

University of Groningen

Therapeutic drug monitoring using saliva as matrix

van den Elsen, Simone H. J.; Akkerman, Onno W.; Jongedijk, Erwin M.; Wessels, Mireille; Ghimire, Samiksha; van der Werf, Tjip S.; Touw, Daan J.; Bolhuis, Mathieu S.; Alffenaar, Jan-Willem C.

Published in:
European Respiratory Journal

DOI:
[10.1183/13993003.01903-2019](https://doi.org/10.1183/13993003.01903-2019)

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version
Final author's version (accepted by publisher, after peer review)

Publication date:
2020

[Link to publication in University of Groningen/UMCG research database](#)

Citation for published version (APA):

van den Elsen, S. H. J., Akkerman, O. W., Jongedijk, E. M., Wessels, M., Ghimire, S., van der Werf, T. S., Touw, D. J., Bolhuis, M. S., & Alffenaar, J.-W. C. (2020). Therapeutic drug monitoring using saliva as matrix: an opportunity for linezolid, but challenge for moxifloxacin. *European Respiratory Journal*, *55*(5), Article 1901903. <https://doi.org/10.1183/13993003.01903-2019>

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: <https://www.rug.nl/library/open-access/self-archiving-pure/taverne-amendment>.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): <http://www.rug.nl/research/portal>. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.



Early View

Research letter

Therapeutic drug monitoring using saliva as matrix: an opportunity for linezolid, but challenge for moxifloxacin

Simone H.J. van den Elsen, Onno W. Akkerman, Erwin M. Jongedijk, Mireille Wessels, Samiksha Ghimire, Tjip S. van der Werf, Daan J. Touw, Mathieu S. Bolhuis, Jan-Willem C. Alffenaar

Please cite this article as: van den Elsen SHJ, Akkerman OW, Jongedijk EM, *et al.* Therapeutic drug monitoring using saliva as matrix: an opportunity for linezolid, but challenge for moxifloxacin. *Eur Respir J* 2020; in press (<https://doi.org/10.1183/13993003.01903-2019>).

This manuscript has recently been accepted for publication in the *European Respiratory Journal*. It is published here in its accepted form prior to copyediting and typesetting by our production team. After these production processes are complete and the authors have approved the resulting proofs, the article will move to the latest issue of the ERJ online.

Therapeutic drug monitoring using saliva as matrix: an opportunity for linezolid, but challenge for moxifloxacin

Simone HJ van den Elsen^a, Onno W Akkerman^{b,c}, Erwin M Jongedijk^a, Mireille Wessels^a, Samiksha Ghimire^a, Tjip S van der Werf^{c,d}, Daan J Touw^a, Mathieu S Bolhuis^a, and Jan-Willem C Alffenaar^{a,e,f,g,#}.

^a University of Groningen, University Medical Center Groningen, Department of Clinical Pharmacy and Pharmacology, Groningen, The Netherlands.

^b University of Groningen, University Medical Center Groningen, Tuberculosis Center Beatrixoord, Haren, The Netherlands.

^c University of Groningen, University Medical Center Groningen, Department of Pulmonary Diseases and Tuberculosis, Groningen, The Netherlands.

^d University of Groningen, University Medical Center Groningen, Department of Internal Medicine, Groningen, The Netherlands.

^e Sydney Pharmacy School, Faculty of Medicine and Health, The University of Sydney, Sydney, Australia.

^f Westmead hospital, Sydney, Australia.

^g Marie Bashir Institute of Infectious Diseases and Biosecurity, Sydney, Australia.

corresponding author: J.W.C. Alffenaar, University of Sydney, Faculty of Medicine and Health, School of Pharmacy, Pharmacy Building A15, NSW 2006, Australia. Email:

johannes.alfenaar@sydney.edu.au

Take home message: Therapeutic drug monitoring using saliva as matrix is a suitable alternative for serum therapeutic drug monitoring of linezolid, but not for moxifloxacin due to a high variability in saliva-plasma ratios.

To the editor:

The World Health Organization (WHO) has listed moxifloxacin and linezolid among the preferred “Group A” drugs in the treatment of multidrug-resistant tuberculosis (MDR-TB).[1] Therapeutic drug monitoring (TDM) could potentially optimize MDR-TB therapy, since moxifloxacin and linezolid show large pharmacokinetic variability.[1–4] TDM of moxifloxacin focuses on identifying patients with low drug exposure who are at risk of treatment failure and acquired fluoroquinolone resistance.[5, 6] Alternatively, TDM of linezolid strives to reduce toxicity while ensuring an adequate drug exposure because of its narrow therapeutic index.[1, 3, 7]

TDM is typically performed using plasma or serum samples, but other biological matrices can be considered as alternatives (e.g. saliva).[8] A benefit of saliva is the easy and non-invasive nature of sampling. Especially in high TB burden areas the option for sampling at home would be advantageous. Penetration of moxifloxacin into saliva has typically been studied in healthy volunteers, but has never been evaluated for the purpose of TDM in MDR-TB patients.[9] Only one study described linezolid concentrations in saliva and found that saliva is a suitable matrix for TDM in MDR-TB patients and that salivary concentrations can be translated to serum concentrations without the need of a correction factor.[7] The aim of this prospective study was to explore the feasibility of saliva-based TDM of moxifloxacin and to determine if earlier results of linezolid in saliva of MDR-TB patients could be confirmed.

