



University of Groningen

Virtual Ontogeny of Cortical Growth Preceding Mental Illness

Patel, Yash; Shin, Jean; Abé, Christoph; Agartz, Ingrid; Alloza, Clara; Alnæs, Dag; Ambrogi, Sonia; Antonucci, Linda A.; Arango, Celso; Arolt, Volker

Published in: Biological Psychiatry

DOI:

10.1016/j.biopsych.2022.02.959

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version Publisher's PDF, also known as Version of record

Publication date: 2022

Link to publication in University of Groningen/UMCG research database

Citation for published version (APA):

Patel, Y., Shin, J., Abé, C., Agartz, I., Alloza, C., Alnæs, D., Ambrogi, S., Antonucci, L. A., Arango, C., Arolt, V., Auzias, G., Ayesa-Arriola, R., Banaj, N., Banaschewski, T., Bandeira, C., Başgöze, Z., Cupertino, R. B., Bau, C. H. D., Bauer, J., ... Paus, T. (2022). Virtual Ontogoto Cortical Growth Preceding Mental Illness. Biological Psychiatry, 92(4), 299-313. https://doi.org/10.1016/j.biopsych.2022.02.959

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: https://www.rug.nl/library/open-access/self-archiving-pure/taverneamendment.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): http://www.rug.nl/research/portal. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

Download date: 30-04-2023

Archival Report

Virtual Ontogeny of Cortical Growth Preceding Mental Illness

Yash Patel, Jean Shin, Christoph Abé, Ingrid Agartz, Clara Alloza, Dag Alnæs, Sonia Ambrogi, Linda A. Antonucci, Celso Arango, Volker Arolt, Guillaume Auzias, Rosa Ayesa-Arriola, Nerisa Banaj, Tobias Banaschewski, Cibele Bandeira, Zeynep Basgöze, Renata Basso Cupertino, Claiton H.D. Bau, Jochen Bauer, Sarah Baumeister, Fabio Bernardoni, Alessandro Bertolino, Caterina del Mar Bonnin, Daniel Brandeis, Silvia Brem, Jason Bruggemann, Robin Bülow, Juan R. Bustillo, Sara Calderoni, Rosa Calvo, Erick J. Canales-Rodríguez, Dara M. Cannon, Susanna Carmona, Vaughan J. Carr, Stanley V. Catts, Sneha Chenji, Qian Hui Chew, David Coghill, Colm G. Connolly, Annette Conzelmann, Alexander R. Craven, Benedicto Crespo-Facorro, Kathryn Cullen, Andreas Dahl, Udo Dannlowski, Christopher G. Davey, Christine Deruelle, Covadonga M. Díaz-Caneja, Katharina Dohm, Stefan Ehrlich, Jeffery Epstein, Tracy Erwin-Grabner, Lisa T. Eyler, Jennifer Fedor, Jacqueline Fitzgerald, William Foran, Judith M. Ford, Lydia Fortea, Paola Fuentes-Claramonte, Janice Fullerton, Lisa Furlong, Louise Gallagher, Bingchen Gao, Si Gao, Jose M. Goikolea, Ian Gotlib, Roberto Goya-Maldonado, Hans J. Grabe, Melissa Green, Eugenio H. Grevet, Nynke A. Groenewold, Dominik Grotegerd, Oliver Gruber, Jan Haavik, Tim Hahn, Ben J. Harrison, Walter Heindel, Frans Henskens, Dirk J. Heslenfeld, Eva Hilland, Pieter J. Hoekstra, Sarah Hohmann, Nathalie Holz, Fleur M. Howells, Jonathan C. Ipser, Neda Jahanshad, Babette Jakobi, Andreas Jansen, Joost Janssen, Rune Jonassen, Anna Kaiser, Vasiliy Kaleda, James Karantonis, Joseph A. King, Tilo Kircher, Peter Kochunov, Sheri-Michelle Koopowitz, Mikael Landén, Nils Inge Landrø, Stephen Lawrie, Irina Lebedeva, Beatriz Luna, Astri J. Lundervold, Frank P. MacMaster, Luigi A. Maglanoc, Daniel H. Mathalon, Colm McDonald, Andrew McIntosh, Susanne Meinert, Patricia T. Michie, Philip Mitchell, Ana Moreno-Alcázar, Bryan Mowry, Filippo Muratori, Leila Nabulsi, Igor Nenadić, Ruth O'Gorman Tuura, Jaap Oosterlaan, Bronwyn Overs, Christos Pantelis, Mara Parellada, Jose C. Pariente, Paul Pauli, Giulio Pergola, Francesco Maria Piarulli, Felipe Picon, Fabrizio Piras, Edith Pomarol-Clotet, Clara Pretus, Yann Quidé, Joaquim Radua, J. Antoni Ramos-Quiroga, Paul E. Rasser, Andreas Reif, Alessandra Retico, Gloria Roberts, Susan Rossell, Diego Luiz Rovaris, Katya Rubia, Matthew D. Sacchet, Josep Salavert, Raymond Salvador, Salvador Sarró, Akira Sawa, Ulrich Schall, Rodney Scott, Pierluigi Selvaggi, Tim Silk, Kang Sim, Antonin Skoch, Gianfranco Spalletta, Filip Spaniel, Dan J. Stein, Olaf Steinsträter, Aleks Stolicyn, Yoichiro Takayanagi, Leanne Tamm, Maria Tavares, Alexander Teumer, Katharina Thiel, Sophia I. Thomopoulos, David Tomecek, Alexander S. Tomyshev, Diana Tordesillas-Gutiérrez, Michela Tosetti, Anne Uhlmann, Tamsyn Van Rheenen, Javier Vazquez-Bourgón, Meike W. Vernooij, Eduard Vieta, Oscar Vilarroya, Cynthia Weickert, Thomas Weickert, Lars T. Westlye, Heather Whalley, David Willinger, Alexandra Winter, Katharina Wittfeld, Tony T. Yang, Yuliya Yoncheva, Jendé L. Zijlmans, Martine Hoogman, Barbara Franke, Daan van Rooij, Jan Buitelaar, Christopher R.K. Ching, Ole A. Andreassen, Elena Pozzi, Dick Veltman, Lianne Schmaal, Theo G.M. van Erp, Jessica Turner, F. Xavier Castellanos, Zdenka Pausova, Paul Thompson, and Tomas Paus

ABSTRACT

BACKGROUND: Morphology of the human cerebral cortex differs across psychiatric disorders, with neurobiology and developmental origins mostly undetermined. Deviations in the tangential growth of the cerebral cortex during pre/perinatal periods may be reflected in individual variations in cortical surface area later in life.

METHODS: Interregional profiles of group differences in surface area between cases and controls were generated using T1-weighted magnetic resonance imaging from 27,359 individuals including those with attention-deficit/hyperactivity disorder, autism spectrum disorder, bipolar disorder, major depressive disorder, schizophrenia, and high general psychopathology (through the Child Behavior Checklist). Similarity of interregional profiles of group differences in surface area and prenatal cell-specific gene expression was assessed.

RESULTS: Across the 11 cortical regions, group differences in cortical area for attention-deficit/hyperactivity disorder, schizophrenia, and Child Behavior Checklist were dominant in multimodal association cortices. The same interregional profiles were also associated with interregional profiles of (prenatal) gene expression specific to proliferative cells, namely radial glia and intermediate progenitor cells (greater expression, larger difference), as well as differentiated cells, namely excitatory neurons and endothelial and mural cells (greater expression, smaller difference). Finally, these cell types were implicated in known pre/perinatal risk factors for psychosis. Genes coexpressed with radial glia were enriched with genes implicated in congenital abnormalities, birth weight, hypoxia, and starvation. Genes coexpressed with endothelial and mural genes were enriched with genes associated with maternal hypertension and preterm birth.

CONCLUSIONS: Our findings support a neurodevelopmental model of vulnerability to mental illness whereby prenatal risk factors acting through cell-specific processes lead to deviations from typical brain development during pregnancy.

https://doi.org/10.1016/j.biopsych.2022.02.959

The majority of symptoms of mental illness, from hallucinations and delusions in psychosis to the impaired attention and cognitive control in attention-deficit/hyperactivity disorder (ADHD), are rooted in disturbances of perceptual, cognitive, and affective processes subserved by the cerebral cortex. The human cerebral cortex is a highly folded sheath of tissue (~1800 cm² of surface area) containing approximately 12 billion neurons and 17 billion non-neuronal cells (1). Both global and regional expansion of the primate cerebral cortex are driven by biological events taking place during fetal development; the phase of symmetrical division of progenitor cells in the proliferative zones during the first trimester is particularly important for tangential growth through addition of ontogenetic columns (2). Although neurogenesis—and related additions of ontogenetic columns-ends before birth, the surface area of the cerebral cortex continues to increase during the first 2 to 4 years of human life (3). But subsequent changes in the surface area of the human cerebral cortex, as estimated with magnetic resonance imaging (MRI), are comparatively minimal (4-6). Quantitively, a majority of the cortical expansion occurs prenatally and perinatally, with the most prominent rate in cortical expansion occurring during prenatal development (Figure S1) (7-10). Moreover, cortical surface area in children, adolescents, and young adults is correlated with birth weight, a common indicator for healthy neurodevelopment (11,12). The genetics of cortical surface area also implicates neurodevelopmental proliferative cells as compared with adult cell types (13,14). Therefore, in the adult brain, measures of cortical surface area provide a window into events shaping prenatal and early postnatal growth of the cerebral cortex that predate a broad array of mental illnesses (13,15-17).

To gain insights into the neurodevelopmental events that may underlie differential growth of the cerebral cortex in individuals with mental illness and/or the presence of clinically significant psychopathology (vs. healthy individuals) and the influence of external risk factors, we first estimated the extent of such group differences between cases¹ and controls in the surface areas of 11 cortical regions (due to corresponding availability of fetal gene expression data). We then identified cellular elements underlying interregional variations in these group differences using virtual ontogeny, through which interregional profiles of group differences in surface area were correlated with interregional profiles of gene expression. The latter were restricted to transcripts expressed during 12 to 22 postconception weeks (PCWs) and to the following cell types: radial glia, intermediate progenitor cells (IPCs), excitatory neurons, interneurons, oligodendrocyte progenitor cells, microglia, and endothelial and mural cells. Finally, we asked which of these cell types might mediate the impact on cortical growth of prenatal factors reported to increase the risk of developing psychosis-risk factors applicable to many mental illnesses in general.

Cases are defined as individuals with a diagnosis of the following conditions: schizophrenia, autism spectrum disorder, attentiondeficit/hyperactivity disorder, bipolar disorder, and major depressive disorder, or by the presence of symptoms of psychopathology as assessed with the Child Behavior Checklist in a large community-based sample of children (the ABCD Study).

METHODS AND MATERIALS

Meta-analytic Group Differences in Cortical Surface Area

T1-weighted MRI scans were acquired in 89 cohorts participating in the ENIGMA (Enhancing Neuro Imaging Genetics through Meta Analysis) Consortium. The ENIGMA Consortium is a collaborative initiative in global neuroscience and focuses on studying the human brain in health and disease through genetics and imaging (18). Sample demographics and MRI acquisition details per cohort are provided in Tables S1-S7. FreeSurfer cortical reconstruction software was used to extract surface area according to a parcellation scheme that intersects with tissue sampling from the PsychENCODE Consortium, described in Supplemental Methods and presented in Figure S2. Individual ENIGMA cohorts performed multiple linear regression analysis, modeling surface area of each cortical region separately as a function of diagnosis status, age, age squared, sex, and site-specific covariates (such as MR scanner, multiple sites). Cohort-specific information regarding diagnostic and sampling criteria are described in previously published ENIGMA reports (19-23). Individual cohorts obtained institutional ethics approval, and informed consent was obtained from study participants or guardians. Cohort-level summary statistics were then meta-analyzed using an inverse variance-weighted random effects model from the "metafor" R package (24). Meta-analytic estimates are provided in Tables S8-S12.

