

Review

Role of the Dietary Phytochemical Curcumin in Targeting Cancer Cell Signalling Pathways

Abhay Prakash Mishra ¹, Swetanshu ², Pratchi Singh ^{2,*}, Shikha Yadav ³, Manisha Nigam ^{4,*},
Veronique Seidel ^{5,*} and Celia Fortuna Rodrigues ^{6,7,8}

- ¹ Department of Pharmacology, Faculty of Health Science, University of Free State, Bloemfontein 9300, South Africa
² Department of Biosciences, School of Basic and Applied Sciences, Galgotias University, Greater Noida 203201, Uttar Pradesh, India
³ Department of Pharmacy, School of Medical and Allied Sciences, Galgotias University, Greater Noida 203201, Uttar Pradesh, India
⁴ Department of Biochemistry, H. N. B. Garhwal University, Srinagar Garhwal 246174, Uttarakhand, India
⁵ Natural Products Research Laboratory, Strathclyde Institute of Pharmacy and Biomedical Sciences, University of Strathclyde, 161 Cathedral Street, Glasgow G4 0RE, UK
⁶ LEPABE—Laboratory for Process Engineering, Environment, Biotechnology and Energy, Faculty of Engineering, University of Porto, Rua Dr. Roberto Frias, 4200-465 Porto, Portugal
⁷ ALiCE—Associate Laboratory in Chemical Engineering, Faculty of Engineering, University of Porto, Rua Dr. Roberto Frias, 4200-465 Porto, Portugal
⁸ TOXRUN—Toxicology Research Unit, Cooperativa de Ensino Superior Politécnico e Universitário—CESPU, 4585-116 Gandra PRD, Portugal
* Correspondence: pratchi.singh@galgotiasuniversity.edu.in (P.S.); m.nigam@hnbgu.ac.in (M.N.); veronique.seidel@strath.ac.uk (V.S.)

Abstract: The diarylheptanoid curcumin [(1*E*,6*E*)-1,7-bis(4-hydroxy-3-methoxyphenyl)hepta-1,6-diene-3,5-dione] is one of the phenolic pigments responsible for the yellow colour of turmeric (*Curcuma longa* L.). This phytochemical has gained much attention in recent years due to its therapeutic potential in cancer. A range of drug delivery approaches have been developed to optimise the pharmacokinetic profile of curcumin and ensure that it reaches its target sites. Curcumin exhibits numerous biological effects, including anti-inflammatory, cardioprotective, antidiabetic, and anti-aging activities. It has also been extensively studied for its role as a cancer chemopreventive and anticancer agent. This review focusses on the role of curcumin in targeting the cell signalling pathways involved in cancer, particularly via modulation of growth factors, transcription factors, kinases and other enzymes, pro-inflammatory cytokines, and pro-apoptotic and anti-apoptotic proteins. It is hoped that this study will help future work on the potential of curcumin to fight cancer.

Keywords: cancer; curcumin; signalling pathways



Citation: Mishra, A.P.; Swetanshu; Singh, P.; Yadav, S.; Nigam, M.; Seidel, V.; Rodrigues, C.F. Role of the Dietary Phytochemical Curcumin in Targeting Cancer Cell Signalling Pathways. *Plants* **2023**, *12*, 1782. <https://doi.org/10.3390/plants12091782>

Academic Editors: Anna Oniszczuk and Karolina Wojtunik-Kulesza

Received: 15 February 2023

Revised: 19 April 2023

Accepted: 22 April 2023

Published: 26 April 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Curcumin (C₂₁H₂₀O₆), also known as [(1*E*,6*E*)-1,7-bis(4-hydroxy-3-methoxyphenyl)hepta-1,6-diene-3,5-dione] or diferuloylmethane, is a crystalline substance with a bright orange-yellow colour that is used as a dye and food colouring agent, mainly in the Indian subcontinent. It is most commonly found, along with related compounds collectively known as the curcuminoids, in the rhizome of the spice turmeric (*Curcuma longa* L.) as well as in other plants from the Zingiberaceae family (Table 1, Figure 1). The amount of curcumin in food such as turmeric is influenced by environmental factors such as climate, soil type, and methods used to process the plant material. The content of curcuminoids in *C. longa* has been estimated to range between 1–2 µg/g [1].

Curcumin has a moderate to low degree of solubility in water and a low bioavailability [1,2] (Table 2). When consumed orally, it is moderately absorbed via the gastrointestinal

tract and gets rapidly metabolized in the liver, small intestine and kidney, mostly by reduction and conjugation as curcumin sulphate, curcumin glucuronide and methylated curcumin. Thereafter, it is excreted out via the faeces and urine. Studies have reported that curcumin metabolites, among which tetra and hexahydrocurcumin and tetrahydrocurcumin are the most predominant, contribute to the various pharmacological properties of curcumin [3–5]. The gender of an individual can affect the pharmacokinetics of curcumin. Studies have revealed that females show 1.4 to 2.1 times higher levels of curcuminoids in their plasma than males after oral administration [6,7]. A significant level research has been carried out attempting to increase the bioavailability of curcumin, including using nanoparticles, liposomes, polymeric micelles, phospholipid complexes, and administering curcumin in combination with other substances such as piperine (Table 3).

Table 1. Natural curcuminoids and analogues of curcumin [1].

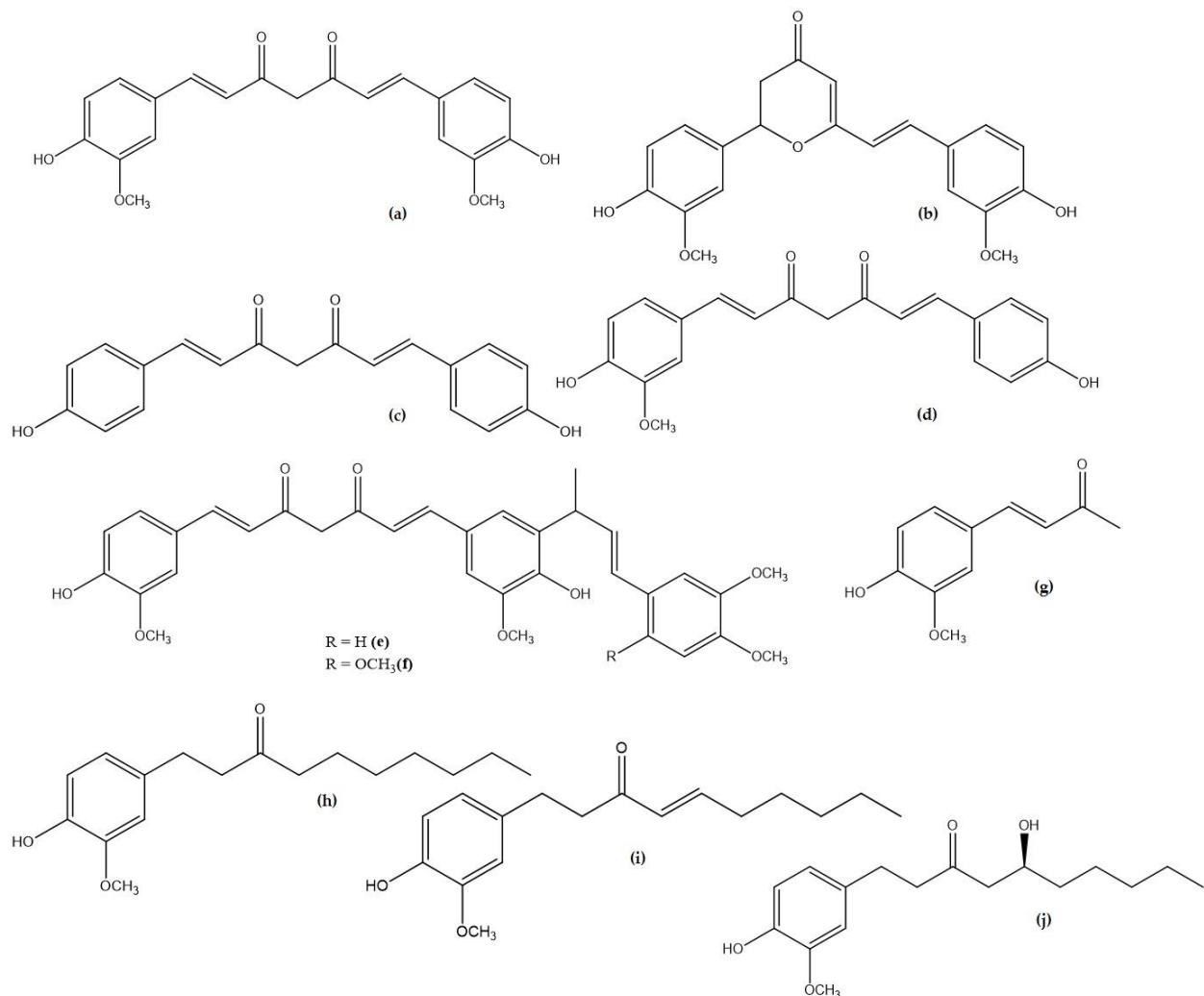
| Compound Name | Plant Origin | Molecular Formula | Pubchem ID |
|--------------------------|--|--|------------|
| Curcumin (a) | <i>Curcuma longa</i> (Turmeric) | C ₂₁ H ₂₀ O ₆ | 969516 |
| Cyclocurcumin (b) | <i>Curcuma longa</i> (Turmeric) | C ₂₁ H ₂₀ O ₆ | 69879809 |
| Bisdemethoxycurcumin (c) | <i>Curcuma longa</i> (Turmeric) | C ₁₉ H ₁₆ O ₄ | 5315472 |
| Demethoxycurcumin (d) | <i>Curcuma longa</i> (Turmeric) | C ₂₀ H ₁₈ O ₅ | 5469424 |
| Cassumunin A (e) | <i>Zingiber cassumunar</i> (Ginger) | C ₃₃ H ₃₄ O ₈ | 10460395 |
| Cassumunin B (f) | <i>Zingiber cassumunar</i> (Ginger) | C ₃₄ H ₃₆ O ₉ | 10054109 |
| Dehydrozingerone (g) | <i>Zingiber officinale Roscoe</i> (Ginger) | C ₁₁ H ₁₂ O ₃ | 5354238 |
| 6-Paradol (h) | <i>Zingiber officinale Roscoe</i> (Ginger) | C ₁₇ H ₂₆ O ₃ | 94378 |
| 6-Shogaol (i) | <i>Zingiber officinale</i> (Ginger) | C ₁₇ H ₂₄ O ₃ | 5281794 |
| 6-Gingerol (j) | <i>Zingiber officinale Roscoe</i> (Ginger) | C ₁₇ H ₂₆ O ₄ | 442793 |

