

University of Groningen



The role of PET/CT in large vessel vasculitis and related disorders

Jamar, François; Gormsen, Lars C.; Yildiz, Halil; Slart, Riemer H.; van der Geest, Kornelis S.; Gheysens, Olivier

Published in: The Quarterly Journal of Nuclear Medicine and Molecular Imaging

DOI: 10.23736/S1824-4785.22.03465-3

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version Publisher's PDF, also known as Version of record

Publication date: 2022

Link to publication in University of Groningen/UMCG research database

Citation for published version (APA): Jamar, F., Gormsen, L. C., Yildiz, H., Slart, R. H., van der Geest, K. S., & Gheysens, O. (2022). The role of PET/CT in large vessel vasculitis and related disorders: diagnosis, extent evaluation and assessment of therapy response. The Quarterly Journal of Nuclear Medicine and Molecular Imaging, 66(3), 182-193. https://doi.org/10.23736/S1824-4785.22.03465-3

Copyright Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: https://www.rug.nl/library/open-access/self-archiving-pure/taverneamendment.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): http://www.rug.nl/research/portal. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

© 2022 EDIZIONI MINERVA MEDICA Online version at https://www.minervamedica.it The Quarterly Journal of Nuclear Medicine and Molecular Imaging 2022 September;66(3):182-93 DOI: 10.23736/S1824-4785.22.03465-3

REVIEW

PET/CT IMAGING IN AUTOIMMUNE DISORDERS

The role of PET/CT in large vessel vasculitis and related disorders: diagnosis, extent evaluation and assessment of therapy response

François JAMAR¹*, Lars C. GORMSEN², Halil YILDIZ³, Riemer H. SLART^{4, 5}, Kornelis S. van der GEEST⁶, Olivier GHEYSENS¹

¹Department of Nuclear Medicine, Saint-Luc University Clinics and Institute of Clinical and Experimental Research (IREC), Catholic University of Louvain (UCLouvain), Brussels, Belgium; ²Department of Nuclear Medicine and PET Center, Aarhus University Hospital, Aarhus, Denmark; ³Department of Internal Medicine and Infectious Diseases, Saint-Luc University Clinics, Brussels, Belgium; ⁴Department of Nuclear Medicine and Molecular Imaging, University Medical Center of Groningen, University of Groningen, Groningen, the Netherlands; ⁵Department of Rheumatology and Clinical Immunology, University Medical Center of Groningen, University of Groningen, University of Groningen, University of Groningen, the Netherlands; ⁶Department of Rheumatology and Clinical Immunology, University Medical Center of Groningen, University of Twente, Enschede, the Netherlands; ⁶Department of Rheumatology and Clinical Immunology, University Medical Center of Groningen, University Of Groningen, Univ

*Corresponding author: François Jamar, Department of Nuclear Medicine, Saint-Luc University Clinics and Institute of Clinical and Experimental Research (IREC), Catholic University of Louvain (UCLouvain), avenue Hippocrate 10, B-1200 Brussels, Belgium. E-mail: francois.jamar@uclouvain.be

ABSTRACT

Large vessel vasculitides (LVV) are defined as chronic inflammatory disorders that affect the arteries with two major variants being distinguished: giant cell arteritis (GCA) and Takayasu's arteritis (TAK). These often present with nonspecific constitutional symptoms which makes an accurate diagnosis often challenging. Nevertheless, timely diagnosis is of utmost importance to initiate treatment and to avoid potential life-threatening complications. [¹⁸F]FDG-PET/CT is nowadays widely accepted as useful tool to aid in the diagnosis of large vessel vasculitis. However, its role to monitor disease activity and to predict disease relapse during follow-up is less obvious since vascular [¹⁸F]FDG uptake can be detected in the absence of clinical or biochemical signs of disease activity. In addition to the two major variants, [¹⁸F]FDG-PET/CT has shown promise in (peri-)aortitis and related disorders. This article aims to provide an update on the current knowledge and limitations of [¹⁸F] FDG-PET/CT for the diagnosis and assessment of treatment response in LVV. Furthermore, other radiopharmaccuticals targeting key components of the vascular immune system are being discussed which could provide an interesting alternative to image vascular inflammation in LVV.

(Cite this article as: Jamar F, Gormsen LC, Yildiz H, Slart RH, van der Geest KS, Gheysens O. The role of PET/CT in large vessel vasculitis and related disorders: diagnosis, extent evaluation and assessment of therapy response. Q J Nucl Med Mol Imaging 2022;66:182-93. DOI: 10.23736/S1824-4785.22.03465-3)

KEY WORDS: Positron emission tomography computed tomography; Fluorodeoxyglucose F18; Vasculitis; Aortitis.

Vasculitides encompass a wide spectrum of disorders, with all in common the feature of inflammation of blood vessels. The primary vasculitides are categorized as a function of the vessels' size: large, medium and small vessels.¹ Medium and small vessel vasculitides usually require pathological confirmation on biopsies. Large vessel vasculitis (LVV) is characterized by inflammation of large to medium-sized vessels and is associated with a triad of

pathophysiological features: arterial occlusion, aneurysm formation and in some cases arterial dissection (aorta). These pathological mechanisms may lead to severe symptoms such as transient or definitive loss of vision, strokes, or acute peripheral occlusive symptoms. Two main forms of LVV are well identified, giant cell arteritis (GCA) and Takayasu arteritis (TKA). Besides, aortitis of various aetiologies have been reported. Large vessel vasculitis (LVV)

PET/CT IN VASCULITIS AND RELATED DISORDERS

occurs in vessels large enough to be accessible to noninvasive imaging. Positron emission tomography with computed tomography using 2-deoxy-2-[18F]fluoro-Dglucose ([18F]FDG-PET/CT) is a whole-body functional imaging modality widely used in oncology and has also demonstrated its versatility in inflammatory disorders, initially by serendipity.^{2, 3} [¹⁸F]FDG-PET was shown in experimental models and in the clinics to enable detection of increased glucose uptake in immune cells and fibroblasts in inflamed vessel walls.^{2, 4, 5} This review deals with the potential of functional imaging with a focus on $[^{18}F]$ FDG-PET/CT, to help in the diagnosis of LVV, but also the extent evaluation and the assessment of response to therapy, essentially in the two major forms, with a few words about atypical vasculitis presentations. In addition, potential future developments of functional imaging using other pathophysiological imaging pathways will be discussed in LVV.

Clinical spectrum of LVV and related disorders

Vasculitis is a group of disorders affecting both arteries and veins. According to the International Chapel Hill consensus conference in 2012, the term "large vessel" refers to the aorta and its major branches.¹ GCA and Takayasu arteritis (TAK) are the most common causes of LVV. Isolated aortitis is also considered as a LVV but classified as single-organ vasculitis compared to TAK and GCA which are potentially (multi)-systemic vasculitides.¹ Both LVV subtypes mainly differ in terms of epidemiological aspects (ethnic distribution, age of onset, gender), distribution pattern, treatment response to the same drug and prognosis. They however share clinical, imaging and histopathological features.6-8

A prompt recognition and treatment of GCA and TAK is essential to avoid irreversible complications/damage. Histopathologically, GCA and TAK are characterized by large vessel wall infiltration by inflammatory cells such as mononuclear cells and macrophages.9, 10 However, the clinical presentation is different. GCA mainly affects patients of ≥ 50 years of age while TAK occurs in young women \leq 40 years of age.^{11, 12} TAK and GCA are characterized by nonspecific symptoms such as fever, fatigue, malaise, weight loss but also more typical symptoms such as headache, jaw claudication, and scalp tenderness in GCA versus limb claudication (with loss of pulse) and carotidynia in TAK. Furthermore, GCA may occur simultaneously with polymyalgia rheumatic (PMR) and TAK with inflammatory bowel disease, sarcoidosis or ankylosing spondylitis.¹¹⁻¹³ The most striking clinical presentation of GCA is loss of vision, sometimes but not always preceded by transient loss, so called amaurosis fugax or just resulting in sudden irreversible blindness. stroke, aorta aneurysm or dissection in large vessel GCA (LvGCA). The main complications of TAK are transitory ischemic attack, stroke and large vessel steno-occlusive lesions as well as renovascular hypertension due to renal artery stenosis, pulmonary thrombosis and aortic regurgitation.¹⁴ Nonspecific constitutional symptoms, together with markers of systemic inflammation such as C-reactive protein (CRP) or erythrocyte sedimentation rate (ESR) make the diagnosis of LVV very challenging and prompt to additional diagnostic criteria.^{15, 16} The American College of Rheumatology 1990 criteria for GCA, are based on clinical and histological features, but are not suitable for individual patients.^{17, 18} In all cases, early imaging, such as ultrasound, MRI, CT and [18F]FDG-PET/CT is highly recommended to avoid delay in diagnosis and therapeutic actions as highlighted in the latest EULAR imaging recommendations, the latter being discussed in this paper.¹⁹ The treatment of these diseases is based on corticosteroids as a first line and followed with diseasemodifying antirheumatic drugs (DMARDS)/immunosuppressive agent(s) as a second or subsequent line.²⁰

