

ACTIVE OPTICAL SENSORS TO DEVELOP NITROGEN FERTILIZER RECOMMENDATIONS FOR POTATO CROP

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ABSTRACT

This study was performed to determine whether active optical sensors could develop an algorithm for N recommendation for the potato crop (*Solanum tuberosum* L.). The experiment was conducted in Maine State, (USA) during the growing season of 2018-2019. Six N rates (0-280 kg ha⁻¹) were applied on eleven locations under a randomized complete block design (RCBD), with four replications. Data of normalized difference vegetation index (NDVI) were collected via active sensors, GreenSeeker-(GS), and Crop Circle-(CC). Sensors measurements collected at the 20th of the leaf stage were significantly associated with tuber yield, where the exponential model exhibited a better fit for the regression curve. Conventionally, 168 kg N ha⁻¹ produced the maximum potato yield. The N rate computed based on in-season sensors reading reduced by about 12-14% from the total N rate that growers use to apply based on the conventional approach. Studying potato cultivars separately in the same soil properties can improve the algorithm accurately.

Keywords: nitrogen, greenseeker, crop circle, in-season estimated yield

زعين وآخرون

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توصية السماد النيتروجيني لمحصول البطاطا باستخدام أجهزة الاستشعار البصرية النشطة

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المستخلص

إجريت هذا البحث لتحديد ما إذا كانت المستشعرات الضوئية النشطة يمكن أن تولد خوارزمية لتوصية النيتروجين لمحصول البطاطا (*Solanum tuberosum* L.). نفذت التجربة في ولاية مين في الولايات المتحدة الأمريكية للموسم الزراعي 2018-2019. تم تطبيق ستة معدلات نيتروجين (0-280 كغم هكتار⁻¹) على أحد عشر موقعًا بتصميم القطاعات العشوائية الكاملة، مع أربعة مكررات. تم الحصول على بيانات فهرس الغطاء النباتي (NDVI) المعياري بواسطة المستشعرات الضوئية النشطة، GreenSeeker و Crop Circle. ارتبطت قياسات المستشعرات التي تم الحصول عليها في المرحلة العشرين (عدد اوراق) ارتباطًا وثيقًا بإنتاجية الدرنات، حيث أظهر النموذج الأسّي ملائمة أفضل لمنحنى الانحدار. تقليديًا ، أنتج المعدل 168 كغم N هكتار⁻¹ أقصى إنتاجية من البطاطا. معدل النيتروجين المحسوب على أساس قراءة أجهزة الاستشعار في الموسم انخفض بنسبة 12-14% تقريبًا من إجمالي معدل النيتروجين الذي استخدمه المزارعون لتطبيقه بناءً على النهج التقليدي. يمكن أن تؤدي دراسة أصناف البطاطا بشكل منفصل في خصائص التربة المماثلة إلى تحسين الخوارزمية بدقة.

الكلمات المفتاحية: المتحسس كرين سيكير، كروب سيركل، الانتاجية المحسوب في الحقل.

INTRODUCTION

It has been a fact that unreasonable application of N fertilizer to the potato crop causes low tuber production due to excessive vegetative growth, lower tuber quality (low specific gravity, large size with hollow heart, delay in maturity, etc.), and lower N use efficiency (NUE) that causes the leaching of a large part of N to groundwater and leads to a high risk of environmental contamination (of the atmosphere by nitrous oxides and water by nitrate, etc.) (6, 17, 51). N deficiency, in contrast, can considerably decrease crop yield (68). Furthermore, the potato production system is well known for low NUE, varying between 50 and 60% (14,55), and this could be due to shallow and poorly developed root systems. Typically, loss of N occurs when mineral N (NH_4^+ and NO_3^-) is present in the soil, in amounts higher than plant requirements (28). Consequently, inadequate synchronization between soil N supply and crop demand is one of the main reasons for low N fertilizer use efficiency (3,4,14,41,38). Potato growers in developed countries are under immense pressure to keep profitability against new environmental restrictions, such as the commitment to the nitrate directive (91/676/EEC) and a recent increment in N fertilizer prices, to motivate them for precise input management. Nevertheless, having adequate food supplies globally is a challenge that fertilizer application cannot be achieved (56). In such a meaning, it is essential to develop instruments and procedures for potato growers that could help them determine “the right N fertilizer rate at the right time and place.” It is generally acknowledged that a temporary field-specific N recommendation for potato at planting time can never be accurate. Furthermore, it is challenging to predict crop N requirements during the growing season (61) due to numerous predictable or unpredictable factors, such as chemical, physical, and biological soil characteristics, soil organic matter, cultural practices, crop maturity time, and weather conditions. Nitrogen fertilization recommendation with estimated requirements during crop growing seasons can essentially aid in matching crop N requirement times and rates with supplies. Accordingly, N fertilizer

efficiency can be improved (5,61). Precision agriculture technology allows growers to apply the correct quantity of fertilizer in real-time based on the crop’s current growth status without negatively affecting the final yield. A modeling strategy (N recommendation at field-specific scale) of crop N status monitoring can lead to helpful decision-support methods to enhance N fertilizer use efficiency. It has been found that the approach of using crop N status assessment to determine crop demands is more reliable than predicting the available soil N supply (48). Plants are often considered a good indicator (mirror) of growing conditions (45). Most of the available crop monitoring techniques depend on the magnitudes of reflected light above the crop canopy (49). A remote sensing approach can be performed at several spatial scales: ground-based, airborne, or space-borne (31,57). All these scales focus on measuring plant canopy formation factors, such as the leaf area index (LAI) and leaf chlorophyll, among others, with well-established science that these factors are strongly related to each other and plant N status (47). The most common precision agriculture tools used for grain crops, such as corn (*Zea mays* L.), wheat (*Triticum aestivum* L.), and sunflower (*Helianthus annuus* L.), among others, are ground-based active optical sensors such as GS, CropScan, N-sensor, and Holland CC (12). The GS and CC are the most prevalent ground-based sensors in North America for research and commercial use. GreenSeeker (GS) has been widely used for developing N recommendations (46), and with which an algorithm for wheat crop increased nitrogen use efficiency by more than 15% (23). In another study in Oklahoma, the coefficient of variation (CV) from NDVI data was used to evaluate plant density in wheat (11). Similar techniques were used in wheat and rice (*Oryza sativa* L.) grown in Northwest India and attained higher NUE than conventional methods (20,47). The CV was further used to adjust the algorithm in wheat (19). Another in-season N uptake was developed for rice, which increased the NUE and yields (9). Crop characteristics have been used in various methods to calculate optimum N requirements (20). Several other research

studies have used plant biomass (11,19) and plant N content (9, 10) to determine N requirements. Spectral measurements have also been used to assess yield potential (YPO) (43,54). Yield potential is a function of the growing environment (28) and is essential for fertilizer N calculation methods. The YPO has been predicted in-season utilizing optical sensors (49). In addition, the NDVI has been used to determine in-season estimated yield (INSEY) (49), which is a measurement of biomass produced per day as NDVI; (Large, 1954) (34) divided by the number of growing degree days (GDD). A few studies have utilized active optical sensors to predict leaf N content (8,25) and yield (47) studies developed an algorithm for N recommendations for potato crops. Therefore, this study aimed to use the data of active optical sensors (NDVI) to develop N recommendation and compare it with what potato growers commonly applied in Maine, USA.

