MARSCHNER REVIEW



Plant-soil feedback: incorporating untested influential drivers and reconciling terminology

Jonathan R. De Long[®] · Robin Heinen · Johannes Heinze · Elly Morriën · G. Kenny Png · Sarah J. Sapsford · François P. Teste · Ellen L. Fry

Received: 4 July 2022 / Accepted: 28 January 2023 © The Author(s) 2023

Abstract

Background Plants condition the soil in which they grow, thereby altering the performance of subsequent plants growing in this soil. This phenomenon, known as plant-soil feedback (PSF), has garnered increasing interest. Experiments are moving from single species soil pairings in the glasshouse to community-level field trials. Consequently, our knowledge of the role PSF plays in shaping ecosystem functions has advanced. However, knowledge gaps remain.

Scope Here, we explore intrinsic and extrinsic abiotic and biotic drivers of PSF such as maternal effects, plant functional traits, self-DNA, plantplant competition, herbivory, interactions between soil organisms, temperature, drought, flooding, greenhouse gases, (micro)nutrients, plant-litter-soil

Responsible Editor: Alexia Stokes.

J. R. De Long (⊠) · E. Morriën Department of Ecosystem and Landscape Dynamics, Institute of Biodiversity and Ecosystem Dynamics (IBED-ELD), University of Amsterdam, P.O. Box 94240, 1090 GE Amsterdam, The Netherlands e-mail: j.r.delong@uva.nl

R. Heinen

Terrestrial Ecology Research Group, Department for Life Science Systems, TUM School of Life Sciences, Technical University of Munich, Freising, Germany

J. Heinze

Berlin-Brandenburg Institute of Advanced Biodiversity Research (BBIB), Berlin, Germany feedback and priority effects. These drivers have begun to feature in experiments, thereby increasing our mechanistic understanding of PSF. Nonetheless, many of these topics have received insufficient coverage to determine general principles across larger temporal and spatial scales. Further, conflicting terminology has excluded PSF studies from reviews and meta-analyses. We review terms such as soil sickness, Janzen-Connell hypothesis, soil-related invasive species work, soil legacies, allelopathy and soil-related succession that overlap with PSF but are generally not named as such.

Conclusion Holistic experimental designs that consider the continual reciprocal feedback between the extrinsic environment, plants and soil, as well as the

G. K. Png

Asian School of the Environment, Nanyang Technological University, 50 Nanyang Avenue, 639798 Singapore, Singapore

S. J. Sapsford Harry Butler Institute, Murdoch University, 6150 Murdoch, Australia

F. P. Teste Agriculture and Agri-Food Canada, Swift Current Research and Development Centre, P.O. Box 1030, Swift Current, SK S9H 3X2, Canada

E. L. Fry

Department of Biology, Edge Hill University, Lancashire, Ormskirk L39 4QP, United Kingdom unification of terminologies are necessary if we are to realise the full potential of PSF for understanding and steering ecosystem processes. Here, we compile outstanding questions related to PSF research that emphasis the aforementioned topics and suggest ways to incorporate them into future research in order to advance plant-soil ecology.

 $\label{eq:constraint} \begin{array}{ll} \textbf{Keywords} & Above-belowground interactions} \\ Allelopathy \cdot Decomposition \cdot Herbivory \cdot Invasive \\ species \cdot Janzen-Connell \cdot Maternal effects \cdot Plant \\ functional traits \cdot Plant-soil-litter feedback \cdot Priority \\ effects \cdot Self-DNA \cdot Soil legacies \cdot Soil sickness \cdot \\ Succession \end{array}$

Introduction

As plants grow, they shape the soil environment. Rooting patterns change the soil structure (Canadell et al. 1996), root exudation recruits the soil microbial community (Canarini et al. 2019) and nutrient uptake can deplete soil pools (Bennett and Klironomos 2019), while senesced root and shoot litter may return or add nutrients, facilitate the build-up of soil organic matter (SOM) and alter soil physical characteristics (Hassan et al. 2021b). In turn, these plant-induced alterations to the soil affect the growth of subsequent generations of plants, a phenomenon termed plantsoil feedback (PSF) (Bever 1994; van der Putten et al. 2013). Such PSF can range from positive to negative and the effects can vary between individuals of the same species (i.e., conspecific feedback) or between species (i.e., heterospecific feedback) (van der Putten et al. 2016). Although this process has been known since the dawn of agriculture (i.e., soil sickness: (Cesarano et al. 2017; Huang et al. 2013; Schreiner and Reed 1907), the complexity of the myriad interactions involved means that understanding and predicting PSF remains difficult (Bennett and Klironomos 2019). Recent decades have seen an uptick in PSF research interest (Fig. 1a). Plant-soil feedback research has evolved from glasshouse work focused on single plant-soil conditioning followed by single plant responses (Kulmatiski and Kardol 2008) to multi-factor studies seeking to understand the mechanisms and context-dependencies of PSF in mixed, natural plant communities (Beals et al. 2020; Casper and Castelli 2007; De Long et al. 2019a; Heinen et al. 2020b; Heinze et al. 2016). As our knowledge base has expanded, it is increasingly recognized that PSF plays a pivotal role in shaping plant community development and ecosystem function (Crawford et al. 2019; Suding et al. 2013; van der Putten et al. 2013).

Despite increased attention, finding general principles of PSF impacts on plant communities remains challenging due to the fact that several potentially important abiotic and biotic drivers have received limited attention (Bennett and Klironomos 2019; De Long et al. 2018; Smith-Ramesh and Reynolds 2017) and due to inconsistent terminology across disciplines (De Long et al. 2018; Smith-Ramesh and Reynolds 2017; Veen et al. 2019). The influences of intrinsic biotic drivers, as well as extrinsic abiotic and biotic drivers, have begun to receive attention in PSF experiments. A better understanding of the impacts of such drivers may help explain the influence of interactions with the wider environment and the mechanisms behind PSF (Beals et al. 2020; De Long et al. 2018; Smith-Ramesh and Reynolds 2017). Further, PSF has gone by many names throughout the years, such as soil sickness. Therefore, results going by another name are often being overlooked during meta-analyses (Crawford et al. 2019; Kulmatiski et al. 2008; but see Meisner et al. 2014) and could greatly add to the mechanistic understanding of PSF.

In this review, we have two aims. The first is to highlight intrinsic (biotic) and extrinsic (abiotic and biotic) drivers that have received minimal attention in PSF studies, but are up and coming areas of interest (Fig. 1b, c). We have deliberately chosen to include drivers that have received minimal attention in previous reviews and those that are underrepresented in PSF research based on literature searches (Fig. 1b, c). The second aim is to discuss research fields that may be considered as part of PSF, but have gone by different names over the years, which has limited inclusion of these topics in PSF experiments and meta-analyses (Fig. 1a). Overall, whenever possible, we have chosen to focus predominantly on the most recent literature published, namely studies published within the last 5 years, to emphasise the state of the art in PSF research. We present a conceptual framework demonstrating the developmental trajectory of PSF research



Fig. 1 Panel A shows the number of articles published each year in plant-soil feedback (PSF) and other related topics that fall under the umbrella of plant-soil feedback, but are listed under an alias. The trends demonstrates that many of the research topics showed a steady uptick in publication over time, but PSF seems to be plateauing, indicating that calling topics that are actually PSF by another name might lead to under- or misrepresentation in the literature and meta-analyses. The year 1994 was chosen as a start year because this is when the term plant-soil feedback was coined (Bever 1994). Panels B and C show the total number of studies published

and the necessity to harmonise terminology across fields of related research, whilst expanding study foci to encompass neglected or lesser explored themes. that included under- and unexplored drivers of PSF according to Web of Science searches. Drivers are grouped by intrinsic (panel B) and extrinsic (panel C). Within extrinsic, drivers are further subdivided into abiotic, biotic or a combination of abiotic and biotic. The studies were searched by including the "driver" AND "plant-soil feedback*" or "driver" AND "plant soil feedback*" and all resultant hits were controlled to ensure that the article actually looked at PSF. Literature searches were performed in Scopus and Web of Science in April and October 2022

We conclude with a list of outstanding questions that the aforementioned drivers and merging of existing terminologies can help to answer (Box 1).

Under- and unexplored terrain

- 1) How does PSF affect the fitness of future generations of plants (e.g., maternal effects), with consequences for their ecology?
- 2) Do microbial and plant functional traits interact to drive PSF and under what contexts do their respective influences wax or wane?
- 3) How do above- and belowground mutualistic and/or antagonistic interactions (i.e., herbivores, microorganisms) affect PSF processes and can generalizations be made for certain taxa or functional groups?
- 4) What is the role of self-DNA (and relic DNA) and other plant-perceived molecular signatures in shaping PSF processes?
- 5) What direct and interactive role does above- and belowground plant litter play in driving PSF?
- 6) How do soil stoichiometry and phytochemistry (i.e., (micro)nutrients)) tie in with PSF?
- 7) Does order of the arrival of plants and microbes (i.e., priority effects) change the strength and direction of PSF?

Plant-soil feedback under an alias

- 1) Overall, what can be learned about PSF from closely related fields that go by a different name?
- 2) Can data be mined from deep and grey literature about PSF concepts that have been known for decades to centuries (i.e., soil sickness, allelopathy, succession) with the end goal of conducting a meta-analysis?
- 3) Are there analogies between different alias fields (i.e., Janzen-Connell, plant invasion, soil legacies) in terms of how (a)biotic interactions influence the PSF process or the processes that affect the (a)biotic environment?
- 4) Is there a hierarchical order to PSF (or alias processes)? For example, soil structure > soil nutrients > soil microbes > plant-plant interactions, etc.?

Box 1 Outstanding questions for consideration in future plant-soil feedback (PSF) research

Under- and unexplored PSF terrain

Most early PSF experiments grew a single plant species in soils under glasshouse conditions and then used soils to grow the same plants (i.e., conspecifics), as well as other, usually co-occurring species (i.e., heterospecifics) (Kulmatiski and Kardol 2008). This was a necessary first step in developing the over-arching concept behind PSF: plants change the soils and these (abiotic and biotic) changes to the soil impact on plants that grow in the soil at a later date. However, the full picture is much more complex. Plant growth is influenced by myriad other drivers such as temperature, moisture, competition, herbivory, priority effects, genetic adaptation and maternal effects, which interact to alter plant growth, reproductive performance and functional trait expression in subsequent generations. While literature is amassing on these drivers, many topics are only now reaching the level of understanding that allows us to make cross-linkages for a holistic picture of influences on plants. Here, we highlight a selection of intrinsic biotic drivers, as well as extrinsic abiotic, biotic and abiotic-biotic drivers that are likely to bear strong, direct influences on PSF, as demonstrated by what is known about plant-soil interactions in general and the recent emergence of these topics in published PSF studies. We focus on understudied drivers that have been rarely or never considered in other reviews and opinion pieces on PSF and provide an assessment of certain other drivers which have been considered previously (e.g., herbivory, plant-soil-litter feedback), but still require further research (Fig. 1b, c). Finally, we suggest ways to take the field forward in each instance.

Intrinsic biotic drivers

Maternal effects

The influences of the genotype or phenotype of a maternal plant on the phenotype of the offspring is referred to as maternal effects (Wolf and Wade 2009). The result of maternal effects could be that plants produce offspring better adapted to cope with the stressful conditions experienced by their mother (Herman and Sultan 2011; Roach and Wulff 1987). Such effects can manifest through changes to seed constitution (e.g., size, dispersal characteristics, nutrient content) (Germain and Gilbert 2014), seed microbiomes (Nelson 2018; Shade et al. 2017) and/or via DNA methylation that switches genes off or on that are related to, e.g., plant height (Hu et al. 2015) or reproduction (Alonso et al. 2018). Therefore, maternal effects are an intrinsic biotic driver that may affect PSF. For example, Taraxacum officinale L. exposed to root herbivory produced seeds with a better dispersal capacity (i.e., lower biomass), thereby enabling their progeny to "escape" hostile soils (Bont et al. 2020). Plants grown with a specific, beneficial soil microbial community may also produce offspring better adapted to grow with this particular soil microbial community, leading to enhanced performance of the next generation of plants in the same soil (Wagg et al. 2015).

However, soil-related maternal effects may not always be adaptive. Specifically, maternal plants that experience negative PSF may produce seeds with lower germination rates (De Long et al. 2021). There is also evidence that soil-induced maternal effects can be overridden by interactions with soil abiotic and biotic properties. For example, Stotz et al. (2018) found that plants originating from maternal lines grown with an invasive plant had decreased competitive ability, but when plants were grown in soil conditioned by the invader, competitive ability was increased. Further, De Long et al. (2019b) showed that maternal plants exposed to drought produced offspring that performed worse (i.e., smaller offspring) under ambient moisture (i.e., negative maternal effect), but this effect disappeared under drought conditions. This means that although PSF-related maternal effects are being detected, their relative importance in driving plant community composition may be limited due, in part, to a greater importance of environmental drivers. However, a comparison between the relative importance of environmental drivers versus PSF-induced maternal effects has yet to be carried out. To date, most studies on PSF maternal effects have focused on a limited number of herbaceous, temperate grassland species. Consequently, general conclusions as to the role PSF maternal effects play across plants with different growth habits and across ecosystems cannot be made.

Plant functional traits

Plant functional traits comprise the morphological, physiological or phenological features measurable at the plant individual level (Violle et al. 2007). It is widely recognized that plant traits determine how plants respond to their abiotic and biotic environment, impact on other above- and belowground trophic levels and affect functions and processes (e.g., nutrient cycling) in ecosystems (Kattge et al. 2020). Plant traits influence abiotic soil properties, as well as the soil biological community, indicating that plant traits are an intrinsic biotic driver that plays an important role in PSF (Bergmann et al. 2016; Xi et al. 2021). Chemical (e.g., plant nutrient and defence compound concentrations) and structural (e.g., specific root length and root diameter) plant traits can determine PSF processes by directly affecting the composition of rhizosphere microbial communities (Spitzer et al. 2021; Wilschut et al. 2019) and the decomposition of root (Zhang et al. 2016) and shoot (Eppinga et al. 2011; Ke et al. 2015; Laughlin et al. 2015) litter.

Despite the uptick of research looking at how plant traits influence PSF, most studies have focused on the individual effects of either shoot or root traits on PSF. However, under natural conditions, shoot and root traits interact to determine the outcome of PSF. A recent meta-analysis revealed that dissimilarities in shoot and root traits have the potential to influence the individual feedback of two plant species in each other's soil (i.e., pairwise PSF; Xi et al. 2021). This study also provided evidence that plant traits along the fast- to slow-growing plant economic spectrum (Reich 2014) might be a powerful tool to predict the outcome of PSF and ties into other work demonstrating the influence of traits on PSF processes (Baxendale et al. 2014; Heinen et al. 2020a; Orwin et al. 2010; Teste et al. 2017).

The relationship between plant traits and PSF is not unidirectional and interactions with other organisms and environmental factors occur in natural settings. For example, floral traits and fecundity have been shown to be under the influence of PSF (Jiao et al. 2021), demonstrating this bidirectional relationship. Plant and microbial traits also exist in a bidirectional relationship. This has been investigated, particularly in the context of soil nutrient cycling (Fry et al. 2018a). Plant traits such as litter quality are known to impact on microbial traits such as carbon use efficiency (Allison 2012; Manzoni 2017). In turn, microbial traits such as colonization efficiency can also influence plant traits involved in nutrient uptake (de Souza et al. 2020; Laliberté 2017), potentially leading to knock-on effects for PSF.

Plant traits can also be driven by environmental factors (Lienin and Kleyer 2012; Milanović et al. 2020), thus affecting PSF processes. For example, Werger et al. (2020) found wind-induced changes in root morphological traits affected the strength and direction of PSF. Similar to wind, aboveground herbivory was found to influence root morphology, which would most likely affect PSF (Heinze 2020). Although the first steps have been made, the next challenge in PSF research is to integrate plant traitbased approaches and to understand how interactions with the wider environment affect the relationship between plant functional traits and PSF.

Self-DNA inhibitory effects

DNA carries the instructions that control cell functioning and genetic inheritance. However, there is an additional functional role for extracellular DNA (ex-DNA): after fragmentation, either by natural or artificial decomposition, small DNA molecules (between ~50 and ~2000 base pairs) can exert an inhibitory effect on individuals of the same species (Mazzoleni et al. 2015a). Although for a long time the exact mechanism was unclear, Mazzoleni et al. (2015b) showed for the first time that exposure to fragmented self-DNA (i.e., conspecific) inhibits root growth in plants, while non-self-DNA does not trigger these effects. This inhibitory effect of ex-DNA is not only found in plants (Singh et al. 1999), but also in bacteria (Trinick and Parker 1982), fungi (Bottone et al. 1998) and animals (from insects to humans (Monticolo et al. 2020)). Such inhibitory effects could depend on the similarity of the plant DNA with the fragmented molecules: the self-DNA toxic effect remained to a lesser extent when exposing plants to decomposing litters of phylogenetically similar plants (Mazzoleni et al. 2015b). Self-DNA effects have been proposed as a contributing mechanism to soil sickness effects in agriculture (Cesarano et al. 2017) and therefore is likely an important intrinsic biotic driver influencing PSF. In support of this, Barbero et al. (2016) showed that fragments of self-DNA induced intracellular calcium signalling and plasma membrane depolarization in Phaseolus lunatus L. and Zea mays L., thereby resolving a potential mechanism by which autotoxicity of self-DNA could work. However, Cartenì et al. (2016) proposed additional mechanisms such as a DNAsensor that orchestrates a wide variety of innate cellular immune responses that could also explain inhibitory effects of extracellular DNA. To complicate this even further, Carini et al. (2016) showed that "relic DNA" could even make-up to 40% of DNA in soils, remaining un-degraded for years, and be involved in legacy effects for the next generation of both plants and microbes. With regards to the implications for PSF, self- or non-self DNA could be an overlooked driver of intergenerational shifts in plant performance and warrants more exploration.

Extrinsic biotic drivers

Plant-plant competition

Virtually all plants are engaged in intra- and/or interspecific above-belowground competition, making it highly likely that this extrinsic biotic factor is playing a pivotal role in PSF. Aggressively competitive plants often dominate an ecosystem, meaning that soil pathogens or herbivores may come to specialise in exploiting such an ample resource. In line with this, it has been proposed that negative PSF is responsible for maintaining species diversity in systems where competitive exclusion would otherwise dominate (Bever 2003; van der Putten et al. 2013). Although there is empirical and modelling support for this hypothesis, (Chung et al. 2019; Mack et al. 2019), such competitive interactions can only explain co-existence under real world conditions for certain species, under certain circumstances (Fitzpatrick et al. 2017; Maron et al. 2016) and the short-term nature of most PSF experiments has limited extrapolation to the field (Beckman et al. 2022). In addition, according to a recent meta-analysis (Crawford et al. 2019), most studies to date have typically considered species pairs and modelling studies suggest that such dynamics are unlikely to occur under scenarios where a multitude of species interact (Miller et al. 2022). Another meta-analysis suggests that competition renders PSF effects inconsequential (Lekberg et al. 2018), calling into question the true influence of PSF in the field. However, real world studies have indeed demonstrated that species coexistence can be predicted in part by PSF and competition (Chung et al. 2019; Kulmatiski 2019). Such mixed evidence inhibits general conclusions to be drawn regarding the role of competition in altering PSF.

Variation can also occur when the type, intensity and circumstances of competition are considered. Further, interspecific competition between co-occurring natives was found to exacerbate negative PSF relative to intraspecific competition, but this effect was species-specific and held over both live and sterile soils (Xue et al. 2018b). A recent meta-analysis showed that interspecific competition in heterospecific soils reduced plant growth more than when plants were exposed to intraspecific competition (Beals et al. 2020). There may be no effects of PSF at low levels of competitive density and positive feedbacks realised at the highest levels (Buerdsell et al. 2021). Heterogeneity in the soil can also dictate the outcome of PSF on a species-specific basis (Xue et al. 2018a), which further complicates our ability to apply PSF-competition dynamics to real world scenarios. Interspecific plant competition with native plants has been shown to effectively short-circuit positive PSF induced by invasive species (Crawford and Knight 2017). However, beneficial synergistic effects between positive PSF in invasive plants and interspecific competition with natives have also been observed (Oduor et al. 2022). Taken together, these findings have implications on the management of plant invasions, which require tailor-made management strategies depending on the species considered.

Studies have begun to consider the mechanisms behind PSF-competitive interactions and to tease apart the relative importance of above- versus belowground competition. For example, Shannon et al. (2014) found that arbuscular mycorrhizal fungi colonisation can be reduced both by previous heterospecific soil conditioning, as well as under directly competitive circumstances. Whether the direct competitive circumstances led to the reduced arbuscular mycorrhizal fungi colonisation as the result of, e.g., allelopathic effects versus direct competition for arbuscular mycorrhizal fungi symbionts remains to be tested. Ectomycorrhizal fungi generate more positive PSF versus arbuscular mycorrhizal fungi under competition in the field, which could help explain patterns of diversity and community assembly (Kadowaki et al. 2018). Competition for light can also affect microbialdriven PSFs, with high light conditions increasing plant competitive intensity in the presence of soil microbes, while this effect disappeared under low light (Xi et al. 2019). Nutrient availability can affect the outcome of PSF and competition, with higher nutrient levels alleviating negative biotic effects under competitive scenarios (Klinerova and Dostal 2020). Root competition can be alleviated by conspecific feedbacks, thereby creating niche space for enhanced root growth of an inferior competitor (Hendriks et al. 2015). In sum, it remains unresolved as to the contexts and ecosystems under which PSF is overridden or enhanced by competition, as well as the precise mechanisms that drive these effects.

Interactions with herbivory

Herbivory is a major driving process in ecosystems. As most plant species are potential food sources for a multitude of organisms, herbivory is an important extrinsic biotic factor shaping PSF. Herbivory changes plant-soil interactions mediated via plant physiological processes (e.g., shifts in leaf primary and secondary metabolites, root exudation (reviewed in Biere and Goverse (2016)), which can determine the strength and direction of PSF processes. It has been shown that herbivory by polyphagous caterpillars can alter soil fungal community composition possibly via changes to plant exudation patterns (Bezemer et al. 2013), and that this modifies PSF dynamics in response plants (Kostenko et al. 2012). Several studies suggest that herbivory may mask or alleviate PSF (Bennett et al. 2020; Dietterich et al. 2019; Heinze and Joshi 2018), and that the importance of PSF for plant species increases along a gradient of diminishing herbivory (Heinze et al. 2019). In addition, a large-scale PSF study using 20 herbaceous plant species demonstrated that the individual PSFs experienced by plants exposed to herbivores during the conditioning phase were better predictors of plant landscape abundance and diversity than those of plants that were shielded from herbivory (Heinze et al. 2020). This may indicate that considering the role of PSF independent of herbivory may be insufficient to predict in situ feedbacks.

