

Washington University School of Medicine

Digital Commons@Becker

2020-Current year OA Pubs

Open Access Publications

12-1-2022

Bone metabolism and incretin hormones following glucose ingestion in young adults with pancreatic insufficient cystic fibrosis

Wang Shin Lei

Marissa J Kilberg

Babette S Zemel

Ronald C Rubenstein

Clea Harris

See next page for additional authors

Follow this and additional works at: https://digitalcommons.wustl.edu/oa_4

 Part of the [Medicine and Health Sciences Commons](#)

Please let us know how this document benefits you.

Authors

Wang Shin Lei, Marissa J Kilberg, Babette S Zemel, Ronald C Rubenstein, Clea Harris, Saba Sheikh, Andrea Kelly, and Joseph M Kindler



Contents lists available at ScienceDirect

Journal of Clinical & Translational Endocrinology

journal homepage: www.elsevier.com/locate/jcte

Original research

Bone metabolism and incretin hormones following glucose ingestion in young adults with pancreatic insufficient cystic fibrosis

Wang Shin Lei^a, Marissa J. Kilberg^{b,c}, Babette S. Zemel^{c,d}, Ronald C. Rubenstein^e, Clea Harris^f, Saba Sheikh^{c,g}, Andrea Kelly^{b,c}, Joseph M. Kindler^{a,*}^a Department of Nutritional Sciences, The University of Georgia, Athens, GA, USA^b Division of Endocrinology and Diabetes, The Children's Hospital of Philadelphia, Philadelphia, PA, USA^c Department of Pediatrics, University of Pennsylvania, Philadelphia, PA, USA^d Division of Gastroenterology, Hepatology, and Nutrition, Children's Hospital of Philadelphia, Philadelphia, PA, USA^e Division of Allergy and Pulmonary Medicine, Department of Pediatrics, Washington University School of Medicine, St. Louis, MO, USA^f Department of Pediatrics, Yale School of Medicine, New Haven, CT, USA^g Division of Pulmonary Medicine, The Children's Hospital of Philadelphia, Philadelphia, PA, USA

ARTICLE INFO

Keywords:

Nutrition
OGTT
Incretins
Bone
Cystic Fibrosis

ABSTRACT

Background: Gut-derived incretin hormones, including glucose-dependent insulinotropic peptide (GIP) and glucagon-like peptide 1 (GLP-1), regulate post-prandial glucose metabolism by promoting insulin production. GIP, GLP-1, and insulin contribute to the acute bone anti-resorptive effect of macronutrient ingestion by modifying bone turnover. Cystic fibrosis (CF) is associated with exocrine pancreatic insufficiency (PI), which perturbs the incretin response. Cross-talk between the gut and bone ("gut-bone axis") has not yet been studied in PI-CF. The objectives of this study were to assess changes in biomarkers of bone metabolism during oral glucose tolerance testing (OGTT) and to test associations between incretins and biomarkers of bone metabolism in individuals with PI-CF.

Methods: We performed a secondary analysis of previously acquired blood specimens from multi-sample OGTT from individuals with PI-CF ages 14–30 years (n = 23). Changes in insulin, incretins, and biomarkers of bone resorption (C-terminal telopeptide of type 1 collagen [CTX]) and formation (procollagen type I N-terminal propeptide [P1NP]) during OGTT were computed.

Results: CTX decreased by 32% by min 120 of OGTT (P < 0.001), but P1NP was unchanged. Increases in GIP from 0 to 30 mins (rho = -0.48, P = 0.03) and decreases in GIP from 30 to 120 mins (rho = 0.62, P = 0.002) correlated with decreases in CTX from mins 0–120. Changes in GLP-1 and insulin were not correlated with changes in CTX, and changes in incretins and insulin were not correlated with changes in P1NP.

Conclusions: Intact GIP response was correlated with the bone anti-resorptive effect of glucose ingestion, represented by a decrease in CTX. Since incretin hormones might contribute to development of diabetes and bone disease in CF, the "gut-bone axis" warrants further attention in CF during the years surrounding peak bone mass attainment.

Introduction

Bone health tracks strongly throughout the growing years and into adulthood, and disruption of normal bone accrual during childhood might increase risk for osteoporosis and fracture in later life [1]. Myriad behavioral and biological factors contribute to peak bone mass,

particularly nutrition and related hormonal mediators [2–4]. The biological mechanisms through which nutrition impacts bone health are not clearly defined.

Bone accretion largely depends on the independent and coordinated actions of the bone-forming osteoblasts and the bone-resorbing osteoclasts [5]. These bone-regulating cells are responsive to a variety of

Abbreviations: CF, cystic fibrosis; PI, pancreatic insufficiency; OGTT, oral glucose tolerance test; GIP, gastric inhibitory polypeptide; GLP-1, glucagon-like peptide-1; CTX, C-terminal telopeptide of type 1 collagen; P1NP, procollagen type I N-terminal propeptide.

* Corresponding author.

E-mail address: kindlerj@uga.edu (J.M. Kindler).

<https://doi.org/10.1016/j.jcte.2022.100304>

Received 27 May 2022; Received in revised form 12 August 2022; Accepted 29 August 2022

Available online 3 September 2022

2214-6237/© 2022 The Author(s). Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

stimuli, including macronutrient ingestion [6,7]. One hypothesized mechanism through which nutrition might influence peak bone mass is through activation and/or deactivation of cells orchestrating bone turnover, thereby leading to modifications in bone accrual [8]. For example, ingestion of a glucose-containing solution (75 g) yielded an approximately 50% reduction in bone resorption as measured via C-terminal telopeptide of type 1 collagen (CTX) [7,9–11]. Others have reported similar effects of glucose ingestion on bone resorption, and that glucose-related effects on bone are more pronounced following oral glucose administration compared to routes of glucose administration that bypass the gastrointestinal tract (e.g., intravenous administration) [10,11]. These collective findings suggest that carbohydrate ingestion has a bone anti-resorptive effect, and that gut-mediated mechanisms are likely involved.

Several hormonal mediators of nutrient metabolism are hypothesized to contribute to this “gut-bone axis.” Following macronutrient ingestion, numerous gut-derived hormones are mobilized to regulate satiety, gastric motility, and metabolism [12]. Glucagon-like peptide 1 (GLP-1) and glucose-dependent insulinotropic peptide (GIP) are the main gut-derived hormones or “incretins” that play an integral role in augmenting insulin secretion [13]. *In vitro* studies suggest that the bone-forming osteoblasts and bone-resorbing osteoclasts possess membrane-bound receptors for GIP, GLP-1, and insulin, and experimental studies in humans indicate that these incretins, as well as insulin, promote increases in bone formation and decreases in bone resorption [14–18]. Accordingly, medical conditions with perturbed incretin and/or insulin response to nutrient ingestion might threaten bone health through modifications of the “gut-bone axis.”

