Check for updates

OPEN ACCESS

EDITED BY Yi Yang, Yangzhou University, China

REVIEWED BY Tianle Xu, Yangzhou University, China Subhasis Batabyal, West Bengal University of Animal and Fishery Sciences, India

*CORRESPONDENCE Xiaodong Kang ⊠ kangxd1971@126.com Kangkang Guo ⊠ guokk2007@nwsuaf.edu.cn

 $^{\dagger}\mbox{These}$ authors have contributed equally to this work

SPECIALTY SECTION This article was submitted to Veterinary Infectious Diseases, a section of the journal Frontiers in Veterinary Science

RECEIVED 31 January 2023 ACCEPTED 30 March 2023 PUBLISHED 17 April 2023

CITATION

Wang D, Gao H, Zhao L, Lv C, Dou W, Zhang X, Liu Y, Kang X and Guo K (2023) Detection of the dominant pathogens in diarrheal calves of Ningxia, China in 2021–2022. *Front. Vet. Sci.* 10:1155061. doi: 10.3389/fvets.2023.1155061

COPYRIGHT

© 2023 Wang, Gao, Zhao, Lv, Dou, Zhang, Liu, Kang and Guo. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Detection of the dominant pathogens in diarrheal calves of Ningxia, China in 2021–2022

Dong Wang^{1†}, Haihui Gao^{1,2†}, Long Zhao^{1†}, Changrong Lv¹, Wei Dou¹, Xiuping Zhang¹, Yong Liu¹, Xiaodong Kang^{2*} and Kangkang Guo^{1*}

¹College of Veterinary Medicine, Northwest A&F University, Xianyang, Shaanxi, China, ²Institute of Animal Science, Ningxia Academy of Agriculture and Forestry Sciences, Yinchuan, China

Introduction: Calf diarrhea is a complex disease that has long been an unsolved problem in the cattle industry. Ningxia is at the forefront of China in the scale of cattle breeding, and calf diarrhea gravely restricts the development of Ningxia's cattle industry.

Methods: From July 2021 to May 2022, we collected diarrhea stool samples from calves aged 1–103 days from 23 farms in five cities in Ningxia, and performed PCR using specific primers for 15 major reported pathogens of calf diarrhea, including bacteria, viruses, and parasites. The effect of different seasons on the occurrence of diarrhea in calves was explored, the respective epidemic pathogens in different seasons were screened, and more detailed epidemiological investigations were carried out in Yinchuan and Wuzhong. In addition, we analyzed the relationship between different ages, river distributions and pathogen prevalence.

Results: Eventually, 10 pathogens were detected, of which 9 pathogens were pathogenic and 1 pathogen was non-pathogenic. The pathogens with the highest detection rate were *Cryptosporidium* (50.46%), Bovine rotavirus (BRV) (23.18%), *Escherichia coli* (*E. coli*) K99 (20.00%), and Bovine coronavirus (BCoV) (11.82%). The remaining pathogens such as Coccidia (6.90%), Bovine Astrovirus (BoAstV) (5.46%), Bovine Torovirus (BToV) (4.09%), and Bovine Kobuvirus (BKoV) (3.18%) primarily existed in the form of mixed infection.

Discussion: The analysis showed that different cities in Ningxia have different pathogens responsible for diarrhea, with *Cryptosporidium* and BRV being the most important pathogens responsible for diarrhea in calves in all cities. Control measures against those pathogens should be enforced to effectively prevent diarrhea in calves in China.