MATERIALS AND METHODS

Research locations: The experimentation was performed in Aroostook County, Maine, USA (Figure1), in the period of 2018 and 2019. Eleven investigation locations, six of them in Presque Isle, Aroostook Farm (AF1, Lat.46.66134° and Long.-68.01808°), New Sweden-1 (NS-1, Lat.46.95156° and Long.-68.14779°), Frenchville (FV, Lat. 47.21676° and Long. -68.41153°), and New Sweden-2 (NS-2, Lat.46.95271° and Long.-68.14572°), Caribou (CA1, Lat. 46.88227° and Long.-68.02895°), and Wood Land (WL, Lat. 46.88520° and Long.-68.12577°), were selected to experiment in 2018. In 2019, five research sites were chosen to conduct the investigation, two in Presque Isle, Aroostook Farm (AF2 and AF3, Lat. 46.66134° and Long. -68.01808°), Limestone (LM, Lat: 46.96186° and Long. -67.83333°), two in Caribou (CA2, Lat. 46.89628° and Long. -68.07750°), and (CA3, Lat: 46.89180° and Long.-68.04055°). All locations have a separate average annual precipitation and temperature, where AF1, 2, and 3 locations had yearly average precipitation of 91.0 cm and an annual mean temperature of 5.15 °C. The sites WL, NS (1 and 2), as well as CA1, 2, and 3 had average yearly rainfall of 97.9 cm and a yearly mean temperature of 4.3 °C,

whereas FV had average annual precipitation of 85.5 cm and an annual mean temperature of 3.6 °C (59).

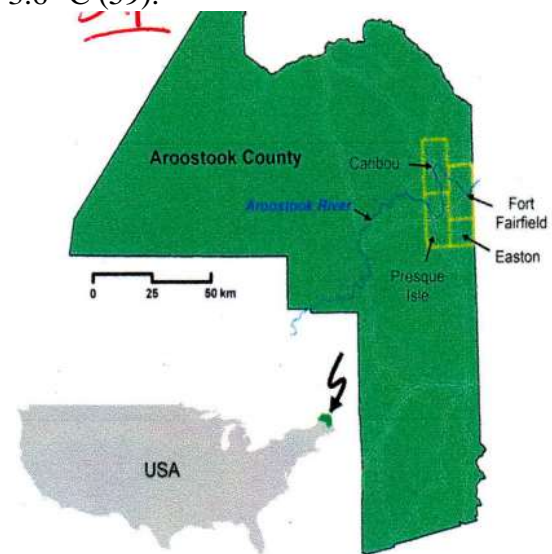


Figure 1. The study area, Aroostook County within Maine state, USA (62).

Experimental materials

The investigation covered three potato cultivars: Russet Burbank, Shepody, and Superior. Russet Burbank was planted in all sites, while Shepody and Superior were planted in AF. The planting space was 30 cm within rows, where the row width was about 90 cm.

Experimental treatments and design

Six N rates of fertilizer, 0, 56, 112, 168, 224, and 280 kg ha⁻¹, as ammonium sulfate ((NH₄)₂SO₄) in the first year and ammonium nitrate (NH₄NO₃) in the second year, were used on all the locations a randomized complete block design, RCBD, with four replications. Phosphorus (P), potassium (K), and sulfur (S) were used as instructed by the University of Maine Soil Laboratory (Maine). Each subplot was 9.14 m long × 3.65 m wide and had four rows. A distance of 1.50 m was left between replicates as a buffer zone. All managing procedures, such as disease, pest, insect control, and weeding, were employed for all locations. Planting was conducted between the middle and end of May; harvesting was accomplished between September and October.

Soil properties

Pre-plant soil samples were sampled from each location, and then samples were examined at the University of Maine Soil Laboratory. The USDA-Natural Resources Conservation

Service-NRCS (United States Department of Agriculture) was utilized for physical soil properties (Table 1). The locations NS-1, NS-2, and WL were subjected to three-year crop rotation (potato-grain-cover crop), whereas CA1 and FV locations had a two-year crop rotation (potato-grain). The AF1, 2, and 3 locations did not follow any crop rotation plan, where the grass was grown continually over seven years. The locations CA2, CA3, and LM locations subjected to two-year crop rotation (potato, (mustard, radish)), (potato, (red, white clover), rye), and (potato, (clover, oat-grains)) respectively, (Table 1).

Measurements

Active sensors and data collection: Active optical sensors (GS and CC) (Figure 2) were employed to assemble NDVI data weekly, where sensing began once plants reached the fourth leaf stage till the twentieth (4, 8, 10, 12, 16, and 20) (22). The NDVI data were acquired at 60 cm over the top of the plant

from the middle row of each plot, where about 40–60 readings were collected from each plot. The excel and in-house macro programs for Visual Basic were utilized to compute the mean of sensing data (21). The NDVI data were normalized by calculating in-season yield estimation (INSEY) and then combined according to leaves number (22), counted during each sense date. Data collection was continued until completing the twentieth leaf stage. After that, plants start laying down, and greenness declines, preparing to enter the maturing stage. Table 2 shows how sensors provided NDVI data during walk-throughs of plant rows; due to the long Excel columns, the table has been truncated to indicate the beginning and end of the data series. Table 2 a shows the beginning of collecting data, while (Table 2 b) with a marked row represents a new data collection for the following plot in the RCBD.

