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### Carbon and nitrogen stable isotopes of well-preserved Middle Pleistocene bone collagen from Schöningen (Germany) and their paleoecological implications





Margot Kuitems <sup>a, \*</sup>, Johannes van der Plicht <sup>a, b</sup>, Dorothée G. Drucker <sup>c</sup>, Thijs Van Kolfschoten <sup>a</sup>, Sanne W.L. Palstra <sup>b</sup>, Hervé Bocherens <sup>c, d</sup>

<sup>a</sup> Faculty of Archaeology, Leiden University, Reuvensplaats 3-4, 2311 BE Leiden, The Netherlands

<sup>b</sup> Center for Isotope Research, Groningen University, Nijenborgh 4, 9747 AG Groningen, The Netherlands

<sup>c</sup> Department of Geosciences, Biogeology, University of Tübingen, Hölderlinstr. 12, 72074, Tübingen, Germany

<sup>d</sup> Senckenberg Center for Human Evolution and Palaeoecology, Hölderlinstr. 12, 72074 Tübingen, Germany

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### ABSTRACT

Carbon and nitrogen stable isotopes in bone collagen can provide valuable information about the diet and habitat of mammal species. However, bone collagen degrades in normal circumstances very rapidly, and isotope analyses are therefore usually restricted to fossil material with a Late Pleistocene or Holocene age. The Middle Pleistocene site of Schöningen, dated to around 300,000 years ago, yielded bones and teeth with an exceptionally good state of collagen preservation. This allowed us to measure reliable biogenic carbon and nitrogen stable isotope ratios for different herbivorous taxa from the families Elephantidae, Rhinocerotidae, Equidae, Cervidae, and Bovidae. The results provide insights regarding the paleoenvironmental setting in which Middle Pleistocene hominins operated. The vegetation consumed by the herbivores from the famous spear horizon originates from open environments. During the climatic Reinsdorf Interglacial optimum, the landscape seems to have been relatively open as well, but certainly included parts that were forested. The results also indicate some niche partitioning; different herbivore species used different plant resources. For instance, the horses seem to have been predominantly browsers, while the straight-tusked elephants were feeding chiefly on grass.

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### Introduction

The stable isotope (<sup>13</sup>C, <sup>15</sup>N) signal of fossil animal bones is a powerful tool for investigating ancient environments, ecosystems, and the reconstruction of the diet and ecological preferences of species (e.g., Bocherens et al., 1991, 1999, 2011; lacumin et al., 1997, 2000; Coltrain et al., 2004; Drucker et al., 2008; Fox-Dobbs et al., 2008). These kinds of isotopic analyses are based on the principle that the organic tissues of an organism (bone collagen and teeth) reflect the chemical composition of the food and water the organism ingested during its life (Kohn, 1999).

However, bone collagen molecules tend to break down rapidly after the animal dies, and all kinds of taphonomic processes can lead to degradation of the collagen. Since bone collagen can be

\* Corresponding author.

E-mail address: m.kuitems@arch.leidenuniv.nl (M. Kuitems).

sensitive to degradation, the majority of stable isotope analyses using bone collagen have been carried out on relatively young material with a Late Pleistocene age (usually within the radiocarbon time scale, i.e., up to 50 ka [thousands of years ago]; e.g., Drucker et al., 2003; Richards and Hedges, 2003; Stevens and Hedges, 2004; Szpak et al., 2010). Reliable results from older material have been published but are very rare (e.g., Bocherens et al., 1999; Jones et al., 2001; Palmqvist et al., 2008; Britton et al., 2012).

Most of the Middle Pleistocene fossils from Schöningen were macroscopically very well preserved, while the preservation at the microscopic level needed further investigation. To determine if the bone collagen was well preserved enough to retrieve reliable stable carbon and nitrogen isotopic data to decipher the environmental conditions hominins experienced at Schöningen, a pilot study was executed in 2009 at Leiden University. Based on the very promising results (Van der Plicht et al., 2011), further isotopic research on material from the site of Schöningen was conducted in a joint project with the University of Tübingen.

## The Schöningen locality and the stratigraphic position of the different assemblages

The Quaternary deposits exposed in the lignite quarry east of the village of Schöningen during the past two decades have yielded a large amount of Paleolithic material, such as artefacts made out of flint and wood, stone and bone tools, and a large number of vertebrate skeletal remains. The exposed Ouaternary deposits cover the late Middle Pleistocene up to the Holocene (Mania, 1995; Lang et al., 2012). The Elsterian till forms the base of the Quaternary sedimentary sequence (Fig. 1C). On top of the Elsterian till lies a series of so-called channels (Mania, 1995; Thieme, 1999) (Fig. 1B). Channels I–III date from the period between the Elsterian and the Saalian (Drenthe) glaciations. The majority of the fossil material studied for this paper is from deposits of the second channel. Channel II contains five depositional levels of organic muds and peats with loess deposits on top, dating from the second half of the Reinsdorf Interglacial and the ensuing Fuhne cold stage. The Reinsdorf Interglacial is generally correlated with Marine Isotope Stage 9 with an age of around 300 ka (Lang et al., 2012; Van Kolfschoten, 2012, 2014). For more detailed information about the Reinsdorf sedimentary sequence the reader is referred to Urban et al. (2011) and Lang et al. (2012). Within Channel II, two spatially separated concentrations of sites are located: Schö 12 and Schö 13, each with a number of sites/depositional levels. The notation of the different sites and depositional levels (e.g., Schö 13 II-4) refers to a specific geographical position in the investigated area (Schö 13), to a particular channel (II), and to a specific depositional level within the sedimentary sequence (4; e.g., Serangeli et al., 2012b). Most of the larger mammal material was collected from the Schö 13 II site, in particular Schö 13 II-4: more than 12,000 large mammal remains are recorded from the Schö 13 II-4 site representing a variety of species. The famous spears are also from the Schö 13 II-4 site.