Table 2. Physicochemical properties of curcumin [1,2].

| Formula | C ₂₁ H ₂₀ O ₆ |
|------------------------------------|---|
| Chemical name | [(1E,6E)-1,7-bis(4-hydroxy-3-methoxyphenyl)hepta-1,6-diene-3,5-dione] |
| Molecular weight | 368.38 g/mol |
| pKa values | First (pKa 7.7–8.5) Second (pKa 8.5–10.4) Third (pKa 9.5–10.7) |
| Stable at pH | Between 1–6 |
| Num. heavy atoms | 27 |
| Num. rotatable bonds | 8 |
| Num. H-bond acceptors | 6 |
| Num. H-bond donors | 2 |
| Molar refractivity | 102.80 |
| Melting temperature | 176 °C to 183 °C |
| Water solubility | 0.4 mg/mL |
| Bioavailability score | 0.55 |
| Gastrointestinal absorption | High |
| Blood–brain barrier (BBB) permeant | No |

Table 3. Approaches used to increase the bioavailability of curcumin [7–9].

| Formulations | Curcumin Dose Administered | Plasma Levels of Curcumin |
|----------------------------------|-------------------------------------|---|
| Use of lipid particles | 650 mg | 22.4 ng/mL at 2.4 h |
| | From 2 to 4 g | 30–40 ng/mL between 2 to 4 h |
| Use of micelles | 500 mg | 1189 ng/mL at 1.1 h |
| | 210 mg/day per 4 days | 253 ng/mL (total curcuminoids) |
| Use of piperine | 2 g + 5 mg | 6.92 ng/mL (mean) |
| | 4 g + 24 mg | 136–176 ng/mL (range) |
| | 2 g/kg + 20 mg/kg | 180 ng/mL at 0.75 h |
| Use of hydrophilic nanoparticles | 30 mg | 1.8 ± 2.8 ng/mL |
| | 376 mg | 27.3 ± 6.4 ng/mL at 1.4 h |
| | 30 mg | 25.5 ± 12.2 ng/mL |
| | Multiple doses of 200 or 400 mg/day | 324 ng/mL with a dose of 200 mg of Theracurmin® and 440 ng/mL with a dose of 400 mg |
| | 150 or 210 mg | 189 ± 48 ng/mL with a dose of 150 mg and 275 ± 7 ng/mL with a dose of 210 mg |

**Figure 1.** Chemical structures of curcumin (a) and other curcuminoids (b–j).

Curcumin has various health benefits, including anti-inflammatory, anti-allergic, antioxidant, and anticancer properties [8]. In India, around 1.4 million people are diagnosed with cancer each year, causing 1.2 million deaths annually. In 2020, it was estimated that around 10 million people died due to cancer worldwide. The majority of cases included deaths from lung cancer in males and from breast or cervical cancer in females [9].

The purpose of this review is to discuss the role of curcumin in cancer, with a particular focus on the cell-signalling pathways targeted by curcumin. Laboratory studies carried out to date on animal models suggest that curcumin might have therapeutic potential in cancer. Although these studies are still in the early stage, curcumin remains a promising phytochemical to consider in cancer discovery and development given its significant role in numerous cancer-cell signalling pathways.

2. Methodology

Search engines including Google Scholar, PubMed and Medline were used to retrieve the relevant literature. Almost 200 articles, including original research, review papers, and book chapters, all published between 2000 and 2022, were used to gather relevant information. The primary search terms were 'curcumin and clinical studies', 'curcumin and bioavailability', 'curcumin and breast cancer', 'curcumin and prostate cancer', 'curcumin and brain cancer', 'curcumin and pancreatic cancer', 'curcumin and gastric cancer', 'curcumin and leukaemia', and 'curcumin and nutraceuticals'. Each article was carefully read, and it was ensured that no information was duplicated. ACD/ChemSketch (2021.2.1) was used to draw all chemical structures.

3. Curcumin and Cancer: In Vitro and In Vivo Studies

Curcumin, either alone or in combination with other anticancer drugs, is able to modulate various molecular targets and signalling pathways involved in cancer (Table 4). The sections below discuss the effects of curcumin on various types of cancer, namely lung, breast, prostate, brain, pancreatic, gastric and leukaemia.

Table 4. Effects of curcumin on cell signalling pathways in different types of cancer.

| Type of Cancer | Cell Signalling Pathway | Effect | Model Used | Dose Administered | References |
|-------------------|--------------------------------|---------------------------|--------------------------------------|------------------------------|------------|
| Lung Cancer | Wnt/ β -catenin | Downregulation/inhibition | Human cell line A549 | 60 μ M | [10–19] |
| | VEGF | Downregulation/inhibition | Nude mice | 100 mg/kg | |
| | NF- κ B | Downregulation/inhibition | Nude mice | 100 mg/kg | |
| | Notch 1 | Downregulation/inhibition | Human lung cancer cell lines | 6 μ M | |
| | ERK 1/2 | Downregulation/inhibition | Human NCI-H1975 line | 10 ng/mL | |
| Breast Cancer | Akt/mTOR | Downregulation/inhibition | Human breast cell lines | 10 or 30 μ M | [20–28] |
| | NF- κ B | Downregulation/inhibition | Human breast cell lines | 20 or 25 μ M | |
| | MDR-1 | Downregulation/inhibition | MCF-7 breast cancer cell line | 1.3 μ M | |
| | Bcl-2 and Bcl- xL | Downregulation/inhibition | T47D human breast cells | 20 μ M | |
| | FEN1 | Downregulation/inhibition | MCF-7 breast cancer cell line | 0–50 μ M | |
| Autocrine GH | Downregulation/inhibition | T47D human breast cells | 20 μ M | | |
| Prostate Cancer | Androgen receptor-dependent | Downregulation/inhibition | LNCaP cell line | 0.25 μ M and 0.5 μ M | [29–39] |
| Brain Cancer | STAT3 | Downregulation/inhibition | Human GBM stem cells | 25 μ M | [40–47] |
| | IAP | Downregulation/inhibition | Human GBM stem cells | 25 μ M | |
| | MAPK | Upregulation/activation | Human GBM stem cells | 25 μ M | |
| Pancreatic cancer | Platelet-derived growth factor | Downregulation/inhibition | Rat pancreatic stellate cells | 25 μ M | [48–54] |
| | PI3 K/Akt | Downregulation/inhibition | Panc-1 human pancreatic cells | 20 μ M | |
| | Cdc20 | Downregulation/inhibition | Patu8988 and Panc-1 human cell lines | 10 or 20 μ M | |
| | IAP | Downregulation/inhibition | PANC-1 human cells | 10/50/100 μ M | |
| Gastric cancer | PI3K | Downregulation/inhibition | Human SGC-7901 and BGC-823 cells | 10/20/40 μ M | [55–61] |
| | BCL-2 | Downregulation/inhibition | Human gastric cell lines | 20 μ M | |
| | Wnt3 a/ β -catenin/EMT | Downregulation/inhibition | Human gastric cell lines | 20 μ M | |

Table 4. Cont.

