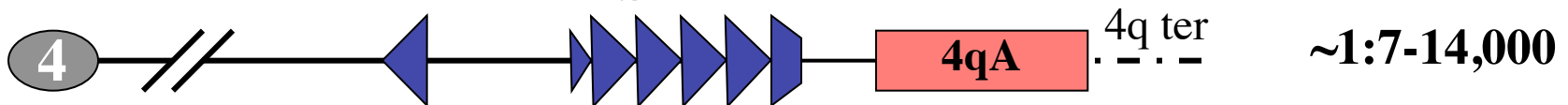


Unaffected



Scionti *et al.* (2012) *J Med Genet* 49:171

FSHD1A

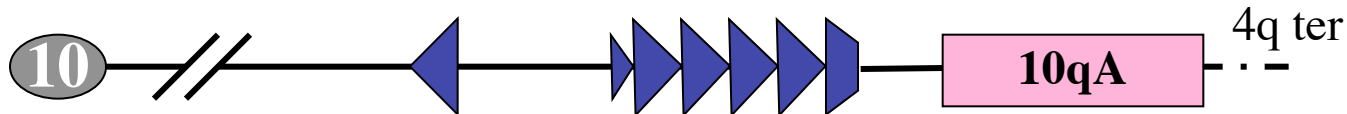
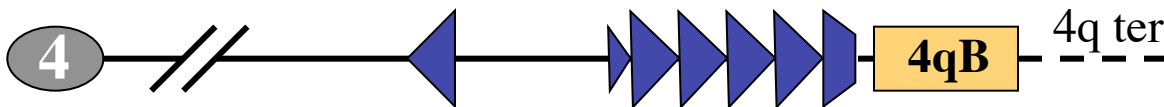
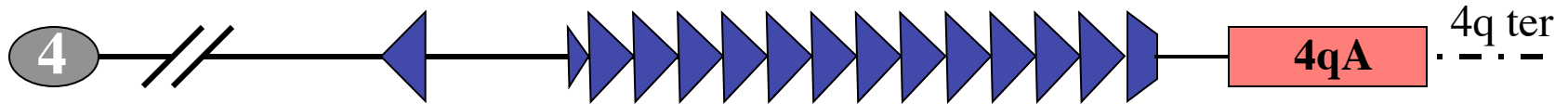


The deletion itself is not pathogenic

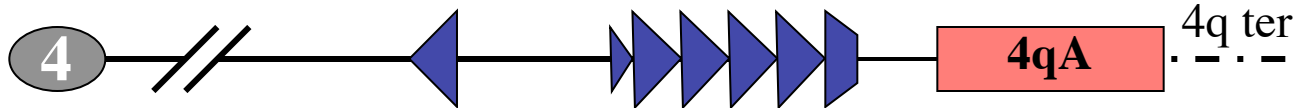
The 4qA sub-telomere is permissive, not pathogenic

FSHD2 is independent of the contraction

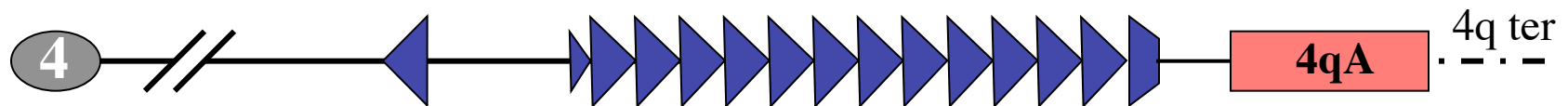
Normal



FSHD1



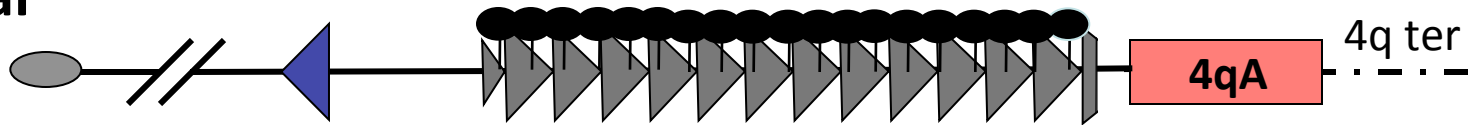
FSHD2



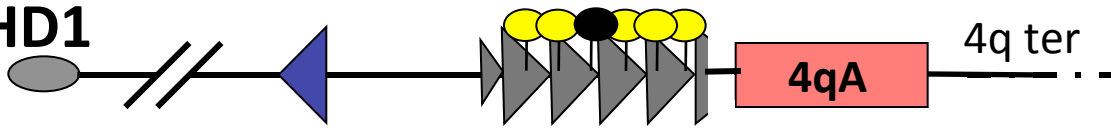
- The 4qA sub-telomere is required for FSHD1 and 2
- At least 1 D4Z4 is required for FSHD1 and 2

FSHD is linked to D4Z4, the A type subtelomere and the epigenetic status of the 4q35 D4Z4 repeat

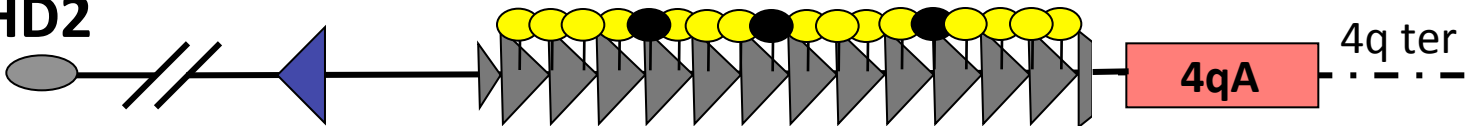
Normal



FSHD1



FSHD2



● = Hypermethylated CpGs
more heterochromatic

● = Hypomethylated CpGs
more euchromatic

The 3 types of FSHD are linked by epigenetic dysregulation

FSHD1: Dominant deletions at 4q35 D4Z4 array

Apparently low penetrance

DNA Hypomethylation of shortened 4q35

FSHD2: SMCHD1 inactivating mutations

--ATPase chromatin remodeling protein

--Modifier of metastable epialleles

DNA Hypomethylation of 4q35 and 10q26 arrays

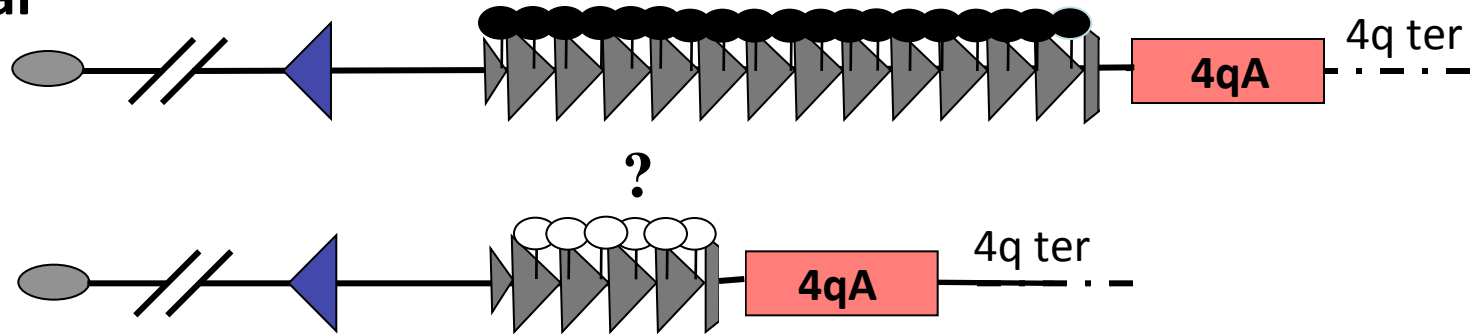
IFSHD: Infantile form of FSHD1 or FSHD2, much more severe

DNA Hypomethylation of FSHD1 or 2

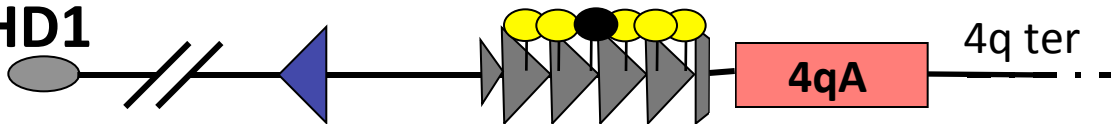


FSHD is linked to the A type subtelomere and the epigenetic status of the 4q35 D4Z4 repeat