The ABCD (Adolescent Brain Cognitive Development) Study is a longitudinal cohort study of brain development on roughly ~11,500 children sampled across the United States from the general community (25). T1-weighted MRI data from the ABCD Study were processed with FreeSurfer version 7.1 on the Compute Canada Niagara server (26). MRI and sample recruitment procedures for the ABCD Study have been described previously (25,27). Psychopathology was indexed by the total problem score from the parent-completed Child Behavior Checklist (CBCL)—a simple index of global psychopathology (28). The top and bottom 20% of the CBCL total score distribution (stratified by sex and ethnicity) was used to classify cases and controls, respectively (Figure S2B). Note that this extremes-only approach minimizes possible noise in CBCL data resulting from the known discrepancies between parental reports (used here) and self-reports. Linear mixedeffects models for each cortical region were run as a function of high/low psychopathology, age, age squared, sex, ethnicity, and random effects (family structure and MRI machine). The "Ime4" R package was used to run mixed-effects models (29).

Virtual Ontogeny

To gain insights into the relationship between prenatal development and postnatal group differences in cortical surface area, we proceeded by following three steps (depicted in Figure 1). First, we identified gene-expression markers specific to a set of cells present in the human cerebral cortex toward the end of the first and throughout the second trimester (30–32). To do so, we used publicly available single-cell data from the developing cerebral cortex of 5 donors, with

postconception age ranging from 5 to 22 PCWs (30) (Figures S4 and S5). Second, we used these cell-specific genes and calculated the median value of their expression (200 genes per cell type) for each of the 11 cortical regions for which group differences in surface area were examined (steps 2 and 3 from Figure 1). These expression values were derived from the PsychENCODE bulk RNA sequencing dataset (14 donors, 12-22 PCWs) (33). The processing of single-cell and bulk RNA sequencing data is described in the Supplement. Third, the interregional profiles of the (median) expression of these marker genes were correlated with the interregional profiles of group differences in cortical surface area from Figure 2A (step 4 from Figure 1). The average MRI-expression correlation was tested for significance using a permutationbased approach with 10,000 resamplings of random gene lists, as described in detail in the Supplement (step 5 from Figure 1). We also performed two additional sensitivity analyses 1) to estimate the distribution of the average correlation coefficient between MRI and cell-specific gene expression by bootstrapping the 200 gene expression profiles per cell type and 2) to use gene set enrichment analysis as a test of overrepresentation of cell-specific genes within the rank-ordered list of MRI-gene expression correlations (34,35).

Gene Coexpression and Enrichment Analyses

The virtual ontogeny analysis focused exclusively on the limited set of cell-specific genes. To expand the focus of genes investigated while simultaneously interjecting findings from our cell-specific approach, we used genome-wide coexpression analysis including all prenatal donors from the PsychENCODE dataset. Modeling of coexpression is presented in the Supplement. Next, coexpressed gene panels for cell types that showed significance from virtual ontogeny were used as inputs for several enrichment analyses, including 1) gene ontology enrichment, 2) disorder-related gene set enrichment, 3) cortical surface area gene enrichment from prior ENIGMA genome-wide association study data, and 4) enrichment with genes associated with risk factors for psychosis. The details for each analysis are presented in the Supplement.

RESULTS

Case-Control Differences in Surface Area and Expression of Proliferative-Cell Genes

Meta-analytic profiles of group differences in cortical surface area were quantified using structural T1-weighted brain MRI scans. Cohorts from the ENIGMA Consortium contributed MRI scans of individuals diagnosed with schizophrenia (SCZ), ADHD, autism spectrum disorder (ASD), bipolar disorder, and major depressive disorder. In addition, children from the ABCD Study were classified into two groups with high or low psychopathology, defined as the top and bottom 20%, respectively, of the CBCL total problem score (Figure S2). This cohort of children allowed us to extend findings obtained in patients with an established clinical diagnosis to young people with emerging psychopathology from the general community (25). In total, 27,359 individuals contributed to group differences in cortical surface area across 11 cortical regions (Figure 2A, B; Tables S2–S6). These specific regions (and time period) were

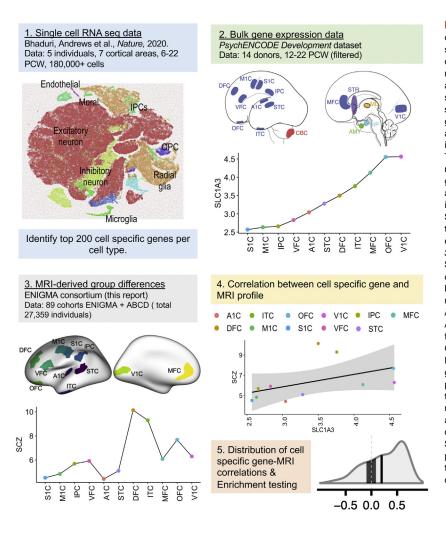


Figure 1. Methodological workflow for virtual ontogeny. Step 1 (top left): identify top 200 cellspecific genes from single-cell RNA sequencing data of the developing neocortex (30). Step 2 (top right): quantify median gene expression (bulk RNA) across donors for each of 11 cortical regions sampled from the PsychENCODE dataset (33). Cell specificity was defined as the ratio of expression of a gene in a given cell type divided by the expression across all cells. For instance, the gene SLC1A3 was in the top 200 specific genes for the radial-glia panel. The expression of this gene is plotted in step 2 (top right). Step 3 (bottom left): quantify meta-analytic group differences in surface area between cases and controls across the 11 cortical regions sampled in the PsychENCODE dataset. Group differences for SCZ are plotted as an example. Step 4 (bottom right, top half): correlation between cell-specific gene expression and an MRI-derived profile, in this case, SLC1A3 expression and case-control differences for SCZ. This is repeated for all 200 genes specific to a cell type (in this case, radial glia) to create a distribution of correlation coefficients in step 5 (bottom right, bottom half). A1C, primary auditory cortex; ABCD, Adolescent Brain Cognitive Development; AMY, amygdala; CBC, cerebral cortex; DFC, dorsal frontal cortex; ENIGMA, Enhancing Neuro Imaging Genetics through Meta Analysis; HIP, hippocampus; IPC, inferior parietal cortex; IPCs, intermediate progenitor cells; ITC, inferior temporal cortex; M1C, primary motor cortex; MD, mediodorsal nucleus of thalamus; MDD, major depressive disorder; MFC, medial frontal cortex; MRI, magnetic resonance imaging: OFC, orbitofrontal cortex: OPC, oligodendrocyte progenitor cell; PCW, postconception week; S1C, primary somatosensory cortex; SCZ, schizophrenia; STC, superior temporal cortex; STR, striatum; V1C, primary visual cortex; VFC, ventral frontal cortex.

selected based on the availability of gene expression data during gestation (Figures S3 and S4) (33).

Case-control differences in surface area were greatest in patients with SCZ and ADHD, and in the community sample of children with high CBCL psychopathology scores (Figure 2A; Tables S7–S12). Interregional profiles across the 11 cortical regions were highly correlated between SCZ and ADHD (Figure 2C). At the nominal level of significance (p < .05), we also observed correlations between the CBCL profile and both the ADHD and SCZ profiles (Figure 2C).

What neurodevelopmental processes might underlie these group differences? To answer this, we related interregional profiles of cell-specific gene expression in the developing cerebral cortex (12–22 PCWs) with interregional profiles of group differences in cortical area across the same 11 regions. These case-control group differences were used as input to the analytic framework depicted in Figure 1. This "virtual ontogeny" analysis revealed positive associations between prenatal expression profiles of proliferative cells, namely radial glia and IPCs, and postnatal profiles of group differences in SCZ, ADHD, CBCL, and ASD (Figure 3A, B; Table S13). Likewise, these group contrasts showed negative associations with a

number of differentiated cells, namely excitatory neurons and endothelial and mural cells². We tested the sensitivity of these findings using two different statistical approaches: 1) bootstrapped estimation of the correlation-coefficient distribution and 2) gene-set enrichment analysis (Figures S6 and S7, respectively). These somewhat more conservative analyses confirm the general opposing pattern of enrichment with radial glia/IPCs and excitatory neurons with ADHD, SCZ, and ASD. This association was nominally significant for CBCL. In the next steps, we focused on results specific to SCZ, ADHD, and CBCL because these profiles presented robust group differences in surface area (Figure 2A).

Multimodal Associative Versus Primary/Unimodal Cortex

Unsupervised hierarchical clustering of interregional profiles of group differences in surface area revealed two distinct sets of

²Undifferentiated (radial glia, intermediate progenitor cells); differentiated (neurons, microglia, oligodendrocytes, and mural and endothelial cells). Oligodendrocyte progenitor cells are a hybrid state.

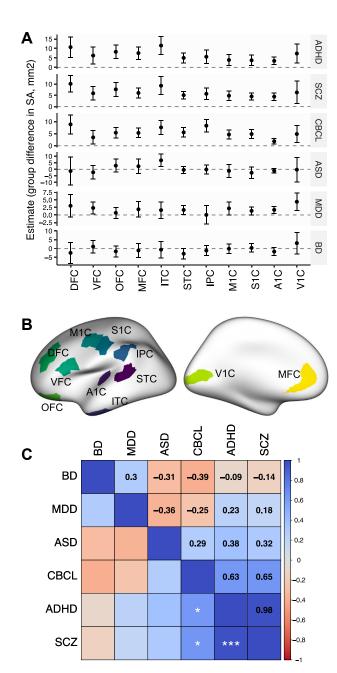


Figure 2. Regional differences in cortical surface area across multiple psychiatric conditions. (A) Meta-analytic estimates of group differences in cortical surface between cases and controls. Contrast shown as controls minus cases, where positive values indicate smaller surface area in cases. (B) Schematic location of regions of interest from which surface area was quantified. (C) Cross-disorder correlation matrix of profiles from panel (A). *Nominal p < .05; ***false discovery ratecorrected p < .05. A1C, primary auditory cortex; ADHD, attentiondeficit/hyperactivity disorder; ASD, autism spectrum disorder; BD, bipolar disorder; CBCL, Child Behavior Checklist; DFC, dorsal frontal cortex; IPC, inferior parietal cortex; ITC, inferior temporal cortex; M1C, primary motor cortex; MDD, major depressive disorder; MFC, medial frontal cortex; OFC, orbitofrontal cortex; SA, surface area; S1C, primary somatosensory cortex; SCZ, schizophrenia; STC, superior temporal cortex; V1C, primary visual cortex; VFC, ventral frontal cortex.

cortical regions (Figure 4A). Cluster 1 consisted of multimodal associative cortices³ while cluster 2 contained mostly primary and unimodal cortices⁴. The group differences in cortical surface area for SCZ, ADHD, and CBCL were greater in multimodal versus primary/unimodal cortices (Figure 4B; Figure S8). Cell-specific gene expression trajectories during gestation also revealed remarkable differences between these two clusters: proliferative (i.e., undifferentiated) cells have greater cell-specific expression in the multimodal cortices while differentiated cells have greater expression in primary/unimodal cortices (Figure 4C; Table S14).