| Type of Cancer | Cell Signalling Pathway | Effect | Model Used | Dose Administered | References |
|----------------|-------------------------|---------------------------|-------------------------------|-------------------|------------|
| Leukaemia-CML | MAPK | Downregulation/inhibition | Human K562 cell line | 5 or 10 mg/L | [62–75] |
| | p210 BCR-ABL | Downregulation/inhibition | Human K562 cell line | 5 or 10 mg/L | |
| | Hsp90 | Downregulation/inhibition | Human K562 cell line | 30 μ M | |
| Leukaemia-CLL | AKT | Downregulation/inhibition | Human CLL B cells | 10–12.5 μ M | |
| | NF- κ B | Downregulation/inhibition | Human CLL B cells | 10–12.5 μ M | |
| | STAT3 | Downregulation/inhibition | Human CLL B cells | 10–12.5 μ M | |
| | XIAP | Downregulation/inhibition | Human CLL B cells | 10–12.5 μ M | |
| | Mcl-1 | Downregulation/inhibition | Human CLL B cells | 10–12.5 μ M | |
| Leukaemia-AML | MMP | Downregulation/inhibition | Human SHI-1 cells | 6.25–25 μ M | |
| Leukaemia-ALL | Bcl-2 | Downregulation/inhibition | Primary human CD34+ AML cells | 0–80 μ M | |
| | MAPK | Downregulation/inhibition | Human SHI-1 cells | 6.25–25 μ M | |
| | AKT/mTOR | Downregulation/inhibition | Human ALL cell lines | 0–40 μ M | |
| | BCR/ABL | Downregulation/inhibition | Human ALL cell lines | 0–40 μ M | |
| | ABL/STAT5 | Downregulation/inhibition | Human ALL cell lines | 0–40 μ M | |

3.1. Lung Cancer

Lung cancer is mostly prevalent in males rather than females [10]. Common treatments for lung cancer involve chemotherapy, radiation therapy, immunotherapy and surgery [11]. Curcumin has been shown to modulate the wingless/integrated Wnt/ β -catenin pathway in A549 lung cancer cells. It downregulates the expression of the nuclear factor- κ B (NF- κ B) and of the vascular endothelial growth factor (VEGF) in that cell line [12]. It also inhibits the expression of the enhancer of zeste homolog 2 (EZH2) in cancerous cells, which eventually downregulates the expression of the gene coding for the neurogenic locus notch homolog protein 1 (Notch 1) [13]. Curcumin has been reported to stop cell division at the G2/M phase, increase cell apoptosis, and show an antiproliferative effect on non-small-cell lung cancer (NSCLC) cells via activating reactive oxidative species (ROS)-DNA damage [14]. The ROS-mediated apoptosis and migration-blocking of lung cancer cells was also reported for a curcumin synthetic derivative [15]. Curcumin has also been shown to inhibit the phosphoinositide 3-kinase (PI3K)/Akt-dependent pathway, leading to apoptosis in various lung cancer cells [16]. This was also observed when administered combined with Paris saponin II (a chemical extracted from the rhizomes of *Paris polyphilla*) [17]. In addition, curcumin enhanced the effects of the cancer chemotherapeutics cisplatin and gefitinib, increasing their antiproliferative ability and inducing apoptosis [18,19].

3.2. Breast Cancer

Breast cancer is the most common type of cancer in women worldwide. Modern treatment approaches involve targeting the production of molecules such as NF- κ B, the human epidermal growth factor receptor 2 (Her-2), Notch, and signal transducer and activator of transcription 3 (STAT-3) [20–22]. The Akt/mTOR-dependent pathway is a predominant signalling pathway associated with breast cancer, and many clinical trials have confirmed that targeting this pathway could lead to promising therapeutic activity [23]. Curcumin has been reported to interfere with the phosphorylation of Akt and the mechanistic target of rapamycin (mTOR) in MCF7 and T47D breast cancer cells [24]. The activation of NF- κ B also plays an important role in cancer and has been linked with the invasion, proliferation, and metastasis of breast cancer cells. Curcumin can inhibit the nuclear translocation of NF- κ B, reducing the levels of p100 and p52 in MCF-7 and MDA-MB-453 breast cancer cells [24]. Its cytotoxicity on MCF-7 cells has been linked with the enhanced expression of the spermidine/spermine N1-acetyltransferase (SSAT) gene, which is also associated with the NF- κ B-dependent signalling pathway [25]. Curcumin has also been reported to inactivate the autocrine growth hormone (GH) signalling pathway in T47D cancer cells as well as reduce the release of anti-apoptotic proteins Bcl-2 and Bcl-xl [26]. Curcumin reduces the overexpression of flap endonuclease 1 (FEN1), an enzyme associated with cisplatin-resistance in breast cancer cells, thereby increasing the sensitivity of cancer cells to this chemotherapeutic agent [27]. Finally, curcumin also downregulates the expression of the multidrug resistance mutation 1 (MDR-1) gene in paclitaxel-resistant cells [28].

3.3. Prostate Cancer

In the western world, prostate cancer ranks second in the types of cancers affecting men [29]. One approach to treat this type of cancer is the use of drugs that inhibit the androgen receptor (AR)-dependent signalling pathway [30,31]. In studies carried out on prostate cancer cells, curcumin has been reported to interact with the mitogen-activated protein kinase (MAPK), epidermal growth factor receptor (EGFR), and NF- κ B signalling pathways [32]. It can inactivate NF- κ B, suppressing the release of inflammatory mediators such as interleukin (IL)-6. It is also able to reduce the levels of cyclooxygenase (COX)-2, Bcl-2, and Bcl-xL [33,34]. In androgen-independent (AI) PC-3 prostate cancer cells, curcumin has been reported to inactivate the NF- κ B pathway and suppress the C-X-C motif chemokine ligand 1 (CXCL-1) and CXCL-2. It can inhibit the MAPKs-activated activator protein (AP-1) transcription factor in prostate cancer cells, eventually suppressing tumour growth [35,36].

It has been demonstrated to significantly reduce the levels of c-Jun N-terminal kinase (JNK) and of the epigenetic marker H3K4 in lymph node carcinoma of the prostate (LNCaP) cells [37]. In both androgen-dependent and androgen-independent prostate cancer cells, curcumin induces apoptosis by downregulating apoptosis suppressor proteins [38]. It has also been shown to block NF- κ B activation and enhance TRAIL-induced cytotoxicity in LNCaP cells [39].

3.4. Brain Cancer

Brain tumours are very resistant to many kinds of therapy [40]. Nearly half of all brain tumours are classified as glioblastoma (GBM) [41,42]. Several studies have been conducted to enhance the delivery of curcumin through the BBB using nanoparticles, as curcumin, in its free form, has low permeability across the BBB [43]. Curcumin has been reported to exert an antiproliferative effect on GBM cells, significantly reducing the levels of non-coding RNAs (miR-21 and miR-378), which play a significant role in the progression of GBM. This reduction in the proliferation of the GBM stem cells by curcumin occurs via activation of the MAPK pathway and inhibition of the inhibitor of apoptosis (IAP) and STAT3-dependent pathways [44]. In many in-vitro studies, curcumin was reported to suppress the proliferation of GBM cells, controlling the expression levels of EGFR, linked to pathways such as the PI3K/Akt and the Janus kinase (JAK)/STAT-dependent pathways [45,46]. Curcumin administered with tyrphostin AG1478 (a type of EGFR kinase inhibitor) causes irreparable damage in DNA, decreasing the viability of GBM cancer cells [47].

3.5. Pancreatic Cancer

The occurrence of pancreatic cancer worldwide is low (3% of all cancers). This type of cancer, with a high level of metastasis, is very difficult to treat and has a high fatality rate [48]. Curcumin has been reported to exert antiproliferative activity on pancreatic stellate cells (PSCs), via suppressing platelet-derived growth factors and the phosphorylation of extracellular signal-related kinases [49]. Recent studies showed that curcumin, together with one of its synthetic derivatives, effectively suppresses tumours by acting on cancer stem cells (CSC) which are the root cause of tumour generation and proliferation [50,51]. Curcumin induces apoptosis in pancreatic cancer cells through the induction of forkhead box O1 and inhibition of the PI3 K/Akt pathway in PANC-1 cancer cells [52]. It downregulates the expression of the key oncogenic factor cell division cycle 20 (cdc20) protein. It increases the expression of p21 and Bcl-2-like protein 11 (Bim), reducing the motility of cancer cells and increasing apoptosis [52,53]. It also shows antiproliferative activity on PANC-1 cancer cells via decreasing the mRNA expression of the IPA protein [54].

3.6. Gastric Cancer

Gastric cancer is the world's third-most lethal cancer [55]. Similarly to other cancers, it is linked to several genes, molecular pathways, signalling molecules, and epigenetic patterns [56]. Curcumin exerts its effect on gastric cancers via inactivation of a number of signalling pathways such as extracellular signal-Regulated Kinases (ERK), Akt, Ras, PI3K, p53, Wnt- β , and MAPKs. Curcumin also inactivates the NF- κ B signalling pathway, reducing the levels of inflammatory mediators including tumour necrosis factor (TNF)- α and various other chemokines and interleukins [57,58]. It has been reported to inhibit the growth of hepatic stellate cells (HSC), promoting p53 gene expression and causing apoptosis [59,60]. It also inhibits the proliferation of BGC-823 and SGC-7901 gastric cancer cells, via interaction with the P13K pathway [60]. Its antiproliferative effect on MKN45, SGC7901, and NCI N87 cells is via regulating Bcl-2 signalling and caspase pathways and inactivating the Wnt3 a/ β -catenin/epithelial-mesenchymal transition (EMT) pathway [61].