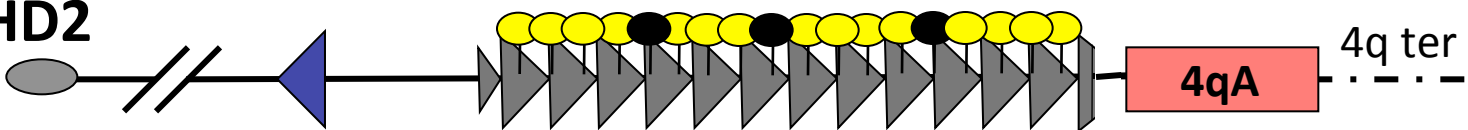
Normal



FSHD1



FSHD2



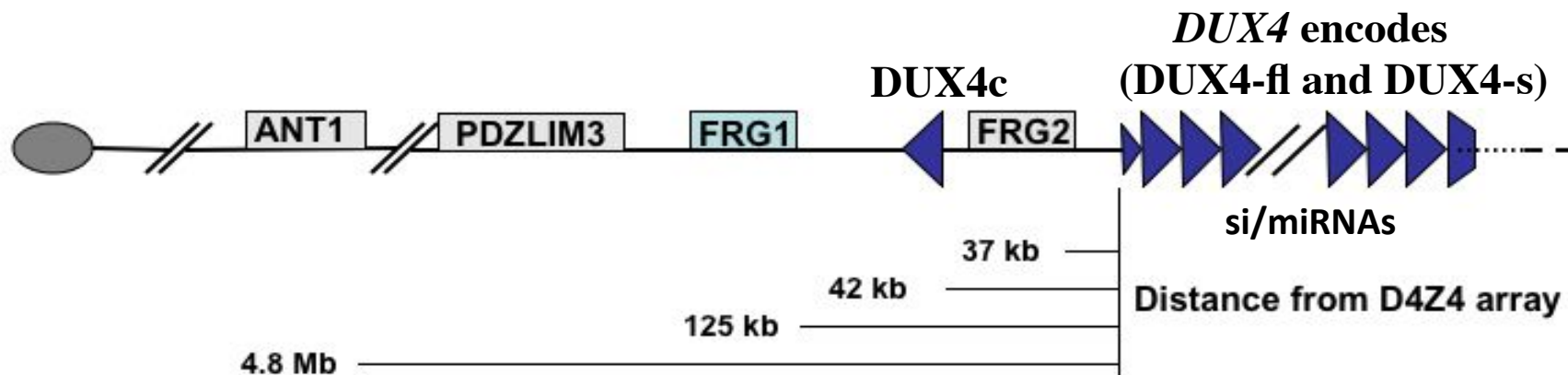
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FSHD results from an epigenetic-mediated dysregulation of gene repression

- **DNA methylation**
- **Histone modifications**
- **Chromatin structure**
- **Long non-coding RNAs**
- **Nuclear organization**
- **High variability within the clinical population**
 - Severity**
 - Age of onset**
 - Gene expression**

Which gene(s) is responsible for FSHD pathology?



The FSHD causal gene:

1. Should be misexpressed in FSHD (Up or down)

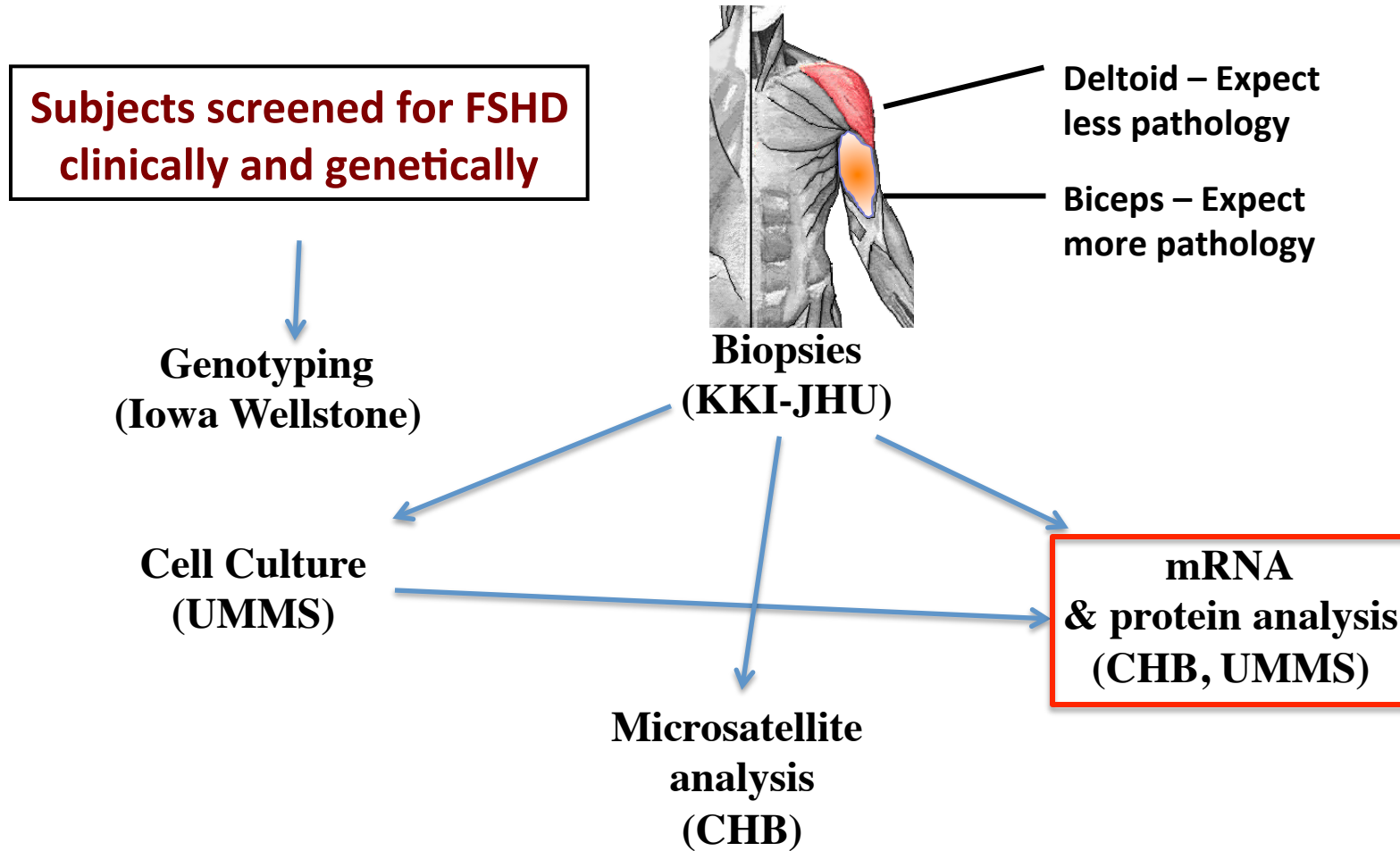
→ mRNA, protein, cell type, developmental timing

→ adversely affect skeletal muscle and potentially vasculature

2. Explain the 4qA linkage and under epigenetic repression

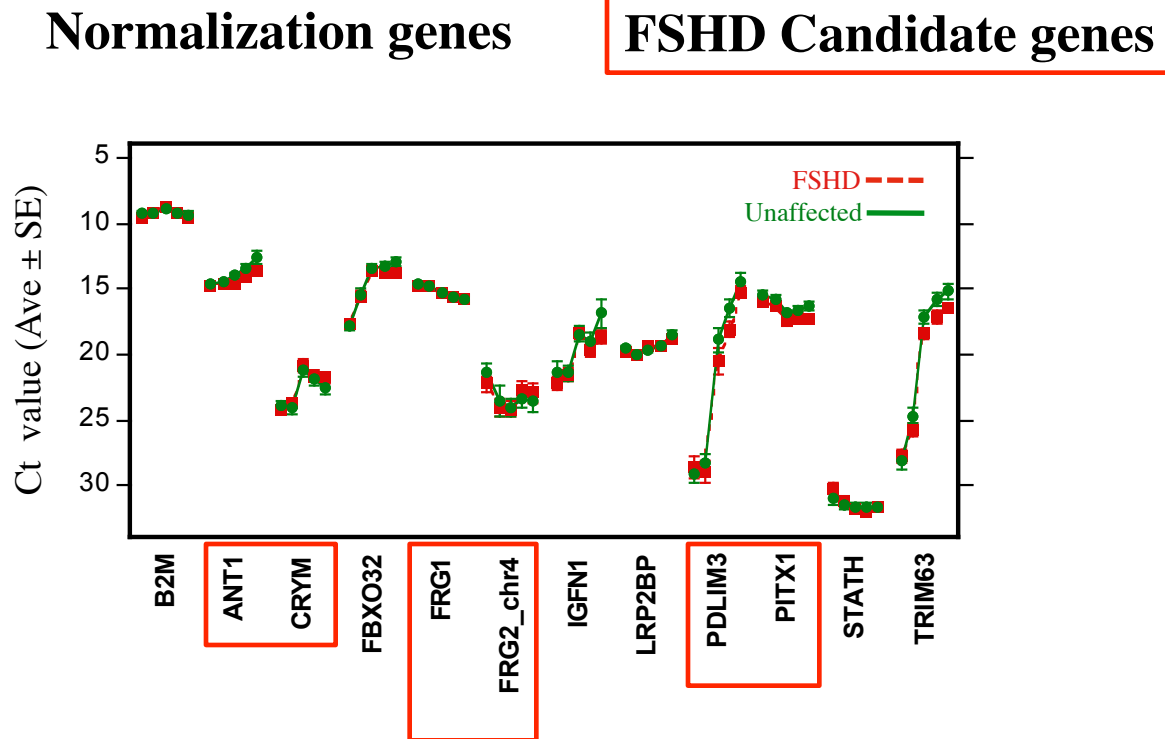
Both FRG1 and DUX4 produce phenotypes consistent with FSHD when overexpressed in animal models

Wellstone family cohorts of muscle biopsies and myogenic cell cultures



FSHD1 affected and genetically unaffected 1st degree relatives

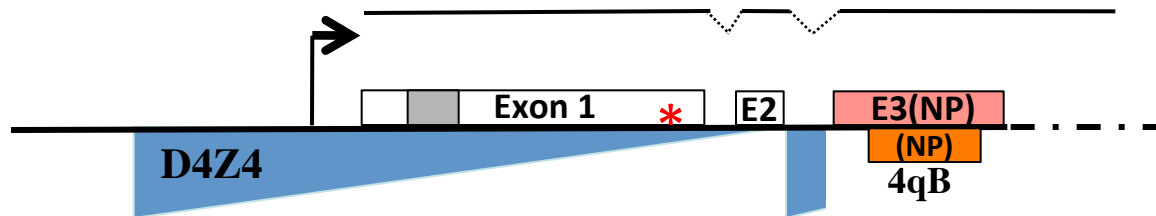
Myogenic cultures from FSHD and unaffected first-degree relatives have similar patterns of gene expression during proliferation and myogenic differentiation



