

Irish Journal of Agricultural and Food Research

Solutions to enteric methane abatement in Ireland

S. Cummins^{1†}, G.J. Lanigan¹, K.G. Richards¹, T.M. Boland², S.F. Kirwan³, P.E. Smith³ and S.M. Waters^{3†}

¹Teagasc, Environment Research Centre, Johnstown Castle, Co. Wexford, Ireland ²School of Agriculture and Food Science, University College Dublin, Belfield, Dublin 4, Ireland ³Teagasc, Animal and Bioscience Research Department, Animal & Grassland Research and Innovation Centre, Grange, Dunsany, Co. Meath, Ireland

Abstract

The efficiency of Ireland's grass-based livestock systems can be attributed to high outputs, low production costs and a low carbon footprint relative to housed systems. Methane (CH.) is a potent greenhouse gas (GHG) of which enteric fermentation from livestock production is a key source, being directly responsible for 57% of Irish agricultural GHG emissions. There are a number of strategies including dietary manipulation and breeding initiatives that have shown promising results as potential mitigation solutions for ruminant livestock production. However, the majority of international research has predominantly been conducted on confined systems. Given the economic viability of Irish livestock systems, it is vital that any mitigation methods are assessed at pasture. Such research cannot be completed without access to suitable equipment for measuring CH, emissions at grazing. This review documents the current knowledge capacity in Ireland (publications and projects) and includes an inventory of equipment currently available to conduct research. A number of strategic research avenues are identified herein that warrant further investigation including breeding initiatives and dietary manipulation. It was notable that enteric CH, research seems to be lacking in Ireland as it constituted 14% of Irish agricultural GHG research publications from 2016 to 2021. A number of key infrastructural deficits were identified including respiration chambers (there are none currently operational in the Republic of Ireland) and an urgent need for more pasture-based GreenFeed™ systems. These deficits will need to be addressed to enable inventory refinement, research progression and the development of effective solutions to enteric CH₄ abatement in Ireland.

Keywords

Abatement • livestock • methane • research • sustainability

Introduction

Although methane (CH₄) has a relatively short atmospheric residence time (half-life of 9.1 yrs), it is a potent greenhouse gas (GHG) with 84 times the global warming potential (GWP) of carbon dioxide (CO₂) over its atmospheric residence time and 28 times that of CO₂ over a 100-yr time horizon (GWP₁₀₀) (IPCC, 2014). Methane is the second-largest contributor to anthropogenic climate change with global emissions estimated at 559 [540–568] Tg CH₄/yr for the decade 2003–2012 (Saunois *et al.*, 2016). The principal anthropogenic sources are fossil fuel extraction and burning, with biogenic contributions associated with eructation from ruminants during fermentation of feed, management of organic wastes and manures and rice paddy cultivation (IPCC, 2014). In 2019, agriculture was responsible for 35.5% of Ireland's GHG emissions, of which enteric CH₄ comprised 57.45% (EPA, 2021).

Agricultural GHG emissions belong to the non-emissions trading sector (non-ETS), meaning that emissions from the sector are not subject to the European Union cap and trade system. This also means that national governments, rather than EU, are directly responsible for emission reductions in these sectors with the extent of each country's reductions established under the 2030 Effort Sharing Regulation (ESR) (PE/3/2018/REV/2). Under the ESR, Ireland is obliged to reduce non-ETS GHG emissions by 30% relative to 2005 over the 2021–2030 commitment period. This will require agriculture to limit emissions to between 17.5 and 19 million tonnes CO_2 equivalent, while also establishing a downward trajectory to reach net zero emissions by 2050. In order to identify pathways for emissions reductions, a marginal abatement cost curve (MACC) analysis was conducted to

[†]Corresponding authors: S. Cummins and S.M. Waters E-mail: saoirse-c@hotmail.com



establish the potential mitigation capacity of the Agriculture, Forestry and Other Land-use (AFOLU) sectors and the associated costs (Lanigan *et al.*, 2018). While other GHG such as nitrous oxide have been well researched regarding point sources and mitigation methods (Harty *et al.*, 2016; Krol *et al.*, 2016), there are many unanswered questions regarding the development of a suitable Irish CH₄ abatement strategy.

There are a number of metrics for the expression of CH, emissions. These include: absolute emissions (total quantity of CH, emissions produced), daily emissions (g CH,/day⁻¹) (Cottle et al., 2015), emissions yield (g CH₄/kg dry matter intake [DMI]) (Y_m) (Hegarty et al., 2007; Løvendahl et al., 2018) and emissions intensity (g CH,/unit output i.e. milk or meat) (Gerber et al., 2011; Haves et al., 2013). Current abatement strategies focus on improving efficiency measures, such as the dairy economic breeding index (EBI), increasing beef live weight gain, extended grazing and the use of sexed semen all of which can significantly reduce CH₄ emissions intensity and associated production costs by increasing overall farm efficiency (Holden and Butler, 2018). However, these measures may not lead to reductions in absolute CH, emissions, as increasing GHG efficiency can result in increased activity (and hence emissions) via so-called "rebound effects" (Paul et al., 2019). Reductions in absolute CH, emissions require reductions in methane output per head and for the overall herd size not to increase. Although there has been progress in achieving this in confinement systems, reducing daily emissions from pastoral systems remains challenging, due to reduced control over animal feed consumption. Researchers in New Zealand are trying to address these challenges and have been doing so for many years (Buddle et al., 2011; Pacheco et al., 2014). In order to achieve carbon neutrality, major mitigation efforts will have to be established and implemented, particularly for CH, emissions resulting from enteric fermentation and offsetting of residual emissions through enhanced carbon sequestration. Thus, there is a requirement in Ireland to conduct research on mitigation strategies to reduce absolute CH, emissions while cattle are at pasture. This will require the development of infrastructure and acquisition of necessary equipment to conduct high-level research.

Therefore, the specific aims of this review are: (1) to identify and critically review CH_4 mitigation strategies that may be suitable for Irish livestock production systems, (2) to review enteric CH_4 publications and projects conducted in Ireland to date, (3) to document the current Irish enteric CH_4 inventories, and (4) to determine the infrastructures required in Ireland to allow for high-level research on decoupling CH_4 emissions from livestock production to progress.

The process of enteric methane production

Enteric CH₄ originates as a by-product of rumen microbial fermentation, the process through which ruminant livestock

digest feed (Tapio et al., 2017). The majority (87%) of enteric CH, is eructated from the forestomach (rumen). A smaller proportion (13%) originates in the hindgut where it is absorbed into the blood and exhaled from the lungs (Hammond et al., 2016). Fermentation is carried out by a complex anaerobic microbial ecosystem, consisting of bacteria, archaea, protozoa and fungi, in the forestomach (rumen) of ruminant livestock (Huws et al., 2018). Individual members of the rumen microbial community play a key role in the fermentation of complex plant structures, and subsequent supply of nutrition, for the ruminant host. Indeed, during enteric (rumen) fermentation, ingested feed is converted into volatile fatty acids (VFA), which provide an estimated 63% of the ruminant animal's energy requirements (Bergman, 1990). In addition to the supply of VFA to the ruminant host, rumen microbial fermentation yields carbon dioxide (CO₂) and hydrogen (H₂) as metabolic endproducts (Newbold & Ramos-Morales, 2020). The production of CH₄, methanogenesis, acts as a H₂ sink in the rumen, which facilitates the progression of further fermentation by preventing an accumulation of excess H₂ which may otherwise hinder electron transport (Morgavi et al., 2010). Methanogens, belonging to the kingdom archaea, are the sole producers of CH, in the rumen with the majority of enteric CH, believed to originate from the hydrogenotrophic methanogenesis pathway (Equation 1), whereby CO₂ is reduced by H₂ (Janssen & Kris, 2008). Finally, the majority of CH₄ originating from ruminant livestock is expelled, to the atmosphere, via the breath of the animal (Hammond et al., 2016).

$$CO_2 + 4H_2 \rightarrow CH_4 + 2H_2O \tag{1}$$

The relationship between the rumen microbial community composition and CH, output has been investigated in a variety of studies (Kittelmann et al., 2014; Wallace et al., 2015; Kamke et al., 2016; Wallace et al., 2019). As highlighted in many reviews (Morgavi et al., 2010; Tapio et al., 2017; Waters et al., 2020), individual members of the rumen microbiome, rather than the overall size of any one microbial kingdom, have been linked to the methanogenic potential of an animal. As a result, practices which are capable of reducing the abundance of microbes that produce substrates for methanogenesis, or alter the methanogen community in favour of a reduced CH, output, offer promise as successful CH, mitigation strategies. Dietary regimes and animal selection are promising CH, mitigation strategies as both dietary management (Carberry et al., 2014; Henderson et al., 2015; Lyons et al., 2017; Liu et al., 2019; Smith et al., 2020a) and host genetics (Weimer et al., 2010; Henderson et al., 2015; Roehe et al., 2016; Li et al., 2019) have been demonstrated to alter rumen microbial composition.

Methods for measuring methane

The main methods to measure CH_4 emissions from ruminants include: (1) respiration chambers (RCs); (2) portable

Method	Advantages	Disadvantages	Approximate cost
Respiration chambers	Measures all CH ₄ emissions from	Does not represent field or grazing	€50,000–€60,000 per chamber including
	the animal	patterns	individual air-conditioning, gas meters and
		Artificial environment for the animal	associated devices
SF ₆	Measures CH ₄ at pasture	Large variability and difficult to	~ €10,000 for 20 sets. Equipment include
		maintain at pasture	individual measurement sets including
			canisters, absorption pipes (excluding gas
			chromatography techniques and pump
			required for analyses)
GreenFeeds	Measures CH_4 at both indoor	Potential bias due to animal	€65,000 per unit for indoor systems (up
	and at pasture	behaviour	to €125,000 in total for additional costs of
	High accuracy of measurements	Short-term measurements require	adding trailers for pasture-based systems)
	Easy to maintain relative to ${\sf SF}_{_6}$	long-term trials	
Portable accumulation	High levels of correlation with RC	Artificial environment for the animal	~ €80,000 per chamber including
chambers (sheep)	Quick measurements cause less	which can cause stress.	individual air-conditioning, gas meters and
	stress on the animal	Unsuited for longer and/or	associated devices
	Easy to operate	repeated daily measurements	

Table 1: Summary of methane measurement equipment and associated cost, advantages and disadvantages

accumulation chambers (PACs) (for smaller livestock); (3) GreenFeed[™] (GF) systems; and (4) the sulphur hexafluoride tracer technique (SF_e). Respiration chambers have been used to estimate energy losses and CH₄ emissions from livestock for over a century (Hammond et al., 2016). The underlying principles of the RC involves keeping an animal in a pressurised chamber, with the enteric emissions of the animal estimated as the difference in the concentration of gases entering and leaving the chamber, with the fluctuation in gas concentrations assumed to be those emitted from the animal (eructated, exhaled and flatulence) (Storm et al., 2012). The experimental time period may vary but is typically ~96 h (Muñoz et al., 2012). Emissions are determined as the difference in CH, gas concentrations entering and leaving the chamber with and corrected for temperature, humidity and pressure. RCs are considered the "gold standard" for recording CH, measurements due to high levels of repeatability, robustness and precision (Gardiner et al., 2015; Patra, 2016). However, animals are required to be contained within a standardised chamber for the experimental period. Therefore, the method is unsuited to estimating emissions from animals under grazing conditions. One of the main disadvantages to RC is the changes in the behaviour of contained animals and DMI can drop, meaning in-chamber CH₄ measurements may not reflect actual animal CH, output (Table 1). Portable accumulation chambers act as airtight chambers that measure CH₄ and CO₂ emissions from small animals such as sheep (Image 1). This method is considered a rapid (1 h), straightforward and highly



