



University of Groningen

General Assembly, Diagnosis, Definitions

Elkins, Jacob M; Kates, Stephen; Lange, Jeffrey; Lange, Jeppe; Lichstein, Paul; Otero, Jesse; Soriano, Alex; Wagner, Christof; Wouthuyzen-Bakker, Marjan

Published in: Journal of Arthroplasty

DOI: 10.1016/j.arth.2018.09.069

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version Publisher's PDF, also known as Version of record

Publication date: 2019

Link to publication in University of Groningen/UMCG research database

Citation for published version (APA): Elkins, J. M., Kates, S., Lange, J., Lange, J., Lichstein, P., Otero, J., Soriano, A., Wagner, C., & Wouthuyzen-Bakker, M. (2019). General Assembly, Diagnosis, Definitions: Proceedings of International Consensus on Orthopedic Infections. Journal of Arthroplasty, 34(2, Supp), S181-S185. https://doi.org/10.1016/j.arth.2018.09.069

Copyright Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: https://www.rug.nl/library/open-access/self-archiving-pure/taverneamendment.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): http://www.rug.nl/research/portal. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

The Journal of Arthroplasty 34 (2019) S181-S185



Contents lists available at ScienceDirect

The Journal of Arthroplasty

journal homepage: www.arthroplastyjournal.org

General Assembly, Diagnosis, Definitions: Proceedings of International Consensus on Orthopedic Infections



THE JOURNAL OF

Check for updates

Jacob M. Elkins¹, Stephen Kates², Jeffrey Lange¹, Jeppe Lange³, Paul Lichstein¹, Jesse Otero¹, Alex Soriano³, Christof Wagner², Marjan Wouthuyzen-Bakker³

ARTICLE INFO

Article history: Available online 19 October 2018

Keywords:

acute versus chronic periprosthetic joint infection (PII) time interval debridement, antibiotics, retention implant (DAIR) symptoms duration implant colonization implant-related infection host interaction immune response pathogenic organisms nonpathogenic organisms next-generation sequencing (NGS) sinus tract fistulogram ultrasound computed tomography magnetic resonance imaging

Question 1: What is the recommended time interval that would divide acute and chronic PJI (4 weeks, 90 days, etc.)?

Recommendation #1:

There is no evidence-based time interval that divides acute from chronic PJI. The natural history of infection is a continuum from initiation to chronicity. Surgical treatment for patients with infection should not solely be based on the duration of symptoms or the time from implantation of the prosthesis. Other factors also should be considered such as implant

stability, presence of sinus tract, virulence of the infective organism, and general health of the patient. It is important to note that the efficacy of surgical intervention, involving retention of the prosthesis, is more likely to fail as one moves more than 4 weeks from the index arthroplasty and/or duration of symptoms of infection.

Level of Evidence: Limited

Delegate Vote: Agree: 84%, Disagree: 15%, Abstain: 1% (Super Majority, Strong consensus)

Recommendation #2:

We recommend to move away from the traditional division between acute and chronic infection that was based solely on time from index arthroplasty or duration of symptoms. Periprosthetic infection is a continuum that leads to establishment of biofilm.

Level of Evidence: Limited

Delegate Vote: Agree: 60%, Disagree: 34%, Abstain: 6% (Super Majority, Weak Consensus)

Recommendation #3:

Should we have specific time limit cutoff between chronic and acute infection?

Delegate Vote: Agree: 60%, Disagree: 37%, Abstain: 3% (Super Majority, Weak Consensus)

Rationale:

According to the Oxford Advanced Learner's Dictionary, the term "acute" in case of illness is defined as "coming quickly to the most severe or critical stage," and the term "chronic" as "lasting for a long time, happening continually." In case of an acute periprosthetic joint infection (PJI), this would be translated in a sudden onset of severe joint pain and/or swelling in a prior symptom-free prosthetic joint, and in case of chronicity as the presence of mild or moderate pain in which its exact onset is hard to establish. In our opinion, this is the most accurate definition to differentiate acute from chronic PJIs and reflects the virulence of the microorganism(s) causing the infection. The reason that a certain time frame was subsequently introduced in the world of PJI to divide acute from chronic infections was primarily based on clinical grounds to identify those patients with a high and low success rate when treated with debridement, antibiotics, and retention of the implant (DAIR) [1–15]. One of the factors associated with DAIR failure is the presence of a mature biofilm in which embedded bacteria are

One or more of the authors of this paper have disclosed potential or pertinent conflicts of interest, which may include receipt of payment, either direct or indirect, institutional support, or association with an entity in the biomedical field which may be perceived to have potential conflict of interest with this work. For full disclosure statements refer to https://doi.org/10.1016/j.arth.2018.09.069.

Question 3. 2

Question 2.