The botanical (Urban, 2007a,b) and malacological (Mania, 2007) record from the successive depositional levels in Channel II levels 1-5 indicate an environmental and/or climatic shift from an interglacial optimum at the base of the sequence (level 1) with vegetation that is characterized by the occurrence of warm deciduous forest to the beginning of the following cold stage at the top of the sequence with a dominance of boreal forest and steppe vegetation (Urban, 2007a). However, the changes in the mammalian record are not as obvious as one would expect (Van Kolfschoten, 2014): the Channel II sequence did not yield mammal assemblages that clearly indicate climatic deterioration and the occurrence of glacial conditions. Several assumed forest dwellers (for example Stephanorhinus kirchbergensis, Sus scrofa) that occur in Channel II level 1 are also represented in the faunal assemblage of Channel II level 4 (Van Kolfschoten et al., 2007; Van Kolfschoten, 2014). The faunal assemblages from level 1 indicate interglacial conditions and a forested environment alternating with areas of more open, steppe vegetation, while the mammalian fauna from Schö 13 II-4 also suggest the occurrence of forested areas in a steppe landscape.

The environmental context of hominin activities at Schöningen is still a matter of debate, in particular the proportion of forest versus open landscape during the deposition of different levels in Schöningen. Stable isotope investigation might deliver a valuable contribution to this discussion, since  $\delta^{13}$ C and  $\delta^{15}$ N values in plants, and consequently in the bone collagen of the consumers, can vary due to several environmental factors. For example, herbivore populations feeding in dense forest usually have more negative  $\delta^{13}$ C values than animals feeding in open environments (e.g., Bocherens et al., 1999; Drucker et al., 2008). This principle was used to examine, for instance, whether the environment was open or forested during the deposition of different layers in the Scladina Cave in Belgium (Bocherens et al., 1999; Bocherens, 2014) and to document the use of dense forest habitat by large herbivores from interglacial periods in southwestern Germany (Pushkina et al., 2014).

The aim of this study is to investigate the ecological preference of the different herbivores represented in the Schöningen assemblages using stable isotope data. This isotopic study was preceded by a thorough examination of the preservation of the collagen in such an old context.

### Materials and methods

### Material

For this study skeletal material of 69 specimens including bones and antlers has been selected. The specimens were from Schöningen 12 and 13 and belonged to different taxonomic families of large herbivorous mammals: Elephantidae, Rhinocerotidae, Equidae, Cervidae, and Bovidae.

The Elephantidae were represented by eight samples of the straight-tusked elephant (*Palaeoloxodon antiquus*). Samples of Rhinocerotidae (n = 18) included Merck's rhinoceros (*Stephanorhinus kirchbergensis*), narrow-nosed rhinoceros (*Stephanorhinus hemitoechus*), and *Stephanorhinus* sp. Skeletal material of 20 horses (*Equus mosbachensis*) was sampled; 10 bone samples originated from Schö 13 II-4. Among the 16 Cervidae samples, red deer (*Cervus elaphus* and cf. *Cervus elaphus*) were most frequent. Furthermore, four samples of giant deer (*Megaloceros giganteus*) were taken. The seven Bovidae samples were represented by bison (*Bison priscus*) and a number of samples that could not be determined as either aurochs or bison (hereafter designated as *Bos*/*Bison*).

### Methods

Collagen extraction and isotopic analysis was undertaken, and the stable isotope concentrations were measured by IRMS (Isotope Ratio Mass Spectrometry) at the Centre for Isotope Research in Groningen following an improved version of the Longin method (Longin, 1971) and at the Department of Geosciences of Tübingen University following the method published by Bocherens et al. (1997a).

The isotopic content of materials is expressed in delta ( $\delta$ ) values, which are defined as the deviation (expressed in per mil) of the rare to abundant isotope ratio from that of a reference material:

$$\delta^{13}C = \frac{({}^{13}C/{}^{12}C)_{sample}}{({}^{13}C/{}^{12}C)_{reference}} - 1(\times 1000\%) \text{ and}$$
  
$$\delta^{15}N = \frac{({}^{15}N/{}^{14}N)_{sample}}{({}^{15}N/{}^{14}N)_{reference}} - 1(\times 1000\%)$$

For carbon, the reference material is belemnite carbonate (V-PDB); for nitrogen, the reference is ambient air. Bone collagen was prepared following an improved version of the Longin method (Longin, 1971; Mook and Streurman, 1983). The collagen was combusted into CO<sub>2</sub> or N<sub>2</sub> gas by an Elemental Analyzer, coupled to an Isotope Ratio Mass Spectrometer (Aerts et al., 2001). The analytical error is 0.1‰ and 0.2‰ for  $\delta^{13}$ C and  $\delta^{15}$ N, respectively.