Image 1. A sheep in a portable accumulation chamber, Teagasc, Athenry, Co. Galway.

effective technique as results show high comparability with RC results (Goopy et al., 2016; Jonker et al., 2018). However, PAC cannot be deployed for longer periods as increased CO, concentration can negatively influence measurements, thus PAC only allows for measurements over a single time point (Hammond et al., 2016). Breath sampling during feeding can be analysed through GreenFeed™ (GF) systems. They operate as open-circuit head-chambers baited with feed pellets to attract the animal. Airflow inside the feed troughs is collected via an extractor fan, with the analyses of CH, and CO, fluxes determined with the use of infra-red sensors, allowing for the calculation of CH, concentration (Hristov et al., 2015; Huhtanen et al., 2015; Jonker et al., 2016). GreenFeed™ units can also be fitted with sensors to measure H₂, O₂ and H₂S. The accuracy of CH, measurements, obtained with the use of the GF system, can potentially be compromised, if visitation to the unit is not reflective of the diurnal pattern of enteric emissions (Hristov et al., 2018). As a result, it is imperative animals are permitted access to the unit at even intervals throughout a 24-h period over the duration of the measurement period. Feeding of bait to attract the animal to the GF may also cause issues, for example, if investigating forage diets, feeding concentrate feed may impact on the results. Similarly, if the cattle are on an ad libitum concentrate diet, they may not visit the GF unit and training the animals may be difficult (Table 1). GreenFeed™ systems are not restricted to housed environments and have been successfully used to measure CH, emissions on intensive pasture-based dairy systems (Waghorn et al., 2016). Although GFs have been used primarily on cattle to date, there has been development of systems for use with sheep and calves (Nguyen et al., 2018; Meale et al., 2021). Another breath sampling method termed "the sniffer technique", where a gas sampling inlet is placed within the feed manger of a robotic milking unit, measures CH, emissions from the animal at milking events. From these measurements, the daily emission rates can be determined. However, as the sniffer technology has primarily been developed for assessing the methanogenic output of dairy cows at milking (Garnsworthy et al., 2012), its use within a pasture setting, to estimate emissions from nonlactating ruminants, is limited.

The SF₆ technique involves placing a small permeation tube containing SF₆ inside the rumen, and collection tubes with sample lines are used to collect breath samples from the animal. This allows the CH₄ emission rate to be calculated from the known SF₆ emission rate and the measured SF₆ and CH₄ concentrations (Johnson *et al.*, 1994). The use of SF₆ is a labour-intensive method for measuring CH₄ as it involves daily gas canister collections, some loss of canisters from animals and animal handling issues (Table 1). In a comparative study by Deighton *et al.* (2013), it was reported that prolonged deployment of SF₆ tubes can result in an overestimation of CH₄ emissions from animals if the declining release rate of SF₆ from

permeation tubes over time is not accounted for. Additionally, Hristov *et al.* (2016) found higher variability of results generated by the SF_a technique compared to the GF method.

The comparative study by Garnsworthy *et al.* (2019) found that all methods are highly correlated with RC, but levels of correlation between non-RC methods are lower. Although Jonker *et al.* (2016) found no difference between average emission yields from the SF₆ technique, the GF system or from those of RC, there are benefits and drawbacks associated with all methods. Additionally, correcting CH₄ emissions for either intake or output can provide a more contextualised comparison between individual animals than daily emissions alone (outlined in section 1). In order for such calculations to be made, individual feeding units for use in housed systems and during experiments are vital to determine the total feed intake and DMI associated with CH₄ output.

Mitigation strategies – farm efficiency

O'Brien & Shalloo (2016) recommended a number of on-farm efficiency measures that can aid in reducing CH_4 emissions from Irish livestock production systems such as extending the length of the grazing season, increasing the daily live weight gain of beef cattle and lambs, optimising the age and rate of calving and lambing, and reducing the age at slaughter. Such strategies are complementary and when used in conjunction can provide effective CH_4 emissions savings. Albeit, these measures only confer a benefit on a short-term basis and at fixed livestock numbers as increasing the stocking rate will increase overall emissions.

Livestock breeding initiatives are an effective farm management practice that can aid in CH, mitigation in a number of ways including the following: (1) breeding more productive animals can reduce emission intensity (less CH₄ emissions per unit milk/meat produced), for example, increasing dairy cow genetic merit via the EBI and therefore milk production; (2) breeding for increased health, fertility and productivity, that is, faster-growing animals reduces the age at slaughter and thus cumulative CH, emissions over the animal's lifespan; and (3) selecting for more CH4-efficient traits in animals, which will in turn reduce overall CH₄ emissions. Regarding the latter, there are a number of studies, particularly in New Zealand, which highlight the association between the rumen microbiome, genotype and phenotype of sheep bred for low CH, output (Xiang et al., 2016; Jonker et al., 2019a; Rowe et al., 2020). Goopy et al. (2014) found that certain physical traits are associated with sheep selectively bred for low CH, output, with such animals having smaller rumens and a shorter rumen retention time. In the study by Smith et al. (2021), cattle ranked low for residual CH, output (difference between animals predicted, and actual level of CH₄ output, based on DMI and body weight) had an ~30% reduction in daily CH₄ emissions and emissions intensity (g/kg of carcass weight) in comparison to their high residual CH_4 counterparts. In addition, the previous authors noted feed intake and animal performance were not compromised by residual CH_4 output ranking. Although it can be slow to select for such low-emitting animals, nevertheless, such breeding strategies offer a long-term, highly effective solution to CH_4 abatement (González-Recio *et al.*, 2020).

Mitigation strategies – dietary manipulation

Dietary manipulation involving additives and feed/sward type are of fundamental importance in CH_4 mitigation strategies as emissions are highly correlated to animal feed intake and digestibility. Thus, dietary manipulation (i.e. changing the amount or proportion of carbohydrate, protein and roughage in animal diets) has been previously explored by numerous studies and remains an important avenue of further investigation.

Feed additives

Historically, halogenic compounds such as bromoform and chloroform have been studied for their efficacy as antimethanogenic compounds (Bauchop, 1967; Russel & Martin, 1984). At present, such compounds are mainly used as experimental controls (Martinez-Fernandez *et al.*, 2018) due to their strong anti-methanogenic albeit toxic and carcinogenic qualities. This has also been the case for ionophores such as monesin (an antibiotic) which deplete the rumen microbiome and therefore methanogenesis. Antibiotics were previously fed to animals as growth promotors, particularly in the USA. However, since 2006, the use of ionophores is prohibited within the EU due to issues with resistance and human health concerns (EC, 2005).

Hristov et al. (2013) list the efficacy of various livestock dietary additives for reducing CH₄ emissions. Their study highlights the efficacy of including seaweed in ruminant diets to substantially reduce CH₄ emissions. It has been found that certain seaweeds, particularly red and brown species such as Asparagopsis taxiformis (red) and Sargassum flavicans (brown), contain compounds with anti-methanogenic properties as outlined in Abbott et al. (2020). Bromoform (a haloform mostly found in red seaweeds) is known to inhibit the enzymes involved in methanogenesis. Various other compounds found in seaweeds such as lipids, peptides and phlorotannins also play a role in reducing CH, emissions, although the modes of action are less understood (Machado et al., 2016; Abbott et al., 2020). Many studies have seen significant reductions in CH, emissions from livestock receiving seaweed-based additives at various administration rates and concentrations (Machado et al., 2016; Li et al., 2018; Roque et al., 2019; Kinley et al., 2020; Roque et al., 2021). However, there are concerns surrounding the potential for compounds such as iodine (toxic at high levels) and bromoform (carcinogen) to carry through the food chain and

adversely affect human health (Antaya *et al.*, 2019; Abbott *et al.*, 2020). Additionally, importing tropical seaweed species such as *A. taxiformis* risks negatively impacting bioactive compounds during transit. There are also monetary costs and risk of "pollution swapping" through increased GHG emissions associated with transport. The supply of native seaweeds (such as temperate brown species) could present issues, for example, harvesting wild crops or commercially growing seaweeds could have negative environmental impacts. To date, seaweed has not been tested in Irish pasture-based production systems as an anti-methanogenic additive. Overall, further work will be required to understand the potential long-term effects of seaweed additives on animal productivity and human health.

The addition of fats and oils as CH₄ abatement compounds to ruminant diets has shown promising results. Lipids can reduce CH, emissions, as they are toxic to and therefore reduce methanogens and protozoan numbers within the rumen (Beauchemin et al., 2009; Broucek, 2018). Overall, results from lipid addition have been variable, but up to 20% reduction in emissions have been reported (Beauchemin et al., 2020). However, fat addition can negatively affect animal feed intake, carbohydrate digestion in the rumen and overall milk quality. As regards plant-based oil seeds, Kliem et al. (2019) found only linseed-based supplements reduced CH, emissions (across production, yield and emissions intensity) when comparing the administration of linseed, palm and rapeseed oil products to dairy cows. Similarly, Boland et al. (2020) reported an 18% decrease in emissions intensity (g CH₄/kg milk) from pasture-fed dairy cows receiving linseed oil-based concentrates compared with cows receiving stearic acid or soy oil-based concentrates.

There are a range of industrially formulated products with the potential to reduce methanogenesis such as Mootral (a feed additive containing allicin from garlic and citrus extracts) and Agolin Ruminant (an essential oil blend). Studies have shown positive, albeit variable, effects of both products on reducing the amount and rate of enteric CH_4 production in both dairy and beef cattle (Castro-Montoya *et al.*, 2015; Hargreaves *et al.*, 2019; Belanche *et al.*, 2020). Agolin Ruminant is an affordable solution that has also been shown to improve livestock productivity (particularly dairy), which can reduce CH_4 emission intensity. However, both of these additives have primarily been tested as part of total mixed ration (TMR) diets and are in need of testing at pasture level.