³ Question 1.

unresponsive to antibiotic treatment because of multiple phenotypic and genotypic changes [16,17]. In such a condition, a PJI cannot be cured with antibiotics alone without removal of the implant. In which time frame a biofilm reaches maturity is not clear. In vitro studies indicate that biofilm already start to form within hours after inoculation of bacteria [18], but these experiments are performed under "optimal" circumstances for bacterial growth and do not include the complexity of the host's environment and the protective effect of its immune system [19]. Carli et al observed in a mouse model with a proximal tibial implant infection using a high initial bacterial inoculum (3 \times 10⁵ CFU) that a biofilm is evident after 2 weeks of injection but extends and is covered by fibrinous tissue and multiple host cells after 6 weeks [20]. A recent mouse model of knee PJI using a low-infecting inoculum of Staphylococcus aureus (10^3 CFU), which is similar to the expected inoculum during surgery [21], demonstrated that after a 2-week incubation period, antibiotics combinations including rifampicin were able to eradicate the infection [22]. These studies suggest that a mature biofilm develops within 2-6 weeks. However, the process of biofilm formation varies greatly among bacterial species, its inoculum, and the host [23,24]. Accordingly, it has been demonstrated that the efficacy of DAIR in acute infections is highest when the DAIR is performed as soon as possible after the onset of symptoms [25–36]. Moreover, it is important to note that, since the success of DAIR is determined by many factors, the decision to perform a DAIR procedure should not solely be based on symptom duration and/or time from index surgery in acute PJIs but should include hostrelated factors, causative microorganisms, and the stability of the implant. For this reason, we propose not to include a time interval in the definition of acute and chronic PJI because the natural history of an infection is a continuum from initiation to chronicity.

Question 2: What is the definition of implant "colonization" versus implant-related infection?

Recommendation:

Colonization is the presence of microbiota in a joint, with growth and multiplication of the organism, but without interaction between the organism and the host's immune response thus avoiding any clinical expression. Infection is the invasion of a joint by disease-causing organisms that results in an interplay with the host's immune response causing a clinical expression and disease state.

Level of Evidence: Limited

Delegate Vote: Agree: 83%, Disagree: 8%, Abstain: 9% (Super Majority, Strong Consensus)

Rationale:

Over the last few years, extensive research efforts have been invested in the diagnosis of implant-related infection or prosthetic joint infection (PJI) and numerous definitions have been proposed [37–39]. Infections result in an immune response, thus all definitions rely on a combination of clinical findings, laboratory results from peripheral blood and synovial fluid, microbiological data, histological evaluation of periprosthetic tissue, and intraoperative findings. The advancements in the field of diagnostics and statistics have allowed us to establish a validated, evidence-based definition for PJI as presented in another chapter.

On the other hand, research into colonization of a prosthetic joint implant is scarce, and currently, there is no universally accepted definition for implant colonization. Colonization and infection are two different processes. There are approximately ten times as many bacterial cells in the human flora as there are human cells in the body thus all multicellular organisms are colonized to some degree by extrinsic organisms. The human microbiome is the collection of all the microorganisms living in association with the human body. Microbiome and host form a complex relationship, where microorganisms can confer symbiotic benefits to the host in many key aspects of life [40]. However, defects in the regulatory circuits of the host-microbiome interaction may disturb this symbiotic relationship and promote disease [41]. The difference between an infection and colonization is often only a matter of circumstance. Nonpathogenic organisms can become pathogenic given specific conditions, and even the most virulent organism requires certain circumstances to cause a compromising infection.

Analysis using next-generation sequencing (NGS) has improved understanding of the microbiome [42,43]. Recent studies suggest the presence of microbiome in aseptic, deep tissue [43–45]. This is a fascinating discovery, as it suggests that microorganisms may inhabit organs previously thought to be sterile, given that they do not communicate with the outside world. In a recent study using NGS, an organism was identified in 6 of 17 patients undergoing primary arthroplasty, with no clinical or laboratory evidence of infection [46]. In another recent study, NGS frequently identified multiple organisms in an infected sample and the question remains whether these infections are the result of a single-dominant organism, or multiple pathogenic organisms [47]. This becomes of particular concern when considering that the majority of patients who fail treatment for infection are infected with a different organism [48,49].

As we forge new alliances in our quest to eliminate PJIs, we should also consider a call to new and mutually beneficial ways of coexisting with the microbial flora of the world. Novel molecular techniques for organism detection provide comprehensive information on the organisms occupying the joint and thus hold the promise for a better understanding of joint colonization.

Question 3: What is the definition of a sinus tract? Recommendation: A sinus tract has the following characteristics:

- 1) It is an abnormal channel through the soft tissues that allows communication between a joint prosthesis and the outside environment, known or presumed to be colonized by bacteria.
- 2) Its presence may be confirmed with direct visualization of an underlying prosthesis, evidence of communication with fistulogram, ultrasound, computed tomography, or MRI.