The atomic C:N ratio (C:N =  $(%C/\%N) \times (14/12)$ ) is a proxy for the integrity of the collagen. The widely accepted range of atomic C:N ratios for well-preserved bone and tooth collagen is 2.9–3.6 (DeNiro, 1985; Ambrose, 1990; Van Klinken, 1999). This range of values was used here for accepting samples as reliable: all



**Figure 1.** Origin of the samples discussed in this paper. A: Geographical position of the locality Schöningen. B: Horizontal distribution of the sites mentioned in the text (after Serangeli et al., 2012b: 14). C: General lithology of the Reinsdorf stratigraphic sequence, discordantly overlying Elsterian glacial sediments. Key: 1) Elsterian till, gravel and fluvioglacial sediments, 2) sand, 3) basin silt, 4) silty and calcareous mud, 5) ice wedges, 6) solifluction layers, 7) fen peat (Urban et al., 2011: 130).



**Figure 2.** Carbon content relative to nitrogen content in the samples. The samples with both acceptable C:N ratios and %C and %N values are shown in black triangles ( $\mathbf{\nabla}$ ), while rejected samples are represented by open circles ( $\bigcirc$ ).

'collagen' samples with C:N ratios outside this range are rejected. In addition, based on comparison with the chemical composition of collagen extracted from fresh bone using the same purification treatment, the carbon content of genuine collagen should be around 30-40% and its nitrogen content around 11-16% for the results to be reliable (e.g., Van Klinken, 1999). Besides examining the carbon and nitrogen content and the atomic C:N ratio with the aim of assessing the quality of the collagen, five representative samples were <sup>14</sup>C dated by AMS in order to confirm the antiquity of the collagen samples and, in particular, determine if the samples have been contaminated by geologically young material. The <sup>14</sup>C activities are reported in conventional activities (<sup>14</sup>a<sub>N</sub>), that is, they are measured relative to oxalic acid standard and corrected for isotopic fractionation using the stable isotope ratio  ${}^{13}C/{}^{12}C$  to  $\delta^{13}C = -25\%$ , using a half-life value of 5568 years (Mook and van der Plicht, 1999).

### **Results and discussion**

### Collagen preservation

A number of preliminary chemical composition determinations following the approach of Bocherens et al. (2005), measuring the

 Table 1

 Overview of samples with accepted collagen, showing the number of samples per taxonomic group and per site layer.

|                | Level 1         |             | Level 2         |                 | Level 3     | Lev             | el 4            | Total           |    |
|----------------|-----------------|-------------|-----------------|-----------------|-------------|-----------------|-----------------|-----------------|----|
|                | Schö<br>13 II-1 | Schö<br>12B | Schö<br>12 II-1 | Schö<br>13 II-2 | Schö<br>12A | Schö<br>13 II-3 | Schö<br>13 II-4 | Schö<br>13 II A |    |
| Elephantidae   | 4               | 1           |                 |                 |             |                 | 1               |                 | 6  |
| Rhinocerotidae | 1               | 4           |                 |                 |             | 1               | 1               |                 | 7  |
| Equidae        | 1               | 1           |                 |                 | 1           | 1               | 10              | 1               | 15 |
| Cervidae       | 1               | 2           | 1               | 1               |             | 4               | 3               |                 | 12 |
| Bovidae        |                 |             |                 | 1               |             | 3               | 3               |                 | 7  |
| Total          | 7               | 8           | 1               | 2               | 1           | 9               | 18              | 1               | 47 |

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| Sample                        | Site layer   | GrA nr | <sup>14</sup> C activity<br><sup>14</sup> a (%)   | <sup>14</sup> C age (BP) |
|-------------------------------|--------------|--------|---|--------------------------|
| Palaeoloxodon antiquus        | Schö 12B     | 49111  | $\begin{array}{c} 0.20 \pm 0.03 \\ 0.00 \pm 0.02 \\ 0.27 \pm 0.03 \\ 0.03 \pm 0.03 \\ -0.02 \pm 0.02 \end{array}$ | >45,000                  |
| Palaeoloxodon antiquus        | Schö 13 II-4 | 49249  |   | >45,000                  |
| Stephanorhinus kirchbergensis | Schö 12B     | 49107  |   | >45,000                  |
| Equus mosbachensis            | Schö 12A     | 49110  |   | >45,000                  |
| Cervus elaphus                | Schö 13 II-3 | 49112  |   | >45,000                  |

percentage of nitrogen in whole bones, were performed and yielded values up to 2-3%, not far from the 4% nitrogen found in fresh bones. Collagen was successfully extracted for 72% of the samples, a very high percentage given the geological age of the fossils. However, not all of these samples meet the collagen quality criteria and to determine which samples have good collagen and thus can be considered to produce reliable stable isotope ratios for ecological interpretation, the %C, %N, and C:N ratios were examined.

Fig. 2 presents the carbon and nitrogen content of the extracted bone collagen and shows that 47 samples have %C between 31.1 and 47.0, %N between 10.2 and 16.3, and an atomic C:N ratio between 3.1 and 3.6. We considered these 47 samples to have good quality collagen, with collagen yield varying between 0.5% and 7.8% (mean = 2.9%). An overview of these samples is given in Table 1. The collagen of the other three samples does not meet the quality criteria, and the stable isotope ratios of these samples are therefore disregarded in the final paleobiological interpretation. The final result is that 68% of the Schöningen samples yielded reliable collagen.

The <sup>14</sup>C activities measured are shown in Table 2. The background for this batch showed an activity of <sup>14</sup>a = 0.20%, corresponding with a radiocarbon age of >50 kBP. Based on laboratory experience, the background for fossil bone is determined as 45 ka BP (thousands of years before present). There is one sample (GrA-49107) showing a finite age of 47 ka BP when using anthracite as a background; we report this age as >45 ka BP. It appears impossible to have proper blanks (i.e., infinite age and the same quality as the bone sample, i.e., degradation properties) for fossil bone of this age. All results are at the <sup>14</sup>C background level, which means they are older than 45 ka BP. This convincingly indicates that the Schöningen bones do not show contamination with modern carbon.