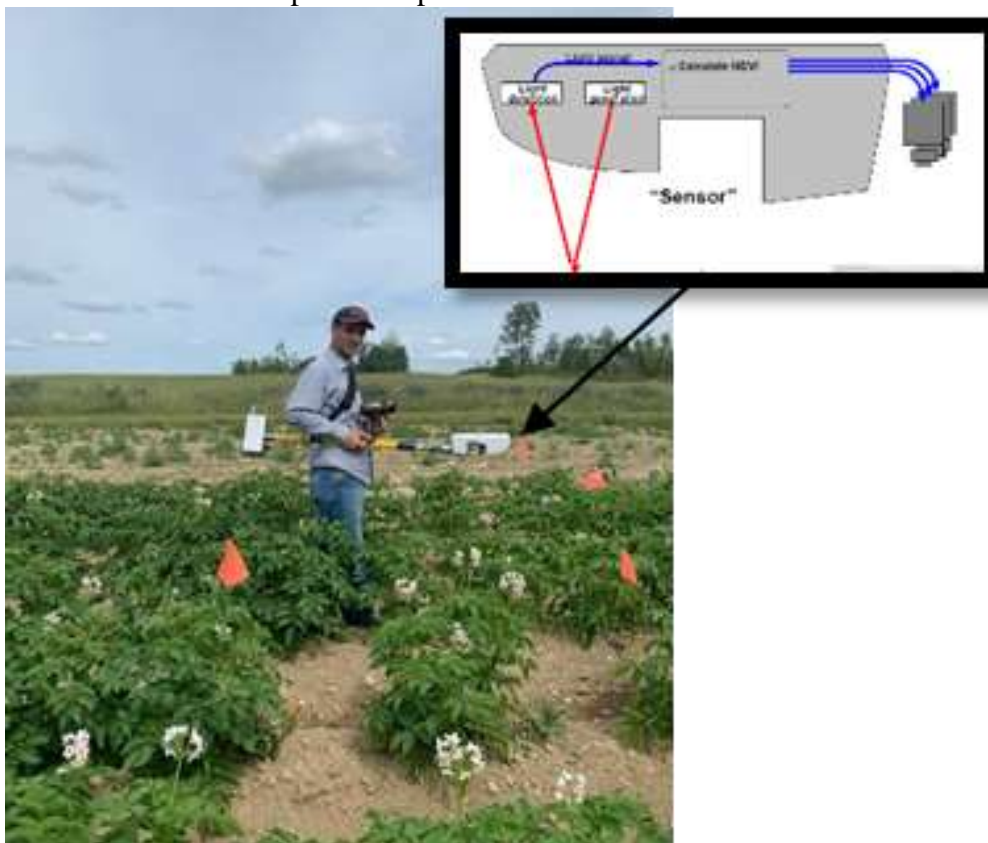


Figure 2. GreenSeeker sensor that was used in the experiment

Table 1. Soil chemical data, crop rotation, and soil series (66)

Site	pH	OM g kg ⁻¹	NO ₃	NH ₄	P	K _{sat}	Ca _{sat}	Mg _{sat}	S	Bo	Cu	Fe	Mn	Zn	CEC me 100g ⁻¹	Crop Rotation	Soil Series
AF1	6.5	27	7	4.0	21.5	98000	680000	222000	4.0	0.2	0.42	2.0	2.9	0.3	7.0	>3 yrs	Fine-Loamy, mixed, Frigid Typic Haplorthods
AF2	7.0	18	6	9.0	17.0	80000	660000	250000	4.0	0.1	0.38	3.3	2.7	0.7	6.2	>3 yrs	Fine-Loamy, mixed, Frigid Typic Haplorthods
AF3	6.0	18	12	8.0	15.0	70000	430000	162000	5.0	0.2	0.57	8.9	4.9	0.5	6.3	>3 yrs	Fine-Loamy, mixed, Frigid Typic Haplorthods
CA1	6.5	37	6	1.0	23.6	128000	692000	180000	8.0	0.3	0.67	5.0	3.2	1.3	7.9	2 yrs	Fine-Loamy, mixed, Frigid Aquic Haplorthods
CA2	5.0	41	8.0	3.0	19.4	90000	270000	96000	19.0	0.4	1.69	23	8.1	2.6	7.8	2 yrs	Gravelly loam, Isotic, Frigid, Typic Haplorthods
CA3	6.0	30	7.0	2.0	19.5	80000	800000	125000	9.0	0.3	0.95	6.2	1.3	1.4	6.2	2 yrs	Gravelly loam, Isotic, Frigid, Typic Haplorthods
FV	5.9	49	5	1.0	19.8	93000	771000	136000	15.0	0.3	0.85	10.0	4.2	1.3	7.3	3 yrs	Fine-Loamy, mixed, Frigid Aquic Haplorthods,
LM	6.0	33	3.0	2.0	19.0	90000	7700000	1370000	7.0	0.2	2.96	8.7	3.4	0.8	6.5	2 yrs	Gravelly loam, Isotic, Frigid, Typic Haplorthods
NS-1	5.4	45	21	6.0	18.2	46000	476000	127000	10.0	0.3	1.12	8.4	7.3	1.9	8.7	3 yrs	Fine-Loamy, mixed, Frigid Typic Haplorthods
NS-2	5.6	41	16	6.0	19.3	66000	541000	138000	6.0	0.3	1.33	11.0	8.8	1.7	7.9	3 yrs	Coarse-Loamy, Isotic, Frigid Oxyaquic Haplorthods
WL	5.8	41	15	5.0	16.5	99000	626000	147000	9.0	0.3	0.71	6.0	8.4	1.6	7.3	3 yrs	Fine-Loamy, mixed, Frigid Aquic Haplorthods

Soil reaction was estimated in a 1:1 ratio of soil to deionized water (63), organic matter was estimated utilizing loss on ignition (LOI) approach (7), micro and macronutrients and were extracted utilizing modified Morgan extraction approach (38), and estimated by ICP-OES (Inductively coupled plasma optical emission spectroscopy) (25), but phosphorus was estimated utilizing colorimetric (34), NO₃ was extracted utilizing KCL (33), cation exchange capacity (CES) was estimated utilizing ammonium acetate approach (25).

Table 2. Represents data (NDVI) collected by the GS*, a) starting a plot, b) ending the previous plot, and starting the next, as the marked row showing.

(a) A	B	C	D	(b) A	B	C	D
Time	Plot	Count	NDVI				
37010	27	1	0.689	440510	27	32	0.854
437110	27	2	0.798	440610	27	33	0.828
437210	27	3	0.829	440710	27	34	0.815
437310	27	4	0.832	440810	27	35	0.864
437410	27	5	0.852	440910	27	36	0.847
437510	27	6	0.722	441010	27	37	0.856
437610	27	7	0.828	441110	27	38	0.868
437710	27	8	0.847	441210	27	39	0.875
437810	27	9	0.851	441310	27	40	0.837
437910	27	10	0.871	441410	27	41	0.839
438010	27	11	0.855	441510	27	42	0.853
438110	27	12	0.871	441610	27	43	0.842
438210	27	13	0.864	441710	27	44	0.795
438310	27	14	0.842	441810	27	45	0.836
438410	27	15	0.838	441910	27	46	0.769
438510	27	16	0.856	442010	27	47	0.43
438610	27	17	0.866	442110	27	48	0.256
438710	27	18	0.865	442210	27	49	0.258
438810	27	19	0.881	442310	27	50	0.256
438910	27	20	0.878	442410	27	51	0.456
439010	27	21	0.856	442510	27	52	0.514
439110	27	22	0.878	442610	27	53	0.589
439210	27	23	0.872	442710	27	54	0.682
439310	27	24	0.876	442810	27	55	0.764
439410	27	25	0.861	442910	27	56	0.668
439510	27	26	0.811	443010	27	57	0.767
439610	27	27	0.874	443110	27	58	0.689
439710	27	28	0.845	444610	27	59	0.874
439810	27	29	0.828	444710	28	60	0.821
439910	27	30	0.809	444810	28	61	0.878
440010	27	31	0.815	444910	28	62	0.787
440110	27	32	0.854	445010	28	1	0.846
440210	27	33	0.828	445110	28	2	0.847
440310	27	34	0.809	445210	28	3	0.846
440410	27	35	0.815	445310	28	4	0.835
				445410	28	5	0.819

GreenSeeker*Sensor description and sensing procedure:**

Two handheld active optical sensors (GS-Trimble Navigation Limited, Sunnyvale, CA, USA) and (CC-ACS-430 sensor Holland Scientific, Inc., Lincoln, NE, USA) were used in this investigation. The GS sensor estimates incident and reflected light from the plant canopy at a wavelength in R and NIR bands (Table 3) (50). In GreenSeeker, a ray is transmitted from light-emitting diodes at different duration, such that the visible source pulses for 1.0 ms, and then the NIR diode

source pulses for 1.0 ms with 40,000 Hz. The light covers about 60 cm in width by 1.0 cm in length, with a long dimension positioned vertically in the running path. The Crop Circle sensor emits white light and employs three types of filters, which are R, red-edge, and NIR (Table 3), in order to segregate the reflected light. The sensor collects nearly 2–20 of NDVI readings per second, so with each recorded value in a 6.0 m plot length with 5.0 km hr⁻¹ of walking speed, there is an average of 4000 NDVI readings. Sensor outputs are

reflectance values that allow the calculation of vegetation indices (50).

