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Evolution and connectivity in the world-wide migration system of the mallard: Inferences from mitochondrial DNA

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Abstract

Background: Main waterfowl migration systems are well understood through ringing activities. However, in mallards (*Anas platyrhynchos*) ringing studies suggest deviations from general migratory trends and traditions in waterfowl. Furthermore, surprisingly little is known about the population genetic structure of mallards, and studying it may yield insight into the spread of diseases such as Avian Influenza, and in management and conservation of wetlands. The study of evolution of genetic diversity and subsequent partitioning thereof during the last glaciation adds to ongoing discussions on the general evolution of waterfowl populations and flyway evolution. Hypothesised mallard flyways are tested explicitly by analysing mitochondrial mallard DNA from the whole northern hemisphere.

Results: Phylogenetic analyses confirm two mitochondrial mallard clades. Genetic differentiation within Eurasia and North-America is low, on a continental scale, but large differences occur between these two land masses ($F_{ST} = 0.51$). Half the genetic variance lies within sampling locations, and a negligible portion between currently recognised waterfowl flyways, within Eurasia and North-America. Analysis of molecular variance (AMOVA) at continent scale, incorporating sampling localities as smallest units, also shows the absence of population structure on the flyway level. Finally, demographic modelling by coalescence simulation proposes a split between Eurasia and North-America 43,000 to 74,000 years ago and strong population growth (~100fold) since then and little migration (not statistically different from zero).

Conclusions: Based on this first complete assessment of the mallard's world-wide population genetic structure we confirm that no more than two mtDNA clades exist. Clade A is characteristic for Eurasia, and clade B for North-America although some representatives of clade A are also found in North-America. We explain this pattern by evaluating competing hypotheses and conclude that a complex mix of historical, recent and anthropogenic factors shaped the current mallard populations. We refute population classification based on flyways proposed by ornithologists and managers, because they seem to have little biological meaning. Our results have implications for wetland management and conservation, with special regard to the release of farmed mallards for hunting, as well as for the possible transmission of Avian Influenza by mallards due to migration.

Background

The large-scale migration systems of temperate waterfowl (Anseriformes:Anatidae) have been extensively studied using ringing, telemetry, morphometrics, radar tracking and isotope analysis [1]. In general, migration

routes are clearly defined [2]. Most have north-south bearings, with populations travelling between northerly breeding areas and more southerly non-breeding areas. Many species follow similar routes and decades of studies on bird migration have led to the delineation of major waterfowl 'flyways' [2-4]. Especially in North-America, flyways are managerial units created by agreements between adjoining states and provinces, and thus are bounded by management requirements. The

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boundaries of true migratory pathways (in a population ecological sense) are much fuzzier. Populations and individuals within species may occupy different flyways, and many migrants are flexible in migration routing [5], even though these migration routes have been in place for relatively long periods of time [6,7]. Especially in ducks, 'irregularities' in migration routes have been described, such as individuals switching migratory trajectories, termed 'abmigration' [8] or 'flyway permeability' [9].

The mallard (*Anas platyrhynchos*) is the most numerous Holarctic waterfowl species and distributed widely over the whole Northern Hemisphere. Northerly breeding birds are mostly migratory, wintering much further south, while birds breeding in temperate regions, especially in parts of Western Europe, can be resident [2]. Migratory mallard populations often do not exhibit clearly defined routes, even when breeding and non-breeding destinations are thousands of kilometres distant [10]. Mallards play a significant role in the management and conservation of wetland habitats [11], are a very common bird in recreational wetland parks as well as the major game species in wetland systems. Hunting of mallards is facilitated in many countries by supplementary restocking wild populations with farmed mallards [12], possibly with large-scale consequences for the population genetic structure, genetic integrity and fitness of the wild populations [13].

Due to its wide range and large population sizes, the mallard is considered the primary natural reservoir of avian influenza (AI) [14]. It has been identified (along with the black-headed gull *Larus ridibundus*) as the species posing the highest risk to transmit AI to farm-birds [15]. Mallards may (among other bird species) contribute to the spread of AI from Eurasia (Old World; OW) into North-America (New World; NW). Some studies propose trans-hemispheric movements of dabbling ducks between these land masses [16] and that these facilitate AI transmission [17,18] (but see [19,20]). The mallard is thus a prime candidate for spreading AI in the wild [21], and perhaps to humans, and it is important to learn more about its ecology, movements and dispersion [22,23]. Analyses of mitochondrial DNA (mtDNA) show just two genetic clades: clade A, mainly found in Asia, and clade B, found in North-America [10,24], indicating widespread mixing within but not between the Old and New World. However, to date no genetic studies have been carried out for the complete native range of mallard, as data from Europe and eastern North America were missing. A range-wide survey of genetic diversity and connectivity between proposed mallard flyways could thus be useful to finally generalise findings of small-scale population genetic and phylogeographic studies. Such a study would also add substance to an increasing body of literature on migration systems, flyways concepts, and genetic population structure in other waterfowl.

In this study, we attempt to close existing gaps in the knowledge of the genetic structure of female mallards by providing large numbers of mtDNA sequences from previously unsampled regions throughout the whole native distribution of the mallard. Due to the mallard's high mobility and examples of flyway permeability in closely related duck species, we hypothesise weak barriers to dispersal between Asia and Europe. Thus, in Europe we expect to find clade A haplotypes, as in Asia, and Europe and Asia to form a joint Eurasian 'Old World' meta-population, with possibly extensive east-west gene flow. Additionally, we present many more samples from central and eastern Canada, which lacked in the most recent analysis [10], to complete an mtDNA sequence data set comprising samples from the whole Holarctic. A combination of population genetic analyses and coalescent simulations in an isolation-with-migration framework [25,26] enables us in this to measure genetic diversity and geographic partitioning thereof, as well as population histories and migration rates between Old World and New World mallards, and to test the currently hypothesised flyways of mallards (see Table 1 and [2-4]).

Results

MtDNA control region sequencing

Our data set is comprised of mtDNA sequences from 346 mallard ducks around the world consisting of 195 newly sampled mallards and 151 sequences from previous studies [10,27] (for details on the sequences and localities see Table 1 and methods section). 155 different haplotypes were found in this data set, of which 101 were already contained in the data set of Kulikova *et al.* [10,27], and 54 were novel. Of the 622 aligned nucleotide positions 93 (15%) were variable, 73 (11%) sites were parsimony informative and four sites showed gaps. Additional file 1 lists the haplotype names with corresponding sample IDs. Of the 155 haplotypes, 108 haplotypes were only represented by a single individual, 44 haplotypes by 2-7 individuals, one haplotype by 16 individuals (Hap 56), one haplotype by 20 individuals (Hap 57), and the most frequent haplotype (Hap A7) was represented by 71 individuals (see Additional file 1 for details). This A7 haplotype is found in both Old and New World, in almost all localities. All new sequences obtained in this study are deposited in GenBank [28] with accession numbers JN029963-JN030157. In Table 1 we further list nucleotide diversity (π) and haplotype diversity (dH). Nucleotide diversity is similar in all Old World flyways (π between 0.003 and 0.005, and 0.008 in Eastern Asia) but consistently higher in the New World ($\pi > 0.013$). Samples from the Aleutians have intermediate nucleotide diversity somewhere between Old and New World ($\pi = 0.009$). The Greenland samples are very low in both nucleotide diversity ($\pi = 0.0004$) and

Table 1 Sampling localities and genetic diversity

land mass ¹⁾	region	flyway ²⁾	π (\pm SD)	dH (\pm SD)	locality	n	lat.	long.		
OW	Europe	North-West EU	0.00344 (0.00038)	0.696 (0.05)	GBAB	21	57.433	-2.393		
					NLFR	23	53.035	5.574		
					NOBE*	22	60.355	5.345		
		Central EU	0.00470 (0.00034)	0.929 (0.018)	ATHO	18	48.607	16.905		
					DEWU	24	50.036	11.972		
					EETA*	22	58.345	27.154		
	East EU	0.00536 (0.00182)	1.000 (0.272)	RUKR	10	60.999	38.556			
				RUAS	3	46.217	47.767			
	Asia	Central Asia	0.00438 (0.00112)	0.833 (0.127)	KZAO	3	44.893	75.122		
					RUOM*	6	55.845	71.853		
		East Asia	0.00835 (0.00093)	0.987 (0.005)	MNDA	1	47.000	119.367		
RUKK*					4	50.388	136.996			
RUPR					1	59.631	149.115			
RUPR					82	45.007	132.432			
NW	Alaska	Pacific NA	0.01323 (0.00151)	0.985 (0.015)	USFI	13	64.825	-147.584		
					USIZ	6	55.358	-162.728		
					USJU	1	58.364	-134.572		
					USKB	2	60.545	-151.148		
					USKI	4	57.491	-153.495		
					USSF	1	61.216	-149.884		
					USYD	1	61.367	-163.717		
					USYR	1	65.821	-149.733		
	Canada	Central NA	0.01364 (0.00175)	0.971 (0.021)	CARM	20	50.628	-101.159		
					CASL*	3	49.666	-112.704		
					USPI	1	72.677	-99.469		
		Atlantic NA	0.01430 (0.00224)	0.934 (0.061)	CACO	3	45.579	-64.345		
					CAJC*	2	42.321	-82.385		
					CALM	9	43.962	-80.400		
	N/A	Aleutians	N/A	0.00870 (0.00243)	0.824 (0.084)	USAD	2	51.762	-176.622	
						USAI	8	52.905	172.906	
						USSI	7	52.723	174.112	
		Greenland	N/A	0.00042 (0.00027)	0.177 (0.106)	GLNU	22	64.190	-51.708	

Nucleotide diversity (π) and haplotype diversity (dH) were calculated per flyway. Sampling locations within each flyway are coded by the two letter country abbreviation (e.g., GB - Great Britain; NL - Netherlands, etc.) and a two letter locality abbreviation (GBAB - Great Britain, Aberdeen; NLFR - Netherlands, Friesland). Details on coding are given in Additional file 2. Samples sizes per locality within flyways (n) are reported, as well as digital GPS coordinates (latitude, lat. and longitude, long.; negative longitudes indicate westward direction).