Level of Evidence: Consensus

Delegate Vote: Agree: 97%, Disagree: 2%, Abstain: 1% (Unanimous, Strongest Consensus)

Rationale:

The presence of a sinus tract communicating with a total joint arthroplasty (TJA) is one of the two major criteria for the diagnosis of proposed by the Musculoskeletal Infection Society and the International Consensus Meeting [37]. Therefore, consistently defining what constitutes a sinus tract in this context has significant implications for the appropriate diagnosis and treatment of PJI. Interestingly, there is a paucity of information in the arthroplasty literature that defines the characteristics of a periprosthetic sinus tract. Many investigations discuss the presence and subsequent surgical management of sinus tracts in the setting of knee and hip arthroplasties but do not provide consistent or detailed descriptions of the cutaneous pathology. Given the paucity of information and evidence, it is important to develop a comprehensive and standardized method for characterizing a soft tissue sinus tract surrounding a total joint prosthesis.

A sinus tract (latin: hollow, cavity) is an abnormal channel connecting a cavity lined with granulation tissue to an epithelial surface [50]. Although a fistula and a sinus tract are technically separate entities, with the former representing an abnormal connecting channel between two epithelialized cavities specifically [50], they are frequently grouped together.

Given the relationship between infection and the development of sinus tracts and vice versa, it is not surprising that there exists a rich accounting of draining wounds and sinus tracts throughout medical history. In fact, a likely description of a draining sinus tract, secondary to chronic shoulder infection and osteomyelitis, is included in the Edwin-Smith Papyrus [51], the oldest surgical treatise in existence. Centuries later, Hippocrates [52] would provide various descriptions of sinus tracts and fistulae and extensive options for remedies, including topical, oral, and surgical.

However, perhaps, the most important of the historical treatments of sinus tracts comes from the Chirurgical Treatises of Richard Wiseman, c. 1686 [53]. In his chapter titled "On Fistulae," which appears in the appendix to his treatise on gunshot wounds, Wiseman describes a fistula as a sinuous ulcer, which has actively been draining for at least 2-3 months. He associates the draining sinus fistula with a "long pipe of skin," and the presence of "callus," which has been "hastened by the transpiration and resolution of the thin and subtill humours." Like Hippocrates, Wiseman advocated for treatment with either medications or surgical debridement. Of note, Wiseman specifically commented on the particular difficulty of curing sinus tracts associated with joints. Since Wiseman, there have been numerous additional descriptions of sinus tracts associated with bones and joints. However, one of the particular interests to the field of arthroplasty dates from the early 1700s [54]. Johanne Daniele Schlichting describes a case report, from 1730, of a 14-year-old girl suffering from disability due to a hip infection associated with a large draining sinus tract. Schlichting also describes his method of treatment, including removal of the femoral head, and in doing so, provided the first report of a proximal femoral resection in the medical literature. Throughout surgical history, a sinus tract has been pathognomonic for deep infection. The same is true in TJA, but the terms of the definition have not been established.

Sinus tracts are currently synonymous with PJI [55]. Fistulas in TJA have been noted to form connections between the prosthesis and vascular channels [56], the ureter [57], bladder [58,59], colon [60], rectum [61], and vagina [55], and are clearly a risk for the development of PJI when associated with bacterially colonized cavities. In addition, there is little information differentiating a communication that originates from inside the joint versus outside the joint.

There has been a significant amount of effort spent in determining the yield of culture samples from sinus tracts and fistulas originating from or terminating at joint arthroplasties [56,61–68]. Although this has provided insight as to the utility of sinus content cultures in the diagnosis of the responsible pathogens, it has not further assisted in defining the pathology. For the purposes of PJI diagnosis, we suggest that sinus tracts and fistulas communicating with bacterially colonized areas be grouped together, regardless of origin from within the joint or not, to fulfill the major criterion for the diagnosis of PJI.

The majority of information regarding the definition of a sinus tract in the presence of musculoskeletal infection has been studied in the context of osteomyelitis. There are multiple classification systems for sinus tracts, with varying degrees of focus on associated soft tissue compromise. The Cierny-Mader classification is perhaps the most commonly referenced system and involves categorical divisions staged by combining anatomic class (I-medullary, II-superficial, III-localized, and IV-diffuse) and host physiologic class (A-normal immune function, B-local or systemic immune compromise, and C-treatment worse than disease) [69]. A sinus tract leading to an exposed bone is the hallmark of stage II (superficial) osteomyelitis and occurs on a continuum with stage III and IV disease. Although further details of sinus tract characteristics aside from direct contact with osseous structures are not included, treatment with thorough debridement is consistently advocated [69,70]. Conceptually similar to the anatomic class used by Cierny and Mader, Ger proposed a classification system in 1984 that focused on the wound, separating simple sinus, chronic superficial ulcer, multiple sinuses, and multiple skin-lined sinuses [64]. Similarly, these pathologic conduits tunneled directly to bone. Currently, no analogous method is used to characterize sinus tracts associated with PJI. However, a patent channel through soft tissue connecting the outside environment directly to a total joint prosthesis should be considered a sinus tract.