“A Unifying Model for FSHD” based on DUX4

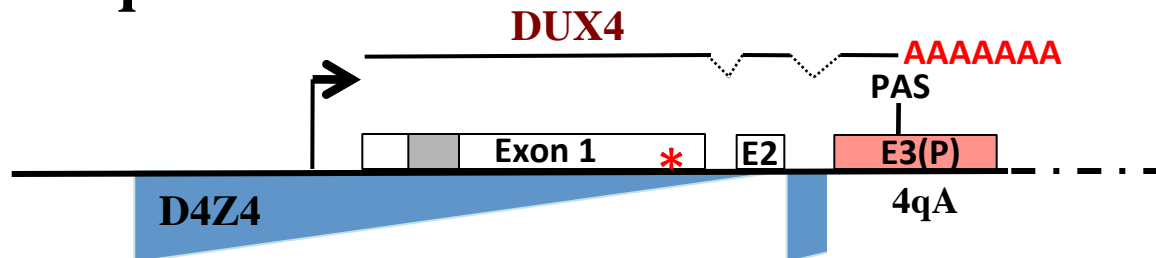
FSHD permissive 4qA subtelomeres encode a third exon containing a polyadenylation site that stabilizes the DUX4 mRNA

Chr 4qB or Chr 10qA or 10qB → Normal



- Unstable mRNA
- DUX4 protein not made
- Nontoxic

Chr 4qA → FSHD



- Stable poly A mRNA
- DUX4 protein produced
- DUX4 expression is exclusive to FSHD

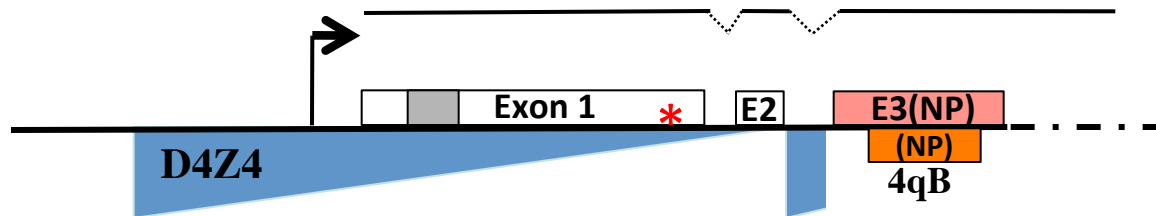
PAS = polyadenylation site
* = translation stop

NP = non-permissive
P = permissive

“A Unifying Model for FSHD” based on DUX4

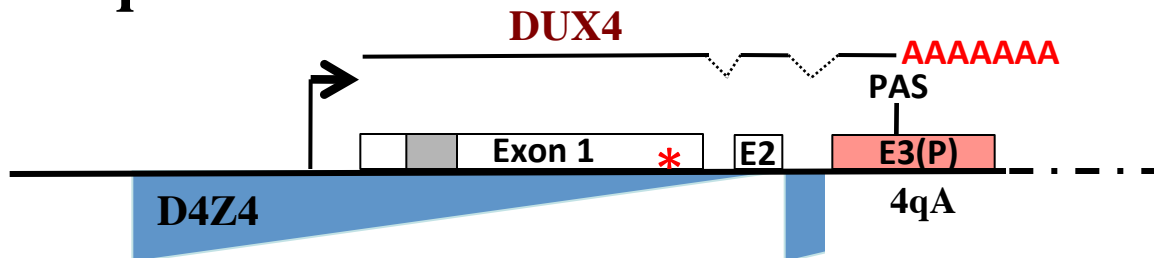
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Chr 4qA → FSHD



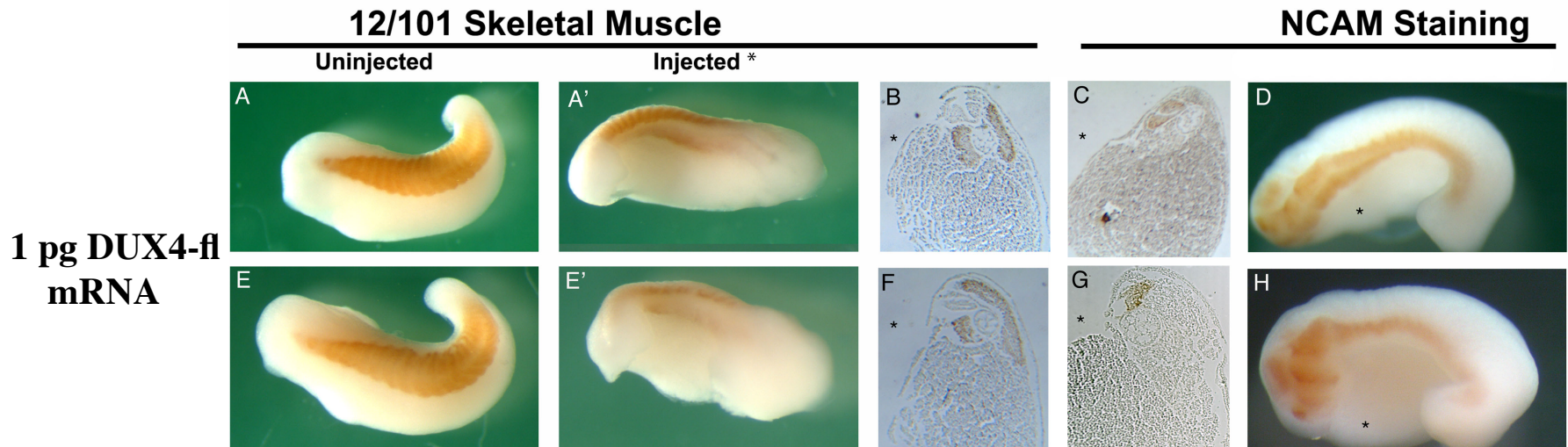
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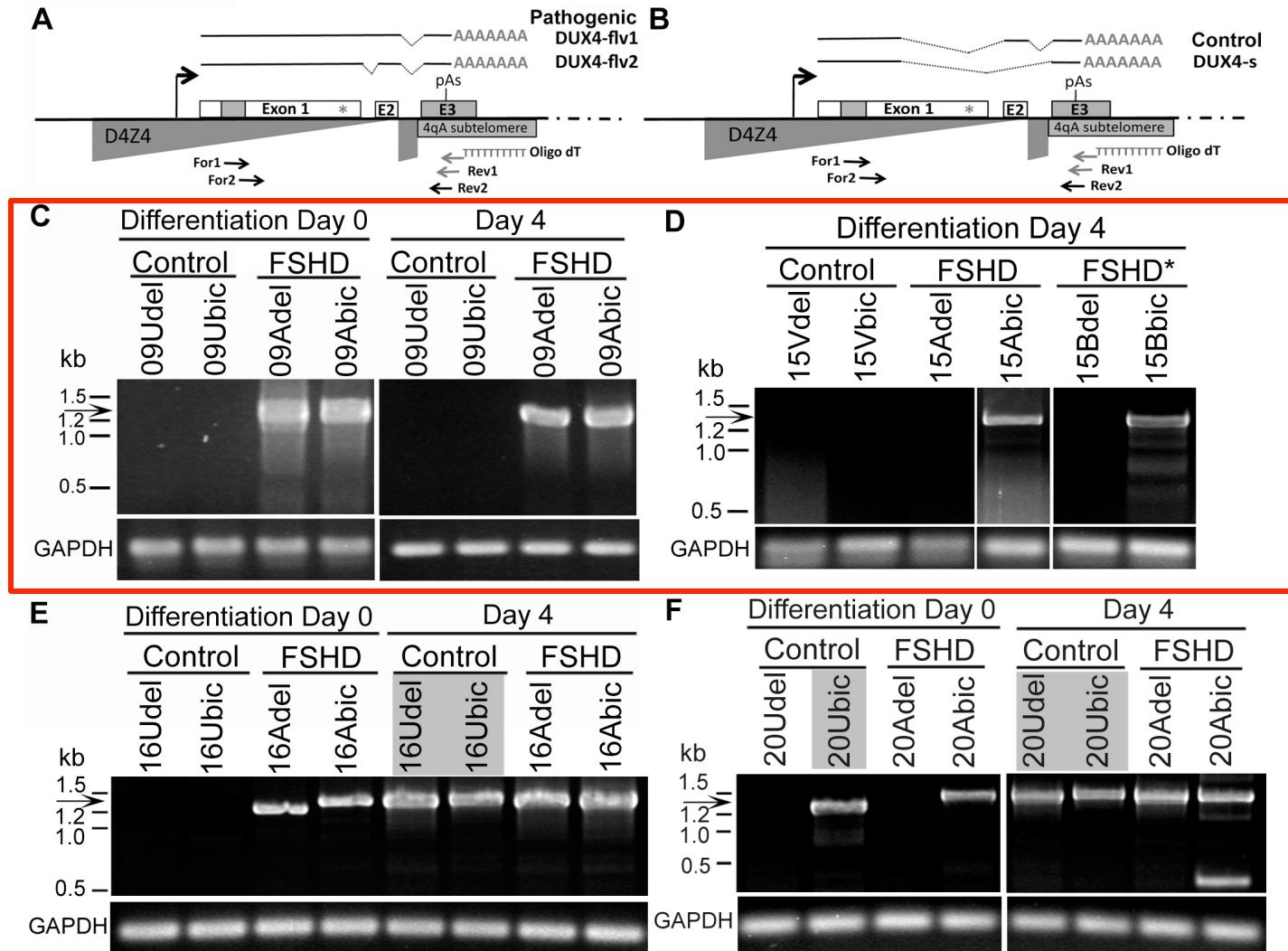
DUX4-FL expression leads to massive loss of developing myogenic cells