¹⁾land masses are defined as Old World (OW, which is Eurasia) and New World (NW, which is the North-American continent). The Aleutian Islands and Greenland are not assigned to either of these land masses

²⁾flyway definitions based on references 2-4

*coordinates are averages of several near-by places, where ducks have been sampled and combined into one sampling locality. Full details for each individual can be found in Additional file 2.

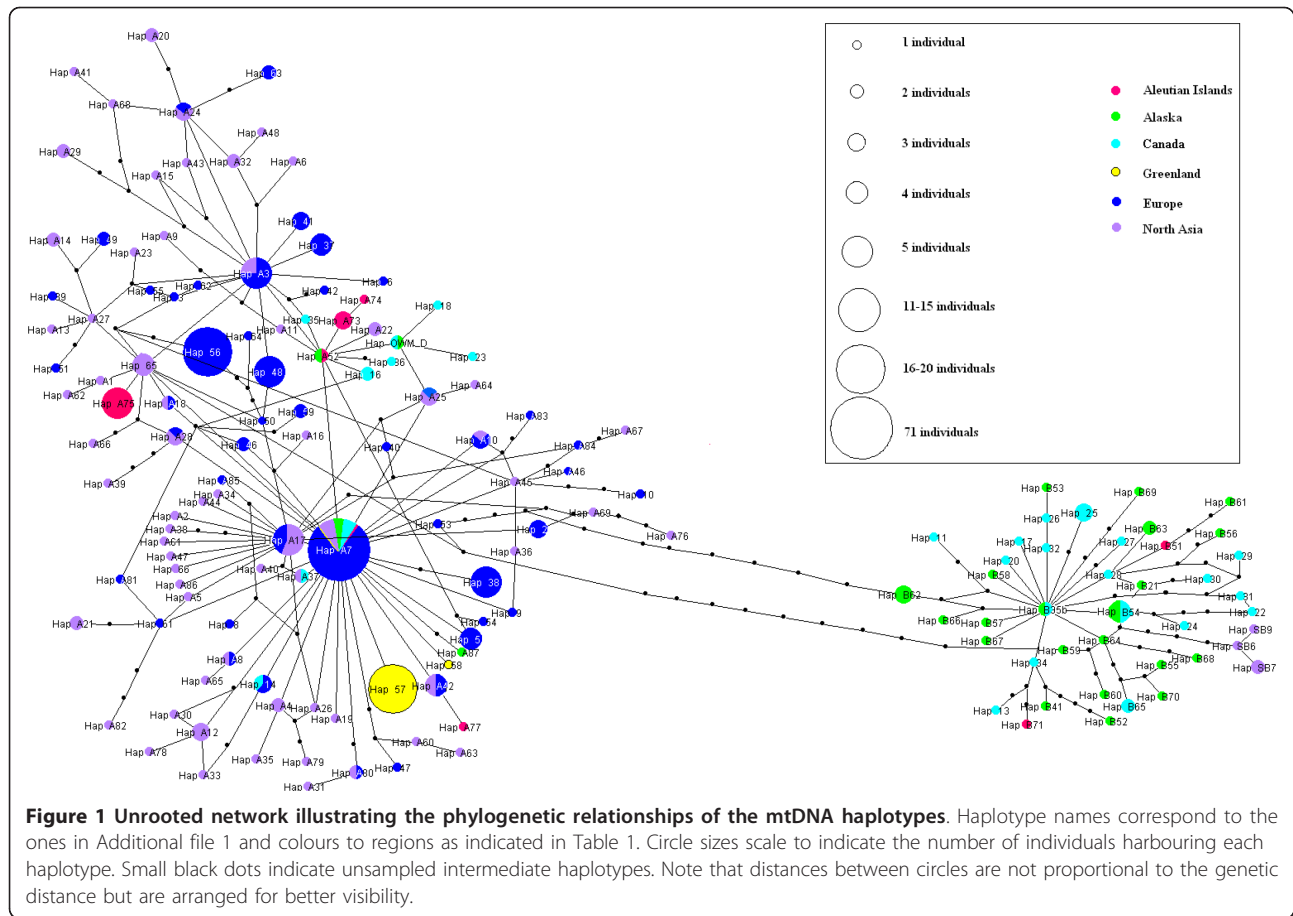
haplotype diversity (dH = 0.2), whereas haplotype diversity is high (dH > 0.8) in all other flyways except in the North-West Europe flyway (dH = 0.7).

Testing neutrality based on the frequency spectrum of the haplotypes with Fu's F_S [29] revealed negative test statistics for both measures when samples were partitioned into Eurasia ($F_S = -25.52$, $p < 0.001$) vs. North-America ($F_S = -24.63$, $p < 0.001$) Splitting Eurasia into

Europe ($F_S = -26.41$, $p < 0.001$) and Asia ($F_S = -25.38$, $p < 0.001$) leads to statistics significantly smaller than zero for both continents.

Two distinct mallard clades

An unrooted haplotype network (Figure 1) shows the presence of two distinct clades. These clades correspond well to a classification of sampling locations into Old



World (clade A haplotypes, mainly Eurasia) and New World (clade B haplotypes, mainly North-America). Within these clades, a great variety of haplotypes was sampled with relatively few missing intermediate ones, represented by small black dots in Figure 1. Clade A and clade B are separated by about ten substitutions. Phylogenetic inference by building a Neighbour-Joining [30] tree supported the split between clade A and clade B haplotypes with 97% of the bootstrap replicates (Figure 2).

Low genetic differentiation within Eurasia and North-America, but large differences between them

Genetic differentiation between Eurasia and North-America (Aleutians and Greenland excluded, see methods section) and between all individual flyways (flyways definitions are in Table 1) was assessed by Wright's F_{ST} statistics [31]. The F_{ST} value between the two land masses was 0.51. Pairwise F_{ST} values on the flyway-level are given in Table 2. Not all flyways were significantly differentiated from each other. In case of the Eastern Europe flyway sample size was too low ($n = 3$) to make any meaningful statement about the amount of

differentiation. None of the intra-North-American flyways was significantly differentiated from the others on that continent. The same was found between the two Asian flyways. In Europe, however, with the exception of the Eastern European flyway which suffered from low sample size, we observed significant structure, albeit low in magnitude (North-West vs. Central Europe: $F_{ST} = 0.08$). Notably, the Aleutian sample was most differentiated from the North-American Pacific and Central flyways but to a lesser extent from the Eurasian ones (in some cases not even significantly; Table 2). Finally, the Greenland sampling location displayed relatively high and significant differentiation with each of the other flyways.