The synthetic, non-toxic, organic compound 3-nitrooxypropanol (3-NOP) has proven to be an effective feed additive for the reduction of enteric CH_4 emissions (up to 30% reductions), without compromising animal performance. 3-NOP inhibits methane formation by binding with the enzyme methyl-coenzyme M reductase (MCR) (the catalyst for methane formation) during the final stages of methanogenesis (Meale *et al.*, 2021). The

efficacy of 3-NOP has been assessed in multiple experimental trials (Romero-Perez et al., 2015; Jayanegara et al., 2018; Martinez-Fernandez et al., 2018; Vyas et al., 2018; McGinn et al., 2019; van Gastelen et al., 2020). The recent study of Meale et al. (2021) found that a daily oral administration of 3-NOP to dairy calves from birth to 3 wks post-weaning (14 wks of age) significantly reduced daily CH, emissions up until at least 1 yr of age. It is likely that this early-life administration imprinted on the rumen microbiome, which resulted in lasting alterations of the methanogenesis process. Similar reductions in CH, emissions from adult cattle receiving 3-NOP have not shown long-term persistence once administration ceased as reported in the study on housed beef cattle by Romero-Perez et al. (2015). Most 3-NOP studies have been carried out on indoor/confined systems; however, beneficial results are harder to achieve at pasture level as additives such as 3-NOP need to be combined within animal feed or administered directly after feeding. Therefore, the development of suitable technologies such as slow-release boluses will be required for pasturebased administration as direct feeding cannot take place outside housed periods (Leahy et al., 2020). Additionally, consumer behaviour will need to be considered before 3-NOP or other synthetic additives are adopted as a mitigation option. that is there may be issues surrounding the consumption of products that arise from animals fed on synthetic compounds (Beauchemin et al., 2020).

Sward type

In Ireland, cattle consume >80% of their DMI requirement from grassland forage (O'Brien et al., 2018) with the majority of improved pastures consisting of perennial ryegrass (Lolium perenne L.) monocultures. In recent years there has been increased interest in the use of multi-species swards (a forage mixture consisting of two or more species from different functional groups) for livestock production. This is due to the associated multi-functional benefits which include increased agronomic productivity, livestock health benefits, drought resilience and environmental benefits (Haughey et al., 2018; Grace et al., 2019; Suter et al., 2021). Legumes including white clover (Trifolium repens L.) are often used in multispecies swards and have been shown to contain high tannin levels (Woodfield et al., 2019). Recently, Ku-Vera et al. (2020) presented the potential of plant secondary metabolites such as tannins to modify the rumen microbiome and its function. Stewart et al. (2019) also found that havs with high tannin content have the potential to reduce enteric CH, emissions from beef cattle. It has been found that tannins can lower CH₄ emissions through a reduction in fibre digestion (which decreases H₂ production), and by inhibiting the growth of methanogens (Naumann et al., 2017; Jafari et al., 2019). However, Patra & Saxena (2010) recommend that the proportion of tannin in the diet does not exceed 50 g/kg DMI to

avoid negative implications on animal performance. It is also worth noting the role of pasture management in CH_4 mitigation with animals grazing on low pre-grazing herbage mass swards having reduced CH_4 emissions intensity due to increased forage digestibility (Wims *et al.*, 2010; Boland *et al.*, 2013).

The benefits of white clover on animal performance could potentially impact CH_4 emissions intensity, that is, increased milk yield/solids (Lee *et al.*, 2004; Egan *et al.*, 2017; Dineen *et al.*, 2018) but with the same/lower level of CH_4 output. White clover also increases the passage rate through the rumen which can impact methanogens (Dewhurst *et al.*, 2003; Smith *et al.*, 2020a). Research has also been conducted on legume species other than white clover; Huyen *et al.* (2016a,b found that Sanfoin (*Onobrychis viciifolia*, a tanniniferous legume forage) inclusion within the diet of dairy cows at 50% silage proportion significantly reduced CH_4 emissions intensity. Aside from emissions intensity, Montoya-Flores *et al.* (2020) found a significant decrease in daily CH_4 emissions from cattle fed high legume (*Leucaena leucocephala*) proportion diets.

Herbaceous species Cichorium intybus and Plantago lanceolata contain high levels of bioactive compounds, in particular condensed tannins (Totty et al., 2013; Peña-Espinoza et al., 2018; Ineichen et al., 2019). A multispecies mixture of sorrel (Rumex acetosa), ox-eye daisy (Leucanthemum vulgare), yarrow (Achillea millefolium), knapweed (Centaurea nigra) and ribwort plantain (Plantago lanceolata) fed as haylage resulted in a 10% reduction in Y_m (SF₆ technique) compared with a perennial ryegrass monoculture (Hammond et al., 2014). Wilson et al. (2020) reported no CH, mitigation effects when lactating dairy cows were fed legume or herb-rich swards, compared to grassbased swards. Jonker et al. (2019b) reported elevated Y_ for cows fed a diverse sward (containing ryegrasses, [both Lolium perenne and Lolium multiflorum], white clover, lucerne, chicory and plantain) compared to a ryegrass (L. perenne and L. multiflorum) and white clover mixture. Similarly, Loza et al. (2021) found that CH, emissions per kg energy corrected milk produced (emissions intensity) to be 11% higher from dairy cows grazing diverse mixtures compared to grass-clover swards. The aforementioned studies present some contrasting results. However, verifying the effect of multi-species swards and different types of species at grazing will be important to determining solutions to CH₄ abatement in Ireland.

Methodology

We undertook a comprehensive literature survey of recent (2016–2021) research publications that focus on enteric CH_4 emissions and rumen microbiome function. This was performed through online searches of relevant databases (SCOPUS and Scholar) using specific key words ("rumen"

AND "livestock" OR "methane" OR "microbiome"). The search was limited to research articles from Irish and Northern Irish research-performing organisations (RPOs). In addition, further relevant published scientific articles were identified from cited references in relevant review or metaanalysis papers including Waters et al. (2020). The relevant papers were added to an excel sheet and the methodology sections were analysed to determine the equipment and methodology used. Contact was then made with the corresponding authors of relevant publications to identify current projects and facilities. The proposals of the main Irish CH, research projects were also evaluated to determine the facilities and equipment available for use. Site visits to the relevant research centres took place where CH, projects, research infrastructure and facilities were documented. The current Irish bovine and ovine GHG inventory methodologies were reviewed according to the United Nations Framework Convention on Climate Change (UNFCCC, 2021) Irish National Inventory Reports.

Results

Global warming potential and inventory

The current metric used to assess the impact of individual gases towards climate change is termed the GWP of CH_4 which is "GWP100". This method equates one GHG unit to its CO_2 equivalent, averaged over a 100-yr time horizon. This results in a GWP of 28 in the case of CH_4 (IPCC, 2014). However, this method fails to account for the short-lived atmospheric persistence of CH_4 (9.1 yrs) relative to other GHGs such as N₂O, which persists in the atmosphere for more than 100 yrs and CO_2 , which has a residence time of several centuries. This means that whilst CH_4 has a strong radiative forcing impact when first emitted, this warming impact diminishes as CH_4 oxidises to CO_2 and H_2O . Therefore, there are concerns that the GWP100 overestimates the contribution of CH_4 to long-term radiative forcing (Allen *et al.*, 2016; Allen *et al.*, 2018).

In terms of CH_4 inventory accounting, sheep are currently reported using Tier 1 methodologies in the Irish National Inventory (UNFCCC, 2021). However, there are data available on Irish sheep populations, finishing ages, concentrate usage, housing periods and manure storage systems which could potentially be utilised in the progression to Tier 2 methodologies (O'Brien & Shalloo, 2016). Ireland currently uses Tier 2 inventory estimation for cattle, which disaggregates enteric and manure CH_4 emissions from the bovine herd between numerous categories as described in O'Mara (2006). The principal subdivisions are made between dairy and non-dairy (beef) animals. Dairy is subsequently divided in terms of calving date and region with a separate category for dairy heifers. The non-dairy herd is partitioned in terms of suckler cows (subdivided as per dairy cows) and heifers, with bovines for finishing classified based on age and gender. Future developments for reporting of CH, emissions and incorporating mitigation may require a Tier 3 or modelling approach. Dynamic models based on the mathematical modelling of rumen processes have the potential to incorporate a myriad of feed strategies such as carbohydrate, protein, fat and fibre metabolism. Such models have been simulated in detail and incorporated into US, Dutch and French inventories (Kebreab et al., 2008; Bannink et al., 2011; Eugène et al., 2019). These models are complex and must describe both the dynamics and interactions between various substrate pools and microbial pools and the consequences for the end-products of fermentation. Turnover rates between different pools are generally based on enzyme kinetics and degradation characteristics of different feed types.

As outlined by O'Brien & Shalloo (2016), there is potential to develop Tier 3 inventories for cattle. Data gathered and held by the CSO, Teagasc NFS, Bord Bia and the animal feed industry could potentially be integrated into the development of more accurate, Tier 3 CH_4 emission factors for cattle. There is already an extensive body of work on the variation in forage quality on production (Hart *et al.*, 2009; Muñoz *et al.*, 2016). Therefore, the national data required for inputting into a Tier 3 system are already available.

Irish enteric methane research to date

Over the last 5 yrs (2016-2021), 14% of Irish agricultural GHG research publications have been based on enteric CH, emissions (Figure 1). In total, 43 research papers have been published on studies using methodologies such as RCs, SF, (Images 2 and 3), RUSITEC in vitro rumen simulation and 16S rRNA amplicon sequencing (Table 2). Work carried out in Ireland, which furthers the understanding of the rumen microbiome, include the study by Kumar et al. (2018) and reviews by Huws et al. (2018) and Leahy et al. (2019). A number of studies have investigated strategies for decoupling enteric CH, emissions from livestock production. The studies by Popova et al. (2017), Smith et al. (2020b) and Boland et al. (2020) examine the effects of dietary supplementations, that is, linseed oil and industrial by-products. Other dietary manipulations strategies including sward composition, sward N application, concentrate type and protein content have been assessed (Hynes et al., 2016; McDonnell et al., 2016; Zhao et al., 2016; Günal et al., 2019; O'Connor et al., 2019; Ferris et al., 2020; Smith et al., 2020a). There have been a number of studies that focused on the genetic selection for production efficiency and low CH, output animals. The traits investigated include phenotype, age and breeding index of cattle (Morrison et al., 2017; Rubino et al., 2017; Quinton et al., 2018; Ferris et al., 2020). Additionally, there has also

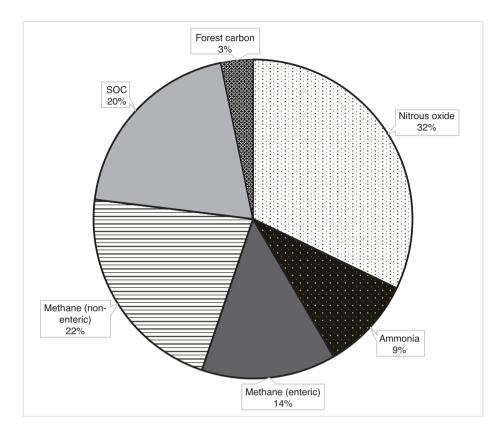


Figure 1. Research publications from Irish and Northern Irish research-performing organisations (2016–2021) that focus on agricultural greenhouse gases (GHGs): nitrous oxide, ammonia (pollutant and indirect GHG), carbon (soil organic carbon (SOC) and forest C) and methane (enteric and non-enteric).



Image 2. SF₆ equipment on grazing sheep, Teagasc, Athenry, Co. Galway.



Image 3. SF $_{\rm 6}$ equipment on grazing dairy cattle, Teagasc, Moorepark, Co. Cork.

been Irish involvement with international enteric CH_4 database and model development (Niu *et al.*, 2018).