Xenopus embryos

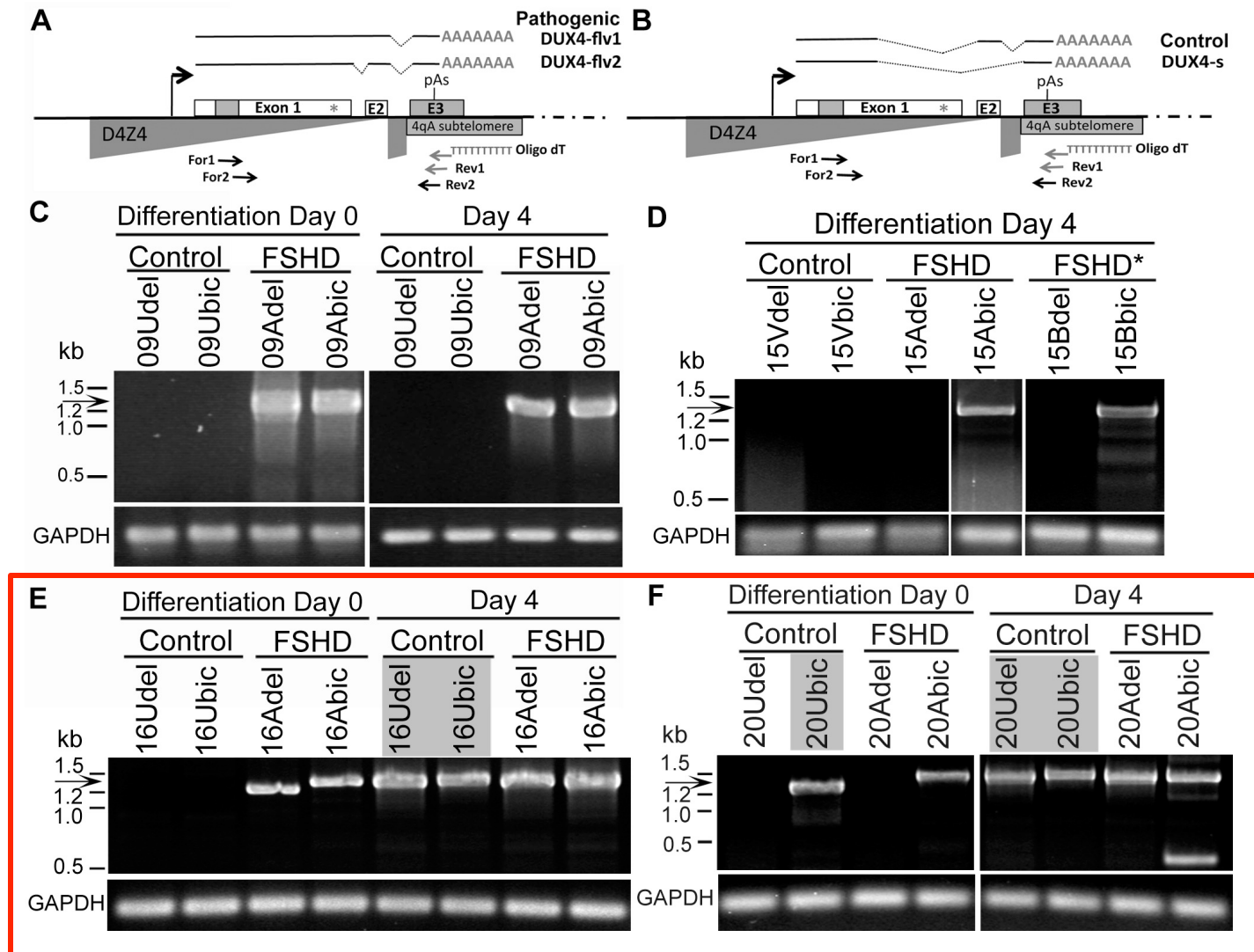


Immunostaining for developing skeletal muscle

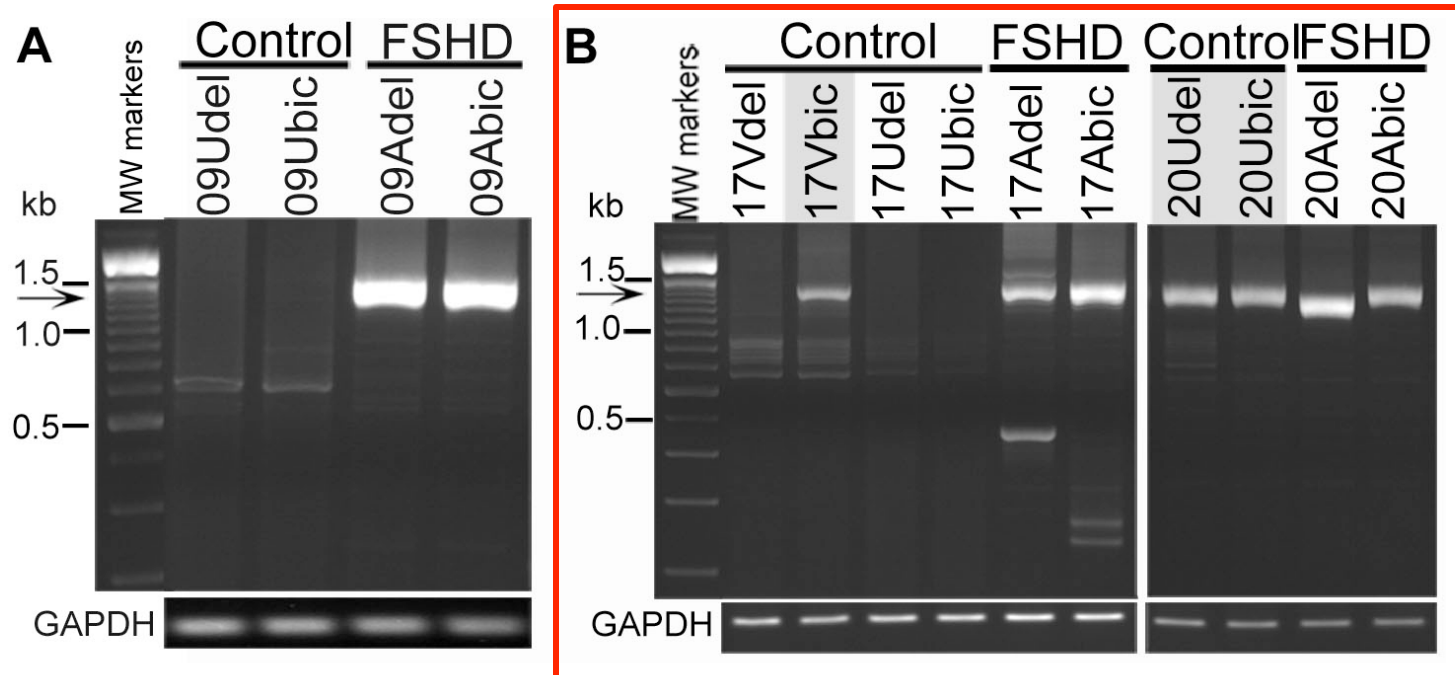
DUX4-fl mRNA is expressed in FSHD1-derived myogenic cells



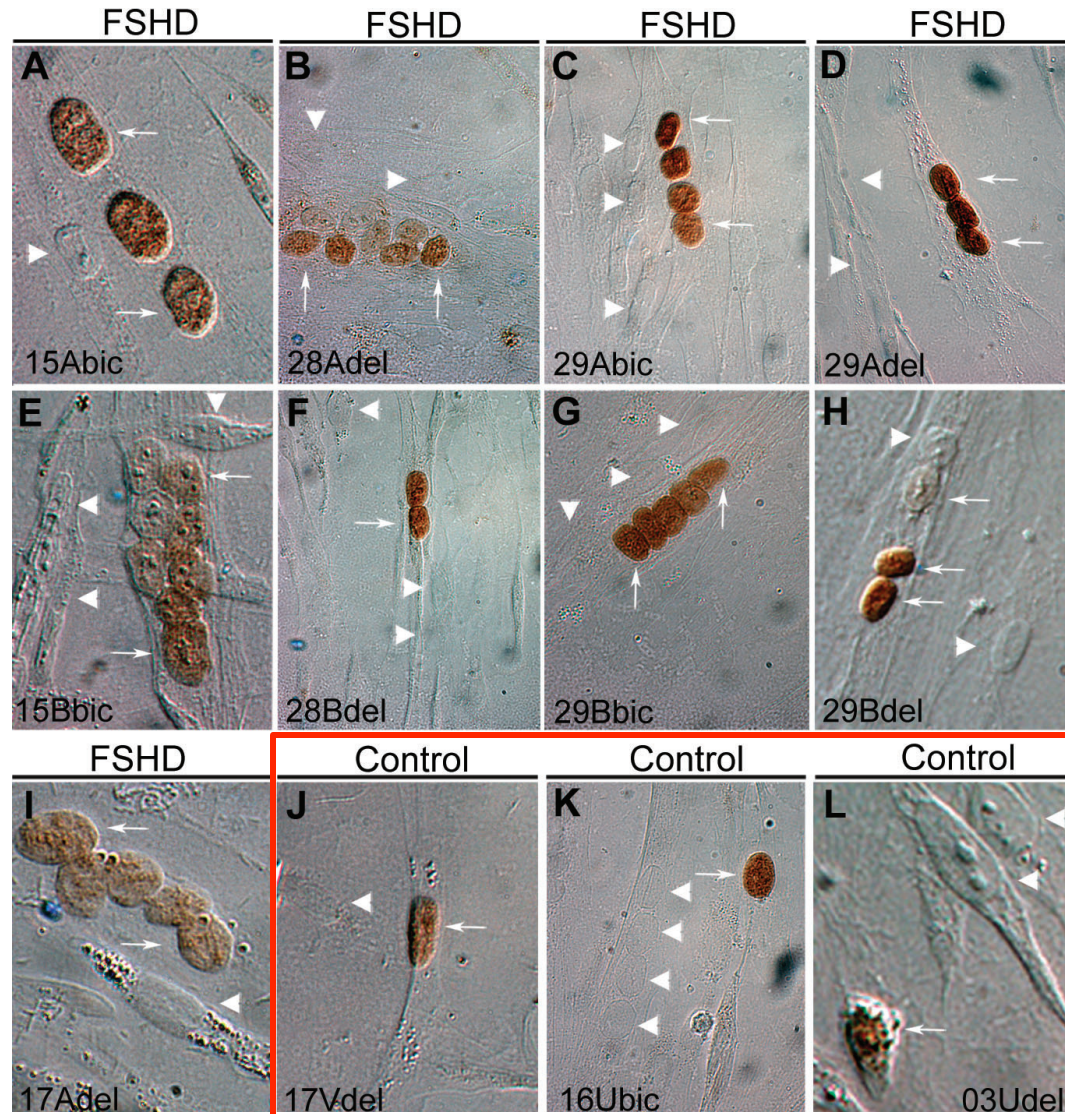
DUX4-fl mRNA expression is not exclusive to FSHD1-derived myogenic cells