An analysis of molecular variance (AMOVA) revealed little genetic variation within the various Eurasian or North-American flyways compared to the amount of variation between these two land masses. 50.2% of the genetic variation lies between North-America and Eurasia, and 46.6% within the sampled localities. Only 3.2% of the genetic variation was partitioned between flyways within the continents (Table 3). Performing an AMOVA in which localities are not pooled into flyways does not

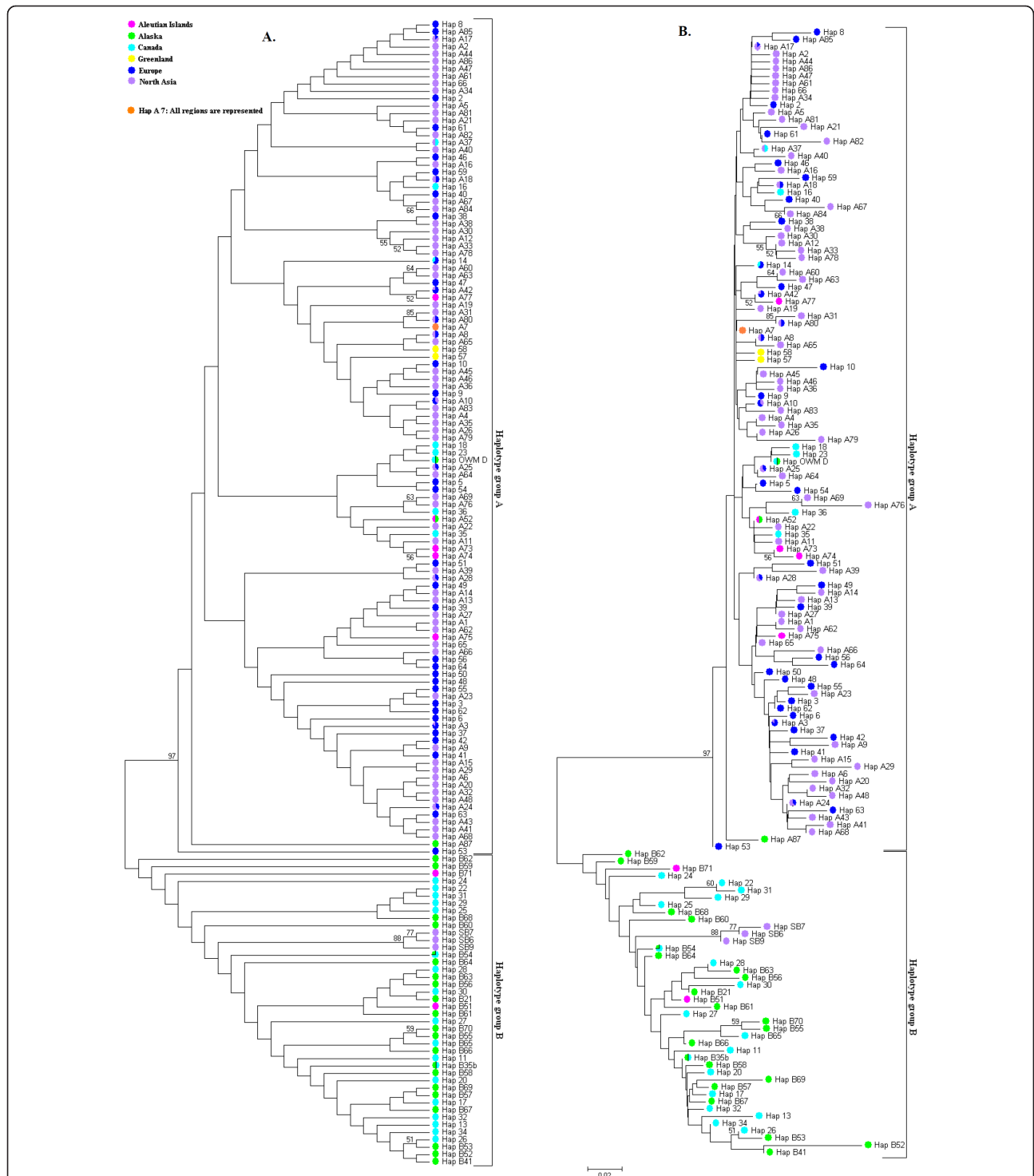


Figure 2 Unrooted Neighbour Joining tree illustrating clade A and B mtDNA control region haplotypes. Bootstrap values (500 replicates) are shown next to the branches if > 50%. Positions containing gaps are removed by pairwise deletion. Haplotype names and colours of the dots correspond to the ones in Figure 1. A: Tree ignoring branch lengths. B: Branch lengths scaled to evolutionary distances (sum of branch lengths = 2.65).

Table 2 Pairwise F_{ST} values for all flyways

	1	2	3	4	5	6	7	8	9	10
1) Pacific NA	-									
2) Central NA	-0.01	-								
3) Atlantic NA	0.19	0.15	-							
4) North-West EU	0.65*	0.63*	0.33*	-						
5) Central EU	0.62*	0.60*	0.28*	0.08*	-					
6) East EU ¹⁾	0.44	0.39	0.03	0.05	-0.09	-				
7) Central Asia	0.50*	0.47*	0.16	0.08	0.00	-0.07	-			
8) East Asia	0.52*	0.49*	0.16*	0.06*	0.01	-0.12	-0.01	-		
9) Greenland	0.64*	0.64*	0.41*	0.36*	0.30*	0.71*	0.54*	0.18	-	
10) Aleutians	0.43*	0.40*	0.10	0.17*	0.09*	-0.06	0.05	0.05	0.37*	-

F_{ST} values printed boldface and marked by an asterisk are statistically significant after Bonferroni correction.

¹⁾note, that the east European flyway is only represented by three samples, and hence the presented values are very crude!

lead to qualitative differences of this outcome (Table 3; values in brackets).

When we assigned localities to flyways within continents, per-continent AMOVA showed that genetic variation was to a large extent partitioned within localities: 85% in North-America, 76.8% in Europe, 98.7% in Asia and if combined into one land mass: 87.7% in Eurasia. In none of these analyses there was a significant amount of genetic variation between flyways ($p > 0.2$). In Europe, there was a significant ($p < 0.001$) genetic variance component of 22.9% between the localities within the flyways, in the other continents this was not statistically significant.

Demographic history and gene flow

The runs of the coalescent simulation program IMA2 converged quickly in an optimal parameter space. Effective sample sizes for all parameters exceeded 300,000 and each newly sampled genealogy was independent from the previous one (autocorrelations in all parameters were 0.05 and less). Pairwise correlations of parameters were between -0.1 and +0.1, except between q_0 and q_1 (the estimates of population size for Old World (OW) and New World (NW)) where it was -0.19.

All parameter estimates had relatively sharp unimodal posterior density distributions (Figure 3), except the migration rates which were poorly estimated, with wide ranges covering three orders of magnitude ($m_{OW \rightarrow NW}$ and $m_{OW \rightarrow NW}$: 5.1×10^{-9} - 4.7×10^{-6} and 5.1×10^{-9} -

1.8×10^{-6} , see Figure 3). Table 4 summarises the results in demographic units. Although 95% highest posterior densities (HPD95) for population sizes of OW and NW samples overlapped IMA2 calculated that there was a 66% probability that the effective population size of the NW mallards (1.3 - 4.2 million) is larger than the OW population (1.5 - 2.9 million). Further, even though the HPD95 ranges of the migration rates peaked almost at zero, there was a 76% probability that the migration rate from OW into NW is higher than the other way around in the few cases in which it may occur.

Discussion

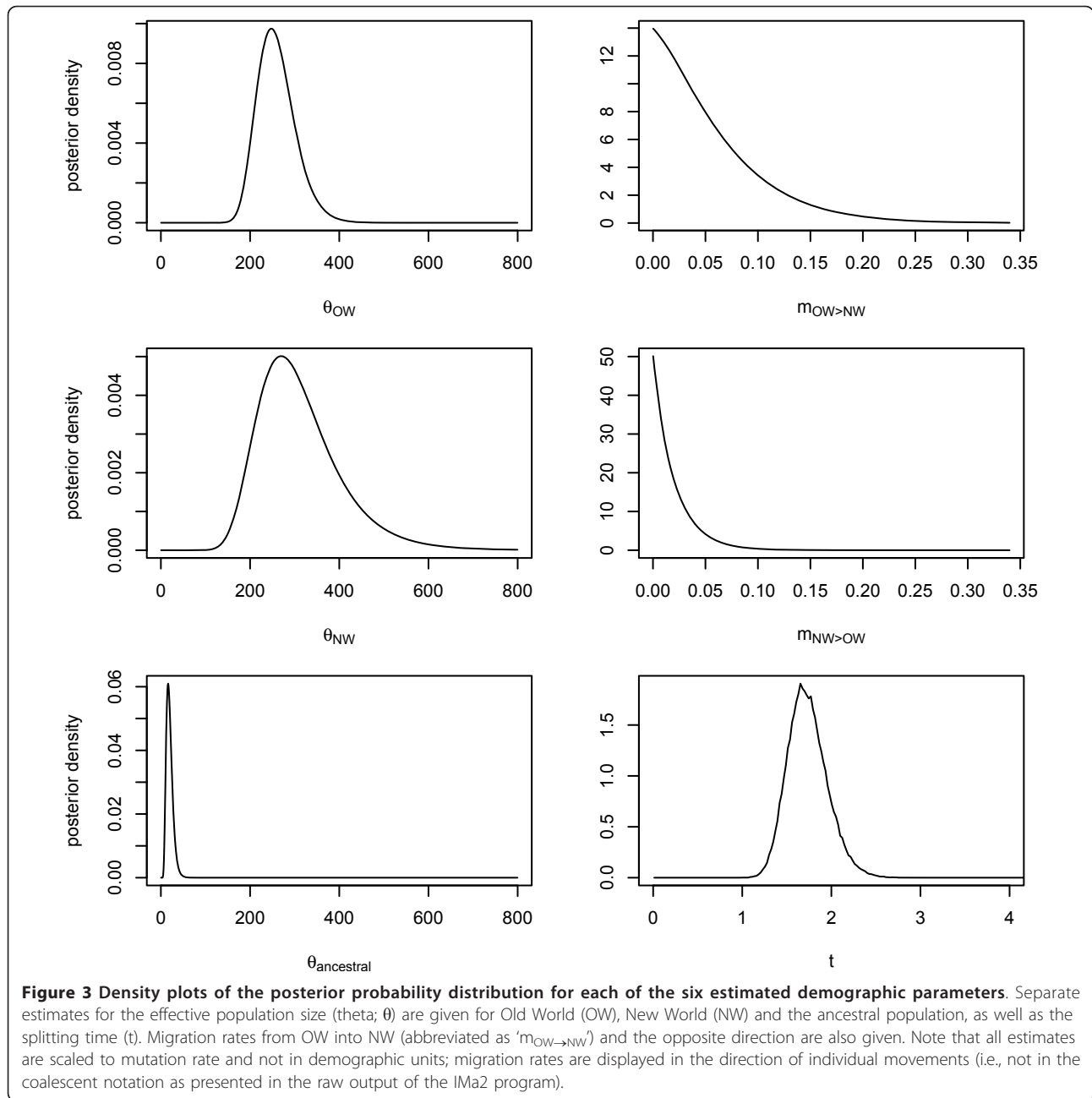
Conclusive assertion: There are two distinct mallard clades