This study evaluates the “gut-bone axis” in a sample of emerging and young adults with PI-CF, a genetic condition associated with compromised pulmonary health, impaired nutritional status, and diabetes [19], as well as reduced bone mass and increased fracture risk [20,21]. PI and poor incretin response to food intake are suspected to contribute to CF-related diabetes (CFRD) [22–24], but might also contribute to CF-related bone disease [20,21]. To interrogate how carbohydrate ingestion may influence bone turnover in people with PI-CF, we evaluated changes in biomarkers of bone metabolism following ingestion of an oral glucose solution and tested the relationships of changes in insulin and incretin hormones with biomarkers of bone metabolism. Based on previous studies in adults [10,11,13,25,26], we hypothesized that in adolescents and young adults with PI-CF, bone resorption (assessed via CTX) would decrease significantly following glucose ingestion, but that bone formation (assessed via procollagen 1 intact N-terminal propeptide [P1NP]) would not be affected. Additionally, we hypothesized that GIP, GLP-1, and insulin responses following glucose ingestion are associated with reductions in bone resorption as indicated by greater reductions in CTX.

Methods

Study design and participants

We performed a secondary analysis of data and biological specimens collected during a prospective study designed to test the underpinnings of post-glucose load hypoglycemia in people with PI-CF [27]. Insulin secretory rates and glucagon results from the multi-sample oral glucose tolerance tests (OGTT) conducted in 23 youth and young adults (39% female) ages 14–30 years receiving treatment for CF at the Children’s Hospital of Philadelphia or University of Pennsylvania Medical Center were previously published [27]. A more detailed description of study participants, protocols, and procedures were published previously [27].

The diagnoses of CF and PI were confirmed using clinical records documenting sweat testing and/or cystic fibrosis transmembrane regulatory mutation analysis, and pancreatic enzyme replacement therapy and/or fecal elastase levels <200 µg/g. Exclusion criteria included history of organ transplantation, CFRD with fasting hyperglycemia (fasting

glucose \geq 126 mg/dL), and systemic glucocorticoid therapy within previous four weeks of the study visit.

Anthropometry

Standing height and weight were assessed using a wall-mounted stadiometer and electronic scale, respectively. Body mass index (BMI; kg/m²) was calculated.

Oral glucose tolerance test

All participants completed an OGTT following an overnight fast. Participants ingested a solution containing 1.75 g of glucose per kg of body weight (75 g of glucose maximum) over a 5-minute period, indicating time point “0.” Using an indwelling intravenous catheter, blood specimens were collected at mins –10, –1, 10, 20, 30, and every subsequent 15 mins until 240 mins. For subjects that experienced symptomatic hypoglycemia with glucose < 65 mg/dL, or glucose \leq 50 mg/dL in the absence of symptoms, testing was terminated early. For the present analyses, only data from mins 0 to 120 are included.

Blood biochemistries

At each time point indicated above, glucose, insulin, GIP, and GLP-1 were assayed in duplicate. Glucose was measured using the YSI 2300 glucose analyzer (Yellow Spring Instruments, Yellow Spring, OH) and, for immediate results, the Nova StatStrip glucometer (Data Sciences International, St. Paul, MN). These glucose assessment methods demonstrate strong agreement with one another [27]. Insulin was measured in duplicate using a double antibody radioimmunoassay [28], and active GLP-1 and total GIP were measured in duplicate by ELISA (Millipore, Billerica, Massachusetts). Biomarkers of bone metabolism, CTX and P1NP, were assessed at time points 0, 30, 60, and 120. CTX was evaluated using the Cobas e411 automated analyzer (Roche Diagnostics International Ltd., Basel, Switzerland), and P1NP was evaluated using a commercially available ELISA kit (Antibodies-Online Inc., Limerick, PA).

Calculations

Percent changes (% Δ) in CTX, P1NP, GIP, GLP-1, insulin, and glucose were calculated. Percent change in CTX and P1NP from mins 0 to 120 were computed and were abbreviated as CTX-% $\Delta_{0-120\text{min}}$ and P1NP-% $\Delta_{0-120\text{min}}$, respectively. Percent change in GIP, GLP-1, insulin, and glucose were calculated for mins 0 to 30, 0 to 120, and 30 to 120. As an example, % Δ in GIP from mins 0 to 30 was abbreviated as GIP-% $\Delta_{0-30\text{min}}$. Percent change was calculated as follows: ((measurement 2 – measurement 1) / measurement 1) \times 100. Using data from all available time points, incremental area under the curve (iAUC) was calculated for GIP, GLP-1, insulin, and glucose for mins 0 to 30 and mins 0 to 120 using the trapezoidal method. iAUCs for CTX and P1NP were calculated for mins 0 to 120. As an example, iAUC for CTX and insulin from mins 0 to 120 are abbreviated as CTX-AUC_{0-120mins} and insulin-AUC_{0-120mins}, respectively.

Statistical analyses

Prior to conducting statistical analyses, all data were visually inspected for outliers, biologically implausible data points, and non-normal distributions. Subject descriptive characteristics are summarized using mean (standard deviation) or median (inter-quartile range) for continuous variables and count (percent) for categorical variables.

To evaluate change in glucose, insulin, incretins, and biomarkers of bone turnover during OGTT, mixed regressions were performed. For each analysis, min 0 of the OGTT was used as the comparison time point. Relationships between incretins, metabolic parameters, and age with

biomarkers of bone turnover during OGTT were evaluated using non-parametric Spearman's rank correlation. We also compared % Δ and iAUC for biomarkers of bone metabolism during OGTT between males and females using linear regression.

Various sensitivity analyses were performed. Linear regression was used to assess associations between changes in insulin/incretins and biomarkers of bone metabolism during OGTT while accounting for min 0 values of insulin/incretins. Additional linear regression analyses were performed to assess age and sex interactions in the association between changes in insulin/incretins and biomarkers of bone metabolism during OGTT.

All statistical analyses were performed using STATA (version 15). P-values < 0.05 were considered statistically significant.