KEYWORDS

diarrhea, calf, epidemic investigation, Ningxia, pathogens

Introduction

Diarrhea is one of the most important diseases that damages the health of calves worldwide. It is considered to be one of the diseases causing the highest economic losses to the cattle industry, with losses of up to 10 million dollars due to calf diarrhea in Norway in 2006, followed by cases of varying degrees of calf diarrhea reported in the United States in 2007, South Korea in 2013, and Pakistan in 2014 (1, 2). The main causes of calf diarrhea are intricate and complex (3, 4). In addition to genetics, age, herd and farm environment, feeding practices, poor management and other complications, the most important factor is infection (5, 6). Many countries, including China, have experienced calf diarrhea outbreaks of differing degrees caused by pathogens, such as *Cryptosporidium*, BRV, BCoV,

E. coli K99 and other pathogens (7-10). According to the annual report of Japan in 2017, the economic losses caused by BRV in the previous years were estimated to be about 1 billion yen (11). In addition to causing diarrhea, Cryptosporidium, BCoV, and E.coli K99 also have different effects on increasing mortality, reducing immunity, and reducing milk production (12, 13).

In China, calf diarrhea outbreaks have been reported in many provinces and regions (14-17), but Ningxia has few reports in the article that has comprehensively and systematically investigated the epidemic situation and pathogen distribution characteristics of calf diarrhea. Ningxia has a natural and favorable breeding environment, coupled with the government policy support for the cattle breeding industry, making it one of the important cattle breeding areas in China. With the growing scale of the cattle industry in Ningxia, diarrhea in calves has become an increasingly serious problem, such as the absence of clinical symptoms in calves carrying the pathogen, the rapid spread of the pathogen, and the effect of different environments on the occurrence of diarrhea, which have not been reported or studied.

In order to investigate the prevalence of calf diarrhea in Ningxia and clarify the main pathogens that cause calf diarrhea prevalence in different cities, and study the effects of different seasons to diarrhea in calves, calf diarrhea fecal samples were collected from 23 large-scale cattle farms in five cities of Yinchuan, Wuzhong, Shizuishan, Zhongwei and Guyuan. Pathogens that have been reported to be associated with calf diarrhea were tested, including E. coli K99 (18), Salmonella (19), Proteus mirabilis (20), Clostridium perfringens (C. perfringens) (21), Bovine Viral Diarrhea Virus (BVDV) (22), BRV (23), BCoV (23), BToV (22), BoAstV (24), BKoV (24), Bovine Norovirus (BNoV) (24), Bovine Enterovirus (BEV) (25), Cryptosporidium (26), Coccidia (27, 28), and Giardia (29). The prevalence and distribution characteristics of these pathogens were analyzed to develop a reasonable and effective treatment plan for diarrhea in calves and to provide basic data for the prevention of diarrhea in calves.

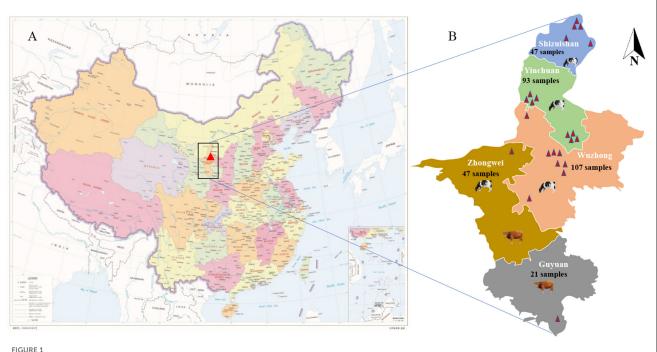
Materials and methods

Sampling

From July 2021 to May 2022, 315 calf stool samples including 220 fresh calf stool samples with diarrhea and 95 fresh normal samples from 23 large-scale cattle farms in 5 cities of Ningxia were collected. Using sterile disposable gloves to collect normal calf rectal stool samples; 4 mL fetal calf serum(FBS)-free DMEM was taken to a sterile 15 mL tube, and the diarrhea stool samples were collected into the tube and stored at 4°C. The common symptoms of diarrheal calves were dehydration, loss of appetite, watery diarrhea, and mental depression. Figure 1A shows the geographical location of the Ningxia Hui Autonomous Region, and Figure 1B shows the geographical location of the cattle farm and the total number of samples collected in each area. Table 1 shows the specific sampling numbers in Ningxia.