Since *Awise et al.* first studied genetic structure in mallards in 1990 [24] it is hypothesised that no more than two mitochondrially distinct clades exist among them. Our phylogenetic analyses confirm this hypothesis for mallard samples collected and analysed from the numerically largest and geographically most complete set of mallard mtDNA sequences to date. In addition to previously analysed North-American and Asian samples [10], we added substantial numbers of European and Central and Eastern North-American mallard samples and thus completed a data set spanning the whole Northern Hemisphere. Through large-scale sampling of mallards from all over their distribution range we can now conclusively state that no more than two main

Table 3 AMOVA analysis of flyway genetic variance in the land masses Eurasia and North America

	d.f.	SS	VC	% var
Between land masses	1 (1)	288.03 (288.03)	2.67 (2.66)	50.20 (49.94)
Between flyways (localities) within land masses	6 (26)	50.71 (168.71)	0.17 (0.43)	3.22 (8.1)
Within flyways	299 (279)	740.26 (622.74)	2.48 (2.23)	46.57 (41.96)
Total	306 1079	5.32	100	

Values in brackets were obtained when treating sampling localities directly as groups (i.e., not pooled into flyways). All variances are statistically significant ($p < 0.05$). Abbreviations: d.f.: degrees of freedom, SS: sum of squares, VC: variance components, % var: percentage of variation.



clades exist in mallard, also not in the previously unsampled European populations.

Gene pool differentiation and diversity

Mallards are highly mobile and some studies suggest that natal philopatry (i.e., philopatry in the sense of [32]), which in waterfowl is usually strongest in females, is less pronounced - even in female mallards - than it is in related species [2,9,33] (but see [34-36]). We found strong support for the hypothesis of 'flyway permeability' [8,9] in mallards. The analyses of F_{ST}

between intra-continental flyways strongly promote the conclusion of female inter-flyway mixing. Neither within North-America nor in Asia did we detect differentiation between the subcontinental flyways. We note that for some comparisons the sample size was on the low side. However, if this was causal to not finding significant differentiation between flyways within a continent, we would also not have found significant differentiation between those flyways with lower sample sizes with flyways on different continents. Hence, we believe our finding of a lack of differentiation is

Table 4 Demographic parameters as estimated under an isolation with migration model

parameter	scaled to mutation rate			in demographic units		
	mode	HPD95L	HPD95H	mode	HPD95L	HPD95H
θ_{ow}	247.6	181.2	345.2	2,073,285	1,517,283	2,890,541
θ_{NW}	270.0	154.8	499.6	2,260,852	1,296,222	4,183,414
$\theta_{ancestral}$	16.4	6.8	35.6	137,326	56,940	298,098
t	1.7	1.3	2.2	55,366	42,002	74,993
$m_{OW \rightarrow NW}$	0.00017	0	0.1586	5.1×10^{-9}	0	4.7×10^{-6}
$m_{NW \rightarrow OW}$	0.00017	0	0.06069	5.1×10^{-9}	0	1.8×10^{-6}

For calculation of demographic units a generation time of one year was used. The mutation rate per locus per year was set to 2.9856×10^{-5} . See main text for further explanations. The demographic units are: population size (θ in effective individuals, splitting time (t) in years, migration rates (Nm) in effective individuals per year. Estimates of the peak of the posterior parameter distributions (mode) are presented along with low and high bounds of the 95% highest posterior densities (HPD95L/H).

valid. Yet, there is population structure in Europe based on F_{ST} analysis (see below).

Confirmative evidence for the pattern of genetic population structuring found above stems from our analysis of molecular variance (AMOVA). Almost no genetic variation in our world-wide data set is explained between flyways within continents. On this global scale, about half the variation lies between North-America and Eurasia, and the other half within the flyways. When analysing genetic variance per continent, in North-America and Asia the largest proportion of variance resides on the level of sampling localities but not within or between flyways. However, notably, in Europe there is an unexpected significant variance component between localities within flyways, pointing at more genetic structure than in the other continents. In general, these findings confirm the absence of strong female philopatry in mallards. The population structure is not shaped by female mallards returning to the place where they were born, and hence lifetime dispersal seems high. In that sense, currently recognised waterfowl flyways do not depict well the movements and dispersal of mallards - a pattern similar to other duck species [33,37,38] - although it cannot be generalized for all ducks [39,40]. However, our findings show that indeed the European mallard population seems to be more structured than their conspecifics in other parts of the world. Geographic, micro-climatic and urbanisation structuring in Europe may be different in Europe when compared to Asia and North-America, shaping mallard population structures in different ways. Future studies employing genetic markers, probably at an even denser sampling scheme, may be able to quantify such effects by making use of GIS and remote sensing technology. A further possibility to explain European mallard population structure may be extensive release of farmed mallards for hunting purposes in many countries in Europe. These originally wild mallards are bred in captivity for many generations, and even though looking quite similar to their wild relatives they can differ subtly in

morphology [12]. Such differences have been proven to leak into wild populations by interbreeding with escaped farmed mallards [41]. Obviously, this introgression of genetic material of farmed animals has consequences for the genetic identity of wild populations, especially when translocation and release at distant localities takes place [42]. Genetic introgression has been demonstrated for mallards in Italy already [43]. Such manipulation of local population structure may well explain our finding of increased genetic structure in Europe.

Demographic history

Mallards from the Old World are different from those in the New World, which is attributable to the differences in the distribution of clade A (Eurasia) and clade B (North-America) haplotypes. In Europe and the largest part of Asia we only found clade A haplotypes with the exception of a handful clade B haplotypes in the far east of Asia (Figures 1, 2). Nevertheless, in a phylogenetic assessment in the original study in which these sequences were determined, they were shown to be characteristic for introgressed eastern spot-billed ducks and likely do not indicate original North-American mallard clade B individuals [10,27] (removing these three curious sequences from the demographic modelling analysis did not alter the results, though; data not shown). On the other hand, in North-America, where clade B dominates, also clade A haplotypes are relatively common, which is reflected in higher nucleotide and haplotype diversities. This suggests two factors shaping the current genetic structure in mallards. From the deep split into clade A and B haplotypes we infer that Old World and New World have become separate with no or little gene flow since then. Further, if genetic exchange occurs - as indicated by clade A haplotypes occurring both in Old and New World - it seems directional with larger rates from Old World into New World. However, this directionality is not statistically robust because in the IM analysis migration rates were only estimated poorly. The split between mallards from