Taken from the perspective of the herbivore, considerable evidence suggests that PSF also affects the herbivores that feed on the affected plants (Kos et al. 2015; Kostenko et al. 2012). For instance, PSF of twelve different plant species affected consumption and growth of the polyphagous caterpillar Mamestra brassicae L. on individual plants (Heinen et al. 2019) and determined its overall feeding patterns in plant communities (Heinen et al. 2018). Further, PSF is positively correlated with herbivore performance (Heinen et al. 2019), indicating that soils that generate more vigorous plants help improve plant resources for herbivores. On the other hand, various studies have shown strong shifts in phytohormonal defences (Zhu et al. 2018), and shifts in the plant metabolomic profile that can repel herbivores (Huberty et al. 2020; Mutyambai et al. 2019; Ristok et al. 2019). These processes also seem to have a strong successional component, with plants inducing successional changes belowground (e.g., decreases in nutrient availability and increasingly fungal-dominated microbial communities), which feedback on plant growth and tissue quality over time, and subsequently, the performance of herbivores (Howard et al. 2020; Huberty et al. 2022).

Finally, herbivory can interact with other environmental factors. In a mesocosm study, Sveen et al. (2021) illustrated that herbivory and nitrogen deposition interact to influence PSF and arbuscular mycorrhizal fungal colonization, with the surprising finding that herbivory was actually a strong, plant speciesspecific mediator of the negative effect of nitrogen deposition. These complex relationships indicate that herbivory can be beneficial to the plant in some instances, which has implications for our understanding of PSF in real-world conditions. Although herbivores can affect PSF and vice versa, how this interaction affects ecosystem dynamics is not fully understood (Smith-Ramesh and Reynolds 2017). The effects of herbivory seem to mask PSF partially and may fade as complexity in interactions with other environmental factors increases. Unravelling how the different components of PSF (i.e., microbial, nutritional, allelopathic) relate to herbivory from the individual to the community level in the field will increase our understanding of these processes, and their importance for ecosystem functioning (Bennett and Klironomos 2019).

Microbial and soil organism interactions

Soil mutualists, saprotrophs and pathogens play important roles as extrinsic biotic drivers of PSF. (Note: microorganisms that inhabit the soil, but also inhabit plant tissue (e.g., mycorrhizae, other endophytes) could be considered *intrinsic* biotic drivers.) Mutualists and saprotrophs tend to have synergistic interactions that aid in plant growth, whereas pathogens tend to be antagonistic for plant growth (Hannula et al. 2020). Soil pathogens are considered the main drivers behind negative PSF (Bever et al. 2015; Domínguez-Begines et al. 2021) and most PSF studies have largely focused on broad pathogen groups rather than the interactions within and between different functional groups of microorganisms. Further, it has recently been proposed that generalist microbiota can have species-specific PSF effects that correlate with phylogenetic relatedness and plant functional diversity in the wider plant community (Semchenko et al. 2022). Essentially, this suggests that generalist soil organisms can have context-dependent, species-specific effects. Given that soil-borne plant pathogens are expected to become an increasingly pressing global problem (Delgado-Baquerizo et al. 2020), it is important to understand how interactions with mutualists may ameliorate the negative effects of pathogens.

Both pathogens and mutualists vary in their host specificity, ranging from highly specific to a single host to generalists, which can affect PSF (Domínguez-Begines et al. 2021; Hannula et al. 2020). For example, the invasive oomycete pathogen *Phytophthora cinnamomi* (Rands) drove negative PSF in the highly susceptible, dominant *Quercus suber* L., but not in the coexisting *Olea europaea* L. in Mediterranean

mixed forests (Domínguez-Begines et al. 2021). These disparate responses could partly be due to their symbiotic associations with contrasting types of mycorrhizal fungi: Q. suber is ectomycorrhizal, while O. europaea, associates with arbuscular mycorrhizal fungi (Domínguez-Begines et al. 2020). These results are somewhat surprising because evidence suggests that ectomycorrhizal fungi are better at protecting their plant hosts against pathogens in comparison to arbuscular mycorrhizal fungi because ectomycorrhizal fungi produce a mantle around the host root, while arbuscular mycorrhizal fungi grow predominantly within the roots (Tedersoo et al. 2020). Alternatively, it could be due to inherent differences in susceptibility or that mycorrhizal colonisation primed a stronger defence response in O. europaea versus Q. suber (Jung et al. 2012).

Considering interactions between mutualists is also gaining traction. For example, it is known that saprotrophic and ectomycorrhizal fungi both decompose organic matter and have been shown to compete with one another (Fernandez and Kennedy 2016; Lindahl et al. 2021). In line with this, studies have shown that Pinus spp. L. co-invade with beneficial ectomycorrhizal fungi from their native range (Dickie et al. 2010; Policelli et al. 2020; Sapsford et al. 2022). As the density of the co-invading pines and ectomycorrhizae increases, saprotrophic diversity decreases drastically, suggesting this loss is due to competitive exclusion of saprotrophs by ectomycorrhizae (Sapsford et al. 2022). The result is correlated with losses in plant community diversity (Nuñez et al. 2017). In contrast, it has been demonstrated in a growth chamber study that native Nothofagus (Blume) trees and non-native pine trees grown in pine-invaded soils and non-invaded soils grew equally well in both soils in terms of biomass and ectomycorrhizal root colonisation, suggesting the presence of each host (and their mutualists) did not necessarily hinder the other (Policelli et al. 2020). The implications of such interactions on future plant communities and the subsequent PSF remains relatively unknown, particularly when designing invasive species management plans (Dickie et al. 2022).

Plant-associated soil organisms (both microbes and soil animals) exist in a complex, heterogeneous environment where myriad interactions occur. Therefore, considering the entire soil community beyond interactions between microbial functional groups is critical. Recent research showed that conditioning plant species can leave long-term impacts on the soil fungal community (in comparison to bacterial communities that fade faster) and both fungal and bacterial legacies are stored inside plant roots, which can affect the growth of the next plant (Hannula et al. 2021; Heinen et al. 2020b). Soil animals such as nematodes have also been implicated in negative PSF (Brinkman et al. 2015; Wilschut and Geisen 2021; Wilschut et al. 2019), but their interactions with the rhizosphere microbial community can induce positive effects on plants (Wilschut and Geisen 2021). Interactions between nematodes and soil microarthropods can also change PSF effects, but effects depend on plant species identity and the previous conditioning plant species (Kuťáková et al. 2018). The logical follow up is to consider additional microbial functional groups and soil animals (Geisen et al. 2022), eventually building to soil food web analyses that can identify all the major players involved behind PSF.

Extrinsic abiotic drivers

Temperature

Temperature is an extrinsic abiotic factor that strongly influences the physiology and enzyme activities of plants (Bita and Gerats 2013; Moore et al. 2021) and soil (micro)organisms (Nottingham et al. 2019; Walker et al. 2018). Hence, temperature impacts on processes like decomposition (Li et al. 2021; Wang et al. 2021) and nutrient uptake (Pregitzer and King 2005) and thus the growth of, and interactions between, plants and soil (micro)organisms (De Long et al. 2018; Hassan et al. 2022). As these interactions affect ecosystem functioning, it is necessary to understand the effects of temperature on PSF (Hassan et al. 2022), especially because temperature is expected to increase during climate change (Pörtner et al. 2022).

Increasing temperatures impact on soil water availability (Osakabe et al. 2014), but to date many studies on PSFs investigated only the direct effects of drought without including temperature (Fry et al. 2018b; Kaisermann et al. 2017; Martorell et al. 2021; Wilschut and van Kleunen 2021). However, some studies have tested the interacting effects of drought and warming on PSFs (Duell et al. 2019; Pugnaire et al. 2019) or the direct effects of temperature on feedback processes between plants and soils (De Long et al. 2015; van Grunsven et al. 2010). For example, increasing temperature was found to affect PSFs in a native forb (Rasmussen et al. 2020) and native grasses along an elevational gradient (De Long et al. 2015). Further studies that tested whether native and non-native plant species showed different PSF responses to increased temperature showed conflicting results. For instance, van Grunsven et al. (2010) found no overall effect of a 5 °C temperature increase on PSF in both native and non-native plants. In contrast, Duell et al. (2019) observed that warmer growing conditions resulted in more positive PSF for non-native species. This finding was also supported by a recent meta-analysis (Hassan et al. 2022). However, the mechanisms behind these temperature-related PSFs, remain open and partly contradictory. For example, it was found that increasing temperature increased arbuscular mycorrhizal colonization (Rasmussen et al. 2020), but negatively affected their activity (Mohan et al. 2014). Warming was also found to affect fungal to bacterial ratios in soils (Cregger et al. 2014), thereby changing interactions between soil biota and plants. Furthermore, there is empirical evidence that increasing soil temperature impacts the effects of soil biota on plants (Heinze et al. 2017; Pineda et al. 2013) and nutrient cycling (Burns et al. 2013; De Long et al. 2018), with potential consequences for plant-litter-soil feedbacks (Veen et al. 2019).

However, two recent meta-analyses revealed no overall effect of temperature on PSF (Beals et al. 2020; Hassan et al. 2022), although plants with different life-cycle traits (De Long et al. 2018) or invasion status (Duell et al. 2019) might differ in their PSF response to warming. Potential differences in PSF might be due to dissimilarities in nutrient uptake rates between fast- and slow-growing species and the fact that temperature impacts on nutrient cycling (Burns et al. 2013; De Long et al. 2018). Furthermore, increasing temperature was found to enhance pathogen accumulation (Crawford et al. 2019; Romero et al. 2022), thus increasing negative PSFs experienced by native plant species and potentially enhancing the enemy release experienced by non-natives (Duell et al. 2019; Hassan et al. 2022).

Besides direct effects on soil biota and thus PSFs, temperature is likely to interact with other drivers of PSFs. For instance, it can be assumed that increasing temperature impacts on PSF via effects on herbivores. Warmer environmental conditions influence the growth and feeding activity of insect herbivores (Lemoine et al. 2014). Such temperature-related effects on herbivory might result in changes in root exudation or plant litter quality via accumulation of defence compounds (Massey et al. 2007) as plants respond to herbivory - that in turn feedback on plant growth via effects on soil biota. Furthermore, by influencing photosynthesis and thus plant functional traits like specific leaf area and leaf dry matter content (Zhu et al. 2020), increasing temperature will likely impact plant-litter-soil feedbacks. As plants species differ in their growth responses to temperature (Gremer et al. 2012), it is also possible that temperature effects might affect competition between plant species and subsequently their relative contribution to PSFs. Overall, more research is needed to get a comprehensive picture of temperature effects on PSF.

Drought and flooding

There has been increasing interest in the effects of drought on PSF. Given that severe drought events are expected to increase as climate change advances (Pörtner et al. 2022), understanding the effect of this extrinsic abiotic driver on PSF has become paramount. Drought leaves a strong signature in the soil, changing nutrient availability and stoichiometry (Gao et al. 2020) and soil microorganism community composition (Canarini et al. 2021; Meisner et al. 2018), consequently influencing plant growth (Buchenau et al. 2022). Such drought effects can have implications for ecosystem processes such as carbon cycling (Fuchslueger et al. 2016). It has also been demonstrated that drought can effectively override PSF (Fry et al. 2018b), create a positive PSF (Rutten and Gomez-Aparicio 2018) or shift PSF in a way that alters plant-plant competition (Kaisermann et al. 2017). Including interactions between drought and PSF can better predict plant landscape abundance versus considering PSF in isolation (Xi et al. 2022). Drought can interact with other global change factors, such as N deposition, leading to disruption in mycorrhizal colonisation, shifts in plant biomass allocation and more negative PSF (Valliere and Allen 2016). In addition, plants from different functional groups can show disparate responses to drought legacies (Hassan et al. 2021a), leading to potential shifts in plant community composition in the long-term. A recent meta-analysis showed that soil microbial feedbacks can enhance plant growth under drought conditions (Beals et al. 2020). However, these results were based on only seven papers, which indicates that no broad conclusions about drought and PSF can be reached as of yet.

In contrast to drought, the impact of flooding on PSF has received very limited attention. Flooding can have direct damaging effects on the soil by enhancing erosion (Julian and Torres 2006) or washing away nutrients and organic matter (Talbot et al. 2018; Yarwood 2018). Positive effects can occur, however, when flood waters recede and deposit nutrient-rich soils (Talbot et al. 2018). With flooding there tends to be a shift from aerobic- to anaerobic-dominated soil microbial communities, with likely cascade effects on plants if these microorganisms produce phytotoxic chemicals or induce shifts in root metabolic profiles (Hartman and Tringe 2019). Enhanced soil moisture after flooding could generate a proliferation of pathogens (Martínez-Arias et al. 2022), with potentially synergistic effects on negative PSF. From the perspective of the plant, flooding changes virtually all aspects and functions, including metabolism, stress hormone production, respiration and overall growth during all life stages (Zhou et al. 2020). The strongest directive negative effect of flooding on plants is the deprivation of oxygen in the soil (Jackson 1985). Species-rich plant communities can mitigate the effect of these anoxic flooding conditions by generating greater soil porosity via diverse rooting strategies (Wright et al. 2017), which could lead to future PSF. One of the few PSF studies that included a flooding component found that negative feedbacks in rice (Oryza sativa L.) increased with time since land conversation to rice paddies (Edwards et al. 2019). However, this study was conducted in a permanently flooded field and did not explicitly explore changes to the soil between flooded versus unflooded fields. Thus, how flooding drives PSF remains unclear.

Greenhouse gases: carbon dioxide, methane, nitrogen oxides and ozone

Anthropogenic greenhouse gas (GHG) emissions (e.g., carbon dioxide (CO₂), methane (CH₄), nitrous oxides (NO_X)) are increasing and there is abundant evidence that many of these have direct effects on plant function and plant-soil interactions (Maček et al. 2019; Rosado-Porto et al. 2022). These GHGs

are likely to directly affect PSF through alteration of litter quality, shifts in plant growth or by changing the strength and direction of plant-microbial relationships (Maček et al. 2019). Elevated CO_2 (eCO₂) has been shown to increase above- and belowground plant biomass (Terrer et al. 2021), photosynthetic rates (Li et al. 2018) and labile carbon inputs to soil (Kuzyakov et al. 2019). Resultantly, more carbon is being fixed without a concomitant increase in (micro)nutrients and long-term CO₂ experiments (e.g., Free Air Carbon Dioxide Enrichment (FACE)) have shown a significant decrease in concentrations of nutrients in plant tissue. This can have cascading effects through the soil food web, potentially changing litter decomposition rates (Loladze 2014; Penuelas et al. 2020). The decomposition of previously stable soil carbon (i.e., carbon priming) can occur with eCO₂, likely due to increased competition between microbes and plants for resources, the activation of different microbial groups or preferential substrate consumption (Reich et al. 2006). eCO_2 also has direct effects on soil microbial community composition. For example, in a scrub-oak system, carbon-degrading fungi and associated enzymatic activities were increased, leading to reduced carbon storage (Carney et al. 2007). Further, there is evidence of the up-regulation of genes related to nitrogen cycling, indicating that PSF could be positive under eCO₂ through increased soil nitrogen availability (Yu et al. 2018). All of these effects are likely to have strong intergenerational impacts on plant growth and community dynamics. Given the contrasting effects of eCO₂ on carbon storage and the microbial community composition, it is still unclear if net PSF will be positive or negative. It seems likely that there will be high context dependency, and various interactions with other global change drivers such as drought and flooding (Walker et al. 2021).

While CH_4 is commonly studied in light of its role in radiative forcing, recent evidence has come to light that plants can produce CH_4 independently of microbes in response to cell damage from reactive oxygen species. CH_4 is produced during the breakdown of plant structural materials such as pectin and lignin (Keppler et al. 2008; Messenger et al. 2009). Methane could play the role of an antioxidant in plant cells, protecting against a range of abiotic stresses (Li et al. 2020). The breakdown of these large molecules and the CH₄-induced resilience to abiotic stress could have interesting implications for plant-litter-soil feedback (PLSF). The resultant alterations to litter quality, plus interspecific variation in the magnitude of these effects, could change decomposition rates and thereby PLSF. Further, CH₄ is implicated in root organogenesis, which would have further effects on the rhizosphere, increasing its breadth and potentially offering more opportunities for root foraging and microbial association (Kou et al. 2018; Zhao et al. 2019). An increased sphere of influence could further increase PSF effects, whether positive or negative.

Ground level ozone (O_3) is an increasing risk to plant health. It forms from NO_x and volatile organic compounds, entering plants through their stomata. There, it will react with the membrane or form reactive oxygen species, which damage cell apparatus (Cho et al. 2011). O_3 is highest on hot sunny days and causes visible injury to the plant. In a timely review, Agathokleous et al. (2020) showed that leaf thickness and other leaf traits can inform vulnerability to O_3 damage, as can plant functional type, with resultant cascading consequences across trophic levels likely. Therefore, the effect of O_3 on PSF could be partly predictable by these criteria. Further, elevated O_3 reduces plant carbon allocation to the soil, which could indirectly affect the soil microbial community. Currently, it is unclear if and how O_3 can directly impact soil microbial communities to influence PSF, and this could form the basis of an important research area. It is clear nevertheless, that GHGs alone and in combination with other drivers can have strong impacts on PSFs and it is urgent that the less studied GHGs are incorporated into PSF experiments.

The role of (micro)nutrients

The role of base cations (e.g., calcium, magnesium and potassium), plant macronutrients (e.g., nitrogen, phosphorus, potassium), plant micronutrients (e.g., iron, manganese and zinc), and important elements such as sulphur and silicon (hereafter collectively plant nutrients) constitute important extrinsic abiotic drivers that have been overlooked in most PSF studies (but see the section on soil sickness). Nitrogen and phosphorus are considered to be the main drivers and often the immediate limiting factors for plant growth (Wang et al. 2018a). Soil nitrogen can change PSF via impacts on the microbial community and soil organic matter (Wang et al. 2022). However, research on individual plant nutrients is increasing, particularly in an agricultural context (Kaspari 2021). For example, necrosis of crop leaves can occur as a result of potassium, sulphur, copper or nitrogen deficiency, which can affect the quality and quantity of carbon exudates and litter (Grundon 2006), likely resulting in alterations to the rhizosphere community composition and changes to PSF.

Micronutrients help determine the quality of the litter and speed of its turnover. There is evidence that micronutrients, when added to soils, can catalyse release of plant-available nitrogen, phosphorus and potassium, which can increase plant and invertebrate abundance (Prather et al. 2020). Measuring local plant nutrients in the rhizosphere to a high resolution has begun to gain momentum (Capstaff et al. 2021; Clode et al. 2009). Some research has linked changes in the plant community with ecosystem scale plant nutrient shifts, although such changes are often not characterised as PSF. For example, woody encroachment in savannah landscapes is associated with an accumulation of copper, sulphur, calcium and strontium in the soil, as well as nitrogen and phosphorus (Zhou et al. 2021). The increase in concentration and availability of these nutrients can create a positive PSF that enables further encroachment (but see D'Angioli et al. (2021)). Nutrient-related PSF is also at play in other regions where shrub encroachment is an issue, namely, in the arctic tundra (Myers-Smith et al. 2011; Wookey et al. 2009).

In order to understand how micronutrients drive PSF, detailed information on plant responses to micronutrients alone and in combination is needed. For example, Wang et al. (2018b) demonstrated a close link between concentrations of plant nutrients in soils and plant tissues on an elevational gradient, finding that soil was a stronger determinant of plant magnesium, calcium, manganese and zinc than elevation, while potassium and iron were decoupled. Further, litter decomposition in a range of forests was found to be tightly correlated with micronutrient concentration. Specifically, in a range of Pinus species, manganese constrains late successional stage litter breakdown because of its importance in the production of lignolytic enzymes by Basidiomycota fungi (Berg et al. 2007), while in tropical tree litter, single micronutrients can have strong effects, either accelerating (zinc; (Powers and Salute 2011); sodium; (Kaspari et al. 2009)) or delaying (magnesium; (Powers and Salute 2011)) decomposition. Shifts in litter decomposition rates can affect PSF by either altering seedling establishment rates through litter thatch or increasing plant growth through rapid release of micronutrients (Veen et al. 2019). Soil pH is also an important determinant of bioavailability of cations and micronutrients, (Wang et al. 2018a). Finally, recent work indicates that there is widespread nutrient dilution in food webs (iron, zinc, manganese, etc.) as a consequence of anthropogenic global change (Kaspari 2021). This is likely to have long-term impacts on plant community dynamics, and it is important to begin to understand the role of heretofore overlooked plant (micro)nutrients in PSF.

Abiotic-biotic extrinsic drivers

Plant-litter-soil feedback (PLSF)

Above- and belowground plant litter impacts various ecosystems processes such as nutrient cycling, carbon sequestration and plant community composition (Xu et al. 2013). Although the importance of litter is recognized as an important extrinsic driver that can have both abiotic and biotic effects on PSF, placing its effects into PSF research has only recently become recognized (Hobbie 2015; Veen et al. 2019). Feedbacks caused by physical, chemical or biotic components of litter could result in broader implications for the dominance of a species within the ecosystem (Capellesso et al. 2016; Mazzoleni et al. 2010).

Physical effects of litter can lead to both negative and positive PLSFs. It is long known that a layer of litter insulates the ground, thereby reducing fluctuations of temperature (MacKinney 1929). Such buffering effects are stronger in grasslands versus forests (Xu et al. 2013), suggesting that the potential feedbacks vary by ecosystem. A thick litter layer can inhibit seeds that require specific wavelengths of light in order to germinate (Vázquez-Yanes et al. 1990), but could also prevent moisture loss under drought (Wellstein 2012). However, whether these effects are seen as negative versus positive remains a question of perspective. For example, suppression of one species could have benefits for another via the alleviation of competitive exclusion (Vaccaro et al. 2009). Further, Wang et al. (2013) found that a layer of pine needle litter impaired atmospheric methane uptake by the soil, but this trend reversed when soil moisture reached a certain threshold (i.e., the soil became a methane sink), suggesting a possible link between litter physio-dynamics and climate change.

Chemical-mediated litter feedback pathways can range from allelochemical influences to nutrient sequestration or release. The section above on allelopathy covers the former, thus here we focus on the latter: nutrient dynamics. As litter decomposes, it can provide valuable nutritional resources for plants (Hobbie 2015). Shifts towards fast-growing herbaceous vegetation that produces labile litter with higher litter nitrogen and lower carbon and lignin content can impact on the soil community, thereby leading to increases in soil nitrogen and generating positive feedbacks (Zhang et al. 2022). In fact, litter traits associated with decomposability (i.e., leaf dry matter and nitrogen content) have been shown to influence landscape-level plant community composition via soil nutrient availability (Laughlin et al. 2015) (see section above on plant functional traits). Typically, shoot litter is more labile than root litter (Freschet et al. 2013), suggesting that root litter might be more likely to generate neutral or negative PLSFs. Belowground biomass allocation tends to be higher compared to aboveground (particularly in harsh ecosystems such as boreal forests and the arctic) (Poorter et al. 2012; Qi et al. 2019), but root decomposition remains underrepresented in the literature.