The occurrence of good quality collagen in bones that are older than 50 ka is remarkable but not unique. Stable isotope data have also been recorded from archaeological sites such as Scladina Cave in Belgium (±120 ka BP, Eemian interglacial [MIS 5e], n = 36/47 [77%]: Bocherens et al., 1999), Stanton Harcourt in southern Britain (200 ka [MIS 7], n = 4/7 [57%]: Jones et al., 2001), Neumark-Nord 2 in Germany (±120 ka BP, Eemian interglacial [MIS 5e], n = 23/42 [55%]: Britton et al., 2012), and Venta Micena in southeastern Spain (±1.5 Ma [millions of years ago], n = 77/105 [73%]: Palmqvist et al., 2003, 2008).

The quality of the collagen of skeletal remains from Stanton Harcourt was estimated by analyzing the amino acid profiles of the mammoth and elephant remains and by considering the atomic C:N ratio. However, two out of four samples that have been considered to yield qualitatively good collagen had low %C and %N values (*Bison priscus*: %C = 23.2 and %N = 8.5; *Palaeoloxodon antiquus*: %C = 23.9 and %N = 8.2). Britton and colleagues (2012) also included samples (n = 7) in their isotope analysis with collagen having low %C (12.6–27.0) and low %N (4.2–9.5), values that are outside of the range of values that are considered in the present study to correspond to good quality collagen. Palmqvist and colleagues (2003) performed amino acid analysis on a small number of samples (n = 4) and only considered samples with C:N ratios between 2.9 and 3.6 to constitute good quality collagen. However, the



Figure 3. Proportions of extracted collagen in bones older than 50 ka in different archaeological sites. Reliable collagen samples (collagen with %C and %N values that are within the range of fresh collagen) are shown in black. Collagen samples classified as reliable in other studies but that are or may be outside the range of fresh collagen are shown in gray. Samples that did not yield collagen are shown in white.

authors did not report the %C and %N for the samples. For the analyses of the Schöningen data we restrict ourselves to collagen with %C and %N values that are within the range of fresh collagen. Using such restrictive conditions, quite a high proportion (68%; Fig. 3) of the Schöningen fossils fulfill that criteria.

The current data indicate that a large part of the Schöningen vertebrate remains is well preserved, not only at a macroscopic level but also at a molecular level. An explanation for the excellent conservation could be the high groundwater level (Lang et al., 2012; Serangeli et al., 2012a). Until very recently the groundwater table at many Schöningen sites was above the find horizons, and the fossil remains were located in waterlogged sediments. This buffered depositional setting offered favorable conditions for the preservation of faunal and botanical remains (Hedges and Millard, 1995; Bocherens et al., 1997b; Prummel and Niekus, 2011; Lang et al., 2012; Serangeli et al., 2012a). Furthermore, the ground water that partly originates from springs in the Elm ridge is rich in calcium carbonate (Huckriede, 1967; Lang et al., (2015)), which also has a positive effect on the preservation of organic material.

### Stable isotopic values

The results of the stable isotope measurements are shown in Table 3. The data set of good quality collagen samples consists of 47 fossil bones (the black down pointing triangle data points in Fig. 2), representing different mammal species and different sites and/or stratigraphical levels. More than a third of these samples are derived from the famous spear horizon (Schöningen 13 II-4, n = 18). In Table 3 the samples that have not been accepted (the open circles in Fig. 2) are crossed out.

The  $\delta^{15}$ N values from all collagen samples ranged from 2.5% to 8.7‰, with an average of 4.8‰ and a standard deviation of 1.6. The  $\delta^{13}C$  values from all collagen samples ranged from -23.6%to -19.8%, with an average of -21.4% and a standard deviation of 0.9. Although the  $\delta^{13}$ C and  $\delta^{15}$ N ranges of several mammal species show some overlap, certain trends among species are visible. For example, the  $\delta^{13}$ C values of Elephantidae (-22.9% to -21.4%) are all lower than those of Bovidae (-21.2% to -19.8%), while the  $\delta^{15}$ N values of Elephantidae (6.6% to 8.7%) exceed the stable nitrogen values of all other analyzed herbivores, except for some Bovidae (3.6‰ to 7.1‰). The significance of the differences observed in  $\delta^{13}$ C and  $\delta^{15}N$  values between the families was determined with the Mann–Whitney U test, where  $H_0$  = not significantly different and  $H_1$  = significantly different (Shennan, 1997). As shown in Table 4, the  $\delta^{13}$ C values were significantly different for Elephantidae compared to Bovidae, Elephantidae compared to Cervidae, Equidae compared to Cervidae, and Equidae compared to Bovidae. Between all families, the  $\delta^{15}$ N values were significantly different, except for these of Rhinocerotidae compared to Equidae, and Rhinocerotidae compared to Bovidae.