Hospitalized adult TB patients in the Tuberculosis Center Beatrixoord (Haren, The Netherlands), who had moxifloxacin or linezolid as part of their TB treatment and had routine TDM using blood samples were eligible for inclusion. All participants signed informed consent. This study was registered at Clinicaltrials.gov (NCT03080012) and approved by the ethical review committee of the University Medical Center Groningen (IRB 2016/069).

After at least 14 days of treatment, saliva samples were taken simultaneously with plasma (moxifloxacin) or serum (linezolid) according to routine TDM schedule which generally included a

sample before and 1, 2, 3, 4, and 8 h after drug administration. All samples were stored at -80°C until analysis.

To collect saliva samples, patients were asked to chew on a cotton roll after rinsing their mouth with water. Subsequently, the samples were processed using one of the following methods. Salivette (Sarstedt, Nümbrecht, Germany) in combination with centrifugation was used for non-contagious patients. Membrane filtration was applied to the saliva samples of the TB patients who still had *Mycobacterium tuberculosis* bacilli in their sputum to minimize infection hazard.[10] Salivary pH values were determined by two independent observers using pH indicator strips with pH range 4.0-7.0 and 2.0-9.0 (Merck KGaA, Darmstadt, Germany), because it could influence drug penetration into saliva.[9] Recovery of both sampling methods was determined similarly to Ghimire *et al* [11], except that moxifloxacin was tested at 1 and 3 mg/L and linezolid at 2 and 20 mg/L. Recovery was comparable for low and high concentrations. Using the Salivette, recovery was 48% (coefficient of variation [CV] 6%) for moxifloxacin or 95% (CV 3%) for linezolid and via membrane filtration 48% (CV 6%) or 98% (CV 3%), respectively. After analysis the salivary concentrations were corrected for recovery accordingly.

All samples were analysed using an updated version of our previously published liquid chromatography tandem mass spectrometry (LC-MS/MS) method.[12, 13] The LC-MS/MS method of linezolid was already cross-validated for saliva.[7] Cross-validation between plasma and saliva was performed for moxifloxacin at low (1 mg/L), medium (2 mg/L), and high (4 mg/L) concentrations as well as at lower limit of quantification (LLOQ; 0.05 mg/L). All concentration levels met the pre-set criteria for accuracy and precision (bias and CV <15%; at LLOQ both <20%).

Area under the concentration-time curve from 0 to 24 h (AUC_{0-24}) in saliva and plasma/serum was calculated using noncompartmental pharmacokinetic analysis (MWPharm version 3.82, Mediware, Groningen, The Netherlands). C_{max} was defined as highest observed concentration and T_{max} as corresponding time of C_{max} . Two different saliva-plasma/serum ratios were calculated; one used the paired drug concentrations, while the other compared AUC_{0-24} in both matrices. Passing-Bablok

regression and Bland-Altman plots (Analyze-it 4.81; Analyze-it Software Ltd., Leeds, United Kingdom) were used to analyse results.

Patient characteristics, pharmacokinetic parameters (C_{max} , T_{max} , AUC_{0-24}), and saliva-serum/plasma ratios are shown in Table 1. All patients on linezolid did also receive moxifloxacin. Individual linezolid concentration-time curves in saliva versus serum were similarly shaped and T_{max} in saliva was not delayed, which suggested that penetration of linezolid into saliva is fast. Passing-Bablok analysis showed a linear regression line of saliva concentration = $0.389 + 0.680 \cdot \text{serum concentration}$ with 95% confidence interval (CI) of intercept -0.14 to 1.06; 95% CI of slope 0.60 to 0.76, $r=0.954$, and $p=0.519$. Bland-Altman demonstrated a mean (95% CI) saliva-serum concentration ratio of 0.76 (0.70-0.82). In general, the linezolid saliva-serum paired concentration ratio was considerably constant at 0.6-0.8 (range 0.25-1.29). Saliva-serum AUC_{0-24} ratios were even less variable with a median of 0.81 (range 0.54-0.96). However, we found a lower saliva-serum ratio than before,[7] which could be caused by differences in sampling method, processing or storage. Because linezolid efficacy is related to the ratio of AUC_{0-24} to minimal inhibitory concentration (AUC_{0-24}/MIC), it is recommended to collect multiple saliva samples to calculate AUC_{0-24} in saliva and afterwards translate to plasma AUC_{0-24} using a correction factor of 1.2. Based on these results, salivary TDM of linezolid indeed might be feasible and is ready for testing in a high burdened TB setting. Although LC-MS/MS was used in our study, HPLC-UV could be a suitable alternative in less resourced settings. Simple point-of-care tests in saliva, centralized drug analysis, stability studies for transport at room temperature conditions, and cross-validation of existing analytical methods in saliva may improve feasibility.[8]