Biological pathways also play a significant role in shaping the PLSF pathway. The "homefield advantage" (HFA) hypothesis proposes that plant litter will break down faster in "home" versus "away" soils due, in part, to co-evolved specialised decomposer communities in "home" soils (Avres et al. 2009; Veen et al. 2015a). This can result in the release of nutrients, which the plant can rapidly recycle, thereby initiating a positive PSF. Although many studies have found evidence for HFA (Asplund et al. 2018; Milcu and Manning 2011; Vivanco and Austin 2008), others have not (Ayres et al. 2006; Gießelmann et al. 2011; Veen et al. 2015a). On occasion, a reverse HFA has been found, where litter breaks-down more easily away from the plant from which it originated (Ding and Wang 2019). Taken together, it appears that HFA is not a universally applicable theory and the mechanisms behind such effects remain unresolved (Austin et al. 2014; Palozzi and Lindo 2017; Veen et al. 2015b). Theories postulate that other factors outside of the soil microbiome could be driving HFA, such as litter traits (Veen et al. 2015a) and soil abiotic properties. Further, fungal endophytes present in leaves that remain active after senescence can begin to function as saprotrophs after the leaf senesces, which could have implications for decomposition, nutrient release from organic matter and thereby act as one of the mechanisms behind HFA (Fanin et al. 2021; Wolfe and Ballhorn 2020). There is evidence that phyllosphere-derived microbiota can remain dominant in abscised leaves for up to eight months, after which this litter becomes dominantly colonised by soil biota (Voříšková and Baldrian 2013). On the other hand, recent work has also shown that repeated introduction of litter can shape the soil fungal community structure (Veen et al. 2021). What precisely these reciprocal influences on soil and litter microbiomes have for PLSFs is beginning to be understood. For example, it has been shown that litter inoculum with no direct contact with the soil or plant can drive negative feedbacks in multiple Asteraceae species (Zaret et al. 2021). Similarly, inoculation of unsterilised conspecific leaf material had positive feedback effects on Leymus chinensis (Trin.) grass grown under sterile

conditions (Li et al. 2022). Root litter is also postulated to harbour pathogens capable of infecting the next generation of plants (Aldorfová et al. 2022). The enhanced understanding of PLSF demonstrates that litter microbiota have the potential to mediate plant species coexistence via, e.g., species-specific negative feedbacks caused by the phyllosphere microbiome (Whitaker et al. 2017).

Priority effects

Priority effects explain how the order or timing of species arrival affects the establishment, growth or reproduction of subsequent later-arriving species. Priority effects are extrinsic abiotic and/or biotic driver(s) that can add an extra layer of complexity to determining PSF effects. In essence, priority effects relate to PSF in that the first species to arrive will be the soil "conditioning" species and subsequent species will act as the "response" species". Priority effects can play important roles in the assembly of communities of various trophic-levels at different scales (Debray et al. 2022; Fukami 2015; Grainger et al. 2019; Kardol et al. 2013b), such as dictating the order of colonization of microorganisms on plant litter that consequently influences litter degradation rates (Veen et al. 2019) and plant community assembly via sequential soil conditioning (Grman and Suding 2010; van de Voorde et al. 2011; Weidlich et al. 2017).

The investigation of priority effects resulting from soil conditioning by a plant species on the subsequent species growing in that same soil (van de Voorde et al. 2011) is akin to the two phases of a typical PSF experiment - "conditioning" and "feedback" phases. Therefore, priority effects could change plant performance in the succeeding plant community (Grman and Suding 2010; van de Voorde et al. 2011; Weidlich et al. 2017). For example, priority effects caused by legumes that arrived before grasses in an experimental grassland in Germany consistently improved belowground productivity of the successive plant community (Weidlich et al. 2018). Thus, the identification of species or plant functional traits that create positive priority effects that allow for successful establishment of target plants or those that create detrimental priority effects can help us to facilitate restoration (Weidlich et al. 2021; Wolfsdorf et al. 2021) or prioritise the removal of certain plants that generate undesirable soil legacies (Grman and Suding 2010). As such, active management of the plant community composition and their priority effects may be required to achieve desirable soil legacy effects in degraded ecosystems.

The incorporation of priority effects into PSF studies could be expanded by having more than two experimental phases and/or varying the intervals of measurements for each phase (Wubs and Bezemer 2017). Certain plant species may generate highly persistent soil legacies that interact with or override the conditioning effects by subsequent laterarriving species (Hannula et al. 2021). For example, in a long-term mesocosm experiment utilising grassland plants of the Netherlands, Hannula et al. (2021) showed that the influence of an initial plant species on the soil fungal community can be persistent in the endosphere of new plants growing in the soil, which temporarily obscured the soil conditioning effects of these new plants. Future studies should consider the order of plant species' soil conditioning and the persistence of their effects to improve the predictability of PSF (Hannula et al. 2021; van de Voorde et al. 2011).

Plant-soil feedback going by an alias

Soil sickness

After humans began to settle and cultivate crops for subsistence, it quickly became evident that planting the same crop repeatedly in the same soils usually resulted in diminished yields over time. Colloquially, this phenomenon became known as soil sickness. In essence, soil sickness is negative PSF. Soil sickness is widespread within agriculture, with at least 111 species documented as demonstrating negative responses when grown in their "own" soils (Cesarano et al. 2017). Historically, the mechanisms behind soil sickness have been attributed to a combination of nutrient depletion, the build-up of soilborne pathogens and the release of allelopathic compounds from the crops directly or via their residue. In modern agriculture, nutrients are likely to play a secondary role, due to high fertilisation rates (Cesarano et al. 2017; Kumar et al. 2017). Soil pathogens and autotoxic chemicals released from the crop can play important individual and interactive roles in driving soil sickness (Huang et al. 2013). However, emerging evidence suggests soil biota and autotoxicity caused by litter (i.e., crop residues) cannot fully explain soil sickness, with self-DNA (i.e., the DNA of the same species that remains in the environment after decomposition) being proposed as an additional driving mechanism (Cesarano et al. 2017; Mazzoleni et al. 2015a) (see section on self-DNA). Given that the underpinning mechanisms behind soil sickness and PSF are the same, recent calls have emerged to better integrate these concepts (Cesarano et al. 2017; Mariotte et al. 2018). Creating crossdiscipline knowledge exchange offers opportunities to better understand and manage natural and agricultural ecosystems.

Janzen-Connell hypothesis

In the 1970's, Daniël Janzen and Joseph Connell developed, in parallel, a theory to help explain tropical tree diversity distribution (Connell 1971; Janzen 1970). Specifically, they observed that host-specific

herbivores and pathogens make the areas near a parent tree inhospitable for the survival of conspecific seedlings. These herbivores and pathogens were named "distance-responsive predators" if they kill seeds or seedlings near the parent tree or "densitydependent predators" if they kill seeds or seedlings where a particular species was most abundant (typically near the parent tree). These negative distance and density dependence effects are widespread in lowland tropical rainforests (Comita et al. 2010; Harms et al. 2000; Metz et al. 2010). Such effects can prevent any one tree species from dominating the landscape, because if that species is too common, there will be fewer safe places for its seedlings to survive. However, since these natural enemies are host-specific, they will not harm other tree species. As a result, if a species becomes very rare, then more enemy-free areas will become available, giving that species' seedlings a competitive advantage. This negative feedback allows tree species to coexist and provides a partial explanation for the high levels of biodiversity in tropical rainforests (Terborgh 2012; Wright 2002). This mechanism is also at play across different ecosystems (Comita et al. 2014) such as grasslands (Petermann et al. 2008), temperate forests (Deniau et al. 2018; Packer and Clay 2000; Ramage et al. 2017; Seiwa et al. 2019; Yao et al. 2020) and savannahs (Rutten et al. 2016). Conversely, host-specific mutualists can increase the abundance of a particular host species, hence reducing community diversity, in line with the mutualistic effects seen in positive PSF, thereby comprising a "reverse Janzen-Connell effect" (Connell and Lowman 1989; Zahra et al. 2021). The Janzen-Connell hypothesis is essentially a PSF process except in cases when aboveground herbivory is the main driver (Norghauer et al. 2016).

Although the importance of soil-borne pathogens are one mechanism behind the Janzen-Connell hypothesis (Bagchi et al. 2010; Bell et al. 2006; Laliberté et al. 2015), it is not fully known which microorganisms are responsible for this strong negative PSF. Thus far, oomycete pathogens (e.g., *Phytophthora*, *Pythium* (Pringsheim)) are suggested to be important causal agents of negative distance- and density-dependent seedling mortality in species-rich tropical rainforests (Bell et al. 2006; Du et al. 2017; Harms et al. 2000) and temperate forests (Comita et al. 2014; Ramage et al. 2017), but perhaps not in Mediterranean woodlands (Teste and Laliberté 2021). Although oomycete-driven mortality of seedlings in Mediterranean woodland was found regardless of proximity to conspecific trees, short-term seedling recruitment patterns in fire-prone ecosystems did not follow the Janzen-Connell hypothesis, with positive density-dependent survival occurring under conspecific trees (Teste and Laliberté 2021). Instead, the seedlings may have benefited from more nutrient-rich litter under conspecific versus heterospecific trees, have been facilitated under conspecific trees via increased nutrients or have been more negatively affected by generalist soil fungal pathogens found away from conspecific trees.

The role of plant-soil feedback in plant invasion

Soil-driven mechanisms are involved in making plant communities more resistant or vulnerable to non-native plant species invasion, with PSF featuring as the underlying mechanism (Dawson and Schrama 2016; Klironomos 2002; Levine et al. 2006). Soil chemical properties can explain invasions and could be considered a component of PSF (Slesak et al. 2022). Recent reviews point to large variation in the effects non-native invasive species have on soil properties (Lee et al. 2017; Lone et al. 2019; Xu et al. 2022). Native species can (but not always) develop soil microbial communities that suppress the growth of non-native species, while soils modified under non-native species tend to benefit recruitment of their own seedlings (Perkins and Nowak 2013; Reinhart and Callaway 2006). However, relationships between non-natives and soil properties are not straightforward. Studies have explicitly used PSF to demonstrate that invasive plants can escape specific soil pathogens from their native range, leading to increased invasibility in the introduced range (Aldorfová et al. 2021; Reinhart et al. 2003); a phenomenon known as The Enemy Release Hypothesis (Keane and Crawley 2002). However, competition between invaders can interact with soil conditioning to generate idiosyncratic release responses from above- and belowground herbivores (Lucero et al. 2020). This finding puts focus on the importance of including multiple drivers (in this case, competition and herbivory) in order to fully understand PSF effects.

Invasive plants can impact on soil biota and physiochemical properties interactively, leading to PSF effects. Invasive plants can affect soil biota via litter and rhizosphere inputs and, with time, support greater abundances of soil decomposers that stimulate nutrient release (Zhang et al. 2019). As a result of these litter or rhizosphere loops, positive feedbacks can develop, which then can facilitate more invasion (Zhang et al. 2019). However, consideration of other interactions is needed. For example, chemical compounds (e.g., allelopathic) released by invasive plants can promote considerable shifts in soil bacteria, that then impact on nutrient cycling, enzymatic activity, mineralization, soil carbon and nitrogen content (Torres et al. 2021).

It is becoming clearer that the important role of PSF in plant invasion is context- and temporallydependent. For example, the study of Slesak et al. (2022) on Scotch broom (Cytisus scoparius L.) suggests strong context-dependency since the effects of Scotch broom on soil physiochemical properties were more pronounced in less fertile sites. In this case, identifying context-dependency of the effects of plant invasion on PSF can help focus management efforts on removing Scotch broom from lowquality sites first. Other studies have demonstrated the role of biotic interactions in regulating early invasion processes at the root level. For example, native tree species (Quercus acutissima Carruth.) can create a negative PSF against non-native invading plants, likely due to the production of allelopathic litter and a distinct soil microbial community (Chen et al. 2021). These interactions were also noted to be context-dependent, in this case the production of allelopathic compounds depended on the degree of landscape openness. The context of competitive interactions can also affect the end result of invasion. For example, the invasive legume, Lespedeza cuneata (Dum.Cours), benefited from positive PSF effects when interspecific competition was mostly absent, but when root competition from native species was present, PSF effects were neutralized (Crawford and Knight 2017). There is also evidence that positive PSF wanes over time as more generalist soil-borne pathogens colonize the non-native-conditioned soil (Gruntman et al. 2017).

We require more long-term tests or assessments of the ongoing role of PSF (i.e., soil legacy effects; see next section) after non-native plant removal has been done by land managers. Recent work suggests the importance of removing non-native plant invaders rapidly during the establishment phase (Chen et al. 2021; de la Peña et al. 2010; Slesak et al. 2022). The strength, direction, context dependency and temporal stability of invasion-related PSF, as well as the biological, chemical, and physical mechanisms behind them, must be taken into account after non-native plant removal to better inform restoration efforts.

Soil legacy effects

The broad definition of soil legacy considers any event that leaves an influence or "legacy" in the soil that can have subsequent impacts on anything occurring in or on that soil. Drivers of soil legacies can be of anthropogenic or natural origin. The effects they generate can change both abiotic and biotic soil properties and, resultantly, may be directly responsible for PSF or indirectly by initiating alternative plant community developmental trajectories. All soil legacies may not technically be a form of PSF. For example, mine tailings (i.e., the waste soils and substrates of metal and mineral extraction) can leave a legacy of toxic elements (e.g., arsenic, lead, nickel, zinc), extremes in pH and poor water infiltration, creating soils that are ruined for agricultural use and poisonous to a plethora of organisms (Kossoff et al. 2014; Xu et al. 2019). Agricultural practices can leave soil legacies leading to knock-on effects for the plant community and how PSF operates. Nonetheless, in many instances, the term soil legacy is used in place of PSF. Numerous examples exist (Cong et al. 2015; De Long et al. 2019b; Heinen et al. 2020b; Nsikani et al. 2017). Gathering literature on soil legacies and bringing it into the main sphere of PSF studies is essential. Including search terms such as "soil legacy" in meta-analyses on PSF is one way that this can be accomplished.

Soil-associated allelopathy

Allelopathy is a biological phenomenon by which one organism has harmful effects on the growth of surrounding organisms, through the production of secondary metabolites (i.e., allelochemicals) (Whittaker and Feeny 1971). Such effects have been explicitly studied in plants, and their role in PSF is becoming more appreciated (Latif et al. 2017). However, allelopathy clearly has an important role in intergenerational plant performance, which makes it an important component of PSF. Allelopathic effects are usually the direct result of plant chemical impacts (Weston and Duke 2003), but can also be generated directly or interactively via microbes (Cipollini et al. 2012; Inderjit 2005). For example, allelopathic compounds may selectively inhibit certain species of mycorrhizal fungi, with consequences for plant species dominance and overall community composition. Chemical transport via common mycorrhizal hyphal networks allows for systemic defence signalling across plant populations and directed allelochemical delivery to target plants (Achatz and Rillig 2014; Barto et al. 2012; Gorzelak et al. 2015). This process is mainly known from ectomycorrhizal networks where plants can warn each other of herbivore attacks or other potential threats (Delaux 2017; Souto et al. 2000). However, arbuscular mycorrhizal fungi are also able to perform this function and even saprobic or parasitic fungi might have the ability to pass allelochemicals through their hyphae (Barto et al. 2012). In plant-fungus mutualistic interactions, the fungus can benefit if adjacent plants are warned of potential threats and therefore remain healthy, allowing them to continue providing resources for the fungus. In recent years, research on the mechanisms associated with allelopathic interactions has increased exponentially (Zhang et al. 2021). In particular, allelopathic mechanisms play a large role in explaining PSF associated with plant invasiveness (Callaway and Ridenour 2004; Hierro and Callaway 2003; Lankau et al. 2009). Further, the rise of -omics techniques has allowed us to unravel the identity of the secondary metabolites involved in these PSF interactions (Scognamiglio et al. 2015). Allelopathy is one of the mechanisms that may lead to soil sickness (Huang et al. 2013) (see section above for further details), but it can also play a beneficial feedback role in agricultural systems by promoting weed suppression (Birkett et al. 2001; Mariotte et al. 2018). Moreover, certain chemical compounds produced by plant roots such as strigolactones can attract mycorrhizal fungi, but also inhibit stem division and even attract parasitic plants such as broomrape (Orobanche spp. L.) (Bouwmeester et al. 2021). Such chemical compounds could result in PSF if they remain active in the soil over longer periods. Collectively, allelochemicals can play a pivotal role in PSF, even though their effects are not always specifically categorized as such.

Primary and secondary plant community succession

Primary and secondary succession of terrestrial plant communities occurs when plants colonise the land after the formation of a new substrate or a significant disturbance event occurs in a pre-existing ecosystem, respectively. Over time, plant community composition changes, generally as a result of the effects the plants have on themselves and each other, as well as their interactions with the wider environment. Changes to the plant community typically occur in concert with changes in soil abiotic and/or biotic properties (Bardgett and Wardle 2010; Peltzer et al. 2010; Walker et al. 2010). By definition, these concomitant changes in the soil and the plant community fit within the PSF framework (Jiang et al. 2010; Kardol et al. 2013a). Studies investigating PSF dynamics during secondary succession show that negative feedbacks typically occur during early stages, which facilitates species turnover, and neutral or positive feedbacks in later stages that promote plant community stability (Bauer et al. 2015; Jing et al. 2015; Kardol et al. 2006; Koziol and Bever 2015; Kulmatiski et al. 2008). Some examples include studies on secondary succession after agricultural abandonment (Kardol et al. 2006), restoration of tallgrass prairies (Bauer et al. 2015), mine site remediation (Orozco-Aceves et al. 2015) and shifts from a tree- to a shrub-dominated ecosystem following pest-induced tree mortality (Pfennigwerth et al. 2018). The drivers of PSF among most of these examples are linked with soil microorganisms, in particular, mycorrhizal and pathogenic fungi (Bauer et al. 2015; Kardol et al. 2006; Pfennigwerth et al. 2018). Types of secondary succession may vary depending on the ecosystem age or form(s) of disturbance. This can lead to disparities in strength and/or direction of PSF effects (Kardol et al. 2022; van de Voorde et al. 2011) and interactions with other factors (e.g., herbivory, water or nutrient availability, climate) that may override the feedback effects (de la Peña et al. 2016; Dietterich et al. 2019; Heinze and Joshi 2018; Yelenik and Levine 2011).

By contrast, primary succession involves a predictable shift from nitrogen to phosphorus limitation, which may strongly favour plant species of certain nutrient-acquisition strategies at each successional stage (Laliberté et al. 2013; Lambers et al. 2008; Peltzer et al. 2010; Walker et al. 2010). This could hypothetically render PSF more predictable during primary versus secondary succession (Cortois et al. 2016; Kardol et al. 2013a). For example, during early stages of primary succession, negative abiotic PSF, due to soil conditioning by pioneer symbiotic nitrogen-fixing plants, is expected to promote the growth and establishment of non-nitrogen-fixing plants in nitrogen-poor soil (Chapin et al. 1994; Png et al. 2019; Titus and del Moral 1998). Increased N fixation early on in succession can also be induced by invasive species (e.g., Myrica faya Ait.), which could have repercussions for subsequent plant community development (Vitousek et al. 1987). On the other hand, owing to a conspecific negative density dependence mechanism assumed in the Janzen-Connell hypothesis (see section above), negative biotic PSF is expected to become more important in primary late-successional ecosystems, thereby promoting species coexistence (Bever et al. 2015; Kardol et al. 2013a; Laliberté et al. 2015).

However, studies examining PSF of plant communities during early- and late primary succession revealed unpredictable effects. For example, negative abiotic and biotic PSF was found to be important during very early stages of sand dune ecosystem development (van der Putten et al. 1993). Furthermore, Castle et al. (2016), Teste et al. (2017), Png et al. (2019) and Trevenen et al. (2022) showed that negative PSF was not predominant during late primary succession of plant communities in glacial forefield and costal dune ecosystems, and that positive and neutral PSF were also commonly detected. The uncertainties of how the type and direction of feedback effects may present or shift during succession encumbers our ability to utilise PSF as a management tool to predict and steer successional trajectories. Experimental designs should consider the use of fieldconditioned soil for ex situ greenhouse studies or the use of phytometer plants in situ, which can incorporate a greater range of "latent" effects (De Long et al. 2018, 2019a). Overall, most literature (see references mentioned above in this subsection) on plant community succession does not explicitly consider the soil-mediated turnover effects to be PSF, despite that these effects play a pivotal role in shifts in plant community composition. However,



Increasing pressure, influence or progression of a particular driver

Fig. 2 Conceptual diagram showing how different intrinsic (i.e., maternal effects, plant functional traits and self-DNA) and extrinsic (i.e., plant-plant competition, herbivory and soil biota interactions) biotic drivers could be shaping plant-soil feedback (PSF) within the different research areas that are actually PSF but have been called by an alias in the literature. The x-axis shows the increasing pressure, influence or progression of a particular driver, while the y-axis shows the direction of feedback and its increasing or decreasing strength. Certain drivers have the capacity to shift the direction of PSF towards either positive or negative, depending on the context.

the increasing body of literature on PSF and succession demonstrates the benefits of continuing to integrate PSF into understanding soil-mediated turnover effects in plant succession.

Conclusions and future directions

Given the aforementioned knowledge gaps, it is evident that deeper, holistic experiments and metaanalyses are needed in order for substantial gains in PSF research to be realised. We hope that the papers Predictions are made based on the current knowledge body, as reviewed in the text. Relative certainty of the predictions based upon available evidence is indicated by a gradient of certainty, with solid, dashed and dotted lines indicating reasonable certainty, limited certainty and uncertainty, respectively. Note: there are many exceptions to the predictions here and this figure is not meant to be a definitive predictor of how drivers influence PSF under these circumstances. Rather, this figure is meant to provide the best possible predictions based on state of the art knowledge and stimulate further research to test these hypotheses

presented here in this Special Issue on Plant-Soil Feedback (S98) will inspire others in PSF research and tangential fields to build upon these results and suggestions. It is important to point out that the topics covered here are not an exhaustive review of the facets and drivers involved in PSF, but here we covered what we consider to be some of the most pressing future research directions, based on what has been emerging in the literature and basic knowledge about drivers of plant-soil interactions. For example, other phenomenon such as hydraulic lift (Horton and Hart 1998), nurse plant facilitation (Di Bella et al.