Results

Descriptive characteristics

Descriptive characteristics of the study participants were previously reported [27]. Their age ranged from 14.6 to 30.6 years, with an average age of 24.6 ± 4.4 years, and there was a greater proportion of males compared to females (57 % male) and Whites compared to Blacks (96 % White). The BMI of study participants ranged from 17.8 to 27.8 kg/m², with an average BMI of 22.4 ± 2.7 kg/m². Nine percent of participants had normal glucose tolerance (n = 2; 1-hour glucose \leq 155 mg/dL and 2-hour glucose < 140 mg/dL), 35 % had early glucose intolerance (n = 8; 1-hour glucose \geq 155 mg/dL and 2-hour glucose < 140 mg/dL), 43 % had impaired glucose tolerance (n = 10; 2-hour glucose 140–199 mg/dL), and 13 % had diabetes (n = 3; 2-hour glucose \geq 200 mg/dL).

Changes in incretins and metabolic outcomes during OGTT

Changes in glucose, insulin, GLP-1, and GIP during OGTT in the total study sample are presented in Fig. 1. Glucose, insulin, GLP-1, and GIP increased during OGTT (all P < 0.001). Glucose and insulin increased until mins 60 and 90, respectively, and declined thereafter. GLP-1 and GIP increased similarly from mins 0 to 30. While GIP remained increased from baseline until min 120, GLP-1 returned to baseline by min 120.

CTX and P1NP during OGTT are displayed in Fig. 2. CTX decreased significantly during OGTT (P < 0.05). CTX at mins 30 (P = 0.39) and 60 (P = 0.050) were not significantly different than min 0, but CTX at min 120 was significantly lower than min 0 (P = 0.004). When expressed in units of % Δ , CTX decreased significantly between mins 0 and 30 (P = 0.004), 0 and 60 (P < 0.001), and 0 and 120 (P < 0.001). From mins 0 to 120, CTX decreased by an average of 32%, ranging from approximately -5% to -62%. P1NP responses varied from mins 0 to 120, ranging from approximately +66% to -33%, with a mean change of about +1%.

Correlations between incretin and metabolic responses and biomarkers of bone turnover during OGTT

Bivariate correlations between % Δ changes in incretins and metabolic parameters and CTX during OGTT are presented in Table 1. GIP-% $\Delta_{0-30\text{min}}$ was negatively correlated with CTX-% $\Delta_{0-120\text{min}}$, suggesting that individuals with greatest increases in GIP from mins 0 to 30 had the greatest reductions in CTX from mins 0 to 120 (Fig. 3). GIP-% $\Delta_{30-120\text{min}}$ was positively correlated with CTX-% $\Delta_{0-120\text{min}}$, suggesting that individuals with the greatest decreases in GIP from mins 30 to 120 had the greatest reductions in CTX from mins 0 to 120. GLP-1-% $\Delta_{30-120\text{min}}$ was negatively correlated with CTX-% $\Delta_{0-120\text{min}}$, but this relationship did not meet significance (P = 0.061; Supplemental Fig. 1). Insulin-% $\Delta_{30-120\text{min}}$ was positively correlated with CTX-% $\Delta_{0-120\text{min}}$, but this relationship was not significant (P = 0.105; Supplemental Fig. 2). Percent change in glucose, insulin, and incretins were not correlated with P1NP during OGTT (data not shown).

Bivariate correlations between iAUCs for incretins and metabolic

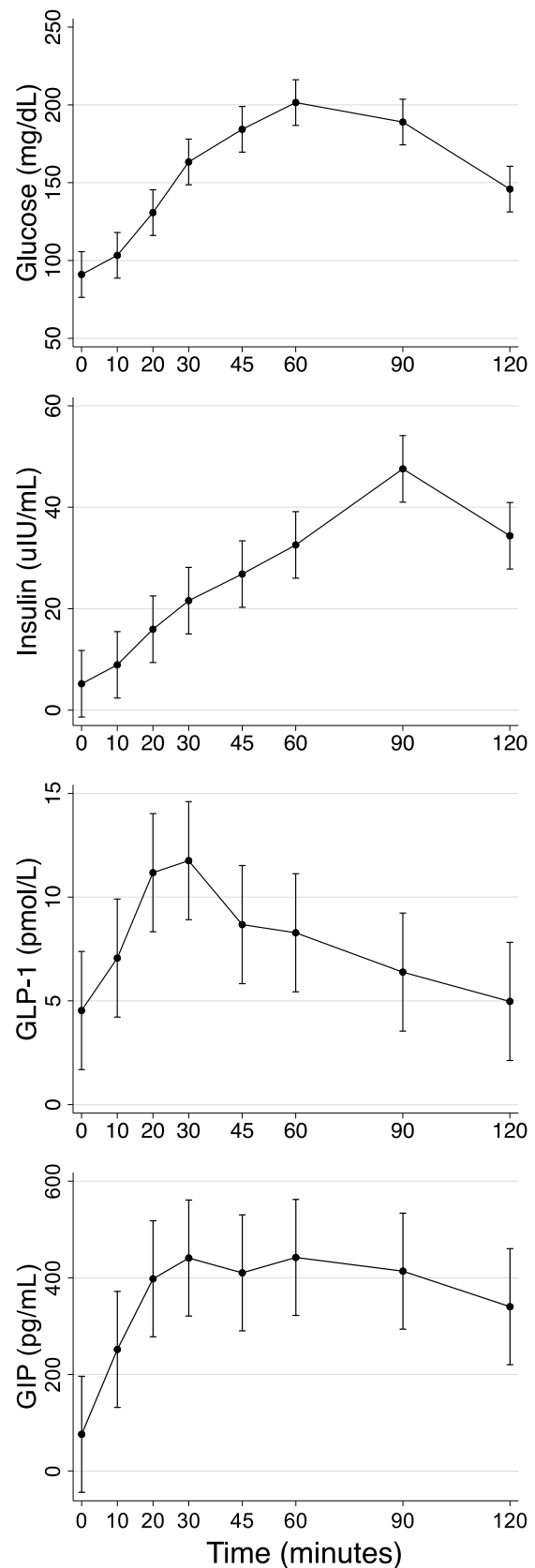


Fig. 1. Changes in glucose, insulin, GLP-1, and GIP during OGTT in youth with PI-CF. Vertical bands represent the 95% confidence interval. OGTT, oral glucose tolerance test; CF, cystic fibrosis.