Chronicity of drainage and of associated symptoms is an important consideration. Although it has been noted that postoperative wound drainage lasting longer than 5-7 days is unlikely to remit without intervention [62], differentiating between simple prolonged postoperative drainage and early sinus tract formation is difficult. Galat et al [63] reviewed the records of over 17,000 primary total knee arthroplasties and identified a 5.3%-6.0% risk of deep infection in knees with persistent wound drainage within a 30-day postoperative time frame. However, "surgeon judgment" rather than objective testing played a significant role in the diagnosis of deep infection in many cases and may have skewed results. Another series of over 11,000 arthroplasty procedures identified 300 patients who developed wound drainage lasting >48 hours after surgery [65]. Although persistent wound drainage was noted to cease in the majority of patients between postoperative days 2 and 4, 28% continued to drain and underwent further surgery. Surgical debridement was adequate to resolve the wound issues in the majority of cases, but 20% required additional intervention in the form of two-stage exchange, resection arthroplasty, or antibiotic suppression. In this series, the mean interval between the onset of drainage and surgical treatment was 10 days in patients who required further intervention. Other studies have suggested that drainage for more than 5 days imparts a 12.5 times greater risk of developing infection [71] and that each day of continued drainage increases the risk of wound infection by 42% in hips and 29% in knees [72]. However, these studies do not subdivide the portion of superficial wound infections that progress to true PII. In addition, surgery on a draining wound performed after 12 days of continuous drainage was noted to yield positive cultures in only 25% of cases [73]. Although the distinction between persistent wound drainage and a developed sinus tract is not defined in the acute setting after surgery, there is likely a time after which persistent drainage should be deemed a sinus tract. Currently, there is no evidence to guide us, to our knowledge, in understanding this distinction. Regardless of the definition, persistent drainage in any form is clearly concerning for PJI.

There is a strong association between chronically draining wound sinus tracts and deep infection of prosthetic hip and knee joints [2]. However, it is important to draw a distinction between the presence of a sinus tract *de facto* as a diagnostic criterion for PJI and the utility of sinus tract cultures in guiding infection treatment. Wound sinus cultures for osteomyelitis have notoriously low sensitivity and specificity [68,74,75]. The same has proven true for deep prosthetic joint infection. Two studies have been conducted to determine the correlation between superficial cultures from wounds or draining sinus tracts and a deep pathogen in the setting of prosthetic joint infection. Cune et al evaluated the usefulness of wound culture results in the treatment of acute postoperative prosthetic joint infection. Authors found 80.3% agreement between superficial and deep surgical cultures in this setting with high sensitivity and specificity for Staphylococcus aureus and gramnegative bacilli [76]. Tetreault et al performed a similar analysis comparing superficial and deep cultures in patients with deep prosthetic joint infection. Their results showed a 47.3% concordance between superficial and deep cultures, and in 41.8% of cases, the superficial organism wound has guided therapy with a different antibiotic than deep cultures [77]. There is likely a gradient of organisms within a sinus tract community, but the biology of the sinus tract microenvironment has not yet been studied. Therefore, although the presence of a sinus tract should be considered equivalent to a deep prosthetic joint infection, cultures of the fluid cannot be relied on to guide treatment.

In general, for the diagnosis of PJI, a sinus tract should demonstrate clear communication between the prosthesis and a nonsterile environment. The most obvious method is to directly visualize the underlying prosthesis through the lumen of the sinus or directly access the prosthesis with a sterile probe. However, to corroborate physical examination findings or evaluate a suspicious channel, various imaging methodologies may be utilized to confirm the presence of a true sinus tract that communicates with a total joint arthroplasty. Conventional radiography may be helpful in identifying areas concerning for infection with a sinus tract in combination with subcutaneous or intra-articular gas. However, plain X-rays may be negative in more than 50% of cases and may be of minimal diagnostic utility in acute infection [78]. Instead, conventional X-ray with the addition of arthrography or fistulography may drastically increase the diagnostic yield by illuminating infectious channels and accumulations [4,79]. Traditionally, more advanced imaging modalities such as computed tomography and magnetic resonance imaging were believed to be of limited use in evaluating the soft tissues immediately around a total joint prosthesis due to large amounts of metal artifact and image distortion. Recent developments, including metal artifact reduction sequence magnetic resonance imaging and three-dimensional reconstruction, allow for a much more detailed evaluation of periarticular structures and the presence of sinus tracts. However, given the dynamic nature of soft tissues and underlying infection, imaging studies may not provide sufficient evidence to verify the existence of a sinus tract as these may fluctuate in their patency and extent. Therefore, imaging modalities should not solely be relied on for the identification of a sinus communicating with a joint prosthesis.

In summary, an established sinus tract or fistulous connection between a deep prosthetic joint and another space known to be colonized with pathogenic microorganisms should be considered tantamount to deep prosthetic infection. Although the literature does not provide clear guidelines regarding the time at which a draining wound becomes a sinus tract, it is clear that prolonged drainage from an arthroplasty wound increases the likelihood that deep infection will occur. Although literature does not support the use of superficial sinus cultures to guide treatment of deep prosthetic joint infection, clinicians should rely on the presence of a sinus to justify surgical treatment. Therefore, any suspected connection between a deep prosthetic joint and an area colonized by pathogenic microorganisms should be considered seriously and evaluated thoroughly.