Increasing pressure, influence or progression of a particular driver

Fig. 3 Conceptual diagram showing how different extrinsic abiotic drivers (i.e., CO_2 , O_3 , temperature, drought, flooding, nutrients) and extrinsic drivers that can be both abiotic and biotic (i.e., plant-litter-soil feedbacks and priority effects) could be shaping plant-soil feedback (PSF) within the different research areas that are actually PSF but have been called by an alias in the literature. The x-axis shows the increasing pressure, influence or progression of a particular driver, while the y-axis shows the direction of feedback and its increasing or decreasing strength. Certain drivers have the capacity to shift the direction of PSF towards either positive or negative, depend-

2019; Gómez-Aparicio et al. 2004) and resource islands (Garcia et al. 2018) no doubt play roles in PSF, but remain understudied. Fully integrating the drivers and PSF going by an alias that are discussed here is one of the next crucial steps. Making certain predictions as to the outcomes of these interactions is possible in some cases (Figs. 2 and 3), but sufficient knowledge is currently lacking in other cases. Bidirectional relationships also exist between external drivers and PSF (Fig. 4). For example, PSF can affect herbivore performance, (Heinen et al. 2018), which could change herbivore population

ing on the context. Predictions are made based on the current knowledge body, as reviewed in the text. Relative certainty of the predictions based upon available evidence is indicated by a gradient of certainty, with solid, dashed and dotted lines indicating reasonable certainty, limited certainty and uncertainty, respectively. Note: there are many exceptions to the predictions here and this figure is not meant to be a definitive predictor of how drivers influence PSF under these circumstances. Rather, this figure is meant to provide the best possible predictions based on state of the art knowledge and stimulate further research to test these hypotheses

dynamics and, in turn, further influence PSF as the next generation of herbivores consumes plant tissue and generates another round of feedbacks. Floral traits and fecundity can be driven by PSF (Jiao et al. 2021), possibly leading to long-terms shifts in plant population and community composition, resulting eventually in different PSF. Future work needs to consider the continual reciprocal feedback between the extrinsic environment, the intrinsic environment of the plant itself and the soil.

Plant-soil feedback research has been on a developmental trajectory that has sought to include



Fig. 4 Conceptual diagram showing how the extrinsic abiotic (e.g., CO_2 , O_3 , temperature, drought, flooding, nutrients, plant-litter-soil feedbacks and priority effects) and biotic (e.g., plant-plant competition, herbivory, soil biota interactions, plant-litter-soil feedbacks and priority effects) environments, as well as the intrinsic biotic environment (e.g., maternal effects, plant functional traits and self-DNA) affect the soil during the con-

ever-increasing levels of complexity in order to better reflect how PSF operates in natural ecosystems. The first reviews on PSF called for studies outside of the greenhouse (Kulmatiski and Kardol 2008). This led to reviews and opinion pieces that highlighted context-dependency (Smith-Ramesh and Reynolds 2017), proposed frameworks to use PSF (i.e., the soil microbiome) to suppress aboveground insect herbivory (Pineda et al. 2017), encouraged merging fundamental and agricultural PSF research lines (Mariotte et al. 2018) and examined the role of temporal PSF dynamics in controlling plant community diversity (Thakur et al. 2021). Still others have pulled focus on the necessity of selecting the best soil sampling designs to answer specific questions (Allen et al. 2021; Gundale et al. 2019; Reinhart and Rinella 2016; Rinella and Reinhart 2017) and the importance of including environmental influencers such as temperature, soil moisture, aboveground herbivory and belowground interactions (De Long et al. 2018). Progress is being made to explore the influences of such external

ditioning process. During the feedback process, the soil affects the extrinsic and intrinsic environments, resulting in a continual reciprocal feedback between environmental drivers, the soil and the plant. Future work on plant-soil feedback should consider these continual reciprocal feedback relationships. The size of the arrows indicates hypothesized relative strength of effect of a particular environmental driver

drivers, as demonstrated by a recent meta-analysis that showed specific types of competition (e.g., interspecific competition in heterospecific soils) can enhance negative PSF, while specific environmental stressors (i.e., drought) can bolster positive microbial effects (Beals et al. 2020). Another metaanalysis solidified the importance of PSF for plant coexistence, citing microbial communities (i.e., pathogens, mycorrhizae) and phylogenetic distance as some of the primary mechanisms behind pair-wise PSF (Crawford et al. 2019). Additional drivers with strong impacts on PSF will continue to emerge in the literature. For example, soil acidification can drive increases in root biomass and inhibit its subsequent decomposition (Wang et al. 2020), leading to possible consequences not only for soil carbon storage, but shifts in plant community composition. No doubt additional avenues of interest in PSF studies will continue to emerge, highlighting the need for collaborative efforts and knowledge exchange between researchers across the globe.

Ultimately, once substantial knowledge has been amassed, it will become possible to begin modelling landscape-wide patterns (Caruso and Rillig n.d.; Collins et al. 2020; Ke and Levine 2021; Mack et al. 2019). Such modelling could be used to predict when and where PSF is most important, particularly when disturbances (e.g., fire, climate, plant pathogens, anthropogenic) alter ecosystem processes. Next, steering PSF might become possible. Due to the continuous reciprocal process of plants influencing soil abiotic and biotic properties and soil properties influencing plants (Fig. 4), several possible routes exist for steering PSF: (1) Direct transplantation of soil with "healthy" microbiomes, analogous with "faecal transplants" in human guts from healthy individuals (Wubs et al. 2016; Yergeau et al. 2015); (2) Spreading of seeds and plant particles which introduces both the plant community as well as the associated plant microbiome (Elzenga et al. 2019); (3) shifting soil biotic communities by planting plant species that might alter the microbial and soil animal communities via their exudates (Clocchiatti et al. 2021; Vieira et al. 2020); (4) shaping soil communities by introducing litter or amendments (ranging from simple sugars to crop residues to manure) (Clocchiatti et al. 2020; Docherty and Gutknecht 2019; Reardon and Wuest 2016); and 5) adjusting agricultural systems in order to minimize negative PSF (Seipel et al. 2019). Various combinations of the above-listed methods can also be employed (Wubs et al. 2019). Steering of PSF can be a valuable tool in a nature restoration context (Moreno-Mateos et al. 2020; Morriën et al. 2017), facilitate soil nutrient retention (Mariotte et al. 2018) and sequester carbon (Bergmann et al. 2016; Hannula and Morriën 2022). No doubt the years to come will continue to see a massive explosion in PSF research, as numerous outstanding questions remain (Box 1). It is imperative that developing lines of research consider the underexplored, interrelated topics discussed above, as well as those that have been proposed by others, and make a concerted effort to include other areas of research that are actually PSF going by an alias. Essentially a "whole ecosystem approach" should be undertaken. Such an approach will further our understanding of the role PSF plays within ecosystem processes.

Authors' contributions All authors contributed to the initial concept and design of the manuscript. J.R.D.L. and E.L.F. led the writing, with substantial input from all co-authors. All authors contributed to polishing the final version.

Data availability This manuscript contains no data.

Declarations

Competing interests The authors declare no competing interests of any nature.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

References

- Achatz M, Rillig MC (2014) Arbuscular mycorrhizal fungal hyphae enhance transport of the allelochemical juglone in the field. Soil Biol Biochem 78:76–82. https://doi.org/ 10.1016/j.soilbio.2014.07.008
- Agathokleous E, Feng Z, Oksanen E, Sicard P, Wang Q, Saitanis CJ, Araminiene V, Blande JD, Hayes F, Calatayud V, Domingos M, Veresoglou SD, Peñuelas J, Wardle DA, De Marco A, Li Z, Harmens H, Yuan X, Vitale M, Paoletti E (2020) Ozone affects plant, insect, and soil microbial communities: a threat to terrestrial ecosystems and biodiversity. Sci Adv 6:eabc1176. https://doi.org/10. 1126/sciadv.abc1176
- Aldorfová A, Dostálek T, Münzbergová Z (2022) Effects of soil conditioning, root and shoot litter addition interact to determine the intensity of plant–soil feedback. Oikos: e09025. https://doi.org/10.1111/oik.09025
- Aldorfová A, Hanzelková V, Drtinová L, Pánková H, Cajthaml T, Münzbergová Z (2021) Plant-soil interactions of an invasive plant species in its native range help to explain its invasion success elsewhere. https://doi.org/10.21203/ rs.3.rs-788590/v1
- Allen WJ, Sapsford SJ, Dickie IA (2021) Soil sample pooling generates no consistent inference bias: a meta-analysis of 71 plant-soil feedback experiments. New Phytol 231:1308–1315. https://doi.org/10.1111/nph.17455
- Allison SD (2012) A trait-based approach for modelling microbial litter decomposition. Ecol Lett 15:1058–1070. https://doi.org/10.1111/j.1461-0248.2012.01807.x
- Alonso C, Perez R, Bazaga P, Medrano M, Herrera CM (2018) Within-plant variation in seed size and inflorescence fecundity is associated with epigenetic mosaicism in the shrub Lavandula latifolia (Lamiaceae).

Ann Bot 121:153–160. https://doi.org/10.1093/aob/ mcx140

- Asplund J, Kauserud H, Bokhorst S, Lie MH, Ohlson M, Nybakken L (2018) Fungal communities influence decomposition rates of plant litter from two dominant tree species. Fungal Ecol 32:1–8. https://doi.org/10.1016/j.funeco. 2017.11.003
- Austin AT, Vivanco L, González-Arzac A, Pérez LI (2014) There's no place like home? An exploration of the mechanisms behind plant litter–decomposer affinity in terrestrial ecosystems. New Phytol 204:307–314. https://doi. org/10.1111/nph.12959
- Ayres E, Dromph KM, Bardgett RD (2006) Do plant species encourage soil biota that specialise in the rapid decomposition of their litter? Soil Biol Biochem 38:183–186. https://doi.org/10.1016/j.soilbio.2005.04.018
- Ayres E, Steltzer H, Simmons BL, Simpson RT, Steinweg JM, Wallenstein MD, Mellor N, Parton WJ, Moore JC, Wall DH (2009) Home-field advantage accelerates leaf litter decomposition in forests. Soil Biol Biochem 41:606–610. https://doi.org/10.1016/j.soilbio.2008.12.022
- Bagchi R, Swinfield T, Gallery RE, Lewis OT, Gripenberg S, Narayan L, Freckleton RP (2010) Testing the Janzen-Connell mechanism: pathogens cause overcompensating density dependence in a tropical tree. Ecol Lett 13:1262– 1269. https://doi.org/10.1111/j.1461-0248.2010.01520.x
- Barbero F, Guglielmotto M, Capuzzo A, Maffei ME (2016) Extracellular self-DNA (esDNA), but not heterologous plant or insect DNA (etDNA), induces plasma membrane depolarization and calcium signaling in lima bean (Phaseolus lunatus) and maize (Zea mays). Int J Mol Sci 17:1659. https://doi.org/10.3390/ijms17101659
- Bardgett RD, Wardle DA (2010) Aboveground-belowground linkages: biotic interactions, ecosystem processes, and global change. Oxford Series in Ecology and Evolution
- Barto EK, Weidenhamer JD, Cipollini D, Rillig MC (2012) Fungal superhighways: do common mycorrhizal networks enhance below ground communication? Trends Plant Sci 17:633–637. https://doi.org/10.1016/j.tplants. 2012.06.007
- Bauer JT, Mack KML, Bever JD (2015) Plant-soil feedbacks as drivers of succession: evidence from remnant and restored tallgrass prairies. Ecosphere 6:art158. https:// doi.org/10.1890/ES14-00480.1
- Baxendale C, Orwin KH, Poly F, Pommier T, Bardgett RD (2014) Are plant-soil feedback responses explained by plant traits? New Phytol 204:408–423. https://doi.org/10. 1111/nph.12915
- Beals KK, Moore JA, Kivlin SN, Bayliss SL, Lumibao CY, Moorhead LC, Patel M, Summers JL, Ware IM, Bailey JK (2020) Predicting plant-soil feedback in the field: meta-analysis reveals that competition and environmental stress differentially influence PSF. Front Ecol Evol 8:191. https://doi.org/10.3389/fevo.2020.00191
- Beckman NG, Dybzinski R, Tilman D (2022) Short-term plant-soil feedback experiment fails to predict outcome of competition observed in long-term field experiment. Ecology: e3883. https://doi.org/10.1002/ecy.3883
- Bell T, Freckleton RP, Lewis OT (2006) Plant pathogens drive density-dependent seedling mortality in a tropical tree.

Ecol Lett 9:569–574. https://doi.org/10.1111/j.1461-0248.2006.00905.x

- Bennett JA, Klironomos J (2019) Mechanisms of plant-soil feedback: interactions among biotic and abiotic drivers. New Phytol 222:91–96. https://doi.org/10.1111/nph. 15603
- Bennett SI, Howard C, Albrecht R, Smith-Ramesh LM, Reynolds H (2020) Simulated herbivory weakens plant-soil feedbacks in competitive mixtures of native and invasive woodland plants. Front Ecol Evol 7:497. https://doi.org/ 10.3389/fevo.2019.00497
- Berg B, Steffen K, McClaugherty C (2007) Litter decomposition rate is dependent on litter mn concentrations. Biogeochemistry 82:29–39. https://doi.org/10.1007/ s10533-006-9050-6
- Bergmann J, Verbruggen E, Heinze J, Xiang D, Chen BD, Joshi J, Rillig MC (2016) The interplay between soil structure, roots, and microbiota as a determinant of plant-soil feedback. Ecol Evol 6:7633–7644. https:// doi.org/10.1002/ece3.2456
- Bever JD (1994) Feedback between plants and their soil communities in an old field community. Ecology 75:1965– 1977. https://doi.org/10.2307/1941601
- Bever JD (2003) Soil community feedback and the coexistence of competitors: conceptual frameworks and empirical tests. New Phytol 157:465–473. https://doi. org/10.1046/j.1469-8137.2003.00714.x
- Bever JD, Mangan SA, Alexander HM (2015) Maintenance of plant species diversity by pathogens. Annu Rev Ecol Evol Syst 46:305–325. https://doi.org/10.1146/annur ev-ecolsys-112414-054306
- Bezemer TM, van der Putten WH, Martens H, van de Voorde TFJ, Mulder PPJ, Kostenko O (2013) Above- and below-ground herbivory effects on below-ground plantfungus interactions and plant-soil feedback responses. J Ecol 101:325–333. https://doi.org/10.1111/1365-2745. 12045
- Biere A, Goverse A (2016) Plant-mediated systemic interactions between pathogens, parasitic nematodes, and herbivores above-and belowground. Annu Rev Phytopathol 54:499–527. https://doi.org/10.1146/annur ev-phyto-080615-100245
- Birkett MA, Chamberlain K, Hooper AM, Pickett JA (2001) Does allelopathy offer real promise for practical weed management and for explaining rhizosphere interactions involving higher plants? Plant Soil 232:31–39. https:// doi.org/10.1023/A:1010325801256
- Bita CE, Gerats T (2013) Plant tolerance to high temperature in a changing environment: scientific fundamentals and production of heat stress-tolerant crops. Front Plant Sci 4:273. https://doi.org/10.3389/fpls.2013.00273
- Bont Z, Pfander M, Robert CA, Huber M, Poelman EH, Raaijmakers CE, Erb M (2020) Adapted dandelions trade dispersal for germination upon root herbivore attack. Proc R Soc B 287:20192930. https://doi.org/10.1098/rspb.2019. 2930
- Bottone EJ, Nagarsheth N, Chiu K (1998) Evidence of selfinhibition by filamentous fungi accounts for unidirectional hyphal growth in colonies. Can J Microbiol 44:390–393. https://doi.org/10.1139/w98-009

- Bouwmeester H, Li C, Thiombiano B, Rahimi M, Dong L (2021) Adaptation of the parasitic plant lifecycle: germination is controlled by essential host signaling molecules. Plant Physiol 185:1292–1308. https://doi.org/10.1093/ plphys/kiaa066
- Brinkman E, Duyts H, Karssen G, Van der Stoel C, Van der Putten WH (2015) Plant-feeding nematodes in coastal sand dunes: occurrence, host specificity and effects on plant growth. Plant Soil 397:17–30. https://doi.org/10. 1007/s11104-015-2447-z
- Buchenau N, van Kleunen M, Wilschut RA (2022) Direct and legacy-mediated drought effects on plant performance are species-specific and depend on soil community composition. Oikos. https://doi.org/10.1111/oik.08959
- Buerdsell SL, Milligan BG, Lehnhoff EA (2021) Invasive plant benefits a native plant through plant-soil feedback but remains the superior competitor. Neobiota: 119–136. https://doi.org/10.3897/neobiota.64.57746
- Burns RG, DeForest JL, Marxsen J, Sinsabaugh RL, Stromberger ME, Wallenstein MD, Weintraub MN, Zoppini A (2013) Soil enzymes in a changing environment: current knowledge and future directions. Soil Biol Biochem 58:216–234. https://doi.org/10.1016/j.soilbio.2012.11. 009
- Callaway RM, Ridenour WM (2004) Novel weapons: invasive success and the evolution of increased competitive ability. Front Ecol Environ 2:436–443. https://doi.org/10. 1890/1540-9295(2004)002[0436:nwisat]2.0.co;2
- Canadell J, Jackson RB, Ehleringer JR, Mooney HA, Sala OE, Schulze ED (1996) Maximum rooting depth of vegetation types at the global scale. Oecologia 108:583–595. https://doi.org/10.1007/bf00329030
- Canarini A, Kaiser C, Merchant A, Richter A, Wanek W (2019) Root exudation of primary metabolites: mechanisms and their roles in plant responses to environmental stimuli. Front Plant Sci 10:157. https://doi.org/10.3389/ fpls.2019.00157
- Canarini A, Schmidt H, Fuchslueger L, Martin V, Herbold CW, Zezula D, Gündler P, Hasibeder R, Jecmenica M, Bahn M, Richter A (2021) Ecological memory of recurrent drought modifies soil processes via changes in soil microbial community. Nat Commun 12:5308. https://doi. org/10.1038/s41467-021-25675-4
- Capellesso ES, Scrovonski KL, Zanin EM, Hepp LU, Bayer C, Sausen TL (2016) Effects of forest structure on litter production, soil chemical composition and litter-soil interactions. Acta Bot Brasilica 30:329–335. https://doi.org/10. 1590/0102-33062016abb0048
- Capstaff NM, Domoney C, Miller AJ (2021) Real-time monitoring of rhizosphere nitrate fluctuations under crops following defoliation. Plant Methods 17:1–13. https://doi. org/10.1186/s13007-021-00713-w
- Carini P, Marsden PJ, Leff JW, Morgan EE, Strickland MS, Fierer N (2016) Relic DNA is abundant in soil and obscures estimates of soil microbial diversity. Nat Microbiol 2:1–6. https://doi.org/10.1038/nmicrobiol.2016.242
- Carney KM, Hungate BA, Drake BG, Megonigal JP (2007) Altered soil microbial community at elevated CO₂ leads to loss of soil carbon. Proc Natl Acad Sci 104:4990– 4995. https://doi.org/10.1073/pnas.0610045104

- Cartenì F, Bonanomi G, Giannino F, Incerti G, Vincenot CE, Chiusano ML, Mazzoleni S (2016) Self-DNA inhibitory effects: underlying mechanisms and ecological implications. Plant Signal Behav 11:e1158381. https://doi.org/ 10.1080/15592324.2016.1158381
- Caruso T, Rillig MC (n.d.) A general stochastic model shows that plant-soil feedbacks can buffer plant species from extinction risks in unpredictable environments. Plant Soil. https://doi.org/10.1007/s11104-022-05698-6
- Casper BB, Castelli JP (2007) Evaluating plant-soil feedback together with competition in a serpentine grassland. Ecol Lett 10:394–400. https://doi.org/10.1111/j.1461-0248. 2007.01030.x
- Castle SC, Lekberg Y, Affleck D, Cleveland CC (2016) Soil abiotic and biotic controls on plant performance during primary succession in a glacial landscape. J Ecol 104:1555–1565. https://doi.org/10.1111/1365-2745. 12615
- Cesarano G, Zotti M, Antignani V, Marra R, Scala F, Bonanomi G (2017) Soil sickness and negative plant-soil feedback: a reappraisal of hypotheses. J Plant Pathol :545–570. http://www.jstor.org/stable/44687125
- Chapin FS, Walker LR, Fastie CL, Sharman LC (1994) Mechanisms of primary succession following deglaciation at Glacier Bay, Alaska. Ecol Monogr 64:149–175. https://doi.org/10.2307/2937039
- Chen P, Huang Q, Zhuge Y, Li C, Zhu P, Hou Y (2021) The effects of plant-soil feedback on invasion resistance are soil context dependent. Oecologia 197:213–222. https://doi.org/10.1007/s00442-021-05004-8
- Cho K, Tiwari S, Agrawal SB, Torres NL, Agrawal M, Sarkar A, Shibato J, Agrawal GK, Kubo A, Rakwal R (2011) Tropospheric ozone and plants: absorption, responses, and Consequences. In: Whitacre DM (ed) Reviews of environmental contamination and toxicology volume 212. Springer, New York
- Chung YA, Collins SL, Rudgers JA (2019) Connecting plantsoil feedbacks to long-term stability in a desert grassland. Ecology 100. https://doi.org/10.1002/ecy.2756
- Cipollini D, Rigsby CM, Barto EK (2012) Microbes as targets and mediators of allelopathy in plants. J Chem Ecol 38:714–727. https://doi.org/10.1007/ s10886-012-0133-7
- Clocchiatti A, Hannula SE, Hundscheid MP, klein Gunnewiek PJ, de Boer W (2021) Stimulated saprotrophic fungi in arable soil extend their activity to the rhizosphere and root microbiomes of crop seedlings. Environ Microbiol 23:6056–6073. https://doi.org/10.1111/1462-2920.15563
- Clocchiatti A, Hannula SE, van den Berg M, Korthals G, De Boer W (2020) The hidden potential of saprotrophic fungi in arable soil: patterns of short-term stimulation by organic amendments. Appl Soil Ecol 147:103434. https://doi.org/10.1016/j.apsoil.2019.103434
- Clode PL, Kilburn MR, Jones DL, Stockdale EA, Cliff JB III, Herrmann AM, Murphy DV (2009) *In situ* mapping of nutrient uptake in the rhizosphere using nanoscale secondary ion mass spectrometry. Plant Physiol 151:1751– 1757. https://doi.org/10.1104/pp.109.141499
- Collins CD, Bever JD, Hersh MH (2020) Community context for mechanisms of disease dilution: insights from linking epidemiology and plant-soil feedback theory. Ann