3.7. Leukaemia

Leukaemia represents 8% of all cancers worldwide, representing 30% of all cancer occurring in children [62]. Leukaemia can be classified into four subtypes, i.e., acute myeloid

leukaemia (AML), acute lymphoblastic leukaemia (ALL), chronic myeloid leukaemia (CML), and chronic lymphocytic leukaemia (CLL) [63]. The aetiology of CML is directly linked to the expression levels of the P210 BCR-ABL protein translated by the breakpoint cluster region-Abelson (BCR-ABL) gene. This protein is involved in the progenesis of cancerous cells due to its association with different pathways such as MAPK, Ras, and Raf [64]. Curcumin inhibits the MAPK pathway by downregulating p210 BCR-ABL [65]. This downregulation, along with that of the heat shock protein 90 (Hsp90), increases the therapeutic effect of imatinib [65]. This downregulation, along with that of the heat shock protein 90 (Hsp90), increases the therapeutic effect of imatinib [64,65]. Curcumin inactivates NF- κ B in KCL-22 myeloid cells, leading to apoptosis. It also upregulates the TNF α -related apoptosis-inducing ligand (TRAIL) in the same cell line [66]. When administered in combination with another plant polyphenol called carnosic acid, it induced a synergistic effect, inducing apoptosis in AML cells [67]. When administered in combination with daunorubicin, it increases the cytotoxicity of daunorubicin in CD34+ AML cells [68]. In AML cells, curcumin has been reported to decrease the levels of STAT5A and FLT3—a biomarker present in AML [69]. Among all hematological cancers, CLL is most common in the western world [70]. In this type of leukaemia, the levels of T cells and natural killer (NK) cells are high, and there is the presence of defective neoplastic B lymphocytes [71]. Curcumin has been reported to target the pathways related to the persistence of neoplastic B lymphocytes. It can downregulate the expression of Mcl-1, an X-linked inhibitor of apoptosis protein (XIAP), and inhibit the AKT, NF- κ B, and STAT3-dependent pathways *in vitro*. It also leads to cleavage of the poly [ADP-ribose] polymerase-1 (PARP1)-dependent pathways. Curcumin targets various other signalling pathways associated with the progenesis of tumours (e.g., MEK/Raf/ERK and mTOR/Akt, STAT5) [72–75].

4. Clinical Trials of Curcumin in Cancer

The potential therapeutic effects of curcumin on cancer continue to draw great interest from the scientific community. There have been a number of clinical studies conducted on human subjects to evaluate the effectiveness and safety of treatment with curcumin, either alone or in combination with other drugs, and in various cancer types. A summary of the clinical trials conducted to date is presented in Tables 5 and 6. So far, most clinical trials have explored the bioavailability of curcumin, how it affects distinct cancer types, and how well it works to mitigate the adverse effects of radiotherapy and chemotherapy. The results of these trials indicate that curcumin has a promising potential in the treatment of cancer. However, it is important to point out that research on the long-term usage of curcumin supplementation is still lacking, making it difficult to predict if this would elicit any chronic adverse effects. Prospective clinical studies ought to investigate the efficacy and bioavailability of various dosages and/or formulations of curcumin as well as confirm its synergistic effects with currently available cancer chemotherapeutics. Results from the current and upcoming clinical trials will provide a strong scientific basis for the clinical use of curcumin in cancer therapy.

Table 5. Clinical studies on the effect of curcumin on different types of cancer.

| Cancer Type | Study Type | Number of Patients in the Study | Treatment | Endpoints | Results | References |
|---------------------------------------|---------------------------------|---------------------------------|--|---|--|------------|
| Breast cancer | Clinical trial | 14 | Docetaxel + Curcumin (0.5–8 g/day for 7 days) | VEGF and tumour markers levels; Maximal tolerated dose of curcumin; Efficacy; Safety; Toxicity | Decreased levels of VEGF; No cancer progression; Low frequency of toxic effects; Partial response in some patients | [76] |
| Chronic Myeloid Leukaemia | Randomized controlled trial | 50 | Imatinib (400 mg twice daily) + Curcumin (5 g three times daily for 6 weeks) | Plasma nitric oxide levels | Reduced nitric oxide levels | [77] |
| Benign Prostatic Hypertrophy | Pilot project | 61 | Curcumin (1 g per day for 24 weeks) | Quality of life; Signs and symptoms | Improved quality of life; Reduced signs and symptoms of the disease | [78] |
| Head and Neck Squamous Cell Carcinoma | Pilot study | 21 | Single dose of curcumin (1 g) | Cytokine levels and I κ B kinase activity in saliva | Reduced I κ B activity in salivary cells | [79] |
| | Dose-escalation pilot study | 15 | Curcumin (40–200 mg per day for 29 days) | PGE2 levels and COX-2 activity | Dose-dependent decrease in PGE2 levels | [80] |
| | Does-escalation trial (Phase I) | 12 | Curcumin (0.45 g, 1.8 g and 3.6 g per day for 7 days) | Concentrations of curcumin and its metabolites in plasma, and colorectal tissue | Concentrations of Biologically active curcumin in the colorectal tissue | [81] |
| | Does-escalation trial (Phase I) | 15 | Curcumin (0.45–3.6 g per day for 120 days) | PGE2 and glutathione S-transferase activity in blood; Concentration of curcumin and its metabolites in plasma, faeces and urine | Very low levels of curcumin and its metabolites in plasma and urine and dose-dependent decrease in PGE2 levels | [82] |
| | Pilot study | 26 | Curcumin (2.35 g per day for 14 days) | Tolerance, safety and levels of curcumin in the colonic mucosa | Prolonged biologically active levels of curcumin achieved in the colon. Safe and well tolerated | [83] |
| | Clinical trial (Phase I) | 126 | Curcumin (360 mg three times daily for 10–30 days) | p53 expression and TNF- α levels in serum and colorectal tissue | Increased expression of p53; Decreased levels of TNF- α in serum and tissue | [84] |
| | Clinical trial (Phase II) | 44 | Curcumin (2 and 4 g per day for 30 days) | Total number and concentration of 5-hydroxyeicosatetraenoic acid and PGE2 within aberrant crypt foci and normal mucosa | Reduced number of aberrant crypt foci with a dose of 4 g per day | [85] |

Table 5. Cont.

| Cancer Type | Study Type | Number of Patients in the Study | Treatment | Endpoints | Results | References |
|--------------------|-----------------------------|---------------------------------|--|--|---|------------|
| Prostate cancer | Randomized controlled trial | 85 | Soy isoflavones (40 mg) + curcumin (100 mg) for 180 days | Prostate-specific antigen levels in serum | Decreased levels of prostate-specific antigen | [86] |
| | Randomized controlled trial | 40 | Radiotherapy + curcumin (3 g per day for 90 days) | Altered activity of antioxidant enzymes and biochemical and clinical progression-free survivals | Decreased levels of prostate-specific antigen and considerable antioxidant effect | [87] |
| | Clinical trial (Phase I) | 16 | Curcumin (200–400 mg per day for 270 days) | Safety, cytokine levels, pharmacokinetics, NF- κ B activity, efficacy and quality of life | No noteworthy changes in NF- κ B activity or cytokine levels, safe, good pharmacokinetics and improved quality of life | [88] |
| Pancreatic cancer | Clinical trial (Phase I/II) | 21 | Gemcitabine + curcumin (8 g per day for 14 days) | Efficacy, patient compliance and toxicity | Median overall survival time of 161 days; Safe and well tolerated | [89] |
| | Clinical trial (Phase II) | 17 | Curcumin (8 g per day for 30 days) | Toxicity profile and time to tumour progression | Tumour progression of 1–12 months and high frequency of side effects | [90] |
| | Clinical trial (Phase II) | 25 | Curcumin (8 g per day for 60 days) | Tumour markers, tumour response, and adverse effects | Biological response in only 2 patients, poor oral bioavailability, and no toxicity | [91] |
| Intestinal Adenoma | Randomized controlled trial | 44 | Curcumin (1.5 g twice a day for 12 months) | Mean polyp size, total number of polyps and adverse effects | No significant clinical response Very few adverse effects | [92] |

Table 6. Recent clinical trials investigating the effect of curcumin on different cancer types [10].

| Cancer Type | Treatment | Project Title | NCT * | Phase | Estimated/Actual Completion Date |
|-------------------|--|---|-------------|-------|----------------------------------|
| Breast Cancer | Curcumin [®] (CUC-01)+ paclitaxel | Curcumin in Combination with Chemotherapy in Advanced Breast Cancer | NCT03072992 | 2 | 30 June 2019 |
| Colorectal Cancer | Avastin/FOLFIRI + curcumin | Avastin/FOLFIRI in Combination with Curcumin in Colorectal Cancer Patients with Unresectable Metastasis | NCT02439385 | 2 | 1 August 2019 |
| Prostate Cancer | Curcumin + radiation | Nanocurcumin for Prostate Cancer Patients Undergoing Radiotherapy (RT) | NCT02724618 | 2 | April 2022 |
| Breast Cancer | Curcumin | A ‘Window Trial’ on Curcumin for Invasive Breast Cancer Primary Tumours | NCT03980509 | 1 | 30 December 2022 |
| Cervical Cancer | Curcumin | Curcumin in Advanced Cervical Cancer | NCT04294836 | 2 | 31 December 2023 |
| Prostate Cancer | Curcumin | Trial of Curcumin to Prevent Progression of Low-risk Prostate Cancer Under Active Surveillance | NCT03769766 | 3 | November 2026 |

* NCT = National Clinical Trial.