Old World and New World was estimated to have occurred between 43,000 and 74,000 years ago. This dating coincides with the drop in sea level during the last glacial period about 110,000 - 10,000 years ago. During this time the Beringia land bridge fell dry and extensive exchange of floras and faunas became possible. When the glaciers melted again, this Beringia land bridge connection got cut off and Eurasian and North-American faunas and floras became separated more strictly. But also during the glacial maximum the continental ice sheet developed in North-America separated northern from southern regions of this continent [44]. The estimated population size at that time ($\theta_{\text{ancestral}}$) was about 30 times smaller than it is today (the sum of θ_{OW} and θ_{NW}). Such high growth rates in waterfowl populations since the last glaciation has already been demonstrated in several other studies, often proven to be exponential [10,16,33,45] and connected to population size bottlenecks. This could explain the excess of rare haplotypes as demonstrated especially by negative Fu's F_S in Eurasia, but also in North-America. The present-day situation is a deep split into an Old World and New World population, with migration rate estimates statistically not significantly different from zero, indicating that the rise of sea levels after the last glacial maximum results in a potent barrier to female gene flow between OW and NW. Numerically, the peak of the density distribution of migration rates suggests 0.04 - 0.05 effective migrants per year in each direction. However, the posterior density plots of the migration rates (Figure 3) are not bell-shaped as they should be. This indicates that IMA2 did not find enough information in the data to reliably estimate migration rates. If prior ranges are too wide, software like IMA2 may estimate parameters to be zero, but in our analysis the range of the prior distribution (0 - 0.34) was not much wider than the range of the estimated posterior distribution (HPD95) for migration of Old World mallards to the New World (0 - 0.16) (Table 4). Re-running the IMA2 analysis with a narrower prior (0 - 0.2) did not resolve the issue (data not shown). We believe this is due to a combination of very large extant population sizes as a result of strong population growth and divergence. Based on HPD95 migration rates could be as low as zero, or as high as 23 (NW→OW) or 80 (OW→NW) effective individuals. The failure of IMA2 to quantify migration rates may be related to this issue. Irrespective of this, the sharing of identical haplotypes indicates the possibility that migrants do travel between the land masses and establish genetic traces in the receiving population. The nucleotide diversity of the Aleutian samples, being intermediate between that of Eurasia and North-America, is a good indicator for this. Even though migration rates as such were poorly estimated

with our data, the shapes of the migration rate curves and the implemented probability test in IMA2 point at directionality of migration from Old World into New World.

Genetic exchange between Old World and New World

Based on the sharing of haplotypes between Old and New World mallards and the results of the demographic modelling, we propose that some genetic exchange of mtDNA between the two land masses possibly occurs. IMA2's estimated migration rates, being not statistically different from zero, may be biased downwards due to a lack of statistical power for the migration rate parameters, or model violation of migration/drift equilibrium, but indicate directionality. Genetic differentiation between Old and New World was high ($F_{ST} = 0.51$) indicating limited gene flow at most. We propose that the route of this migration is mainly, if not completely, via the Aleutian Islands. On these islands individuals with clade A and B haplotypes intermix and form a haplotype cline [10], and genetic diversity (π) is intermediate between Eurasia and North-America. F_{ST} values between the Aleutians and the flyways indicate a closer relationship between the Aleutians and Old World populations than with the New World. This may be explained by the Westerly winds (more regularly blowing from the west into the east), governing the Ferrel cell of the global climatic circulation system. The Aleutian Islands are all well within the Ferrel cell that reaches up to higher than 60° north [46,47], and characteristic winds are especially strong during the time in which mallards migrate [48]. A note of caution is warranted here, because smaller scale terrestrial weather conditions can be more important than the large atmospheric systems. Even if weak when compared to the strength of flight abilities this effect can have a profound impact on the regular "drifting off" of migrating birds in general [48] and waterfowl specifically [49].

The alternative route, from Europe via the Atlantic Ocean, along Iceland and Greenland, could be deemed less likely on basis of this "drifting by wind" proposition. Mallards travelling to North-America by the Atlantic route would face headwinds more regularly [48]. However, the mallards sampled from Greenland all bear clade A haplotypes implying a Eurasian origin. Additionally, genetic differentiation between mallards from the Atlantic part of Canada and European mallards is only half of what we measured between central or western Canada and Europe. Once arrived in Greenland it would not be hard to imagine crossing the last few hundred kilometres to Canada. Unfortunately, we were not able to analyse more samples from Greenland, especially from different localities. Interpretations based on the current Greenland data can only be tentative. Just two

different haplotypes were found there and the sampling location was a pond near the city of Nuuk, likely inhabited by closely related mallard families. Little is known about the origin and movements of Greenland mallards [2,50], and hardly anything (except from data in this study) about their genetics (for a preliminary report, see [51]). More studies into this population are certainly needed to draw firm conclusions. Sampling of mallards from Iceland and the Faroe Islands would make it possible to perform detailed analyses of a potential Atlantic route of genetic exchange of mallard populations. Note, however, that all results presented in this paper are based on the analysis of mtDNA which is a maternally inherited marker. Patterns discerned from our data are thus only valid for the female part of the population. Males are sometimes suggested to also possess a homing instinct [52] but in contrast to females this is generally believed to be much less pronounced [53]. It will thus be important to study nuclear markers in mallard [43,54-57] on large geographical scales which depict the genetic structure of both sexes (Kraus *et al.*, manuscript in preparation). Some sex-bias in migration is not unusual [58] but can be extreme if males are much more dispersive than females, as demonstrated in a famous example of the white shark *Carcharodon carcharias* [59].

Post-glacial colonization

Clade A is characteristic for Eurasia, and clade B for North-America, although many individuals in North-America belong to clade A. Kulikova *et al.* [10] proposed two phylogeographic hypotheses to explain such a haplotype distribution, termed "Asian invasion" and "incomplete lineage sorting" (initially suggested by Avise *et al.* [24]). In an Asian invasion scenario the occurrence of clade B haplotypes in mallards is explained by Asian mallards moving into a previously mallard-free North-America, followed by acquiring B-haplotypes by introgressive hybridisation with closely related indigenous duck species (such as the black duck *A. rubripes* [60]) resulting in frequently observed mtDNA paralogy [27,61,62]. The incomplete lineage sorting hypothesis rests on the proposed occurrence of a polymorphic ancestral gene pool (at least with respect to mtDNA clades) which is facilitated further by large populations [63,64] and [Kraus *et al.*, manuscript submitted]. These two hypotheses offer different predictions about the expected distribution of clade A and clade B haplotypes in North-America: As a result of an Asian invasion one would expect to find a gradual decline of clade A haplotypes from western to eastern North-America if we assume that all immigration from Eurasia took place via the Aleutian Islands and continues so. We cannot confirm this scenario; clade A haplotypes today occur in the

whole of North-America. This, however, can also be the result of large dispersal abilities of mallards in North-America, which can diminish genetic clines relative quickly. Hence, an Asian invasion cannot be clearly rejected, either. Kulikova *et al.* [10] imagine this further to be due the impact of mallard farming, resulting in anthropogenic translocation also of clade A mallards. This explanation resembles one of the possibilities we propose to explain genetic structure in Europe.

As an alteration of the Asian invasion scenario we offer additional thoughts based on the extent of the most recent continental glaciation in North-America. The opening of the Beringia land bridge on the one hand enabled exchange of fauna and flora between East Asia and Alaska because these regions were to a large extent ice-free. Coincident with the emergence of this connection, though, the considerable North-America ice sheet built up south of Alaska, what is today southern Canada. An Asian invasion thus took place probably only in the very north of North-America. The clade B haplotype may have become fixed south of that ice sheet due to the population bottlenecks. After the retreat of the glaciers clade B mallards from the south got into secondary contact with northern clade A mallards. It would thus be interesting to sample mallards from central and southern North-America with this hypothesis in mind. To account for and further study mtDNA paralogy in duck species such a study would have to include samples especially from Black Ducks [24,60], and if possible from other *Anas* species harbouring clade B haplotypes, too [61,62]. Avise *et al.*'s incomplete lineage sorting hypothesis [24] would naturally account for a more even distribution of clades and is consistent with the idea that a dichromatic (and mitochondrially polymorphic) mallard population gave rise to its monochromatic sister species by peripatric isolation [65]. But as Kulikova *et al.* [10] point out it seems unlikely that only the clade B haplotype gets fixed in all sister species but not the "original" mallard. We thus concur with Kulikova *et al.* [10] in concluding that a complicated mix of historical, recent and anthropogenic factors shaped the current world-wide mallard population structure. The results we can add from analysing >100 additional European mallards and the outcome of our demographic modelling study substantiate this claim.

Conclusions

Many aspects of the biology of ducks are known when it concerns ecological and management parameters (see [11] and references therein for a recent overview), and also their phylogenetic placement has frequently been assessed, e.g. [61,66]. Surprisingly, only few studies have contributed to discern population genetic patterns for

ducks, especially mallards. The information collected in this study is essentially representing the first complete assessment of the world-wide mallard population genetic structure in this duck. With more than 300 samples from the whole distribution range of the mallard we do not find intermediate haplotypes between clade A and B, or additional discernable clades. Our data and analyses corroborate the conclusion of Kulikova *et al.* [10] that a complicated mix of historical, recent and anthropogenic factors shaped the current world-wide mallard population structure. Further, we offer an additional hypothesis on how the current haplotype distribution emerged: a partial Asian invasion that took place only in northern North-America, which was ice-free at the last glacial maximum, followed by secondary contact with the southerly mallard population in North-America in which clade B haplotypes could have been fixed. A study to address this hypothesis would need to additionally analyse mallard samples from several well-spaced localities throughout central and southern North-America, as well to pay attention to *Anas* species closely related to mallards, often bearing clade B haplotypes themselves [61,62]. Mallards form an enormously large population. In the northern hemisphere, the population is structured deeply into two major clades, but within these landmasses the populations are very homogeneous (and perhaps panmictic) in which the concept of flyway do not contribute to a further understanding of mallard population genetic structure. Homogeneity over thousands of kilometres facilitates the spread of diseases such as Avian Influenza. Between the two landmasses there may be a little gene flow, apparently in west - east direction across the Bering Strait. The genetic distinction between the OW and the NW is not eroded away by this gene flow, and apparently arose during the last Glacial Maximum.