Current research projects in Ireland

There are currently a number of projects focusing on CH_4 abatement underway in Ireland (Table 3). Both EU-funded projects, RumenPredict (ERA-GAS) and *MASTER* (Horizon 2020 – European Commission), have members of Teagasc, University College Dublin (UCD) and the Irish Cattle Breeding Federation (ICBF) working in collaboration to better understand the link between the rumen microbiome composition and CH_4 output. There are a number of other ERA-GAS-funded projects including *SeaSolutions*, a Teagasc co-ordinated project which aims to determine the potential of seaweeds to reduce enteric CH_4 emissions from sheep, beef and dairy cattle, METHlab which focuses on the use of lactic acid bacteria as an approach to reduce CH_4 emissions from ruminant livestock and GrassToGas which aims to identify individual animal, feed and

environmental attributes associated with feed and water intake efficiency for pasture-based sheep production systems using portable accumulation chambers (Image 1). The Teagasc-led Department of Agriculture, Food and the Marine (DAFM)funded Meth-Abate project seeks to develop new technologies to reduce enteric CH, emissions from ruminants and emissions from stored manure and slurry. This project is focused on investigating the potential of feed and slurry additives as CH, mitigation solutions. Meth-Abate has research partners from National University of Ireland, Galway (NUIG), Teagasc, Agri-Food and Biosciences Institute (AFBI), Queens University as well as a number of industry stakeholders. Greenbreed (DAFM funded) is a collaborative project between UCD, Teagasc and the ICBF which aims to determine strategies for the breeding of more CH,-efficient animals. There are a number of GF systems in place at the confinement facilities at the ICBF used to carry out this research (Images 4 and 5). VistaMilk is an Science Foundation Ireland (SFI) research centre, with the aim to "facilitate the development and deployment of new knowledge, new technologies and new decision support tools to maximise the efficiency and effectiveness of the entire dairy production chain". VistaMilk funds a number of projects investigating feed additives at pasture in dairy cattle at Teagasc, Moorepark (Image 6). APC microbiome is also an SFI-funded centre which has links with CH, abatement projects such as METHlab. SMARTSWARD (DAFM funded) is a collaborative project between UCD, TUD and AFBI and is investigating the impact of multi-species swards on enteric CH₄ emissions of beef cattle and lactating dairy cows using the SF₆ technique. The current (as of 2021) infrastructure available in Ireland to conduct CH, research is listed in Table 2.

Discussion

Irish livestock systems are primarily grass-based owing to the temperate climate that promotes grass growth (O'Donovan

Table 2: Irish methane research infrastructure as of 2021: a summary of current research projects in Ireland that focus on enteric methane

Research centre	Facilities			
Teagasc Grange	5 × GreenFeed (GF) systems, 4 × Rusitec in vitro simulators, SF ₆ equipment, digestibility crates for measurements			
Irish Cattle Breeding Federation Tully	10 × GF systems (indoor), cattle digestibility units			
Teagasc Moorepark	4 × GF (pasture), SF ₆ equipment			
Teagasc Athenry	Portable accumulation chambers for sheep, SF ₆ equipment for sheep			
UCD	4 × GF (pasture/indoor), SF ₆ equipment, 4 × Rusitec in vitro simulators, digestibility crates for measurements			
	(dairy and sheep)			
AFBI	2 × large respiration chambers, 6 × medium respiration chambers, 3 × GF systems, SF ₆ equipment, dairy cow			
	digestibility units, digestibility crates for measurements			
Queens University Belfast	Rumen microbiology laboratory facilities			

Project title	Coordinator	Funder	Duration	Collaborators
Rumenpredict	Prof Sharon Huws	ERA-GAS	3 yrs	QUB, Teagasc, ICBF, UCD, Natural Resources Institute
				Finland, Agresearch NZ, Swedish University of Agricultural
				Sciences, Wageningen University, INRA, France
MASTER	Prof Paul Cotter	EU Horizon 2020	4 yrs	Teagasc, ICBF
Seasolutions	Dr Maria Hayes	ERA-GAS	3 yrs	Teagasc, IT Sligo, QUB, AFBI, Norwegian Institute
				of Bioeconomy Research, Agriculture and Agri-Food
				Canada, Department of Agricultural Sciences Sweden,
				Friedrich-Loeffler-Institut Germany, SINTEF Norway
Methlab	Prof Catherine	ERA-gas	3 yrs	Teagasc, UCC, Wageningen University, INRA, France,
	Stanton			Agresearch NZ, SACCO Italy
Meth-Abate	Prof Sinead Waters	DAFM/DAERA	4 yrs	Teagasc, NUIG, AFBI, Queens, Industry
Greenbreed	Prof Donagh Berry	DAFM	4 yrs	Teagasc, ICBF, UCD, WIT, CIT
Grasstogas	Joanne Conington	ERA-GAS	3 yrs	Sheep Ireland, Teagasc, INRA France, National
				Agriculture Research Institute Uruguay,
				Norwegian University of Life Sciences, AgResearch NZ,
				International Center for Livestock Research and Training
				Turkey
Greengrowth	Dr Fiona McGovern	Teagasc	4 yrs	Teagasc, UCD
SMARTSWARD	Prof Tommy Boland	DAFM/DAERA	4 yrs	UCD, Technological University of Dublin, AFBI
VistaMilk SFI Research	Prof Donagh Berry	Science Foundation Ireland	Ongoing	Teagasc, UCC, UCD, WIT, multiple national and
Centre		(SFI) and DAFM, national and		international industry collaborators
		international industry funding		
Water-based delivery	Prof Tommy Boland	Enterprise Ireland	2 yrs	UCD
of rumen modifiers				
to enhance the				
sustainability of ruminant				
production systems				

Table 3: A summary of current research projects in Ireland that focus on enteric methane

et al., 2021). There are also lower economic and environmental costs associated with grass-based systems relative to confinement/feedlot systems (O'Brien *et al.*, 2012; O'Brien *et al.*, 2014; Herron *et al.*, 2021). Thus, investigating how different sward compositions impact CH₄ emissions/emissions intensity from grazing livestock is certainly an avenue worth further investigation. Some research has shown that multispecies swards can decrease daily CH₄ emissions/head and/ or directly lower CH₄ emissions intensity through the improved animal performance achieved with grazing multi-species swards (e.g. Hammond *et al.*, 2014). Although, some research does not support these findings, e.g. the study of Loza *et al.*, 2021. Nonetheless, legume-based and multi-species swards require less N fertiliser inputs to maintain yield production and have lower associated N₂O emissions and emissions intensity

than conventional *L. perenne* monocultures (Cummins *et al.*, 2021). Thus, there is the potential that more diverse swards could give dual GHG abatement.

Anti-methanogenic feed additives could potentially have a role in Irish CH_4 abatement strategies. However, issues with social acceptance (Beauchemin *et al.*, 2020), cost and on-farm delivery should be taken into account. Due to the constant turnover of the ruminal contents (van Soest, 2018), the efficacy of additives may depend on their residency time in the rumen/frequency of intake. It is easier to deliver additives as part of a TMR diet in housed systems where additives can be supplemented at feeding. This is harder to achieve at pasture level where grazed forage makes up the majority of the animals' diet. To overcome this, early-life supplementation strategies with additives such as 3-NOP may offer an effective



Image 4. A beef steer visiting a GreenFeed[™] system at the ICBF facilities, Tully, Co. Kildare.



Image 5. Teagasc GreenFeed™ systems at the ICBF facilities, Tully, Co. Kildare.

solution as recently established by Meale *et al.* (2021). The technological development of slow-release boluses could also provide an option for pasture-based delivery as is often the case with mineral supplementations (Grace & Knowles,

2012; Aliarabi *et al.*, 2019). The effects of additives on animal productivity and welfare will need to be evaluated alongside farm-level cost-effectiveness (i.e. life cycle assessments) to determine the most effective and practical mitigation strategy at farm level.

Selecting for low CH, emissions in livestock breeding programmes, that is, including animal CH, output in breeding indexes, is a cumulative, permanent and effective CH, abatement strategy (González-Recio et al., 2020). In addition, some authors have advocated the benefits of selecting more feed-efficient ruminants as a mitigation strategy (Basarab et al., 2013). Residual CH, output is a useful phenotype for use in selection processes, as it is strongly related to daily CH, output but also independent of body weight and feed intake (Bird-Gardiner et al., 2017). The work, currently ongoing with Teagasc and the ICBF, will contribute to the development of breeding values for low CH, output for both beef and dairy sires as presented in Smith et al. (2021). The benefits of selecting for lower CH, output animals would be further complemented with CH₄-inhibiting additives and grazing on multi-species swards. However, breeding indexes will require validation at pasture as there is the possibility of a genetic × feed type effect. Therefore, it is vital that further resources are allocated to continuing and expanding on this research.

A major increase in CH, research capacity and output is required. For example, between 2016 and 2021, only 14% of agricultural GHG research publications in Ireland were focused on enteric CH₄ (Figure 1). Therefore, research outputs seem to be disjointed from research requirements given that enteric CH, emissions from livestock production constitute 57% of Irish agricultural GHG emissions. Thus, considerable investment towards research infrastructure and facilities is urgently required if the Irish CH, research capacity is to expand and progress. Regarding specific infrastructures (Table 2), there are currently no RCs in use in the Republic of Ireland. These will be required as RCs are a fundamentally important methodology for measuring CH, emissions and for validating other CH, measurement methodologies. More individual feeding and feed intake recording systems would be useful to assess further the relationship between genotype, feed use efficiency and CH, emissions from a large cohort of animals for breeding purposes. There is also a need for more accurate methodologies for measuring feed intake from grazing animals and further investment in pasture-based GF systems for large-scale field trials. Additionally, there is a need to investigate and further develop upcoming technologies for measuring CH, emissions (Neethirajan, 2020) for application to grazing systems. Research would ideally take place at a multi-institutional level and capture CH₄ emissions from dairy, beef and sheep production systems (both intensive and extensive) across Ireland.



Image 6. Pasture-based GreenFeed™ system, Teagasc, Moorepark, Co. Cork.

In order to improve and refine the national enteric CH_4 inventory, the working data (i.e. yearly livestock numbers from the CSO) should be combined with existing research output to develop a dynamic CH_4 model for inclusion within the national inventory. This would ideally be performed for sheep, beef and dairy cattle aiming to progress sheep inventory from Tier 1 to Tier 2 (with the aim of further progression to Tier 3) and cattle from Tier 2 to Tier 3. Further emission factor and inventory development will provide increased accuracy in Ireland's GHG emissions estimations and therefore mitigation efforts. An accurate inventory accounting is essential for determining the efficacy of mitigation efforts and meeting carbon neutrality in Ireland by 2050.