5. Concluding Remarks and Future Perspectives

The activity of curcumin on different types of cancer, including breast cancer, chronic myeloid leukaemia, head and neck squamous cell carcinoma, colorectal cancer, prostate cancer, intestinal adenomas, and cervical cancer, has been demonstrated in numerous *in vitro*, *in vivo* and clinical studies. This effect is mediated via various pathways, including PI3K/Akt, JAK/STAT, MAPK, Wnt/ β -catenin, p53, NF- κ B, and apoptosis-related cell signalling. Curcumin has so far shown a promising role in cancer chemoprevention and chemotherapy. Future research is warranted to identify the most suitable formulation/dosage to be used to guarantee optimal concentrations of curcumin in the blood and tissues and achieve the best outcome.

Author Contributions: P.S., S., A.P.M. and M.N. conceptualized and wrote the original draft. S.Y., C.F.R. and V.S. reviewed and edited. A.P.M. and V.S. participated in drafting the final version. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Data Availability Statement: Not applicable.

Acknowledgments: We are grateful to all the authors whose work is cited in this manuscript.

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

ALL: Acute lymphoblastic leukaemia; AML: Acute myeloid leukaemia; BBB: Blood–brain barrier; Bcl: B Cell Leukemia; BCR-ABL: Breakpoint Cluster Region-Abelson; Bim: Bcl-2 Interacting Mediator; CLL: Chronic lymphocytic leukaemia; CML: Chronic myeloid leukaemia; CSC: Cancer stem cells; DNA: Deoxyribonucleic Acid; EGFR: Epidermal growth factor; ERK: Extracellular signal-regulated kinase; ERK: Extracellular signal-regulated kinases; EZH: Enhancer of zeste homolog; FEN: Flap Endonuclease; FLT: FMS-like tyrosine kinase 3; GBM: Glioblastoma; GH: Growth hormone; Her: Human epidermal growth factor receptor; HSC: Hepatic stellate cells; Hsp: Heat shock protein; IAP: Inhibitor of apoptosis; IPA: Ingenuity Pathways Analysis; JAK/STAT: Janus kinase/signal transducers and activators of transcription; JNK: c-Jun N-terminal kinases; LNCaP: Lymph node carcinoma of the prostate; MAPK: Mitogen-activated protein kinase; MCF-7: Michigan Cancer Foundation—7; MDR: Multidrug resistance mutation; mRNA: Messenger Ribonucleic Acid; NK Cell: natural killer cell; NSCLC: Non-small-cell lung cancer; PI3K: Phosphoinositide 3-kinase; PSC: Pancreatic stellate cells; ROS: Reactive oxygen species; SSAT: Spermidine/spermine N1-acetyltransferase; STAT: Signal transducer and activator of transcription; TNF: Tumour necrosis factor; TRAIL: TNF α -related apoptosis-inducing ligand; VEGF: Vascular endothelial growth factor; XIAP: X-linked inhibitor of apoptosis protein.

References

1. Sohn, S.-I.; Priya, A.; Balasubramaniam, B.; Muthuramalingam, P.; Sivasankar, C.; Selvaraj, A.; Valliammai, A.; Jothi, R.; Pandian, S. Biomedical applications and bioavailability of Curcumin—An updated overview. *Pharmaceutics* **2021**, *13*, 2102. [[CrossRef](#)]
2. Dei Cas, M.; Ghidoni, R. Dietary Curcumin: Correlation between bioavailability and health potential. *Nutrients* **2019**, *11*, 2147. [[CrossRef](#)] [[PubMed](#)]
3. Prasad, S.; Tyagi, A.K.; Aggarwal, B.B. Recent developments in delivery, bioavailability, absorption and metabolism of Curcumin: The golden pigment from golden spice. *Cancer Res. Treat.* **2014**, *46*, 2–18. [[CrossRef](#)] [[PubMed](#)]
4. Stohs, S.J.; Chen, C.Y.O.; Preuss, H.G.; Ray, S.D.; Bucci, L.R.; Ji, J.; Ruff, K.J. The fallacy of enzymatic hydrolysis for the determination of bioactive curcumin in plasma samples as an indication of bioavailability: A comparative study. *BMC Complement. Altern. Med.* **2019**, *19*, 293. [[CrossRef](#)] [[PubMed](#)]
5. Pandey, A.; Chaturvedi, M.; Mishra, S.; Kumar, P.; Somvanshi, P.; Chaturvedi, R. Reductive metabolites of curcumin and their therapeutic effects. *Heliyon* **2020**, *6*, e05469. [[CrossRef](#)]
6. Mahale, J.; Singh, R.; Howells, L.M.; Britton, R.G.; Khan, S.M.; Brown, K. Detection of plasma curcuminoids from dietary intake of turmeric-containing food in human volunteers. *Mol. Nutr. Food Res.* **2018**, *62*, e1800267. [[CrossRef](#)] [[PubMed](#)]