N Y Acad Sci 1469:65–85. https://doi.org/10.1111/nyas. 14325

- Comita LS, Muller-Landau HC, Aguilar S, Hubbell SP (2010) Asymmetric density dependence shapes species abundances in a Tropical Tree Community. Science 329:330– 332. https://doi.org/10.1126/science.1190772
- Comita LS, Queenborough SA, Murphy SJ, Eck JL, Xu K, Krishnadas M, Beckman N, Zhu Y (2014) Testing predictions of the Janzen–Connell hypothesis: a metaanalysis of experimental evidence for distance-and density-dependent seed and seedling survival. J Ecol 102:845–856. https://doi.org/10.1111/1365-2745.12232
- Cong W, Ruijven vJ W, vdW D, dGB, Mommer L, Berendse F, Hoffland E (2015) Plant species richness leaves a legacy of enhanced root litter-induced decomposition in soil. Soil Biol Biochem 80:341–348. https://doi.org/10.1016/j. soilbio.2014.10.017
- Connell JH (1971) On the role of natural enemies in preventing competitive exclusion in some marine animals and in rain forest trees. Dyn Popul 298:312
- Connell JH, Lowman MD (1989) Low-diversity tropical rain forests: some possible mechanisms for their existence. Am Nat 134:88–119. https://doi.org/10.1086/284967
- Cortois R, Schröder-Georgi T, Weigelt A, van der Putten WH, De Deyn GB (2016) Plant–soil feedbacks: role of plant functional group and plant traits. J Ecol 104:1608–1617. https://doi.org/10.1111/1365-2745.12643
- Crawford KM, Bauer JT, Comita LS, Eppinga MB, Johnson DJ, Mangan SA, Queenborough SA, Strand AE, Suding KN, Umbanhowar J (2019) When and where plant-soil feedback may promote plant coexistence: a meta-analysis. Ecol Lett 22:1274–1284. https://doi.org/10.1111/ele. 13278
- Crawford KM, Knight TM (2017) Competition overwhelms the positive plant–soil feedback generated by an invasive plant. Oecologia 183:211–220. https://doi.org/10.1007/ s00442-016-3759-2
- Cregger MA, Sanders NJ, Dunn RR, Classen AT (2014) Microbial communities respond to experimental warming, but site matters. Peerj 2:1–13. https://doi.org/10.7717/peerj. 358
- D'Angioli AM, Dantas VL, Lambais M, Meir P, Oliveira RS (2021) No evidence of positive feedback between litter deposition and seedling growth rate in neotropical savannas. Plant Soil. https://doi.org/10.1007/ s11104-021-05163-w
- Dawson W, Schrama M (2016) Identifying the role of soil microbes in plant invasions. J Ecol 104:1211–1218. https://doi.org/10.1111/1365-2745.12619
- de la Peña E, Baeten L, Steel H, Viaene N, De Sutter N, De Schrijver A, Verheyen K (2016) Beyond plant-soil feedbacks: mechanisms driving plant community shifts due to land-use legacies in post-agricultural forests. Funct Ecol 30:1073–1085. https://doi.org/10.1111/1365-2435. 12672
- de la Peña E, de Clercq N, Bonte D, Roiloa S, Rodríguez-Echeverría S, Freitas H (2010) Plant-soil feedback as a mechanism of invasion by *Carpobrotus edulis*. Biol Invasions 12:3637–3648. https://doi.org/10.1007/ s10530-010-9756-1

- De Long JR, Fry EL, Veen GF, Kardol P (2018) Why are plantsoil feedbacks so unpredictable, and what to do about it? Funct Ecol 33:118–128. https://doi.org/10.1111/1365-2435.13232
- De Long JR, Heinen R, Jongen R, Hannula SE, Huberty M, Kielak AM, Steinauer K, Bezemer TM (2021) How plant–soil feedbacks influence the next generation of plants. Ecol Res 36:32–44. https://doi.org/10.1111/1440-1703.12165
- De Long JR, Heinen R, Steinauer K, Hannula SE, Huberty M, Jongen R, Vandenbrande S, Wang M, Zhu F, Bezemer TM (2019a) Taking plant-soil feedbacks to the field in a temperate grassland. Basic Appl Ecol. https://doi.org/10. 1016/j.baae.2019.08.001
- De Long JR, Kardol P, Sundqvist MK, Veen GF, Wardle DA (2015) Plant growth response to direct and indirect temperature effects varies by vegetation type and elevation in a subarctic tundra. Oikos 124:772–783. https://doi.org/ 10.1111/oik.01764
- De Long JR, Semchenko M, Pritchard WJ, Cordero I, Fry EL, Jackson BG, Kurnosova K, Ostle NJ, Johnson D, Baggs EM, Bardgett RD (2019b) Drought soil legacy overrides maternal effects on plant growth. Funct Ecol 33:1400– 1410. https://doi.org/10.1111/1365-2435.13341
- de Souza RSC, Armanhi JSL, Arruda P (2020) From microbiome to traits: designing synthetic microbial communities for improved crop resiliency. Front Plant Sci 11:1179. https://doi.org/10.3389/fpls.2020.01179
- Debray R, Herbert RA, Jaffe AL, Crits-Christoph A, Power ME, Koskella B (2022) Priority effects in microbiome assembly. Nat Rev Microbiol 20:109–121. https://doi.org/10.1038/s41579-021-00604-w
- Delaux PM (2017) Comparative phylogenomics of symbiotic associations. New Phytol 213:89–94. https://doi.org/10. 1111/nph.14161
- Delgado-Baquerizo M, Guerra CA, Cano-Díaz C, Egidi E, Wang J-T, Eisenhauer N, Singh BK, Maestre FT (2020) The proportion of soil-borne pathogens increases with warming at the global scale. Nat Clim Change 10:550– 554. https://doi.org/10.1038/s41558-020-0759-3
- Deniau M, Jung V, Le Lann C, Kellner H, Béchade B, Morra T, Prinzing A (2018) Janzen–Connell patterns can be induced by fungal-driven decomposition and offset by ectomycorrhizal fungi accumulated under a closely related canopy. Funct Ecol 32:785–798. https://doi.org/ 10.1111/1365-2435.13003
- Di Bella CE, Garcia-Parisi PA, Lattanzi FA, Druille M, Schnyder H, Grimoldi AA (2019) Grass to legume facilitation in saline-sodic steppes: influence of vegetation seasonality and root symbionts. Plant Soil 443:509–523. https:// doi.org/10.1007/s11104-019-04247-y
- Dickie I, Sprague R, Peltzer DA, Green J, Orwin K, Sapsford S (2022) Beyond control: applying ecological research to improve long-term outcomes of wilding conifer management. Authorea Preprints. https://doi.org/10.22541/au. 162428605.59992740/v3
- Dickie IA, Bolstridge N, Cooper JA, Peltzer DA (2010) Coinvasion by Pinus and its mycorrhizal fungi. New Phytol 187:475–484. https://doi.org/10.1111/j.1469-8137.2010. 03277.x

- Dietterich LH, Li A, Garvey SM, Casper BB (2019) Aboveground competition and herbivory overpower plant-soil feedback contributions to succession in a remediated grassland. Front Ecol Evol 7:459. https://doi.org/10. 3389/fevo.2019.00459
- Ding S, Wang D (2019) The effects of litter quality and living plants on the home-field advantage of aquatic macrophyte decomposition in a eutrophic urban lake, China. Sci Total Environ 650:1529–1536. https://doi.org/10. 1016/j.scitotenv.2018.09.104
- Docherty KM, Gutknecht JL (2019) Soil microbial restoration strategies for promoting climate-ready prairie ecosystems. Ecol Appl 29:e01858. https://doi.org/10.1002/eap. 1858
- Domínguez-Begines J, Ávila JM, García LV, Gómez-Aparicio L (2020) Soil-borne pathogens as determinants of regeneration patterns at community level in Mediterranean forests. New Phytol 227:588–600. https://doi.org/10.1111/ nph.16467
- Domínguez-Begines J, Ávila JM, García LV, Gómez-Aparicio L (2021) Disentangling the role of oomycete soil pathogens as drivers of plant–soil feedbacks. Ecology 102:e03430. https://doi.org/10.1002/ecy.3430
- Du Y, Queenborough SA, Chen L, Wang Y, Mi X, Ma K, Comita LS (2017) Intraspecific and phylogenetic densitydependent seedling recruitment in a subtropical evergreen forest. Oecologia 184:193–203. https://doi.org/10. 1007/s00442-017-3842-3
- Duell EB, Zaiger K, Bever JD, Wilson GW (2019) Climate affects plant-soil feedback of native and invasive grasses: negative feedbacks in stable but not in variable environments. Front Ecol Evol 7:419. https://doi.org/10.3389/ fevo.2019.00419
- Edwards J, Santos-Medellín C, Nguyen B, Kilmer J, Liechty Z, Veliz E, Ni J, Phillips G, Sundaresan V (2019) Soil domestication by rice cultivation results in plant-soil feedback through shifts in soil microbiota. Genome Biol 20:221. https://doi.org/10.1186/s13059-019-1825-x
- Elzenga J, Bekker R, Pritchard H (2019) Maximising the use of native seeds in restoration projects. Plant Biol (Stuttgart Germany) 21:377. https://doi.org/10.1111/plb.12984
- Eppinga MB, Kaproth MA, Collins AR, Molofsky J (2011) Litter feedbacks, evolutionary change and exotic plant invasion. J Ecol 99:503–514. https://doi.org/10.1111/j.1365-2745.2010.01781.x
- Fanin N, Lin D, Freschet GT, Keiser AD, Augusto L, Wardle DA, Veen G (2021) Home-field advantage of litter decomposition: from the phyllosphere to the soil. New Phytol 231:1353–1358. https://doi.org/10.1111/nph. 17475
- Fernandez CW, Kennedy PG (2016) Revisiting the 'Gadgil effect': do interguild fungal interactions control carbon cycling in forest soils? New Phytol 209:1382–1394. https://doi.org/10.1111/nph.13648
- Fitzpatrick CR, Gehant L, Kotanen PM, Johnson MTJ (2017) Phylogenetic relatedness, phenotypic similarity and plant-soil feedbacks. J Ecol 105:786–800. https://doi.org/ 10.1111/1365-2745.12709
- Freschet GT, Cornwell WK, Wardle DA, Elumeeva TG, Liu WD, Jackson BG, Onipchenko VG, Soudzilovskaia NA, Tao JP, Cornelissen JHC (2013) Linking litter

decomposition of above- and below-ground organs to plant-soil feedbacks worldwide. J Ecol 101:943–952. https://doi.org/10.1111/1365-2745.12092

- Fry EL, De Long JR, Álvarez Garrido L, Alvarez N, Carrillo Y, Castañeda-Gómez L, Chomel M, Dondini M, Drake JE, Hasegawa S, Hortal S, Jackson BG, Jiang M, Lavallee JM, Medlyn BE, Rhymes J, Singh BK, Smith P, Anderson IC, Bardgett RD, Baggs EM, Johnson D (2018a) Using plant, microbe and soil fauna traits to improve the predictive power of biogeochemical models. Methods Ecol Evol 10:146–157. https://doi.org/10.1111/2041-210X.13092
- Fry EL, Johnson GN, Hall AL, Pritchard WJ, Bullock JM, Bardgett RD (2018b) Drought neutralises plant–soil feedback of two mesic grassland forbs. Oecologia 186:1113– 1125. https://doi.org/10.1007/s00442-018-4082-x
- Fuchslueger L, Bahn M, Hasibeder R, Kienzl S, Fritz K, Schmitt M, Watzka M, Richter A (2016) Drought history affects grassland plant and microbial carbon turnover during and after a subsequent drought event. J Ecol 104:1453–1465. https://doi.org/10.1111/1365-2745. 12593
- Fukami T (2015) Historical contingency in Community Assembly: integrating niches, species pools, and priority effects. In: Futuyma DJ (ed) Annual review of ecology, evolution, and systematics, vol 46. Annual Reviews, Palo Alto
- Gao D, Bai E, Li M, Zhao C, Yu K, Hagedorn F (2020) Responses of soil nitrogen and phosphorus cycling to drying and rewetting cycles: a meta-analysis. Soil Biol Biochem 148:107896. https://doi.org/10.1016/j.soilbio. 2020.107896
- Garcia DE, Lopez BR, de-Bashan LE, Hirsch AM, Maymon M, Bashan Y (2018) Functional metabolic diversity of the bacterial community in undisturbed resource island soils in the southern Sonoran Desert. Land Degrad Dev 29:1467–1477. https://doi.org/10.1002/ldr.2905
- Geisen S, Heinen R, Andreou E, van Lent T, ten Hooven FC, Thakur MP (2022) Contrasting effects of soil microbial interactions on growth-defence relationships between early- and mid-successional plant communities. New Phytol 233:1345–1357. https://doi.org/10.1111/nph. 17609
- Germain RM, Gilbert B (2014) Hidden responses to environmental variation: maternal effects reveal species niche dimensions. Ecol Lett 17:662–669. https://doi.org/10. 1111/ele.12267
- Gießelmann UC, Martins KG, Brändle M, Schädler M, Marques R, Brandl R (2011) Lack of home-field advantage in the decomposition of leaf litter in the Atlantic Rainforest of Brazil. Appl Soil Ecol 49:5–10. https://doi. org/10.1016/j.apsoil.2011.07.010
- Gómez-Aparicio L, Zamora R, Gómez JM, Hódar JA, Castro J, Baraza E (2004) Applying plant facilitation to forest restoration: a meta-analysis of the use of shrubs as nurse plants. Ecol Appl 14:1128–1138. https://doi.org/10.1890/03-5084
- Gorzelak MA, Asay AK, Pickles BJ, Simard SW (2015) Interplant communication through mycorrhizal networks mediates complex adaptive behaviour in plant communities. AoB Plants 7. https://doi.org/10.1093/aobpla/plv050

- Grainger TN, Letten AD, Gilbert B, Fukami T (2019) Applying modern coexistence theory to priority effects. Proc Natl Acad Sci 116:6205–6210. https://doi.org/10.1073/ pnas.1803122116
- Gremer JR, Kimball S, Angert AL, Venable DL, Huxman TE (2012) Variation in photosynthetic response to temperature in a guild of winter annual plants. Ecology 93:2693– 2704. https://doi.org/10.1890/12-0006.1
- Grman E, Suding KN (2010) Within-year soil legacies contribute to strong priority effects of exotics on native California grassland communities. Restor Ecol 18:664–670. https://doi.org/10.1111/j.1526-100X.2008.00497.x
- Grundon NJ (2006) Nutrient deficiency and toxicity symptoms. Encyclopedia of soil science, 1144–1146. https://doi.org/ 10.1081/e-ess3
- Gruntman M, Segev U, Glauser G, Tielbörger K (2017) Evolution of plant defences along an invasion chronosequence: defence is lost due to enemy release – but not forever. J Ecol 105:255–264. https://doi.org/10.1111/1365-2745. 12660
- Gundale MJ, Wardle DA, Kardol P, Nilsson MC (2019) Comparison of plant-soil feedback experimental approaches for testing soil biotic interactions among ecosystems. New Phytol 221:577–587. https://doi.org/10.1111/nph. 15367
- Hannula SE, Heinen R, Huberty M, Steinauer K, De Long JR, Jongen R, Bezemer TM (2021) Persistence of plantmediated microbial soil legacy effects in soil and inside roots. Nat Commun 12:1–13. https://doi.org/10.1038/ s41467-021-25971-z
- Hannula SE, Ma Hk, Pérez-Jaramillo JE, Pineda A, Bezemer TM (2020) Structure and ecological function of the soil microbiome affecting plant–soil feedbacks in the presence of a soil-borne pathogen. Environ Microbiol 22:660–676. https://doi.org/10.1111/1462-2920.14882
- Hannula SE, Morriën E (2022) Will fungi solve the carbon dilemma? Geoderma 413:115767. https://doi.org/10. 1016/j.geoderma.2022.115767
- Harms KE, Wright SJ, Calderón O, Hernández A, Herre EA (2000) Pervasive density-dependent recruitment enhances seedling diversity in a tropical forest. Nature 404:493–495. https://doi.org/10.1038/35006630
- Hartman K, Tringe SG (2019) Interactions between plants and soil shaping the root microbiome under abiotic stress. Biochem J 476:2705–2724. https://doi.org/10.1042/ BCJ20180615
- Hassan K, Carrillo Y, Nielsen UN (2021a) The effect of prolonged drought legacies on plant-soil feedbacks. https:// doi.org/10.1111/jvs.13100
- Hassan K, Dastogeer KG, Carrillo Y, Nielsen UN (2022) Climate change driven shifts in plant-soil feedbacks–A meta-analysis. https://doi.org/10.21203/rs.3.rs-1548188/ v1
- Hassan N, Sher K, Rab A, Abdullah I, Zeb U, Naeem I, Shuaib M, Khan H, Khan W, Khan A (2021b) Effects and mechanism of plant litter on grassland ecosystem: a review. Acta Ecol Sin. https://doi.org/10.1016/j.chnaes.2021.02. 006
- Heinen R, Biere A, Bezemer TM (2019) Plant functional group and growth rate interactively shape soil legacy effects on

individual plant-insect interactions. Soil legacy effects on aboveground plant-insect interactions, 83

- Heinen R, Biere A, Bezemer TM (2020a) Plant traits shape soil legacy effects on individual plant–insect interactions. Oikos 129:261–273. https://doi.org/10.1111/oik.06812
- Heinen R, Hannula SE, De Long JR, Huberty M, Jongen R, Kielak A, Steinauer K, Zhu F, Bezemer TM (2020b) Plant community composition steers grassland vegetation via soil legacy effects. Ecol Lett 23:973–982. https://doi. org/10.1111/ele.13497
- Heinen R, van der Sluijs M, Biere A, Harvey JA, Bezemer TM (2018) Plant community composition but not plant traits determine the outcome of soil legacy effects on plants and insects. J Ecol 106:1217–1229. https://doi.org/10. 1111/1365-2745.12907
- Heinze J (2020) Herbivory by aboveground insects impacts plant root morphological traits. Plant Ecol 221:725– 732. https://doi.org/10.1007/s11258-020-01045-w
- Heinze J, Gensch S, Weber E, Joshi J (2017) Soil temperature modifies effects of soil biota on plant growth. J Plant Ecol 10:808–821. https://doi.org/10.1093/jpe/rtw097
- Heinze J, Joshi J (2018) Plant-soil feedback effects can be masked by aboveground herbivory under natural field conditions. Oecologia 186:235–246. https://doi.org/10. 1007/s00442-017-3997-y
- Heinze J, Simons NK, Seibold S, Wacker A, Weithoff G, Gossner MM, Prati D, Bezemer TM, Joshi J (2019) The relative importance of plant-soil feedbacks for plant-species performance increases with decreasing intensity of herbivory. Oecologia 190:651–664. https:// doi.org/10.1007/s00442-019-04442-9
- Heinze J, Sitte M, Schindhelm A, Wright J, Joshi J (2016) Plant-soil feedbacks: a comparative study on the relative importance of soil feedbacks in the greenhouse versus the field. Oecologia 181:559–569. https://doi. org/10.1007/s00442-016-3591-8
- Heinze J, Wacker A, Kulmatiski A (2020) Plant-soil feedback effects altered by aboveground herbivory explain plant species abundance in the landscape. Ecology 101:e03023. https://doi.org/10.1007/ s00442-017-3997-y
- Hendriks M, Visser EJW, Visschers IGS, Aarts BHJ, de Caluwe H, Smit-Tiekstra AE, van der Putten WH, de Kroon H, Mommer L (2015) Root responses of grassland species to spatial heterogeneity of plant-soil feedback. Funct Ecol 29:177–186. https://doi.org/10.1111/1365-2435.12367
- Herman JJ, Sultan SE (2011) Adaptive transgenerational plasticity in plants: case studies, mechanisms, and implications for natural populations. Front Plant Sci 2:1–10. https://doi.org/10.3389/fpls.2011.00102
- Hierro JL, Callaway RM (2003) Allelopathy and exotic plant invasion. Plant Soil 256:29–39. https://doi.org/10. 1023/A:1026208327014
- Hobbie SE (2015) Plant species effects on nutrient cycling: revisiting litter feedbacks. Trends Ecol Evol 30:357–363. https://doi.org/10.1016/j.tree.2015.03.015
- Horton JL, Hart SC (1998) Hydraulic lift: a potentially important ecosystem process. Trends Ecol Evol 13:232–235. https://doi.org/10.1016/S0169-5347(98)01328-7

- Howard MM, Kao-Kniffin J, Kessler A (2020) Shifts in plantmicrobe interactions over community succession and their effects on plant resistance to herbivores. New Phytol 226:1144–1157. https://doi.org/10.1111/nph.16430
- Hu YD, Morota G, Rosa GJM, Gianola D (2015) Prediction of plant height in *Arabidopsis thaliana* using DNA methylation data. Genetics 201:779–793. https://doi.org/10.1534/ genetics.115.177204
- Huang LF, Song LX, Xia XJ, Mao WH, Shi K, Zhou YH, Yu JQ (2013) Plant-soil feedbacks and Soil Sickness: from mechanisms to application in Agriculture. J Chem Ecol 39:232–242. https://doi.org/10.1007/s10886-013-0244-9
- Huberty M, Choi YH, Heinen R, Bezemer TM (2020) Aboveground plant metabolomic responses to plant–soil feedbacks and herbivory. J Ecol 108:1703–1712. https://doi. org/10.1111/1365-2745.13394
- Huberty M, Steinauer K, Heinen R, Jongen R, Hannula SE, Choi YH, Bezemer TM (2022) Temporal changes in plant-soil feedback effects on microbial networks, leaf metabolomics and plant-insect interactions. J Ecol 110:1328–1343. https://doi.org/10.1111/1365-2745. 13872
- Inderjit (2005) Soil microorganisms: an important determinant of allelopathic activity. Plant Soil :227–236. https://www.jstor.org/stable/24129045
- Jackson MB (1985) Ethylene and responses of plants to soil waterlogging and submergence. Annu Rev Plant Physiol 36:145–174
- Janzen DH (1970) Herbivores and the number of tree species in tropical forests. Am Nat 104:501–528. https://doi. org/10.1086/282687
- Jiang L, Han X, Zhang G, Kardol P (2010) The role of plantsoil feedbacks and land-use legacies in restoration of a temperate steppe in northern China. Ecol Res 25:1101– 1111. https://doi.org/10.1007/s11284-010-0735-x
- Jiao L, Sun T, Zhang P, Yang W, Shao D, Zheng S (2021) Effect of floral traits mediated by plant-soil feedback on the relationship between plant density and fecundity: case study of Tamarix chinensis in the Yellow River Delta, China. Global Ecol Conserv 26:e01479. https://doi.org/10.1016/j.gecco.2021.e01479
- Jing J, Bezemer TM, van der Putten WH (2015) Complementarity and selection effects in early and mid-successional plant communities are differentially affected by plant-soil feedback. J Ecol 103:641–647. https://doi. org/10.1111/1365-2745.12388
- Julian JP, Torres R (2006) Hydraulic erosion of cohesive riverbanks. Geomorphology 76:193–206. https://doi.org/ 10.1016/j.geomorph.2005.11.003
- Jung SC, Martinez-Medina A, Lopez-Raez JA, Pozo MJ (2012) Mycorrhiza-induced resistance and priming of plant defenses. J Chem Ecol 38:651–664. https://doi. org/10.1007/s10886-012-0134-6
- Kadowaki K, Yamamoto S, Sato H, Tanabe AS, Hidaka A, Toju H (2018) Mycorrhizal fungi mediate the direction and strength of plant-soil feedbacks differently between arbuscular mycorrhizal and ectomycorrhizal communities. Commun Biol 1. https://doi.org/10.1038/ s42003-018-0201-9
- Kaisermann A, de Vries FT, Griffiths RI, Bardgett RD (2017) Legacy effects of drought on plant-soil feedbacks and

plant-plant interactions. New Phytol 215:1413-1424. https://doi.org/10.1111/nph.14661