Thus in summary, research is required in the following areas: (1) on-pasture deliverance of anti-methanogenic substances in the form of additives, for example, 3-NOP, oilseeds, seaweeds; (2) the effects of sward type on CH_4 emissions and emissions intensity at pasture level including the effects of different species; and (3) breeding efficiency: selecting for low CH_4 -emitting genetics while retaining production and

profitability. The capabilities and limitations of all mitigation options should be considered during the development of a national CH_4 abatement strategy and a holistic approach should be taken rather than a "silver bullet" solution. The meeting of GHG commitments will be dependent on policy drivers and technology adaptation. Knowledge transfer and advisory will also have key roles with the on-farm delivery of abatement strategies.

Acknowledgements

The authors wish to thank the DAFM for funding of the Towards an Agricultural Greenhouse Gas Research & Innovation Centre (AGGRIC) scoping exercise. Thanks to Dr Fiona McGovern, Dr Tianhai Yan, Dr Laurence Shalloo, Dr Ben Lahart and Katie Starsmore for their guidance and supplying of information. Thanks and credit are due to Dr Rachel Power and Dr Noirín McHugh for their supplying of photographic material.

References

- Abbott, D.W., Aasen, I.M., Beauchemin, K.A., Grondahl, F., Gruninger, R., Hayes, M., Huws, S., Kenny, D.A., Krizsan, S.J., Kirwan, S.F., Lind, V., Meyer, U., Ramin, M., Theodoridou, K., von Soosten, D., Walsh, P.J., Waters, S. and Xing, X. 2020. Seaweed and seaweed bioactives for mitigation of enteric methane: challenges and opportunities. *Animals* **10**: 2432.
- Aliarabi, H., Fadayifar, A., Alimohamady, R. and Dezfoulian, A.H. 2019. The effect of maternal supplementation of zinc, selenium, and cobalt as slow-release ruminal bolus in late pregnancy on some blood metabolites and performance of ewes and their lambs. *Biological Trace Element Research* **187**: 403–410.
- Allen, M.R., Fuglestvedt, J.S., Shine, K.P., Reisinger, A., Pierrehumbert, R.T. and Forster, P.M. 2016. New use of global warming potentials to compare cumulative and short-lived climate pollutants. *Nature Climate Change* 6: 773–776.
- Allen, M.R., Shine, K.P., Fuglestvedt, J.S., Millar, R.J., Cain, M., Frame, D.J. and Macey, A.H. 2018. A solution to the misrepresentations of CO 2-equivalent emissions of short-lived climate pollutants under ambitious mitigation. *npj Climate and Atmospheric Science* 1: 1–8.
- Antaya, N.T., Ghelichkhan, M., Pereira, A.B.D., Soder, K.J. and Brito, A.F. 2019. Production, milk iodine, and nutrient utilization in Jersey cows supplemented with the brown seaweed Ascophyllum nodosum (kelp meal) during the grazing season. *Journal of Dairy Science* **102**: 8040–8058.
- Bannink, A., Van Schijndel, M.W. and Dijkstra, J., 2011. A model of enteric fermentation in dairy cows to estimate methane emission for the Dutch National Inventory Report using the IPCC Tier 3 approach. *Animal Feed Science and Technology* **166**: 603–618.
- Basarab, J.A., Beauchemin, K.A., Baron, V.S., Ominski, K.H., Guan, L.L., Miller, S.P. and Crowley, J.J. 2013. Reducing GHG emissions through genetic improvement for feed efficiency: effects on economically important traits and enteric methane production. *Animal* 7: 303–315.
- Bauchop, T. 1967. Inhibition of rumen methanogenesis by methane analogues. *Journal of Bacteriology* 94: 171–175.
- Beauchemin, K.A., McAllister, T.A. and McGinn, S.M. 2009. Dietary mitigation of enteric methane from cattle. CAB Reviews: Perspectives in Agriculture, Veterinary Science, Nutrition and Natural Resources 4: 1–18.
- Beauchemin, K.A., Ungerfeld, E.M., Eckard, R.J. and Wang, M. 2020. Fifty years of research on rumen methanogenesis: lessons learned and future challenges for mitigation. *Animal* 14: s2–s16.
- Belanche, A., Newbold, C.J., Morgavi, D.P., Bach, A., Zweifel, B. and Yáñez-Ruiz, D.R. 2020. A meta-analysis describing the effects of the essential oils blend agolin ruminant on performance, rumen fermentation and methane emissions in dairy cows. *Animals* **10**: 620.
- Bergman, E.N. 1990. Energy contributions of volatile fatty acids from the gastrointestinal tract in various species. *Physiological Reviews* **70**: 567–590.

- Bird-Gardiner, T., Arthur, P.F., Barchia, I.M., Donoghue, K.A. and Herd, R.M. 2017. Phenotypic relationships among methane production traits assessed under ad libitum feeding of beef cattle. *Journal of Animal Science* **95**: 4391–4398.
- Boland, T.M., Quinlan, C., Pierce, K.M., Lynch, M.B., Kenny, D.A., Kelly, A.K. and Purcell, P.J. 2013. The effect of pasture pregrazing herbage mass on methane emissions, ruminal fermentation, and average daily gain of grazing beef heifers. *Journal of Animal Science* 91: 3867–3874.
- Boland, T.M., Pierce, K.M., Kelly, A.K., Kenny, D.A., Lynch, M.B., Waters, S.M., Whelan, S.J. and McKay, Z.C. 2020. Feed intake, methane emissions, milk production and rumen methanogen populations of grazing dairy cows supplemented with various C 18 fatty acid sources. *Animals.* **10**: 2380.
- Broucek, J. 2018. Options to methane production abatement in ruminants: a review. *Journal of Animal and Plant Sciences* 28: 348–364.
- Buddle, B.M., Denis, M., Attwood, G.T., Altermann, E., Janssen, P.H., Ronimus, R.S., Pinares-Patiño, C.S., Muetzel, S. and Wedlock, D.N. 2011. Strategies to reduce methane emissions from farmed ruminants grazing on pasture. *The Veterinary Journal* **188**: 11–17.
- Carberry, C.A., Kenny, D.A., Kelly, A.K. and Waters, S.M. 2014. Quantitative analysis of ruminal methanogenic microbial populations in beef cattle divergent in phenotypic residual feed intake (RFI) offered contrasting diets. *Journal of Animal Science* and Biotechnology 5: 41.
- Castro-Montoya, J., Peiren, N., Cone, J.W., Zweifel, B., Fievez, V. and De Campeneere, S. 2015. In vivo and in vitro effects of a blend of essential oils on rumen methane mitigation. *Livestock Science*. **180**: 134–142.
- Cottle, D.J., Velazco, J., Hegarty, R.S. and Mayer, D.G. Estimating daily methane production in individual cattle with irregular feed intake patterns from short-term methane emission measurements. *Animal* 9, no. 12 (2015): 1949–1957.
- Cummins, S., Finn, J.A., Richards, K.G., Lanigan, G.J., Grange, G., Brophy, C., Cardenas, L.M., Misselbrook, T.H., Reynolds, C.K. and Krol, D.J. 2021. Beneficial effects of multi-species mixtures on N_2O emissions from intensively managed grassland swards. *Science of The Total Environment* **792**: 148163.
- Deighton, M.H., O'Loughlin, B.M., Williams, S.R.O., Moate, P.J., Kennedy, E., Boland, T.M. and Eckard, R.J. 2013. Declining sulphur hexafluoride permeability of polytetrafluoroethylene membranes causes overestimation of calculated ruminant methane emissions using the tracer technique. *Animal Feed Science and Technology* **183**: 86-95.
- Dewhurst, R.J., Fisher, W.J., Tweed, J.K. and Wilkins, R.J. 2003. Comparison of grass and legume silages for milk production. 1. Production responses with different levels of concentrate. *Journal* of Dairy Science 86: 2598–2611.
- Dineen, M., Delaby, L., Gilliland, T. and McCarthy, B. 2018. Metaanalysis of the effect of white clover inclusion in perennial ryegrass swards on milk production. *Journal of Dairy Science*. **101**: 1804–1816.

- EC European Commission. 2005. Press Corner. Available online: https://ec.europa.eu/commission/presscorner/detail/en/ IP_05_1687 [Accessed 14 January 2021].
- Egan, M., Lynch, M.B. and Hennessy, D. 2017. Including white clover in nitrogen fertilized perennial ryegrass swards: effects on dry matter intake and milk production of spring calving dairy cows. *The Journal of Agricultural Science* **155**: 657–658.
- EPA, 2021. Agriculture: Environmental Protection Agency Ireland. Available online: https://www.epa.ie/ghg/ agriculture/#:~:text=Greenhouse%20gas%20emissions%20 from%20agriculture%20in%202019,variety%20of%20 processes%20or%20activities [Accessed 9 April 2021].
- Eugène, M., Sauvant, D., Nozière, P., Viallard, D., Oueslati, K., Lherm, M., Mathias, E. and Doreau, M., 2019. A new Tier 3 method to calculate methane emission inventory for ruminants. *Journal of Environmental Management* 231: 982–988.
- Ferris, C.P., Jiao, H., Murray, S., Gordon, A. and Laidlaw, S. 2020. Effect of dairy cow genotype and concentrate feed level on cow performance and enteric methane emissions during grazing. *Agricultural and Food Science* 29: 130–138.
- Gardiner, T.D., Coleman, M.D., Innocenti, F., Tompkins, J., Connor,
 A., Garnsworthy, P.C., Moorby, J.M., Reynolds, C.K., Waterhouse,
 A. and Wills, D. 2015. Determination of the absolute accuracy of
 UK chamber facilities used in measuring methane emissions from
 livestock. *Measurement* 66: 272–279.
- Garnsworthy, P.C., Craigon, J., Hernandez-Medrano, J.H. and Saunders, N. 2012. On-farm methane measurements during milking correlate with total methane production by individual dairy cows. *Journal of Dairy Science* **95**: 3166–3180.
- Garnsworthy, P.C., Difford, G.F., Bell, M.J., Bayat, A.R., Huhtanen, P., Kuhla, B., Lassen, J., Peiren, N., Pszczola, M., Sorg, D. and Visker, M.H. 2019. Comparison of methods to measure methane for use in genetic evaluation of dairy cattle. *Animals* **9**: 837.
- Gerber, P., Vellinga, T., Opio, C. and Steinfeld H. 2011. Productivity gains and greenhouse gas emissions intensity in dairy systems. *Livestock Science* **139**: 100–108.
- González-Recio, O., López-Paredes, J., Ouatahar, L., Charfeddine, N., Ugarte, E., Alenda, R. and Jiménez-Montero, J.A. 2020. Mitigation of greenhouse gases in dairy cattle via genetic selection: 2. Incorporating methane emissions into the breeding goal. *Journal of Dairy Science* **103**: 7210–7221.
- Goopy, J.P., Donaldson, A., Hegarty, R., Vercoe, P.E., Haynes, F., Barnett, M. and Oddy, V.H. 2014. Low-methane yield sheep have smaller rumens and shorter rumen retention time. *British Journal* of Nutrition. 111: 578–585.
- Goopy, J.P., Robinson, D.L., Woodgate, R.T., Donaldson, A.J., Oddy, V.H., Vercoe, P.E. and Hegarty, R.S. 2016. Estimates of repeatability and heritability of methane production in sheep using portable accumulation chambers. *Animal Production Science* 56: 116–122.
- Grace, C., Lynch, M.B., Sheridan, H., Lott, S., Fritch, R. and Boland, T.M. 2019. Grazing multispecies swards improves ewe and lamb performance. *Animal* 13: 1721–1729.