References

- Coventry MB. Treatment of infections occurring in total hip surgery. Orthop Clin North Am 1975;6:991–1003.
- [2] Fitzgerald Jr RH, Nolan DR, Ilstrup DM, Van Scoy RE, Washington JA 2nd, Coventry MB. Deep wound sepsis following total hip arthroplasty. J Bone Joint Surg Am 1977;59:847–55.
- [3] Toms AD, Davidsom D, Masri BA, Duncan CP. The management of periprosthetic infection in total joint arthroplasty. J Bone Joint Surg Br 2006;88: 149–55.
- [4] Zimmerli W, Ochsner PE. Management of infection associated with prosthetic joints. Infection 2003;31:99–108.
- [5] Tsukayama DT, Estrada R, Gustilo RB. Infection after total hip arthroplasty. A study of the treatment of one hundred and six infections. J Bone Joint Surg Am 1996;78:512–23.
- [6] McPherson EJ, Woodson C, Holtom P, Roidis N, Shufelt C, Patzakis M. Periprosthetic total hip infection: outcomes using a staging system. Clin Orthop Relat Res 2002:8–15.
- [7] Zimmerli W, Widmer AF, Blatter M, Frei R, Ochsner PE. Role of rifampin for treatment of orthopedic implant-related staphylococcal infections: a

randomized controlled trial. Foreign-Body Infection (FBI) Study Group. JAMA 1998;279:1537-41.

- [8] Cierny III G, DiPasquale D. Periprosthetic total joint infections: staging, treatment, and outcomes. Clin Orthop Relat Res 2002:23–8.
- [9] Maillet M, Pavese P, Bruley D, Seigneurin A, François P. Is prosthesis retention effective for chronic infections in hip arthroplasties? A systematic literature review. Eur J Clin Microbiol Infec Dis 2015;34:1495–502.
- [10] Barberan J, Águilar L, Carroquino G, Giménez MJ, Sánchez B, Martínez D, et al. Conservative treatment of staphylococcal prosthetic joint infections in elderly patients. Am J Med 2006;119. 993.e7-993.e10.
- [11] Betsch BY, Eggli S, Siebenrock KA, Täuber MG, Mühlemann K. Treatment of joint prosthesis infection in accordance with current recommendations improves outcome. Clin Infect Dis 2008;46:1221–6.
- [12] Westberg M, Grøgaard B, Snorrason F. Early prosthetic joint infections treated with debridement and implant retention. Acta Orthop 2012;83:227–32.
- [13] Geurts JA, Janssen DM, Kessels AG, Walenkamp GH. Good results in postoperative and hematogenous deep infections of 89 stable total hip and knee replacements with retention of prosthesis and local antibiotics. Acta Orthop 2013;84:509–16.
- [14] Odum SM, Fehring TK, Lombardi AV, Zmistowski BM, Brown NM, Luna JT, et al. Irrigation and debridement for periprosthetic infections: does the organism matter? J Arthroplasty 2011;26(6 Suppl):114–8.
- [15] Fehring TK, Odum SM, Berend KR, Jiranek WA, Parvizi J, Bozic KJ, et al. Failure of irrigation and debridement for early postoperative periprosthetic infection. Clin Orthop Relat Res 2013;471:250–7.
- [16] Lebeaux D, Ghigo JM, Beloin C. Biofilm-related infections: bridging the gap between clinical management and fundamentel aspects of recalcitrance toward antibiotics. Microbiol Mol Biol Rev 2014;78:510–43.
- [17] Davies D. Understanding biofilm resistance to antibacterial agents. Nat Rev Drug Discov 2003;2:114–22.
- [18] Veerachamy S, Yarlagadda T, Manivasagam G, Yarlagadda PK. Bacterial adherence and biofilm formation on medical implants: a review. Proc Inst Mech Eng H 2014;228:1083–99.
- [19] Bandyk DF, Kinney EV, Riefsnyder TI, Kelly H, Towne JB. Treatment of bacteriabiofilm graft infection by in situ replacement in normal and immune-deficient states. J Vasc Surg 1993;18:398–405.