The current data show interesting patterns in respect to the environmental conditions (forest versus open landscape)

experienced by hominins at Schöningen. In Figs. 4 and 5, the  $\delta^{15}N$ and  $\delta^{13}$ C values, respectively, are shown for the different taxonomic families during depositional level 1 (indicated with black circles; samples derived from Schö 12B, Schö 12 II-1, and Schö 13 II-1) and depositional level 4 (indicated with open circles; samples derived from Schö 13 II A and Schö 13 II-4). Based on, for example, botanical and malacological data, depositional level 1 is assumed to represent the climatic Reinsdorf Interglacial optimum during which the environment was quite closed with forest dominated by oak. ash. and linden, whereas depositional level 4 is characterized by a boreal continental climate and an open steppe-like landscape with some pine, spruce, larch, and birch (Urban, 2007b). As Fig. 4 shows, no divergent pattern in  $\delta^{15}$ N values exists between the two depositional levels. The  $\delta^{15}$ N values of rhinoceroses, horses, and cervids seem to alternate with each other in the different periods. The highest  $\delta^{15}$ N value (8.7‰) was measured from the sample of a young straight-tusked elephant from level 4 and might reflect the effect of suckling on a not yet weaned individual, as has been observed in woolly mammoths (Metcalfe et al., 2010; Bocherens et al., 2013). The three bovids in Figs. 4 and 5 are all derived from level 4, and no samples were taken from Bovidae living during the Reinsdorf Interglacial optimum. The Rhinocerotidae and Elephantidae have low values (-23.6% to -22.1% and -22.9%)to -21.8‰, respectively) during the Reinsdorf Interglacial optimum and higher values in level 4 (-20.4% and -21.4%, respectively; Fig. 5). Since only one measurement is available for each of these species from level 4, stating a significant difference in  $\delta^{13}$ C values between the two periods might be too oversimplified. The stable carbon values of the horses and cervids from level 1 overlap with those from level 4.

Since the  $\delta^{15}$ N and  $\delta^{13}$ C values in bone collagen are related to the food consumed by the animal, the variation in plant  $\delta^{15}N$  and  $\delta^{13}C$ values is reflected in the tissues of the herbivore eating the plants. For example, fungi and grasses/sedges have relatively higher  $\delta^{15}N$ values than lichens, mosses, and shrubs/trees (see summary in Bocherens [2003] and Drucker et al. [2010]), and understory vegetation in dense forest usually has more negative  $\delta^{13}$ C values (i.e., the so-called 'canopy effect') than plants growing in open environments (e.g., Drucker et al., 2008). The fact that the  $\delta^{15}N$ values do not seem to diverge between depositional level 1 and 4 points to a quite similar dietary plant preference for these mammal species during both periods. The  $\delta^{13}$ C values indicate relative stability in the diet despite climatic change but also point to an environmental change. As shown in Fig. 5, seven samples have  $\delta^{13}$ C values of -22.5% and lower, suggested to reflect the canopy effect. This canopy effect is only seen in samples from level 1, the Reinsdorf Interglacial optimum. The stable carbon values thus indicate that the landscape was more densely forested during depositional level 1 and more open during level 4. However, the majority of samples from level 1 have  $\delta^{13}$ C values between -22.4% and -20.5‰, indicating that these animals mainly fed in an environment that was not densely forested. Thus, the  $\delta^{13}$ C values

| Table 3  |
|--|
| Stable isotope ratios $\delta^{13}$ C and $\delta^{15}$ N for fossil bones from Schöningen with accepted |
| collagen and rejected collagen (crossed out).  |