The equation for red-NDVI and red-edge NDVI is :

$$\text{Red NDVI} = \frac{\text{NIR}-\text{Red}}{\text{NIR}+\text{Red}} \dots(1) \quad \text{Red Edge}$$

$$\text{NDVI} = \frac{\text{NIR}-\text{Red Edge}}{\text{NIR}+\text{Red Edge}} \dots(2) \quad \text{Where NIR=near-}$$

infrared band

Red= red band

RedEdge= rededge band

Table 3. Spectral properties for sensors

Properties	Sensors	
	GreenSeeker	Crop Circle
R (nm)	656	630
Red-edge (nm)	-	730
NIR (nm)	774	780
Total covered area (cm ²)	92	741

Due to the insignificant differences in the growth stages among locations, NDVI data were normalized employing the INSEY procedure. The in-season estimate of yield (INSEY) could be beneficial if NDVI data were combined from different sites and years. The in-season estimate of yield (INSEY) (43) was calculated by dividing NDVI data on the GDD that started from the planting date to the date of taking sensor readings (58), as shown in equation (3).

$$\text{GDD} = [(\text{T}_{\text{max}}+\text{T}_{\text{min}})/2] - \text{C} \dots(3)$$

where: Tmax. and Tmin. describes the daily maximum and minimum temperatures, which is the base growing temperature for potato (10°C). Sensing was performed by passing GS and CC at an approximate space of 60 cm over the plant canopy, resulting in a similar portion of reflectance at all locations and each growth stage (21).

Yield harvesting and calculation

A three 3.0-m random plot length selected from the two middle rows (6.0 m total) of each subplot was harvested mechanically utilizing a potato digger machine. Potato tubers were collected into individual paper containers of 23 kg capacity, then cleaned from plant and soil debris, and graded to four different sizes using a potato grading machine. The length of the two middle rows (6.0 m total length) of each subplot was converted to 3.0 m length and then utilized to estimate total yield production employing the equation supplied by North Dakota and Minnesota (equation 4) (16). The

certain weight/acre (cwt/acre) = [lb/(10 ft). × multiplication Factor](4) The multiplication factor relies on the row width, where 14.5 is used when planting a row at a width of 36 inches (90cm). Equation (4) was used to calculate the total yield per area and then converted to the standard units (Mg ha⁻¹). The total weight per plant was estimated by dividing the total weight of tubers from each subplot by the number of plants in a row. Tuber yield data has been combined from the two years of research based on N rates and statistical analysis.

Data analysis

Analysis of variance test (ANOVA) was used to examine the effect of nitrogen rates on potato tuber yield using SPSS software (52). Microsoft Excel (40) was used to plot the relationships between potato tuber yields and a series of nitrogen rates. The bar graph (Figure 1) shows the difference between the control treatment (0 N kg ha⁻¹) and other treatments in addition to the N rate that maximized the potato yield and the rate after which potato yield did not respond significantly. Regression analysis was conducted between potato yield and sensors data (INSEY) to generate models for yield prediction.

RESULTS AND DISCUSSION

A large gap was noticed among yield data from the 11 sites. Therefore, a multiple regression analysis was conducted (data not shown) among soil characteristics and yield, where OM was found to be the main factor that had a high correlation with crop yield ($R^2 = 0.78^{**}$) at P-value <0.01. Therefore, all sites were divided into soil OM ≤30 g kg⁻¹ and ≥30 g kg⁻¹. The sites LM, WD, NS-1, NS-2, FV, CA1, CA2, and CA3, were categorized as ≥ 30.0 g kg⁻¹ OM, while the sites AF1, AF2, and AF3 were categorized as ≤30 g kg⁻¹ OM. It is crucial to note that the Superior and Shepody potato cultivars had only one location each that came under ≤30 g kg⁻¹ OM.

Yield responses to nitrogen rates

Potato yields at various N rates are shown in (Figure 3), showing the association between N rates and potato yields for locations with ≤ 30 g kg⁻¹ of OM, ≥ 30 g kg⁻¹ of soil OM, and an average of all locations combined. The potato yield remarkably enhanced with N fertilizer applications at all the abovementioned

locations. Compared with the control remedy, 0 kg N ha⁻¹, the yields under 56, 112, 168 kg ha⁻¹ treatments were enhanced by 10.75%, 20.71%, and 18.76%, concerning the rates of 56 kg N ha⁻¹; 13.3%, 28.8%, and 25.4%, respectively, for 112 kg N ha⁻¹, 21.72%, 42.73%, and 37.74%, respectively for 168 kg

N ha⁻¹. For all locations, potato yields enhanced as the N rate increased from 0 kg N ha⁻¹ to 168 kg N ha⁻¹. Nonetheless, no substantial increase was seen for 224 kg N ha⁻¹, indicating that the 168 kg N ha⁻¹ was the highest economic rate for potato yield production.

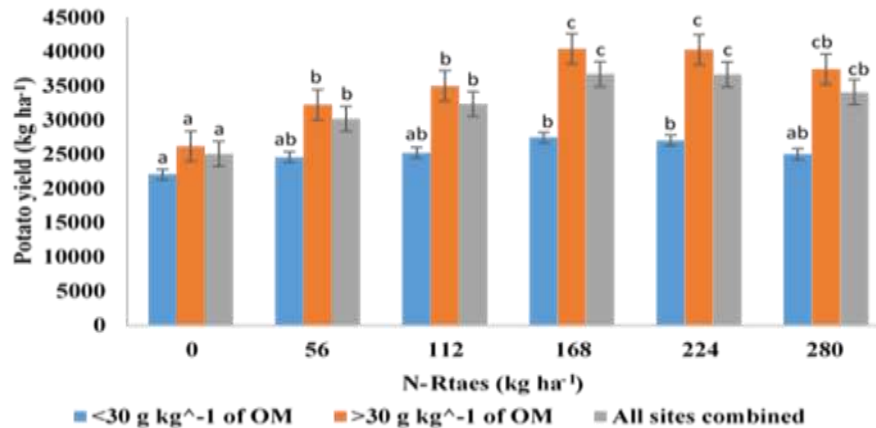


Figure 3. The potato yield response to various N fertilizer rates from all locations during two growing seasons (2018-2019) when P-value < 0.05 (2).