- Kardol P, Bezemer TM, van der Putten WH (2006) Temporal variation in plant-soil feedback controls succession. Ecol Lett 9:1080–1088. https://doi.org/10.1111/j.1461-0248.2006.00953.x
- Kardol P, De Deyn GB, Laliberte E, Mariotte P, Hawkes CV (2013a) Biotic plant-soil feedbacks across temporal scales. J Ecol 101:309–315. https://doi.org/10.1111/ 1365-2745.12046
- Kardol P, Souza L, Classen AT (2013b) Resource availability mediates the importance of priority effects in plant community assembly and ecosystem function. Oikos 122:84–94. https://doi.org/10.1111/j.1600-0706.2012. 20546.x
- Kardol P, Yang T, Arroyo DN, Teste FP (2022) Plant-soil feedback in the 'real world': how does fire fit into all of this? Plant Soil. https://doi.org/10.1007/s11104-022-05778-7
- Kaspari M (2021) The invisible hand of the periodic table: how micronutrients shape ecology. Annu Rev Ecol Evol Syst 52:199–219. https://doi.org/10.1146/annurev-ecols ys-012021-090118
- Kaspari M, Yanoviak SP, Dudley R, Yuan M, Clay NA (2009) Sodium shortage as a constraint on the carbon cycle in an inland tropical rainforest. Proc Natl Acad Sci 106: 19405–19409. https://doi.org/10.1073/pnas.0906448106
- Kattge J, Bönisch G, Díaz S, Lavorel S, Prentice IC, Leadley P, Tautenhahn S, Werner GDA, Aakala T, Abedi M, Acosta ATR, Adamidis GC, Adamson K, Aiba M, Albert CH, Alcántara JM, Alcázar C C, Aleixo I, Ali H, Amiaud B, Ammer C, Amoroso MM, Anand M, Anderson C, Anten N, Antos J, Apgaua DMG, Ashman T-L, Asmara DH, Asner GP, Aspinwall M, Atkin O, Aubin I, Baastrup-Spohr L, Bahalkeh K, Bahn M, Baker T, Baker WJ, Bakker JP, Baldocchi D, Baltzer J, Banerjee A, Baranger A, Barlow J, Barneche DR, Baruch Z, Bastianelli D, Battles J, Bauerle W, Bauters M, Bazzato E, Beckmann M, Beeckman H, Beierkuhnlein C, Bekker R, Belfry G, Belluau M, Beloiu M, Benavides R, Benomar L, Berdugo-Lattke ML, Berenguer E, Bergamin R, Bergmann J, Bergmann Carlucci M, Berner L, Bernhardt-Römermann M, Bigler C, Bjorkman AD, Blackman C, Blanco C, Blonder B, Blumenthal D, Bocanegra-González KT, Boeckx P, Bohlman S, Böhning-Gaese K, Boisvert-Marsh L, Bond W, Bond-Lamberty B, Boom A, Boonman CCF, Bordin K, Boughton EH, Boukili V, Bowman DMJS, Bravo S, Brendel MR, Broadley MR, Brown KA, Bruelheide H, Brumnich F, Bruun HH, Bruy D, Buchanan SW, Bucher SF, Buchmann N, Buitenwerf R, Bunker DE, Bürger J, Burrascano S, Burslem DFRP, Butterfield BJ, Byun C, Marques M, Scalon MC, Caccianiga M, Cadotte M, Cailleret M, Camac J, Camarero JJ, Campany C, Campetella G, Campos JA, Cano-Arboleda L, Canullo R, Carbognani M, Carvalho F, Casanoves F, Castagneyrol B, Catford JA, Cavender-Bares J, Cerabolini BEL, Cervellini M, Chacón-Madrigal E, Chapin K, Chapin FS, Chelli S, Chen S-C, Chen A, Cherubini P, Chianucci F, Choat B, Chung K-S, Chytrý M, Ciccarelli D, Coll L, Collins CG, Conti L, Coomes D, Cornelissen JHC, Cornwell WK, Corona P, Coyea M, Craine J, Craven D,

Cromsigt JPGM, Csecserits A, Cufar K, Cuntz M, da Silva AC, Dahlin KM, Dainese M, Dalke I, Dalle Fratte M, Dang-Le AT, Danihelka J, Dannoura M, Dawson S, de Beer AJ, De Frutos A, De Long JR, Dechant B, Delagrange S, Delpierre N, Derroire G, Dias AS, Diaz-Toribio MH, Dimitrakopoulos PG, Dobrowolski M, Doktor D, Dřevojan P, Dong N, Dransfield J, Dressler S, Duarte L, Ducouret E, Dullinger S, Durka W, Duursma R, Dymova O, E-Vojtkó A, Eckstein RL, Ejtehadi H, Elser J, Emilio T, Engemann K, Erfanian MB, Erfmeier A, Esquivel-Muelbert A, Esser G, Estiarte M, Domingues TF, Fagan WF, Fagúndez J, Falster DS, Fan Y, Fang J, Farris E, Fazlioglu F, Feng Y, Fernandez-Mendez F, Ferrara C, Ferreira J, Fidelis A, Finegan B, Firn J, Flowers TJ, Flynn DFB, Fontana V, Forey E, Forgiarini C, François L, Frangipani M, Frank D, Frenette-Dussault C, Freschet GT, Fry EL, Fyllas NM, Mazzochini GG, Gachet S, Gallagher R, Ganade G, Ganga F, García-Palacios P, Gargaglione V, Garnier E, Garrido JL, de Gasper AL, Gea-Izquierdo G, Gibson D, Gillison AN, Giroldo A, Glasenhardt M-C, Gleason S, Gliesch M, Goldberg E, Göldel B, Gonzalez-Akre E, Gonzalez-Andujar JL, González-Melo A, González-Robles A, Graae BJ, Granda E, Graves S, Green WA, Gregor T. Gross N. Guerin GR. Günther A. Gutiérrez AG, Haddock L, Haines A, Hall J, Hambuckers A, Han W, Harrison SP, Hattingh W, Hawes JE, He T, He P, Heberling JM, Helm A, Hempel S, Hentschel J, Hérault B, Hereş A-M, Herz K, Heuertz M, Hickler T, Hietz P, Higuchi P, Hipp AL, Hirons A, Hock M, Hogan JA, Holl K, Honnay O, Hornstein D, Hou E, Hough-Snee N, Hovstad KA, Ichie T, Igić B, Illa E, Isaac M, Ishihara M, Ivanov L, Ivanova L, Iversen CM, Izquierdo J, Jackson RB, Jackson B, Jactel H, Jagodzinski AM, Jandt U, Jansen S, Jenkins T, Jentsch A, Jespersen JRP, Jiang G-F, Johansen JL, Johnson D, Jokela EJ, Joly CA, Jordan GJ, Joseph GS, Junaedi D, Junker RR, Justes E, Kabzems R, Kane J, Kaplan Z, Kattenborn T, Kavelenova L, Kearsley E, Kempel A, Kenzo T, Kerkhoff A, Khalil MI, Kinlock NL, Kissling WD, Kitajima K, Kitzberger T, Kjøller R, Klein T, Kleyer M, Klimešová J, Klipel J, Kloeppel B, Klotz S, Knops JMH, Kohyama T, Koike F, Kollmann J, Komac B, Komatsu K, König C, Kraft NJB, Kramer K, Kreft H, Kühn I,Kumarathunge D, Kuppler J, Kurokawa H, Kurosawa Y, Kuyah S, Laclau J-P, Lafleur B, Lallai E, Lamb E, Lamprecht A, Larkin DJ, Laughlin D, Le Bagousse-Pinguet Y, le Maire G, le Roux PC, le Roux E, Lee T, Lens F, Lewis SL, Lhotsky B, Li Y, Li X, Lichstein JW, Liebergesell M, Lim JY, Lin Y-S, Linares JC, Liu C, Liu D, Liu U, Livingstone S, Llusià J, Lohbeck M, López-García A, Lopez-Gonzalez G, Lososová Z, Louault F, Lukács BA, Lukeš P, Luo Y, Lussu M, Ma S, Maciel Rabelo Pereira C, Mack M, Maire V, MäkeläA, Mäkinen H, Malhado ACM, Mallik A, Manning P, Manzoni S, Marchetti Z, Marchino L, Marcilio-Silva V, Marcon E, Marignani M, Markesteijn L, Martin A, Martínez-Garza C, Martínez-Vilalta J, Mašková T, Mason K, Mason N, Massad TJ, Masse J, Mayrose I, McCarthy J, McCormack ML, McCulloh K, McFadden IR, McGill BJ, McPartland MY, Medeiros JS, Medlyn

B, Meerts P, Mehrabi Z, Meir P, Melo FPL, Mencuccini M, Meredieu C, Messier J, Mészáros I, Metsaranta J, Michaletz ST, Michelaki C, Migalina S, Milla R, Miller JED, Minden V, Ming R, Mokany K, Moles AT, Molnár V A, Molofsky J, Molz M, Montgomery RA, Monty A, Moravcová L, Moreno-Martínez A, Moretti M, Mori AS, Mori S, Morris D, Morrison J, Mucina L, Mueller S, Muir CD, Müller SC, Munoz F, Myers-Smith IH, Myster RW, Nagano M, Naidu S, Narayanan A, Natesan B, Negoita L, Nelson AS, Neuschulz EL, Ni J, Niedrist G, Nieto J, Niinemets Ü, Nolan R, Nottebrock H, Nouvellon Y, Novakovskiy A, The Nutrient N, Nystuen KO, O'Grady A, O'Hara K, O'Reilly-Nugent A, Oakley S, Oberhuber W, Ohtsuka T, Oliveira R, Öllerer K, Olson ME, Onipchenko V, Onoda Y, Onstein RE, Ordonez JC, Osada N, Ostonen I, Ottaviani G, Otto S, Overbeck GE, Ozinga WA, Pahl AT, Paine CET, Pakeman RJ, Papageorgiou AC, Parfionova E, Pärtel M, Patacca M, Paula S, Paule J, Pauli H, Pausas JG, Peco B, Penuelas J, Perea A, Peri PL, Petisco-Souza AC, Petraglia A, Petritan AM, Phillips OL, Pierce S, Pillar VD, Pisek J, Pomogaybin A, Poorter H,Portsmuth A, Poschlod P, Potvin C, Pounds D, Powell AS, Power SA, Prinzing A, Puglielli G, Pyšek P, Raevel V, Rammig A, Ransijn J, Ray CA, Reich PB, Reichstein M, Reid DEB, Réjou-Méchain M, de Dios VR, Ribeiro S, Richardson S, Riibak K, Rillig MC, Riviera F, Robert EMR, Roberts S, Robroek B, Roddy A, Rodrigues AV, Rogers A, Rollinson E, Rolo V, Römermann C, Ronzhina D, Roscher C, Rosell JA, Rosenfield MF, Rossi C, Roy DB, Royer-Tardif S, Rüger N, Ruiz-Peinado R, Rumpf SB, Rusch GM, Ryo M, Sack L, Saldaña A, Salgado-Negret B, Salguero-Gomez R, Santa-Regina I, Santacruz-García AC, Santos J, Sardans J, Schamp B, Scherer-Lorenzen M, Schleuning M, Schmid B, Schmidt M, Schmitt S, Schneider JV, Schowanek SD, Schrader J, Schrodt F, Schuldt B, Schurr F, Selaya Garvizu G, Semchenko M, Seymour C, Sfair JC, Sharpe JM, Sheppard CS, Sheremetiev S, Shiodera S, Shipley B, Shovon TA, Siebenkäs A, Sierra C, Silva V, Silva M, Sitzia T, Sjöman H, Slot M, Smith NG, Sodhi D, Soltis P, Soltis D, Somers B, Sonnier G, Sørensen MV, Sosinski Jr EE, Soudzilovskaia NA, Souza AF, Spasojevic M, Sperandii MG, Stan AB, Stegen J, Steinbauer K, Stephan JG, Sterck F, Stojanovic DB, Strydom T, Suarez ML, Svenning J-C, Svitková I, Svitok M, Svoboda M, Swaine E, Swenson N, Tabarelli M, Takagi K, Tappeiner U, Tarifa R, Tauugourdeau S, Tavsanoglu C, te Beest M, Tedersoo L, Thiffault N, Thom D, Thomas E, Thompson K, Thornton PE, Thuiller W, Tichý L, Tissue D, Tjoelker MG, Tng DYP, Tobias J, Török P, Tarin T, Torres-Ruiz JM, Tóthmérész B, Treurnicht M, Trivellone V, Trolliet F, Trotsiuk V, Tsakalos JL, Tsiripidis I, Tysklind N, Umehara T, Usoltsev V, Vadeboncoeur M, Vaezi J, Valladares F, Vamosi J, van Bodegom PM, van Breugel M, Van Cleemput E, van de Weg M, van der Merwe S, van der Plas F,van der Sande MT, van Kleunen M, Van Meerbeek K, Vanderwel M, Vanselow KA, Vårhammar A, Varone L, Vasquez Valderrama MY, Vassilev K, Vellend M, Veneklaas EJ, Verbeeck H, Verheyen K, Vibrans A, Vieira I, Villacís J, Violle C, Vivek P, Wagner K, Waldram M, Waldron A, Walker AP, Waller M, Walther G, Wang H, Wang F, Wang W, Watkins H, Watkins J, Weber U, Weedon JT, Wei L, Weigelt P, Weiher E, Wells AW, Wellstein C, Wenk E,Westoby M, Westwood A, White PJ, Whitten M, Williams M, Winkler DE, Winter K, Womack C, Wright IJ, Wright SJ, Wright J, Pinho BX, Ximenes F, Yamada T, Yamaji K, Yanai R, Yankov N, Yguel B, Zanini KJ, Zanne AE, Zelený D, Zhao Y-P, Zheng J, Zheng J, Ziemińska K, Zirbel CR, Zizka G, Zo-Bi IC, Zotz G, Wirth C (2020) TRY plant trait database – enhanced coverage and open access. Global Change Biol 26:119–188. https://doi.org/10.1111/gcb.14904

- Ke P-J, Levine JM (2021) The temporal dimension of plantsoil microbe interactions: mechanisms promoting feedback between generations. Am Nat 198:E80–E94. https://doi.org/10.1086/715577
- Ke PJ, Miki T, Ding TS (2015) The soil microbial community predicts the importance of plant traits in plant-soil feedback. New Phytol 206:329–341. https://doi.org/10.1111/ nph.13215
- Keane RM, Crawley MJ (2002) Exotic plant invasions and the enemy release hypothesis. Trends Ecol Evol 17:164–170. https://doi.org/10.1016/s0169-5347(02)02499-0
- Keppler F, Hamilton JTG, McRoberts WC, Vigano I, Braß M, Röckmann T (2008) Methoxyl groups of plant pectin as a precursor of atmospheric methane: evidence from deuterium labelling studies. New Phytol 178:808–814. https:// doi.org/10.1111/j.1469-8137.2008.02411.x
- Klinerova T, Dostal P (2020) Nutrient-demanding species face less negative competition and plant-soil feedback effects in a nutrient-rich environment. New Phytol 225:1343– 1354. https://doi.org/10.1111/nph.16227
- Klironomos JN (2002) Feedback with soil biota contributes to plant rarity and invasiveness in communities. Nature 417:67–70. https://doi.org/10.1038/417067a
- Kos M, Tuijl MAB, de Roo J, Mulder PPJ, Bezemer TM (2015) Species-specific plant-soil feedback effects on aboveground plant-insect interactions. J Ecol 103:904–914. https://doi.org/10.1111/1365-2745.12402
- Kossoff D, Dubbin W, Alfredsson M, Edwards S, Macklin M, Hudson-Edwards KA (2014) Mine tailings dams: characteristics, failure, environmental impacts, and remediation. Appl Geochem 51:229–245. https://doi.org/10. 1016/j.apgeochem.2014.09.010
- Kostenko O, van de Voorde TFJ, Mulder PPJ, van der Putten WH, Bezemer TM (2012) Legacy effects of aboveground–belowground interactions. Ecol Lett 15:813–821. https://doi.org/10.1111/j.1461-0248.2012.01801.x
- Kou N, Xiang Z, Cui W, Li L, Shen W (2018) Hydrogen sulfide acts downstream of methane to induce cucumber adventitious root development. J Plant Physiol 228:113–120. https://doi.org/10.1016/j.jplph.2018.05.010
- Koziol L, Bever JD (2015) Mycorrhizal response trades off with plant growth rate and increases with plant successional status. Ecology 96:1768–1774. https://doi.org/10. 1890/14-2208.1
- Kulmatiski A (2019) Plant-soil feedbacks predict native but not non-native Plant Community Composition: a 7-Year

common-garden experiment. Front Ecol Evol 7. https:// doi.org/10.3389/fevo.2019.00326

- Kulmatiski A, Beard KH, Stevens JR, Cobbold SM (2008) Plant-soil feedbacks: a meta-analytical review. Ecol Lett 11:980–992. https://doi.org/10.1111/j.1461-0248.2008. 01209.x
- Kulmatiski A, Kardol P (2008) Getting plant-soil feedbacks out of the greenhouse: experimental and conceptual approaches. In: Lüttge U, Beyschlag W (eds) Progress in Botany. Springer, Berlin, Heidelberg
- Kumar S, Jakhar SR, Dahiya S, Jangir C, Meena R (2017) Soil sickness and productivity from ecological aspects. J Pharma Phytochem SPI 6:827–831
- Kuťáková E, Cesarz S, Münzbergová Z, Eisenhauer N (2018) Soil microarthropods alter the outcome of plant-soil feedback experiments. Sci Rep 8:1–11. https://doi.org/ 10.1038/s41598-018-30340-w
- Kuzyakov Y, Horwath WR, Dorodnikov M, Blagodatskaya E (2019) Review and synthesis of the effects of elevated atmospheric CO₂ on soil processes: no changes in pools, but increased fluxes and accelerated cycles. Soil Biol Biochem 128:66–78. https://doi.org/10.1016/j. soilbio.2018.10.005
- Laliberté E (2017) Below-ground frontiers in trait-based plant ecology. New Phytol 213:1597–1603. https://doi. org/10.1111/nph.14247
- Laliberté E, Grace JB, Huston MA, Lambers H, Teste FP, Turner BL, Wardle DA (2013) How does pedogenesis drive plant diversity? Trends Ecol Evol 28:331–340. https://doi.org/10.1016/j.tree.2013.02.008
- Laliberté E, Lambers H, Burgess TI, Wright SJ (2015) Phosphorus limitation, soil-borne pathogens and the coexistence of plant species in hyperdiverse forests and shrublands. New Phytol 206:507–521. https://doi.org/ 10.1111/nph.13203
- Lambers H, Raven JA, Shaver GR, Smith SE (2008) Plant nutrient-acquisition strategies change with soil age. Trends Ecol Evol 23:95–103. https://doi.org/10.1016/j. tree.2007.10.008
- Lankau RA, Nuzzo V, Spyreas G, Davis AS (2009) Evolutionary limits ameliorate the negative impact of an invasive plant. Proc Natl Acad Sci USA 106:15362– 15367. https://doi.org/10.1073/pnas.0905446106
- Latif S, Chiapusio G, Weston L (2017) Allelopathy and the role of allelochemicals in plant defence. Advances in Botanical Research. Elsevier, Amsterdam
- Laughlin DC, Richardson SJ, Wright EF, Bellingham PJ (2015) Environmental filtering and positive plant litter feedback simultaneously explain correlations between leaf traits and soil fertility. Ecosystems 18:1269–1280
- Lee MR, Bernhardt ES, van Bodegom PM, Cornelissen JHC, Kattge J, Laughlin DC, Niinemets U, Penuelas J, Reich PB, Yguel B, Wright JP (2017) Invasive species' leaf traits and dissimilarity from natives shape their impact on nitrogen cycling: a meta-analysis. New Phytol 213:128–139. https://doi.org/10.1111/nph.14115
- Lekberg Y, Bever James D, Bunn Rebecca A, Callaway Ragan M, Hart Miranda M, Kivlin Stephanie N, Klironomos J, Larkin Beau G, Maron John L, Reinhart Kurt O, Remke M, Putten Wim H (2018) Relative importance of competition and plant-soil feedback,

their synergy, context dependency and implications for coexistence. Ecol Lett 21:1268–1281. https://doi.org/ 10.1111/ele.13093

- Lemoine NP, Burkepile DE, Parker JD (2014) Variable effects of temperature on insect herbivory. PeerJ 2:e376. https:// doi.org/10.7717/peerj.376
- Levine JM, Pachepsky E, Kendall BE, Yelenik SG, Lambers JHR (2006) Plant-soil feedbacks and invasive spread. Ecol Lett 9:1005–1014. https://doi.org/10.1111/j.1461-0248.2006.00949.x
- Li H, Yang S, Semenov MV, Yao F, Ye J, Bu R, Ma R, Lin J, Kurganova I, Wang X (2021) Temperature sensitivity of SOM decomposition is linked with a K-selected microbial community. Glob Change Biol 27:2763–2779. https://doi.org/10.1111/gcb.15593
- Li L, Wei S, Shen W (2020) The role of methane in plant physiology: a review. Plant Cell Rep 39:171–179. https://doi. org/10.1007/s00299-019-02478-y
- Li Q, Lu X, Wang Y, Huang X, Cox PM, Luo Y (2018) Leaf area index identified as a major source of variability in modeled CO₂ fertilization. Biogeosciences 15:6909– 6925. https://doi.org/10.5194/bg-15-6909-2018
- Li X, Png GK, Sun S, Shi H, Jin K, Li Y (2022) Positive microbial legacy and short-term clonal plasticity aid grazing tolerance of a widespread grass species. Plant Soil 473:291–303. https://doi.org/10.1007/ s11104-021-05281-5
- Lienin P, Kleyer M (2012) Plant trait responses to the environment and effects on ecosystem properties. Basic Appl Ecol 13:301–311. https://doi.org/10.1016/j.baae.2012.05. 002
- Lindahl BD, Kyaschenko J, Varenius K, Clemmensen KE, Dahlberg A, Karltun E, Stendahl J (2021) A group of ectomycorrhizal fungi restricts organic matter accumulation in boreal forest. Ecol Lett 24:1341–1351. https://doi. org/10.1111/ele.13746
- Loladze I (2014) Hidden shift of the ionome of plants exposed to elevated CO_2 depletes minerals at the base of human nutrition. elife 3:e02245. https://doi.org/10.7554/eLife. 02245
- Lone PA, Dar JA, Subashree K, Raha D, Pandey PK, Ray T, Khare PK, Khan ML (2019) Impact of plant invasion on physical, chemical and biological aspects of ecosystems: a review. Trop Plant Res 6:528–544. https://doi.org/10. 22271/tpr.2019.v6.i3.067
- Lucero JE, Arab NM, Meyer ST, Pal RW, Fletcher RA, Nagy DU, Callaway RM, Weisser WW (2020) Escape from natural enemies depends on the enemies, the invader, and competition. Ecol Evol 10:10818–10828. https://doi.org/ 10.1002/ece3.6737
- Maček I, Clark DR, Šibanc N, Moser G, Vodnik D, Müller C, Dumbrell AJ (2019) Impacts of long-term elevated atmospheric CO₂ concentrations on communities of arbuscular mycorrhizal fungi. Mol Ecol 28:3445–3458. https://doi.org/10.1111/mec.15160
- Mack KML, Eppinga MB, Bever JD (2019) Plant-soil feedbacks promote coexistence and resilience in multi-species communities. PLoS ONE 14:e0211572. https://doi. org/10.1371/journal.pone.0211572