Even though we refute the flyway concept for mallards as a representation of biological realities of geographic structuring of populations, we would like to point out that waterfowl can very well be managed on the basis of a flyway concept: the North American success story of waterfowl population increases over the last century underpins that success. In that sense, flyways ought to be viewed as 'problemsheds' in the sense of the Ecosystem Approach of the Convention on Biological Diversity and not so much as biological realities.

Methods

Sample collection

Blood from 195 mallards was collected on FTA cards [67], in most cases by hunters. Exceptions are the localities from Greenland and Norway. There, mallards were trapped, blood drops on FTA cards were sampled from the wing or foot vein, and mallards were released again (animal handling approved by the animal ethical committee of Wageningen University - DEC, the Greenland

Home Rule and the Norwegian Food Safety Authority - Forsøksdyrvalget). Details on sampling localities and samples can be found in Table 1 and Additional file 2. A cautionary note is needed for the Greenland samples. The trapping of these animals most likely resulted in the catching of a few ducks with several of their chicks which often entered the trap as a group. This may have affected some measures of population genetic parameter (see results and discussion).

For phylogenetic analyses samples were assigned to continents to visualise their geographic region of origin. Greenland and the Aleutian Islands were not assigned to a specific continent, and Alaska and Canada were treated as separate regions within North-America. For population genetic analyses, sampling localities were pooled into biological populations based on hypothesised mallard flyways in Europe [2], Asia [3], and North-America [4] (Table 1), allowing us to test our data against these human-made classifications. Samples from Greenland and the Aleutian Islands were classified as separate populations for their intermediate status.

DNA isolation and sequencing

DNA was extracted from FTA cards using the Gentra Systems 'Puregene DNA purification Kit' (Qiagen, Valencia, California). The manufacturer's instructions were followed with slight modifications for handling the FTA cards: up to a quarter of the encircled area of the FTA cards (depending on how much blood was preserved) was cut into small pieces of approximately 2 mm² and digested with 60 µg Proteinase K (Sigma- Aldrich, St. Louis, Missouri) in 600 µl Cell Lysis Solution (Gentra Systems) at 55°C over night. Subsequently, proteins were precipitated with 200 µl Protein Precipitation Solution (Gentra Systems) and spun down together with the FTA card material. DNA in the supernatant was precipitated with isopropanol and washed with 70% ethanol. Quantity and purity of the DNA were measured using a Nanodrop ND1000.

PCR amplification targeted the 5' end of the mtDNA control region which is homologous to positions 79-773 in the chicken (*Gallus gallus*) mitochondrial genome [68]. In some duck species the presence of 'numts' (nuclear copies of mtDNA) was proposed in this region [69] but previous examination of mallard sequences did not reveal evidence for this [10]. Reactions were performed in 12 µl containing 30 ng genomic DNA as template, 3 µl STE buffer, 5.5 µl Abgene Mastermix (ThermoScientific) and 0.25 µl of each primer (10 mM): L78 [69] (forward) and H774 [70] (reverse). PCR amplification was done in a BioMetra Thermocycler (Biometra, Göttingen, Germany) under the following cycling conditions: 7 minutes initial denaturation at 94°C, followed by 45 cycles of 20 seconds at 94°C, 20 seconds at 49°C and 1 minute at 72°C, completed by 7 minutes

final elongation at 72°C. Quality and quantity of the PCR product was determined by gel electrophoresis and the product was purified by vacuum filtration on a Millipore Multiscreen PCR plate. Forward and reverse DNA strands were cycle-sequenced using the ABI Big Dye Terminator Cycle Sequencing Kit 3.1 under the following cycling conditions: 1 minute initial denaturation at 96°C followed by 25 cycles of 10 seconds at 96°C, 5 seconds at 49°C, and final elongation of 4 minutes at 60°C. The sequencing reaction products were precipitated by sodium acetate and ethanol to purify the product, followed by capillary sequencing on an ABI 3730 DNA Analyzer. The forward sequences were verified with the sequence of the reverse strand in MEGA4 [71]; some manual corrections were needed. If the forward sequence was absent, or only partially resolved, the reverse strand was used and aligned with the other sequences for verification. Additionally, 151 published sequences from studies of Kulikova *et al.* [10,27] were downloaded from GenBank [28] (accession numbers: AY506868-AY506870; AY506873-AY506901; AY506904-AY506908; AY506910-AY506917; AY506919-AY506944; AY506974-AY506984; AY928831-AY928899). Altogether, these sequences were aligned in MEGA4 [71] using the ClustalW algorithm [72] under default settings. Names of haplotypes defined in the studies by Kulikova *et al.* [10,27] were preserved.

Phylogenetic analyses

A phylogenetic tree of the haplotype sequences was constructed in MEGA4 [71], using the Neighbour-Joining algorithm [30] with 500 bootstrap replicates. Evolutionary distances were computed under the Tajima Nei model [73]. All positions containing alignment gaps and missing data were eliminated from the dataset for tree construction (complete deletion option). Further, a phylogenetic network was constructed in TCS [74] (version 1.21) by statistical parsimony, here, treating alignment gaps as fifth state.

Population genetic analyses

The basic population genetic parameters nucleotide diversity (π) and haplotype diversity (dH) for each flyway was calculated with DnaSP [75]. Fu's F_S [29] was calculated in Arlequin 3.5.1.2 [76], and evaluated for statistical significance by 16,000 simulated samples in order to guarantee a less than 1% difference with the exact probability in 99% of the cases [77]. Population differentiation was assessed by Wright's F -statistics [31], and partitioning of genetic variance among and within groups was investigated by analyses of molecular variance (AMOVA [78-80]). Calculations were also performed in Arlequin 3.5.1.2 [76] from pairwise nucleotide differences, and statistical significance tested by 16,000 permutations.

Demographic modelling

To make inferences about the extent of migration between Old World (OW) and New World (NW) we modelled the demographic history of mallards by coalescent simulations under an "Isolation with Migration" (IM) model [25,26], as implemented in the program IMA2 (Linux version 10.13.10). OW and NW samples were treated as belonging to distinct populations based on their sampling locality. Greenland and Aleutian samples were excluded because of their possible intermediate status. Upper bounds for parameter priors were estimated during consecutive preliminary runs of the program, based on initial estimates of theta as advised in the manual of IMA2. The final values used for population size, migration rate and splitting time were: $-q$ 800, $-m$ 0.34, $-t$ 23.448. We ran 60 Markov chains in parallel under a geometric heating scheme (option `-hfg`), with the hottest chain being $\beta = 0.5$ and the coldest chain $\beta = 0.975$. Estimated parameters in IMA2 are scaled to the mutation rate. To convert them into demographic units we used a mutation rate of 4.8×10^{-8} (confidence interval $3.1-6.9 \times 10^{-8}$) substitutions per site per year initially published for a wood duck [39]. This rate also produced sensible results in a study of two other ducks of the genus *Anas* [81,82] and needs to be multiplied by the number of nucleotides in the sequence alignment (here, 622) to obtain the substitutions per locus per year to be used for IM analysis. From the two sequence mutation models available in IMA2 we chose HKY [83] which is the applicable model for mtDNA control region sequences [84]. Several run time settings with different heating schemes and durations were explored, all yielding essentially the same outcome. The final simulation, for which we report the results here, was run for a burn-in period of 360,000 steps (they reached convergence already after a few 10,000 steps), and afterwards 26,000 genealogies were sampled every 100 steps from a total 2,600,000 steps.

Additional material

Additional file 1: MS Excel sheet giving information which samples had which haplotype.

Additional file 2: MS Excel sheet with all details for each mallard samples and sequenced for this study. Details given include internal sampling ID, sampling date, sampling country, names of sample collectors, name of sampling location, decimal latitude and longitude coordinates and determined sex of the sampled mallard.

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Authors' contributions

RHSK and AZ designed the study, carried out and coordinated sampling efforts, analysed the data and wrote the paper. PvH designed the study, analysed the data and wrote the paper. DS and SAS coordinated sample collection and discussed the paper. HHTP and RCY revised the manuscript. All authors read and approved this paper.