Procedure (1) for nitrogen recommendation Generating the nitrogen fertilizer optimization algorithm (NFOA)

Algorithms for managing N rates for numerous crops and regions have been established (28). They can be practiced in a sensor-based N rate calculator produced by agronomists at Oklahoma State University to feed in zone-specific sensor data for determining the in-season crop yield and N response index (RI). The algorithm of N rate recommendation for sensor-based information is (41);

$$N_{rate} = \frac{[(YPO \times RI) - YPO] \times N\%}{NUE} \dots(5)$$

where YPO is the maximum achievable crop yield with no applied N.

RI is the response index

N% is the percentage of N in the yield, 0.026 (2.6%) (1).

NUE is the nitrogen use efficiency (yield ratio to fertilizers) (18).

Yield potential (YPO)

YPO is defined as the maximum achievable crop yield with no applied N. Considered the backbone of any N fertilizer rate measurements, YPO can be predicted from the relationship between crop yield and INSEY (35,43). The yield potential (YPO) is presented by

$$YPO = Ae b \frac{NDVI}{GDD} \dots(6)$$

where: A and b indicate the intercept and slope, respectively, of the exponential function due to the regression analysis between potential yield and INSEY (43,52). The regression analysis between tuber yield (kg ha⁻¹) from the plots of 0 N kg ha⁻¹ and INSEY data was used to generate the prediction equation. All growth stages were tested to generate the best-fitted model. The end of the tuber initiation of growth stage (20th of leaves stages) was the best time to produce a significant coefficient of determination (R²=0.24) at P-value <0.05. The tuber initiation stage is when that plant uptake more N for tuber growth, while the stage before (vegetative growth) is the time to leave and stems from growing, so most of the N is only for vegetative growth. Next is the tuber bulking, where the N uptake is less than the tuber initiation stage (66). The INSEY derived from GS-NDVI data showed a significant relationship between the INSEY derived from CC-NDVI and CC-NDRE data.

Response index

The NDVI readings from the high N (N-rich) plot divided by the NDVI of the test plot is referred to as the response index (RI), which refers to the possibility of increment in crop yield with added N (30). The RI also is a valuable indicator of crop yield response and a guide to reduce Type II errors (31,41).

Hodgen et al. (2005) (27) concluded that when $1 < RI < 1.1$, N application will likely result in no yield response to any added fertilizer

$1.1 < RI < 1.25$, N application will likely result in marginal responses

$RI > 1.25$, N application will result in a response

Johnson and Raun (2003) (31) first defined yield response to applied N as the proportion of crop yield of an N-reference plot to that of a non-N treated plot given by,

$$RI_{\text{Harvest}} = \frac{\text{Yield}_{\text{N-Reference strip}}}{\text{Yield}_{\text{non-treated strip}}} \dots\dots(7)$$

The mean tuber yield produced from the N-rich plot (280 kg N ha^{-1}) was $34080.91 \text{ kg ha}^{-1}$, while untreated plots (0 kg N ha^{-1}) produced a mean of $25071.85 \text{ kg ha}^{-1}$. As a result, the response index (RI) was equal to 1.36, more than 1.25; the N application will respond. Raun et al. (2002) (44) stated that the combined advantage of the RI concept and INSEY allowed an accurate top-dressed N rate for wheat. Total grain (yield) N removed from each area is measured, and the difference between the N-rich and farmer's application values were divided by a calculated NUE value.

$$RI_{\text{NDVI}} = \frac{\text{NDVI}_{\text{Reference strip}}}{\text{NDVI}_{\text{non-treated strip}}}$$

Nitrogen use efficiency

The most fundamental description of nutrient use efficiency is a crop yield per unit of available nutrients (52), while Teboh et al. (2012) (53) described it as the portion of N input, indicating that it corresponds to the portion of N taken up to fulfill further yield needs. The information can be utilized to assess the nutrient use efficiency of a given cropping operation on an annual or multi-year basis. Nitrogen use efficiency (NUE) can be computed as defined by Baligar et al. (2001) (6) as follows:

$$NUE = \frac{(\text{Crop yield in N fertilized plot} - \text{Crop yield in no N plot})}{(\text{Quantity of N fertilizer applied in N fertilized plot})} \dots\dots (9)$$

Thus, applying the data of potato yield and sensors in equation (5) resulted in 195 kg ha^{-1} being the N recommendation for the potato crop, which is about 14% lower than the amount that potato growers have previously applied, 224 kg N ha^{-1} .

Procedure (2) nitrogen recommendation

This procedure differs from procedure number (1) mathematically; however, the materials (yield and sensor data) are still the same. Sharma (2014) (47) used procedure number (2) to generate an N recommendation for the corn crop, as in equation (10).

$$N \text{ rate in } \text{kg ha}^{-1} = \frac{[(Y1 - Y2) \times N\%]}{NUE} \dots\dots(10)$$

where: Y1 is the predicted yield from the N-rich plot in kg ha^{-1}

Y2 is the predicted yield from farmer practice plot in kg ha^{-1}

N% is nitrogen percent in potato tuber, 0.026 (2.6%) (1).

NUE is the nitrogen use efficiency. As mentioned before, the N-rich plot is the plot that has been provided with a complete fertilizer to be an unlimited N area. Nitrogen (N) at 280 kg N ha^{-1} was applied to fulfill the N-rich plot, while 224 kg N ha^{-1} was the rate practiced by potato growers in Maine. A regression analysis was conducted between potato tuber yield and sensor data (INSEY) to generate an algorithm for potato yield prediction at p-value < 0.05 . The exponential model was the best to fit that curve for both Y1 and Y2, respectively. The twentieth leaf stage was most likely to have a significant relationship between yield data and INSEY. The GS and CC sensors showed a significant association with yield data, but the determination coefficient for the CC-NDRE was higher than those for other wavelengths (NDVI from GS and CC) (Figure 5 a-d). At the 20th leaf stage, the plant vegetation density is maximum, called the NDVI saturation condition. The red (R) wavelength from GS and CC is sensitive only for a low range of chlorophyll ($3-5 \mu\text{g.cm}^{-2}$) in comparison to the CC red-edge wavelength that is sensitive to a wider range ($0.3-45 \mu\text{g cm}^{-2}$) (23). As a result, applying the potato yield data and sensor data in equation (10) resulted in 199 kg ha^{-1} being the N recommendation for potato crops, which is about 12% lower than the amount that potato growers used to apply, 224 kg N ha^{-1} . Although the coefficient of determination (R^2) was statistically significant but not very strong (0.13, 0.24, 0.27, and 0.38), it could still be considered a step toward utilizing active optical sensors for the nitrogen recommendation potato crop. Experimenting with sites with different soil properties is a

reason to have representative samples from multiple locations. However, having a massive gap among sites could be a problem, especially for statistical analysis. That was our problem; there was a significant gap in the yield data between locations. Classifying or grouping

sites is an excellent idea to overcome this issue, but running a regression analysis for a single N rate (0, 224, 280 kg N ha⁻¹) using a few points are considered insufficient. Therefore, experimenting with enough numbers sites would be the solution to this issue