| Species                         | Layer        | Material | %C              | %N              | C:N        | $\delta^{13}C$ | $\delta^{15}N$ |
|---------------------------------|--------------|----------|-----------------|-----------------|------------|----------------|----------------|
| Flonbantidao                    |              |          |                 |                 |            | _              |                |
| Palaeoloxodon antiquus          | Schö 12 B    | hone     | 40.6            | 144             | 22         | _22.0          | 73             |
| Palaeoloxodon antiquus          | Schö 13 II-1 | bone     | 36.6            | 12.2            | 35         | -22.4          | 7.8            |
| Palaeoloxodon antiquus          | Schö 13 II-1 | bone     | 30.1            | 10.7            | 3.3        | -22.9          | 6.6            |
| Palaeoloxodon antiquus          | Schö 13 II-1 | bone     | 35.3            | 11.4            | 3.6        | -22.5          | 7.0            |
| Palaeoloxodon antiquus          | Schö 13 II-1 | bone     | 37.6            | 13.2            | 3.3        | -21.9          | 7.2            |
| Palaeoloxodon antiquus          | Schö 13 II-4 | bone     | 41.6            | 15.2            | 3.2        | -21.4          | 8.7            |
|                                 |              |          |                 |                 | Av.        | -22.2          | 7.4            |
|                                 |              |          |                 |                 | SD         | 0.5            | 0.7            |
| Rhinocerotidae                  |              |          |                 |                 |            |                |                |
| Stephanorhinus                  | Scho 12 B    | molar    | 43.2            | 14./            | 3.4        | -22.1          | 6.4            |
| Kirchbergensis                  | Schö 12 P    | root     | 40 G            | 15 2            | 21         | 22.4           | 57             |
| kirchhergensis                  | 3CH0 12 B    | root     | 40.0            | 15.2            | 5.1        | -22.4          | 5.7            |
| Stenhanorhinus sp               | Schö 12 B    | bone     | 363             | 12.8            | 33         | -22.7          | 65             |
| (cf. S. kirchbergensis)         | 56110 12 5   | bone     | 50.5            | 12.0            | 5.5        | 2217           | 0.0            |
| Stephanorhinus sp.              | Schö 12 B    | bone     | 40.3            | 14.6            | 3.2        | -22.9          | 3.7            |
| (cf. S. kirchbergensis)         |              |          |                 |                 |            |                |                |
| Stephanorhinus sp.              | Schö 13 II-1 | bone     | 35.6            | 12.6            | 3.3        | -23.6          | 3.4            |
| Stephanorhinus sp.              | Schö 13 II-3 | bone     | 42.4            | 14.8            | 3.3        | -20.4          | 3.1            |
| Stephanorhinus sp.              | Schö 13 II-4 | bone     | 44.0            | 15.2            | 3.4        | -20.4          | 3.8            |
| Stephanorhinus sp.              | Schö 13 II-3 | bone     | 32.1            | <u>9.2</u>      | 4.1        | - <u>21.2</u>  | 4.3            |
| Stephanorhinus sp.              | Schö 13 II-3 | bone     | <del>33.2</del> | <del>10.6</del> | 3.7        | -22.2          | 2.8            |
|                                 |              |          |                 |                 | Av.        | -22.1          | 4.7            |
| Fauidao                         |              |          |                 |                 | SD         | 1.2            | 1.5            |
| Equiuae<br>Fauus moshachensis   | Schö 12 A    | hone     | 38.4            | 132             | 34         | _22.7          | 36             |
| Equus mosbachensis              | Schö 12 R    | bone     | 343             | 12.1            | 33         | -23.2          | 4.6            |
| Equus mosbachensis              | Schö 13 A    | bone     | 37.1            | 12.4            | 3.5        | -21.9          | 3.2            |
| Equus mosbachensis              | Schö 13 II-1 | bone     | 35.4            | 12.4            | 3.3        | -20.9          | 2.5            |
| Equus mosbachensis              | Schö 13 II-3 | bone     | 33.7            | 11.7            | 3.4        | -21.5          | 3.6            |
| Equus mosbachensis <sup>a</sup> | Schö 13 II-4 | bone     | 46.5            | 15.1            | 3.6        | -21.2          | 3.1            |
| Equus mosbachensis <sup>a</sup> | Schö 13 II-4 | bone     | 44.3            | 15.0            | 3.4        | -21.4          | 3.1            |
| Equus mosbachensis <sup>a</sup> | Schö 13 II-4 | bone     | 44.1            | 14.6            | 3.5        | -21.4          | 3.0            |
| Equus mosbachensis <sup>a</sup> | Schö 13 II-4 | bone     | 34.6            | 12.6            | 3.2        | -21.6          | 4.2            |
| Equus mosbachensis <sup>a</sup> | Schö 13 II-4 | bone     | 47.0            | 15.5            | 3.5        | -21.9          | 2.7            |
| Equus mosbachensis <sup>a</sup> | Scho 13 II-4 | bone     | 41.1            | 15.1            | 3.2        | -21.1          | 2.8            |
| Equus mosbachensis <sup>a</sup> | Schö 13 II-4 | bone     | 45.8            | 10.2            | 3.3        | -22.1          | 2.8            |
| Equus mosbachensis <sup>a</sup> | Schö 13 II-4 | bone     | 41.0<br>37.4    | 13.2            | 3.0<br>3.4 | -20.0          | 4.5            |
| Equus mosbachensis <sup>a</sup> | Schö 13 II-4 | bone     | 39.5            | 13.0            | 34         | -21.0          | 2.9            |
| Equus mosbachensis              | Schö 13 B    | bone     | 28.8            | 14.3            | 2.3        | -25.7          | 3.7            |
| 1                               |              |          |                 |                 | Av.        | -21.6          | 3.3            |
|                                 |              |          |                 |                 | SD         | 0.8            | 0.7            |
| Cervidae                        |              |          |                 |                 |            |                |                |
| Cervus elaphus                  | Schö 12 B    | antler   | 30.5            | 11.0            | 3.2        | -23.0          | 4.8            |
| Cervus elaphus                  | Schö 12 B    | bone     | 36.5            | 13.9            | 3.1        | -20.7          | 3.9            |
| Megaloceros giganteus           | Schö 12 II-1 | bone     | 42.2            | 14.9            | 3.3        | -21.5          | 5.4            |
| Cervus elaphus                  | Scho 13 II-1 | antier   | 30.7            | 11.4            | 3.2        | -20.5          | 3.5            |
| Cervus elaphus                  | Schö 13 II-2 | Done     | 38.U            | 12.4            | 3.0<br>2.4 | -20.9          | 4.9            |
| Cervus elaphus                  | Schö 13 II-3 | antler   | 41.0            | 14.5            | 3.4        | -20.1          | 1.2            |
| Cervus elaphus                  | Schö 13 II-3 | antler   | 41.6            | 14.7            | 33         | -20.0          | 4.0            |
| Megaloceros giganteus           | Schö 13 II-3 | bone     | 37.4            | 12.7            | 3.4        | -20.5          | 5.1            |
| cf. Cervus elaphus              | Schö 13 II-4 | antler   | 35.4            | 12.9            | 3.2        | -21.2          | 4.3            |
| cf. Cervus elaphus              | Schö 13 II-4 | antler   | 35.2            | 12.1            | 3.4        | -21.8          | 3.7            |
| Megaloceros giganteus           | Schö 13 II-4 | molar    | 31.9            | 10.9            | 3.4        | -20.6          | 5.5            |
|                                 |              | root     |                 |                 |            |                |                |
|                                 |              |          |                 |                 | Av.        | -21            | 4.7            |
| Deside -                        |              |          |                 |                 | SD         | 0.8            | 0.7            |
| Biogram                         | Cab = 10 U 0 | har -    | 40.1            | 1 4 1           | 2.2        | 20.4           | c =            |
| DISOTI SP.                      | Schö 12 II-3 | bone     | 40.1            | 14.1            | う.う<br>2 1 | -20.4          | 0.5<br>7 1     |
| Bison sp.<br>Ros/Rison          | Schö 13 II-2 | hone     | 45.0            | 15.0            | 33         | -21.2          | 36             |
| Bos/Bison                       | Schö 13 II-3 | bone     | 41.1            | 144             | 3.3        | -19.8          | 66             |
| Bos/Bison                       | Schö 13 II-3 | bone     | 41.4            | 14.6            | 3.3        | -20.2          | 6.4            |
| Bos/Bison                       | Schö 13 II-4 | bone     | 41.0            | 15.3            | 3.1        | -21.1          | 5.5            |
| Bos/Bison                       | Schö 13 II-4 | bone     | 41.2            | 14.9            | 3.2        | -20.9          | 5.2            |
|                                 |              |          |                 |                 | Av.        | -20.6          | 5.8            |
|                                 |              |          |                 |                 | SD         | 0.5            | 12             |