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References

1. Wink M: Use of DNA markers to study bird migration. *J Ornith* 2006, **147**:234-244.
2. Scott DA, Rose PM: *Atlas of Anatidae populations in Africa and Western Eurasia* Wageningen, The Netherlands: Wetlands International Publication No. 41, Wetlands International; 1996.
3. Miyabayashi Y, Mundkur T: *Atlas of Key Sites for Anatidae in the East Asian Flyway* Wetlands International - Japan, Tokyo, and Wetlands International-Asia Pacific, Kuala Lumpur; 1999.
4. Anon: U.S. Fish and Wildlife Service.[<http://www.flyways.us/flyways/info#flyways-bio>].
5. Bowlin MS, Bisson IA, Shamoun-Baranes J, Reichard JD, Sapir N, Marra PP, Kunz TH, Wilcove DS, Hedenström A, Guglielmo CG, Åkesson S, Ramenofsky M, Wikelski M: Grand challenges in migration biology. *Integr Comp Biol* 2010, **50**:261-279.
6. Jonker RM, Eichhorn G, van Langevelde F, Bauer S: Predation danger can explain changes in timing of migration: The case of the Barnacle goose. *PLoS ONE* 2010, **5**:e11369.
7. Jonker RM, Kraus RHS, Zhang Q, Van Hooft P, Larsson K, Van Der Jeugd HP, Kurvers RHJM, Van Wieren SE, Loonen MJJE, Crooijmans RPMA, Ydenberg RC, Groenen MAM, Prins HHT: Genetic consequences of breaking migratory traditions in barnacle geese. In *Revolutionary non-migratory migrants*, PhD Thesis, Wageningen University Edited by: Jonker RM 2011.
8. Thompson AL: On 'abmigration' among the ducks, an anomaly shown by the results of bird-marking. *Proc Int Ornithol Congr* 1931, **7**:382-388.
9. Guillemain M, Sadouli N, Simon G: European flyway permeability and abmigration in Teal *Anas crecca*, an analysis based on ringing recoveries. *Ibis* 2005, **147**:688-696.
10. Kulikova IV, Drovetski SV, Gibson DD, Harrigan RJ, Rohwer S, Sorenson MD, Winker K, Zhuravlev YN, McCracken KG: Phylogeography of the Mallard (*Anas platyrhynchos*): Hybridization, dispersal, and lineage sorting contribute to complex geographic structure. *Auk* 2005, **122**:949-965.
11. Elmerberg J: Are dabbling ducks major players or merely noise in freshwater ecosystems? A European perspective, with references to population limitation and density dependence. *Wildfowl* 2009, **9**:2-3.
12. Laikre L, Palmé A, Josefsson M, Utter F, Ryman N: Release of alien populations in Sweden. *Ambio* 2006, **35**:255-261.
13. Champagnon J, Guillemain M, Gauthier-Clerc M, Lebreton JD, Elmerberg J: Consequences of massive bird releases for hunting purposes: Mallard *Anas platyrhynchos* in the Camargue, southern France. *Wildfowl* 2009, **184**:191.
14. Kaleta EF, Hergarten G, Yilmaz A: Avian influenza A viruses in birds - an ecological, ornithological and virological view. *Dtsch Tierärztl Wochenschr* 2005, **112**:448-456.
15. Atkinson PW, Clark JA, Delany S, Diagona CH, du Feu C, Fiedler W, Fransson T, Gauthier-Clerc M, Grantham M, Gschweg M, Hagemeyer W, Helmink T, Johnson A, Khomenko S, Martakis G, Overdijk O, Robinson RA, Solokha A, Spina F, Sylla SJV, Visser D: Urgent preliminary assessment of ornithological data relevant to the spread of Avian Influenza in Europe Wageningen, The Netherlands: Wetlands International; 2006.
16. Edgell MCR: Trans-hemispheric movements of Holarctic Anatidae - The Eurasian wigeon (*Anas penelope* L.) in North-America. *J Biogeogr* 1984, **11**:27-39.
17. Koehler AV, Pearce JM, Flint PL, Avian JC, Ip HS: Genetic evidence of intercontinental movement of avian influenza in a migratory bird: The northern pintail (*Anas acuta*). *Mol Ecol* 2008, **17**:4754-4762.
18. Ramey AM, Pearce JM, Ely CR, Sheffield Guy LM, Irons DB, Derksen DV, Ip HS: Transmission and reassortment of avian influenza viruses at the Asian-North American interface. *Virology* 2011, **406**:352-359.
19. Winker K, McCracken KG, Gibson DD, Pruett CL, Meier R, Huettmann F, Wege M, Kulikova IV, Zhuravlev YN, Perdue ML, Spackman E, Suarez DL, Swayne DE: Movements of birds and avian influenza from Asia into Alaska. *Emerg Infect Dis* 2007, **13**:547-552.
20. Si Y, Skidmore AK, Wang T, De Boer WF, Debba P, Toxopeus AG, Li L, Prins HHT: Spatio-temporal dynamics of global H5N1 outbreaks match bird migration patterns. *Geospat Health* 2009, **4**:65-78.
21. Nishiura H, Hoyer B, Klaassen M, Bauer S, Heesterbeek H: How to find natural reservoir hosts from endemic prevalence in a multi-host population: A case study of influenza in waterfowl. *Epidemics* 2009, **1**:118-128.
22. Webster RG, Bean WJ, Gorman OT, Chambers TM, Kawaoka Y: Evolution and ecology of influenza A viruses. *Microbiol Rev* 1992, **56**:152-179.
23. Olsen B, Munster VJ, Wallensten A, Waldenström J, Osterhaus ADME, Fouchier RAM: Global patterns of influenza A virus in wild birds. *Science* 2006, **312**:384-388.
24. Avise JC, Ankney CD, Nelson WS: Mitochondrial gene trees and the evolutionary relationship of Mallard and Black Ducks. *Evolution* 1990, **44**:1109-1119.
25. Hey J, Nielsen R: Multilocus methods for estimating population sizes, migration rates and divergence time, with applications to the divergence of *Drosophila pseudoobscura* and *D. persimilis*. *Genetics* 2004, **167**:747-760.
26. Hey J, Nielsen R: Integration within the Felsenstein equation for improved Markov chain Monte Carlo methods in population genetics. *Proc Natl Acad Sci USA* 2007, **104**:2785-2790.
27. Kulikova IV, Zhuravlev YN, McCracken KG: Asymmetric hybridization and sex-biased gene flow between Eastern Spot-billed Ducks (*Anas zonorhynchos*) and Mallards (*A. platyrhynchos*) in the Russian Far East. *Auk* 2004, **121**:930-949.
28. Benson DA, Karsch-Mizrachi I, Lipman DJ, Ostell J, Sayers EW: GenBank. *Nucleic Acids Res* 2011, **39**:D32-D37.
29. Fu YX: Statistical tests of neutrality of mutations against population growth, hitchhiking and background selection. *Genetics* 1997, **147**:915-925.
30. Saitou N, Nei M: The neighbor-joining method - a new method for reconstructing phylogenetic trees. *Mol Biol Evol* 1987, **4**:406-425.
31. Wright S: Evolution in Mendelian populations. *Genetics* 1931, **16**:97-159.
32. Pearce JM: Philopatry: A return to origins. *Auk* 2007, **124**:1085-1087.