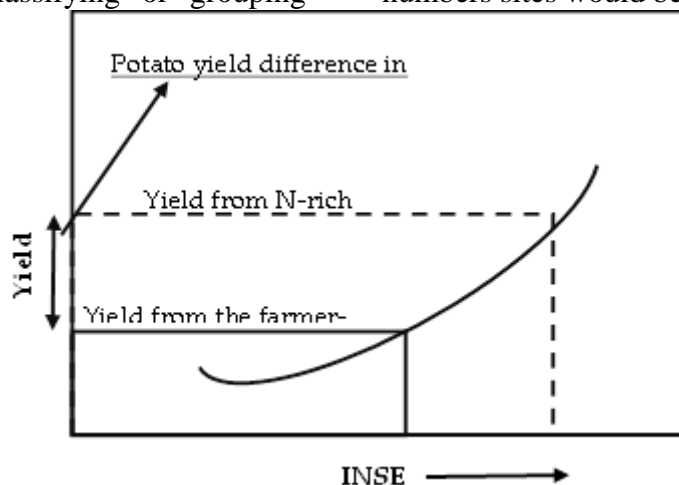


Figure 4. The schematic demonstrates the algorithm of nitrogen recommendation’s work

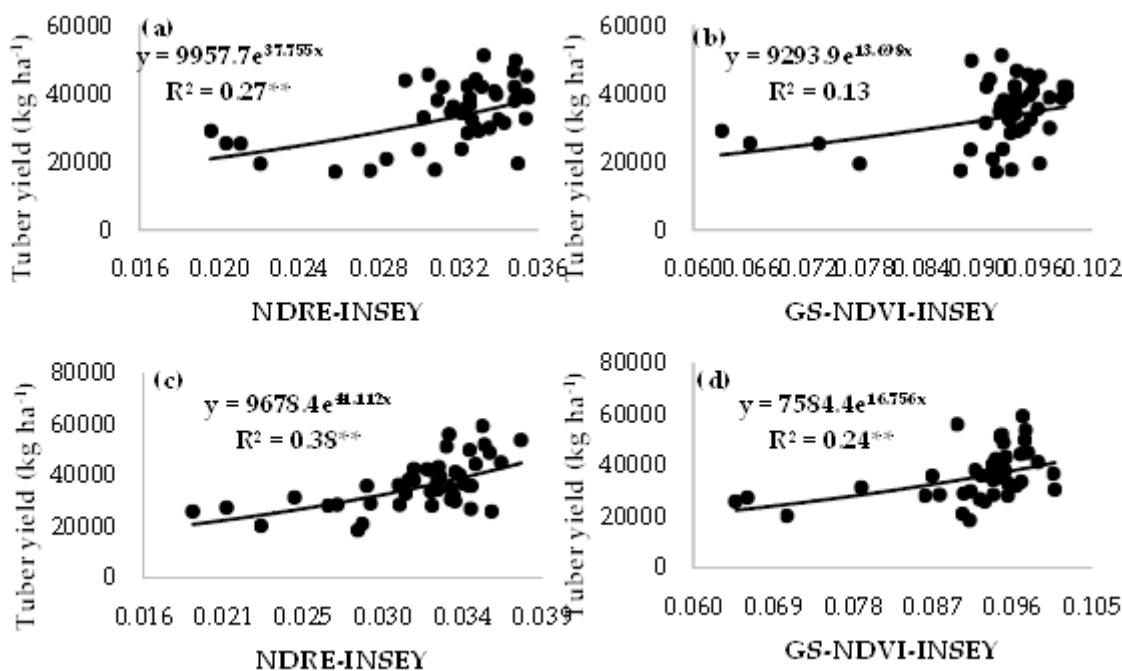


Figure 5. The relationship between potato tuber yield (kg ha⁻¹) and the sensor reading (INSEY) derived from a) NDRE, b) GS-NDVI for the N-rich plots, and c) NDRE, d) GS-NDVI for the farmers practiced rate r, at p-value<0.05

CONCLUSIONS

At the twentieth leaf stage, the sensing time has been observed to give a significant yield estimation. Although the determination coefficient between tubers yield and sensor data is not very strong due to uncontrolled factors such as temperature and rain, it still can give good results regarding yield prediction. Despite satellite and aerial images covering

large areas; still, there are some uncontrolled restrictions, such as clouds issues and temporal resolution (16 days for Landsat-8). Ground-based sensors have advantages that let users overcome some of these disadvantages. For instance, there is no problem with cloudy day’s condition that affects the sunlight to reach ground targets, where ground-based sensors do not depend on sunlight as a source

of light; it sends and collects its lights. Ground-based sensors can be used anytime unless there is no rain or muddy soil (64). The disadvantages of active sensors are the high initial cost. The used sensors may be somewhat expensive (\$4000.0), but a small GS can do the same job. GS provides only the NDVI resulting from the NIR and R wavelengths, and it is cheap (\$400.0) and easy to use. There is news that Trimble, the producer of the handheld GS, is working on updating the sensor; we hope that they will add the red edge wavelength as well. In procedure (1), the calculation depended totally on the predicted yield from the control treatment (0 kg N ha⁻¹). There was no chlorophyll saturation issue, so the R wavelength showed a considerable association with yield data. In contrast, procedure (2) calculations depended on the N-rich and farmer-practice plots (280 and 224 kg N ha⁻¹), respectively. The chlorophyll saturation issue happens commonly, so the red-edge wavelength was the best to overcome this issue and showed a significant relationship with yield data. The N-recommendation rates from both procedures (1&2) (195 and 199 kg N ha⁻¹), respectively, were lower than the average rate that potato growers in Maine are applying annually (224 kg ha⁻¹). Procedure (1) can save about 14% of the rate that potato growers apply, while procedure (2) can save about 12%. So far, these were valuable results and a good step toward utilizing active optical sensors to generate N recommendations. However, to be more accurate, the sites had to be classified into the soil with high OM content and soils with low OM content, but according to literature from other crops, calculations cannot be conducted with a small number of sites. Therefore, separating our sites into two classes will be scientifically a good idea, but when we have at least ten to fifteen sites experimenting with different soil types can help determine whether soil properties have a significant effect on the N-recommendation outcome or not. The same issue holds for potato cultivars when planting specific cultivars in a particular soil type; it can expose whether potato cultivars significantly affect the N-recommendation outcome.