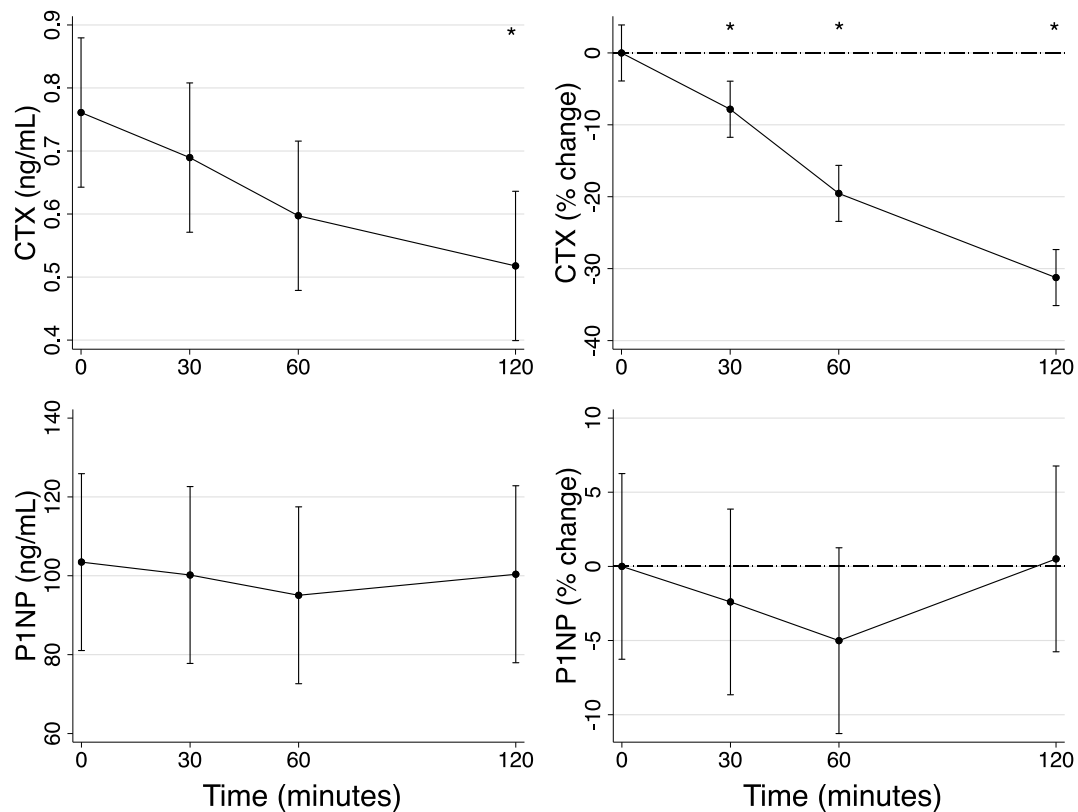


Fig. 2. CTX (top) and P1NP (bottom) % Δ during OGTT in youth with PI-CF. Vertical bands represent the 95% confidence interval. *Significantly different than minute 0. % Δ , percent change; CTX, C-terminal telopeptide of type 1 collagen.

Table 1

Associations between % Δ in incretin and metabolic parameters and CTX (minutes 0–120) at varying timepoints during OGTT.

	Time (minutes)	Spearman's rho	P
GIP	0–30	−0.54	0.008
	0–120	−0.16	0.471
	30–120	0.50	0.016
GLP-1 ^a	0–30	−0.25	0.268
	0–120	0.06	0.805
	30–120	0.40	0.061
Glucose	0–30	−0.14	0.517
	0–120	−0.03	0.879
	30–120	−0.01	0.964
Insulin	0–30	−0.16	0.452
	0–120	0.15	0.506
	30–120	0.35	0.105

% Δ , percent change; CTX, C-terminal telopeptide of type 1 collagen; OGTT, oral glucose tolerance test; GIP, glucose-dependent insulinotropic peptide; GLP-1, glucagon-like peptide 1.

^a n = 22.

parameters and CTX during OGTT are presented in **Supplemental Table 1**. Insulin-iAUC₀₋₃₀ was negatively correlated with CTX-iAUC_{0-120min}, but this relationship was not statistically significant ($P = 0.063$). Glucose, GIP, and GLP-1 iAUCs were not correlated with CTX-iAUC_{0-120min}. Glucose, insulin, GIP, and GLP-1 iAUCs were also not significantly correlated with P1NP-iAUC_{0-120min} (data not shown).

Correlations between age and sex with biomarkers of bone turnover during OGTT

Age correlated positively with CTX-% Δ _{0-120min} ($\rho = 0.34$, $P = 0.105$) and CTX-iAUC_{0-120min} ($\rho = 0.40$, $P = 0.061$), but relationships were not significant. Age did not correlate with changes in P1NP during

OGTT (data not shown). Additionally, CTX-iAUC_{0-120min} differed between males and females (males: -22.9 ± 12.6 , females: -8.9 ± 6.7 ; $P = 0.005$), and this difference persisted after adjusting for age ($P = 0.011$). Changes in P1NP during OGTT did not differ between males and females.

Sensitivity analyses

To determine whether relationships between changes in insulin, GIP, and GLP-1 and changes in CTX and P1NP were confounded by min 0 values of insulin or incretin hormones, linear regression analyses were performed. Associations remained consistent when including min 0 values for the respective predictor variable (insulin, GIP, or GLP-1) as an additional regression model parameter (data not shown). Additionally, no age or sex interactions with insulin, GIP, or GLP-1 were found in the relationships of these hormones with bone turnover markers (data not shown).

Discussion

The “gut-bone axis” is a hypothesized contributor to nutrition and diabetes-related effects on bone [3,7,13,29]. Experimental studies suggest that consumption of macronutrients results in acute anti-resorptive effects on bone, and that hormones involved in nutrient metabolism, such as the gut-derived incretins GIP and GLP-1, as well as insulin contribute to these effects [9,30–32]. Cystic fibrosis is associated with a unique form of diabetes that involves deranged incretin and insulin responses to macronutrient consumption [33,34]. Deficits in bone density and increased fracture risk are reported in patients with CF [35,36]; thus, invoking disruption to the “gut-bone axis” as a plausible contributor to CF related bone disease. The current study addresses these gaps in knowledge by evaluating the “gut-bone axis” in a sample of adolescents and young adults with PI-CF. Similar to prior studies in healthy