7. Schiborr, C.; Kocher, A.; Behnam, D.; Jandasek, J.; Toelstede, S.; Frank, J. The oral bioavailability of curcumin from micronized powder and liquid micelles is significantly increased in healthy humans and differs between sexes. *Mol. Nutr. Food Res.* **2014**, *58*, 516–527. [[CrossRef](#)]
8. Arslan, A.K.K.; Uzunhisarcikli, E.; Yerer, M.B.; Bishayee, A. The golden spice curcumin in cancer: A perspective on finalized clinical trials during the last 10 years. *J. Cancer Res. Ther.* **2022**, *18*, 19–26. [[CrossRef](#)]
9. Kunnumakkara, A.B.; Harsha, C.; Banik, K.; Vikkurthi, R.; Sailo, B.L.; Bordoloi, D.; Gupta, S.C.; Aggarwal, B.B. Is curcumin bioavailability a problem in humans: Lessons from clinical trials. *Expert Opin. Drug Metab. Toxicol.* **2019**, *15*, 705–733. [[CrossRef](#)]
10. Memon, H.; Patel, B.M. Immune checkpoint inhibitors in non-small cell lung cancer: A bird's eye view. *Life Sci.* **2019**, *233*, 116713. [[CrossRef](#)]
11. Wang, J.Y.; Wang, X.; Wang, X.J.; Zheng, B.Z.; Wang, Y.; Wang, X.; Liang, B. Curcumin inhibits the growth via Wnt/beta-catenin pathway in non-small-cell lung cancer cells. *Eur. Rev. Med. Pharmacol. Sci.* **2018**, *22*, 7492–7499. [[CrossRef](#)]
12. Li, X.; Ma, S.; Yang, P.; Sun, B.; Zhang, Y.; Sun, Y.; Hao, M.; Mou, R.; Jia, Y. Anticancer effects of curcumin on nude mice bearing lung cancer A549 cell subsets SP and NSP cells. *Oncol. Lett.* **2018**, *16*, 6756–6762. [[CrossRef](#)] [[PubMed](#)]
13. Wu, G.Q.; Chai, K.Q.; Zhu, X.M.; Jiang, H.; Wang, X.; Xue, Q.; Zheng, A.H.; Zhou, H.Y.; Chen, Y.; Chen, X.C.; et al. Anti-cancer effects of curcumin on lung cancer through the inhibition of EZH2 and NOTCH1. *Oncotarget* **2016**, *7*, 26535–26550. [[CrossRef](#)] [[PubMed](#)]
14. Wang, C.; Song, X.; Shang, M.; Zou, W.; Zhang, M.; Wei, H.; Shao, H. Curcumin exerts cytotoxicity dependent on reactive oxygen species accumulation in non-small-cell lung cancer cells. *Future Oncol.* **2019**, *15*, 1243–1253. [[CrossRef](#)] [[PubMed](#)]
15. Zhou, G.Z.; Li, A.F.; Sun, Y.H.; Sun, G.C. A novel synthetic curcumin derivative MHMM-41 induces ROS-mediated apoptosis and migration blocking of human lung cancer cells A549. *Biomed. Pharmacother.* **2018**, *103*, 391–398. [[CrossRef](#)] [[PubMed](#)]
16. Jin, H.; Qiao, F.; Wang, Y.; Xu, Y.; Shang, Y. Curcumin inhibits cell proliferation and induces apoptosis of human non-small cell lung cancer cells through the upregulation of miR-192-5p and suppression of PI3K/Akt signaling pathway. *Oncol. Rep.* **2015**, *34*, 2782–2789. [[CrossRef](#)] [[PubMed](#)]
17. Man, S.; Zhang, L.; Cui, J.; Yang, L.; Ma, L.; Gao, W. Curcumin enhances the anti-cancer effects of Paris Saponin II in lung cancer cells. *Cell Prolif.* **2018**, *51*, e12458. [[CrossRef](#)]
18. Jin, X.; Wang, J.; Shen, H.; Ran, R.; Xu, K.; Zhang, W.; Tong, X.; Feng, L. Curcumin co-treatment ameliorates resistance to gefitinib in drug-resistant NCI-H1975 lung cancer cells. *J. Tradit. Chin. Med.* **2017**, *37*, 355–360.
19. Baharuddin, P.; Satar, N.; Fakiruddin, K.S.; Zakaria, N.; Lim, M.N.; Yusoff, N.M.; Zakaria, Z.; Yahaya, B.H. Curcumin improves the efficacy of cisplatin by targeting cancer stem-like cells through p21 and cyclin D1-mediated tumour cell inhibition in non-small cell lung cancer cell lines. *Oncol. Rep.* **2016**, *35*, 13–25. [[CrossRef](#)]
20. Lheureux, S.; Denoyelle, C.; Ohashi, P.S.; De Bono, J.S.; Mottaghy, F.M. Molecularly targeted therapies in cancer: A guide for the nuclear medicine physician. *Eur. J. Nucl. Med. Mol. Imaging* **2017**, *44*, 41–54. [[CrossRef](#)]
21. Yang, S.X.; Polley, E.; Lipkowitz, S. New insights on PI3K/AKT pathway alterations and clinical outcomes in breast cancer. *Cancer Treat. Rev.* **2016**, *45*, 87–96. [[CrossRef](#)]
22. Januskeviciene, I.; Petrikaite, V. Heterogeneity of breast cancer: The importance of interaction between different tumor cell populations. *Life Sci.* **2019**, *239*, 117009. [[CrossRef](#)]
23. Steelman, L.S.; Martelli, A.M.; Cocco, L.; Libra, M.; Nicoletti, F.; Abrams, S.L.; McCubrey, J.A. The therapeutic potential of mTOR inhibitors in breast cancer. *Br. J. Clin. Pharmacol.* **2016**, *82*, 1189–1212. [[CrossRef](#)]
24. Hu, S.; Xu, Y.; Meng, L.; Huang, L.; Sun, H. Curcumin inhibits proliferation and promotes apoptosis of breast cancer cells. *Exp. Ther. Med.* **2018**, *16*, 1266–1272. [[CrossRef](#)] [[PubMed](#)]
25. Coker-Gurkan, A.; Celik, M.; Ugur, M.; Arisan, E.D.; Obakan-Yerlikaya, P.; Durdu, Z.B.; Palavan-Unsal, N. Curcumin inhibits autocrine growth hormone-mediated invasion and metastasis by targeting NF-kappaB signaling and polyamine metabolism in breast cancer cells. *Amino Acids* **2018**, *50*, 1045–1069. [[CrossRef](#)] [[PubMed](#)]
26. Coker-Gurkan, A.; Bulut, D.; Genc, R.; Arisan, E.D.; Obakan-Yerlikaya, P.; Palavan-Unsal, N. Curcumin prevented human autocrine growth hormone (GH) signaling mediated NF-kappaB activation and miR-183-96-182 cluster stimulated epithelial mesenchymal transition in T47D breast cancer cells. *Mol. Biol. Rep.* **2019**, *46*, 355–369. [[CrossRef](#)]
27. Zou, J.; Zhu, L.; Jiang, X.; Wang, Y.; Wang, Y.; Wang, X.; Chen, B. Curcumin increases breast cancer cell sensitivity to cisplatin by decreasing FEN1 expression. *Oncotarget* **2018**, *9*, 11268–11278. [[CrossRef](#)]
28. Attia, Y.M.; El-Kersh, D.M.; Ammar, R.A.; Adel, A.; Khalil, A.; Walid, H.; Eskander, K.; Hamdy, M.; Reda, N.; Mohsen, N.E.; et al. Inhibition of aldehyde dehydrogenase-1 and p-glycoprotein-mediated multidrug resistance by curcumin and vitamin D3 increases sensitivity to paclitaxel in breast cancer. *Chem. Biol. Interact.* **2020**, *315*, 108865. [[CrossRef](#)] [[PubMed](#)]
29. Siegel, R.L.; Miller, K.D.; Jemal, A. Cancer statistics, 2020. *CA Cancer J. Clin.* **2020**, *70*, 7–30. [[CrossRef](#)]
30. Lonergan, P.E.; Tindall, D.J. Androgen receptor signaling in prostate cancer development and progression. *J. Carcinog.* **2011**, *10*, 20. [[CrossRef](#)]
31. Watson, P.A.; Arora, V.K.; Sawyers, C.L. Emerging mechanisms of resistance to androgen receptor inhibitors in prostate cancer. *Nat. Rev. Cancer* **2015**, *15*, 701–711. [[CrossRef](#)] [[PubMed](#)]
32. Tomeh, M.A.; Hadianamrei, R.; Zhao, X. A Review of Curcumin and Its Derivatives as Anticancer Agents. *Int. J. Mol. Sci.* **2019**, *20*, 1033. [[CrossRef](#)] [[PubMed](#)]