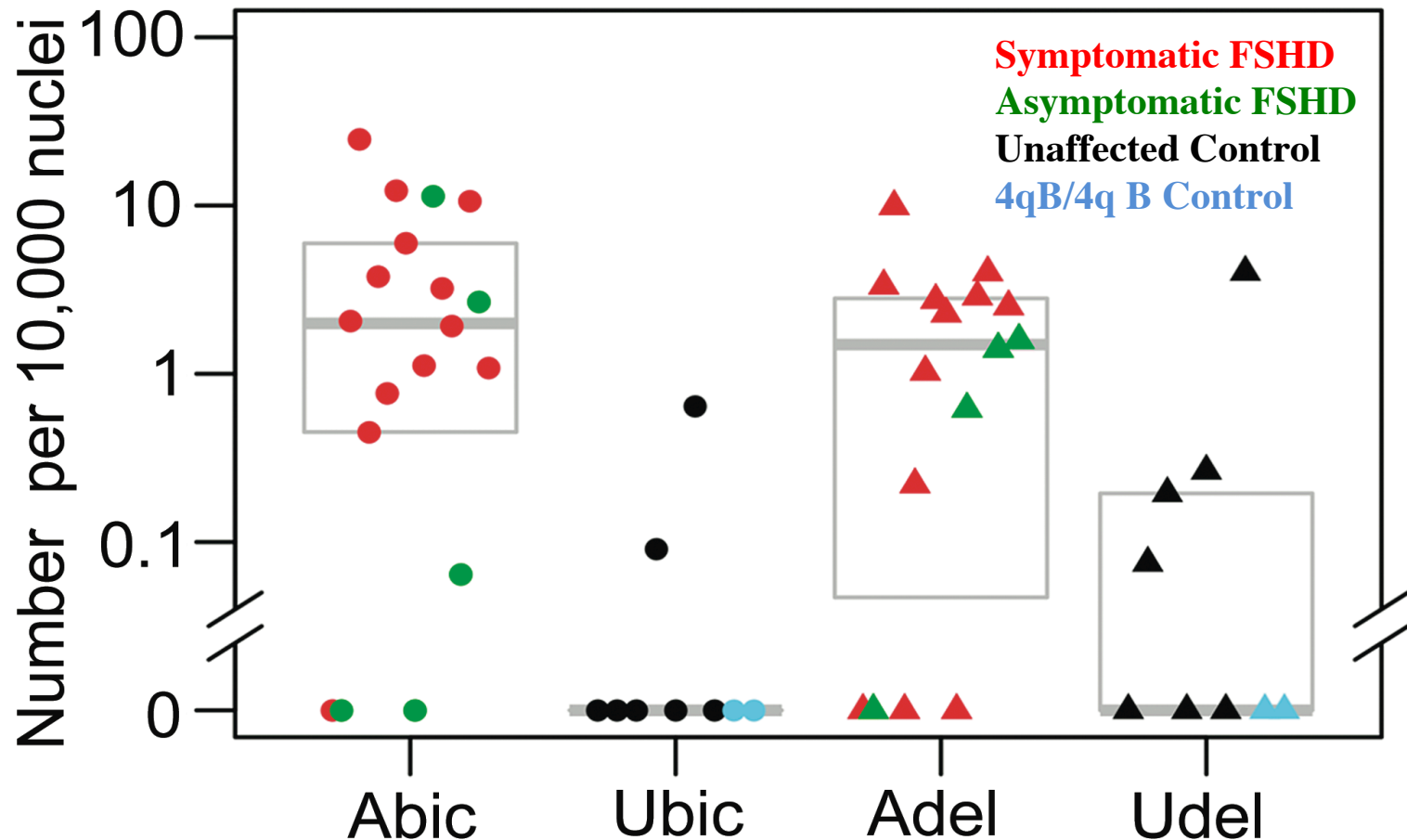
DUX4-fl mRNA expression is expressed in both FSHD1-derived and unaffected muscle biopsies



Differentiated myogenic cells from genetically FSHD1 and control subjects express DUX4-FL protein

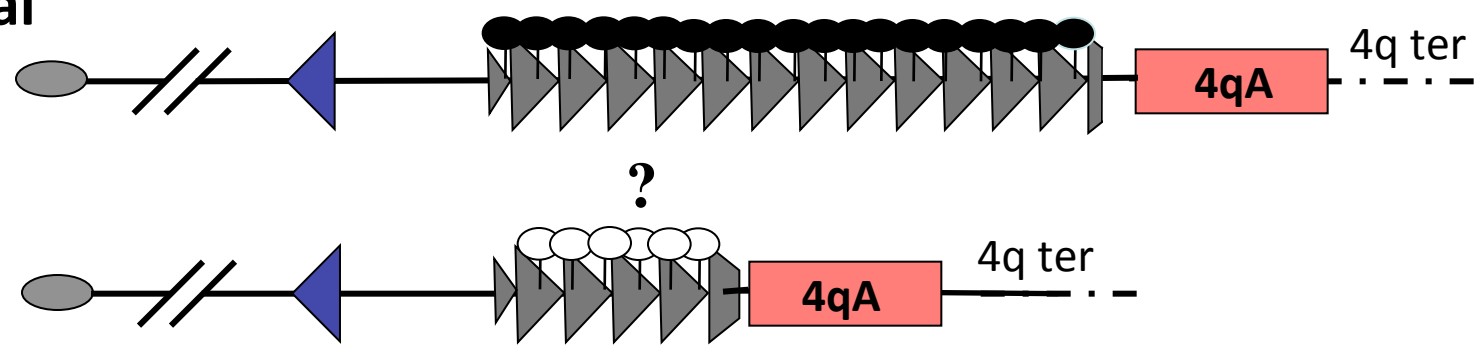


DUX4-FL expression in myogenic cells from FSHD affected unaffected subjects suggests a quantitative model of pathogenesis



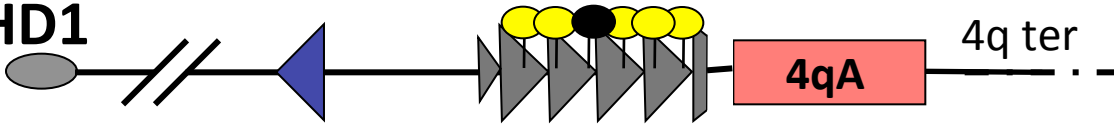
FSHD is linked to the permissive 4qA subtelomere, the epigenetic status of the 4q35 D4Z4 repeat and DUX4-fl expression