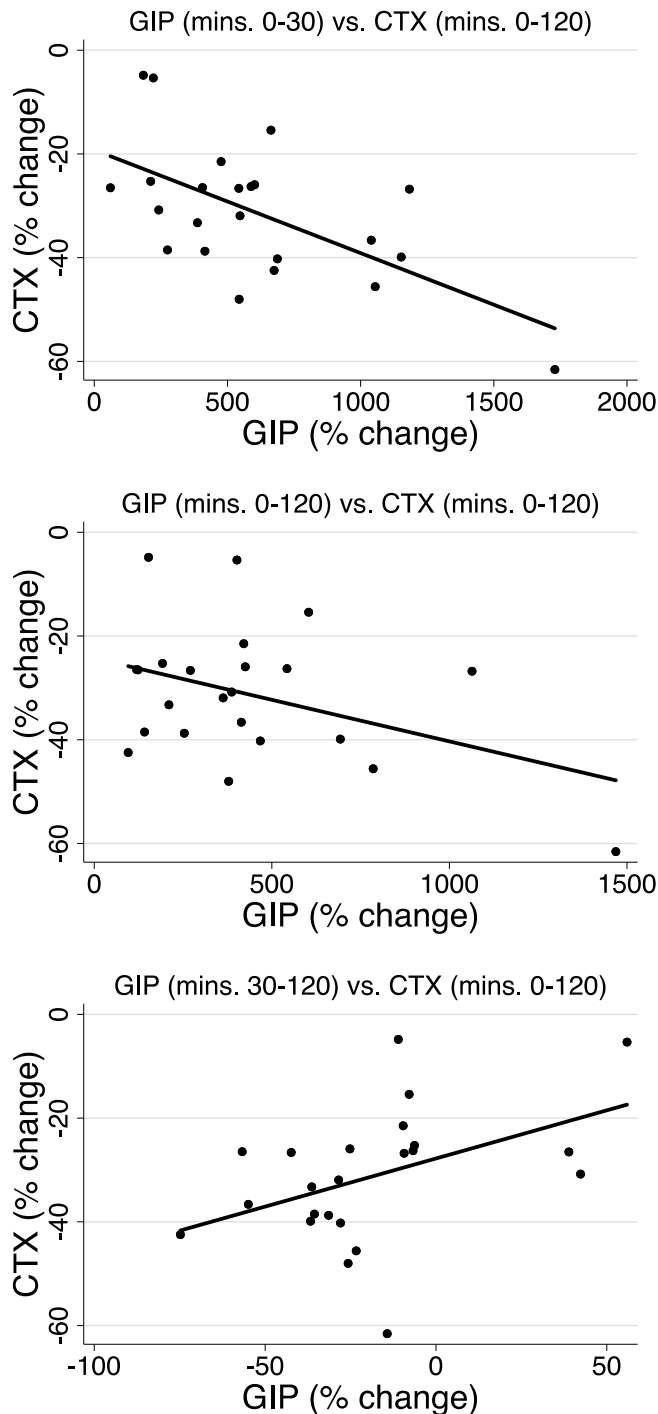


Fig. 3. Associations between $\% \Delta$ in GIP and CTX during OGTT in youth with PI-CF. $\% \Delta$, percent change; CTX, C-terminal telopeptide of type 1 collagen.

adults [10,13,25,26], our data suggest that acute glucose ingestion results in a significant decrease in CTX, a biomarker of bone resorption, but did not have an effect on P1NP, a biomarker of bone formation. Our data also suggest that the incretin hormones, notably GIP, might contribute to these changes in bone metabolism following glucose ingestion. Interpretation of our findings is limited by the lack of a healthy control group and individuals with pancreatic sufficient CF. The pathophysiology of CFRD includes a perturbed incretin and insulin response to nutrient intake [37,38]. These data support the need for future research involving the “gut-bone axis” in the convergence of diabetes and bone disease in CF.

Previous data suggest that the bone forming osteoblasts and bone resorbing osteoclasts are responsive to consumption of macronutrients [39], resulting in a shift in the balance between bone formation and resorption, predominantly by decreasing bone resorption [29]. Consumption of a fixed amount of carbohydrate (75 g of glucose) in adults results in a significant reduction in CTX, a well-characterized biomarker of bone resorption, by 120 mins following ingestion [7,9,10]. We also found significant decreases in CTX following glucose ingestion in our study, as demonstrated by a median $\text{CTX-}\% \Delta_{0-120\text{min}}$ of about -32% by min 120, which is lesser in magnitude than the approximately 50% reduction in CTX by min 120 of OGTT that has been consistently reported previously in healthy adults [7,9,10]. Other studies comparing bone metabolism following macronutrient ingestion have reported suppressed effects on bone resorption in adults with hypothyroidism, nonalcoholic fatty liver disease, and type 2 diabetes compared to healthy controls [10,25,40]. Together, these studies suggest that the biological responses of bone to macronutrient and/or food consumption might be modified in individuals with chronic health conditions that exert more global effects on metabolism.

Our study sample included adolescents and young adults with PI-CF, a condition which leads to impaired nutrient metabolism and nutritional status [41]. Endocrine pancreatic dysfunction is also common in people with CF, and patients with CF are at increased risk for developing diabetes [19]. CF-related diabetes is at least in part attributed to progressive pancreatic β -cell functional decline resulting in abnormal insulin secretion, including a diminished early-phase insulin response followed by a pronounced late-phase insulin response [27]. In addition to dysglycemia, the perturbed insulin response might also impact bone health since insulin inhibits osteoclastogenesis [42], which might contribute to decreased bone resorption during insulin infusion, as reported in a mouse model of type 1 diabetes [43]. In the current study, late-phase insulin was marginally associated with changes in CTX, suggesting that bone-augmenting effects of insulin might be perturbed in youth with the greatest risk of developing CFRD. The metabolic health status among our study sample was variable, ranging from normal glucose tolerance to diabetes. The small sample size prevented comparisons across groups based on glucose control.

Our data suggest a potential role of incretins in decreasing bone resorption, as a greater increase in GIP from mins 0 to 30 and a greater decrease in GIP from mins 30 to 120 was associated with more pronounced reductions in CTX. In contrast, greater decreases in GLP-1 from mins 30 to 120 were only marginally associated with decreases in CTX. These data are consistent with findings of others that infusion of GIP and/or GLP-1 significantly decreases bone resorption [13,26], and that GIP and GLP-1 receptor knockout mice have decreased bone mass [15,44]. ‘Incretin mimetic’ drugs (e.g., GLP-1 receptor agonists) increase insulin and decrease glucagon production in the pancreas, and ‘dipeptidyl peptidase-4 (DPP-4) inhibitors’ reduce degradation of incretin hormones, GIP and GLP-1, by the DPP-4 enzyme [45]. While insulin therapy is the first-line treatment for CFRD [46], recent data suggest a potential benefit of incretin-based therapies on insulin secretion following a meal [47]. Incretin-based therapies are suspected to benefit bone health [48], but effects on the gut-bone axis have not yet been explored in CF.

Most studies evaluating acute effects of nutrient ingestion on bone metabolism reported decreases in bone resorption, but inconsistent responses in bone formation [10,11,13,25,26,30,49]. Our data are consistent with these studies; change in P1NP following glucose ingestion was highly variable. In fact, most studies evaluating bone formation via P1NP reported null effects [10,11,13,25,26], as did other studies utilizing alternate bone formation markers such as osteocalcin [11,49,50]. Extended experimental protocols and inclusion of complementary biomarkers of bone formation should be considered in future studies.