Besides the two typical LVV, non-infectious aortitis is a third entity that can be either isolated or related to several systemic disorders.²¹ Such association has been shown for instance with Behçet's disease, Cogan's syndrome, immunoglobulin G4-related diseases (IgG4-RD), relapsing polychondritis, granulomatosis with polyangiitis (GPA), spondyloarthritis, PMR, etc.¹³ Aortitis can also be the only (initial) manifestation of a systemic LVV and if no etiology is found, the aortitis is classified as "clinically isolated aortitis."1 The constitutional symptoms are similar to those encountered in GCA or TAK. More specific symptoms can be present according to the underlying systemic disease, for example, 1) chondritis of ear and nasal cartilage with or without scleritis in relapsing polychondritis; 2) genital and mouth ulcers, uveitis in Behçet's disease; 3) interstitial keratitis and vestibuloauditory symptoms in Cogan's syndrome; 4) bilateral swelling of lacrymal or salivary glands, orbital pseudotumor in IgG4-RD. Histopathological findings are usually similar to those reported in GCA or TAK. In IgG4-RD though, the pathological findings comprise dense lymphoplasmocytic inflammation, storiform fibrosis, obliterative phlebitis and eosinophilia.²² The main complications of aortitis are the formation of aneurysm and dissection.

ъ

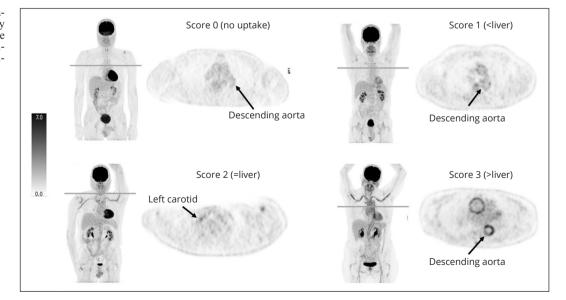
The role of PET-CT in LVV: diagnosis, extent evaluation and therapy response assessment

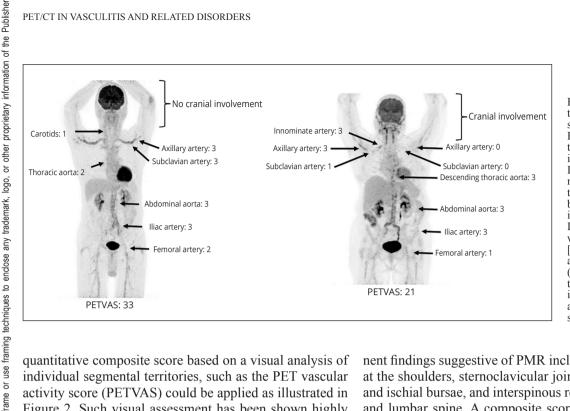
Diagnostic value of [18F]FDG-PET/CT and extent evaluation

[18F]FDG-PET/CT imaging is considered a good diagnostic tool for LVV, even though its exact diagnostic accuracy is difficult to determine due to a lack of an independent reference standard. In addition, meta-analyses on the diagnostic performance of [18F]FDG-PET or [18F] FDG-PET/CT are mainly based on retrospective data. The meta-analysis by Soussan et al. including 4 studies in GCA patients and 7 studies in TAK patients reported a pooled sensitivity of 90% and 87% and pooled specificity of 98% and 73%, respectively.23 Another meta-analysis of 8 studies reported an overall pooled sensitivity of 76% and pooled specificity of 93% for diagnosing LVV (GCA and TAK) and a sensitivity of 83% and specificity of 90% for the GCA subgroup.²⁴ The diagnostic performance of [¹⁸F] FDG-PET/CT was further investigated in two more recent studies that included patients undergoing [18F]FDG-PET/ CT for suspected LVV. Larivière et al. found a sensitivity of 67% and specificity of 100% of [18F]FDG-PET/CT for a diagnosis of LvGCA and Moragas Solanes et al. reported a sensitivity of 96% and specificity of 97% for a diagnosis of LVV (GCA and TAK).^{25, 26} Although FDG uptake patterns may show strong similarities between individual patients with GCA and TAK, it has been shown that differences in distribution exist between the two main forms of LVV on a group level.^{27, 28} Aortic involvement is a prominent feature in both conditions. However, involvement of abdominal arteries and predominant involvement of the subclavian / carotid arteries is more frequently observed in TAK. Patients with GCA more often present with diffuse vascular involvement or bilateral axillary/subclavian artery involvement. Pulmonary artery involvement on [18F]FDG-PET/CT has also been described in a series of patients with TAK.²⁹ Till recently, studies on [18F]FDG-PET/CT have been restricted to assess LVV involvement in patients with systemic vasculitis, although increasing evidence indicates that improved resolution of the newest generation (digital) PET scanners allows for assessing inflammation of cranial arteries, such as the vertebral, temporal and maxillary arteries, in patients with cranial GCA (cGCA). Two independent studies reported a sensitivity of 79%-83% and specificity of 75-100% of [18F]FDG-PET/CT for cGCA.30,31

To date, there is no sufficient evidence that semiquantitative [¹⁸F]FDG-PET/CT parameters may outperform visual assessment to diagnose (extracranial) LVV.³² Visual analysis (arterial uptake compared to a reference organ) should be preferred in routine clinical practice. The recommended visual analysis is a standardized 4-point grading scale with grade 0: no uptake, grade 1: vascular uptake less than liver, grade 2: vascular uptake equal to liver and grade 3: vascular uptake greater than liver as illustrated in Figure 1. When applying this 4-point grading scale, a diffuse and segmental grade 3 uptake in the wall of the aorta and/or its main branches (including the carotid bifurcation and vertebral arteries) is considered positive for LVV while a grade 2 uptake may be indicative of LVV.³³ In addition, a

Figure 1.—Representative maximum intensity projection and transverse [¹⁸F]FDG PET images illustrating the 4-point visual grading scale.





2.-Representa-Figure tive maximum intenprojection [18F]FDG sitv PÉT images illustrating the PET vascular activity score (PETVAS) in a LvGCA patient with symmetric [18F]FDG uptake in the aorta and most larger branches without cranial involvement (left panel). LvGCA and cGCA patient with more asymmetrical [18F]FDG uptake in the aorta and its branches (right panel). In addition, there was extensive and involvement of the cranial arteries (typically not considered in the PETVAS).

quantitative composite score based on a visual analysis of individual segmental territories, such as the PET vascular activity score (PETVAS) could be applied as illustrated in Figure 2. Such visual assessment has been shown highly reproducible among observers, which is essential in daily practice for clinical decision-making based on the scan report.³⁴ In this regard, it is important to acknowledge that the diagnostic accuracy of [18F]FDG-PET/CT is impaired in patients on immunosuppressive treatment. Stellingwerff et al. demonstrated an increase in sensitivity from 83% to 92% without affecting specificity when patients receiving glucocorticoids were excluded from the analysis.35 This can, at least partially, be explained by a treatment-induced reduction or normalization of arterial wall uptake in combination with a trend towards higher liver uptake. Several groups have indeed argued that the liver uptake might not be the most suitable reference organ in those patients.^{36, 37} Nevertheless, Nielsen et al. have shown that the sensitivity of [18F]FDG-PET/CT for diagnosing LVV was not affected after 3 days of high-dose oral glucocorticoid treatment while nearly two-thirds of patients showed a normalization of wall uptake after 10 days. This suggests that [18F] FDG-PET/CT can be performed within 72 hours of glucocorticoid initiation without attenuating its diagnostic performance.³⁸ Prospective data in the 3-10 days window are currently lacking: accordingly, as of today, adhering to the 72 hours window is recommended.