DNA extraction

After the collected fresh stool was transported back to the laboratory at 4°C, 200 g of stool were dispensed into 2 mL sterile EP



Cattle farm location and sampling information. (A) The geographical location of Ningxia Hui Autonomous Region (red marked as Ningxia Hui Autonomous Region, red star marked as Beijing, the capital of China). (B) The geographical location of the cattle farm and the total number of samples collected in each city (different colors represent different cities).

02

TABLE 1 The total number of samples with diarrhea and non-diarrhea in five cities.

Location	Number of the calves of diarrhea	Number of non-diarrhea calves
Yinchuan	68	25
Wuzhong	70	37
Shizuishan	35	12
Zhongwei	38	9
Guyuan	9	12
Total	220	95

tubes on a sterile clean bench, and total DNA was extracted using stool DNA kit (OMEGA, Georgia, USA), and then PCR detection was performed to detect *E. coli* K99, *Salmonella*, *Proteus mirabilis*, *C. perfringens*, *Coccidia*, *Cryptosporidium*, *Giardia*.

RNA extraction and reverse transcription

The collected fresh diarrhea stool samples were diluted with 0.9% sterile normal saline. After repeated freezing and thawing at -80° C for three times, the samples were centrifuged at 4°C, 12,000 r/min for 5 min, and the supernatant was collected. Total RNA was extracted from stool using Trizol reagent *AG RNA ex Pro* (Accurate Biotechnology, Hunan, China). According to the manufacturer's operating rules, 2 µg total RNA was reverse transcribed into cDNA using *Evo M-MLV* RT Mix kit with gDNase. The cDNA was used to detect viruses that caused bovine diarrhea such as BVDV, BRV, BCoV, BToV, BoAstV, BKoV, BNoV, and BEV.

Identification and detection of pathogens by PCR

The primers used to detect the above pathogens are shown in Table 2. The extracted RNA was measured using NanoDrop One (Thermo Fisher Scientific, Waltham, MA, USA) and the RNA concentration was in the normal range. Each sample was taken 2 μ g RNA for reverse transcription to obtain the same concentration of cDNA. Nested PCR was performed to detect Cryptosporidium and Giardia using $2 \times \text{Taq}$ Master Mix (Vazyme Biotech, Nanjing, China), the specific PCR system was 2 \times Taq Master mix 10 µL, upstream and downstream primers 1 µL, template 2 µL, supplemented with ddH₂O to 20 µL, the primer concentration was 10 µM. PCR amplification of other pathogens was carried out using 2 × M5 HiPer plus Taq HiFi PCR mix (Mei5 biotechnology, Beijing, China), the specific PCR system was $2 \times M5$ HiPer plus Taq HiFi PCR mix 10 μ L, upstream and downstream primers 1 μ L, template 2 µL, supplemented with ddH2O to 20 µL, the primer concentration was 10 µM.

Detection Coccidia

Take 2 g stools sample of diarrheal calves (\geq 18 d), put it into a beaker, add 5 mL of water first, stir and mix well, add saturated saline to 60 mL, filter through a copper mesh after mixing, absorb the stool liquid, and inject it into McMaster Egg Slide Counting Chamber, after stewing for 5 min, count the number of EPG (Egg Per Gram) or OPG (Oocysts Per Gram) in the two graduated chambers under the microscope (27, 28).

The average A of the number of eggs in the two counting chambers multiplied by 200 is the number of eggs or oocysts per gram of stool. Compute the amount of EPG or OPG of oocysts per gram of stool according to the following formula:

 $EPG/OPG = [(n1 + n2)/(2 \times 0.15)] \times 60 \div 2 = A \times 200$

Statistical analysis

All PCR products were visualized on a 1.0% agarose gel. All positive samples were purified and sequenced by Tsingke Biotechnology (Beijing, China). The sequence results were aligned in GenBank.

The correlation between the pathogen detection rate and the distance between the cattle farm and the river was analyzed using GraphPad, version 9.0.0. Statistical analyses of pathogen detection rates in different seasons throughout Ningxia and in different seasons in Yinchuan and Wuzhong were performed using GraphPad, version 9.0.0. Chi-square tests were performed at a 5% level of significance in SPSS 20.