Few studies address age or sex-related differences in acute changes in bone metabolism following macronutrient ingestion [51,52]. Bone

modeling, which involves an uncoupling of bone formation and resorption, is dominant during the years preceding peak bone mass, whereas bone remodeling, which involves a coordination between bone formation and resorption, is dominant in adulthood. Thus, biological effects of macronutrient ingestion on bone are likely dependent on both age and sex. We only found a marginal and not statistically significant association of younger age with greater decreases in CTX following glucose ingestion, and that males had a significantly greater CTX iAUC compared to females. CF-related complications such as diabetes and bone disease increase with age, and perhaps these complications involve the gut-bone axis. Further, bone metabolism occurs at a more accelerated rate in childhood and adolescence compared to young adulthood [52], suggesting that the gut-bone axis is amplified during the growing years and might contribute to sex differences in bone density and fracture.

Strengths and limitations

This study used previously acquired data and blood specimens to investigate a novel biological mechanism that likely contributes to nutrition and diabetes effects on bone health in a vulnerable population during the years surrounding peak bone mass attainment. Although the original study included a healthy control group to compare against individuals with PI-CF [27], blood specimens for assessment of incretins and bone biomarkers were not available from the healthy control group for the current study. This represents a main limitation of this study. Additionally, all participants had exocrine PI necessitating pancreatic enzyme replacement therapy. PI is among the most common complication of CF, occurring in upwards of 85% of patients [53]. To understand the contribution of maldigestion resulting from PI, future studies in people with PI-CF should also consider including individuals with normal pancreatic function. Moreover, about half of our sample had reactive hypoglycemia during the OGTT [27], which might have contributed to the wide variation in bone responses to glucose ingestion. Coupled with the small sample size, the variability in incretin, insulin, and bone responses to glucose ingestion likely limited our statistical power.

The OGTT method used in this study has both strengths and limitations. Our use of a standardized 75 g OGTT, performed following an overnight fast, helped minimize confounding resulting from nocturnal and daily variation in metabolic response to food ingestion, and also helped facilitate comparisons with previously published studies that followed a similar approach [7,9–11]. Rather than ingesting single nutrients in isolation, humans consume mixtures of foods and nutrients while engaging in normal activities such as sleep, exercise, and sedentary behavior. Specific to CF, high-fat diets are often recommended to ensure adequate caloric intake [54]. Future studies involving the gut-bone axis in CF should apply translatable experimental approaches (e.g., high-fat mixed meal tolerance tests) while encompassing normal nocturnal and daily variation in components of the gut-bone axis.

Conclusions

This study provides novel insights into the role of nutrition on influencing peak bone mass and the development of CF-related bone disease in adolescents and young adults with PI-CF. We report significant decreases in bone resorption following glucose ingestion, and a potential underlying role of incretin hormones and insulin in these antiresorptive effects. Although our study is limited by the lack of a healthy control group, these novel data give insight into potential mechanisms linking complex co-morbidities in CF, namely diabetes and bone disease. Since people with CF typically consume a high-fat diet that might contribute to these developments, the impact of habitual dietary intake on the gut-bone axis warrants consideration. Furthermore, future studies should seek to define the “normal” post-prandial changes in bone metabolism during the years surrounding peak bone mass, notably with respect to age, sex, race, and puberty.

Funding: This work was supported by the Cystic Fibrosis Foundation

grants KILBER18B0 (M.J.K.), KILBER19D0 (M.J.K.), and LEI22H0 (W.S.L.); the Pediatric Endocrine Society (M.J.K.); and Public Health Services Research grants R01DK97830 (A.K.), UL1TR001878 (CHOP Center for Human Phenomic Science), DK19525 (University of Pennsylvania Diabetes Research Center), UL1TR002378, and KL2TR002381.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

The authors would like to acknowledge the individuals with CF, nursing staff, and laboratory staff at the Penn and CHOP Center for Human Phenomic Science for their contribution to the original project.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.jcte.2022.100304>.

References

- [1] Gordon CM, et al. “The Determinants of Peak Bone Mass,” (in eng). *J Pediatr Jan* 2017;180:261–9. <https://doi.org/10.1016/j.jpeds.2016.09.056>.
- [2] Weaver CM. “The role of nutrition on optimizing peak bone mass,” (in eng). *Asia Pac J Clin Nutr* 2008;17(Suppl 1):135–7.
- [3] S. P. Schiellerup et al., “Gut Hormones and Their Effect on Bone Metabolism. Potential Drug Therapies in Future Osteoporosis Treatment,” (in eng). *Front Endocrinol (Lausanne)*, vol. 10, p. 75, 2019, 10.3389/fendo.2019.00075.
- [4] Soyka LA, Fairfield WP, Klibanski A. Hormonal Determinants and Disorders of Peak Bone Mass in Children. *The Journal of Clinical Endocrinology & Metabolism* 2000;85(11):3951–63. <https://doi.org/10.1210/jcem.85.11.6994>.
- [5] Levine MA. “Assessing bone health in children and adolescents,” (in eng). *Indian J Endocrinol Metab Dec* 2012;16(Suppl 2):S205–12. <https://doi.org/10.4103/2230-8210.104040>.
- [6] Redmond J, Fulford AJ, Jarjou L, Zhou B, Prentice A, Schoenmakers I. “Diurnal Rhythms of Bone Turnover Markers in Three Ethnic Groups,” (in eng). *The Journal of clinical endocrinology and metabolism* 2016;101(8):3222–30. <https://doi.org/10.1210/jc.2016-1183>.
- [7] N. H. Bjarnason, E. E. G. Henriksen, P. Alexandersen, S. Christgau, D. B. Henriksen, and C. Christiansen, “Mechanism of circadian variation in bone resorption: 1 Data from studies contained in this report were submitted in abstract form at the 22nd meeting of the American Society of Bone and Mineral Research, Toronto, Ontario, Canada, September 2000,” *Bone*, vol. 30, no. 1, pp. 307–313, 2002/01/01/ 2002, 10.1016/S8756-3282(01)00662-7.
- [8] Hassager C, Risteli J, Risteli L, Jensen SB, Christiansen C. “Diurnal variation in serum markers of type I collagen synthesis and degradation in healthy premenopausal women,” (in eng). *J Bone Miner Res Nov* 1992;7(11):1307–11. <https://doi.org/10.1002/jbmr.5650071110>.
- [9] Henriksen DB, et al. “Role of gastrointestinal hormones in postprandial reduction of bone resorption,” (in eng). *J Bone Miner Res Dec* 2003;18(12):2180–9. <https://doi.org/10.1359/jbmr.2003.18.12.2180>.
- [10] Westberg-Rasmussen S, et al. Differential impact of glucose administered intravenously or orally on bone turnover markers in healthy male subjects. *Bone Apr* 2017;97:261–6. <https://doi.org/10.1016/j.bone.2017.01.027>.
- [11] Maagensen H, Junker AE, Jørgensen NR, Gluud LL, Knop FK, Vilsbøll T. Bone Turnover Markers in Patients With Nonalcoholic Fatty Liver Disease and/or Type 2 Diabetes During Oral Glucose and Isoglycemic Intravenous Glucose. *The Journal of Clinical Endocrinology & Metabolism* 2018;103(5):2042–9. <https://doi.org/10.1210/jc.2018-00176>.
- [12] Martin AM, Sun EW, Keating DJ. “Mechanisms controlling hormone secretion in human gut and its relevance to metabolism,” (in eng). *The Journal of endocrinology* 2019;244(1):R1–15. <https://doi.org/10.1530/JOE-19-0399>.
- [13] Helsted MM, et al. The role of endogenous GIP and GLP-1 in postprandial bone homeostasis. *Bone* 2020;11/01/ 2020;140:115553. <https://doi.org/10.1016/j.bone.2020.115553>.
- [14] Tsukiyama K, et al. “Gastric inhibitory polypeptide as an endogenous factor promoting new bone formation after food ingestion,” (in eng). *Mol Endocrinol Jul* 2006;20(7):1644–51. <https://doi.org/10.1210/me.2005-0187>.
- [15] Yamada C, et al. “The murine glucagon-like peptide-1 receptor is essential for control of bone resorption,” (in eng). *Endocrinology Feb* 2008;149(2):574–9. <https://doi.org/10.1210/en.2007-1292>.
- [16] Fulzele K, et al. “Insulin receptor signaling in osteoblasts regulates postnatal bone acquisition and body composition,” (in eng). *Cell* 2010;142(2):309–19. <https://doi.org/10.1016/j.cell.2010.06.002>.