Since [18F]FDG-PET/CT is a whole-body imaging modality and considerable overlap exists between GCA and PMR, typical uptake patterns for PMR are frequently found in patients eventually diagnosed with GCA. Prominent findings suggestive of PMR include [18F]FDG uptake at the shoulders, sternoclavicular joints, hips, trochanteric and ischial bursae, and interspinous regions of the cervical and lumbar spine. A composite score of visual uptake at these sites provided a sensitivity of 85-90% and specificity of 84-88% according to two reports.39,40 In addition, bilateral diffuse capsular uptake at the knees is present in a large proportion of PMR patients.⁴¹

Taken together, [18F]FDG-PET/CT is a valuable tool for diagnosis and assessment of disease extent in patients with GCA and TAK.

[18F]FDG-PET/CT to evaluate disease activity and response to treatment in LVV

Whereas the value of [18F]FDG-PET/CT to aid in the diagnosis of LVV is well-established, it is far less obvious to what extent – if any – [18F]FDG-PET/CT could play a role in monitoring disease activity. There are several reasons for this, not least the generally low quality of studies investigating the impact of immunosuppressive treatment on vascular wall [18F]FDG uptake in LVV patients. Most studies are cross-sectional, drug dosing varies widely or may even not be reported, disease duration at the time of the [18F]FDG-PET/CT differs, and the interpretation criteria (visual or semi-quantitative metrics) used to evaluate pathological vessel uptake are often not properly described. Furthermore, various criteria for assessment of clinical disease activity are used in these studies. For patients in clinical remission, it is often unclear how long this has been the case at time of the follow-up scan. Nevertheless, a few longitudinal, observational studies have

or any other means which may allow access permitted. It is not permitted to remove,

use is not

The production of reprints for personal or commercial

permitted.

the Article. It is not permitted to t

post on

and save only

download

No additional reproduction is authorized.

laws.

This

to make additional copies (either sporadically

one file and print only one copy of this Article. It is not permitted

been published from which some interesting lessons may be drawn (reviewed recently by van der Geest *et al.*).⁴²

The plethora of arguments for introducing imaging to evaluate treatment response in patients with LVV is compelling though. If not properly controlled, LVV is associated with serious vascular complications such as aortic dilation (and dissection), claudication and potentially strokes.^{43, 44} For decades, the mainstav of treatment has been corticosteroids, which are associated with a number of adverse effects, in particular diabetes and osteoporosis.²⁰ Treating physicians will therefore try to taper and ultimately discontinue corticosteroid treatment as soon as possible. However, there are currently no reliable biomarkers or definite clinical signs of relapsing or residual active LVV disease, and patient and physician perception of disease does not always align.45,46 Consequently, treatment with corticosteroids or corticosteroid-sparing drugs such as methotrexate or tocilizumab is often continued well beyond the necessary time-period.⁴⁷ In this context, functional imaging with [18F]FDG-PET/CT appears to be a logical choice to aid in the decision-making, since $[^{18}F]$ FDG is a marker of cell metabolism and generally reflects disease activity in both malignant and non-malignant diseases.

Ideally, [¹⁸F]FDG-PET/CT performed in LVV patients during treatment should therefore facilitate: 1) differentiation between active and inactive disease; 2) prediction of relapse risk; and 3) choosing whether to continue, escalate or de-escalate instituted treatment. Using [¹⁸F]FDG-PET/ CT in these scenarios would closely mimic the use of [¹⁸F] FDG-PET/CT in malignant lymphoma, which is currently a widely accepted clinical indication for using [¹⁸F]FDG-PET/CT in treatment response evaluations.⁴⁸ However, previously published [¹⁸F]FDG-PET/CT studies in LVV only contain limited data concerning the first point and a few scattered observations concerning the second point. To the best of our knowledge, no properly conducted studies have been performed, which could potentially back up the third point.

Vessel wall [18F]FDG-uptake during treatment

Vessel wall [¹⁸F]FDG uptake in patients with active LVV is thought to mostly reflect infiltrating activated macrophages and lymphocytes causing both transmural panarteritis and adventitial periarteritis.⁹ Therefore, vessel wall uptake should decrease in tandem with decreased leukocyte activity when immunosuppressive therapy is instituted. Since the aim of immunosuppressive therapy is to control the inflammatory activity rather than to cure it, the [¹⁸F]FDG- PET/CT report should provide an overall measurement/ assessment of inflammatory disease burden, similar to the metabolic tumor volume used in oncology. To address this issue, the PETVAS has been proposed in which [18F]FDG uptake is rated in the most commonly affected vascular territories using a visual 4-point scale ranging from 0-3 allowing for a maximum score of 27.49 Depending on the number of segments chosen (typically between 7 and 15), the PETVAS Score can yield a maximum score of 21 to 45. This approach has proven robust with little inter-observer variability and is currently widely endorsed for disease evaluation in treatment response studies.³³ As an alternative, various target-to-background [18F]FDG-PET ratios of the most active vessel may be used as proposed by Meller et al., but these are less suitable for longitudinal studies compared to the PETVAS since they do not convey the extent of disease, but only the severity.50

Overall, a few short- and long-term longitudinal studies in patients with both GCA and TAK have demonstrated a rapid reduction of vessel wall [18F]FDG uptake in response to treatment. A first proof-of-concept study was performed by Blockmans et al. who showed a significant decrease in vascular [18F]FDG uptake at 3 months after glucocorticoid initiation in patients with GCA.51 However, no further changes in vascular [18F]FDG uptake were observed at 6 months even though all patients were in clinical remission and PETVAS was not able to identify patients at high risk for relapse. Similarly, Nielsen et. al demonstrated that 10 days treatment with 60 mg prednisone reduces vascular [18F]FDG uptake by ~40%, and Rimlandet al. observed a ~20% reduction in PETVAS after a median of 6 months corticosteroid treatment.^{38, 52} The treatment effect on vessel wall [18F]FDG uptake is not restricted to corticosteroids as attested by the strikingly similar reduction observed in patients treated with tocilizumab.53-55 Compared to the effect of corticosteroids, methotrexate or tocilizumab revealed a tendency towards a greater reduction in vascular inflammation as reflected by PETVAS.56 Regardless of the PET metric used, vascular [18F]FDG uptake generally tends to decrease between 20% and 60% from diagnosis to time of treatment response evaluation but in a manner not linearly correlated with the duration of treatment.56,57 However, in contrast to the often completely absent [18F]FDG uptake by residual lymphomatous lesions during/after chemotherapy, vessel wall [18F]FDG uptake is only rarely completely normalized in patients with LVV during treatment. As a consequence, the pooled proportion of [18F]FDG-PET scans still indicative of active LVV in patients in clinical remission (putatively false

PET/CT IN VASCULITIS AND RELATED DISORDERS

positive scans) has been estimated to be at least $\sim 25\%$ (as reviewed by van der Geest et al.42) with values up to 58% as reported in a recent study.58 To add further complexity to the read-out of [18F]FDG-PET scans during treatment, biomarkers that are often used to substantiate the patient's remission status (CRP and ESR) have been demonstrated to correlate poorly with vessel wall [18F]FDG uptake.59,60 The pathophysiological importance of persistent vascular [18F]FDG uptake and the lack of association between this uptake and biochemical inflammatory markers remains unknown. The residual [18F]FDG uptake observed on follow-up PET studies may relate to other processes than the inflammatory cell associated vessel wall [18F] FDG uptake at diagnosis or prior to treatment. Vessel wall fibrosis and remodeling, low-grade inflammation caused by atherosclerosis, and hypoxia have been suggested as possible sources of persistent vessel wall [18F]FDG uptake in the absence of clinical or biochemical signs of disease relapse.⁶¹⁻⁶³ Regardless of the underlying cause, a recent meta-analysis on the diagnostic performance of [18F]FDG-PET/CT during immunosuppressive treatment reported a modest pooled sensitivity of 77% and specificity of 71% to detect active LVV disease.42 Such a moderate diagnostic accuracy (based on only four studies and with wide confidence intervals) does not merit using $[^{18}F]$ FDG-PET/CT to singularly either rule-in or rule-out active LVV.