33. Thapa, D.; Ghosh, R. Chronic inflammatory mediators enhance prostate cancer development and progression. *Biochem. Pharmacol.* **2015**, *94*, 53–62. [[CrossRef](#)] [[PubMed](#)]
34. Hayden, M.S.; Ghosh, S. NF-kappaB, the first quarter-century: Remarkable progress and outstanding questions. *Genes Dev.* **2012**, *26*, 203–234. [[CrossRef](#)]
35. Kavya, K.; Kumar, M.N.; Patil, R.H.; Hegde, S.M.; Kiran Kumar, K.M.; Nagesh, R.; Babu, R.L.; Ramesh, G.T.; Chidananda Sharma, S. Differential expression of AP-1 transcription factors in human prostate LNCaP and PC-3 cells: Role of Fra-1 in transition to CRPC status. *Mol. Cell Biochem.* **2017**, *433*, 13–26. [[CrossRef](#)]
36. Killian, P.H.; Kronski, E.; Michalik, K.M.; Barbieri, O.; Astigiano, S.; Sommerhoff, C.P.; Pfeffer, U.; Nerlich, A.G.; Bachmeier, B.E. Curcumin inhibits prostate cancer metastasis in vivo by targeting the inflammatory cytokines CXCL1 and -2. *Carcinogenesis* **2012**, *33*, 2507–2519. [[CrossRef](#)]
37. Zhao, W.; Zhou, X.; Qi, G.; Guo, Y. Curcumin suppressed the prostate cancer by inhibiting JNK pathways via epigenetic regulation. *J. Biochem. Mol. Toxicol.* **2018**, *32*, e22049. [[CrossRef](#)]
38. Dorai, T.; Gehani, N.; Katz, A. Therapeutic potential of curcumin in human prostate cancer-I. curcumin induces apoptosis in both androgen-dependent and androgen-independent prostate cancer cells. *Prostate Cancer Prostatic. Dis.* **2000**, *3*, 84–93. [[CrossRef](#)]
39. Deeb, D.; Jiang, H.; Gao, X.; Hafner, M.S.; Wong, H.; Divine, G.; Chapman, R.A.; Dulchavsky, S.A.; Gautam, S.C. Curcumin sensitizes prostate cancer cells to tumor necrosis factor-related apoptosis-inducing ligand/Apo2L by inhibiting nuclear factor-kappaB through suppression of IkappaBalpha phosphorylation. *Mol. Cancer Ther.* **2004**, *3*, 803–812. [[CrossRef](#)]
40. Klinger, N.V.; Mittal, S. Therapeutic potential of curcumin for the treatment of brain tumors. *Oxid. Med. Cell Longev.* **2016**, *2016*, 9324085. [[CrossRef](#)]
41. Weathers, S.P.; Gilbert, M.R. Advances in treating glioblastoma. *F1000Prime Rep.* **2014**, *6*, 46. [[CrossRef](#)]
42. Lathia, J.D.; Mack, S.C.; Mulkearns-Hubert, E.E.; Valentim, C.L.; Rich, J.N. Cancer stem cells in glioblastoma. *Genes Dev.* **2015**, *29*, 1203–1217. [[CrossRef](#)]
43. Anand, P.; Kunnumakkara, A.B.; Newman, R.A.; Aggarwal, B.B. Bioavailability of curcumin: Problems and promises. *Mol. Pharm.* **2007**, *4*, 807–818. [[CrossRef](#)]
44. Wong, S.C.; Kamarudin, M.N.A.; Naidu, R. Anticancer Mechanism of Curcumin on Human Glioblastoma. *Nutrients* **2021**, *13*, 950. [[CrossRef](#)]
45. Zanutto-Filho, A.; Braganhol, E.; Klafke, K.; Figueiro, F.; Terra, S.R.; Paludo, F.J.; Morrone, M.; Bristot, I.J.; Battastini, A.M.; Forcelini, C.M.; et al. Autophagy inhibition improves the efficacy of curcumin/temozolomide combination therapy in glioblastomas. *Cancer Lett.* **2015**, *358*, 220–231. [[CrossRef](#)]
46. Zhao, J.; Zhu, J.; Lv, X.; Xing, J.; Liu, S.; Chen, C.; Xu, Y. Curcumin potentiates the potent antitumor activity of ACNU against glioblastoma by suppressing the PI3K/AKT and NF-kappaB/COX-2 signaling pathways. *Onco. Targets Ther.* **2017**, *10*, 5471–5482. [[CrossRef](#)]
47. Bojko, A.; Cierniak, A.; Adamczyk, A.; Ligeza, J. Modulatory Effects of Curcumin and Tyrphostins (AG494 and AG1478) on Growth Regulation and Viability of LN229 Human Brain Cancer Cells. *Nutr. Cancer* **2015**, *67*, 1170–1182. [[CrossRef](#)] [[PubMed](#)]
48. Oettle, H. Progress in the knowledge and treatment of advanced pancreatic cancer: From benchside to bedside. *Cancer Treat. Rev.* **2014**, *40*, 1039–1047. [[CrossRef](#)] [[PubMed](#)]
49. Masamune, A.; Suzuki, N.; Kikuta, K.; Satoh, M.; Satoh, K.; Shimosegawa, T. Curcumin blocks activation of pancreatic stellate cells. *J. Cell Biochem.* **2006**, *97*, 1080–1093. [[CrossRef](#)] [[PubMed](#)]
50. Singh, A.; Settleman, J. EMT, cancer stem cells and drug resistance: An emerging axis of evil in the war on cancer. *Oncogene* **2010**, *29*, 4741–4751. [[CrossRef](#)]
51. Bao, B.; Ali, S.; Banerjee, S.; Wang, Z.; Logna, F.; Azmi, A.S.; Kong, D.; Ahmad, A.; Li, Y.; Padhye, S.; et al. Curcumin analogue CDF inhibits pancreatic tumor growth by switching on suppressor microRNAs and attenuating EZH2 expression. *Cancer Res.* **2012**, *72*, 335–345. [[CrossRef](#)] [[PubMed](#)]
52. Zhao, Z.; Li, C.; Xi, H.; Gao, Y.; Xu, D. Curcumin induces apoptosis in pancreatic cancer cells through the induction of forkhead box O1 and inhibition of the PI3K/Akt pathway. *Mol. Med. Rep.* **2015**, *12*, 5415–5422. [[CrossRef](#)]
53. Wang, L.; Zhang, J.; Wan, L.; Zhou, X.; Wang, Z.; Wei, W. Targeting Cdc20 as a novel cancer therapeutic strategy. *Pharmacol. Ther.* **2015**, *151*, 141–151. [[CrossRef](#)]
54. Diaz Osterman, C.J.; Gonda, A.; Stiff, T.; Sigaran, U.; Valenzuela, M.M.; Ferguson Bennit, H.R.; Moyron, R.B.; Khan, S.; Wall, N.R. Curcumin Induces Pancreatic Adenocarcinoma Cell Death Via Reduction of the Inhibitors of Apoptosis. *Pancreas* **2016**, *45*, 101–109. [[CrossRef](#)]
55. Ruge, M.; Fassan, M.; Graham, D.Y. Epidemiology of Gastric Cancer. In *Gastric Cancer*; Strong, V.E., Ed.; Springer: Cham, Switzerland, 2015; pp. 23–34.
56. Yamashita, K.; Sakuramoto, S.; Watanabe, M. Genomic and epigenetic profiles of gastric cancer: Potential diagnostic and therapeutic applications. *Surg. Today* **2011**, *41*, 24–38. [[CrossRef](#)] [[PubMed](#)]
57. Kasi, P.D.; Tamilselvam, R.; Skalicka-Wozniak, K.; Nabavi, S.F.; Daglia, M.; Bishayee, A.; Pazoki-Toroudi, H.; Nabavi, S.M. Molecular targets of curcumin for cancer therapy: An updated review. *Tumour. Biol.* **2016**, *37*, 13017–13028. [[CrossRef](#)]
58. Basnet, P.; Skalko-Basnet, N. Curcumin: An anti-inflammatory molecule from a curry spice on the path to cancer treatment. *Molecules* **2011**, *16*, 4567–4598. [[CrossRef](#)] [[PubMed](#)]