Conflicts of Interest: The authors declare no conflict of interest

REFERENCES

1. Ahmed, A., M. A. El-Baky., A. Ghoname., G. Riad and S. El-Abd. 2009. Potato tuber quality is affected by nitrogen form and rate. *Middle Eastern Russian Journal of Plant Sciences and Biotechnology*, 3, 47-52.
2. Ahmed, A. Z., S. Lakesh., J. Ahmed., B. Sukhwinder., B. Aaron and A. Andrei. 2020. In-season potato yield prediction with active optical sensors. *Agrosystems, Geosciences and Environment*, 3(3), 1-15.
3. Ali, A. M., H. S. Thind., S. Sharma and Y. Singh. 2015. Site-specific nitrogen management in dry direct-seeded rice using chlorophyll meter and leaf colour chart. *Pedosphere*, 25(1), 72-81.
4. Alkhafaji, A. R and N. H. Khalil. 2019. Effect Of Fertilization, Rootstocks and Growth Stimulant On Growth Of Citrus Limon L. Sapling. *Iraqi Journal of Agricultural Science*, 50(3), 990-1000
<https://doi.org/10.36103/ijas.v50i4.743>
5. Alva, L. 2004. Potato nitrogen management. *Journal of Vegetable Crop Production*, 10(1), 97-132.
6. Baligar, V., N. Fageria and Z. He. 2001. Nutrient use efficiency in plants. *Communications in Soil Science and Plant Analysis*, 32(7-8), 921-950.
7. Ball, D. F. 1964. Loss-on-ignition as an estimate of organic matter and organic carbon in non-calcareous soils. *Journal of Soil Science*, 15(1), 84-92
8. Basyouni, R and B. Dunn. 2013. Use of optical sensors to monitor plant nitrogen status in horticultural plants. *Division of Agricultural Sciences and Natural Resources, Oklahoma State University*. HLA-6719:1-4pp
9. Blackmer, T., J. Schepers., G. Varvel and Walter-Shea, E. 1996. Nitrogen deficiency detection using reflected shortwave radiation from irrigated corn canopies. *Agronomy Journal*, 88(1), 1-5.
10. Blackmer, T. M and J. S. Schepers. 1996. Aerial photography to detect nitrogen stress in corn. *Journal of plant physiology*, 148(3-4), 440-444.
11. Bronson, K. F., T. T. Chua., J. Booker., J. W. Keeling and R. J. Lascano. 2003. In-season nitrogen status sensing in irrigated cotton. *Soil*

- science society of America Journal, 67(5), 1439-1448.
12. Bu, H., L. K. Sharma., A. Denton and D. W. Franzen. 2016. Sugar beet yield and quality prediction at multiple harvest dates using active-optical sensors. *Agronomy Journal*, 108(1), 273-284.
 13. Butchee, K. S., J. May and B. Arnall. 2011. Sensor-based nitrogen management reduced nitrogen and maintained yield. *Crop Management*, 10(1), 0-0.
 14. Cassman, K. G., A. Dobermann and D. T. Walters. 2002. Agroecosystems, nitrogen-use efficiency, and nitrogen management. *AMBIO: A Journal of the Human Environment*, 31(2), 132-141.
 15. Dilz, K. 1988. Efficiency of uptake and utilization of fertilizer nitrogen by plants. *Nitrogen efficiency in agricultural soils*, 1-26.
 16. Donavon, J., P. Diane., P. Todd., K. Ted and R. Andy. 1946. Yield estimates -northern plains potato growers association-NPPGA. Retrieved from http://nppga.org/crop_science/measurements.php
 17. Errebhi, M., C. J. Rosen., S. C. Gupta and D. E. Biring. 1998. Potato yield response and nitrate leaching as influenced by nitrogen management. *Agronomy Journal*, 90(1), 10-15.
 18. Fageria, N. 2009. The use of nutrients in crop plants. In *Earth Sciences, Environment and Agriculture*. pp: 448. doi:<https://doi.org/10.1201/9781420075113>
 19. Felton, W. L., C. L. Alston., B. M. Haigh., P. G. Nash., G. A. Wicks and G. E. Hanson. 2002. Using reflectance sensors in agronomy and weed science. *Weed Technology*, 16(3), 520-527.
 20. Franzen, D. W., L. K. Sharma and H. Bu. 2014. Active optical sensor algorithms for corn yield prediction and a corn side-dress nitrogen rate aid: NDSU Extension Service, North Dakota State University. NDSU Extension Circular SF1176-5
 21. Bean, G. M., N. R. Kitchen., J. J. Camberato., R. B. Ferguson., F. G. Fernandez., D. W. Franzen and J. S. Shanahan. 2018. Active-optical reflectance sensing corn algorithms evaluated over the United States Midwest Corn Belt. *Agronomy Journal*, 110(6), 2552.
 22. Liu, N., R. Zhao., L. Qiao., Y. Zhang., M. Li., H. Sun and X. Wang. 2020. Growth stages classification of potato crop based on analysis of spectral response and variables optimization. *Sensors*, 20(14), 3995
 23. Gitelson, A., A. Viña., T. Arkebauer., D. Rundquist., G. Keydan and B. Leavitt. 2003. Remote estimation of leaf area index and green leaf biomass in maize canopies. *Geophysical Research Letters*, 30(5).
 24. Govender, M., K. Chetty and H. Bulcock. 2007. A review of hyperspectral remote sensing and its application in vegetation and water resource studies. *Water Sa*, 33(2), 145-151.
 25. Hendershot, W. H and M. Duquette. 1986. A simple barium chloride method for determining cation exchange capacity and exchangeable cations. *Soil science society of America journal*, 50(3), 605-608.
 26. Herrmann, I., A. Karnieli., D. J. Bonfil., Y. Cohen and V. Alchanatis. 2010. SWIR-based spectral indices for assessing nitrogen content in potato fields. *International Journal of Remote Sensing*, 31(19), 5127-5143
 27. Hodgen, P., W. Raun., G. Johnson., R. Teal., K. Freeman., K. Brixey and M. Stone. 2005. Relationship between response indices measured in-season and at harvest in winter wheat. *Journal of Plant Nutrition*, 28(2), 221-235.
 28. Holzapfel, C., G. Lafond., S. Brandt., P. Bullock., R. Irvine., M. Morrison and D. James. 2009. Estimating canola (*Brassica napus L.*) yield potential using an active optical sensor. *Canadian Journal of Plant Science*, 89(6), 1149-1160.
 29. Johnson, G. 1991. A general model for predicting crop response to fertilizer. *Agronomy Journal*, 83(2), 367-373.
 30. Johnson, G and W. Raun. 1995. Nitrate leaching in continuous winter wheat: use of a soil-plant buffering concept to account for fertilizer nitrogen. *Journal of Production Agriculture*, 8(4), 486-491.
 31. Johnson, G and W. Raun. 2003. Nitrogen response index as a guide to fertilizer management. *Journal of Plant Nutrition*, 26(2), 249-262
 32. Jongschaap, R. E. E. 2006. Integrating crop growth simulation and remote sensing to improve resource use efficiency in farming systems. Wageningen University and Research. 1-24pp