Normal

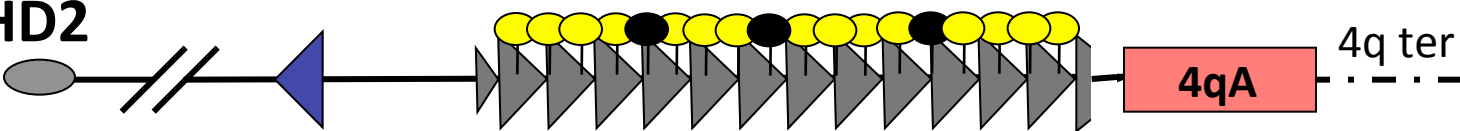


DUX4-fl
-/+
?
++++
++++

FSHD1



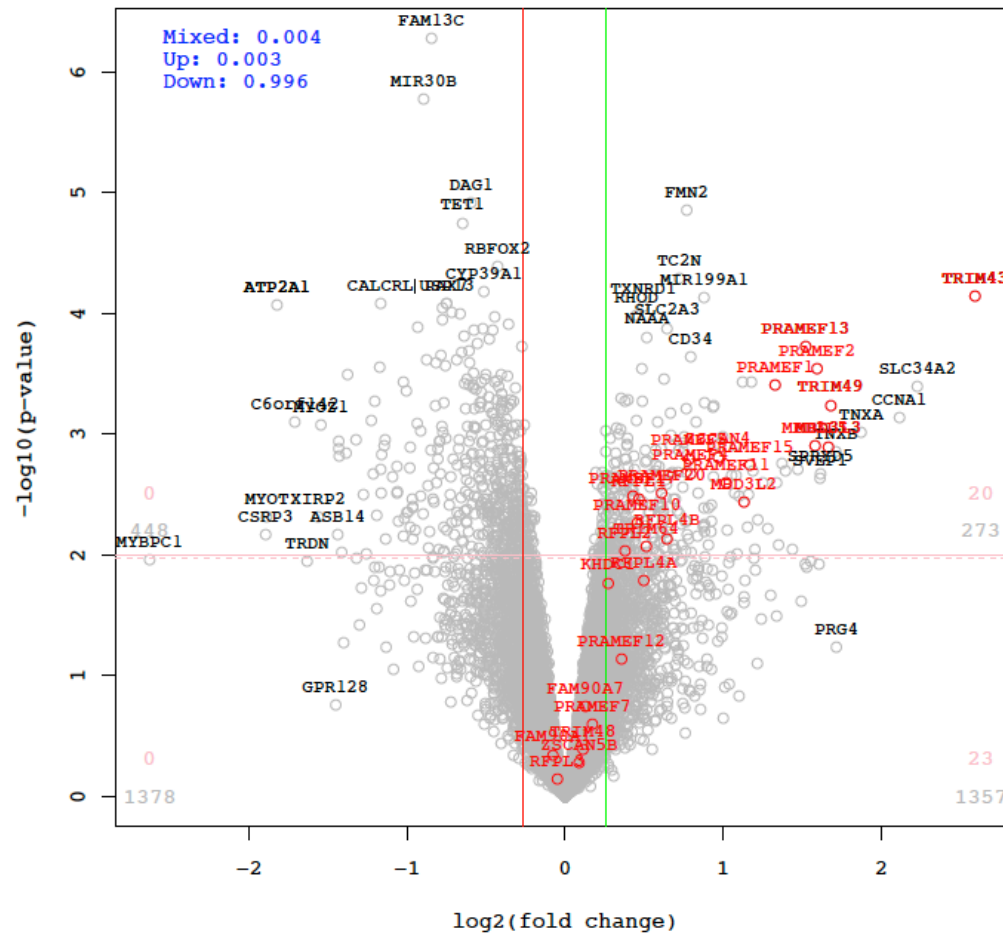
FSHD2



● = Hypermethylated CpGs
more heterochromatic

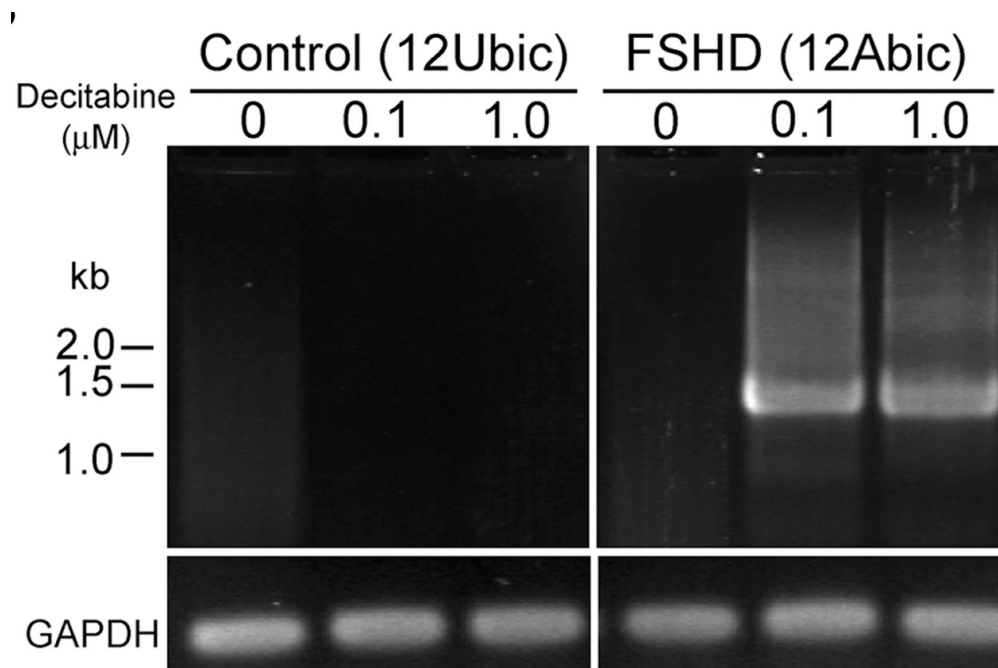
● = Hypomethylated CpGs
more euchromatic

Many DUX4-FL responsive genes are upregulated in FSHD myotubes (biomarkers)



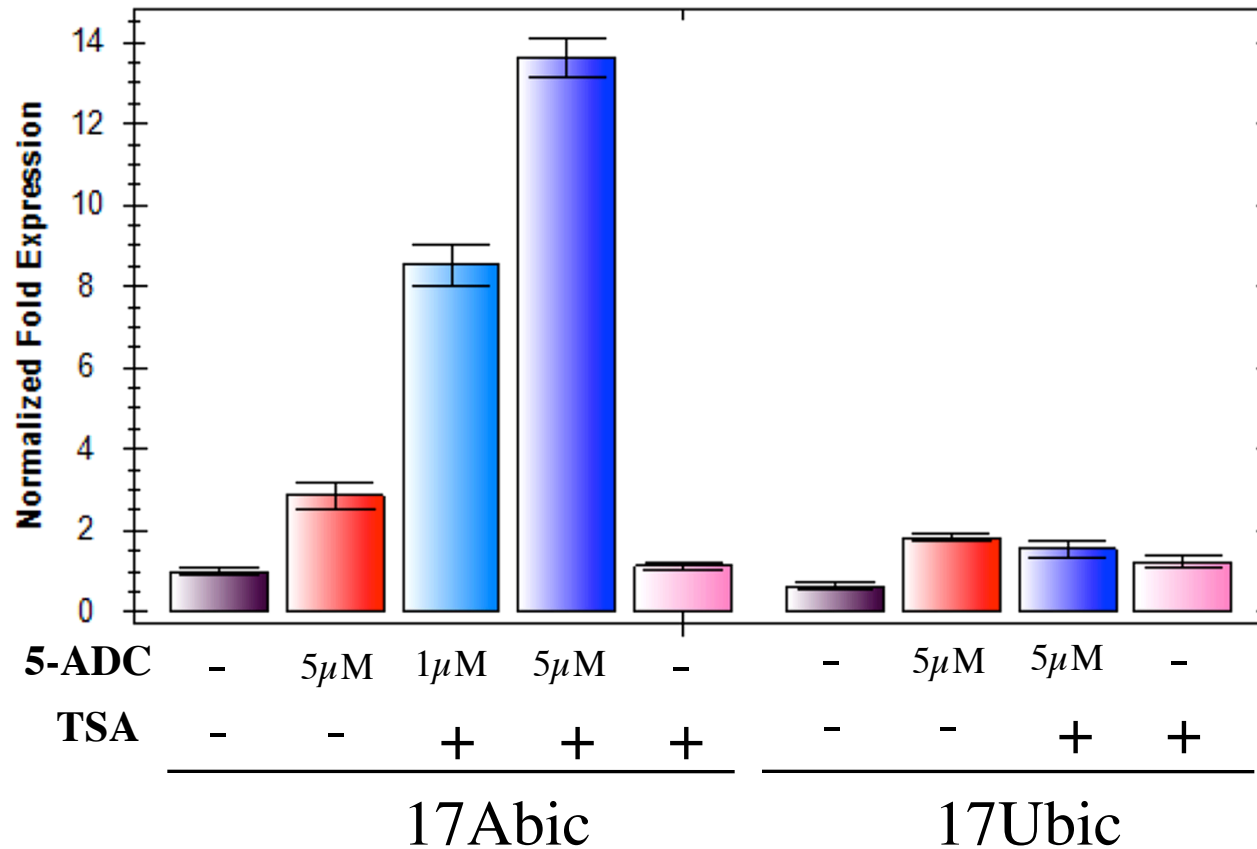
**Increased DUX4-fl expression
appears necessary* but alone is
not sufficient for FSHD**