In addition to monitoring changes in vascular uptake during treatment, several studies have explored whether [18F]FDG-PET/CT performed during clinical remission may predict disease relapse. Gravson et. al. observed that patients with a high PETVAS (>20) were more likely to experience a relapse (45% vs. 11%) than patients with low PETVAS (<20) with a median follow-up time of 15 months.58 Along the same line, Rimland et. al. observed disease relapse in 11 patients in clinical remission who had PETVAS scores above 20 with a median follow-up time of 6 months.⁵² It should be noted though, that a PETVAS score of 20 (with 27 being the highest PETVAS value) implies that at least some of the vascular territories still had a significant [18F]FDG uptake (above liver uptake). Vascular FDG uptake of that magnitude would in many studies have been interpreted as active disease. In this context, it is thought-provoking that PETVAS did not significantly increase from clinical remission to clinical relapse in the study by Rimland et al.52 Therefore, evidence of increased vessel wall [18F]FDG uptake during clinical remission does not allow discriminating between subclinical active disease and *risk* of future relapse.

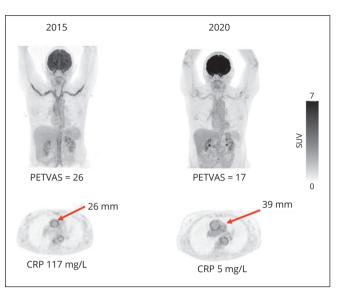


Figure 3.--Patient diagnosed with LvGCA in 2015, treated with corticosteroids and methotrexate. Inflammatory disease burden assessed by PETVAS was 26 (left panel). Five years after the pre-treatment [18F] FDG-PET/CT and in a period of clinical and biochemical remission, she was referred for a staging [18F]FDG-PET/CT for an unrelated disease. Vessel wall FDG uptake was still above liver in the aorta and the PET-VAS had not normalized (score of 17). Of note, an increasing diameter of the ascending aorta was noted on the second scan reflecting progressive aortic dilation despite immunosuppressive treatment.

In summary, vessel wall [18F]FDG uptake decreases during immunosuppressive treatment in patients with LVV, but not to an extent that allows for robust discrimination between active and inactive disease as illustrated in Figures 3, 4. Some data indicate that a high PETVAS during clinical remission is associated with an elevated risk of later disease relapse, but it is evenly possible that a high PETVAS during clinical remission simply reflects subclinical active LVV. Finally, there are currently no studies available that support the use of [18F]FDG-PET/CT to de-escalate or escalate immunosuppressive treatment. As outlined above, a complex relationship exists between clinical, laboratory and imaging findings in assessing disease activity during treatment in patients with LVV and these different disease activity parameters do not always align. Treatment decisions for individual patients should thus be based on all disease activity parameters available and guided by the disease features and existence of comorbidities in those patients. Therefore, future studies are warranted to develop and validate more comprehensive composite outcome measurements for assessing disease activity during treatment, to define the optimal timing at which repeat imaging should be performed, to guide treatment, and to predict disease relapse.

cover.

This ъ

JAMAR

PET/CT IN VASCULITIS AND RELATED DISORDERS

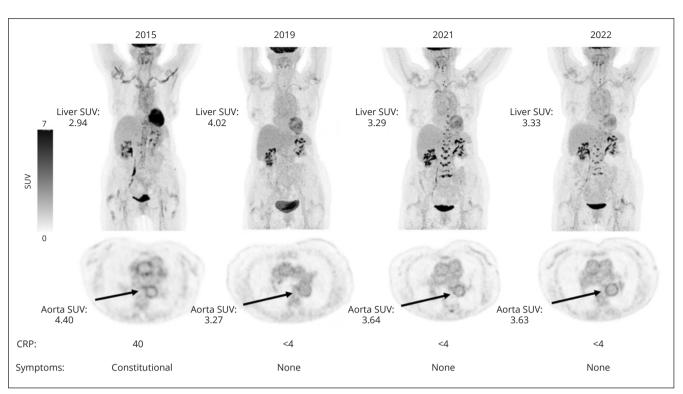


Figure 4.—Patient referred for a [¹⁸F]FDG-PET/CT scan for suspicion of LvGCA, which was confirmed (left panel). She was treated with corticosteroids for a total of 18 months. In the following 7 years, three additional [¹⁸F]FDG-PET/CT's were performed as surveillance of an unrelated disease. In these, aortic vessel wall [¹⁸F]FDG uptake gradually increased to above liver level. In addition, marked [¹⁸F]FDG uptake was noted in the interspinal ligamentous bursae. At no point after the initial tapering of corticosteroid therapy did the patient experience clinical relapse and inflammatory markers were not elevated. CRP is in mg/L.

[18F]FDG-PET/CT in other vasculitides and peri-aortitis

Besides the two main forms of vasculitis, there are numerous reports of the use of [18F]FDG-PET/CT in aortitis and related disorders. Most are anecdotal or merged in heterogeneous series comprising mainly LVV and TAK. Chen et al., recently reported the performance of [18F]FDG-PET/CT in large-, medium- and small-vessel vasculitis: in a series of 71 patients, [18F]FDG-PET/CT was positive in 93% of the patients, showing vascular and extravascular involvement in 64% and 53%, respectively.64 In Behçet's disease, most prospective publications deal with brain involvement. An interesting case report showed multiple cardiovascular involvements in a patient with Behçet's disease.65 The interest of [18F]FDG-PET/CT however goes well beyond vascular hypermetabolism, for it enables to show both vascular and extravascular involvement as illustrated in a small series by Cho et al.66 Behçet's disease was widely present in series of patients with pyrexia of unknown origin, although to the best of our knowledge, no prospective trial has been conducted in this particular disease.67

[¹⁸F]FDG-PET/CT also showed its efficacy in detecting extravascular and vascular involvement in anti-neutrophil cytoplasmic antibodies (ANCA)-associated vasculitis.⁶⁸

Recently, there has been growing interest in IgG4-RD. This versatile disorder may affect large and medium vessels but also various organs. The differential diagnosis stays among that of inflammation of unknown origin and hence a whole-body functional imaging method such as [18F]FDG-PET/CT is of interest. Tsuji et al. showed a correlation between [18F]FDG uptake and histopathology in 21 patients with IgG4-RD.⁶⁹ Interestingly, they reported a SUVmean of 4.07 as being the threshold for lesions to be biopsied for a definite diagnosis. This was confirmed by other authors who identified the role of [18F]FDG-PET/ CT in IgG4-RD to assess organ involvement, guide biopsy and monitor disease response.⁷⁰ There is now ample data about the potential of [18F]FDG-PET/CT in IgG4-RD, but limited evidence suggests that vasculitis is relatively uncommon in IgG4-RD.71 In a recent report, only 15% of the patients with IgG4 presented with aortitis or periaortitis, mainly with involvement of the abdominal aorta.72

PET/CT IN VASCULITIS AND RELATED DISORDERS

Using the same 2011 comprehensive diagnostic criteria, Yabusaki *et al.* found a prevalence of 41% of aortitis using [¹⁸F]FDG-PET/CT, affecting mainly the iliac arteries and infra-renal aorta.^{73, 74} They also demonstrated higher target-to-background ratios in patients with IgG4-aortitis-positive regions than in the negative regions (2.1 *vs.* 1.2, P<0.0001). During the last few years, PET/CT using a [⁶⁸Ga] labelled fibroblast activating protein inhibitor ([⁶⁸Ga]-FAPI) showed promising results in a few case reports of IgG4-RD and a small series of 26 patients in whom a comparison with [¹⁸F]FDG-PET/CT was performed. [⁶⁸Ga]-FAPI PET/CT showed additional sites that were not detected by [¹⁸F]FDG-PET/CT.⁷⁵

Peri-aortitis is a relatively rare disease characterized by infiltration of the tissues surrounding the aorta. It is most often affecting the abdominal aorta and the iliac arteries.⁷⁶ It may be secondary to several causes such as IgG4-RD, neoplastia, histiocytosis (Erdheim-Chester), infections, systemic lupus erythematosus, ankylosing spondylitis and granulomatosis with polyangiitis, also referred to as Wegener's disease. Systemic symptoms are possibly due to the underlying diseases as well as local symptoms secondary to the retroperitoneal fibrosis (RPF). Peri-aortitis due to idiopathic or secondary retroperitoneal fibrosis was well studied with [18F]FDG-PET/CT. An example is given in Figure 5. Einspieler et al. demonstrated that PET-MRI was superior to biochemical markers of systemic inflammation (i.e. CRP and ESR) to evaluate the disease extent.⁷⁷ Another paper reported that [18F]FDG-PET/CT augmented with contrast-enhanced-CT could be a one-step procedure to evaluate patients with RPF.⁷⁸ This paper showed a better identification of patients who would respond to treatment based on the baseline SUV_{max}. Finally, the study by Fernando *et al.* reported on the negative and positive predictive value in RPF.⁷⁹ They concluded that [¹⁸F]FDG-PET/ CT enables to discriminate between cancer-related *versus* not cancer related RPF. Ruhlmann *et al.* showed that [¹⁸F] FDG-PET/MRI had a potential as a one-stop diagnostic procedure for the assessment of active RPF.⁸⁰ More recently, [⁶⁸Ga]-FAPI PET/CT may also prove useful in RPF.⁸¹