Genetics of Psychiatric Conditions and Cortical Growth: Intersection With Cell-Specific Gene Coexpression Networks

As described above, we observed a certain degree of similarity in interregional profiles of group differences in the cortical surface area among the different mental health conditions (particularly with SCZ, ADHD and CBCL) (Figure 2C). To capture these similarities, we carried out principal component (PC) analysis of the interregional profiles. This analysis revealed clear demarcation between the multimodal and primary/ unimodal clusters, respectively (Figure 5A), with PC1 explaining 50% of the variance and PC1 correlating highly with SCZ, ADHD, and CBCL (Figure 5B), As expected from the conditionspecific analyses (Figure 3), virtual ontogeny of the PC1 loadings showed positive associations with radial glia and IPCs and showed negative associations with excitatory neurons and endothelial and mural cells (Figure 5C; Figure S9). Sensitivity analyses confirmed significant associations with radial glia, IPCs, and excitatory neurons, with a weaker finding for the mural cells (Figure S10). To investigate further the processes underlying the association between PC1 and cellspecific genes, we generated coexpression panels of genes for each cell type associated with PC1, expanding the scope of our work from cell-specific genes to all related genes. Gene Ontology enrichment analysis revealed a number of specific biological processes associated with each cell type-specific coexpressed panel. Thus, radial alia and IPC genes were highly enriched for biological processes relating to cell division. while vasculature-forming endothelial and mural cells as well as excitatory neurons were enriched, respectively, for blood vessel morphogenesis and synaptic signaling/organization (Figure 5D-F). Genes associated with schizophrenia, as derived from genetic variant studies (36), were enriched in coexpression networks of the radial glia and excitatory neurons (Figure 5G). Genes associated with the cortical expansion of multimodal cortices, as derived from genome-wide association studies (14), were enriched in coexpression networks of the radial glia and IPCs (Figure 5H; Table S14). Note that the latter enrichment was not found in the case of unimodal cortices, pointing again at the distinction of the two types of

³Multimodal associative cortices in cluster 1 (intermediate progenitor cell, orbital frontal cortex, medial frontal cortex, dorsal frontal cortex).

⁴Primary/unimodal cortices in cluster 2 (primary visual cortex, ventral frontal cortex, primary motor cortex, primary somatosensory cortex, primary auditory cortex, superior temporal cortex).

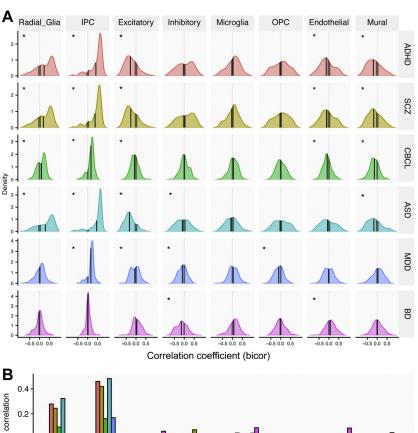
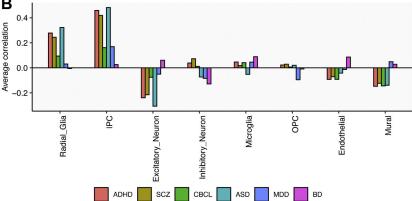


Figure 3. Virtual ontogeny. (A) Distribution of correlation coefficients between prenatal cellspecific gene expression and postnatal group differences in cortical surface area. Gray box around zero represents 99% confidence intervals from the null distribution generated through 10,000 resamplings of gene expression and group-difference profiles. Black vertical line represents the mean correlation coefficient (biweight midcorrelation) of the distribution, also plotted in panel (B). *False discovery rate-corrected p value < .01. ADHD, attention-deficit/hyperactivity disorder; ASD, autism spectrum disorder; BD, bipolar disorder; bicor, biweight midcorrelation; CBCL, Child Behavior Checklist; IPC, intermediate progenitor cell; MDD, major depressive disorder; OPC, oligodendrocyte progenitor cell; SCZ, schizophrenia.



cerebral cortices with respect to their neurodevelopmental characteristics and/or developmental timing.

Cell Types and Prenatal Risk for Psychosis

Experimental studies have pointed to a number of external factors that may interfere with typical development of the cerebral cortex in nonhuman primates (37,38). Similarly, epidemiological studies have identified a number of pre/perinatal risk factors associated with later emergence of psychosis (such as low birth weight and preterm birth) (39). These risk factors can be generalizable to most neurodevelopmental disorders.

Here, we tested which of the cell types associated with the PC1 profile of group differences in surface area might mediate the impact of risk factors for psychosis on prenatal growth of the human cerebral cortex. Prenatal risk factors for psychosis were identified from a systematic review and meta-analysis that included 152 studies (Figure 6A) (39). We selected, a priori, sets of genes linked to each of these risk factors using either relevant Gene Ontology terms (40) or genes associated with a particular condition (e.g., congenital abnormalities), as identified in curated datasets based on genome-wide association study catalogs, animal models, and the greater scientific literature (Table S15) (36,41). The results showed that genes implicated in congenital abnormalities were enriched with the radial glia, IPCs, and mural cell-specific coexpressed panels (Figure 6B; Table S17). Genes pertaining to birth weight, hypoxia, and famine were also enriched in the radial glia panel. In contrast, genes pertaining to the regulation of blood pressure (and, therefore, relevant to maternal hypertension during pregnancy), as well as genes associated with preterm birth,

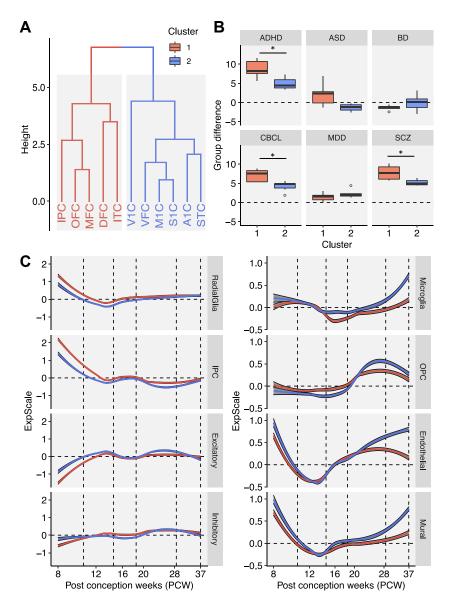


Figure 4. Differences in cortical surface area cluster into associative and primary/unimodal cortex. (A) Hierarchical clustering dendrogram of group differences in cortical surface area with k = 2 clusters. (B) Boxplot depicting group differences between clusters for each of the six profiles investigated. (C) LOESS model fits of cell-specific gene expression trajectories stratified by cortical cluster. Expression (y-axis) is unit scaled. Shaded gray region around the model fit represents 95% confidence intervals. Vertical black dashed lines represent prominent windows of neurodevelopment reported previously (33). A1C, primary auditory cortex; ADHD, attention-deficit/hyperactivity disorder; ASD, autism spectrum disorder; BD, bipolar disorder: CBCL. Child Behavior Checklist: DFC. dorsal frontal cortex; IPC, intermediate progenitor cell; IPC, inferior parietal cortex; ITC, inferior temporal cortex; M1C, primary motor cortex; MDD, major depressive disorder: MFC. medial frontal cortex: OFC. orbitofrontal cortex; OPC, oligodendrocyte progenitor cell; PCW. postconception week; S1C, primary somatosensory cortex; SCZ, schizophrenia; STC, superior temporal cortex; V1C, primary visual cortex; VFC, ventral frontal cortex.

were enriched in the mural panel. Although preeclampsia was not a significant risk factor for psychosis [odds ratio = 1.32, p = .059 from (39)], genes associated with this condition intersected with those included in the endothelial and mural panels (Figure S11).

DISCUSSION

It appears that the differential growth of the cerebral cortex preceding mental illness and general psychopathology in childhood 1) is more pronounced in multimodal (vs. primary/unimodal) cortical regions, 2) is related to the spatial pattern of prenatal expression of genes underlying neuro- and angiogenesis, and 3) might be reflective of influences of known risk factors acting on these cellular processes during prenatal development.

Cortical regions that show the largest case-control group differences in surface area are regions with greater prenatal expression of proliferative cells (radial glia, IPCs) and lower expression of differentiated cells such as excitatory neurons and endothelial and mural cells during the first trimester. This implies potential disruption in processes of progenitor expansion and subsequent differentiation, with possible cascading effects in later (postnatal) developmental periods. Radial glia serve as a key progenitor population driving neurogenesis and creating a vertical scaffold for neuronal migration from proliferative zones to the cortical plate (2). According to the radial unit hypothesis, the cortical surface area of a given region depends on the number of contributing proliferative units (2); experimental enhancement of the neural progenitor population results in greater surface expansion and folding (42). Subtle deviations in progenitor cell division may have a profound

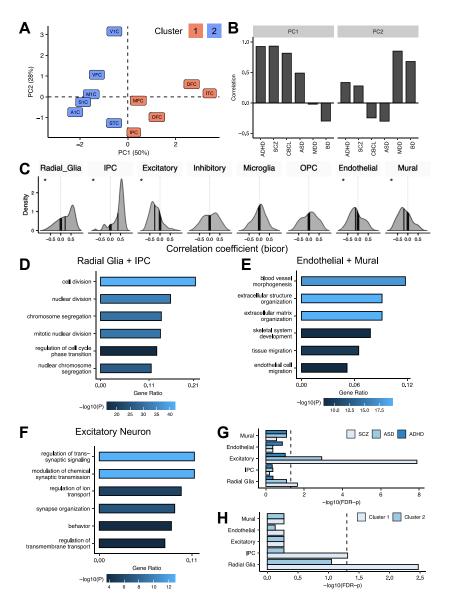


Figure 5. Enrichment of cell-specific gene panels. (A) Principal component analysis plot of regional loadings of PC1 and PC2. (B) Correlation between disorder-specific profiles and PC1/PC2 loadings. (C) Virtual ontogeny analysis depicting distributions of correlation between interregional variation in cellspecific gene expression and PC1 loadings (across the 11 regions). *FDR p < .01. (D-F) Gene Ontology enrichment analysis of coexpressed cell-specific gene panels. Gene ratio represents the proportion of genes in the cell-specific panel that intersect with a Gene Ontology term with the total size of the gene set. (G) Enrichment analysis for disorder-associated genes for the three disorders loading strongest on PC1 (SCZ, ADHD, and ASD) and for (H) cortical surface area-associated genes of clusters 1 and 2. A1C, primary auditory cortex; ADHD, attentiondeficit/hyperactivity disorder; ASD, autism spectrum disorder; BD, bipolar disorder; CBCL, Child Behavior Checklist: DFC, dorsal frontal cortex: FDR. false discovery rate; IPC, intermediate progenitor cell: IPC, inferior parietal cortex: ITC, inferior temporal cortex; M1C, primary motor cortex; MDD, major depressive disorder; MFC, medial frontal cortex; OFC, orbitofrontal cortex; OPC, oligodendrocyte progenitor cell; PC, principal component; S1C, primary somatosensory cortex; SCZ, schizophrenia; STC, superior temporal cortex; V1C, primary visual cortex; VFC, ventral frontal cortex.

impact on the resulting neuronal population owing to the selfrenewing (amplifying) nature of radial glia and IPCs: two radial glia cells may generate more than 80 neurons following eight rounds of cellular division (43). For instance, loss of the DISC1 gene, a genetic locus of relevance for schizophrenia among other mental illnesses, reduces neural-progenitor proliferation, leading to premature differentiation (44). This parallels the observed intersection between genes associated with SCZ and genes in the radial glia coexpression network associated with group differences in cortical surface area between patients with SCZ and healthy control subjects (Figures 2A and 4H). We also observed associations with endothelial and mural cells, components of the developing cortical blood vessels. The development, growth, and maturation of cerebral vasculature and neural structures occurs simultaneously with bidirectional signaling and influences [reviewed in (45)]. Neural-

derived signals control angiogenesis and blood vessel patterning, while vascularization modulates the extent of neurogenesis and progenitor differentiation. Given that neurogenic niches require hypoxic conditions for progenitor cell expansion, a spatiotemporal balance between expansion and differentiation is controlled, in part, by blood vessel formation and subsequent oxygenation (45,46).