33. Keeney, D. R and D. W. Nelson. 1982. Nitrogen—Inorganic Forms 1. Methods of soil analysis. Part 2. Chemical and microbiological properties (methods of soil an2), 643-698.
34. Knudsen, D and D. Beegle. 1988. Recommended phosphorus tests. Recommended chemical soil tests procedures for the north central region. Bulletin No. 499 (Revised). p. 12-15.
35. Large, E. C. 1954. Growth stages in cereals illustration of the Feekes scale. Plant pathology, 3(4), 128-129.
36. Lukina, E., K. Freeman., K. Wynn., W. Thomason., R. Mullen., M. Stone and R. Elliott. 2001. Nitrogen fertilization optimization algorithm based on in-season estimates of yield and plant nitrogen uptake. Journal of Plant Nutrition, 24(6), 885-898.
37. Maine, U. O. Analytical Lab and Maine Soil Testing Service. Retrieved from <https://umaine.edu/soiltestinglab/>
38. Masood, T. K and S.S. Shahadha. 2021. Simulating The Effect Of Climate Change On Winter Wheat Production And Water/Nitrogen Use Efficiency In Iraq: Case Study. The Iraqi Journal of Agricultural Science, 52(4), 999-1007
39. McIntosh, J. L. 1969. bray and morgan soil extractants modified for testing acid soils from different parent materials. Agronomy Journal, 61, 259-265.
40. Microsoft-Corporation. 2018. Microsoft Excel. Retrieved from <https://office.microsoft.com/excel>
41. Mullen, R. W., K. W. Freeman., W. R. Raun., G. V. Johnson., M. L. Stone and J. B. Solie. 2003. Identifying an in-season response index and the potential to increase wheat yield with nitrogen. Agronomy Journal, 95(2), 347-351.
42. Raun, W. R and G. V. Johnson. 1999. Improving nitrogen use efficiency for cereal production. Agronomy Journal, 91(3), 357-363.
43. Raun, W. R., J. B. Solie., G. V. Johnson., M. L. Stone., E. V. Lukina., W. E. Thomason and J. S. Schepers. 2001. In-season prediction of potential grain yield in winter wheat using canopy reflectance. Agronomy Journal, 93(1), 131-138.
44. Raun, W. R., J. B. Solie., G. V. Johnson., M. L. Stone., R. W. Mullen., K. W. Freeman and E. V. Lukina. 2002. Improving nitrogen use efficiency in cereal grain production with optical sensing and variable rate application. Agronomy Journal, 94(4), 815-820.
45. Sharma, L and S. Bali. 2017. A review of methods to improve nitrogen use efficiency in agriculture. Sustainability, 10(1), 51.
46. Sharma, L., S. Bali., J. Dwyer., A. Plant and A. Bhowmik. 2017. A case study of improving yield prediction and sulfur deficiency detection using optical sensors and relationship of historical potato yield with weather data in maine. Sensors, 17(5), 1095.
47. Sharma, L., A. Zaeen., S. Bali and J. Dwyer. 2017. Improving nitrogen and phosphorus efficiency for optimal plant growth and yield. in new visions in plant science: IntechOpen. New Vision in Plant Science, 13-40pp
48. Sharma, L. K. 2014. Evaluation of active optical ground-based sensors to detect early Nitrogen Deficiencies in Corn. North Dakota State University. ProQuest Dissertations.1-224 pp
49. Sharma, L. K., H. Bu., A. Denton and D. Franzen. 2015. Active-optical sensors using red NDVI compared to red edge NDVI for prediction of corn grain yield in North Dakota, USA. Sensors, 15(11), 27832-27853.
50. Sharma, L. K., H. Bu and D. W. Franzen. 2016. Comparison of two ground-based active-optical sensors for in-season estimation of corn (*Zea mays, L.*) yield. Journal of Plant Nutrition, 39(7), 957-966.
51. Shayaa H, Hussein W. A. 2019. Effect of Neem (*Azadirachta indica*) leaves extract and organic fertilizer in the productivity and quality of two potatoes Varieties, Iraqi Journal of Agricultural Sciences, 50(1): 275- 285. <https://doi.org/10.36103/ijas.v50i1.293>
52. SPSS-IBM-Corp. 2017. IBM SPSS Statistics for Windows, Version 25.0. Armonk, NY: IBM Corp. Released 2017. (Version 25.0). Armonk, NY
53. Swain, E. Y., L. Rempelos., C. H. Orr., G. Hall., R. Chapman., M. Almadni and J. M. Cooper, J. M. 2014. Optimizing nitrogen use efficiency in wheat and potatoes: interactions between genotypes and agronomic practices. Euphytica, 199 (1-2), 119-136.
54. Teal, R., B. Tubana., K. Girma., K. Freeman., D. Arnall., O. Walsh and W. Raun.

2006. In-season prediction of corn grain yield potential using normalized difference vegetation index. *Agronomy Journal*, 98(6), 1488-1494.
55. Teboh, J. M., B. S. Tubaña., T. K. Udeigwe., Y. Y. Emendack and J. Lofton. 2012. Applicability of ground-based remote sensors for crop N management in Sub Saharan Africa. *Journal of Agricultural Science*, 4(3), 175.
56. Thind, H., A. Kumar and M. Vashistha. 2011. Calibrating the leaf colour chart for need based fertilizer nitrogen management in different maize (*Zea mays L.*) genotypes. *Field Crops Research*, 120(2), 276-282.
57. Tilman, D., K. G. Cassman., P. A. Matson., R. Naylor and S. Polasky. 2002. Agricultural sustainability and intensive production practices. *Nature*, 418(6898), 671.
58. Tremblay, N. 2004. Determining nitrogen requirements from crops characteristics. Benefits and challenges. *Recent Research Developments in Agronomy and Horticulture*, 157-182.
59. Tyler, K., F. Broadbent and J. Bishop. 1983. Efficiency of nitrogen uptake by potatoes. *American Potato Journal*, 60(4), 261-269.
60. United States Climate Data. 2018. Climate data for Maine, ME, Temperature-Precipitation-Sunshine-Snowfall. version 2.3. Retrieved from <http://www.usclima-tedata.com>
61. United States Department of Agriculture. 2019. Natural Resources Conservation Service. Web Soil Survey. Retrieved from <https://www.nrcs.usda.gov/wps/portal/nrcs/main/national/contact/>
62. Vos, J and D. MacKerron. 2000. Basic concepts of the management of supply of nitrogen and water in potato production.
63. Wang, C., J. Johnston., D. Vail., J. Dickinson., and D. Putnam 2015. High-Precision Land-Cover-Land-Use GIS Mapping and Land Availability and Suitability Analysis for Grass Biomass Production in the Aroostook River Valley, Maine, USA. *Land*, 4(1), 231-254
64. Wang, H and X. Xu. 2018. Cloud classification in wide-swath passive sensor images aided by narrow-swath active sensor data. *Remote Sensing*, 10(6), 812.
65. Watson, D and J. R. Brown. 1998. pH and Lime Requirement, p. 13-16. In: *Recommended Chemical Soil Test Procedures for the North Central Region*. NCR Publication No. 221. Missouri Agricultural Experiment Station, Columbia, MO, USA
66. Westermann, D. 1993. Fertility management. In: RC Rowe (ed), *Potato Health Management*. APS Press, St. Paul, MN (pp. 77-86).
67. Zaeen, A. A., L. K. Sharma., A. Jasim., S. Bali., A. Buzza and A. Alyokhin. 2020. Yield and quality of three potato cultivars under series of nitrogen rates. *Agrosystems, Geosciences & Environment*, 3(1), e20062.
68. Zainaldeen M, Abdul Rasool E. J. 2018. Effect of foliar application of gibberellin and nutrients on growth and yield of potato var. "burren", *Iraqi Journal of Agricultural Sciences*,49(2):168-176.
<https://doi.org/10.36103/ijas.v49i2.232>