Other PET/CT tracers in LVV

As discussed above, [18F]FDG-PET/CT is an established technique to diagnose patients with LVV, but its role for evaluating response to treatment and identifying residual active disease needs to be further clarified. [18F]FDG is taken up by all metabolically active cells that primarily use glucose as substrate and as such is not limited to inflammatory cells (e.g. lymphocytes and macrophages). As consequence, metabolically active cells that are not part of the inflammatory response may, at least to some degree, contribute to the origin of the vascular [18F]FDG uptake. Lower background activity, thereby increasing target-tobackground ratio and the imaging accuracy is of additional value. Also aiding the design of personalized treatment regimens especially with emerging novel immunotherapies is more important these days. Therefore, alternative radiotracers targeting key components of the vascular immune system could provide an interesting alternative to image vascular inflammation in general. Furthermore, one

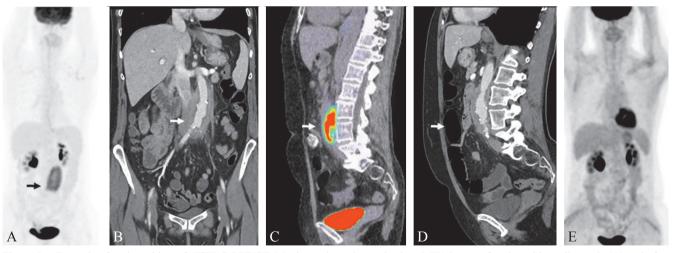


Figure 5.—Example of peri-aortitis on [¹⁸F]FDG-PET/CT. Maximum intensity projection (MIP) image of peri-aortitis at diagnosis (A) and after 6-months of low-dose steroid therapy (E), showing complete remission. Coronal and sagittal contrast-enhanced CT images at the time of diagnosis (B, D), showing a tissular infiltration surrounding the infrarenal aorta with both shrinking and moderate dilation of its lumen. These CT findings correlated with the hypermetabolic activity observed on [¹⁸F]FDG-PET/CT (C).

Vol. 66 - No. 3

This document is protected by international copyright laws. No additional reproduction is authorized. It is permitted for personal use to download and save only one file and print only one copy of this Article. It is not permitted to make additional copies (either sporadically or systematically, either printed or electronic) of the Article for any purpose. It is not permitted to distribute the electronic copy of the article through online internet and/or intranet file sharing systems, electronic mailing or any other means which may allow access The production of reprints for personal or commercial use is not permitted. It is not permitted to remove, one file and print only one copy of this Article. It is not permitted to make additional copies (either sporadically proprietary information of the Publisher other logo, or trademark. enclose any 9 use framing techniques permitted. rame or The creation of derivative works from the Article is not the Article. It is not permitted to post on terms of use which the Publisher may The use of all or any part of the Article for any Commercial Use is not permitted. change any copyright notices or overlay, obscure, block, or to the Article. cover.

JAMAR

can envision the development of PET tracers targeted to specific cell receptors that are important during the inflammatory or healing process, but the major challenge remains the exquisite selectivity of such a PET tracer for a particular cell type. Nevertheless, many novel radiotracers for inflammation imaging have been applied in the field of cardiovascular diseases including vasculitis.

Since LVV is a chronic inflammatory disease characterized by the presence of macrophages, several groups have evaluated PET tracers targeting macrophages in patients with LVV.82-84 Pugliese et al. reported that the TSPO ligand [11C]-PK11195 was taken up in the arterial wall of all symptomatic patients (N.=6) with clinical suspicion of vasculitis while none of the asymptomatic control patients showed any tracer uptake.82 Since PK11195 has a moderate affinity for TSPO and low specific binding, second generation TSPO radioligands with higher affinity have been developed.⁸⁵ However, a recent pilot study evaluating such novel generation TSPO ligand, [11C]-PBR06, did not show any uptake in three patients with LVV.86 Another target on macrophages is the somatostatin receptor 2 (SST²) which can be interrogated by [68Ga]DOTATATE or $[^{18}F]$ FET- β AG-TOCA. In the first two patients with TAK, SST² PET/MRI imaging accurately identified arteritis with a pattern similar to the findings on [18F]FDG or MRI.87 Other interesting targets on macrophages include the folate receptor β that is exclusively expressed on activated macrophages, or the CXCR4 receptor that could be imaged using [18F]-PEG-folate and [68Ga]-Pentixafor, respectively.^{88, 89} Other targets may be used in the future such as the IL-2 receptor agonists or antagonists, or B-cell targeting radiotracers, *i.e.* to CD20, with radiolabelled rituximab. However, further studies are warranted to evaluate their usefulness for the diagnosis and treatment monitoring of patients with vasculitis.

Conclusions

Functional imaging, *per se*, has become part of standard of care in patients with suspected vasculitis. Vasculitis encompasses a wide spectrum of disorders, among which GCA and TAK have been widely studied in the past decade. Even though expert opinion has highlighted its use to aid in the diagnosis of LVV, [¹⁸F]FDG-PET/CT is currently not recommended for follow-up evaluation in recent clinical guidelines. Visual assessment and semi-quantitative evaluation for diagnosis and assessment of disease activity were presented herein with their advantages and drawbacks. In addition, [¹⁸F]FDG-PET has also shown

promising in other types of vasculitides, including periaortitis and related disorders. Other tracers may become available in the near future, for this constellation of disorders, such as those targeting activated macrophages or the fibroblast activating protein.

In all cases, regardless of the tracer used, prospective and multicentric studies are necessary to strengthen the role of functional and metabolic imaging across the wide spectrum of vasculitides and related disorders.

References

1. Jennette JC, Falk RJ, Bacon PA, Basu N, Cid MC, Ferrario F, *et al.* 2012 revised International Chapel Hill Consensus Conference Nomenclature of Vasculitides. Arthritis Rheum 2013;65:1–11.

2. Kubota R, Yamada S, Kubota K, Ishiwata K, Tamahashi N, Ido T. Intratumoral distribution of fluorine-18-fluorodeoxyglucose in vivo: high accumulation in macrophages and granulation tissues studied by microautoradiography. J Nucl Med 1992;33:1972–80.

3. Gamelli RL, Liu H, He LK, Hofmann CA. Augmentations of glucose uptake and glucose transporter-1 in macrophages following thermal injury and sepsis in mice. J Leukoc Biol 1996;59:639–47.

4. Matsui T, Nakata N, Nagai S, Nakatani A, Takahashi M, Momose T, *et al.* Inflammatory cytokines and hypoxia contribute to 18F-FDG uptake by cells involved in pannus formation in rheumatoid arthritis. J Nucl Med 2009;50:920–6.

5. Yamada S, Kubota K, Kubota R, Ido T, Tamahashi N. High accumulation of fluorine-18-fluorodeoxyglucose in turpentine-induced inflammatory tissue. J Nucl Med 1995;36:1301–6.

6. Kermani TA, Crowson CS, Muratore F, Schmidt J, Matteson EL, Warrington KJ. Extra-cranial giant cell arteritis and Takayasu arteritis: how similar are they? Semin Arthritis Rheum 2015;44:724–8.

7. Langford CA, Cuthbertson D, Ytterberg SR, Khalidi N, Monach PA, Carette S, *et al.*; Vasculitis Clinical Research Consortium. A Randomized, Double-Blind Trial of Abatacept (CTLA-4Ig) for the Treatment of Giant Cell Arteritis. Arthritis Rheumatol 2017;69:837–45.

8. Langford CA, Cuthbertson D, Ytterberg SR, Khalidi N, Monach PA, Carette S, *et al.*; Vasculitis Clinical Research Consortium. A Randomized, Double-Blind Trial of Abatacept (CTLA-4Ig) for the Treatment of Takayasu Arteritis. Arthritis Rheumatol 2017;69:846–53.