Multimodal (association) cortices appear to stand out, with regard to both the observed group differences in their surface area and the spatiotemporal pattern of prenatal expression of genes specific to undifferentiated (proliferative) and differentiated (neurons, vasculature) cells. Generally speaking, these cortical regions subserve complex perceptual and cognitive processes, building on information received from unimodal cortices. Previous studies have pointed to a prolonged developmental time course as one of the characteristics

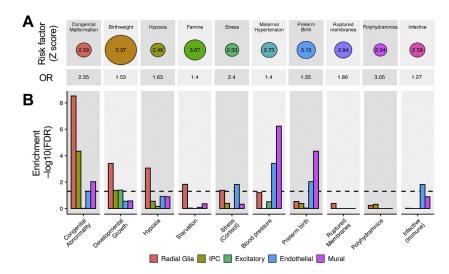


Figure 6. Risk factors of psychosis with implicated cell types. **(A)** *Z* scores for pre/perinatal risk factors for psychosis from Davies *et al.* (39) are represented by the size of the circle, and the corresponding odds ratio is in the text below. **(B)** Enrichment between genes implicated in risk factors for psychosis and coexpressed cell-specific gene panels identified to be related to group differences in cortical surface area. Horizontal dashed line represents FDR < .05. FDR, false discovery rate; IPC, intermediate progenitor cell; OR, odds ratio.

distinguishing multimodal and primary cortices. Evidence supporting this view includes a prolonged existence of the transient associative subplate as compared with primary cortices (47), less dendritic shaft/spine growth at birth (48), and a delayed maturation of projection fibers in associative white matter (49). The prolonged existence of the associative subplate may be of particular importance for disorders characterized by alterations in complex perceptual and cognitive processes because these neurons play key roles in axonal pathfinding, cell survival, and guiding cortical circuitry maturation and, as such, in the development of corticocortical associative fibers [see review in (50)]. Postnatally, functional MRI and structural (tract tracing) studies in humans and macaques, respectively, have shown a principal gradient in cortical connectivity of multimodal regions distinct from the primary cortex (51). These regions are also situated in key nodes of the default mode network, in which aberrant activity is implicated in many, if not all, psychiatric conditions (51,52). Taken together, delayed maturation of association cortices correlates with greater vulnerability to genetic or environmental perturbations.

The neurodevelopmental theory of schizophrenia, as per Murray (53) and Weinberger (54,55), has sparked intense interest in early events that may increase the risk of developing this mental illness later in life. As summarized recently, a number of prenatal and perinatal factors appear to increase the risk to developing psychosis (39). Here, we provide initial evidence that links, albeit indirectly, such risk factors to SCZ via cellular processes underpinning cortical growth during prenatal development (Figure 5). We have identified two possible-mutually nonexclusive-pathways. The first oneat play in cases of low birth weight, hypoxia, and famineinvolves radial glia (i.e., proliferation). The other one-at play in cases of maternal hypertension, preeclampsia, and preterm birth-involves endothelial and mural cells (i.e., vasculature). Nutrient restriction in animal models (nonhuman primates and other vertebrates) produces impaired function of progenitor cells, cell-cycle arrest, and increased cell death (38,56). Likewise, rat models of hypoxia-ischemia-related injury in the developing cortex show marked reduction in the population of neural stem cells (57). In contrast, experimental models of preeclampsia (a hypertensive syndrome) have shown abnormal cerebrovascular morphology and permeability/growth [reviewed in (58)]. The latter parallels our intersection between maternal hypertension (and preeclampsia) and endothelial and mural cells. Finally, the broad classification of congenital malformations was strongly associated with radial glia/IPC genes as well as endothelial and mural cells, hinting at the close (likely bidirectional) relationship between corticogenesis and developing blood vessels [reviewed in (45)].

Limitations and Considerations

It is important to qualify the findings from this report, given the nature of the comparisons between different datasets and periods in time. Group differences in cortical surface area likely indicate a general vulnerability to developing psychopathology, but it is not a feature that distinguishes what kind of disorder an individual may manifest later in postnatal life.

These findings allow us and others to formulate follow-up hypotheses to be tested experimentally, possibly with the advancement in cortical organoid modeling (59). The findings were limited by the availability of prenatal gene-expression data given the limited sampling of cortical regions (only 11 regions) and the limited number of donors from various periods of gestation (missing data from very early and later stages of prenatal development). Statistically, it would be most straightforward to relate the spatial profile of group differences in surface area with the average gene-expression profile specific to cell types; with only 11 regions, however, there is little statistical power. To address this limitation, we have used resampling-based approaches along with sensitivity analyses to test for cell-specific associations. Likewise, the geneexpression dataset was sampled from the cortical plate, while cellular division, differentiation, and maturation take place within the ventricular, subventricular, and intermediate zones of the developing cerebral cortex. This necessitates the assumption of similar interregional expression profiles reflected across developing lamina, as postulated in the protomap hypothesis (2).

We investigated exclusively the prenatal period in relation to group differences in cortical surface area for several reasons: 1) the dominance of prenatal period vis-a-vis the tangential growth of the cerebral cortex (surface area) as shown from experimental (37,60) and genetic (13,14) studies, 2) epidemiologic evidence implicating birth weight (an index of healthy brain growth) and risk for psychiatric disorder diagnosis (16), and 3) enrichment of neurodevelopmental cell types/processes in genetic variants associated with multiple psychiatric disorders (13,15,17). Even so, this is not to say that developmental disturbances during postnatal life, especially during infancy, may not contribute to the surface area sampled later in life. There are three key periods of cortical expansion: 1) greatest expansion during gestation, 2) expansion from birth to the first 2 years of life, and 3) subtle increases until the end of childhood (depicted in Figure S1) (7-10). It is very likely, however, that different processes underly cortical expansion in these different stages of brain development. Prenatally (before birth), expansion is determined through addition of ontogenetic columns (2,43). Between birth and the first 2 years of life, cortical growth may be a consequence of the expansion in neuropil and cortical minicolumns (61-63). Following 2 years of age, cortical expansion may be related to the growth of underlying white matter (64). The processes governing cortical expansion after birth have not been systematically evaluated. Nonetheless, we observe signals relevant to neurodevelopmental cells (radial glia/IPCs) in cohorts with vastly different age ranges such as those in the ENIGMA ASD, ENIGMA ADHD, and ABCD CBCL groups, which were predominately younger, as compared with the (older) ENIGMA SCZ group. This supports our assumption about the importance of the pre/perinatal environment and cortical surface area. Taken together, it is likely that perturbations of early development may have a sizable impact on cortical surface area measured later in life, primarily through neurogenesis and subsequent expansion of neuropil.

Conclusions

In summary, we show that a simple in vivo measure of brain structure, namely surface area of a set of cortical regions, acquired many years after birth provides an anchor for identifying developmental processes at play before birth and for suggesting cellular mechanisms that may mediate the known associations between common pre- and perinatal risk factors and severe mental illness.

ACKNOWLEDGMENTS AND DISCLOSURES

Data used in the preparation of this article were obtained from the Adolescent Brain Cognitive Development (ABCD) Study (https://abcdstudy.org), held in the National Institute of Mental Health (NIMH) Data Archive. This is a multisite, longitudinal study designed to recruit more than 10,000 children age 9 to 10 and follow them over 10 years into early adulthood. The ABCD Study is supported by the National Institutes of Health (NIH) and additional federal partners under Grant Nos. U01DA041048, U01DA050989, U01DA051016, U01DA041022, U01DA051018, U01DA051037, U01DA050987.

U01DA041028, U01DA041134, U01DA050988, U01DA051039, U01DA041156, U01DA041025, U01DA041120, U01DA051038, U01DA041148, U01DA041093, U01DA041089, U24DA041123, and U24DA041147. A full list of supporters is available at https://abcdstudy.org/federal-partners.html. A listing of participating sites and a complete listing of the study investigators can be found at https://abcdstudy.org/consortium_members/. ABCD Consortium investigators designed and implemented the study and/or provided data but did not necessarily participate in the analysis or writing of this report.

Other acknowledgments specific to individual authors are provided in parentheses. Natural Sciences and Engineering Research Council of Canada Graduate Scholarships - Doctoral program (YP); funding by the Research Council of Norway (Grant No. 223273) and KG Jebsen Stiftelsen (to IA, OAA) and by the South-Eastern Norway Regional Health Authority (Grant Nos. 2019107, 2020086 [to DA]). The Neurofeedback study was partly funded by the project D8 of the Deutsche Forschungsgesellschaft collaborative research center 636. We would like to acknowledge Isabella Wolf and Regina Boecker-Schlier. (TB, SBa, DB, SH, NH, AK). The study cohort was financed by Conselho Nacional de Desenvolvimento Científico e Tecnológico (Grant Nos. 476529/2012-3, 466722/2014-1, 424041/2016-2; 310619/2019-0), the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior - Brasil (CAPES) - Finance Code 001 and FIPE-HCPA 160600, and Fundo de Amparo à Pesquisa do Rio Grande do Sul (Grant No. 19/2551-0001731-6 [to CB, EHG, FPic, MTa]). The study was funded by the National Institute of Mental Health (Grant No. K23MH090421), the National Alliance for Research on Schizophrenia and Depression, the University of Minnesota Graduate School, the Minnesota Medical Foundation, and the Biotechnology Research Center (Grant No. P41 RR008079 to the Center for Magnetic Resonance Research), University of Minnesota, and the Deborah E. Powell Center for Women's Health Seed Grant, University of Minnesota (to ZB, KC). The study cohort was financed by Conselho Nacional de Desenvolvimento Científico e Tecnológico (Grant Nos. 476529/2012-3, 466722/ 2014-1, and 424041/2016-2), CAPES - Finance Code 001 and FIPE-HCPA 160600, and the Fundação de Amparo à Pesquisa do Estado do Rio Grande do Sul (Grant Nos. PqG-19/2551-0001731-6 and PqG-19/2551-0001668-9 [CHDB]). This study was supported by the Swiss National Science Foundation (Grant No. #320030_130237 [to SBr]) and the Hartmann Müller Foundation (Grant No. 1460 [to SBr]). This study was supported by the Italian Ministry of Health (Grant No. RC 2768566/2020-2021 [to SCal, FM]); the Spanish Ministry of Science and Innovation (Grant No. Pl09/1588 Ito RCI) ISCIII and the Fondo Europeo de Desarrollo Regional (FEDER) (Grant No. 091510 from La Marató-TV3 Foundation [to RC]); and Health Research Board (HRA-POR-324 [to DMC, LN]). This work was supported by research grants from the National Healthcare Group, Singapore (Grant Nos. SIG/ 05004, SIG/05028, SIG /1103 [to KS]), and the Singapore Bioimaging Consortium (RP C009/2006 [to KS, QHC]), TENOVUS Dundee (to DC), the German Research Foundation (Grant Nos. FOR2107 DA1151/5-1 and DA1151/5-2: SFB-TRR58: Projects C09 and Z02 [to UD]) and the Interdisciplinary Center for Clinical Research (IZKF) of the medical faculty of Münster (Grant No. Dan3/012/17 [to UD]). Data collection and sharing for this project was partially funded by the Multimodal Treatment Study for (National Institutes of Drug Abuse Contract HHSN271200800009C [to JE, LT]). Support was provided by the German Federal Ministry of Education and Research (Bundesministerium fuer Bildung und Forschung Grant No. 01 ZX 1507, "PreNeSt - e:Med" [to TE-G]) and NIH MH083968 and Desert-Pacific Mental Illness Research, Education, and Clinical Center (to LTE). This work was supported by the Department of Veterans Affairs (Senior Research Career Scientist award and VA 1101 CX0004971 [to JMF]). CIBERSAM, AGAUR (to PF-C, EP-C, SS). The "Kids and Sibs" Study was supported by the Australian National Medical and Health Research Council (Program Grant No. 1037196 and Investigator Grant No. 1177991 [to PBM], Project Grant No. 1066177 [to JMF]), the Lansdowne Foundation, Good Talk, and the Keith Pettigrew Family Bequest (to PM). JMF gratefully acknowledges the Janette Mary O'Neil Research Fellowship (to JFu, PM, BO, GR). LFu was supported by an Australian Rotary Health/lan Parker Bipolar Research Fund postgraduate scholarship. Collection of the COGSBD cohort was funded by the Jack Brockhoff Foundation, University of Melbourne, Barbara Dicker Brain Sciences Foundation, Rebecca L Cooper Foundation and the Society of Mental

Health Research (to LFu). The Meath Foundation, Tallaght University Hospital, and The National Children's Hospital Foundation (to LG). This study was supported by Grant Nos. R01AA012207, R01MH123163, RO1NS114628, R01 EB015611, and U01 MH108148 (to SG, PK); NIH (Grant No. R37MH101495 [to IG]), the German Federal Ministry of Education and Research (Bundesministerium fuer Bildung und Forschung: Grant No. 01 ZX 1507, "PreNeSt - e:Med" [to RG-M]). The Study of Health in Pomerania (SHIP) is part of the Community Medicine Research net (CMR) (http://www. medizin.uni-greifswald.de/icm) of the University Medicine Greifswald, which is supported by the German Federal State of Mecklenburg- West Pomerania. MRI scans in SHIP and SHIP-TREND have been supported by a joint grant from Siemens Healthineers, Erlangen, Germany, and the Federal State of Mecklenburg-West Pomerania. This study was further supported by the EU-JPND Funding for BRIDGET (FKZ:01ED1615 [to HJG]), National Health and Medical Research Council of Australia (NHMRC) Project (Grant No. 630471 [to MG]), a Carnegie Developing Emerging Academic Leaders Fellowship (to NAG), and Stiftelsen KGJebsen (Grant No. SKGJ-MED02 [to JH]).