- Grace, N.D. and Knowles, S.O. 2012. Trace element supplementation of livestock in New Zealand: meeting the challenges of freerange grazing systems. *Veterinary Medicine International* 2012: 639472.
- Günal, M., McCourt, A., Zhao, Y., Yan, Z.G. and Yan, T. 2019. The effect of silage type on animal performance, energy utilisation and enteric methane emission in lactating dairy cows. *Animal Production Science* **59**: 499–505.
- Hammond, K.J., Crompton, L.A., Bannink, A., Dijkstra, J., Yáñez-Ruiz, D.R., O'Kiely, P., Kebreab, E., Eugène, M.A., Yu, Z., Shingfield, K.J. and Schwarm, A. 2016. Review of current in vivo measurement techniques for quantifying enteric methane emission from ruminants. *Animal Feed Science and Technology* 219: 13–30.
- Hammond, K.J., Humphries, D.J., Westbury, D.B., Thompson, A., Crompton, L.A., Kirton, P., Green, C. and Reynolds, C.K. 2014.
 The inclusion of forage mixtures in the diet of growing dairy heifers: impacts on digestion, energy utilisation, and methane emissions. *Agriculture, Ecosystems & Environment* **197**: 88–95.
- Hargreaves, P.R., Sunkel, M. and Towers, E. 2019, September. Impact of Mootral Ruminant[©] on methane emissions and performance on Dairy Cows in real farm conditions. In: "XIIIth International Symposium on Ruminant Physiology".
- Hart, K.J., Martin, P.G., Foley, P.A., Kenny, D.A. and Boland, T.M. 2009. Effect of sward dry matter digestibility on methane production, ruminal fermentation, and microbial populations of zero-grazed beef cattle. *Journal of Animal Science* 87: 3342–3350.
- Harty, M.A., Forrestal, P.J., Watson, C.J., McGeough, K.L., Carolan, R., Elliot, C., Krol, D., Laughlin, R.J., Richards, K.G. and Lanigan, G.J. 2016. Reducing nitrous oxide emissions by changing N fertiliser use from calcium ammonium nitrate (CAN) to urea based formulations. *Science of the Total Environment* **563**: 576–586.
- Haughey, E., Suter, M., Hofer, D., Hoekstra, N.J., McElwain, J.C., Lüscher, A. and Finn, J.A. 2018. Higher species richness enhances yield stability in intensively managed grasslands with experimental disturbance. *Scientific Reports* 8: 1–10.
- Hayes, B.J., Lewin, H.A. and Goddard, M.E. 2013. The future of livestock breeding: genomic selection for efficiency, reduced emissions intensity, and adaptation. *Trends in Genetics* 29: 206–214.
- Hegarty, R.S., Goopy, J.P., Herd, R.M. and McCorkell, B. 2007. Cattle selected for lower residual feed intake have reduced daily methane production. *Journal of Animal Science* 85: 1479–1486.
- Henderson, G., Cox, F., Ganesh, S., Jonker, A., Young, W., Abecia, L., Angarita, E., Aravena, P., Arenas, G.N., Ariza, C. and Attwood, G.T. 2015. Rumen microbial community composition varies with diet and host, but a core microbiome is found across a wide geographical range. *Scientific Reports* **5**: Article number: 14567.
- Herron, J., Hennessy, D., Curran, TP., Moloney, A. and O'Brien, D. 2021. The simulated environmental impact of incorporating white clover into pasture-based dairy production systems. *Journal of Dairy Science* 104: 7902–7918.

- Holden, S.A. and Butler, S.T. 2018. Applications and benefits of sexed semen in dairy and beef herds. *Animal* **12**: s97–s103.
- Hristov, A.N., Oh, J., Firkins, J.L., Dijkstra, J., Kebreab, E., Waghorn, G., Makkar, H.P.S., Adesogan, A.T., Yang, W., Lee, C., Gerber, P.J., Henderson, B. and Tricarico, J.M. 2013. Special topics – Mitigation of methane and nitrous oxide emissions from animal operations: I. a review of enteric methane mitigation options. *Journal of Animal Science* **91**: 5045–5069.
- Hristov, A.N., Oh, J., Giallongo, F., Frederick, T., Weeks, H., Zimmerman, P.R., Harper, M.T., Hristova, R.A., Zimmerman, R.S. and Branco, A.F. 2015. The use of an automated system (GreenFeed) to monitor enteric methane and carbon dioxide emissions from ruminant animals. *Journal of Visualized Experiments* **103**, e52904.
- Hristov, A.N., Oh, J., Giallongo, F., Frederick, T., Harper, M.T., Weeks, H., Branco, A.F., Price, W.J., Moate, P.J., Deighton, M.H. and Williams, S.R.O. 2016. Comparison of the GreenFeed system with the sulfur hexafluoride tracer technique for measuring enteric methane emissions from dairy cows. *Journal of Dairy Science* **99**: 5461–5465.
- Hristov, A.N., Kebreab, E., Niu, M., Oh, J., Bannink, A., Bayat, A.R., Boland, T.M., Brito, A.F., Casper, D.P., Crompton, L.A. and Dijkstra, J., 2018. Symposium review: uncertainties in enteric methane inventories, measurement techniques, and prediction models. *Journal of Dairy Science* **101**: 6655–6674.
- Huhtanen, P., Cabezas-Garcia, E.H., Utsumi, S. and Zimmerman, S. 2015. Comparison of methods to determine methane emissions from dairy cows in farm conditions. *Journal of Dairy Science* **98**: 3394–3409.
- Huws, S.A., Creevey, C.J., Oyama, L.B., Mizrahi, I., Denman, S.E., Popova, M., Muñoz-Tamayo, R., Forano, E., Waters, S.M., Hess, M. and Tapio, I. 2018. Addressing global ruminant agricultural challenges through understanding the rumen microbiome: past, present, and future. *Frontiers in Microbiology* 9: 2161.
- Huyen, N.T., Desrues, O., Alferink, S.J.J., Zandstra, T., Verstegen, M.W.A., Hendriks, W.H. and Pellikaan, W.F. 2016a. Inclusion of sainfoin (Onobrychis viciifolia) silage in dairy cow rations affects nutrient digestibility, nitrogen utilization, energy balance, and methane emissions. *Journal of Dairy Science* **99**: 3566–3577.
- Hynes, D.N., Stergiadis, S., Gordon, A. and Yan, T. 2016b. Effects of concentrate crude protein content on nutrient digestibility, energy utilization, and methane emissions in lactating dairy cows fed fresh-cut perennial grass. *Journal of Dairy Science* **99**: 8858–8866.
- Ineichen, S., Marquardt, S., Wettstein, H.R., Kreuzer, M. and Reidy, B. 2019. Milk fatty acid profile and nitrogen utilization of dairy cows fed ryegrass-red clover silage containing plantain (*Plantago lanceolata* L.). *Livestock Science* 221: 123–132.
- IPCC. 2014: Climate Change 2014: Synthesis Report. In: (eds. Core Writing Team, R.K. Pachauri and L.A. Meyer). Contribution of Working Groups I, II and III to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. IPCC, Geneva, Switzerland, pages 151.

- Jafari, S., Ebrahimi, M., Goh, Y.M., Rajion, M.A., Jahromi, M.F. and Al-Jumaili, W.S. 2019. Manipulation of ruMen ferMentation and Methane gas production by plant secondary Metabolites (saponin, tannin and essential oil) – a review of ten-year studies. *Annals of Animal Science* **19**: 3–29.
- Janssen, P.H. and Kirs, M. 2008. Structure of the archaeal community of the rumen. *Applied Environmental Microbiology* 74: 3619–3625.
- Jayanegara, A., Sarwono, K.A., Kondo, M., Matsui, H., Ridla, M., Laconi, E.B. and Nahrowi. 2018. Use of 3-nitrooxypropanol as feed additive for mitigating enteric methane emissions from ruminants: a meta-analysis. *Italian Journal of Animal Science* **17**: 650–656.
- Johnson, K., Huyler, M., Westberg, H., Lamb, B. and Zimmerman, P. 1994. Measurement of methane emissions from ruminant livestock using a sulfur hexafluoride tracer technique. *Environmental Science & Technology* 28: 359–362.
- Jonker, A., Molano, G., Antwi, C. and Waghorn, G.C. 2016. Enteric methane and carbon dioxide emissions measured using respiration chambers, the sulfur hexafluoride tracer technique, and a GreenFeed head-chamber system from beef heifers fed alfalfa silage at three allowances and four feeding frequencies–. *Journal of Animal Science* 94: 4326–4337.
- Jonker, A., Hickey, S.M., Rowe, S.J., Janssen, P.H., Shackell, G.H., Elmes, S., Bain, W.E., Wing, J., Greer, G.J., Bryson, B. and MacLean, S. 2018. Genetic parameters of methane emissions determined using portable accumulation chambers in lambs and ewes grazing pasture and genetic correlations with emissions determined in respiration chambers. *Journal of Animal Science* **96**: 3031–3042.
- Jonker, A., Hickey, S.M., McEwan, J.C., Rowe, S.J., Janssen, P.H., MacLean, S., Sandoval, E., Lewis, S., Kjestrup, H., Molano, G. and Agnew, M. 2019a. Genetic parameters of plasma and ruminal volatile fatty acids in sheep fed alfalfa pellets and genetic correlations with enteric methane emissions. *Journal of Animal Science* 97: 2711–2724.
- Jonker, A., Farrell, L., Scobie, D., Dynes, R., Edwards, G., Hague, H., McAuliffe, R., Taylor, A., Knight, T. and Waghorn, G. 2019b. Methane and carbon dioxide emissions from lactating dairy cows grazing mature ryegrass/white clover or a diverse pasture comprising ryegrass, legumes and herbs. *Animal Production Science* 59: 1063–1069.
- Kamke, J., Kittelmann, S., Soni, P., Li, Y., Tavendale, M., Ganesh, S., Janssen, P.H., Shi, W., Froula, J., Rubin, E.M. and Attwood, G.T. 2016. Rumen metagenome and metatranscriptome analyses of low methane yield sheep reveals a Sharpea-enriched microbiome characterised by lactic acid formation and utilisation. *Microbiome* 4: 56.
- Kebreab, E., Johnson, K.A., Archibeque, S.L., Pape, D. and Wirth, T., 2008. Model for estimating enteric methane emissions from United States dairy and feedlot cattle. *Journal of Animal Science* 86: 2738–2748.
- Kinley, R.D., Martinez-Fernandez, G., Matthews, M.K., de Nys, R., Magnusson, M. and Tomkins, N.W. 2020. Mitigating the carbon

footprint and improving productivity of ruminant livestock agriculture using a red seaweed. *Journal of Cleaner Production* **259**: 120836.