9. Weyand CM, Liao YJ, Goronzy JJ. The immunopathology of giant cell arteritis: diagnostic and therapeutic implications. J Neuroophthalmol 2012;32:259–65.

10. Schmidt J, Kermani TA, Bacani AK, Crowson CS, Cooper LT, Matteson EL, *et al.* Diagnostic features, treatment, and outcomes of Takayasu arteritis in a US cohort of 126 patients. Mayo Clin Proc 2013;88:822–30.

11. Buttgereit F, Matteson EL, Dejaco C. Polymyalgia Rheumatica and Giant Cell Arteritis. JAMA 2020;324:993–4.

12. Podgórska D, Podgórski R, Aebisher D, Dąbrowski P. Takayasu arteritis - epidemiology, pathogenesis, diagnosis and treatment. J Appl Biomed 2019;17:20.

13. Saadoun D, Vautier M, Cacoub P. Medium- and Large-Vessel Vasculitis. Circulation 2021;143:267–82.

14. Seyahi E. Takayasu arteritis: an update. Curr Opin Rheumatol 2017;29:51-6.

15. van der Geest KS, Sandovici M, Brouwer E, Mackie SL. Diagnostic Accuracy of Symptoms, Physical Signs, and Laboratory Tests for Giant Cell Arteritis: A Systematic Review and Meta-analysis. JAMA Intern Med 2020;180:1295–304.

PET/CT IN VASCULITIS AND RELATED DISORDERS

16. Maksimowicz-McKinnon K, Clark TM, Hoffman GS. Takayasu arteritis and giant cell arteritis: a spectrum within the same disease? Medicine (Baltimore) 2009;88:221-6.

17. Hunder GG, Arend WP, Bloch DA, Calabrese LH, Fauci AS, Fries JF, et al. The American College of Rheumatology 1990 criteria for the classification of vasculitis. Introduction. Arthritis Rheum 1990;33:1065-7.

18. Seeliger B. Sznaid J. Robson JC. Judge A. Craven A. Gravson PC. et al. Are the 1990 American College of Rheumatology vasculitis classification criteria still valid? Rheumatology (Oxford) 2017;56:1154-61.

19. Dejaco C, Ramiro S, Duftner C, Besson FL, Bley TA, Blockmans D, et al. EULAR recommendations for the use of imaging in large vessel vasculitis in clinical practice. Ann Rheum Dis 2018;77:636-43

20. Hellmich B, Agueda A, Monti S, Buttgereit F, de Bovsson H. Brouwer E, et al. 2018 Update of the EULAR recommendations for the management of large vessel vasculitis. Ann Rheum Dis 2020;79:19-30.

21. Cinar I, Wang H, Stone JR. Clinically isolated aortitis: pitfalls, progress, and possibilities. Cardiovasc Pathol 2017;29:23-32.

22. Maritati F, Peyronel F, Vaglio A. IgG4-related disease: a clinical perspective. Rheumatology (Oxford) 2020;59(Suppl 3):iii123–31.

23. Soussan M, Nicolas P, Schramm C, Katsahian S, Pop G, Fain O, et al. Management of large-vessel vasculitis with FDG-PET: a systematic literature review and meta-analysis. Medicine (Baltimore) 2015;94:e622.

24. Lee YH, Choi SJ, Ji JD, Song GG. Diagnostic accuracy of 18F-FDG PET or PET/CT for large vessel vasculitis : A meta-analysis. Z Rheumatol 2016;75:924-31.

25. Lariviere D, Benali K, Coustet B, Pasi N, Hyafil F, Klein I, et al. Positron emission tomography and computed tomography angiography for the diagnosis of giant cell arteritis: A real-life prospective study. Medicine (Baltimore) 2016;95:e4146.

26. Moragas Solanes M, Andreu Magarolas M, Martín Miramon JC, Caresia Aróztegui AP, Monteagudo Jiménez M, Oliva Morera JC, et al. Estudio comparativo de la PET/TC con 18F-FDG y la angiografía por TC en la detección de la vasculitis de grandes vasos. Rev Esp Med Nucl Imagen Mol (Engl Ed) 2019;38:280-9.

27. Soriano A, Pazzola G, Boiardi L, Casali M, Muratore F, Pipitone N, et al. Distribution patterns of 18F-fluorodeoxyglucose in large vessels of Takayasu's and giant cell arteritis using positron emission tomography. Clin Exp Rheumatol 2018;36(Suppl 111):99-106.

28. Gribbons KB, Ponte C, Carette S, Craven A, Cuthbertson D, Hoffman GS, et al. Patterns of Arterial Disease in Takayasu Arteritis and Giant Cell Arteritis. Arthritis Care Res (Hoboken) 2020;72:1615-24.

29. Gao W, Gong JN, Guo XJ, Wu JY, Xi XY, Ma ZH, et al. Value of 18F-fluorodeoxyglucose positron emission tomography/computed tomography in the evaluation of pulmonary artery activity in patients with Takayasu's arteritis. Eur Heart J Cardiovasc Imaging 2021;22:541-50.

30. Nienhuis PH. Sandovici M. Glaudemans AW. Slart RH. Brouwer E. Visual and semiquantitative assessment of cranial artery inflammation with FDG-PET/CT in giant cell arteritis. Semin Arthritis Rheum 2020;50:616-23

31. Nielsen BD, Hansen IT, Kramer S, Haraldsen A, Hjorthaug K, Bogsrud TV, et al. Simple dichotomous assessment of cranial artery inflammation by conventional 18F-FDG PET/CT shows high accuracy for the diagnosis of giant cell arteritis: a case-control study. Eur J Nucl Med Mol Imaging 2019;46:184-93.

32. Gheysens O, Jamar F, Glaudemans AW, Yildiz H, van der Geest KS. Semi-Quantitative and Quantitative [18F]FDG-PET/CT Indices for Di-agnosing Large Vessel Vasculitis: A Critical Review. Diagnostics (Basel) 2021:11:2355

33. Slart RH, Glaudemans AW, Chareonthaitawee P, Treglia G, Besson FL, et al.; Writing group; Reviewer group; Members of EANM Cardiovascular; Members of EANM Infection & Inflammation; Members of Committees, SNMMI Cardiovascular; Members of Council, PET Interest Group; Members of ASNC; EANM Committee Coordinator. FDG-PET/ CT(A) imaging in large vessel vasculitis and polymyalgia rheumatica: joint procedural recommendation of the EANM, SNMMI, and the PET Interest Group (PIG), and endorsed by the ASNC. Eur J Nucl Med Mol Imaging 2018;45:1250-69.

34. Lensen KD, Comans EF, Voskuyl AE, van der Laken CJ, Brouwer E, Zwijnenburg AT, et al. Large-vessel vasculitis: interobserver agreement and diagnostic accuracy of 18F-FDG-PET/CT. BioMed Res Int 2015:2015:914692

35. Stellingwerff MD, Brouwer E, Lensen KD, Rutgers A, Arends S, van der Geest KS, et al. Different Scoring Methods of FDG PET/CT in Giant Cell Arteritis: need for Standardization. Medicine (Baltimore) 2015;94:e1542

36. Besson FL, de Boysson H, Parienti JJ, Bouvard G, Bienvenu B, Agostini D. Towards an optimal semiquantitative approach in giant cell arteritis: an (18)F-FDG PET/CT case-control study. Eur J Nucl Med Mol Imaging 2014;41:155-66.

37. Prieto-González S, Depetris M, García-Martínez A, Espígol-Frigolé G, Tavera-Bahillo I, Corbera-Bellata M, et al. Positron emission tomography assessment of large vessel inflammation in patients with newly diagnosed, biopsy-proven giant cell arteritis: a prospective, case-control study. Ann Rheum Dis 2014;73:1388-92.

38. Nielsen BD, Gormsen LC, Hansen IT, Keller KK, Therkildsen P, Hauge EM. Three days of high-dose glucocorticoid treatment attenuates large-vessel 18F-FDG uptake in large-vessel giant cell arteritis but with a limited impact on diagnostic accuracy. Eur J Nucl Med Mol Imaging 2018;45:1119-28.

39. van der Geest KS, van Sleen Y, Nienhuis P, Sandovici M, Westerdijk N, Glaudemans AW, et al. Comparison and validation of FDG-PET/CT scores for polymyalgia rheumatica. Rheumatology (Oxford) 2022;61:1072-82.