FSHD-derived myoblasts are epigenetically poised to express DUX4-fl



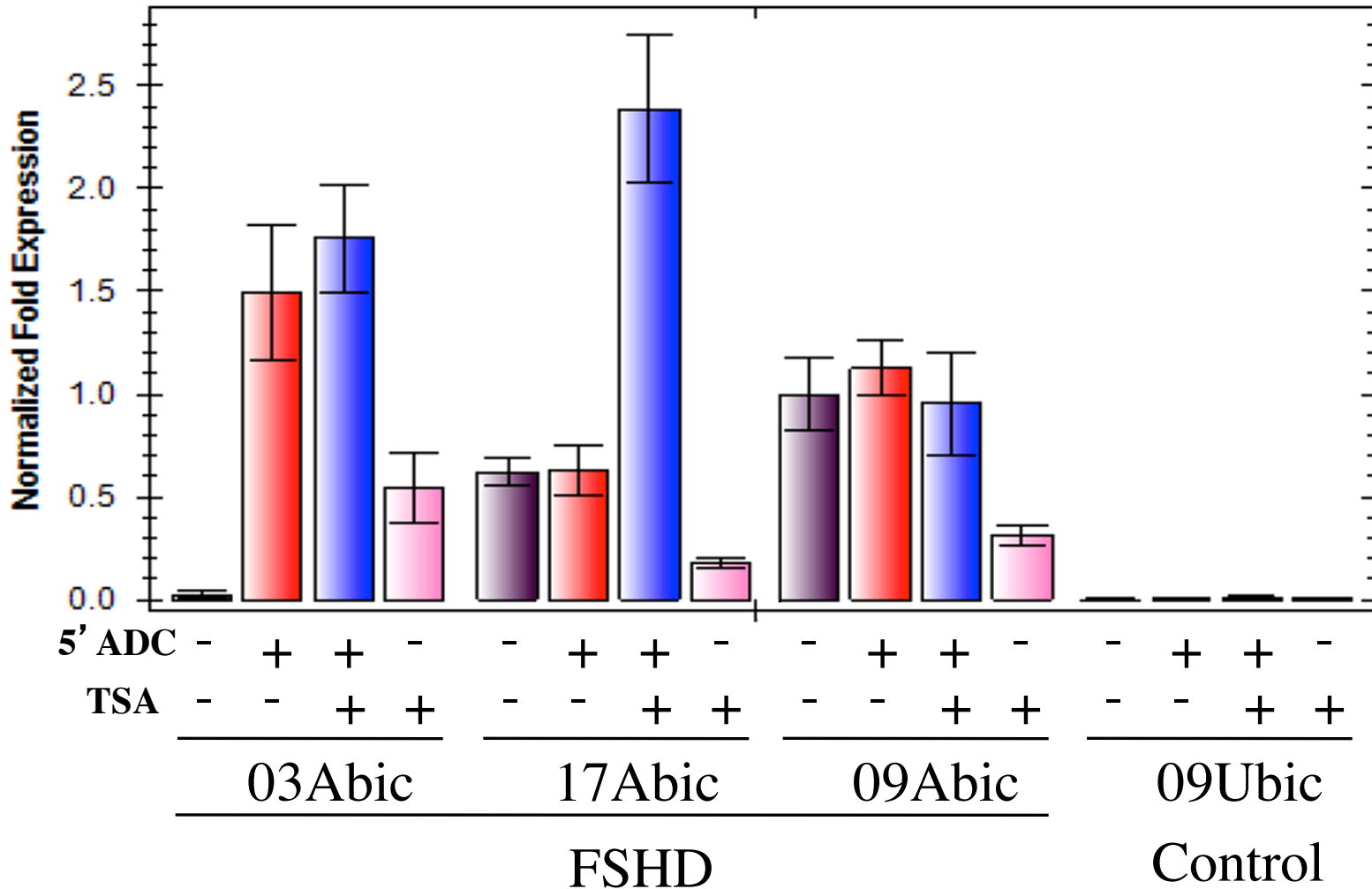
Decitabine treatment leads to DNA demethylation

FSHD derived myogenic cells are epigenetically poised to express DUX4-fl mRNA



DUX4-fl qRT-PCR Analysis

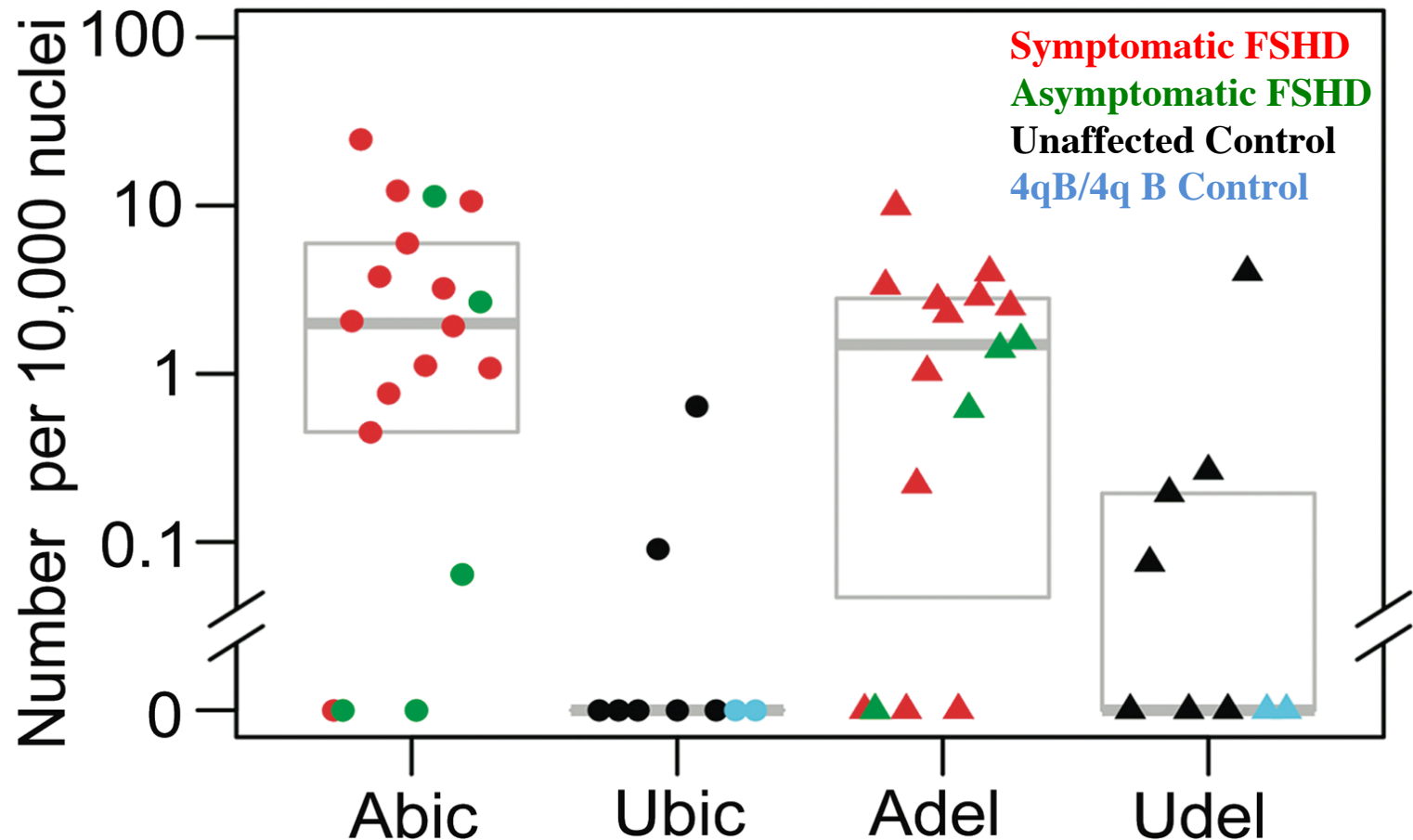
FSHD subjects show individual variability in the stability of DUX4-fl epigenetic repression



The 4q35 D4Z4 in FSHD exists as differentially metastable epialleles among affected subjects
→ epigenetically poised for DUX4 expression

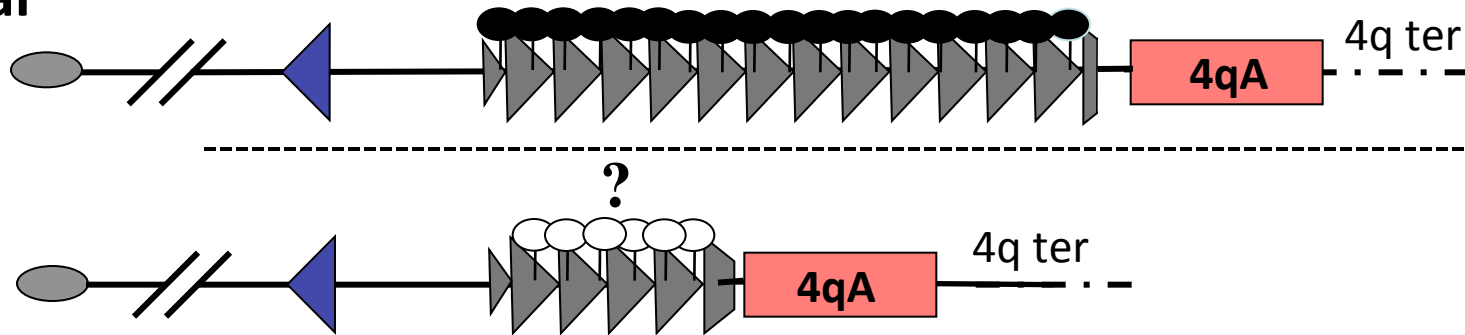
The 4q35 D4Z4 in normal subjects exhibits stable epigenetic repression

DUX4-FL expression in myogenic cells from FSHD1 subjects that show no clinical manifestation of the disease suggests modifiers of disease