- MacKinney A (1929) Effects of forest litter on soil temperature and soil freezing in autumn and winter. Ecology 10:312– 321. https://doi.org/10.2307/1929507
- Manzoni S (2017) Flexible carbon-use efficiency across litter types and during decomposition partly compensates nutrient imbalances—results from analytical stoichiometric models. Front Microbiol 8:661. https://doi.org/10. 3389/fmicb.2017.00661
- Mariotte P, Mehrabi Z, Bezemer TM, De Deyn GB, Kulmatiski A, Drigo B, Veen GF, van der Heijden MGA, Kardol P (2018) Plant–soil feedback: bridging natural and agricultural sciences. Trends Ecol Evol 33:129–142. https://doi. org/10.1016/j.tree.2017.11.005
- Maron JL, Smith AL, Ortega YK, Pearson DE, Callaway RM (2016) Negative plant-soil feedbacks increase with plant abundance, and are unchanged by competition. Ecology 97:2055–2063. https://doi.org/10.1002/ecy.1431
- Martínez-Arias C, Witzell J, Solla A, Martin JA, Rodríguez-Calcerrada J (2022) Beneficial and pathogenic plantmicrobe interactions during flooding stress. Plant Cell Environ 45:2875–2897. https://doi.org/10.1111/pce. 14403
- Martorell C, MartÍnez-Blancas A, García-Meza D (2021) Plant–soil feedbacks depend on drought stress, functional group, and evolutionary relatedness in a semiarid grassland. Ecology :e03499. https://doi.org/10.1002/ecy.3499
- Massey FP, Ennos AR, Hartley SE (2007) Grasses and the resource availability hypothesis: the importance of silicabased defences. J Ecol 95:414–424. https://doi.org/10. 1111/j.1365-2745.2007.01223.x
- Mazzoleni S, Bonanomi G, Giannino F, Incerti G, Dekker SC, Rietkerk M (2010) Modelling the effects of litter decomposition on tree diversity patterns. Ecol Model 221:2784–2792. https://doi.org/10.1016/j.ecolmodel. 2010.08.007
- Mazzoleni S, Bonanomi G, Incerti G, Chiusano ML, Termolino P, Mingo A, Senatore M, Giannino F, Carteni F, Rietkerk M, Lanzotti V (2015a) Inhibitory and toxic effects of extracellular self-DNA in litter: a mechanism for negative plant-soil feedbacks? New Phytol 205:1195–1210. https://doi.org/10.1111/nph.13121
- Mazzoleni S, Carteni F, Bonanomi G, Senatore M, Termolino P, Giannino F, Incerti G, Rietkerk M, Lanzotti V, Chiusano ML (2015b) Inhibitory effects of extracellular self-DNA: a general biological process? New Phytol 206:127–132. https://doi.org/10.1111/nph.13306
- Meisner A, Hol WG, de Boer W, Krumins JA, Wardle DA, van der Putten WH (2014) Plant–soil feedbacks of exotic plant species across life forms: a meta-analysis. Biol Invasions 16:2551–2561. https://doi.org/10.1007/ s10530-014-0685-2
- Meisner A, Jacquiod S, Snoek BL, ten Hooven FC, van der Putten WH (2018) Drought legacy effects on the composition of soil fungal and prokaryote communities. Front Microbiol 9. https://doi.org/10.3389/fmicb.2018.00294
- Messenger DJ, McLeod AR, Fry SC (2009) The role of ultraviolet radiation, photosensitizers, reactive oxygen species and ester groups in mechanisms of methane formation from pectin. Plant Cell Environ 32:1–9. https://doi.org/ 10.1111/j.1365-3040.2008.01892.x

- Metz MR, Sousa WP, Valencia R (2010) Widespread densitydependent seedling mortality promotes species coexistence in a highly diverse amazonian rain forest. Ecology 91:3675–3685. https://doi.org/10.1890/08-2323.1
- Milanović M, Knapp S, Pyšek P, Kühn I (2020) Trait–environment relationships of plant species at different stages of the introduction process. NeoBiota 58:55. https://doi.org/ 10.3897/neobiota.58.51655
- Milcu A, Manning P (2011) All size classes of soil fauna and litter quality control the acceleration of litter decay in its home environment. Oikos 120:1366–1370. https://doi. org/10.1111/j.1600-0706.2010.19418.x
- Miller ZR, Lechon-Alonso P, Allesina S (2022) No robust multispecies coexistence in a canonical model of plant-soil feedbacks. Ecol Lett 25:1690–1698. https://doi.org/10. 1111/ele.14027
- Mohan JE, Cowden CC, Baas P, Dawadi A, Frankson PT, Helmick K, Hughes E, Khan S, Lang A, Machmuller M, Taylor M, Witt CA (2014) Mycorrhizal fungi mediation of terrestrial ecosystem responses to global change: minireview. Fungal Ecol 10:3–19. https://doi.org/10.1016/j. funeco.2014.01.005
- Monticolo F, Palomba E, Termolino P, Chiaiese P, De Alteriis E, Mazzoleni S, Chiusano ML (2020) The role of DNA in the extracellular environment: a focus on NETs, RETs and biofilms. Front Plant Sci: 2045. https://doi.org/10. 3389/fpls.2020.589837
- Moore CE, Meacham-Hensold K, Lemonnier P, Slattery RA, Benjamin C, Bernacchi CJ, Lawson T, Cavanagh AP (2021) The effect of increasing temperature on crop photosynthesis: from enzymes to ecosystems. J Exp Bot 72:2822–2844. https://doi.org/10.1093/jxb/erab090
- Moreno-Mateos D, Alberdi A, Morriën E, van der Putten WH, Rodríguez-Uña A, Montoya D (2020) The long-term restoration of ecosystem complexity. Nat Ecol Evol 4:676– 685. https://doi.org/10.1038/s41559-020-1154-1
- Morriën E, Hannula SE, Snoek LB, Helmsing NR, Zweers H, de Hollander M, Soto RL, Bouffaud M-L, Buée M, Dimmers W, Duyts H, Geisen S, Girlanda M, Griffiths RI, Jørgensen H-B, Jensen J, Plassart P, Redecker D, Schmelz RM, Schmidt O, Thomson BC, Tisserant E, Uroz S, Winding A, Bailey MJ, Bonkowski M, Faber JH, Martin F, Lemanceau P, de Boer W, van Veen JA, van der Putten WH (2017) Soil networks become more connected and take up more carbon as nature restoration progresses. Nat Commun 8:14349. https://doi.org/10.1038/ncomm s14349
- Mutyambai DM, Bass E, Luttermoser T, Poveda K, Midega CAO, Khan ZR, Kessler A (2019) More than "Push" and "Pull"? Plant-soil feedbacks of maize companion cropping increase chemical plant defenses against herbivores. Front Ecol Evol 7. https://doi.org/10.3389/fevo.2019. 00217
- Myers-Smith IH, Forbes BC, Wilmking M, Hallinger M, Lantz T, Blok D, Tape KD, Macias-Fauria M, Sass-Klaassen U, Levesque E, Boudreau S, Ropars P, Hermanutz L, Trant A, Collier LS, Weijers S, Rozema J, Rayback SA, Schmidt NM, Schaepman-Strub G, Wipf S, Rixen C, Menard CB, Venn S, Goetz S, Andreu-Hayles L, Elmendorf S, Ravolainen V, Welker J, Grogan P, Epstein HE, Hik DS (2011) Shrub expansion in tundra ecosystems:

dynamics, impacts and research priorities. Environ Res Lett 6:15. https://doi.org/10.1088/1748-9326/6/4/045509

- Nelson EB (2018) The seed microbiome: Origins, interactions, and impacts. Plant Soil 422:7–34. https://doi.org/10. 1007/s11104-017-3289-7
- Norghauer JM, Free CM, Landis RM, Grogan J, Malcolm JR, Thomas SC (2016) Herbivores limit the population size of big-leaf mahogany trees in an amazonian forest. Oikos 125:137–148. https://doi.org/10.1111/oik.02324
- Nottingham AT, Bååth E, Reischke S, Salinas N, Meir P (2019) Adaptation of soil microbial growth to temperature: using a tropical elevation gradient to predict future changes. Glob Change Biol 25:827–838. https://doi.org/ 10.1111/gcb.14502
- Nsikani MM, Novoa A, Van Wilgen B, Keet JH, Gaertner M (2017) Acacia saligna's soil legacy effects persist longer than ten years after clearing. South Afr J Bot 109:361– 361. https://doi.org/10.1016/j.sajb.2017.01.151
- Nuñez MA, Chiuffo MC, Torres A, Paul T, Dimarco RD, Raal P, Policelli N, Moyano J, García RA, Van Wilgen BW (2017) Ecology and management of invasive Pinaceae around the world: progress and challenges. Biol Invasions 19:3099–3120. https://doi.org/10.1007/ s10530-017-1483-4
- Oduor AMO, Adomako MO, Yuan YG, Li JM (2022) Older populations of the invader Solidago canadensis exhibit stronger positive plant-soil feedbacks and competitive ability in China. Am J Bot 109:1230–1241. https://doi. org/10.1002/ajb2.16034
- Orozco-Aceves M, Standish RJ, Tibbett M (2015) Soil conditioning and plant-soil feedbacks in a modified forest ecosystem are soil-context dependent. Plant Soil 390:183– 194. https://doi.org/10.1007/s11104-015-2390-z
- Orwin KH, Buckland SM, Johnson D, Turner BL, Smart S, Oakley S, Bardgett RD (2010) Linkages of plant traits to soil properties and the functioning of temperate grassland. J Ecol 98:1074–1083. https://doi.org/10.1111/j. 1365-2745.2010.01679.x
- Osakabe Y, Osakabe K, Shinozaki K, Tran L (2014) Response of to water stress. Front Plant Sci 5:1–8. https://doi.org/ 10.3389/fpls.2014.00086
- Packer A, Clay K (2000) Soil pathogens and spatial patterns of seedling mortality in a temperate tree. Nature 404:278– 281. https://doi.org/10.1038/35005072
- Palozzi JE, Lindo Z (2017) Pure and mixed litters of *Sphagnum* and *Carex* exhibit a home-field advantage in Boreal peatlands. Soil Biol Biochem 115:161–168. https://doi.org/ 10.1016/j.soilbio.2017.08.026
- Peltzer DA, Wardle DA, Allison VJ, Baisden WT, Bardgett RD, Chadwick OA, Condron LM, Parfitt RL, Porder S, Richardson SJ, Turner BL, Vitousek PM, Walker J, Walker LR (2010) Understanding ecosystem retrogression. Ecol Monogr 80:509–529. https://doi.org/10.1890/ 09-1552.1
- Penuelas J, Fernández-Martínez M, Vallicrosa H, Maspons J, Zuccarini P, Carnicer J, Sanders TG, Krüger I, Obersteiner M, Janssens IA (2020) Increasing atmospheric CO₂ concentrations correlate with declining nutritional status of european forests. Commun Biology 3:1–11. https://doi.org/10.1038/s42003-020-0839-y

- Perkins LB, Nowak RS (2013) Native and non-native grasses generate common types of plant–soil feedbacks by altering soil nutrients and microbial communities. Oikos 122:199–208. https://doi.org/10.1111/j.1600-0706.2012. 20592.x
- Petermann JS, Fergus AJ, Turnbull LA, Schmid B (2008) Janzen-Connell effects are widespread and strong enough to maintain diversity in grasslands. Ecology 89:2399– 2406. https://doi.org/10.1890/07-2056.1
- Pfennigwerth AA, Van Nuland ME, Bailey JK, Schweitzer JA (2018) Plant-soil feedbacks mediate shrub expansion in declining forests, but only in the right light. J Ecol 106:179–194. https://doi.org/10.1111/1365-2745.12833
- Pineda A, Dicke M, Pieterse CMJ, Pozo MJ (2013) Beneficial microbes in a changing environment: are they always helping plants to deal with insects? Funct Ecol 27:574– 586. https://doi.org/10.1111/1365-2435.12050
- Pineda A, Kaplan I, Bezemer TM (2017) Steering soil microbiomes to suppress aboveground insect pests. Trends Plant Sci 22:770–778. https://doi.org/10.1016/j.tplants. 2017.07.002
- Png GK, Lambers H, Kardol P, Turner BL, Wardle DA, Laliberte E (2019) Biotic and abiotic plant-soil feedback depends on nitrogen-acquisition strategy and shifts during long-term ecosystem development. J Ecol 107:142–153. https://doi.org/10.1111/1365-2745. 13048
- Policelli N, Horton TR, García RA, Naour M, Pauchard A, Nuñez MA (2020) Native and non-native trees can find compatible mycorrhizal partners in each other's dominated areas. Plant Soil 454:285–297. https://doi.org/10. 1007/s11104-020-04609-x
- Poorter H, Niklas KJ, Reich PB, Oleksyn J, Poot P, Mommer L (2012) Biomass allocation to leaves, stems and roots: meta-analyses of interspecific variation and environmental control. New Phytol 193:30–50. https://doi.org/10. 1111/j.1469-8137.2011.03952.x
- Pörtner HO, Roberts DC, Adams H, Adler C, Aldunce P, Ali E, Begum RA, Betts R, Kerr RB, Biesbroek R (2022) Climate change 2022: impacts, adaptation and vulnerability. https://doi.org/10.1017/9781009325844
- Powers JS, Salute S (2011) Macro- and micronutrient effects on decomposition of leaf litter from two tropical tree species: inferences from a short-term laboratory incubation. Plant Soil 346:245–257. https://doi.org/10.1007/ s11104-011-0815-x
- Prather RM, Castillioni K, Kaspari M, Souza L, Prather CM, Reihart RW, Welti EA (2020) Micronutrients enhance macronutrient effects in a meta-analysis of grassland arthropod abundance. Glob Ecol Biogeogr 29:2273– 2288. https://doi.org/10.1111/geb.13196
- Pregitzer K, King J (2005) Effects of soil temperature on nutrient uptake. Nutrient acquisition by plants. Springer, Berlin
- Pugnaire FI, Morillo JA, Peñuelas J, Reich PB, Bardgett RD, Gaxiola A, Wardle DA, Van Der Putten WH (2019) Climate change effects on plant-soil feedbacks and consequences for biodiversity and functioning of terrestrial ecosystems. Sci Adv 5:eaaz1834. https://doi.org/10. 1126/sciadv.aaz1834

- Qi Y, Wei W, Chen C, Chen L (2019) Plant root-shoot biomass allocation over diverse biomes: a global synthesis. Global Ecol Conserv 18:e00606. https://doi.org/10.1016/j.gecco. 2019.e00606
- Ramage BS, Johnson DJ, Gonzalez-Akre E, McShea WJ, Anderson-Teixeira KJ, Bourg NA, Clay K (2017) Sapling growth rates reveal conspecific negative density dependence in a temperate forest. Ecol Evol 7:7661–7671. https://doi.org/10.1002/ece3.3298
- Rasmussen PU, Bennett AE, Tack AJM (2020) The impact of elevated temperature and drought on the ecology and evolution of plant–soil microbe interactions. J Ecol 108:337–352. https://doi.org/10.1111/1365-2745.13292
- Reardon CL, Wuest SB (2016) Soil amendments yield persisting effects on the microbial communities—a 7-year study. Appl Soil Ecol 101:107–116. https://doi.org/10. 1016/j.apsoil.2015.12.013
- Reich PB (2014) The world-wide 'fast-slow' plant economics spectrum: a traits manifesto. J Ecol 102:275–301. https:// doi.org/10.1111/1365-2745.12211
- Reich PB, Hungate BA, Luo Y (2006) Carbon-nitrogen interactions in terrestrial ecosystems in response to rising atmospheric carbon dioxide. Annu Rev Ecol Evol Syst 37:611–636. http://www.jstor.org/stable/30033845
- Reinhart KO, Callaway RM (2006) Soil biota and invasive plants. New Phytol 170:445–457. https://doi.org/10. 1111/j.1469-8137.2006.01715.x
- Reinhart KO, Packer A, Van der Putten WH, Clay K (2003) Plant-soil biota interactions and spatial distribution of black cherry in its native and invasive ranges. Ecol Lett 6:1046–1050. https://doi.org/10.1046/j.1461-0248. 2003.00539.x
- Reinhart KO, Rinella MJ (2016) A common soil handling technique can generate incorrect estimates of soil biota effects on plants. New Phytol 210:786–789. https://doi. org/10.1111/nph.13822
- Rinella MJ, Reinhart KO (2017) Mixing soil samples across experimental units ignores uncertainty and generates incorrect estimates of soil biota effects on plants. New Phytol 216:15–17. https://doi.org/10.1111/nph.14432
- Ristok C, Poeschl Y, Dudenhöffer JH, Ebeling A, Eisenhauer N, Vergara F, Wagg C, van Dam NM, Weinhold A (2019) Plant species richness elicits changes in the metabolome of grassland species via soil biotic legacy. J Ecol 107:2240–2254. https://doi.org/10.1111/1365-2745.13185
- Roach DA, Wulff RD (1987) Maternal effects in plants. Annu Rev Ecol Syst 18:209–235. https://doi.org/10.1146/ annurev.ecolsys.18.1.209
- Romero F, Cazzato S, Walder F, Vogelgsang S, Bender SF, van der Heijden MGA (2022) Humidity and high temperature are important for predicting fungal disease outbreaks worldwide. New Phytol 234:1553–1556. https://doi.org/10.1111/nph.17340
- Rosado-Porto D, Ratering S, Cardinale M, Maisinger C, Moser G, Deppe M, Müller C, Schnell S (2022) Elevated atmospheric CO₂ modifies mostly the metabolic active rhizosphere soil microbiome in the Giessen FACE Experiment. Microb Ecol 83:619–634. https:// link.springer.com/article/10.1007/s00248-021-01791-y

- Rutten G, Gomez-Aparicio L (2018) Plant-soil feedbacks and root responses of two Mediterranean oaks along a precipitation gradient. Plant Soil 424:221–231. https://doi. org/10.1007/s11104-018-3567-z
- Rutten G, Prati D, Hemp A, Fischer M (2016) Plant-soil feedback in East-African savanna trees. Ecology 97:294–301. https://doi.org/10.1890/15-1316.1
- Sapsford S, Wakelin A, Peltzer D, Dickie I (2022) Pine invasion drives loss of soil fungal diversity. Biol Invasions 24:401–414. https://doi.org/10.1007/ s10530-021-02649-7
- Schreiner O, Reed HS (1907) The production of deleterious excretions by roots. Bull Torrey Bot Club 34:279–303
- Scognamiglio M, D'Abrosca B, Esposito A, Fiorentino A (2015) Metabolomics: an unexplored tool for allelopathy studies. J Allelochemical Interact 1:9–23
- Seipel T, Ishaq SL, Menalled FD (2019) Agroecosystem resilience is modified by management system via plant–soil feedbacks. Basic Appl Ecol 39:1–9. https:// doi.org/10.1016/j.baae.2019.06.006
- Seiwa K, Masaka K, Konno M, Iwamoto S (2019) Role of seed size and relative abundance in conspecific negative distance-dependent seedling mortality for eight tree species in a temperate forest. For Ecol Manag 453:117537. https://doi.org/10.1016/j.foreco.2019.117537
- Semchenko M, Barry KE, de Vries FT, Mommer L, Moora M, Maciá-Vicente JG (2022) Deciphering the role of specialist and generalist plant-microbial interactions as drivers of plant-soil feedback. New Phytol 234:1929–1944. https://doi.org/10.1111/nph.18118
- Shade A, Jacques MA, Barrett M (2017) Ecological patterns of seed microbiome diversity, transmission, and assembly. Curr Opin Microbiol 37:15–22. https://doi.org/10.1016/j. mib.2017.03.010
- Shannon SM, Bauer JT, Anderson WE, Reynolds HL (2014) Plant-soil feedbacks between invasive shrubs and native forest understory species lead to shifts in the abundance of mycorrhizal fungi. Plant Soil 382:317–328. https:// doi.org/10.1007/s11104-014-2158-x
- Singh H, Batish DR, Kohli R (1999) Autotoxicity: concept, organisms, and ecological significance. CRC Crit Rev Plant Sci 18:757–772. https://doi.org/10.1080/07352 689991309478
- Slesak RA, Harrington TB, D'Amato AW, Peter DH (2022) Removal of invasive Scotch broom increases its negative effects on soil chemistry and plant communities. Oecologia 198:243–254. https://doi.org/10.1007/ s00442-021-05099-z
- Smith-Ramesh LM, Reynolds HL (2017) The next frontier of plant-soil feedback research: unraveling context dependence across biotic and abiotic gradients. J Veg Sci 28:484–494. https://doi.org/10.1111/jvs.12519
- Souto C, Pellissier F, Chiapusio G (2000) Allelopathic effects of humus phenolics on growth and respiration of mycorrhizal fungi. J Chem Ecol 26:2015–2023. https://doi.org/ 10.1023/A:1005551912405
- Spitzer CM, Lindahl B, Wardle DA, Sundqvist MK, Gundale MJ, Fanin N, Kardol P (2021) Root trait–microbial relationships across tundra plant species. New Phytol 229:1508–1520. https://doi.org/10.1111/nph.16982

- Stotz GC, Gianoli E, Cahill JF (2018) Maternal experience and soil origin influence interactions between resident species and a dominant invasive species. Oecologia 186:247– 257. https://doi.org/10.1007/s00442-017-3996-z
- Suding KN, Stanley Harpole W, Fukami T, Kulmatiski A, MacDougall AS, Stein C, van der Putten WH (2013) Consequences of plant–soil feedbacks in invasion. J Ecol 101:298–308. https://doi.org/10.1111/1365-2745.12057
- Sveen TR, Netherway T, Juhanson J, Oja J, Borgström P, Viketoft M, Strengbom J, Bommarco R, Clemmensen K, Hallin S, Bahram M (2021) Plant-microbe interactions in response to grassland herbivory and nitrogen eutrophication. Soil Biol Biochem 156:108208. https://doi.org/10. 1016/j.soilbio.2021.108208
- Talbot CJ, Bennett EM, Cassell K, Hanes DM, Minor EC, Paerl H, Raymond PA, Vargas R, Vidon PG, Wollheim W (2018) The impact of flooding on aquatic ecosystem services. Biogeochemistry 141:439–461. https://doi.org/ 10.1007/s10533-018-0449-7
- Tedersoo L, Bahram M, Zobel M (2020) How mycorrhizal associations drive plant population and community biology. Science 367:eaba1223. https://doi.org/10.1126/scien ce.aba1223
- Terborgh J (2012) Enemies maintain hyperdiverse tropical forests. Am Nat 179:303–314. https://doi.org/10.1086/ 664183
- Terrer C, Phillips RP, Hungate BA, Rosende J, Pett-Ridge J, Craig ME, van Groenigen KJ, Keenan TF, Sulman BN, Stocker BD, Reich PB, Pellegrini AFA, Pendall E, Zhang H, Evans RD, Carrillo Y, Fisher JB, Van Sundert K, Vicca S, Jackson RB (2021) A trade-off between plant and soil carbon storage under elevated CO2. Nature 591:599–603. https://doi.org/10.1038/ s41586-021-03306-8
- Teste FP, Kardol P, Turner BL, Wardle DA, Zemunik G, Renton M, Laliberté E (2017) Plant-soil feedback and the maintenance of diversity in Mediterranean-climate shrublands. Science 355:173–176. https://doi.org/10. 1126/science.aai8291
- Teste FP, Laliberté E (2021) A test of the Janzen-Connell hypothesis in a species-rich Mediterranean woodland. Ecosphere 12:e03821. https://doi.org/10.1002/ecs2.3821
- Thakur MP, van der Putten WH, Wilschut RA, Veen GF, Kardol P, van Ruijven J, Allan E, Roscher C, van Kleunen M, Bezemer TM (2021) Plant–soil feedbacks and temporal dynamics of plant diversity–productivity relationships. Trends Ecol Evol 36:651–661. https://doi.org/10. 1016/j.tree.2021.03.011
- Titus JH, del Moral R (1998) Seedling establishment in different microsites on Mount St. Helens, Washington, USA. Plant Ecol 134:13–26. https://doi.org/10.1023/A:10097 65502741
- Torres N, Herrera I, Fajardo L, Bustamante RO (2021) Metaanalysis of the impact of plant invasions on soil microbial communities. BMC Ecol Evol 21. https://doi.org/10. 1186/s12862-021-01899-2
- Trevenen EJ, Veneklaas EJ, Teste FP, Dobrowolski Mark P, Mucina L, Renton M (2022) Positive heterospecific interactions can increase long-term diversity of plant communities more than negative conspecific interactions alone.