- Kittelmann, S., Pinares-Patino, C.S., Seedorf, H., Kirk, M.R., Ganesh, S., McEwan, J.C. and Janssen, P.H. 2014. Two different bacterial community types are linked with the low-methane emission trait in sheep. *PLoS One* 9.
- Kliem, K.E., Humphries, D.J., Kirton, P., Givens, D.I. and Reynolds, C.K. 2019. Differential effects of oilseed supplements on methane production and milk fatty acid concentrations in dairy cows. *Animal* 13: 309–317.
- Krol, D.J., Carolan, R., Minet, E., McGeough, K.L., Watson, C.J., Forrestal, P.J., Lanigan, G.J. and Richards, K.G. 2016. Improving and disaggregating N2O emission factors for ruminant excreta on temperate pasture soils. *Science of the Total Environment* 568: 327–338.
- Kumar, S., Treloar, B.P., Teh, K.H., McKenzie, C.M., Henderson, G., Attwood, G.T., Waters, S.M., Patchett, M.L. and Janssen, P.H. 2018. Sharpea and Kandleria are lactic acid producing rumen bacteria that do not change their fermentation products when cocultured with a methanogen. *Anaerobe* 54: 31–38.
- Ku-Vera, J.C., Jiménez-Ocampo, R., Valencia-Salazar, S.S., Montoya-Flores, M.D., Molina-Botero, I.C., Arango, J., Gómez-Bravo, C.A., Aguilar-Pérez, C.F. and Solorio-Sánchez, F.J. 2020. Role of secondary plant metabolites on enteric methane mitigation in ruminants. *Frontiers in Veterinary Science* 7: 584.
- Lanigan, G., Donnellan, T., Hanrahan, K., Carsten, P., Shalloo, L., Krol, D., Forrestal, P., Farrelly, N., O'Brien, D., Ryan, M. and Murphy, P. 2018. "An analysis of abatement potential of Greenhouse Gas emissions in Irish agriculture 2021–2030". Teagasc.
- Lee, J.M., Woodward, S.L., Waghorn, G.C. and Clark, D.A. 2004. Methane emissions by dairy cows fed increasing proportions of white clover (Trifolium repens) in pasture. *Proceedings of the New Zealand Grassland Association*, January, pages 151–155.
- Leahy, S.C., Doyle, N., Mbandlwa, P., Attwood, G.T., Li, Y., Ross, P. and Stanton, C. 2019. Use of lactic acid bacteria to reduce methane production in ruminants, a critical review. *Frontiers in Microbiology* **10**: 2207.
- Leahy, S., Clark, H. and Reisinger, A. 2020. Challenges and prospects for agricultural greenhouse gas mitigation pathways consistent with the Paris Agreement. *Frontiers in Sustainable Food Systems* **4**: 69.
- Li, X., Norman, H.C., Kinley, R.D., Laurence, M., Wilmot, M., Bender, H., de Nys, R. and Tomkins, N. 2018. Asparagopsis taxiformis decreases enteric methane production from sheep. *Animal Production Science* 58: 681–688.
- Li, F., Li, C., Chen, Y., Liu, J., Zhang, C., Irving, B., Fitzsimmons, C. and Plastow, G. 2019. Host genetics influence the rumen microbiota and heritable rumen microbial features associate with feed efficiency in cattle. *Microbiome* **7**: 92.
- Liu, C., Wu, H., Liu, S., Chai, S., Meng, Q. and Zhou, Z. 2019. Dynamic alterations in yak rumen Bacteria community and Metabolome characteristics in response to feed type. *Frontiers in Microbiology* **10**: 1116.

- Løvendahl, P., Difford, G.F., Li, B., Chagunda, M.G.G., Huhtanen, P., Lidauer, M.H., Lassen, J. and Lund, P. 2018. Selecting for improved feed efficiency and reduced methane emissions in dairy cattle. *Animal* **12**: s336–s349.
- Loza, C., Reinsch, T., Loges, R., Taube, F., Gere, J.I., Kluß, C., Hasler, M. and Malisch, C.S. 2021. Methane emission and milk production from jersey cows grazing perennial ryegrass – white clover and multispecies forage mixtures. *Agriculture* **11**: 175.
- Lyons, T., Boland, T., Storey, S. and Doyle, E. 2017. Linseed oil supplementation of lambs' diet in early life leads to persistent changes in rumen microbiome structure. *Frontiers in Microbiology* 8: 1656.
- Machado, L., Magnusson, M., Paul, N.A., Kinley, R., de Nys, R. and Tomkins, N. 2016. Dose-response effects of Asparagopsis taxiformis and Oedogonium sp. on in vitro fermentation and methane production. *Journal of Applied Phycology* 28: 1443–1452.
- Martinez-Fernandez, G., Duval, S.M., Kindermann, M., Schirra, H.J., Denman, S.E. and McSweeney, C.S. 2018. 3-NOP vs Halogenated compound: Methane production, ruminal fermentation and microbial community response in forage fed cattle. *Frontiers in Microbiology* 9: 1582.
- McDonnell, R.P., Hart, K.J., Boland, T.M., Kelly, A.K., McGee, M. and Kenny, D.A. 2016. Effect of divergence in phenotypic residual feed intake on methane emissions, ruminal fermentation, and apparent whole-tract digestibility of beef heifers across three contrasting diets. *Journal of Animal Science* 94: 1179–1193.
- McGinn, S.M., Flesch, T.K., Beauchemin, K.A., Shreck, A. and Kindermann, M. 2019. Micrometeorological methods for measuring methane emission reduction at beef cattle feedlots: evaluation of 3-nitrooxypropanol feed additive. *Journal of Environmental Quality* 48: 1454–1461.
- Meale, S.J., Popova, M., Saro, C., Martin, C., Bernard, A., Lagree, M., Yáñez-Ruiz, D.R., Boudra, H., Duval, S. and Morgavi, D.P. 2021.
 Early life dietary intervention in dairy calves results in a long-term reduction in methane emissions. *Scientific Reports* 11: 1–13.
- Montoya-Flores, M.D., Molina-Botero, I.C., Arango, J., Romano-Muñoz, J.L., Solorio-Sánchez, F.J., Aguilar-Pérez, C.F. and Ku-Vera, J.C. 2020. Effect of Dried Leaves of Leucaena leucocephala on Rumen Fermentation, Rumen Microbial Population, and Enteric Methane Production in Crossbred Heifers. *Animals* **10**: 300.
- Morgavi, D.P., Forano, E., Martin, C. and Newbold, C.J. 2010. Microbial ecosystem and methanogenesis in ruminants. *Animal* 4: 1024–1036.
- Morrison, S.J., McBride, J., Gordon, A.W., Wylie, A.R. and Yan, T. 2017. Methane emissions from grazing Holstein-Friesian heifers at different ages estimated using the sulfur hexafluoride tracer technique. *Engineering* **3**: 753–759.
- Muñoz, C., Yan, T., Wills, D.A., Murray, S. and Gordon, A.W. 2012. Comparison of the sulfur hexafluoride tracer and respiration chamber techniques for estimating methane emissions and correction for rectum methane output from dairy cows. *Journal of Dairy Science* **95**: 3139–3148.

- Muñoz, C., Letelier, P.A., Ungerfeld, E.M., Morales, J.M., Hube, S. and Pérez-Prieto, L.A. 2016. Effects of pregrazing herbage mass in late spring on enteric methane emissions, dry matter intake, and milk production of dairy cows. *Journal of Dairy Science* **99**: 7945–7955.
- Naumann, H.D., Tedeschi, L.O., Zeller, W.E. and Huntley, N.F. 2017. The role of condensed tannins in ruminant animal production: advances, limitations and future directions. *Revista Brasileira de Zootecnia* 46: 929–949.
- Neethirajan, S. 2020. Transforming the adaptation physiology of farm animals through sensors. *Animals* **10**: 1512.
- Newbold, C.J. and Ramos-Morales, E. 2020. Ruminal microbiome and microbial metabolome: effects of diet and ruminant host. *Animal* 14: s78–s86.
- Nguyen, S.H., Nguyen, H.D.T., Bremner, G. and Hegarty, R.S. 2018. Methane emissions and productivity of defaunated and refaunated sheep while grazing. *Small Ruminant Research* **161**: 28–33.
- Niu, M., Kebreab, E., Hristov, A.N., Oh, J., Arndt, C., Bannink, A., Bayat, A.R., Brito, A.F., Boland, T., Casper, D. and Crompton, L.A. 2018. Prediction of enteric methane production, yield, and intensity in dairy cattle using an intercontinental database. *Global Change Biology* 24: 3368–3389.
- O'Brien, D., Shalloo, L., Patton, J., Buckley, F., Grainger, C. and Wallace, M. 2012. A life cycle assessment of seasonal grassbased and confinement dairy farms. *Agricultural Systems* 107: 33–46.
- O'Brien, D., Brennan, P., Humphreys, J., Ruane, E. and Shalloo, L. 2014. An appraisal of carbon footprint of milk from commercial grass-based dairy farms in Ireland according to a certified life cycle assessment methodology. *The International Journal of Life Cycle Assessment* **19**: 1469–1481.
- O'Brien, D. and Shalloo, L. 2016. A Review Of Livestock Methane Emission Factors (2016-CCRP-DS.11). EPA.
- O'Brien, D., Moran, B. and Shalloo, L. 2018. A national methodology to quantify the diet of grazing dairy cows. *Journal of Dairy Science* **101**: 8595–8604.
- O'Connor, A., Moloney, A.P., O'Kiely, P., Boland, T.M. and McGee, M. 2019. Effects of fertiliser nitrogen rate to spring grass on apparent digestibility, nitrogen balance, ruminal fermentation and microbial nitrogen production in beef cattle and in vitro rumen fermentation and methane output. *Animal Feed Science and Technology* 254: 114198.
- O'Mara, F. 2006. "Development of Emission Factors for the Irish Cattle Herd". Environmental Protection Agency, Johnstown Castle, Co, Wexford.
- O'Donovan, M., Hennessy, D. and Creighton, P. 2021. Ruminant grassland production systems in Ireland. *Irish Journal of Agricultural and Food Research* **59**: 225-232.
- Pacheco, D., Waghorn, G. and Janssen, P.H. 2014. Decreasing methane emissions from ruminants grazing forages: a fit with productive and financial realities? *Animal Production Science* 54: 1141–1154.