DUX4 expression alone is not necessarily causal for FSHD

Normal

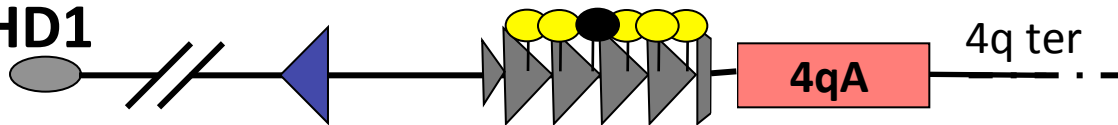


DUX4-fl

-/+

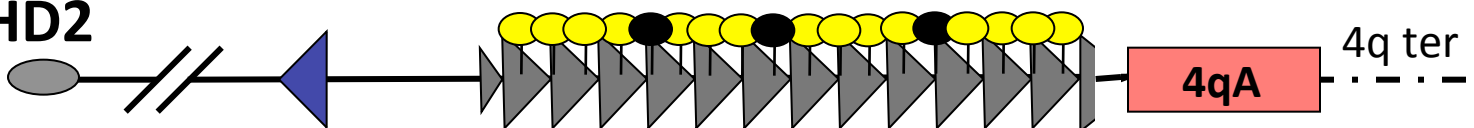
-/+++++

FSHD1



+++++

FSHD2

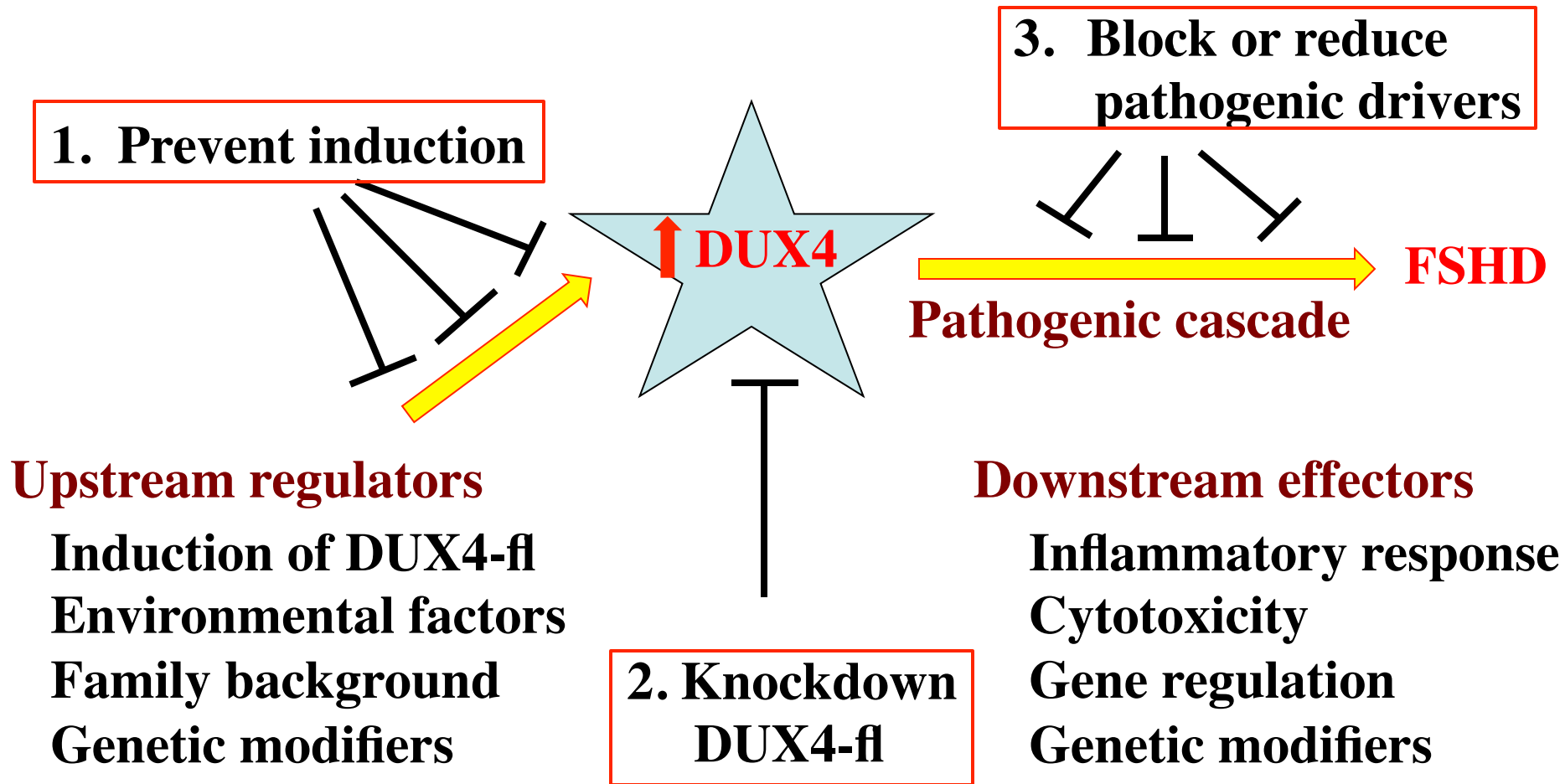


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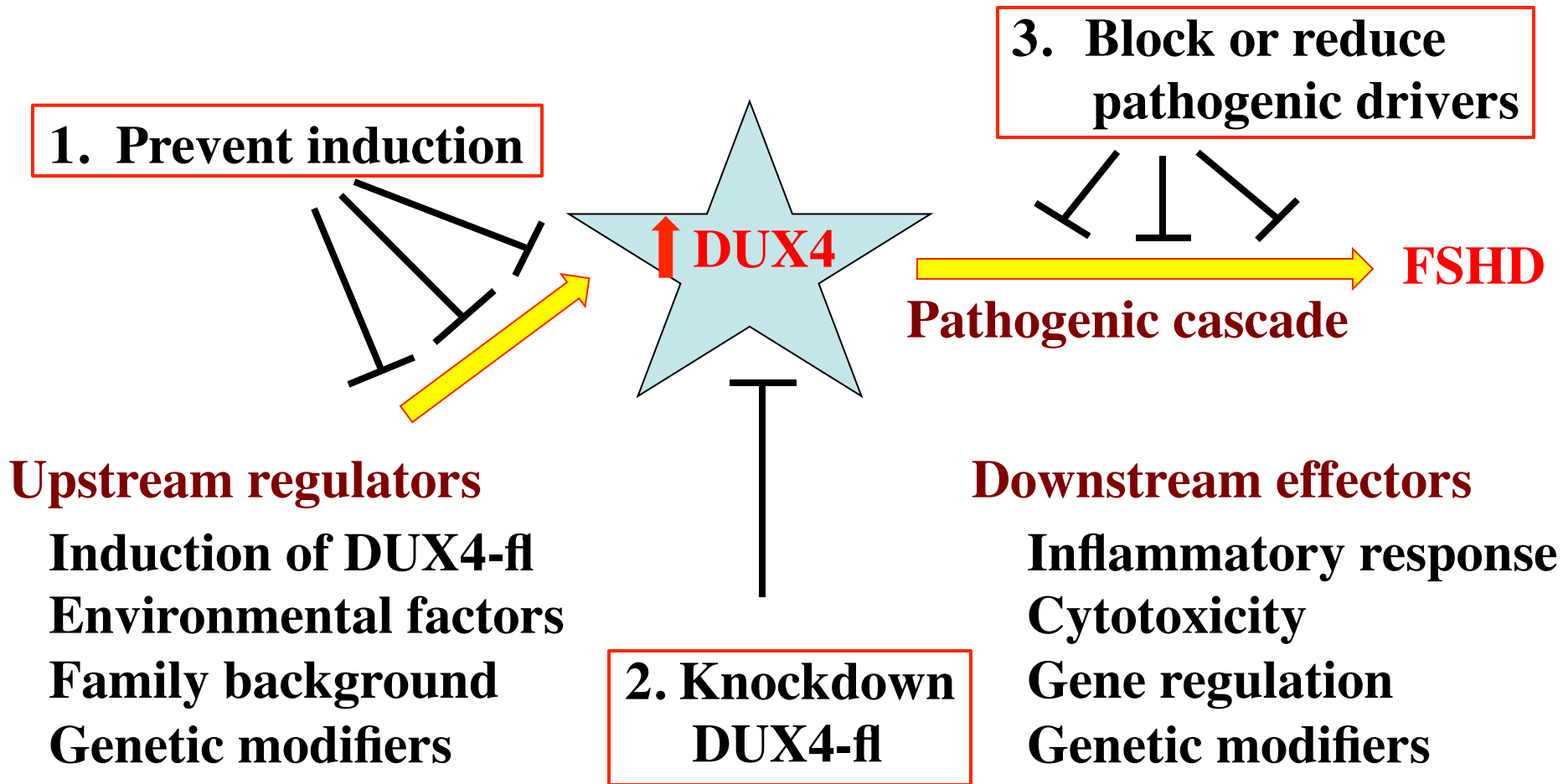
● = Hypermethylated CpGs
more heterochromatic

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more euchromatic

Multiple therapeutic targets for FSHD



Multiple therapeutic targets for FSHD



Key to therapy may lie with identifying the disease modifiers

Analysis of DUX4 mRNA and protein expression in muscles and myogenic cells from FSHD subjects and unaffected relatives

At University of Massachusetts Medical School (and formerly BBRI)

Takako Jones*, Jennifer Chen*, Oliver King, Charles P. Emerson Jr., and Peter L. Jones

At Kennedy Krieger Institute and Johns Hopkins University

Kathryn R. Wagner

At Children's Hospital – Boston and Harvard Medical School

Fedik Rahimov and Louis M. Kunkel

At Boston University School of Medicine (and formerly BBRI)

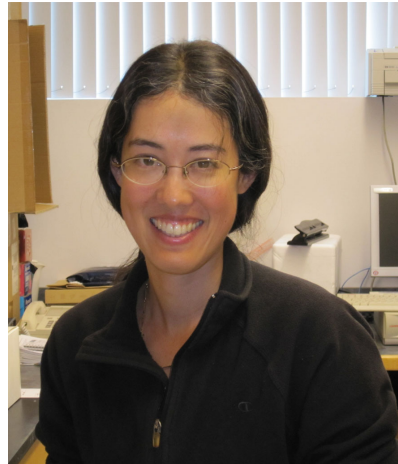
Sachiko Homma, Mary Lou Beerman and Jeffrey Boone Miller



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Charis Himeda



Celine Debarnot