Funct Ecol 36:159–173. https://doi.org/10.1111/1365-2435.13941

- Trinick M, Parker C (1982) Self-inhibition of rhizobial strains and the influence of cultural conditions on microbial interactions. Soil Biol Biochem 14:79–86. https://doi. org/10.1016/0038-0717(82)90048-7
- Vaccaro LE, Bedford BL, Johnston CA (2009) Litter accumulation promotes dominance of invasive species of cattails (*Typha* spp.) in Lake Ontario wetlands. Wetlands 29:1036–1048. https://doi.org/10.1672/08-28.1
- Valliere JM, Allen EB (2016) Interactive effects of nitrogen deposition and drought-stress on plant-soil feedbacks of *Artemisia californica* seedlings. Plant Soil 403:277–290. https://doi.org/10.1007/s11104-015-2776-y
- van de Voorde TFJ, van der Putten WH, Bezemer TM (2011) Intra- and interspecific plant-soil interactions, soil legacies and priority effects during old-field succession. J Ecol 99:945–953. https://doi.org/10.1111/j.1365-2745. 2011.01815.x
- van der Putten WH, Bardgett RD, Bever JD, Bezemer TM, Casper BB, Fukami T, Kardol P, Klironomos JN, Kulmatiski A, Schweitzer JA, Suding KN, Van de Voorde TFJ, Wardle DA (2013) Plant-soil feedbacks: the past, the present and future challenges. J Ecol 101:265–276. https://doi.org/10.1111/1365-2745.12054
- van der Putten WH, Bradford MA, Brinkman EP, van de Voorde TFJ, Veen GF (2016) Where, when and how plant-soil feedback matters in a changing world. Funct Ecol 30:1109–1121. https://doi.org/10.1111/1365-2435.12657
- van der Putten WH, Vandijk C, Peters BAM (1993) Plantspecific soil-borne diseases contribute to succession in foredune vegetation. Nature 362:53–56. https://doi.org/ 10.1038/362053a0
- van Grunsven RHA, van der Putten WH, Bezemer TM, Veenendaal EM (2010) Plant-soil feedback of native and range-expanding plant species is insensitive to temperature. Oecologia 162:1059–1069. https://doi. org/10.1007/s00442-009-1526-3
- Vázquez-Yanes C, Orozco-Segovia A, Rincon Eet a, Sánchez-Coronado M, Huante P, Toledo J, Barradas V (1990) Light beneath the litter in a tropical forest: effect on seed germination. Ecology 71:1952–1958. https://doi.org/10.2307/1937603
- Veen G, ten Hooven FC, Weser C, Hannula SE (2021) Steering the soil microbiome by repeated litter addition. J Ecol 109:2499–2513. https://doi.org/10.1111/1365-2745.13662
- Veen GF, Freschet GT, Ordonez A, Wardle DA (2015a) Litter quality and environmental controls of homefield advantage effects on litter decomposition. Oikos 124:187–195. https://doi.org/10.1111/oik.01374
- Veen GF, Fry EL, ten Hooven FC, Kardol P, Morriën E, De Long JR (2019) The role of plant litter in driving plantsoil feedbacks. Front Environ Sci 7. https://doi.org/10. 3389/fenvs.2019.00168
- Veen GF, Sundqvist MK, Wardle DA (2015b) Environmental factors and traits that drive plant litter decomposition do not determine home-field advantage effects. Funct Ecol 29:981–991. https://doi.org/10.1111/1365-2435. 12421

- Vieira S, Sikorski J, Dietz S, Herz K, Schrumpf M, Bruelheide H, Scheel D, Friedrich MW, Overmann J (2020) Drivers of the composition of active rhizosphere bacterial communities in temperate grasslands. ISME J 14:463–475. https://doi.org/10.1038/s41396-019-0543-4
- Violle C, Navas M-L, Vile D, Kazakou E, Fortunel C, Hummel I, Garnier E (2007) Let the concept of trait be functional! Oikos 116:882–892. https://doi.org/10.1111/j.0030-1299. 2007.15559.x
- Vitousek PM, Walker LR, Whiteaker LD, Mueller-Dombois D, Matson PA (1987) Biological invasion by *Myrica faya* alters ecosystem development in Hawaii. Science 238:802–804. https://doi.org/10.1126/science.238.4828. 802
- Vivanco L, Austin AT (2008) Tree species identity alters forest litter decomposition through long-term plant and soil interactions in Patagonia, Argentina. J Ecol 96:727–736. https://doi.org/10.1111/j.1365-2745.2008.01393.x
- Voříšková J, Baldrian P (2013) Fungal community on decomposing leaf litter undergoes rapid successional changes. ISME J 7:477–486. https://doi.org/10.1038/ismej.2012. 116
- Wagg C, Boller B, Schneider S, Widmer F, van der Heijden MG (2015) Intraspecific and intergenerational differences in plant-soil feedbacks. Oikos 124:994–1004. https://doi.org/10.1111/oik.01743
- Walker AP, De Kauwe MG, Bastos A, Belmecheri S, Georgiou K, Keeling RF, McMahon SM, Medlyn BE, Moore DJP, Norby RJ, Zaehle S, Anderson-Teixeira KJ, Battipaglia G, Brienen RJW, Cabugao KG, Cailleret M, Campbell E, Canadell JG, Ciais P, Craig ME, Ellsworth DS, Farquhar GD, Fatichi S, Fisher JB, Frank DC, Graven H, Gu L, Haverd V, Heilman K, Heimann M, Hungate BA, Iversen CM, Joos F, Jiang M, Keenan TF, Knauer J, Körner C, Leshyk VO, Leuzinger S, Liu Y, MacBean N, Malhi Y, McVicar TR, Penuelas J, Pongratz J, Powell AS, Riutta T, Sabot MEB, Schleucher J, Sitch S, Smith WK, Sulman B, Taylor B, Terrer C, Torn MS, Treseder KK, Trugman AT, Trumbore SE, van Mantgem PJ, Voelker SL, Whelan ME, Zuidema PA (2021) Integrating the evidence for a terrestrial carbon sink caused by increasing atmospheric CO2. New Phytol 229:2413-2445. https://doi.org/10.1111/nph. 16866
- Walker LR, Wardle DA, Bardgett RD, Clarkson BD (2010) The use of chronosequences in studies of ecological succession and soil development. J Ecol 98:725–736. https:// doi.org/10.1111/j.1365-2745.2010.01664.x
- Walker TW, Kaiser C, Strasser F, Herbold CW, Leblans NI, Woebken D, Janssens IA, Sigurdsson BD, Richter A (2018) Microbial temperature sensitivity and biomass change explain soil carbon loss with warming. Nat Clim Change 8:885–889. https://doi.org/10.1038/ s41558-018-0259-x
- Wang C, Morrissey EM, Mau RL, Hayer M, Piñeiro J, Mack MC, Marks JC, Bell SL, Miller SN, Schwartz E (2021) The temperature sensitivity of soil: microbial biodiversity, growth, and carbon mineralization. ISME J 15:2738–2747. https://doi.org/10.1038/ s41396-021-00959-1

- Wang H-R, Zhang J-M, Zhao X-Y, Feng F (2022) N limit as a switch node between positive and negative plant-soil feedback: a meta-analysis based on the covariant diagnosis of plant growth and soil factors. Ecotoxicol Environ Saf 237:113557. https://doi.org/10.1016/j.ecoenv.2022. 113557
- Wang M, Chen L, Li Y, Chen L, Liu Z, Wang X, Yan P, Qin S (2018a) Responses of soil microbial communities to a short-term application of seaweed fertilizer revealed by deep amplicon sequencing. Appl Soil Ecol 125:288–296. https://doi.org/10.1016/j.apsoil.2018.02.013
- Wang P, Guo J, Xu X, Yan X, Zhang K, Qiu Y, Zhao Q, Huang K, Luo X, Yang F (2020) Soil acidification alters root morphology, increases root biomass but reduces root decomposition in an alpine grassland. Environ Pollut 265:115016. https://doi.org/10.1016/j.envpol.2020. 115016
- Wang R, Wang X, Jiang Y, Cerdà A, Yin J, Liu H, Feng X, Shi Z, Dijkstra FA, Li M-H (2018b) Soil properties determine the elevational patterns of base cations and micronutrients in the plant–soil system up to the upper limits of trees and shrubs. Biogeosciences 15:1763–1774. https://doi.org/10.5194/bg-15-1763-2018
- Wang Y, Wang H, Ma Z, Dai X, Wen X, Liu Y, Wang Z-L (2013) The litter layer acts as a moisture-induced bidirectional buffer for atmospheric methane uptake by soil of a subtropical pine plantation. Soil Biol Biochem 66:45–50. https://doi.org/10.1016/j.soilbio.2013.06.018
- Weidlich EW, Nelson CR, Maron JL, Callaway RM, Delory BM, Temperton VM (2021) Priority effects and ecological restoration. Restor Ecol 29:e13317. https://doi.org/10. 3389/fpls.2016.02008
- Weidlich EWA, von Gillhaussen P, Delory BM, Blossfeld S, Poorter H, Temperton VM (2017) The importance of being First: Exploring Priority and Diversity Effects in a Grassland Field Experiment. Front Plant Sci 7. https:// doi.org/10.3389/fpls.2016.02008
- Weidlich EWA, von Gillhaussen P, Max JFJ, Delory BM, Jablonowski ND, Rascher U, Temperton VM (2018) Priority effects caused by plant order of arrival affect belowground productivity. J Ecol 106:774–780. https://doi.org/ 10.1111/1365-2745.12829
- Wellstein C (2012) Seed–litter–position drives seedling establishment in grassland species under recurrent drought. Plant Biol 14:1006–1010. https://doi.org/10.1111/j.1438-8677.2012.00635.x
- Werger L, Bergmann J, Weber E, Heinze J (2020) Wind intensity affects fine root morphological traits with consequences for plant-soil feedback effects. AoB Plants 12:plaa050. https://doi.org/10.1093/aobpla/plaa050
- Weston LA, Duke SO (2003) Weed and crop allelopathy. CRC Crit Rev Plant Sci 22:367–389. https://doi.org/10.1080/ 713610861
- Whitaker BK, Bauer JT, Bever JD, Clay K (2017) Negative plant-phyllosphere feedbacks in native Asteraceae hosts – a novel extension of the plant-soil feedback framework. Ecol Lett 20:1064–1073. https://doi.org/10.1111/ele. 12805
- Whittaker RH, Feeny PP (1971) Allelochemics: Chemical interactions between species: Chemical agents are of major significance in the adaptation of species and

organization of communities. Science 171:757–770. https://doi.org/10.1126/science.171.3973.757

- Wilschut RA, Geisen S (2021) Nematodes as drivers of plant performance in natural systems. Trends Plant Sci 26:237–247. https://doi.org/10.1016/j.tplants.2020.10. 006
- Wilschut RA, van der Putten WH, Garbeva P, Harkes P, Konings W, Kulkarni P, Martens H, Geisen S (2019) Root traits and belowground herbivores relate to plant-soil feedback variation among congeners. Nat Commun 10:1–9. https://doi.org/10.1038/s41467-019-09615-x
- Wilschut RA, van Kleunen M (2021) Drought alters plant-soil feedback effects on biomass allocation but not on plant performance. Plant Soil 462:285–296. https://doi.org/10. 1007/s11104-021-04861-9
- Wolf JB, Wade MJ (2009) What are maternal effects (and what are they not)? Philos Trans Royal Soc B: Biol Sci 364:1107–1115. https://doi.org/10.1098/rstb.2008.0238
- Wolfe ER, Ballhorn DJ (2020) Do foliar endophytes matter in litter decomposition? Microorganisms 8:446. https://doi. org/10.3390/microorganisms8030446
- Wolfsdorf G, Abrahão A, D'Angioli AM, de Sá Dechoum M, Meirelles ST, Pecoral LFL, Rowland L, da Silveira Verona L, Schmidt IB, Sampaio AB, Oliveira RS (2021) Inoculum origin and soil legacy can shape plant-soil feedback outcomes for tropical grassland restoration. Restor Ecol 29:e13455. https://doi.org/10. 1111/rec.13455
- Wookey PA, Aerts R, Bardgett RD, Baptist F, Brathen KA, Cornelissen JHC, Gough L, Hartley IP, Hopkins DW, Lavorel S, Shaver GR (2009) Ecosystem feedbacks and cascade processes: understanding their role in the responses of Arctic and alpine ecosystems to environmental change. Glob Change Biol 15:1153–1172. https://doi.org/10.1111/j.1365-2486.2008.01801.x
- Wright AJ, de Kroon H, Visser EJW, Buchmann T, Ebeling A, Eisenhauer N, Fischer C, Hildebrandt A, Ravenek J, Roscher C, Weigelt A, Weisser W, Voesenek LACJ, Mommer L (2017) Plants are less negatively affected by flooding when growing in species-rich plant communities. New Phytol 213:645–656. https://doi.org/10. 1111/nph.14185
- Wright JS (2002) Plant diversity in tropical forests: a review of mechanisms of species coexistence. Oecologia 130:1–14. https://doi.org/10.1007/s004420100809
- Wubs E, Van Heusden T, Melchers PD, Bezemer TM (2019) Soil inoculation steers plant-soil feedback, suppressing ruderal plant species. Front Ecol Evol 7:451. https:// doi.org/10.3389/fevo.2019.00451
- Wubs ERJ, Bezemer TM (2017) Temporal carry-over effects in sequential plant-soil feedbacks. Oikos: 220–229. https://doi.org/10.1111/oik.04526
- Wubs ERJ, van der Putten WH, Bosch M, Bezemer TM (2016) Soil inoculation steers restoration of terrestrial ecosystems. Nat Plants 2:1–5. https://doi.org/10.1038/ nplants.2016.107
- Xi N, Adler PB, Chen D, Wu H, Catford JA, van Bodegom PM, Bahn M, Crawford KM, Chu C (2021) Relationships between plant-soil feedbacks and functional traits. J Ecol 109:3411–3423. https://doi.org/10.1111/ 1365-2745.13731

- Xi N, Crawford KM, De Long JR (2022) Plant landscape abundance and soil fungi modulate drought effects on plant-soil feedbacks. Oikos: e08836. https://doi.org/10. 1111/oik.08836
- Xi NX, Bloor JMG, Wang Y, Chu CJ (2019) Contribution of conspecific soil microorganisms to tree seedling light responses: insights from two tropical species with contrasting shade tolerance. Environ Exp Bot 166. https:// doi.org/10.1016/j.envexpbot.2019.103826
- Xu D-M, Zhan C-L, Liu H-X, Lin H-Z (2019) A critical review on environmental implications, recycling strategies, and ecological remediation for mine tailings. Environ Sci Pollut Res 26:35657–35669. https://doi. org/10.1007/s11356-019-06555-3
- Xu HW, Liu Q, Wang SY, Yang GS, Xue S (2022) A global meta-analysis of the impacts of exotic plant species invasion on plant diversity and soil properties. Sci Total Environ 810. https://doi.org/10.1016/j.scitotenv. 2021.152286
- Xu S, Liu L, Sayer EJ (2013) Variability of above-ground litter inputs alters soil physicochemical and biological processes: a meta-analysis of litterfall-manipulation experiments. Biogeosciences 10:7423–7433. https:// doi.org/10.5194/bg-10-7423-2013
- Xue W, Berendse F, Bezemer TM (2018a) Spatial heterogeneity in plant-soil feedbacks alters competitive interactions between two grassland plant species. Funct Ecol 32:2085–2094. https://doi.org/10.1111/1365-2435.13124
- Xue W, Bezemer TM, Berendse F (2018b) Density-dependency and plant-soil feedback: former plant abundance influences competitive interactions between two grassland plant species through plant-soil feedbacks. Plant Soil 428:441–452. https://doi.org/10.1007/ s11104-018-3690-x
- Yao J, Bachelot B, Meng L, Qin J, Zhao X, Zhang C (2020) Abiotic niche partitioning and negative density dependence across multiple life stages in a temperate forest in northeastern China. J Ecol 108:1299–1310. https://doi. org/10.1111/1365-2745.13335
- Yarwood SA (2018) The role of wetland microorganisms in plant-litter decomposition and soil organic matter formation: a critical review. FEMS Microbiol Ecol 94:fiy175. https://doi.org/10.1093/femsec/fiy175
- Yelenik SG, Levine JM (2011) The role of plant-soil feedbacks in driving native-species recovery. Ecology 92:66–74. https://doi.org/10.1890/10-0465.1
- Yergeau E, Bell T, Champagne J, Maynard C, Tardif S, Tremblay J, Greer C (2015) Transplanting soil microbiomes leads to lasting effects on willow growth, but not on the rhizosphere microbiome. Front Microbiol 6. https://doi. org/10.3389/fmicb.2015.01436
- Yu H, Deng Y, He Z, Van Nostrand JD, Wang S, Jin D, Wang A, Wu L, Wang D, Tai X (2018) Elevated CO₂ and warming altered grassland microbial communities in soil top-layers. Front Microbiol 9:1790. https://doi.org/10. 3389/fmicb.2018.01790
- Zahra S, Novotny V, Fayle TM (2021) Do reverse Janzen-Connell effects reduce species diversity? Trends Ecol Evol 36:387–390. https://doi.org/10.1016/j.tree.2021.02.002

- Zaret MM, Bauer JT, Clay K, Whitaker BK (2021) Conspecific leaf litter induces negative feedbacks in Asteraceae seedlings. Ecology 102:e03557. https://doi.org/10.1002/ecy. 3557
- Zhang NL, Van der Putten WH, Veen GF (2016) Effects of root decomposition on plant-soil feedback of early- and midsuccessional plant species. New Phytol 212:220–231. https://doi.org/10.1111/nph.14007
- Zhang P, Li B, Wu J, Hu S (2019) Invasive plants differentially affect soil biota through litter and rhizosphere pathways: a meta-analysis. Ecol Lett 22:200–210. https://doi.org/ 10.1111/ele.13181
- Zhang Y-j, Jin Y-h, Xu J-w, He H-s, Tao Y, Yang Z-p, Zhao J, Diao Y-x, Sun C-h, Li M-H (2022) Responses and feedback of litter properties and soil mesofauna to herbaceous plants expansion into the alpine tundra on Changbai Mountain, China. J Mt Sci 19:403–417. https://doi.org/10.1007/s11629-021-6751-1
- Zhang Z, Liu Y, Yuan L, Weber E, van Kleunen M (2021) Effect of allelopathy on plant performance: a meta-analysis. Ecol Lett 24:348–362. https://doi.org/10.1111/ele. 13627
- Zhao Y, Zhang Y, Liu F, Wang R, Huang L, Shen W (2019) Hydrogen peroxide is involved in methane-induced tomato lateral root formation. Plant Cell Rep 38:377– 389. https://doi.org/10.1007/s00299-019-02372-7
- Zhou W, Chen F, Meng Y, Chandrasekaran U, Luo X, Yang W, Shu K (2020) Plant waterlogging/flooding stress responses: from seed germination to maturation. Plant Physiol Biochem 148:228–236. https://doi.org/10.1016/j. plaphy.2020.01.020
- Zhou Y, Taylor RJ, Boutton TW (2021) Divergent patterns and spatial heterogeneity of soil nutrients in a complex and dynamic savanna landscape. J Geophys Research: Biogeosciences 126:e2021JG006575. https://doi.org/10.1029/ 2021JG006575
- Zhu F, Heinen R, van der Sluijs M, Raaijmakers C, Biere A, Bezemer TM (2018) Species-specific plant-soil feedbacks alter herbivore-induced gene expression and defense chemistry in *Plantago lanceolata*. Oecologia 188:801–811. https://doi.org/10.1007/ s00442-018-4245-9
- Zhu J, Zhu H, Cao Y, Li J, Zhu Q, Yao J, Xu C (2020) Effect of simulated warming on leaf functional traits of urban greening plants. BMC Plant Biol 20:139. https://doi.org/ 10.1186/s12870-020-02359-7

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.