This work was supported by the German Research Foundation (DFG Grant Nos. HA7070/2-2, HA7070/3, HA7070/4 [to TH]) and IZKF of the medical faculty of Münster (Grants No. Dan3/012/17 [to UD] and MzH 3/020/ 20 [to TH]), and NHMRC projects (Grant No. 1064643 [to BJH] and 1024570 [to CGD]). The CIAM study was supported by the University of Cape Town Research Committee, South African National Research Foundation, and the South African Medical Research Council (principal investigator [PI], Fleur M. Howells) and Grant No. R01MH117601 (to NJ). This work was funded by the German Research Foundation (Grant Nos. FOR2107 JA 1890/7-1 and FOR2107 JA 1890/7-2 [to AJ]) and Swinburne University scholarship/ Australian Postgraduate Award (to JK). Collection of the COGSBD cohort was funded by the Jack Brockhoff Foundation, University of Melbourne, Barbara Dicker Brain Sciences Foundation, Rebecca L Cooper Foundation, and the Society of Mental Health Research (to JK). This work was funded by the German Research Foundation (Grant Nos. FOR2107 KI588/14-1 and FOR2107 KI588/14-2 [to TK]). The St. Göran study was supported by grants from the Swedish Research Council (Grant No. 2018-02653 [to ML]), the Swedish foundation for Strategic Research (Grant No. KF10-0039 [to ML]), the Swedish Brain foundation (Grant No. FO2020-0261 [to ML]), and the Swedish Government under the LUA/ALF agreement (Grant Nos. ALF 20170019 and ALFGBG-716801 [to ML]). This work was supported by RFBR (Grant No. 20-013-00748 [to IL, AST]) and funded by the Health Research Board (Grant No. HRA_POR/2011/100 [to CM]). The Australian Schizophrenia Research Bank (ASRB) was supported by the NHMRC (Enabling Grant No. 386500), the Pratt Foundation, Ramsay Health Care, the Viertel Charitable Foundation, and the Schizophrenia Research Institute. Chief investigators for ASRB were VC, US, RSc, AJ, BM, PTM, SVC, FH, and CPa. This work was supported by Deutsche Forschungsgemeinschaft (Grant Nos. DFG NE 2254/2-1, NE 2254/3-1, NE2254/4-1 [to IN]), the University Research Priority Program "Integrative Human Physiology" at the University of Zurich (to ROT), an NHMRC Senior Principal Research Fellowship (Grant No. 1105825 [to CPa]), an NHMRC L3 Investigator Grant (Grant No. 1196508 [to CPa]), and NHMRC Program Grant (Grant No. 1150083 [to CPa]). ASRB was supported by the NHMRC (Enabling Grant No. 386500), the Pratt Foundation, Ramsay Health Care, the Viertel Charitable Foundation and the Schizophrenia Research Institute. Chief investigators for ASRB were VC, US, RSc, AJ, BM, PTM, SVC, FH, and CPa. This project has received funding from the European Union's Horizon 2020 research and innovation programme under the Marie Skłodowska-Curie grant (Agreement No. 798181 [to GP]). ASRB was supported by the NHMRC (Enabling Grant, ID 386500), the Pratt Foundation, Ramsay Health Care, the Viertel Charitable Foundation and the Schizophrenia Research Institute. Chief investigators for ASRB were VC, US, RSc, AJ, BM, PTM, SVC, FH, and CPa. The Imaging Genetics in Psychosis study was funded by Project Grants from the NHMRC (Grant Nos. APP630471 and APP1081603 [to YQ]) and the Macquarie University's Australian Research Council (ARC) Centre of Excellence in Cognition and its Disorders (Grant No. CE110001021 [to YQ]). This work was supported by the Spanish Ministry of Science, Innovation and Universities/ Economy and Competitiveness/Instituto de Salud Carlos III (Grant Nos. PI11/01766 and CPII19/00009 [to JR]), co-financed by European Regional Development Fund funds from the European Commission ("A Way of

Making Europe"). ASRB was supported by the NHMRC (Enabling Grant No. 386500), the Pratt Foundation, Ramsay Health Care, the Viertel Charitable Foundation, and the Schizophrenia Research Institute. Chief investigators for ASRB were VC, US, RSc, AJ, BM, PTM, SVC, FH, and CPa. SR was supported by an NHMRC Senior Fellowship (Grant No. GNT1154651). Collection of the COGSBD cohort was funded by the Jack Brockhoff Foundation, University of Melbourne, Barbara Dicker Brain Sciences Foundation, Rebecca L Cooper Foundation, and the Society of Mental Health Research (to SR). This work was supported by NIHR; MRC (to KR), NIH (Grant Nos. MH-094268, MH-105660, and MH-107730 [to ASS]). The Neuroimaging of the Children's Attention Project cohort was funded by NHMRC, Australia (Grant No. 1065895). Earlier funding for the cohort as also provided by NHMRC (Grant No. 1008522) and a grant from the Collier Foundation. The ACPU cohort was funded by NHMRC, Australia (Project Grant Nos. 384419 and 569533 [to TS]). This work was supported by research grants from the National Healthcare Group, Singapore (Grant Nos. SIG/05004, SIG/05028, and SIG /1103), and the Singapore Bioimaging Consortium (RP C009/2006 [to KS]), and the Ministry of Health, Czech Republic - conceptual development of research organization ("Institute for Clinical and Experimental Medicine - IKEM, IN 00023001" [to ASk]). This study was supported by the Italian Ministry of Health (Grant No. RC/17-18-19-20-21/A [to GS]) and Ministry of Health of the Czech Republic (Grant No. NU20-04-00393 [to FS]). The Drakenstein Child Health Study (DCHS) cohort is funded by the Bill and Melinda Gates Foundation (Grant No. OPP 1017641) and the South African Medical Research Council. This DCHS contribution was made possible in part by a grant from Carnegie Corporation of New York. The statements made and views expressed are solely the responsibility of the author (DJS). STRADL study was supported and funded by the Wellcome Trust Strategic Award "Stratifying Resilience and Depression Longitudinally" (Grant No. 104036/Z/14/Z), and the Medical Research Council Mental Health Pathfinder Award "Leveraging routinely collected and linked research data to study the causes and consequences of common mental disorders" (MRC, Grant No. MC_PC_17209). Scottish Bipolar Family Study (SBFS) was supported by National Health Service Research Scotland, through the Scottish Mental Health Research Network (www.smhrn.org.uk), who provided assistance with subject recruitment and assessments. SBFS was conducted at the Brain Research Imaging Centre (http://www.bric.ed.ac.uk), which is supported by SINAPSE (Scottish Imaging Network, a Platform for Scientific Excellence, http://www.sinapse.ac. uk). Processing of the datasets used the resources provided by the Edinburgh Compute and Data Facility (http://www.ecdf.ed.ac.uk/) (ASt). TVR was supported by an NHMRC Early Career Fellowship (Grant No. GNT1088785). Collection of the COGSBD cohort was funded by the Jack Brockhoff Foundation, University of Melbourne, Barbara Dicker Brain Sciences Foundation, Rebecca L Cooper Foundation, and the Society of Mental Health Research (to TVR). EV was supported by the Spanish Ministry of Science and Innovation (PI18/00805) integrated into the Plan Nacional de I+D+I and co-financed by the ISCIII-Subdirección General de Evaluación and the FEDER; the Instituto de Salud Carlos III; the CIBERSAM (Centro de Investigación Biomédica en Red de Salud Mental): by the CERCA Programme/Generalitat de Catalunya and the Secretaria d'Universitats i Recerca del Departament d'Economia I Coneixement (Grant No. 2017SGR1355). This study was also supported by the Departament de Salut de la Generalitat de Catalunya, Pla Estratègic de Recerca i Innovació en Salut (PERIS) 2016-2020 (Grant No. SLT006/17/00345) and the European Union Horizon 2020 research and innovation program (EU.3.1.1. Understanding health, wellbeing and disease: Grant Nos. 754907 and EU.3.1.3 [to EB]; Treating and managing disease: Grant No. 945151 [to EV]). This study was supported by the National Center for Complementary and Integrative Health (Grant Nos. R21AT009173 and R61AT009864 [to TTY]); by the National Center for Advancing Translational Sciences (CTSI), National Institutes of Health, through UCSF-CTSI (Grant No. UL1TR001872 [to TTY]); by the American Foundation for Suicide Prevention (Grant No. SRG-1-141-18 [to TTY]); by UCSF Research Evaluation and Allocation Committee (REAC) and J. Jacobson Fund (to TTY); by the NIMH (Grant No. R01MH085734 [to TTY]); and by the Brain and Behavior Research Foundation (formerly NARSAD) (to TTY).

This work was supported by a personal Veni grant to MH from the Netherlands Organization for Scientific Research (NWO, Grant No.

91619115 [to MH]) and European Community's Horizon 2020 Programme (H2020/2014 - 2020) (Grant Agreements Nos. 667302 [CoCA], 728018 [Eat2beNICE], and 847879 [PRIME] [to BF]). JBu has been supported by the EU-AIMS (European Autism Interventions) and AIMS-2-TRIALS programmes, which receive support from Innovative Medicines Initiative Joint Undertaking Grant Nos. 115300 and 777394, the resources of which are composed of financial contributions from the European Union's FP7 and Horizon 2020 Programmes, and from the European Federation of Pharmaceutical Industries and Associations companies' in-kind contributions. and AUTISM SPEAKS, Autistica and SFARI; and by the Horizon 2020-supported programme CANDY (Grant No. 847818 [to JBu]). This work is supported by Grant No. NIA T32AG058507 and NIH Grant No. U54EB020403 from the Big Data to Knowledge (BD2K) Program (to CRKC). ENIGMA MDD work is supported by NIH (Grant Nos. U54 EB020403 [to PT], R01 MH116147 [to PT], and R01 MH117601 [to NJ and LS]). LS was supported by an NHMRC Career Development Fellowship (Grant No. 1140764). This work was supported by the National Center for Research Resources at the NIH (Grant Nos. NIH 1 U24 RR021992 [Function Biomedical Informatics Research Network], NIH 1 U24 RR025736-01 [Biomedical Informatics Research Network Coordinating Center; http://www.birncommunity.org]). TGMvE is supported by ENIGMA's NIH BD2K initiative (Grant No. U54 EB020403), ENIGMA Sex Differences (Grant No. R01MH116147), and ENIGMA-COINSTAC: Advanced Worldwide Transdiagnostic Analysis of Valence System Brain Circuits (Grant No. R01MH121246). This work was supported by the NIH (Grant No. R01MH121246 [to JT, Calhoun, and TGMvE]). This work was supported in part by NIH (Grant No. U54 EB020403 [to PT]).