Moxifloxacin paired saliva-plasma concentration ratios were highly variable with a range of 0.15-2.81 (median 1.00) which does not favour saliva as a sampling matrix for TDM. Passing-Bablok showed a linear relation of saliva concentration = $-0.620 + 1.49 \cdot \text{serum concentration}$, 95% CI intercept -0.97 to -0.33, 95% CI slope 1.32 to 1.74, $r=0.796$, and $p=0.103$. As moxifloxacin saliva-plasma concentration ratios were not normally distributed according to Shapiro-Wilk test ($p=0.0003$), Bland-Altman

analysis could not be used. Unfortunately, saliva-plasma AUC_{0-24} ratios showed similar results (range 0.30-2.00), but the underlying cause remains unclear. Both inter-individual as well as intra-individual variation was observed. No effect of salivary pH on saliva-plasma ratios of moxifloxacin could be detected. A limitation of our study was that we did not measure the unbound concentrations. Therefore, variation in protein binding could have affected the saliva-plasma ratios. Interestingly, salivary concentrations higher than plasma concentrations were observed suggesting possibilities of active transport in addition to passive diffusion.[9] Moxifloxacin also shows excellent penetration into diseased lung tissue with a median free-tissue/free-serum ratio of 3.2.[14] It would be interesting to investigate whether salivary concentrations are related to tissue concentrations at the site of infection and if penetration is driven by similar mechanisms. If closely related, it might be possible to determine infection site moxifloxacin concentrations without the need of invasive tissue sampling. Clearly, it is the free drug concentrations at the site of action that is predictive of treatment efficacy, while plasma moxifloxacin concentrations serve not more than proxy markers.

Salivary TDM could be an alternative method for traditional linezolid TDM using plasma or serum, and future studies can focus on improving the feasibility. However, for moxifloxacin our data does not support saliva as suitable matrix for TDM using the described method. Future studies should investigate moxifloxacin protein binding, salivary flow, and transport mechanisms to gain more insight in the feasibility of moxifloxacin TDM in saliva. As shown before for amikacin [15], saliva will likely not be a universal but only a selective matrix for TDM of anti-TB drugs.

References

1. World Health Organization. WHO consolidated guidelines on drug-resistant tuberculosis treatment. 2019.
2. Pranger AD, van Altena R, Aarnoutse RE, van Soolingen D, Uges DRA, Kosterink JGW, van der Werf TS, Alffenaar JWC. Evaluation of moxifloxacin for the treatment of tuberculosis: 3 years of experience. *Eur. Respir. J.* 2011; 38: 888–894.
3. Bolhuis MS, Akkerman OW, Sturkenboom MGG, Ghimire S, Srivastava S, Gumbo T, Alffenaar J-WC. Linezolid-based Regimens for Multidrug-resistant Tuberculosis (TB): A Systematic Review to Establish or Revise the Current Recommended Dose for TB Treatment. *Clin. Infect. Dis.* 2018; 67: S327–S335.
4. Srivastava S, Peloquin CA, Sotgiu G, Migliori GB. Therapeutic drug management: is it the future of multidrug-resistant tuberculosis treatment? *Eur. Respir. J.* 2013; 42: 1449–1453.
5. Davies Forsman L, Bruchfeld J, Alffenaar J-WC. Therapeutic drug monitoring to prevent acquired drug resistance of fluoroquinolones in the treatment of tuberculosis. *Eur. Respir. J.* 2017; 49: 1700173.
6. Pranger AD, van der Werf TS, Kosterink JGW, Alffenaar JWC. The Role of Fluoroquinolones in the Treatment of Tuberculosis in 2019. *Drugs* 2019; 79: 161–171.
7. Bolhuis MS, van Altena R, van Hateren K, de Lange WCM, Greijdanus B, Uges DRA, Kosterink JGW, van der Werf TS, Alffenaar JWC. Clinical validation of the analysis of linezolid and clarithromycin in oral fluid of patients with multidrug-resistant tuberculosis. *Antimicrob. Agents Chemother.* 2013; 57: 3676–3680.
8. Ghimire S, Bolhuis MS, Sturkenboom MGG, Akkerman OW, de Lange WCM, van der Werf TS, Alffenaar J-WC. Incorporating therapeutic drug monitoring into the World Health