- [20] Carli AV, Bhimani S, Yang X, Shirley MB, de Mesy Bentley KL, Ross FP, et al. Quantification of peri-implant bacterial load and in vivo biofilm formation in an innovative, clinically representative mouse model of periprosthetic joint infection. J Bone Joint Surg Am 2017;99:e25.
- [21] Menzies BE, Kourteva Y, Kaiser AB, Kernodle DS. Inhibition of staphylococcal wound infection and potentiation of antibiotic prophylaxis by a recombinant fragment of the fibronectin-binding protein of Staphylococcus aureus. J Infect Dis 2002;185:937–43.
- [22] Thompson JM, Saini V, Ashbaugh AG, Miller RJ, Ordonez AA, Ortines RV, et al. Oral-only linezolid-rifampin is highly effective compared with other antibiotics for periprosthetic joint infection: study of a mouse model. J Bone Joint Surg Am 2017;99:656–65.
- [23] Lovati AB, Bottagisio M, Vecchi de E, Gallazzi E, Drago L. Animal models of implant-related low-grade infections. A twenty year review. Adv Exp Med Biol 2017;971:29–50.
- [24] Vidlak D, Kielian T. Infectious dose dictates the host response during Staphylococcus aureus orthopedic-implant biofilm infection. Infect Immun 2016;84(7):1957–65.
- [25] Grammatopoulos G, Bolduc ME, Atkins BL, Kendrick BJL, McLardy-Smith P, Murray DW, et al. Functional outcome of debridement, antibiotics and implant retention in periprosthetic joint infection involving the hip: a casecontrol study. Bone Joint J 2017;99-B:614–22.
- [26] Urish KL, Bullock AG, Kreger AM, Shah NB, Jeong K, Rothenberger SD. A multicenter study of irrigation and debridement in total knee arthroplasty periprosthetic joint infection: treatment failure is high. J Arthroplasty 2018;33:1154–9.
- [27] Koh IJ, Han SB, In Y, Oh KJ, Lee DH, Kim TK. Open debridement and prosthesis retention is a viable treatment option for acute periprosthetic joint infection after total knee arthroplasty. Arch Orthop Trauma Surg 2015;135: 847–55.
- [28] Triantafyllopoulos GK, Poultsides LA, Sakellariou VI, Zhang W, Sculco PK, Ma Y, et al. Irrigation and debridement for periprosthetic infections of the hip and factors determining outcome. Int Orthop 2015;39:1203–9.
- [29] Triantafyllopoulos GK, Poultsides LA, Zhang W, Sculco PK, Ma Y, Sculco TP. Periprosthetic knee infections treated with irrigation and debridement: outcomes and preoperative predictive factors. J Arthroplasty 2015;30:649–57.
- [30] Kuiper JW, Vos SJ, Saouti R, Vergroesen DA, Graat HC, Debets-Ossenkopp YJ, et al. Prosthetic joint-associated infections treated with DAIR (debridement, antibiotics, irrigation, and retention): analysis of risk factors and local antibiotic carriers in 91 patients. Acta Orthop 2013;84:380–6.
- [31] Marculescu CE, Berbari EF, Hanssen AD, Steckelberg JM, Harmsen SW, Mandrekar JN, et al. Outcome of prosthetic joint infections treated with debridement and retention of components. Clin Infect Dis 2006;142:471–8.
- [32] Buller LT, Sabry FY, Easton RW, Klika AK, Barsoum WK. The preoperative prediction of success following irrigation and debridement with polyethylene exchange for hip and knee prosthetic joint infections. J Arthroplasty 2012;27:857–64.
- [33] Hsieh PH, Lee MS, Hsu KY, Chang YH, Shih HN, Ueng SW, et al. Gram-negative prosthetic joint infections: risk factors and outcome of treatment. Clin Infect Dis 2009;49:1036–43.