40. Henckaerts L, Gheysens O, Vanderschueren S, Goffin K, Blockmans D. Use of 18F-fluorodeoxyglucose positron emission tomography in the diagnosis of polymyalgia rheumatica-A prospective study of 99 patients. Rheumatology (Oxford) 2018;57:1908-16.

41. Cimmino MA, Camellino D, Paparo F, Morbelli S, Massollo M, Cutolo M, et al. High frequency of capsular knee involvement in polymyalgia rheumatica/giant cell arteritis patients studied by positron emission tomography. Rheumatology (Oxford) 2013;52:1865-72

42. van der Geest KS, Treglia G, Glaudemans AW, Brouwer E, Sandovici M, Jamar F, et al. Diagnostic value of [18F]FDG-PET/CT for treatment monitoring in large vessel vasculitis: a systematic review and meta-analysis. Eur J Nucl Med Mol Imaging 2021;48:3886-902.

43. Muratore F, Crescentini F, Spaggiari L, Pazzola G, Casali M, Boiardi L, et al. Aortic dilatation in patients with large vessel vasculitis: A longitudinal case control study using PET/CT. Semin Arthritis Rheum 2019;48:1074-82.

44. Therkildsen P, de Thurah A, Nielsen BD, Hansen IT, Eldrup N, Nørgaard M, et al. Increased risk of thoracic aortic complications among patients with giant cell arteritis: a nationwide, population-based cohort study. Rheumatology (Oxford) 2021;keab871.

45. Kermani TA, Schmidt J, Crowson CS, Ytterberg SR, Hunder GG, Matteson EL, et al. Utility of erythrocyte sedimentation rate and C-reactive protein for the diagnosis of giant cell arteritis. Semin Arthritis Rheum 2012;41:866-71

46. Herlyn K, Hellmich B, Seo P, Merkel PA. Patient-reported outcome assessment in vasculitis may provide important data and a unique perspective. Arthritis Care Res (Hoboken) 2010;62:1639-45.

47. Therkildsen P, de Thurah A, Hansen IT, Nørgaard M, Nielsen BD, Hauge EM. Giant cell arteritis: A nationwide, population-based cohort study on incidence, diagnostic imaging, and glucocorticoid treatment. Semin Arthritis Rheum 2021;51:360-6.

48. Cheson BD, Fisher RI, Barrington SF, Cavalli F, Schwartz LH, Zucca E, et al.; Alliance, Australasian Leukaemia and Lymphoma Group; Eastern Cooperative Oncology Group; European Mantle Cell Lymphoma Consortium; Italian Lymphoma Foundation; European Organisation for Research; Treatment of Cancer/Dutch Hemato-Oncology Group; Grupo Español de Médula Ósea; German High-Grade Lymphoma Study Group;

Ŀ 2

JAMAR

or any other means which may allow access

to make additional copies (either sporadically

copy of this Article. It is not permitted

No additional reproduction is authorized. It is permitted for personal use to download and save only one file and print only one copy of this Article. It is not permitted for any purpose. It is not permitted to distribute the electronic copy of the article through online internet and/or intranet file sharing systems, electronic mailing

permitted to remove,

not

permitted. It is

not

use is r

Б

for personal

production of reprints

The

permitted.

the Article is not

from

works 1 Article.

derivative the /

The creation of

permitted.

any Commercial Use is not

is document is protected by international copyright laws. No a systematically, either printed or electronic) of the Article for

proprietary

other

P

logo, commercial

trademark.

any

enclose

9

techniques

iraming

ISe 1

P

rame

9

permitted

lt is not

Ч

post

may

the Publisher

use which t

terms of

ъ

notices

copyright the Article for

any

ъ

block. Ъ

part of t change

any

all use of

The

the Article.

Fhis ъ 2 cover.

information of the Publisher

German Hodgkin's Study Group; Japanese Lymphorra Study Group; Lymphoma Study Association; NCIC Clinical Trials Group; Nordic Lymphoma Study Group; Southwest Oncology Group; United Kingdom National Cancer Research Institute. Recommendations for initial evaluation. staging, and response assessment of Hodgkin and non-Hodgkin lymphoma: the Lugano classification. J Clin Oncol 2014;32:3059-68

49. Walter MA, Melzer RA, Schindler C, Müller-Brand J, Tyndall A, Nitzsche EU. The value of [18F]FDG-PET in the diagnosis of large-vessel vasculitis and the assessment of activity and extent of disease. Eur J Nucl Med Mol Imaging 2005;32:674-81.

50. Meller J, Strutz F, Siefker U, Scheel A, Sahlmann CO, Lehmann K, et al. Early diagnosis and follow-up of aortitis with [(18)F]FDG PET and MRI. Eur J Nucl Med Mol Imaging 2003;30:730-6.

51. Blockmans D, de Ceuninck L, Vanderschueren S, Knockaert D, Mortelmans L, Bobbaers H. Repetitive 18F-fluorodeoxyglucose positron emission tomography in giant cell arteritis: a prospective study of 35 patients. Arthritis Rheum 2006;55:131-7.

52. Rimland CA. Ouinn KA. Rosenblum JS. Schwartz MN. Bates Gribbons K, Novakovich E, et al. Outcome Measures in Large Vessel Vasculitis: Relationship Between Patient-, Physician-, Imaging-, and Laboratory-Based Assessments. Arthritis Care Res (Hoboken) 2020;72:1296-304.

53. Prieto Peña D, Martínez-Rodríguez I, Atienza-Mateo B, Calderón-Goercke M, Banzo I, González-Vela MC, *et al.* Evidence for uncou-pling of clinical and 18-FDG activity of PET/CT scan improvement in tocilizumab-treated patients with large-vessel giant cell arteritis. Clin Exp Rheumatol 2021;39(Suppl 129):69-75.

54. Quinn KA, Dashora H, Novakovich E, Ahlman MA, Grayson PC. Use of 18F-fluorodeoxyglucose positron emission tomography to monitor tocilizumab effect on vascular inflammation in giant cell arteritis. Rheumatology (Oxford) 2021;60:4384-9.

55. Sebastian A, Kayani A, Prieto-Pena D, Tomelleri A, Whitlock M, Mo J, et al. Efficacy and safety of tocilizumab in giant cell arteritis: a single centre NHS experience using imaging (ultrasound and PET-CT) as a diag-nostic and monitoring tool. RMD Open 2020;6:e001417.

56. Schönau V, Roth J, Tascilar K, Corte G, Manger B, Rech J, et al. Resolution of vascular inflammation in patients with new-onset giant cell arteritis: data from the RIGA study. Rheumatology (Oxford) 2021;60:3851-61.

57. de Boysson H, Aide N, Liozon E, Lambert M, Parienti JJ, Monteil J, et al. Repetitive 18F-FDG-PET/CT in patients with large-vessel giant-cell arteritis and controlled disease. Eur J Intern Med 2017;46:66-70

58. Grayson PC, Alehashemi S, Bagheri AA, Civelek AC, Cupps TR, Kaplan MJ, et al. 18 F-Fluorodeoxyglucose-Positron Emission Tomography As an Imaging Biomarker in a Prospective, Longitudinal Cohort of Patients With Large Vessel Vasculitis. Arthritis Rheumatol 2018:70:439-49

59. Banerjee S, Quinn KA, Gribbons KB, Rosenblum JS, Civelek AC, Novakovich E, et al. Effect of Treatment on Imaging, Clinical, and Serologic Assessments of Disease Activity in Large-vessel Vasculitis. J Rheumatol 2020:47:99-107.

60. Gomez L, Chaumet-Riffaud P, Noel N, Lambotte O, Goujard C, Durand E, et al. Effect of CRP value on 18F-FDG PET vascular positivity in Takayasu arteritis: a systematic review and per-patient based meta-analysis. Eur J Nucl Med Mol Imaging 2018;45:575-81.

61. Miller D, Isotalo PA, Weyand CM, Edwards WD, Aubry MC, Tazelaar HD. Surgical Pathology of Noninfectious Ascending Aortitis: A Study of 45 Cases With Emphasis on an Isolated Variant. Am J Surg Pathol 2006;30:1150-8.