- [17] E. Ceccarelli et al., "Beyond glycemic control in diabetes mellitus: effects of incretin-based therapies on bone metabolism," (in eng), *Frontiers in endocrinology*, vol. 4, pp. 73-73, 2013, doi: 10.3389/fendo.2013.00073.
- [18] Dicembrini I, Mannucci E, Rotella CM. "Bone: incretin hormones perceiver or receiver?," (in eng). *Exp Diabetes Res* 2012;2012:519784. <https://doi.org/10.1155/2012/519784>.
- [19] Moran A, et al. Clinical Care Guidelines for Cystic Fibrosis-Related Diabetes. *Diabetes Care* 2010;33(12):2697. <https://doi.org/10.2337/dc10-1768>.
- [20] Stalvey MS, Clines GA. "Cystic fibrosis-related bone disease: insights into a growing problem," (in eng). *Curr Opin Endocrinol Diabetes Obes* 2013;20(6):547-52. <https://doi.org/10.1097/01.med.0000436191.87727.ec>.
- [21] Aris RM, et al. Guide to Bone Health and Disease in Cystic Fibrosis. The Journal of Clinical Endocrinology & Metabolism 2005;90(3):1888-96. <https://doi.org/10.1210/jc.2004-1629>.
- [22] Granados A, Chan CL, Ode KL, Moheet A, Moran A, Holl R. Cystic fibrosis related diabetes: Pathophysiology, screening and diagnosis. *J Cyst Fibros* 2019/10/01/2019;18:S3-9. <https://doi.org/10.1016/j.jcf.2019.08.016>.
- [23] Sheikh S, et al. Reduced β -Cell Secretory Capacity in Pancreatic-Insufficient, but Not Pancreatic-Sufficient, Cystic Fibrosis Despite Normal Glucose Tolerance. *Diabetes* 2017;66(1):134. <https://doi.org/10.2337/db16-0394>.
- [24] Hillman M, Eriksson L, Mared L, Helgesson K, Landin-Olsson M. Reduced levels of active GLP-1 in patients with cystic fibrosis with and without diabetes mellitus. *J Cyst Fibros* 2012/03/01/2012;11(2):144-9. <https://doi.org/10.1016/j.jcf.2011.11.001>.
- [25] Yavropoulou MP, et al. Response of biochemical markers of bone turnover to oral glucose load in diseases that affect bone metabolism. *Eur J Endocrinol* Jun 2011;164(6):1035-41. <https://doi.org/10.1530/EJE-11-0128>.
- [26] Bergmann NC, et al. Separate and Combined Effects of GIP and GLP-1 Infusions on Bone Metabolism in Overweight Men Without Diabetes. *The Journal of Clinical Endocrinology & Metabolism* 2019;104(7):2953-60. <https://doi.org/10.1210/jc.2019-00008>.
- [27] M. J. Kilberg et al., "Hypoglycemia and Islet Dysfunction Following Oral Glucose Tolerance Testing in Pancreatic-Insufficient Cystic Fibrosis," *J Clin Endocrinol Metab*, vol. 105, no. 10, Oct 1 2020, 10.1210/clinem/dgaa448.
- [28] Bak MJ, et al. "Specificity and sensitivity of commercially available assays for glucagon and oxyntomodulin measurement in humans," (in eng). *Eur J Endocrinol* Apr 2014;170(4):529-38. <https://doi.org/10.1530/eje-13-0941>.
- [29] A. Nissen et al., "A Pilot Study Showing Acute Inhibitory Effect of GLP-1 on the Bone Resorption Marker CTX in Humans," *JBM R Plus*, <https://doi.org/10.1002/jbm4.10209> vol. 3, no. 10, p. e10209, 2019/10/01 2019, 10.1002/jbm4.10209.
- [30] Fuglsang-Nielsen R, et al. Consumption of nutrients and insulin resistance suppress markers of bone turnover in subjects with abdominal obesity. *Bone* Apr 2020;133:115230. <https://doi.org/10.1016/j.bone.2020.115230>.
- [31] U. Razny et al., "High Fat Mixed Meal Tolerance Test Leads to Suppression of Osteocalcin Decrease in Obese Insulin Resistant Subjects Compared to Healthy Adults," *Nutrients*, vol. 10, no. 11, Nov 1 2018, 10.3390/nu10111611.
- [32] Valderas JP, Padilla O, Solari S, Escalona M, Gonzalez G. Feeding and bone turnover in gastric bypass. *J Clin Endocrinol Metab* Feb 2014;99(2):491-7. <https://doi.org/10.1210/jc.2013-1308>.
- [33] Kilberg MJ, et al. Dysregulated insulin in pancreatic insufficient cystic fibrosis with post-prandial hypoglycemia. *J Cyst Fibros* Mar 2020;19(2):310-5. <https://doi.org/10.1016/j.jcf.2019.07.006>.
- [34] Frost F, Jones GH, Dyce P, Jackson V, Nazareth D, Walshaw MJ. "Loss of incretin effect contributes to postprandial hyperglycaemia in cystic fibrosis-related diabetes," (in eng). *Diabet Med* Nov 2019;36(11):1367-74. <https://doi.org/10.1111/dme.14121>.
- [35] Aris RM, et al. "Increased rate of fractures and severe kyphosis: sequelae of living into adulthood with cystic fibrosis," (in eng). *Ann Intern Med* 1998;128(3):186-93. <https://doi.org/10.7326/0003-4819-128-3-199802010-00004>.