<sup>a</sup> Indicates horses from the 'spear horizon'.

### Table 4

Results of Mann–Whitney U test for  $\delta^{13}C$  and  $\delta^{15}N$  values between the different families.ª

| Families                    | $\delta^{13}C$ | $\delta^{15}N$ |
|-----------------------------|----------------|----------------|
| Elephantidae-Rhinocerotidae | H <sub>0</sub> | H <sub>1</sub> |
| Elephantidae-Equidae        | H <sub>0</sub> | H <sub>1</sub> |
| Elephantidae-Cervidae       | $H_1$          | H <sub>1</sub> |
| Elephantidae-Bovidae        | $H_1$          | H <sub>1</sub> |
| Rhinocerotidae-Equidae      | H <sub>0</sub> | H <sub>1</sub> |
| Rhinocerotidae-Cervidae     | H <sub>0</sub> | H <sub>0</sub> |
| Rhinocerotidae-Bovidae      | H <sub>0</sub> | H <sub>0</sub> |
| Equidae-Cervidae            | $H_1$          | H <sub>1</sub> |
| Equidae-Bovidae             | $H_1$          | H <sub>1</sub> |
| Cervidae-Bovidae            | H <sub>0</sub> | $H_1$          |

<sup>a</sup>  $H_0 = not$  significantly different values, and  $H_1 = significantly different values.$ 

suggest that during level 1 at least part of the landscape was forested and alternated with a substantial part that was more open, whereas during level 4 the landscape was more open. However, although the  $\delta^{13}$ C values of several species point to a relatively open landscape, these  $\delta^{13}$ C values seem to be too low to reflect a steppe landscape similar to the mammoth steppe-tundra (Bocherens, 2003). This is supported by the zoological data, since remains of typical steppe dwellers such as ground squirrels and hamsters have so far not been recorded in the Schöningen mammalian record.

The current isotopic data suggest that the Schöningen landscape was inhabited by herbivorous species with different ecological niches, which is reflected, for example, in habitat choice (densely forested versus open landscape) and dietary preferences (browsing, grazing, and mixed feeding). The horses from Schöningen have the lowest  $\delta^{15}$ N values of all species. The dental wear pattern observed in the horse molars diverges from that seen in horses from other periods and sites, with evidence that browsing formed a substantial part of the horse diet at Schöningen (see Rivals et al. [2015] for a microwear and mesowear analysis on the horse molars from Schö 13 II-4). The low  $\delta^{15}$ N values also support a browsing diet for the Schöningen horses. This browsing role is remarkable, as horses are generally considered to be typical obligate grazers.



**Figure 4.**  $\delta^{15}N$  values for Elephantidae (El), Rhinocerotidae (Rh), Equidae (Eq), Cervidae (Ce), and Bovidae (Bo) for the Reinsdorf Interglacial optimum ( $\bullet$ ) and the colder, fourth depositional phase ( $\bigcirc$ ). More grass in the diet leads to higher  $\delta^{15}N$  values, while a browsing diet results in lower  $\delta^{15}N$  values.



**Figure 5.**  $\delta^{13}$ C values for Elephantidae (El), Rhinocerotidae (Rh), Equidae (Eq), Cervidae (Ce), and Bovidae (Bo) for the Reinsdorf Interglacial optimum ( $\bullet$ ) and the colder, fourth depositional level ( $\bigcirc$ ). Below the dotted line (at –22.5‰), values are expected to reflect the canopy effect.

The  $\delta^{15}$ N values of the straight-tusked elephants are remarkably higher than those of the other species, except for one bovid. Several studies (e.g., Bocherens et al., 1994; Iacumin et al., 2000) demonstrated that the extinct woolly mammoth (Mammuthus pri*migenius*) had remarkably high  $\delta^{15}N$  values in comparison with other contemporary living herbivores. The results of the current study suggest a similar pattern for the stable nitrogen values of straight-tusked elephants from Schöningen. The driving force behind the discrepancy of  $\delta^{15}$ N values between woolly mammoth and contemporaneous herbivores is not precisely understood but may be induced by dietary selection, physiology, or coprophagy (e.g., Bocherens, 2003; Kuitems et al., 2012). The elevation of  $\delta^{15}$ N values of straight-tusked elephants in the current study might be explained by these factors as well. In contrast to the woolly mammoth, the straight-tusked elephant is classically assumed to be a browser preferring wooded environments (Stewart, 2004; Stuart, 2005). Usually, browsers tend to have lower  $\delta^{15}$ N values than grazers (and lower  $\delta^{13}$ C values; e.g., Drucker et al., 2010). However, recent investigations of microwear patterns on the teeth of Pleistocene proboscideans showed that Palaeoloxodon antiauus exhibits variable dietary preferences similar to those of Mammuthus primigenius (Rivals et al., 2012). Therefore it seems possible that the high  $\delta^{15}$ N values of the Schöningen elephants might be linked to a high proportion of <sup>15</sup>N-enriched grass in their diet.