- [34] Crockarell JR, Hanssen AD, Osmon DR, Morrey BF. Treatment of infection with debridement and retention of the components following hip arthroplasty. J Bone Joint Surg Am 1998;80:1306–13.
- [35] Brandt CM, Sistrunk WW, Duffy MC, Hanssen AD, Steckelberg JM, Ilstrup DM, et al. Staphylococcus aureus prosthetic joint infection treated with debridement and prosthesis retention. Clin Infect Dis 1997;24:914–9.
- [36] Tattevin P, Cremieux A-C, Pottier P, Huten D, Carbon C. Prosthetic joint infection: when can prosthesis salvage Be considered? Clin Infect Dis 1999;29:292–5.
- [37] Parvizi J, Zmistowski B, Berbari EF, Bauer TW, Springer BD, Della Valle CJ, et al. New definition for periprosthetic joint infection: from the workgroup of the musculoskeletal infection society. Clin Orthop Relat Res 2011;469:2992–4. https://doi.org/10.1007/s11999-011-2102-9.
- [38] Osmon DR, Berbari EF, Berendt AR, Lew D, Zimmerli W, Steckelberg JM, et al. Executive summary: diagnosis and management of prosthetic joint infection: clinical practice guidelines by the Infectious Diseases Society of America. Clin Infect Dis 2013;56:1–10. https://doi.org/10.1093/cid/cis966.
- [39] Parvizi J, Gehrke T, Chen AF. Proceedings of the international Consensus on periprosthetic joint infection. Bone Joint J 2013;95-B:1450–2. https://doi.org/ 10.1302/0301-620X.95B11.33135.
- [40] Jones S. Symbiosis: who does what in the microbiome? Nat Rev Microbiol 2008;6:256–7. https://doi.org/10.1038/nrmicro1880.
- [41] Eloe-Fadrosh EA, Rasko DA. The human microbiome: from symbiosis to pathogenesis. Annu Rev Med 2013;64:145–63. https://doi.org/10.1146/ annurev-med-010312-133513.
- [42] The placenta harbors a unique microbiome. Sci Transl Med 2014;6:237ra65. https://doi.org/10.1126/scitranslmed.3008599.
- [43] Hieken TJ, Chen J, Hoskin TL, Walther-Antonio M, Johnson S, Ramaker S, et al. The microbiome of aseptically collected human breast tissue in benign and malignant disease. Sci Rep 2016;6:30751. https://doi.org/10.1038/srep30751.
- [44] Urbaniak C, Gloor GB, Brackstone M, Scott L, Tangney M, Reid G. The microbiota of breast tissue and its association with breast cancer. Appl Environ Microbiol 2016;82:5039–48. https://doi.org/10.1128/AEM.01235-16.
- [45] Urbaniak C, Cummins J, Brackstone M, Macklaim JM, Gloor GB, Baban CK, et al. Microbiota of human breast tissue. Appl Environ Microbiol 2014;80:3007–14. https://doi.org/10.1128/AEM.00242-14.
- [46] Tarabichi M, Shohat N, Goswami K, Alvand A, Silibovsky R, Belden K, et al. Diagnosis of periprosthetic joint infection: the potential of next-generation sequencing. J Bone Joint Surg Am 2018;100:147–54. https://doi.org/10. 2106/JBJS.17.00434.
- [47] Tarabichi M, Shohat N, Goswami K, Parvizi J. Can next generation sequencing play a role in detecting pathogens in synovial fluid? Bone Joint J 2018;100-B: 127–33. https://doi.org/10.1302/0301-620X.100B2.BJJ-2017-0531.R2.
- [48] Mittal Y, Fehring TK, Hanssen A, Marculescu C, Odum SM, Osmon D. Twostage reimplantation for periprosthetic knee infection involving resistant organisms. J Bone Joint Surg Am 2007;89:1227–31. https://doi.org/10.2106/ JBJS.E.01192.
- [49] Zmistowski B, Tetreault MW, Alijanipour P, Chen AF, Della Valle CJ, Parvizi J. Recurrent periprosthetic joint infection: persistent or new infection? J Arthroplasty 2013;28:1486–9. https://doi.org/10.1016/j.arth.2013.02.021.
- [50] Full text of "Bailey & Love's Short Practice Of Surgery 26 E. CRC Press; 2013. https://archive.org/stream/WilliamsBaileyAndLovesShortPracticeOfSurgery NormanWilliamsChristopherBulstrodePR/%28Williams%2C+Bailey+and+Love %27s+Short+Practice+of+Surgery%29+Norman+Williams%2C+Christopher+ Bulstrode%2C+P+Ronan+0%27Connell-Bailey+%26+Love%27s+Short+Practice+ of+Surgery+26E-CRC+Press+%282013%29_ujvu.txt [accessed 9.08.18].
- [51] The Edwin Smith Surgical Papyrus, Volume 1: Hieroglyphic Transliteration, Translation, and Commentary. The Oriental Institute of the University of Chicago, https://oi.uchicago.edu/research/publications/oip/edwin-smith-surgicalpapyrus-volume-1-hieroglyphic-transliteration [accessed 9.08.18].
- [52] Soliman F, Sturgeon G, Hargest R. Revisiting an ancient treatment for transphincteric fistula-in-ano 'There is nothing new under the sun' Ecclesiastes 1v9. J R Soc Med 2015;108:482–9. https://doi.org/10.1177/0141076815588322.
- [53] Russell KF. Richard wiseman and his several chirurgical treatises. ANZ J Surg 1940;9:223-7. https://doi.org/10.1111/j.1445-2197.1940.tb06713.x.
- [54] Schlichting J. Observationes Variae Medico-Chirurgicae a Johanne Daniele Schlichting, Med. & Chir. Doctore, Acad. Caesareo-Leopoldin. Nat. Curios. Membro, & Commercii Literarii Norimberg, London: Socio. Royal Society of London; 1753.
- [55] Palmer SW, Luu HH, Finn HA. Hip-vagina fistula after acetabular revision. J Arthroplasty 2003;18:533–6.
- [56] Guyard M, Vaz G, Aleksic I, Guyen O, Carret J-P, Béjui-Hugues J. Aspergillar prosthetic hip infection with false aneurysm of the common femoral artery