- Patra, A.K. and Saxena, J. 2010. A new perspective on the use of plant secondary metabolites to inhibit methanogenesis in the rumen. *Phytochemistry* **71**: 1198–1222.
- Patra, A.K. 2016. Recent advances in measurement and dietary mitigation of enteric methane emissions in ruminants. *Frontiers in Veterinary Science* **3**: 39.
- Paul, C., Techen, A.K., Robinson, J.S. and Helming, K., 2019. Rebound effects in agricultural land and soil management: Review and analytical framework. *Journal of Cleaner Production* 227: 1054–1067.
- Peña-Espinoza, M., Valente, A.H., Thamsborg, S.M., Simonsen, H.T., Boas, U., Enemark, H.L., López-Muñoz, R. and Williams, A.R. 2018. Antiparasitic activity of chicory (*Cichorium intybus*) and its natural bioactive compounds in livestock: a review. *Parasites & Vectors* **11**: 475.
- Popova, M., McGovern, E., McCabe, M.S., Martin, C., Doreau, M., Arbre, M., Meale, S.J., Morgavi, D.P. and Waters, S.M. 2017. The structural and functional capacity of ruminal and cecal microbiota in growing cattle was unaffected by dietary supplementation of linseed oil and nitrate. *Frontiers in Microbiology* 8: 937.
- Quinton, C.D., Hely, F.S., Amer, P.R., Byrne, T.J. and Cromie, A.R. 2018. Prediction of effects of beef selection indexes on greenhouse gas emissions. *Animal* **12**: 889–897.
- Roehe, R., Dewhurst, R.J., Duthie, C.A., Rooke, J.A., McKain, N., Ross, D.W., Hyslop, J.J., Waterhouse, A., Freeman, T.C., Watson, M. and Wallace, R.J. 2016. Bovine host genetic variation influences rumen microbial methane production with best selection criterion for low methane emitting and efficiently feed converting hosts based on metagenomic gene abundance. *PLoS Genetics* **12**, e1005846.
- Romero-Perez, A., Okine, E.K., McGinn, S.M., Guan, L.L., Oba, M., Duval, S.M., Kindermann, M. and Beauchemin, K.A. 2015. Sustained reduction in methane production from long-term addition of 3-nitrooxypropanol to a beef cattle diet. *Journal of Animal Science* **93**: 1780–1791.
- Roque, B.M., Salwen, J.K., Kinley, R. and Kebreab, E. 2019. Inclusion of Asparagopsis armata in lactating dairy cows' diet reduces enteric methane emission by over 50 percent. *Journal of Cleaner Production* 234: 132–138.
- Roque, B.M., Venegas, M., Kinley, R.D., de Nys, R., Duarte, T.L., Yang, X. and Kebreab, E. 2021. Red seaweed (*Asparagopsis taxiformis*) supplementation reduces enteric methane by over 80 percent in beef steers. *Plos One* **16**, e0247820.
- Rowe, S.J., Hess, M., Zetouni, L., Hickey, S., Brauning, R., Henry, H., Flay, H., Budel, J., Bryson, B., Janssen, P. and Jonker, A. 2020. Breeding low emitting ruminants: predicting methane from microbes. *Multidisciplinary Digital Publishing Institute Proceedings* **36**: 177.
- Rubino, F., Carberry, C., Waters, S.M., Kenny, D., McCabe, M.S. and Creevey, C.J. 2017. Divergent functional isoforms drive niche specialisation for nutrient acquisition and use in rumen microbiome. *The ISME Journal* **11**: 932–944.
- Russell, J.B. and Martin, S.A. 1984. Effects of various methane inhibitors on the fermentation of amino acids by mixed rumen microorganisms in vitro. *Journal of Animal Science* 59: 1329–1338.

- Saunois, M., Bousquet, P., Poulter, B., Peregon, A., Ciais, P., Canadell, J.G., Dlugokencky, E.J., Etiope, G., Bastviken, D., Houweling, S., Janssens-Maenhout, G., Tubiello, F.N., Castaldi, S., Jackson, R.B., Alexe, M., Arora, V.K., Beerling, D.J., Bergamaschi, P., Blake, D., Brailsford, G., Brovkin, V., Bruhwiler, L., Crevoisier, C., Crill, P., Covey, K., Curry, C., Frankenberg, C., Gedney, N., Höglund-Isaksson, L., Ishizawa, M., Ito, A., Joos, F., Kim, H.S., Kleinen, T., Krummel, P., Lamargue, J.F., Langenfelds, R., Locatelli, R., Machida, T., Maksyutov, S., McDonald, K.C., Marshall, J., Melton, J.R., Morino, I., Naik, V., O'Doherty, S., Parmentier, F.J.W., Patra, P.K., Peng, C., Peng, S., Peters, G.P., Pison, I., Prigent, C., Prinn, R., Ramonet, M., Riley, W.J., Saito, M., Santini, M., Schroeder, R., Simpson, I.J., Spahni, R., Steele, P., Takizawa, A., Thornton, B.F., Tian, H., Tohjima, Y., Viovy, N., Voulgarakis, A., van Weele, M., van der Werf, G.R., Weiss, R., Wiedinmyer, C., Wilton, D.J., Wiltshire, A., Worthy, D., Wunch, D., Xu, X., Yoshida, Y., Zhang, B., Zhang, Z. and Zhu, Q. 2016. The global methane budget 2000-2012. Earth System Science Data 8: 697-751.
- Smith, P.E., Enriquez-Hidalgo, D., Hennessy, D., McCabe, M.S., Kenny, D.A., Kelly, A.K. and Waters, S.M. 2020a. Sward type alters the relative abundance of members of the rumen microbial ecosystem in dairy cows. *Scientific Reports* **10**: 9317.
- Smith, P.E., Waters, S.M., Kenny, D.A., Boland, T.M., Heffernan, J. and Kelly, A.K. 2020b. Replacing barley and soybean meal with byproducts, in a pasture based diet, alters daily methane output and the rumen microbial community in vitro using the rumen simulation technique (RUSITEC). *Frontiers in Microbiology* **11**: 1614.
- Smith, P.E., Waters, S.M., Kenny, D.A., Kirwan, S.F., Conroy, S. and Kelly, A.K. 2021. Effect of divergence in residual methane emissions on feed intake and efficiency, growth and carcass performance, and indices of rumen fermentation and methane emissions in finishing beef cattle. *Journal of Animal Science* **99**: skab275.
- Stewart, E.K., Beauchemin, K.A., Dai, X., MacAdam, J.W., Christensen, R.G. and Villalba, J.J., 2019. Effect of tannin-containing hays on enteric methane emissions and nitrogen partitioning in beef cattle. *Journal of Animal Science* 97: 3286–3299.
- Storm, I.M.L.D., Hellwing, A.L.F., Nielsen, N.I. and Madsen, J. 2012. Methods for measuring and estimating methane emission from ruminants. *Animals (Basel)* 2: 160–183.
- Suter, M., Huguenin-Elie, O. and Lüscher, A. 2021. Multispecies for multifunctions: combining four complementary species enhances multifunctionality of sown grassland. *Scientific Reports* **11**: 3835.
- Tapio, I., Snelling, T.J., Strozzi, F. and Wallace, R.J. 2017. The ruminal microbiome associated with methane emissions from ruminant livestock. *Journal of Animal Science and Biotechnology* 8: 7.
- Totty, V.K., Greenwood, S.L., Bryant, R.H. and Edwards, G.R. 2013. Nitrogen partitioning and milk production of dairy cows grazing simple and diverse pastures. *Journal of Dairy Science* 96: 141–149.
- UNFCCC. 2021. National Inventory Reports Ireland. Available online: https://unfccc.int/documents/225982 [Accessed 16 April 2021].

- van Gastelen, S., Dijkstra, J., Binnendijk, G., Duval, S.M., Heck, J.M.L., Kindermann, M., Zandstra, T. and Bannink, A. 2020. 3-Nitrooxypropanol decreases methane emissions and increases hydrogen emissions of early lactation dairy cows, with associated changes in nutrient digestibility and energy metabolism. *Journal of Dairy Science* **103**: 8074–8093.
- van Soest, P.J. 2018. 23. Digestive flow. In: "Nutritional Ecology of the Ruminant". Cornell University Press, pages 371–384.
- Vyas, D., Alemu, A.W., McGinn, S.M., Duval, S.M., Kindermann, M. and Beauchemin, K.A. 2018. The combined effects of supplementing monensin and 3-nitrooxypropanol on methane emissions, growth rate, and feed conversion efficiency in beef cattle fed high-forage and high-grain diets. *Journal of Animal Science* **96**: 2923–2938.
- Waghorn, G.C., Jonker, A. and Macdonald, K.A. 2016. Measuring methane from grazing dairy cows using GreenFeed. *Animal Production Science* 56: 252–257.
- Wallace, R.J., Rooke, J.A., McKain, N., Duthie, C.A., Hyslop, J.J., Ross, D.W., Waterhouse, A., Watson, M. and Roehe, R. 2015. The rumen microbial metagenome associated with high methane production in cattle. *BMC Genomics* **16**: 839.
- Wallace, R.J., Sasson, G., Garnsworthy, P.C., Tapio, I., Gregson, E., Bani, P., Huhtanen, P., Bayat, A.R., Strozzi, F., Biscarini, F., Snelling, T.J., Saunders, N., Potterton, S.L., Craigon, J., Minuti, A., Trevisi, E., Callegari, M.L., Cappelli, F.P., Cabezas-Garcia, E.H., Vilkki, J., Pinares-Patino, C., Fliegerová, K.O., Mrázek, J., Sechovcová, H., Kopečný, J., Bonin, A., Boyer, F., Taberlet, P., Kokou, F., Halperin, E., Williams, J.L., Shingfield, K.J. and Mizrahi, I. 2019. A heritable subset of the core rumen microbiome dictates dairy cow productivity and emissions. *Science Advances* 5: eaav8391.
- Waters, S.M., Kenny, D.A. and Smith, P.E. 2020. Role of the rumen microbiome in pasture-fed ruminant production systems. In:
 "Improving Rumen Function", (eds. C. McSweeney and R. Mackie), Burleigh Dodds Science Publishing, pages 591–650.
- Weimer, P.J., Stevenson, D.M., Mantovani, H.C. and Man, S.L.C. 2010. Host specificity of the ruminal bacterial community in the dairy cow following near-total exchange of ruminal contents. *Journal of Dairy Science* **93**: 5902–5912.
- Wilson, R.L., Bionaz, M., MacAdam, J.W., Beauchemin, K.A., Naumann, H.D. and Ates, S. 2020. Milk production, nitrogen utilization, and methane emissions of dairy cows grazing grass, forb, and legumebased pastures. *Journal of Animal Science* **98**: skaa220.
- Wims, C.M., Deighton, M.H., Lewis, E., O'loughlin, B., Delaby, L., Boland, T.M. and O'donovan, M. 2010. Effect of pregrazing herbage mass on methane production, dry matter intake, and milk production of grazing dairy cows during the mid-season period. *Journal of Dairy Science* **93**: 4976–4985.
- Woodfield, D.R., Roldan, M.B., Voisey, C.R., Cousins, G.R. and Caradus, J.R. 2019. Improving environmental benefits of white clover through condensed tannin expression. *Journal of New Zealand Grasslands* 81: 195–202.
- Xiang, R., McNally, J., Rowe, S., Jonker, A., Pinares-Patino, C.S., Oddy, V.H., Vercoe, P.E., McEwan, J.C. and Dalrymple, B.P.

2016. Gene network analysis identifies rumen epithelial cell proliferation, differentiation and metabolic pathways perturbed by diet and correlated with methane production. *Scientific Reports* **6**: 39022.

Zhao, Y.G., O'Connell, N.E. and Yan, T. 2016. Prediction of enteric methane emissions from sheep offered fresh perennial ryegrass (*Lolium perenne*) using data measured in indirect open-circuit respiration chambers. *Journal of Animal Science* 94: 2425–2435.