- [36] Stahl M, et al. Multiple prevalent fractures in relation to macroscopic bone architecture in patients with cystic fibrosis. *J Cyst Fibros* 2018;17(1):114-20. <https://doi.org/10.1016/j.jcf.2016.06.004>.
- [37] Kelly A, Moran A. "Update on cystic fibrosis-related diabetes," (in eng). *J Cyst Fibros* Jul 2013;12(4):318-31. <https://doi.org/10.1016/j.jcf.2013.02.008>.
- [38] K. Kayani, R. Mohammed, and H. Mohiaddin, "Cystic Fibrosis-Related Diabetes," (in eng), *Front Endocrinol (Lausanne)*, vol. 9, p. 20, 2018, 10.3389/fendo.2018.00020.
- [39] J. A. Clowes, S. Khosla, and R. Eastell, "Potential Role of Pancreatic and Enteric Hormones in Regulating Bone Turnover," *Journal of Bone and Mineral Research*, <https://doi.org/10.1359/JBMR.050524> vol. 20, no. 9, pp. 1497-1506, 2005/09/01 2005, <https://doi.org/10.1359/JBMR.050524>.
- [40] Chailurkit LO, Chanprasertyothin S, Rajatanavin R, Ongphiphadhanakul B. "Reduced attenuation of bone resorption after oral glucose in type 2 diabetes," (in eng). *Clin Endocrinol (Oxf)* Jun 2008;68(6):858-62. <https://doi.org/10.1111/j.1365-2265.2007.03159.x>.
- [41] J. Ockenga, "Importance of nutritional management in diseases with exocrine pancreatic insufficiency," (in eng), *HPB (Oxford)*, vol. 11 Suppl 3, no. Suppl 3, pp. 11-5, Dec 2009, 10.1111/j.1477-2574.2009.00134.x.
- [42] Xu F, Ye YP, Dong YH, Guo FJ, Chen AM, Huang SL. "Inhibitory effects of high glucose/insulin environment on osteoclast formation and resorption in vitro," (in eng). *J Huazhong Univ Sci Technol Med Sci* Apr 2013;33(2):244-9. <https://doi.org/10.1007/s11596-013-1105-z>.
- [43] J. S. Nyman, E. Kalaitzoglou, R. Clay Bunn, S. Uppuganti, K. M. Thraikill, and J. L. Fowlkes, "Preserving and restoring bone with continuous insulin infusion therapy in a mouse model of type 1 diabetes," (in eng), *Bone Rep*, vol. 7, pp. 1-8, Dec 2017, 10.1016/j.bonr.2017.07.001.
- [44] Xie D, et al. "Glucose-dependent insulinotropic polypeptide receptor knockout mice have altered bone turnover," (in eng). *Bone* Dec 2005;37(6):759-69. <https://doi.org/10.1016/j.bone.2005.06.021>.
- [45] Sharma D, Verma S, Vaidya S, Kalia K, Tiwari V. Recent updates on GLP-1 agonists: Current advancements & challenges. *Biomed Pharmacother* 2018/12/01/2018;108:952-62. <https://doi.org/10.1016/j.biopha.2018.08.088>.
- [46] G. M. Onady and A. Stolfi, "Drug treatments for managing cystic fibrosis-related diabetes," (in eng), *Cochrane Database Syst Rev*, vol. 10, no. 10, p. Cd004730, Oct 19 2020, 10.1002/14651858.CD004730.pub5.
- [47] Kelly A, et al. "Effect of Sitagliptin on Islet Function in Pancreatic Insufficient Cystic Fibrosis With Abnormal Glucose Tolerance," (in eng). *J Clin Endocrinol Metab* 2021;106(9):2617-34. <https://doi.org/10.1210/clinem/dgab365>.
- [48] Chakhtoura M, Azar ST. "Incretin based therapies: bone protective effects," (in eng). *Endocr Metab Immune Disord Drug Targets* Dec 2013;13(4):289-94. <https://doi.org/10.2174/18715303113136660046>.
- [49] Paldanius PM, et al. "The effect of oral glucose tolerance test on serum osteocalcin and bone turnover markers in young adults," (in eng). *Calcif Tissue Int* Feb 2012;90(2):90-5. <https://doi.org/10.1007/s00223-011-9551-8>.
- [50] J. Starup-Linde et al., "Glucose Tolerance Tests and Osteocalcin Responses in Healthy People," (in English), *Frontiers in Endocrinology*, Brief Research Report vol. 9, no. 356, 2018-July-13 2018, 10.3389/fendo.2018.00356.
- [51] Sherk VD, Schauer I, Shah VN. "Update on the Acute Effects of Glucose, Insulin, and Incretins on Bone Turnover In Vivo," (in eng). *Curr Osteoporos Rep* Aug 2020;18(4):371-7. <https://doi.org/10.1007/s11914-020-00598-z>.
- [52] Kindler JM, et al. "Bone geometry and microarchitecture deficits in children with Alagille syndrome," (in eng). *Bone* Dec 2020;141:115576. <https://doi.org/10.1016/j.bone.2020.115576>.
- [53] V. K. Singh and S. J. Schwarzenberg, "Pancreatic insufficiency in Cystic Fibrosis," *Journal of Cystic Fibrosis*, vol. 16, pp. S70-S78, 2017/11/01/2017, 10.1016/j.jcf.2017.06.011.
- [54] D. Borowitz, R. D. Baker, and V. Stallings, "Consensus Report on Nutrition for Pediatric Patients With Cystic Fibrosis," *Journal of Pediatric Gastroenterology and Nutrition*, vol. 35, no. 3, pp. 246-259, 2002. [Online]. Available: https://journals.lww.com/jpgn/Fulltext/2002/09000/Consensus_Report_on_Nutrition_for_Pediatric.4.aspx.