Results

Detection of different pathogens by PCR

Cryptosporidium

PCR detection using primers designed by Xiao et al. (6), 111 (50.46%) of 220 stool samples were positive, of which 53 (24.09%) were infected by *Cryptosporidium* alone, and the rest were mixed infection (26.36%). The two highest proportions of mixed infections were *Cryptosporidium* and *E. coli* K99 (5.91%), followed by *Cryptosporidium* and BRV (5.45%), and then *Cryptosporidium* and Giardia (4.09%).

Giardia

PCR detection using primers designed by Sulaiman (30), 30 (13.64%) of 220 stool samples were positive, of which 9 (4.09%) were infected by *Giardia* alone and the rest were mixed infection (9.55%). The two highest proportions of mixed infections were *Giardia* and *Cryptosporidium* (4.09%), followed by *Giardia* & *Cryptosporidium* & *E. coli* K99, *Giardia* & *Cryptosporidium* & BRV, *Giardia* & *Cryptosporidium* & BCoV, with a detection rate of 0.91%.

E. coli K99

PCR detection using the primers reported by Keykhaei (18), among the 220 stool samples, 44 (20.00%) were positive, of which

TABLE 2 Primers used for PCR.

Pathogens species	Primer	Sequence $(5'-3')$	Product size (bp)	References
E. coli K99	F5	F: TATTATCTTAGGTGGTATGG	314	(18)
		R: GGTATCCTTTAGCAGCAGTATTTC		
BCoV	Nsp10 of ORF1a	F: CGAGTTGAACACCCAGAT	230	(23)
		R: GAGACGGGCATCTACACT		
BRV	VP6	F: CCACCAGGTATGAATTGGAC	231	
		R: GAGTAATCACTCAGATGGCG		
BNoV	RdRp	F: AGTTAYTTTTCCTTYTAYGGBGA	532	(20)
		R: AGTGTCTCTGTCAGTCATCTTCAT		
BKoV	3D	F: TGGAYTACAAGRATGTTTTGATGC	216	
		R: TGTTGTTRATGATGGTGTTGA		
BoAstV	ORF1a	F: GAYTGGACBCGHTWTGATGG	432	
		R: KYTTRACCCACATNCCAA		
BEV	5 [′] -UTR	F: AGCAACACTGGATTGTGCG	416	(25)
		R: GGAGTAGTCCGACTCCGC		
BVDV	5 [′] -UTR	F: GCTAGCCATGCCCTTAG	290	(22)
		R: CCATGTGCCATGTACAG		
BToV	М	F: TTCTTACTACACTTTTTGGA	603	
		R: ACTCAAACTTAACACTAG AC		
Cryptosporidium	18S rRNA F1	F: TTCTAGAGCTAATACATGCG	1325	(6)
	18S rRNA R1	R: CCCATTTCCTTCGAAACAGGA		
	18S rRNAF2	F: GGAAGGGTTGTATTTATTAGATAAAG	830	
	18S rRNA R2	R: AAGGAGTAAGGAACAACCTCCA		
Giardia	TPI AL3543	F: AAATIATGCCTGCTCGTCG	605	(30)
	TPI AL3546	R: CAAACCTTITCCGCAAACC		
	TPI AL3544	F: CCCTTCATCGGIGGTAACTT	530	
	TPI AL3545	R: GTGGCCACCACICCCGTGCC		

14 (6.36%) were infected by *E. coli* K99 alone, and the rest were mixed infection (13.64%). The two highest proportions of mixed infections are *E. coli* K99 and *Cryptosporidium* (7.73%), followed by *E. coli* K99 and BRV (4.09%), and the proportion of simultaneous infection of *E. coli* K99, *Cryptosporidium*, and BRV is 1.82%. In the stool samples in which *E. coli* K99 was detected, it was only coinfected with *Cryptosporidium* and BRV, and no other pathogens were detected.