62. Rosenbaum D, Millon A, Favad ZA. Molecular imaging in atherosclerosis: FDG PET. Curr Atheroscler Rep 2012;14:429-37.

63. Folco EJ, Sheikine Y, Rocha VZ, Christen T, Shvartz E, Sukhova GK, et al. Hypoxia but not inflammation augments glucose uptake in human macrophages: implications for imaging atherosclerosis with 18fluorinelabeled 2-deoxy-D-glucose positron emission tomography. J Am Coll Cardiol 2011;58:603-14.

64. Chen Z, Zhao Y, Wang Q, Li Y, Li H, Zhou Y. Imaging features of

18F-FDG PET/CT in different types of systemic vasculitis. Clin Rheumatol 2022;41:1499-509.

65. Xi XY, Gao W, Guo XJ, Jiang W, Yang YH, Gong JN, *et al.* Multiple cardiovascular involvements in Behçet's disease: unique utility of 18F-FDG PET/CT in diagnosis and follow-up. Eur J Nucl Med Mol Imaging 2019:46:2210-1

66. Cho SB, Yun M, Lee JH, Kim J, Shim WH, Bang D, Detection of cardiovascular system involvement in Behçet's disease using fluorodeoxyglucose positron emission tomography. Semin Arthritis Rheum 2011;40:461-6.

67. Mulders-Manders CM, Simon A, Bleeker-Rovers CP, Rheumatologic diseases as the cause of fever of unknown origin. Best Pract Res Clin Rheumatol 2016;30:789-801.

68. Kemna MJ, Vandergheynst F, Vöö S, Blocklet D, Nguyen T, Timmermans SA, et al. Positron emission tomography scanning in anti-neutrophil cytoplasmic antibodies-associated vasculitis. Medicine (Baltimore) 2015;94:e747.

69. Tsuji S, Iwamoto N, Horai Y, Fujikawa K, Fujita Y, Fukui S, et al. Comparison of the quantitative measurement of 18F-FDG PET/CT and histopathological findings in IgG4-related disease. Clin Exp Rheumatol 2021;39:1338-44

70. Tang CY, Chua WM, Cheng LT, Fong W, Zaheer S, Lam WW. 18F-FDG PET/CT Manifestations of IgG4-related Disease. Br J Radiol 2021;94:20210105.

71. Ahpin C, de Oliveira Brito JB, Chow B, Leung E. Aortitis and coronary artery vasculitis of unusual etiology: IgG4-related disease defined by 18F-fluorodeoxyglucose positron emission tomography (FDG-PET). J Nucl Cardiol 2021. [Epub ahead of print]

72. Peng L, Zhang P, Li J, Liu Z, Lu H, Zhu L, et al. IgG4-related aortitis/ periaortitis and periarteritis: a distinct spectrum of IgG4-related disease. Arthritis Res Ther 2020;22:103.

73. Yabusaki S, Oyama-Manabe N, Manabe O, Hirata K, Kato F, Mi-yamoto N, et al. Characteristics of immunoglobulin G4-related aortitis/ periaortitis and periarteritis on fluorodeoxyglucose positron emission tomography/computed tomography co-registered with contrast-enhanced computed tomography. EJNMMI Res 2017;7:20.

74. Umehara H, Okazaki K, Masaki Y, Kawano M, Yamamoto M, Saeki T, et al. Comprehensive diagnostic criteria for IgG4-related disease (IgG4-RD), 2011. Mod Rheumatol 2012;22:21-30.

75. Luo Y, Pan Q, Yang H, Peng L, Zhang W, Li F. Fibroblast Activation Protein-Targeted PET/CT with 68Ga-FAPI for Imaging IgG4-Related Disease: comparison to 18F-FDG PET/CT. J Nucl Med 2021;62:266–71.

76. Marvisi C, Accorsi Buttini E, Vaglio A. Aortitis and periaortitis: the puzzling spectrum of inflammatory aortic diseases. Presse Med 2020;49:104018.

77. Einspieler I, Henninger M, Mergen V, Wendorff H, Haller B, Beyer LP, et al. 18F-FDG PET/MRI compared with clinical and serological markers for monitoring disease activity in patients with aortitis and chronic periaortitis. Clin Exp Rheumatol 2020;38(Suppl 124):99-106.

78. Guignard R. Simukoniene M. Garibotto V. Ratib O. 18F-FDG PET/ CT and contrast-enhanced CT in a one-stop diagnostic procedure: a better strategy for management of patients suffering from retroperitoneal fibrosis? Clin Nucl Med 2012;37:453-9.

79. Fernando A, Pattison J, Horsfield C, D'Cruz D, Cook G, O'Brien T. [18F]-Fluorodeoxyglucose Positron Emission Tomography in the Diagnosis, Treatment Stratification, and Monitoring of Patients with Retroperitoneal Fibrosis: A Prospective Clinical Study. Eur Urol 2017;71:926-33.

80. Ruhlmann V. Poeppel TD. Brandt AS. Grueneisen J. Ruhlmann M. Theysohn JM, et al. Erratum to: 18F-FDG PET/MRI evaluation of retroperitoneal fibrosis: a simultaneous multiparametric approach for diagnosing active disease. Eur J Nucl Med Mol Imaging 2016;43:1395.

81. Pan Q, Luo Y, Zhang W. Idiopathic Retroperitoneal Fibrosis With Intense Uptake of 68Ga-Fibroblast Activation Protein Inhibitor and 18F-FDG. Clin Nucl Med 2021;46:175-6.

PET/CT IN VASCULITIS AND RELATED DISORDERS

82. Pugliese F, Gaemperli O, Kinderlerer AR, Lamare F, Shalhoub J, Davies AH, *et al.* Imaging of vascular inflammation with [11C]-PK11195 and positron emission tomography/computed tomography angiography. J Am Coll Cardiol 2010;56:653–61.

83. Lamare F, Hinz R, Gaemperli O, Pugliese F, Mason JC, Spinks T, *et al.* Detection and quantification of large-vessel inflammation with 11C-(R)-PK11195 PET/CT. J Nucl Med 2011;52:33–9.

84. Jiemy WF, Heeringa P, Kamps JA, van der Laken CJ, Slart RH, Brouwer E. Positron emission tomography (PET) and single photon emission computed tomography (SPECT) imaging of macrophages in large vessel vasculitis: current status and future prospects. Autoimmun Rev 2018;17:715–26.

85. Cumming P, Burgher B, Patkar O, Breakspear M, Vasdev N, Thomas P, *et al.* Sifting through the surfeit of neuroinflammation tracers. J Cereb Blood Flow Metab 2018;38:204–24.

86. Schollhammer R, Lepreux S, Barthe N, Vimont D, Rullier A, Sibon I, *et al.* In vitro and pilot in vivo imaging of 18 kDa translocator protein (TSPO) in inflammatory vascular disease. EJNMMI Res 2021;11:45.

87. Tarkin JM, Wall C, Gopalan D, Aloj L, Manavaki R, Fryer TD, *et al.* Novel Approach to Imaging Active Takayasu Arteritis Using Somatostatin Receptor Positron Emission Tomography/Magnetic Resonance Imaging. Circ Cardiovasc Imaging 2020;13:e010389.

88. Verweij NJ, Yaqub M, Bruijnen ST, Pieplenbosch S, Ter Wee MM, Jansen G, *et al.* First in man study of [18F]fluoro-PEG-folate PET: a novel macrophage imaging technique to visualize rheumatoid arthritis. Sci Rep 2020;10:1047.

89. Weiberg D, Thackeray JT, Daum G, Sohns JM, Kropf S, Wester HJ, *et al.* Clinical Molecular Imaging of Chemokine Receptor CXCR4 Expression in Atherosclerotic Plaque Using 68Ga-Pentixafor PET: Correlation with Cardiovascular Risk Factors and Calcified Plaque Burden. J Nucl Med 2018;59:266–72.

Conflicts of interest.—The authors certify that there is no conflict of interest with any financial organization regarding the material discussed in the manuscript. Authors' contributions.—François Jamar has given substantial contributions to the conception and the design of the manuscript, François Jamar, Lars C. Gormsen, Halil Yildiz, Riemer H. Slart, Kornelis S. van der Geest and Olivier Gheysens to acquisition, analysis and interpretation of the data. All authors have participated to drafting the manuscript, Olivier Gheysens and François Jamar revised it critically. All authors read and approved the final version of the manuscript.

History.—Manuscript accepted: June 13, 2022. - Manuscript revised: May 31, 2022. - Manuscript received: April 5, 2022.