33. Peters JL, Omland KE: Population structure and mitochondrial polyphyly in North American Gadwalls (*Anas strepera*). *Auk* 2007, **124**:444-462.
34. Lokemoen JT, Duebber HF, Sharp DE: Homing and reproductive habits of mallards, gadwalls, and blue-winged teal. *Wildl Monogr* 1990, **106**:3-28.
35. Robertson GJ, Cooke F: Winter philopatry in migratory waterfowl. *Auk* 1999, **116**:20-34.
36. Bishop RA, Andrews RD: Survival and homing of female Mallards. *J Wildl Manage* 1978, **42**:192-196.
37. Cronin MA, Grand JB, Esler D, Derksen DV, Scribner KT: Breeding populations of northern pintails have similar mitochondrial DNA. *Can J Zool* 1996, **74**:992-999.
38. Pearce JM, Talbot SL, Pierson BJ, Petersen MR, Scribner KT, Dickson DL, Mosbech A: Lack of spatial genetic structure among nesting and wintering King Eiders. *Condor* 2004, **106**:229-240.
39. Peters JL, Gretes W, Omland KE: Late Pleistocene divergence between eastern and western populations of wood ducks (*Aix sponsa*) inferred by the 'isolation with migration' coalescent method. *Mol Ecol* 2005, **14**:3407-3418.
40. Gay L, Defos Du Rau P, Mondain-Monval JY, Crochet PA: Phylogeography of a game species: The red-crested pochard (*Netta rufina*) and consequences for its management. *Mol Ecol* 2004, **13**:1035-1045.
41. Champagnon J, Guillemin M, Elmberg J, Folkesson K, Gauthier-Clerc M: Changes in Mallard *Anas platyrhynchos* bill morphology after 30 years of supplemental stocking. *Bird Study* 2010, **57**:344-351.
42. Laikre L, Schwartz MK, Waples RS, Ryman N: Compromising genetic diversity in the wild: Unmonitored large-scale release of plants and animals. *Trends Ecol Evol* 2010, **25**:520-529.
43. Baratti M, Cordaro M, Dessi-Fulgheri F, Vannini M, Fratini S: Molecular and ecological characterization of urban populations of the mallard (*Anas platyrhynchos* L.) in Italy. *Ital J Zool* 2009, **76**:330-339.
44. Hofreiter M, Stewart J: Ecological change, range fluctuations and population dynamics during the Pleistocene. 2009, **19**:R584-R594.
45. Peters JL, McCracken KG, Zhuravlev YN, Lu Y, Wilson RE, Johnson KP, Omland KE: Phylogenetics of wigeons and allies (Anatidae: *Anas*): The importance of sampling multiple loci and multiple individuals. *Mol Phylogenet Evol* 2005, **35**:209-224.
46. Li J, Wang JXL: A modified zonal index and its physical sense. *Geophys Res Lett* 2003, **30**:34.
47. Lamb HH: Our understanding of global wind circulation and climatic variations. *Bird Study* 1975, **22**:121-141.
48. Liechti F: Birds: 'Blowin' by the wind? *J Ornithol* 2006, **147**:202-211.
49. Richardson WJ: Timing and amount of bird migration in relation to weather: a review. *Oikos* 1978, **30**:224-272.
50. Lyngs P: Migration and winter ranges of birds in Greenland. An analysis of ringing recoveries. *Dan Ornitol Foren Tidsskr* 2003, **97**:1-167.
51. Zeddeman A, van Hooft P, Prins HHT, Kraus RHS: Mallard (*Anas platyrhynchos*) gene pool connectivity between Greenland and Eastern Canada, Great Britain and The Netherlands. *Nuuk Ecological Research Operations, 2nd Annual Report, 2008* Jensen LM, Rasch M. Aarhus: National Environmental Research Institute, Aarhus University - Denmark; 2009, 67.
52. Evrard JO: Male philopatry in Mallards. *Condor* 1990, **92**:247-248.
53. Rohwer FC, Anderson MG: Female-biased philopatry, monogamy, and the timing of pair formation in migratory waterfowl. In *Current Ornithology. Volume 5*. Edited by: Johnston RF. New York: Plenum Press; 1988:187-221.
54. Huang Y, Zhao YH, Haley CS, Hu SQ, Hao JP, Wu CX, Li N: A genetic and cytogenetic map for the duck (*Anas platyrhynchos*). *Genetics* 2006, **173**:287-296.
55. Kraus RHS, Kerstens HHD, van Hooft P, Crooijmans RPMA, van Der Poel JJ, Elmberg J, Vignal A, Huang Y, Li N, Prins HHT, Groenen MAM: Genome wide SNP discovery, analysis and evaluation in mallard (*Anas platyrhynchos*). *BMC Genomics* 2011, **12**:150.
56. Mesa CM, Thulien KJ, Moon DA, Veniamin SM, Magor KE: The dominant MHC class I gene is adjacent to the polymorphic TAP2 gene in the duck, *Anas platyrhynchos*. *Immunogenetics* 2004, **56**:192-203.
57. Moon DA, Veniamin SM, Parks-Dely JA, Magor KE: The MHC of the duck (*Anas platyrhynchos*) contains five differentially expressed class I genes. *J Immunol* 2005, **175**:6702-6712.
58. Greenwood PJ: Mating systems, philopatry and dispersal in birds and mammals. *Anim Behav* 1980, **28**:1140-1162.
59. Pardini AT, Jones CS, Noble LR, Kreiser B, Malcolm H, Bruce BD, Stevens JD, Cliff G, Scholl MC, Francis M, Duffy CAJ, Martin AP: Sex-biased dispersal of great white sharks. *Nature* 2001, **412**:139-140.
60. Mank JE, Carlson JE, Brittingham MC: A century of hybridization: Decreasing genetic distance between American black ducks and mallards. *Conserv Genet* 2004, **5**:395-403.
61. Johnson KP, Sorenson MD: Phylogeny and biogeography of dabbling ducks (genus: *Anas*): A comparison of molecular and morphological evidence. *Auk* 1999, **116**:792-805.
62. McCracken KG, Johnson WP, Sheldon FH: Molecular population genetics, phylogeography, and conservation biology of the mottled duck (*Anas fulvigula*). *Cons Gen* 2001, **2**:87-102.
63. Funk DJ, Omland KE: Species-level paraphyly and polyphyly: Frequency, causes, and consequences, with insights from animal mitochondrial DNA. *Ann Rev Ecol Syst* 2003, **34**:397-423.
64. Maddison W, Knowles L: Inferring phylogeny despite incomplete lineage sorting. *Syst Biol* 2006, **55**:21-30.
65. Omland KE: Examining two standard assumptions of ancestral reconstructions: Repeated loss of dichromatism in dabbling ducks (*Anatini*). *Evolution* 1997, **51**:1636-1646.
66. Gonzalez J, Düttmann H, Wink M: Phylogenetic relationships based on two mitochondrial genes and hybridization patterns in Anatidae. *J Zool* 2009, **279**:310-318.
67. Smith LM, Burgoyne LA: Collecting, archiving and processing DNA from wildlife samples using FTA® databasing paper. *BMC Ecol* 2004, **4**:4.
68. Desjardins P, Morais R: Sequence and gene organization of the chicken mitochondrial genome. A novel gene order in higher vertebrates. *J Mol Biol* 1990, **212**:599-634.
69. Sorenson MD, Fleischer RC: Multiple independent transpositions of mitochondrial DNA control region sequences to the nucleus. *Proc Natl Acad Sci USA* 1996, **93**:15239-15243.
70. Sorenson MD, Ast JC, Dimcheff DE, Yuri T, Mindell DP: Primers for a PCR-based approach to mitochondrial genome sequencing in birds and other vertebrates. *Mol Phylogenet Evol* 1999, **12**:105-114.
71. Tamura K, Dudley J, Nei M, Kumar S: MEGA4: Molecular Evolutionary Genetics Analysis (MEGA) software version 4.0. *Mol Biol Evol* 2007, **24**:1596-1599.
72. Thompson JD, Higgins DG, Gibson TJ: Clustal W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Res* 1994, **22**:4673-4680.
73. Tajima F, Nei M: Estimation of evolutionary distance between nucleotide sequences. *Mol Biol Evol* 1984, **1**:269-285.
74. Clement M, Posada D, Crandall KA: TCS: A computer program to estimate gene genealogies. *Mol Ecol* 2000, **9**:1657-1659.
75. Rozas J, Sánchez-DelBarrio JC, Messeguer X, Rozas R: DnaSP, DNA polymorphism analyses by the coalescent and other methods. *Bioinformatics* 2003, **19**:2496-2497.
76. Excoffier L, Lischer HEL: Arlequin suite ver 3.5: A new series of programs to perform population genetics analyses under Linux and Windows. *Mol Ecol Res* 2010, **10**:564-567.
77. Guo SW, Thompson EA: Performing the exact Test of Hardy-Weinberg proportion for multiple alleles. *Biometrics* 1992, **48**:361-372.
78. Weir BS, Cockerham CC: Estimating F-statistics for the analysis of population structure. *Evolution* 1984, **38**:1358-1370.
79. Excoffier L, Smouse PE, Quattro JM: Analysis of molecular Variance inferred from metric distances among DNA haplotypes: Application to human mitochondrial-DNA restriction data. *Genetics* 1992, **131**:479-491.
80. Weir BS: *Genetic Data Analysis II: Methods for Discrete Population Genetic Data* Sunderland, MA, USA: Sinauer Assoc., Inc; 1996.
81. Peters JL, Zhuravlev Y, Fefelov I, Logie A, Omland KE: Nuclear loci and coalescent methods support ancient hybridization as cause of mitochondrial paraphyly between gadwall and falcated duck (*Anas spp.*). *Evolution* 2007, **61**:1992-2006.
82. Peters JL, Zhuravlev YN, Fefelov I, Humphries EM, Omland KE: Multilocus phylogeography of a Holarctic duck: Colonization of North America from Eurasia by gadwall (*Anas strepera*). *Evolution* 2008, **62**:1469-1483.
83. Hasegawa M, Kishino H, Yano T: Dating of the human-ape splitting by a molecular clock of mitochondrial DNA. *J Mol Evol* 1985, **22**:160-147.
84. Palsbøll PJ, Bérubé M, Aguilar A, Notarbartolo-Di-Sciara G, Nielsen R: Discerning between recurrent gene flow and recent divergence under a finite-site mutation model applied to North Atlantic and Mediterranean Sea fin whale (*Balaenoptera physalus*) populations. *Evolution* 2004, **58**:670-675.

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