This article reflects the views of the authors and may not reflect the opinions or views of the NIH or ABCD Consortium investigators.

Investigation (MRI data specific): YP, JSa, CAb, IA, CAr, DA, SA, LAA, CAI, VA, GA, RA-A, NB, TB, CB, ZB, RB, CHDB, JBa, SBa, FB, AB, CdMB, DB, SBr, JBr, RBC, JRB, SCal, RC, EJC-R, DC, SCar, VJC, SVC, SCh, QHC, DC, CGC, AC, ARC, BC-F, KC, UD, CGD, CD, CMD-C, KD, SE, JE, TE-G, LTE, JFe, JFi, WF, JMF, LFu, PF-C, JFu, LFo, LG, BG, SG, JMG, IG, RG-M, HJG, MG, EHG, NAG, DG, OG, JH, TH, BJH, WH, FH, DJH, EH, PJH, SH, NH, FMH, JCI, NJ, BJ, AJ, JJ, RJ, AK, VK, JK, JAK, TK, PK, S-MK, ML, NL, SL, IL, BL, AJL, FPM, LAM, DHM, CM, AM-A, SM, PTM, PM, AM, BM, FM, LN, IN, RO, JO, BO, CPa, MP, JCP, PP, GP, FMP, FPic, FPir, EP-C, CPr, YQ, JR, JAR-Q, PER, ARei, ARet, GR, SR, DLR, KR, MDS, JSh, RSa, SS, ASa, US, RSc, PS, TS, KS, ASk, GS, FS, DJS, OS, ASt, YT, LT, MTa, AT, KT, SIT, DT, AST, DT-G, MTo, AU, TV, JV-B, MWV, EV, OV, CW, TW, LTW, HW, DW, AW, KW, TTY, YY, JLZ, MH, BF, DvR, JBu, CRKC, OAA, EP, LS, TGMvE, JT, FXC, ZP, PT, TP. Visualization: YP, JSh. Funding Acquisition: CAb, IA, CAI, DA, SA, LAA, CAr, VA, GA, RA, NB, TB, CB, ZB, RB, CHDB, JBa, SBa, FB, AB, CdMB, DB, SBr, JBr, RB, JRB, SCal, RC, EJC-R, DC, SCar, VJC, SVC, SCh, QHC, DC, CGC, AC, ARC, BC-F, KC, UD, CGD, CD, CMD-C, KD, SE, JE, TE-G, LTE, JFe, JFi, WF, JMF, LFu, PF-C, JFu, LFo, LG, BG, SG, JMG, IG, RG-M, HJG, MG, EHG, NAG, DG, OG, JH, TH, BJH, WH, FH, DJH, EH, PJH, SH, NH, FMH, JCI, NJ, BJ. AJ. JJ. RJ. AK. VK. JK. JAK. TK. PK. S-MK. ML. NL. SL. IL. BL. AJL. FPM. LAM, DHM, CM, AM-A, SM, PTM, PM, AM, BM, FM, LN, IN, RO, JO, BO, CPa, MP, JCP, PP, GP, FMP, FPic, FPir, EP-C, CPr, YQ, JR, JAR-Q, PER, ARei, ARet, GR, SR, DLR, KR, MDS, JSa, RSa, SS, ASa, US, RSc, PS, TS, KS, ASk, GS, FS, DJS, OS, ASt, YT, LT, MTa, AT, KT, SIT, DT, AST, DT-G, MTo, AU, TV, JV-B, MWV, EV, OV, CW, TW, LTW, HW, DW, AW, KW, TTY, YY, JLZ, MH, BF, DvR, JBu, CRKC, OAA, EP, LS, TGMvE, JT, FXC, ZP, PT, TP. Supervision: TP, ZP. Writing - original draft: YP, ZP, TP. Writing - review and editing: YP, JSh, CAb, IA, CAI, DA, SA, LAA, CAr, VA, GA, RA, NB, TB, CB, ZB, RB, CHDB, JBa, SBa, FB, AB, CdMB, DB, SBr, JBr, RB, JRB, SCal, RC, EJC-R, DC, SCar, VJC, SVC, SCh, QHC, DC, CGC, AC, ARC, BC-F, KC, UD, CGD, CD, CMD-C, KD, SE, JE, TE-G, LTE, JFe, JFe, WF, JMF, LFu, PF-C, JFu, LFo, LG, BG, SG, JMG, IG, RG-M, HJG, MG, EHG, NAG, DG, OG, JH, TH, BJH, WH, FH, DJH, EH, PJH, SH, NH, FMH, JCI, NJ, BJ, AJ, JJ, RJ, AK, VK, JK, JAK, TK, PK, S-MK, ML, NL, SL, IL, BL, AJL, FPM, LAM, DHM, CM, AM-A, SM, PTM, PM, AM, BM, FM, LN, IN, RO, JO, BO, CPa, MP, JCP, PP, GP, FMP, FPic, FPir, EP-C, CPr, YQ, JR, JAR-Q, PER, ARei, ARet, GR, SR, DLR, KR, MDS, JSa, RSa, SS, ASa, US, RSc, PS, TS, KS, ASk, GS, FS, DJS, OS, ASt, YT, LT, MTa, AT, KT, SIT, DT, AST, DT-G, MTo, AU, TV, JV-B, MWV, EV, OV, CW, TW, LTW, HW, DW, AW, KW, TTY, YY, JLZ, MH, BF, DvR, JBu, CRKC, OAA, EP, LS, TGMvE, JT, FXC, ZP, PT, TP. Conceptualization: YP, JSh, ZP, TP. Methodology: YP, JSh, ZP, TP.

VA, HJB, KD, DG, and WH express gratitude to all participants of the Münster Neuroimaging Study. EV and RC acknowledge the support of the Spanish Ministry of Science and Innovation and FEDER. PTM and CPa thank C. Loughland, the ASRB Manager, and acknowledges the help of Jason Bridge for ASRB database queries.

CAr has been a consultant to or has received honoraria or grants from Acadia, Angelini, Boehringer, Gedeon Richter, Janssen-Cilag, Lundbeck, Minerva, Otsuka, Roche, Sage, Servier, Shire, Ethiopia, Schering Plough, Sumitomo Dainippon Pharma, Sunovion, and Takeda. TB served in an advisory or consultancy role for ADHS digital, Infectopharm, Lundbeck, Medice, Neurim Pharmaceuticals, Oberberg GmbH, Roche, and Takeda; received conference support or speaker's fees from Medice and Takeda; and received royalties from Hogrefe, Kohlhammer, CIP Medien, Oxford University Press. This work is unrelated to these relationships. DB serves as an unpaid scientific consultant for an EU-funded neurofeedback trial. DC served on the advisory board or speaker for Takeda, Medice, Novartis, and Sunovian, unrelated to this work. HJG has received travel grants and speakers honoraria from Fresenius Medical Care, Neuraxpharm, Servier, and Janssen-Cilag as well as research funding from Fresenius Medical Care. EHG has received lecture honoraria from Takeda and Sandoz, NJ is MPI of a research grant from Biogen Inc. for work unrelated to the contents of this article. ML has received lecture honoraria from Lundbeck. DHM serves as a consultant for Boehringer Ingelheim, Cadent Therapeutics, Syndisi, and Recognify. CPa has received honoraria for talks at educational meetings and has served on an advisory board for Lundbeck, Australia Ptv Ltd. KR reports a grant from Takeda Pharmaceuticals and consultancy fees from Lundbeck. EV has received grants and served as consultant, advisor, or CME speaker for the following entities (unrelated to the present work): AB-Biotics, AbbVie, Aimentia, Angelini, Celon, Dainippon Sumitomo Pharma, Ferrer, Gedeon Richter, GH, (Research), GlaxoSmithKline, Janssen, Lundbeck, Organon, Otsuka, Sage, Sanofi-Aventis, Sunovion, and Takeda. BF has received educational speaking fees from Medice GmbH. JBu has been in the past 3 years a consultant to, member of advisory board of, and/or speaker for Takeda/Shire, Roche, Medice, Angelini, Janssen, and Servier. He is not an employee of any of these companies and not a stock shareholder of any of these companies. He has no other financial or material support, including expert testimony, patents, and royalties. CRKC has received partial research support from Biogen, Inc for work unrelated to the topic of this article. OAA has served as a consultant to HealthLytix and received speaker's honoraria from Ludbeck and Sunovion. FXC has served in the past on a BOL Pharma scientific advisory board, unrelated to this work. PT received partial funding support from Biogen, Inc for research unrelated to this article. All other authors report no biomedical financial interests or potential conflicts of interest

ARTICLE INFORMATION

From the Institute of Medical Sciences (YP, TP); The Hospital for Sick Children and Departments of Physiology and Nutritional Sciences (JSh, ZP), University of Toronto, Toronto, Ontario; Department of Psychiatry (SCh) and Departments of Psychiatry and Pediatrics (FPM), University of Calgary, Calgary, Alberta; Departments of Psychiatry and Neuroscience (TP), Faculty of Medicine and Centre Hospitalier Universitaire Sainte-Justine. University of Montréal, Montreal, Quebec, Canada: Department of Clinical Neuroscience (CAb), Karolinska Institutet, Stockholm; Institute of Neuroscience and Physiology (ML), University of Gothenburg, Gothenburg, Sweden; NORMENT Centre (IA, DA, LTW, OAA) and Norwegian Centre for Mental Disorders Research NORMENT (EH), Institute of Clinical Medicine; Department of Psychology (AD, NIL); Department for Data Capture and Collections Management (LAM), University Center for Information Technology, University of Oslo; Division of Mental Health and Addiction (IA, DA, LTW, OAA), Oslo University Hospital; Faculty of Health Sciences (RJ), Oslo Metropolitan University, Oslo; Department of Biological and Medical Psychology (ARC, AJL) and Department of Biomedicine (JH), University of Bergen, Bergen, Norway; Department of Child and Adolescent Psychiatry (CAI, CAr, JJ), Institute of Psychiatry and Mental Health, Hospital General Universitario Gregorio Marañón, Instituto de Investigación Sanitaria del Hospital Gregorio Marañón, Centro de Investigación Biomédica en Red de Salud Mental (CIBERSAM); School of Medicine (CAr, CMD-C, MP),