59. Jin, H.; Lian, N.; Zhang, F.; Chen, L.; Chen, Q.; Lu, C.; Bian, M.; Shao, J.; Wu, L.; Zheng, S. Activation of PPAR γ /P53 signaling is required for curcumin to induce hepatic stellate cell senescence. *Cell Death Dis.* **2016**, *7*, e2189. [[CrossRef](#)]
60. Fu, H.; Wang, C.; Yang, D.; Wei, Z.; Xu, J.; Hu, Z.; Zhang, Y.; Wang, W.; Yan, R.; Cai, Q. Curcumin regulates proliferation, autophagy, and apoptosis in gastric cancer cells by affecting PI3K and P53 signaling. *J. Cell Physiol.* **2018**, *233*, 4634–4642. [[CrossRef](#)]
61. Liu, W.H.; Yuan, J.B.; Zhang, F.; Chang, J.X. Curcumin inhibits proliferation, migration and invasion of gastric cancer cells via Wnt3a/beta-catenin/EMT signaling pathway. *Zhongguo Zhong Yao Za Zhi* **2019**, *44*, 3107–3115. [[CrossRef](#)]
62. Rafiq, S.; Raza, M.H.; Younas, M.; Naeem, F.; Adeeb, R.; Iqbal, J.; Anwar, P.; Sajid, U.; Manzoor, H.M. Molecular Targets of Curcumin and Future Therapeutic Role in Leukemia. *J. Biosci. Med.* **2018**, *06*, 33–50. [[CrossRef](#)]
63. Jabbour, E.; Kantarjian, H. Chronic myeloid leukemia: 2016 update on diagnosis, therapy, and monitoring. *Am. J. Hematol.* **2016**, *91*, 252–265. [[CrossRef](#)] [[PubMed](#)]
64. Wu, L.X.; Xu, J.H.; Wu, G.H.; Chen, Y.Z. Inhibitory effect of curcumin on proliferation of K562 cells involves down-regulation of p210(bcr/abl) initiated Ras signal transduction pathway. *Acta Pharmacol. Sin.* **2003**, *24*, 1155–1160. [[PubMed](#)]
65. Mukherjee, A.; Sarkar, R.; Mukherjee, S.; Biswas, J.; Roy, M. Curcumin boosts up the efficacy of imatinib mesylate in chronic myelogenous leukemia cell line K-562 by modulation of various markers. *Int. J. Curr. Microbiol. Appl. Sci.* **2016**, *5*, 240–255. [[CrossRef](#)]
66. Gupta, S.C.; Sundaram, C.; Reuter, S.; Aggarwal, B.B. Inhibiting NF- κ B activation by small molecules as a therapeutic strategy. *Biochim. Biophys. Acta (BBA)-Gene Regul. Mech.* **2010**, *1799*, 775–787. [[CrossRef](#)]
67. Pesakhov, S.; Nachliely, M.; Barvish, Z.; Aqaq, N.; Schwartzman, B.; Voronov, E.; Sharoni, Y.; Studzinski, G.P.; Fishman, D.; Danilenko, M. Cancer-selective cytotoxic Ca²⁺ overload in acute myeloid leukemia cells and attenuation of disease progression in mice by synergistically acting polyphenols curcumin and carnosic acid. *Oncotarget* **2016**, *7*, 31847–31861. [[CrossRef](#)]
68. Rao, J.; Xu, D.-R.; Zheng, F.-M.; Long, Z.-J.; Huang, S.-S.; Wu, X.; Zhou, W.-H.; Huang, R.-W.; Liu, Q. Curcumin reduces expression of Bcl-2, leading to apoptosis in daunorubicin-insensitive CD34+ acute myeloid leukemia cell lines and primary sorted CD34+ acute myeloid leukemia cells. *J. Transl. Med.* **2011**, *9*, 71. [[CrossRef](#)]
69. Tima, S.; Ichikawa, H.; Ampasavate, C.; Okonogi, S.; Anuchapreeda, S. Inhibitory effect of turmeric curcuminoids on FLT3 expression and cell cycle arrest in the FLT3-overexpressing EoL-1 leukemic cell line. *J. Nat. Prod.* **2014**, *77*, 948–954. [[CrossRef](#)]
70. Hallek, M. Chronic lymphocytic leukemia: 2013 update on diagnosis, risk stratification and treatment. *Am. J. Hematol.* **2013**, *88*, 803–816. [[CrossRef](#)]
71. Gonzalez-Rodriguez, A.P.; Contesti, J.; Huergo-Zapico, L.; Lopez-Soto, A.; Fernandez-Guizan, A.; Acebes-Huerta, A.; Gonzalez-Huerta, A.J.; Gonzalez, E.; Fernandez-Alvarez, C.; Gonzalez, S. Prognostic significance of CD8 and CD4 T cells in chronic lymphocytic leukemia. *Leuk Lymphoma* **2010**, *51*, 1829–1836. [[CrossRef](#)]
72. Ghosh, A.K.; Kay, N.E.; Secreto, C.R.; Shanafelt, T.D. Curcumin inhibits prosurvival pathways in chronic lymphocytic leukemia B cells and may overcome their stromal protection in combination with EGCG. *Clin. Cancer Res.* **2009**, *15*, 1250–1258. [[CrossRef](#)] [[PubMed](#)]
73. Mishra, D.; Singh, S.; Narayan, G. Curcumin induces apoptosis in Pre-B acute lymphoblastic leukemia cell lines Via PARP-1 cleavage. *Asian Pac. J. Cancer Prev.* **2016**, *17*, 3865–3869.
74. Guo, Y.; Li, Y.; Shan, Q.; He, G.; Lin, J.; Gong, Y. Curcumin potentiates the anti-leukemia effects of imatinib by downregulation of the AKT/mTOR pathway and BCR/ABL gene expression in Ph+ acute lymphoblastic leukemia. *Int. J. Biochem. Cell Biol.* **2015**, *65*, 1–11. [[CrossRef](#)]
75. Zoi, V.; Galani, V.; Lianos, G.D.; Voulgaris, S.; Kyritsis, A.P.; Alexiou, G.A. The role of curcumin in cancer treatment. *Biomedicines* **2021**, *9*, 1086. [[CrossRef](#)] [[PubMed](#)]
76. Bayet-Robert, M.; Kwiatkowski, F.; Leheurteur, M.; Gachon, F.; Planchat, E.; Abrial, C.; Mouret-Reynier, M.A.; Durando, X.; Barthomeuf, C.; Chollet, P. Phase I dose escalation trial of docetaxel plus curcumin in patients with advanced and metastatic breast cancer. *Cancer Biol. Ther.* **2010**, *9*, 8–14. [[CrossRef](#)] [[PubMed](#)]
77. Ghalaut, V.S.; Sangwan, L.; Dahiya, K.; Ghalaut, P.S.; Dhankhar, R.; Saharan, R. Effect of imatinib therapy with and without turmeric powder on nitric oxide levels in chronic myeloid leukemia. *J. Oncol. Pharm. Pract.* **2012**, *18*, 186–190. [[CrossRef](#)]
78. Ledda, A.; Belcaro, G.; Dugall, M.; Luzzi, R.; Scoccianti, M.; Togni, S.; Appendino, G.; Ciammaichella, G. Meriva®, a lecithinized curcumin delivery system, in the control of benign prostatic hyperplasia: A pilot, product evaluation registry study. *Panminerva Med.* **2012**, *54*, 17. [[PubMed](#)]
79. Kim, S.G.; Veena, M.S.; Basak, S.K.; Han, E.; Tajima, T.; Gjertson, D.W.; Starr, J.; Eidelman, O.; Pollard, H.B.; Srivastava, M.; et al. Curcumin treatment suppresses IKK β kinase activity of salivary cells of patients with head and neck cancer: A pilot study. *Clin. Cancer Res.* **2011**, *17*, 5953–5961.
80. Plummer, S.M.; Hill, K.A.; Festing, M.F.; Steward, W.P.; Gescher, A.J.; Sharma, R.A. Clinical development of leukocyte cyclooxygenase 2 activity as a systemic biomarker for cancer chemopreventive agents. *Cancer Epidemiol. Prev. Biomark.* **2001**, *10*, 1295–1299.
81. Garcea, G.; Berry, D.P.; Jones, D.J.; Singh, R.; Dennison, A.R.; Farmer, P.B.; Sharma, R.A.; Steward, W.P.; Gescher, A.J. Consumption of the putative chemopreventive agent curcumin by cancer patients: Assessment of curcumin levels in the colorectum and their pharmacodynamic consequences. *Cancer Epidemiol. Prev. Biomark.* **2005**, *14*, 120–125. [[CrossRef](#)]

82. Sharma, R.A.; Euden, S.A.; Platton, S.L.; Cooke, D.N.; Shafayat, A.; Hewitt, H.R.; Marczylo, T.H.; Morgan, B.; Hemingway, D.; Plummer, S.M.; et al. Phase I clinical trial of oral curcumin: Biomarkers of systemic activity and compliance. *Clin. Cancer Res.* **2004**, *10*, 6847–6854. [[CrossRef](#)] [[PubMed](#)]
83. Irving, G.R.; Howells, L.M.; Sale, S.; Kralj-Hans, I.; Atkin, W.S.; Clark, S.K.; Britton, R.G.; Jones, D.J.; Scott, E.N.; Berry, D.P.; et al. Prolonged biologically active colonic tissue levels of curcumin achieved after oral administration—a clinical pilot study including assessment of patient acceptability. *Cancer Prev. Res.* **2013**, *6*, 119–128. [[CrossRef](#)] [[PubMed](#)]
84. He, Z.Y.; Shi, C.B.; Wen, H.; Li, F.L.; Wang, B.L.; Wang, J. Upregulation of p53 expression in patients with colorectal cancer by administration of curcumin. *Cancer Investig.* **2011**, *29*, 208–213. [[CrossRef](#)] [[PubMed](#)]
85. Carroll, R.E.; Benya, R.V.; Turgeon, D.K.; Vareed, S.; Neuman, M.; Rodriguez, L.; Kakarala, M.; Carpenter, P.M.; McLaren, C.; Meyskens, F.L., Jr.; et al. Phase IIa clinical trial of curcumin for the prevention of colorectal neoplasia. *Cancer Prev. Res.* **2011**, *4*, 354–364. [[CrossRef](#)] [[PubMed](#)]
86. Ide, H.; Tokiwa, S.; Sakamaki, K.; Nishio, K.; Isotani, S.; Muto, S.; Hama, T.; Masuda, H.; Horie, S. Combined inhibitory effects of soy isoflavones and curcumin on the production of prostate-specific antigen. *Prostate* **2010**, *70*, 1127–1133. [[CrossRef](#)] [[PubMed](#)]
87. Hejazi, J.; Rastmanesh, R.; Taleban, F.A.; Molana, S.H.; Hejazi, E.; Ehtejab, G.; Hara, N. Effect of Curcumin Supplementation During Radiotherapy on Oxidative Status of Patients with Prostate Cancer: A Double Blinded, Randomized, Placebo-Controlled Study. *Nutr. Cancer* **2016**, *68*, 77–85. [[CrossRef](#)]
88. Kanai, M.; Otsuka, Y.; Otsuka, K.; Sato, M.; Nishimura, T.; Mori, Y.; Kawaguchi, M.; Hatano, E.; Kodama, Y.; Matsumoto, S.; et al. A phase I study investigating the safety and pharmacokinetics of highly bioavailable curcumin (Theracurmin) in cancer patients. *Cancer Chemother. Pharmacol.* **2013**, *71*, 1521–1530. [[CrossRef](#)]
89. Kanai, M.; Yoshimura, K.; Asada, M.; Imaizumi, A.; Suzuki, C.; Matsumoto, S.; Nishimura, T.; Mori, Y.; Masui, T.; Kawaguchi, Y.; et al. A phase I/II study of gemcitabine-based chemotherapy plus curcumin for patients with gemcitabine-resistant pancreatic cancer. *Cancer Chemother. Pharmacol.* **2011**, *68*, 157–164. [[CrossRef](#)]
90. Epelbaum, R.; Schaffer, M.; Vziel, B.; Badmaev, V.; Bar-Sela, G. Curcumin and gemcitabine in patients with advanced pancreatic cancer. *Nutr. Cancer* **2010**, *62*, 1137–1141. [[CrossRef](#)]
91. Dhillon, N.; Aggarwal, B.B.; Newman, R.A.; Wolff, R.A.; Kunnumakkara, A.B.; Abbruzzese, J.L.; Ng, C.S.; Badmaev, V.; Kurzrock, R. Phase II trial of curcumin in patients with advanced pancreatic cancer. *Clin. Cancer Res.* **2008**, *14*, 4491–4499. [[CrossRef](#)]
92. Cruz-Correa, M.; Hylind, L.M.; Marrero, J.H.; Zahurak, M.L.; Murray-Stewart, T.; Casero, R.A., Jr.; Montgomery, E.A.; Iacobuzio-Donahue, C.; Brosens, L.A.; Offerhaus, G.J.; et al. Efficacy and Safety of Curcumin in Treatment of Intestinal Adenomas in Patients with Familial Adenomatous Polyposis. *Gastroenterology* **2018**, *155*, 668–673. [[CrossRef](#)] [[PubMed](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.