Merck's rhinoceros (*Stephanorhinus kirchbergensis*) is also believed to have been a browser supplementing its diet by grazing, dwelling in both forest and open landscape (Loose, 1975). In contrast, the narrow-nosed rhinoceros (*Stephanorhinus hemitoechus*) was a true grazer (Loose, 1975). Fossils of both species have been found in Schöningen. Unfortunately, it was not possible to identify all skeletal elements in the sample to the species level. The rhinoceroses have quite a large range in  $\delta^{15}$ N values. Within this spread, it looks as if two groups exist: four lower values (3.1‰ to 3.8‰) and three values that are more enriched (5.7‰ to 6.5‰). Of course, the number of values is limited and there is the risk of over interpretation. However, since the samples with a  $\delta^{15}$ N value of 3.4‰ and 6.4‰ both belong to Merck's rhinoceros, it is unlikely that the difference in values

should (only) be explained by grazing versus browsing. The  $\delta^{13}$ C values of the rhinoceroses diverge in another way than the nitrogen values, having five lower values (-23.6% to -22.1%) and two (*Stephanorhinus* sp.) higher values of both -20.4%.

The cervids have quite low  $\delta^{15}$ N values. One should take into account, however, that seven out of the nine red deer samples have been derived from antler instead of bone. Since antlers re-grow every year, the isotope values in antler reflect a shorter time period than the isotope values in bone material (which reflect the mean isotope signal of several years). Based on stomach content analysis, the red deer is assumed to be a mixed feeder (Gebert and Verheyden-Tixier, 2001) able to incorporate both browsing and grazing in its diet. This is supported by the isotope data of the current study. While the  $\delta^{13}$ C value of one red deer (Schö 12B) is low and seems to reflect the canopy effect (-23.0‰, measured in antler), the 11 other values from red deer (n = 8) and giant deer (n = 3)representing various sites/levels (including another red deer sample from Schö 12B: -20.7‰) are considerably higher (-21.8‰ to -20.1%). Among the cervids, giant deer seem to have higher  $\delta^{15}$ N values than most red deer. This could indicate that the diet of giant deer included grass in a higher proportion than that of red deer.

Six of the seven bovids (from Schö 13 II-3 and Schö 13 II-4) have both high  $\delta^{15}N$  (5.2‰ to 7.1‰) and  $\delta^{13}C$  (-21.2‰ to -20.5‰) values compared with the other species. Both stable nitrogen and carbon values indicate a grazing diet based in an open environment. One bovid (from Schö 13 II-2) has a  $\delta^{15}N$  value of 3.6‰ and  $\delta^{13}C$  value of -20.6‰ and may have supplemented its diet by browsing.

The results of the stable isotope investigations are an important contribution to the debate on the interpretation of the large concentration of fossil horse remains in Schö 13 II-4. It was hypothesized that the Schö 13 II-4 horse assemblage may be the result of a single event during which Paleolithic hominins killed a group of horses. However, the large variation in the isotope values of the horse bones does not support this hypothesis. The 10 horse samples from Schö 13 II-4, the famous spear horizon, have  $\delta^{13}$ C values ranging from -22.1% to -20.0% and  $\delta^{15}N$  values ranging from 2.7% to 4.5%. These ranges of isotopic values seem too large for a homogenous population (Lovell et al., 1986), indicating that more than one horse population is represented in the Schö 13 II-4 assemblage. This conclusion supports the suggestion that we are dealing with multiple events and that, in turn, suggests a scenario in which there is a large concentration of horse remains in a landscape with scattered vertebrate remains from a variety of large mammal species including horses (van Kolfschoten, personal communication, August 2011).

### Conclusions

The current study presented remarkably well-preserved bone collagen of skeletal remains from Schöningen, Germany with an age of 300–400 ka. This indicates that a large part of the vertebrate remains exhibit excellent preservation not only at a macroscopic level but also at a molecular level. We measured stable isotope ratios ( $\delta^{13}$ C and  $\delta^{15}$ N) for five taxonomic groups: Elephantidae, Rhinocerotidae, Equidae, Cervidae, and Bovidae.

The isotopic data indicate that during the Reinsdorf Interglacial the hominins at Schöningen lived in a mosaic-like landscape: during the climatic Reinsdorf Interglacial optimum the landscape seems to have been forested, alternating with open areas, whereas during depositional level 4 the isotopic data give the impression that the landscape was more open, although not steppe-like. These mosaic landscapes were inhabited by herbivorous species with different ecological niches, which is reflected in habitat choice (forest versus open landscape) and diet (browsing, grazing, and mixed feeding). All studied herbivorous mammals, including the horses, from the famous spear horizon (Schö 13 II-4) were living in environments that seem to have been more open than during the climatic Reinsdorf Interglacial optimum, versus the stable isotope values of the horses from the spear horizon seem too variable to represent one homogenous population. This indicates that the Schö 13 II-4 horses did not die during a single event.

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