Universidad Complutense; Instituto de Investigación Sanitaria Gregorio Marañón (SCar), Madrid; Department of Psychiatry (RA-A, JV-B), Marques de Valdecilla University Hospital, Instituto de Investigación Valdecilla, CIBERSAM, School of Medicine, University of Cantabria; Department of Radiology (DT-G), University Hospital Marqués de Valdecilla, Instituto de Investigación Valdecilla, Santander; Institut d'Investigacions Biomèdiques August Pi i Sunyer (CdMB, LFo, JMG, JR), Centro de Investigación Biomédica en Red de Salud Mental (CIBERSAM); Institute of Neuroscience (RC, EV), Hospital Clinic, University of Barcelona, Institut d'Investigacions Biomèdiques August Pi i Sunyer, CIBERSAM; FIDMAG Research Foundation (EJC-R, PF-C, EP-C, RSa, SS); Magnetic Resonance Imaging core facility (JCP), Institut d'Investigacions Biomèdiques August Pi i Sunyer; FIDMAG Germanes Hospitalàries Research Foundation (AM-A, JSa), Biomedical Network Research Centre on Mental Health (CIBERSAM); Department of Psychiatry and Forensic Medicine (CPr), Universitat Autònoma de Barcelona; Department of Psychiatry (JAR-Q), Hospital Universitari Vall d'Hebrón, CIBERSAM, Universitat Autònoma de Barce-Iona, Barcelona; Department of Psychiatry (BC-F), Virgen del Rocio University Hospital, Universidad de Sevilla, Instituto de Biomedicina de Sevilla, CIBERSAM, Sevilla; Department of Psychiatry (OV), Autonomous University of Barcelona, Cerdanyola del Valles, Spain; Laboratory of Neuropsychiatry (SA, NB, FPir, GS), Santa Lucia Foundation Scientific Institute for Research, Hospitalization and Healthcare, Rome; Departments of Education Science, Psychology, Communication Science (LAA) and Basic Medical Sciences, Neuroscience and Sense Organs (AB, GP, FMP), University of Bari Aldo Moro; Azienda Ospedaliero-Universitaria Consorziale Policlinico di Bari (PS). Bari: Department of Developmental Neuroscience (SCal. FM) and Laboratory of Medical Physics and Magnetic Resonance (MTo), Scientific Institute for Research, Hospitalization and Healthcare Stella Maris Foundation; National Institute for Nuclear Physics (ARet), Pisa Division, Pisa, Italy; Institute for Translational Psychiatry (VA, UD, KD, DG, TH, SM, KT, AW) and Department of Clinical Radiology (JBa, WH), University of Münster, Münster; Department of Child and Adolescent Psychiatry and Psychotherapy (TB, SBa, DB, SH, NH, AK), Central Institute of Mental Health, Medical Faculty Mannheim, Heidelberg University, Mannheim; Translational Developmental Neuroscience Section (FB, SE, JAK), Division of Psychological and Social Medicine and Developmental Neurosciences, Faculty of Medicine, and Department of Child and Adolescent Psychiatry (AU), Faculty of Medicine, TU Dresden, Dresden; Institute of Diagnostic Radiology and Neuroradiology (RB), Department of Psychiatry and Psychotherapy (HJG), and Institute for Community Medicine (AT), University Medicine Greifswald; German Center for Neurodegenerative Diseases (KW), Site Rostock/Greifswald, Greifswald; Department of Child and Adolescent Psychiatry, Psychosomatics and Psychotherapy (AC), University of Tübingen, Tübingen; Laboratory of Systems Neuroscience and Imaging in Psychiatry (TE-G, RG-M), Department of Psychiatry and Psychotherapy, University Medical Center Göttingen, Göttingen; Section for Experimental Psychopathology and Neuroimaging (OG), Department of General Psychiatry, Heidelberg University Hospital, Heidelberg; Core Facility Brain imaging (AJ), Faculty of Medicine, University of Marburg; Department of Psychiatry (TK), Marburg University; Department of Psychiatry and Psychotherapy (IN, OS), Philipps-Universität Marburg, Marburg; Department of Psychology (Biological Psychology, Clinical Psychology, and Psychotherapy) (PP), University of Würzburg, Würzburg; Department of Psychiatry, Psychosomatic Medicine and Psychotherapy (ARei), University Hospital Frankfurt-Goethe University, Frankfurt am Main, Germany; National Centre for Scientific Research (GA, CD), Aix-Marseille University, Marseille, France; Department of Genetics (CB, CHDB, MTa), Instituto de Biociências; Department of Psychiatry (EHG), Faculty of Medicine; Graduate Program in Psychiatry and Behavioral Sciences (FPic), Faculty of Medicine, Universidade Federal do Rio Grande do Sul, Porto Alegre; Department of Physiology and Biophysics (DLR), Instituto de Ciencias Biomédicas Universidade de São Paulo, São Paulo, Brazil; Department of Psychiatry and Behavioral Sciences (ZB, KC), University of Minnesota Medical School, Minneapolis, Minnesota; Department of Psychiatry (RBC), University of Vermont, Burlington, Vermont; Department of Psychiatry (JRB), University of New Mexico, Albuquerque, New Mexico; Department of Biomedical Sciences (CGC), Florida State University, Tallahassee, Florida;

Cincinnati Children's Hospital Medical Center (JE, LT), Cincinnati, Ohio; Department of Psychiatry (JFe, WF, BL), University of Pittsburgh, Pittsburgh, Pennsylvania; Department of Psychiatry (LTE), University of California San Diego, San Diego; San Francisco Veterans Affairs Medical Center (JMF); Department of Psychiatry and Behavioral Sciences (DHM); Department of Psychiatry and Behavioral Sciences (TTY), Division of Child and Adolescent Psychiatry, University of California San Francisco, San Francisco; Clinical Translational Neuroscience Laboratory (BG, TGMvE), Department of Psychiatry and Human Behavior, University of California Irvine, Irvine; Department of Psychology (IG), Stanford University, Stanford; USC Mark and Mary Stevens Neuroimaging and Informatics Institute (NJ); Imaging Genetics Center (SIT, PT, CRKC), USC Mark and Mary Stevens Neuroimaging & Informatics Institute, University of Southern California, Marina del Rey, California; Maryland Psychiatric Research Center (SG, PK), Department of Psychiatry, University of Maryland School of Medicine; Johns Hopkins University School of Medicine (ASa), Baltimore, Maryland; Center for Depression, Anxiety, and Stress Research (MDS), McLean Hospital, Harvard Medical School, Belmont, Massachusetts; SUNY Upstate Medical University (TW), Syracuse; NYU Grossman School of Medicine (YY, FXC), New York, New York: Georgia State University (JT), Atlanta. Georgia; Department of Child and Adolescent Psychiatry and Psychotherapy (SBr), University Hospital of Psychiatry; Department of Child and Adolescent Psychiatry and Psychotherapy (DW), Psychiatric University Hospital, University of Zürich; University Children's Hospital Zürich (ROT), Zurich, Switzerland; School of Psychiatry (JBr, VJC, MG, PM, YQ, GR) and Neuroscience Research Australia (JFu, BO); Department of Neuroscience and Physiology (CW), University of New South Wales, Sydney; School of Medicine & Public Health (FH); School of Psychology (PTM), College of Engineering, Science and Environment; Priority Centre for Brain & Mental Health Research (PER, US), The University of Newcastle, Callaghan; Hunter Medical Research Institute (RSc), Newcastle, New South Wales; School of Medicine (SVC) and Queensland Brain Institute (BM), The University of Queensland, Brisbane, Queensland; Departments of Paediatrics (DC) and Psychiatry (DC, CGD); Melbourne Neuropsychiatry Centre (LFu, JK), Department of Psychiatry, University of Melbourne, Parkville; Melbourne Neuropsychiatry Centre (TVR), Department of Psychiatry, and Centre for Youth Mental Health (EP, LS), University of Melbourne, Melbourne; Melbourne Neuropsychiatry Centre (BJH), Department of Psychiatry, University of Melbourne, Melbourne; Melbourne Neuropsychiatry Centre (CPa), Department of Psychiatry, University of Melbourne, Carlton South, Victoria, Australia; Centre for Mental Health (SR), School of Health Sciences, Swinburne University, Melbourne, Victoria, Australia; School of Psychology (TS), Deakin University, Geelong, Victoria, Australia; Clinical Neuroimaging Lab (DMC, LN) and Galway Neuroscience Centre (CM), Center for Neuroimaging, Cognition and Genomics, Galway Neuroscience Centre, College of Medicine, Nursing, and Health Sciences, National University of Ireland Galway, Galway; Trinity Institute of Neuroscience (JFi) and Discipline of Psychiatry (LG), School of Medicine, Trinity College Dublin, Dublin, Ireland: Research Division (QHC) and West Region (KS), Institute of Mental Health, Singapore, Singapore: Department of Psychiatry & Mental Health (NAG. JCI, S-MK, DJS), Neuroscience Institute and the Department of Psychiatry and Mental Health (FMH), University of Cape Town, Cape Town, South Africa; Experimental and Clinical Psychology (DJH) and Clinical Neuropsychology Section (JO), Vrije Universiteit Amsterdam; Department of Psychiatry (DV), Amsterdam UMC, VUMC, Amsterdam; Department of Child and Adolescent Psychiatry (PJH), University of Groningen, University Medical Center Groningen, Groningen; Department of Human Genetics (BJ), Departments of Human Genetics and Psychiatry (BF), and Department of Cognitive Neuroscience (DvR); Department of Human Genetics (MH) and Department of Cognitive Neuroscience (JBu), Donders Institute for Brain, Cognition and Behavior, Radboud University Medical Center, Nijmegen; Department of Radiology & Nuclear Medicine (MWV) and Department of Epidemiology (JLZ), Erasmus MC University Medical Center Rotterdam, Rotterdam, The Netherlands; Mental Health Research Center (VK, IL, AST), Moscow, Russia; Division of Psychiatry (SL, AM, ASt, HW), Centre for Clinical Brain Sciences, University of Edinburgh, Edinburgh; Child & Adolescent Psychiatry (KR), King's College London, London, United Kingdom; National Institute of Mental Health (ASk, FS, DT), Klecany, Czech Republic; and the Department of Neuropsychiatry (YT), Graduate School of Medicine and Pharmaceutical Sciences, University of Toyama, Toyama, Japan.

Address correspondence to Tomas Paus, M.D., Ph.D., at tpausresearch@gmail.com.

Received Oct 19, 2021; revised Feb 2, 2022; accepted Feb 23, 2022. Supplementary material cited in this article is available online at https://doi.org/10.1016/j.biopsych.2022.02.959.

REFERENCES

- Azevedo FAC, Carvalho LRB, Grinberg LT, Farfel JM, Ferretti REL, Leite REP, et al. (2009): Equal numbers of neuronal and nonneuronal cells make the human brain an isometrically scaled-up primate brain. J Comp Neurol 513:532–541.
- Rakic P (1988): Specification of cerebral cortical areas. Science 241:170–176.
- Gilmore JH, Knickmeyer RC, Gao W (2018): Imaging structural and functional brain development in early childhood. Nat Rev Neurosci 19:123–137.
- Storsve AB, Fjell AM, Tamnes CK, Westlye LT, Overbye K, Aasland HW, Walhovd KB (2014): Differential longitudinal changes in cortical thickness, surface area and volume across the adult life span: Regions of accelerating and decelerating change. J Neurosci 34:8488–8498.
- Duerden EG, Chakravarty MM, Lerch JP, Taylor MJ (2020): Sex-based differences in cortical and subcortical development in 436 individuals aged 4–54 years. Cereb Cortex 30:2854–2866.
- Brown TT, Kuperman JM, Chung Y, Erhart M, McCabe C, Hagler DJ Jr, et al. (2012): Neuroanatomical assessment of biological maturity. Curr Biol 22:1693–1698.
- Kapellou O, Counsell SJ, Kennea N, Dyet L, Saeed N, Stark J, et al. (2006): Abnormal cortical development after premature birth shown by altered allometric scaling of brain growth. PLoS Med 3:e265.
- Li G, Nie J, Wang L, Shi F, Lin W, Gilmore JH, Shen D (2013): Mapping region-specific longitudinal cortical surface expansion from birth to 2 years of age. Cereb Cortex 23:2724–2733.
- Tamnes CK, Herting MM, Goddings AL, Meuwese R, Blakemore SJ, Dahl RE, et al. (2017): Development of the cerebral cortex across adolescence: A multisample study of inter-related longitudinal changes in cortical volume, surface area, and thickness. J Neurosci 37:3402–3412.
- Wierenga LM, Langen M, Oranje B, Durston S (2014): Unique developmental trajectories of cortical thickness and surface area. Neuroimage 87:120–126.
- Raznahan A, Greenstein D, Lee NR, Clasen LS, Giedd JN (2012): Prenatal growth in humans and postnatal brain maturation into late adolescence. Proc Natl Acad Sci U S A 109:11366–11371.
- Walhovd KB, Fjell AM, Brown TT, Kuperman JM, Chung Y, Hagler DJ Jr, et al. (2012): Long-term influence of normal variation in neonatal characteristics on human brain development. Proc Natl Acad Sci U S A 109:20089–20094.
- Liang D, Elwell AL, Aygün N, Krupa O, Wolter JM, Kyere FA, et al. (2021): Cell-type-specific effects of genetic variation on chromatin accessibility during human neuronal differentiation. Nat Neurosci 24:941–953.
- Grasby KL, Jahanshad N, Painter JN, Colodro-Conde L, Bralten J, Hibar DP, et al. (2020): The genetic architecture of the human cerebral cortex [published correction appears in Science 2021; 374: eabm7211]. Science 367:eaay6690.
- Cross-Disorder Group of the Psychiatric Genomics Consortium (2019): Genomic relationships, novel loci, and pleiotropic mechanisms across eight psychiatric disorders. Cell 179:1469–1482.e11.
- Abel KM, Wicks S, Susser ES, Dalman C, Pedersen MG, Mortensen PB, Webb RT (2010): Birth weight, schizophrenia, and adult mental disorder: Is risk confined to the smallest babies? Arch Gen Psychiatry 67:923–930.
- Schork AJ, Won H, Appadurai V, Nudel R, Gandal M, Delaneau O, et al. (2019): A genome-wide association study of shared risk across