and cup migration into the pelvis. Rev Chir Orthop Reparatrice Appar Mot 2006;92:606–9.

- [57] Schäfer D, Mattarelli G, Morscher E. Ureteroarticular fistula after total hip replacement. A case report. Arch Orthop Trauma Surg 1994;114:35–6.
- [58] Jones ALC, Acher P, Cynk M. Vesico-acetabular cutaneous fistula: a delayed complication of hip surgery. Urology 2011;78:323-4. https://doi.org/10.1016/ j.urology.2010.06.007.
- [59] Russell RD, Incavo SJ, Mineo MT, Dinh T. Vesicoacetabular fistula in a chronically infected total hip arthroplasty. J Arthroplasty 2010;25. https://doi.org/ 10.1016/ji.arth.2009.04.017. 659.e9-659.e12.
- [60] Long SS, Tawa NE, Ayres DK, Abdeen A, Wu JS. Coloarticular fistula: a rare complication of revision total hip arthroplasty. Radiol Case Rep 2011;6:533. https://doi.org/10.2484/rcr.v6i3.533.
- [61] Bach CM, Nogler M, Wimmer C, Stoeckel B, Ogon M. Fistula between a total hip arthroplasty and the rectum: a case report. Clin Orthop Relat Res 2001: 143–6.
- [62] Dennis DA. Wound complications in total knee arthroplasty. Instr Course Lect 1997;46:165–9.
- [63] Galat DD, McGovern SC, Larson DR, Harrington JR, Hanssen AD, Clarke HD. Surgical treatment of early wound complications following primary total knee arthroplasty. J Bone Joint Surg Am 2009;91:48–54. https://doi.org/10.2106/ JBJS.G.01371.
- [64] Ger R. Muscle transposition for treatment and prevention of chronic posttraumatic osteomyelitis of the tibia. J Bone Joint Surg Am 1977;59:784–91.
- [65] Jaberi FM, Parvizi J, Haytmanek CT, Joshi A, Purtill J. Procrastination of wound drainage and malnutrition affect the outcome of joint arthroplasty. Clin Orthop Relat Res 2008;466:1368–71. https://doi.org/10.1007/s11999-008-0214-7.
- [66] Lauschke FH, Frey CT. Hematogenous osteomyelitis in infants and children in the northwestern region of Namibia. Management and two-year results. J Bone Joint Surg Am 1994;76:502–10.
- [67] Levine SE, Esterhai JL, Heppenstall RB, Calhoun J, Mader JT. Diagnoses and staging. Osteomyelitis and prosthetic joint infections. Clin Orthop Relat Res 1993:77–86.
- [68] Mackowiak PA, Jones SR, Smith JW. Diagnostic value of sinus-tract cultures in chronic osteomyelitis. JAMA 1978;239:2772–5.
- [69] Cierny G, Mader JT. Adult chronic osteomyelitis. Orthopedics 1984;7: 1557–64. https://doi.org/10.3928/0147-7447-19841001-07.
- [70] Cierny G, Mader JT, Penninck JJ. A clinical staging system for adult osteomyelitis. Clin Orthop Relat Res 2003:7–24. https://doi.org/10.1097/01.blo. 0000088564.81746.62.
- [71] Saleh K, Olson M, Resig S, Bershadsky B, Kuskowski M, Gioe T, et al. Predictors of wound infection in hip and knee joint replacement: results from a 20 year surveillance program. J Orthop Res 2002;20:506–15. https://doi.org/10.1016/ S0736-0266(01)00153-X.
- [72] Patel VP, Walsh M, Sehgal B, Preston C, DeWal H, Di Cesare PE. Factors associated with prolonged wound drainage after primary total hip and knee arthroplasty. J Bone Joint Surg Am 2007;89:33–8. https://doi.org/10.2106/ JBJS.F.00163.
- [73] Weiss AP, Krackow KA. Persistent wound drainage after primary total knee arthroplasty. J Arthroplasty 1993;8:285–9.
- [74] Patzakis MJ, Wilkins J, Kumar J, Holtom P, Greenbaum B, Ressler R. Comparison of the results of bacterial cultures from multiple sites in chronic osteomyelitis of long bones. A prospective study. J Bone Joint Surg Am 1994;76: 664-6.
- [75] Perry CR, Pearson RL, Miller GA. Accuracy of cultures of material from swabbing of the superficial aspect of the wound and needle biopsy in the preoperative assessment of osteomyelitis. J Bone Joint Surg Am 1991;73:745–9.
- [76] Cuñé J, Soriano A, Martínez JC, García S, Mensa J. A superficial swab culture is useful for microbiologic diagnosis in acute prosthetic joint infections. Clin Orthop Relat Res 2009;467:531–5. https://doi.org/10.1007/s11999-008-0553-4.
- [77] Tetreault MW, Wetters NG, Aggarwal VK, Moric M, Segreti J, Huddleston JI, et al. Should draining wounds and sinuses associated with hip and knee arthroplasties be cultured? J Arthroplasty 2013;28:133–6. https://doi.org/ 10.1016/j.arth.2013.04.057.
- [78] Tigges S, Stiles RG, Roberson JR. Appearance of septic hip prostheses on plain radiographs. AJR Am J Roentgenol 1994;163:377–80. https://doi.org/10.2214/ ajr.163.2.8037035.
- [79] Jain CU, Yang DC, Patel DM, Gudi KA, Giovanniello J. Cutaneous fistula communicating with the hip in a patient with a painful total hip prosthesis. Demonstration by radionuclide arthrography. Clin Nucl Med 1988;13: 820–2.