- Organization hierarchy of tuberculosis diagnostics. *Eur. Respir. J.* 2016; 47: 1867–1869.
9. van den Elsen SHJ, Oostenbrink LM, Heysell SK, Hira D, Touw DJ, Akkerman OW, Bolhuis MS, Alffenaar J-WC. Systematic Review of Salivary Versus Blood Concentrations of Antituberculosis Drugs and Their Potential for Salivary Therapeutic Drug Monitoring. *Ther. Drug Monit.* 2018; 40: 17–37.
 10. van den Elsen SHJ, van der Laan T, Akkerman OW, van der Zanden AGM, Alffenaar J-WC, van Soolingen D. Membrane Filtration Is Suitable for Reliable Elimination of Mycobacterium tuberculosis from Saliva for Therapeutic Drug Monitoring. *J. Clin. Microbiol.* 2017; 55: 3292–3293.
 11. Ghimire S, Maharjan B, Jongedijk EM, Kosterink JGW, Ghimire GR, Touw DJ, van der Werf TS, Shrestha B, Alffenaar J-WC. Evaluation of Saliva as a Potential Alternative Sampling Matrix for Therapeutic Drug Monitoring of Levofloxacin in Patients with Multidrug-Resistant Tuberculosis. *Antimicrob. Agents Chemother.* 2019; 63: e02379-18.
 12. Harmelink IM, Alffenaar JW, Wessels M, Greijdanus B, Uges D. A rapid and simple liquid chromatography-tandem mass spectrometry method for the determination of linezolid in human serum. *EJHP Sci.* 2008; 14: 3–7.
 13. Pranger AD, Alffenaar J-WC, Wessels AMA, Greijdanus B, Uges DRA. Determination of moxifloxacin in human plasma, plasma ultrafiltrate, and cerebrospinal fluid by a rapid and simple liquid chromatography- tandem mass spectrometry method. *J. Anal. Toxicol.* 2010; 34: 135–141.
 14. Heinrichs MT, Vashakidze S, Nikolaishvili K, Sabulua I, Tukvadze N, Bablshvili N, Gogishvili S, Little BP, Bernheim A, Guarner J, Peloquin CA, Blumberg HM, Derendorf H, Kempker RR. Moxifloxacin target site concentrations in patients with pulmonary TB utilizing microdialysis: a clinical pharmacokinetic study. *J. Antimicrob. Chemother.* 2018; 73: 477–483.

15. van den Elsen SHJ, Akkerman OW, Huisman JR, Touw DJ, van der Werf TS, Bolhuis MS, Alffenaar J-WC. Lack of penetration of amikacin into saliva of tuberculosis patients. *Eur. Respir. J.* 2018; 51: 1702024.

Table 1. Characteristics of the linezolid and moxifloxacin study populations, pharmacokinetic (PK) parameters in serum/plasma and saliva, and saliva-serum/plasma ratios using paired concentrations as well as AUC₀₋₂₄.

	Linezolid (n=7)	Moxifloxacin (n=15)
Study population		
Male [n(%)]	6 (86%)	11 (73%)
Age (years)	44 (37-55)	34 (25-55)
Bodyweight (kg)	67.1 (60.5-68.4)	67.1 (57.5-70.5)
Creatinine concentration (μmol/L)	73 (72-90)	(72 (63-90)
Dose (mg/kg)	8.85 (7.42-9.93)	5.96 (5.68-7.08)
Serum/plasma PK^a		
C _{max} (mg/L)	12.45 (8.84-15.78)	2.28 (1.62-2.80)
T _{max} (h)	3 (2-4)	2 (1-2)
AUC ₀₋₂₄ (mg*h/L)	119.4 (116.2-128.2)	21.3 (15.8-31.0)
Saliva PK		
C _{max} (mg/L)	7.93 (7.55-12.38)	3.20 (2.51-4.25)
T _{max} (h)	3 (2-3)	2 (1-2)
AUC ₀₋₂₄ (mg*h/L)	93.6 (91.7-108.0)	21.3 (13.7-28.3)
Saliva-serum/plasma ratio^a		
Paired concentration ratio	0.76 (0.64-0.85)	1.00 (0.68-1.35)
AUC ₀₋₂₄ ratio	0.81 (0.74-0.88)	0.89 (0.61-1.14)

All parameters are presented as median (interquartile range) unless stated otherwise.

^a Serum for linezolid and plasma for moxifloxacin.