- psychiatric disorders implicates gene regulation during fetal neuro-development. Nat Neurosci 22:353–361.
- Thompson PM, Jahanshad N, Ching CRK, Salminen LE, Thomopoulos SI, Bright J, et al. (2020): ENIGMA and global neuroscience: A decade of large-scale studies of the brain in health and disease across more than 40 countries. Transl Psychiatry 10:100.
- Hoogman M, Muetzel R, Guimaraes JP, Shumskaya E, Mennes M, Zwiers MP, et al. (2019): Brain imaging of the cortex in ADHD: A coordinated analysis of large-scale clinical and population-based samples. Am J Psychiatry 176:531–542.
- van Rooij D, Anagnostou E, Arango C, Auzias G, Behrmann M, Busatto GF, et al. (2018): Cortical and subcortical brain morphometry differences between patients with autism spectrum disorder and healthy individuals across the lifespan: Results from the ENIGMA ASD Working Group. Am J Psychiatry 175:359–369.
- Hibar DP, Westlye LT, Doan NT, Jahanshad N, Cheung JW, Ching CRK, et al. (2018): Cortical abnormalities in bipolar disorder: An MRI analysis of 6503 individuals from the ENIGMA bipolar disorder Working Group. Mol Psychiatry 23:932–942.
- van Erp TGM, Walton E, Hibar DP, Schmaal L, Jiang W, Glahn DC, et al. (2018): Cortical brain abnormalities in 4474 individuals with schizophrenia and 5098 control subjects via the Enhancing Neuro Imaging Genetics through Meta Analysis (ENIGMA) Consortium. Biol Psychiatry 84:644–654.
- Schmaal L, Hibar DP, Sämann PG, Hall GB, Baune BT, Jahanshad N, et al. (2017): Cortical abnormalities in adults and adolescents with major depression based on brain scans from 20 cohorts worldwide in the ENIGMA Major Depressive Disorder Working Group. Mol Psychiatry 22:900–909.
- Viechtbauer W (2010): Conducting meta-analyses in R with the metafor package. J Stat Softw 36:1–48.
- Garavan H, Bartsch H, Conway K, Decastro A, Goldstein RZ, Heeringa S, et al. (2018): Recruiting the ABCD sample: Design considerations and procedures. Dev Cogn Neurosci 32:16–22.
- Ponce M, van Zon R, Northrup S, Gruner D, Chen J, Ertinaz F, et al. (2019): Deploying a top-100 supercomputer for large parallel work-loads: The niagara supercomputer. In: Presented at PEARC: Practice and Experience in Advanced Research Computing, July 28–August 1, Chicago. Illinois.
- Casey BJ, Cannonier T, Conley MI, Cohen AO, Barch DM, Heitzeg MM, et al. (2018): The Adolescent Brain Cognitive Development (ABCD) study: Imaging acquisition across 21 sites. Dev Cogn Neurosci 32:43–54.
- 28. Fried EI, Greene AL, Eaton NR (2021): The p factor is the sum of its parts, for now. World Psychiatry 20:69–70.
- Bates D, Sarkar D, Bates MD (2007): The Ime4 Package. R Package Version 2. Available at: https://github.com/Ime4/Ime4/. Accessed December 1, 2020.
- Bhaduri A, Andrews MG, Mancia Leon W, Jung D, Shin D, Allen D, et al. (2020): Cell stress in cortical organoids impairs molecular subtype specification. Nature 578:142–148.
- Skene NG, Grant SGN (2016): Identification of vulnerable cell types in major brain disorders using single cell transcriptomes and expression weighted cell type enrichment. Front Neurosci 10:16.
- Bryois J, Skene NG, Hansen TF, Kogelman LJA, Watson HJ, Liu Z, et al. (2020): Genetic identification of cell types underlying brain complex traits yields insights into the etiology of Parkinson's disease. Nat Genet 52:482–493.
- Li M, Santpere G, Imamura Kawasawa Y, Evgrafov OV, Gulden FO, Pochareddy S, et al. (2018): Integrative functional genomic analysis of human brain development and neuropsychiatric risks. Science 362: aaat7615
- Korotkevich G, Sukhov V, Budin N, Shpak B, Artyomov MN, Sergushichev A (2021): Fast gene set enrichment analysis. bioRxiv. https://doi.org/10.1101/060012.
- Yu G (2018): clusterProfiler: An universal enrichment tool for functional and comparative study. bioRxiv. https://doi.org/10.1101/256784.

- Piñero J, Queralt-Rosinach N, Bravo À., Deu-Pons J, Bauer-Mehren A, Baron M, et al. (2015): DisGeNET: A discovery platform for the dynamical exploration of human diseases and their genes. Database (Oxford) 2015:bav028.
- Selemon LD, Ceritoglu C, Ratnanather JT, Wang L, Harms MP, Aldridge K, et al. (2013): Distinct abnormalities of the primate prefrontal cortex caused by ionizing radiation in early or midgestation. J Comp Neurol 521:1040–1053.
- Antonow-Schlorke I, Schwab M, Cox LA, Li C, Stuchlik K, Witte OW, et al. (2011): Vulnerability of the fetal primate brain to moderate reduction in maternal global nutrient availability. Proc Natl Acad Sci U S A 108:3011–3016.
- Davies C, Segre G, Estradé A, Radua J, De Micheli A, Provenzani U, et al. (2020): Prenatal and perinatal risk and protective factors for psychosis: A systematic review and meta-analysis. Lancet Psychiatry 7:399–410.
- Gene Ontology Consortium (2001): Creating the gene ontology resource: Design and implementation. Genome Res 11:1425–1433.
- Barbitoff YA, Tsarev AA, Vashukova ES, Maksiutenko EM, Kovalenko LV, Belotserkovtseva LD, Glotov AS (2020): A data-driven review of the genetic factors of pregnancy complications. Int J Mol Sci 21:3384.
- Li Y, Muffat J, Omer A, Bosch I, Lancaster MA, Sur M, et al. (2017): Induction of expansion and folding in human cerebral organoids. Cell Stem Cell 20:385–396.e3.
- Lui JH, Hansen DV, Kriegstein AR (2011): Development and evolution of the human neocortex [published correction appears in Cell 2011; 146:332]. Cell 146:18–336.
- Mao Y, Ge X, Frank CL, Madison JM, Koehler AN, Doud MK, et al. (2009): Disrupted in schizophrenia 1 regulates neuronal progenitor proliferation via modulation of GSK3β/β-catenin signaling. Cell 136:1017–1031.
- Paredes I, Himmels P, Ruiz de Almodóvar C (2018): Neurovascular communication during CNS development. Dev Cell 45:10–32.
- Lange C, Turrero Garcia M, Decimo I, Bifari F, Eelen G, Quaegebeur A, et al. (2016): Relief of hypoxia by angiogenesis promotes neural stem cell differentiation by targeting glycolysis. EMBO J 35:924–941.
- Kostović I, Jovanov-Milošević N, Radoš M, Sedmak G, Benjak V, Kostović-Srzentić M, et al. (2014): Perinatal and early postnatal reorganization of the subplate and related cellular compartments in the human cerebral wall as revealed by histological and MRI approaches. Brain Struct Funct 219:231–253.
- **48.** Travis K, Ford K, Jacobs B (2005): Regional dendritic variation in neonatal human cortex: A quantitative Golgi study. Dev Neurosci 27:277–287.
- Keunen K, van der Burgh HK, de Reus MA, Moeskops P, Schmidt R, Stolwijk LJ, et al. (2018): Early human brain development: Insights into macroscale connectome wiring. Pediatr Res 84:829–836.

- Hoerder-Suabedissen A, Molnár Z (2015): Development, evolution and pathology of neocortical subplate neurons. Nat Rev Neurosci 16: 133–146.
- Margulies DS, Ghosh SS, Goulas A, Falkiewicz M, Huntenburg JM, Langs G, et al. (2016): Situating the default-mode network along a principal gradient of macroscale cortical organization. Proc Natl Acad Sci U S A 113:12574–12579.
- Broyd SJ, Demanuele C, Debener S, Helps SK, James CJ, Sonuga-Barke EJS (2009): Default-mode brain dysfunction in mental disorders: A systematic review. Neurosci Biobehav Rev 33:279–296.
- Murray RM, Lewis SW, Reveley AM (1985): Towards an aetiological classification of schizophrenia. Lancet 1:1023–1026.
- Weinberger DR (1987): Implications of normal brain development for the pathogenesis of schizophrenia. Arch Gen Psychiatry 44:660– 669.
- Marenco S, Weinberger DR (2000): The neurodevelopmental hypothesis of schizophrenia: Following a trail of evidence from cradle to grave. Dev Psychopathol 12:501–527.
- McKeown CR, Cline HT (2019): Nutrient restriction causes reversible G2 arrest in Xenopus neural progenitors [published correction appears in Development 2020; 147:dev195479]. Development 146:dev178871.
- Levison SW, Rothstein RP, Romanko MJ, Snyder MJ, Meyers RL, Vannucci SJ (2001): Hypoxia/ischemia depletes the rat perinatal subventricular zone of oligodendrocyte progenitors and neural stem cells. Dev Neurosci 23:234–247.
- Gumusoglu SB, Chilukuri ASS, Santillan DA, Santillan MK, Stevens HE (2020): Neurodevelopmental outcomes of prenatal preeclampsia exposure. Trends Neurosci 43:253–268.
- Velasco S, Paulsen B, Arlotta P (2020): 3D brain organoids: Studying brain development and disease outside the embryo. Annu Rev Neurosci 43:375–389.
- Nonaka-Kinoshita M, Reillo I, Artegiani B, Martínez-Martínez MÁ., Nelson M, Borrell V, Calegari F (2013): Regulation of cerebral cortex size and folding by expansion of basal progenitors. EMBO J 32:1817– 1828
- Conel JL (1959): The Postnatal Development of the Human Cerebral Cortex: The Cortex of the Twenty-Four-Month Infant. Cambridge, MA: Harvard University Press.
- **62.** Buxhoeveden DP, Casanova MF (2002): The minicolumn hypothesis in neuroscience. Brain 125:935–951.
- Hill J, Inder T, Neil J, Dierker D, Harwell J, Van Essen D (2010): Similar patterns of cortical expansion during human development and evolution. Proc Natl Acad Sci U S A 107:13135–13140.
- Cafiero R, Brauer J, Anwander A, Friederici AD (2019): The concurrence of cortical surface area expansion and white matter myelination in human brain development. Cereb Cortex 29:827–837.