Bovine rotavirus

Using the VP6 gene primers of BRV designed by Guo (23) for PCR detection, 51 (23.18%) of 220 stools were positive, of which 16 (7.28%) were infected by BRV alone, and the rest were mixed infection (15.91%). The two highest proportions of mixed infections were BRV and *Cryptosporidium* (7.73%), followed by BRV and *E. coli* K99 (4.09%), and then BRV and BCoV (2.73%).

Bovine coronavirus

Using the Nsp10 gene primers in ORF1a of BCoV designed by Guo (23), 26 (11.82%) of 220 stools were positive, of which 8 (3.64%) were infected alone and 18 (8.18%) were infected with mixed infection. The two highest proportions of mixed infections were BCoV and *Cryptosporidium* (4.55%), followed by BCoV and BRV (0.91%), and then BCoV and *E. coli* K99 (0.91%).

Bovine kobuvirus

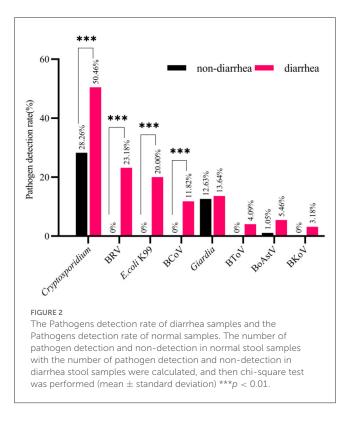
Using the 3D gene primers of BKoV designed by Shi et al. (20) for PCR detection, 7 (3.18%) of 220 stools were positive, all of which were mixed infections. BKoV was predominantly coinfected with *Cryptosporidium* (1.82%) and BRV (1.36%).

Bovine astrovirus

PCR detection using the primers of the ORF1a gene of BoAstV reported by Shi (20) showed that 12 (5.46%) of

Cities	Number of detected calves	Number of infected calves	Infection rate (%)	OPG (pcs/g)	
				Range	Average
Wuzhong	23	2	8.70	4,100-8,600	6,350
Yinchuan	12	2	16.67	17,200-19,000	18,100
Shizuishan	6	0	0	0	0
Zhongwei	8	0	0	0	0
Guyuan	9	0	0	0	0
Total	58	4	6.90	4,100–19,000	12,225





220 stools were positive, of which 1 was infected alone (0.45%), and the rest were mixed infection (5.00%). The two highest proportions of mixed infections were BoAstV and *Cryptosporidium* (2.73%), followed by BoAstV and BRV (1.36%).

Bovine torovirus

The detection was executed using primers designed by Park (22) for the M gene of BToV to PCR. The results showed that 9 of 220 stools (4.09%) were positive, of which 1 was a single infection (0.45%), and the rest were mixed infections (3.64%), suggesting BToV is likely coinfected with two or more pathogens. In a sense, significant diarrhea symptoms only occur when BToV is coinfected with other pathogens.

TABLE 4 The details of calf diarrhea single infection.

Pathogens	Number	Percent (%)
Cryptosporidium	53	24.09
BRV	17	7.73
E. coli K99	14	6.36
Giardia	9	4.09
BCoV	8	3.64
BToV	1	0.46
BoAstV	1	0.46
Coccidia	1	0.46
BKoV	0	0
Total	104	47.27

Coccidia

Through the McMaster Egg Slide Counting Chamber, four cases (6.90%) of 58 calves (\geq 18 days) were detected positive for *coccidia* in this study, including two cases in Wuzhong and two cases in Yinchuan. The OPG levels of the two cases were 4,100 and 8,600 in WuZhong, and the calf ages were 89 and 84 d. The OPG content of the 2 cases was 17,200 and 19,000 in Yinchuan, and the age of the calf was 28 and 27 d. The specific results are shown in Table 3.

Together, a total of nine pathogens causing diarrhea including bacteria, viruses and parasites were detected in this study. The single infection rate of the detected pathogens is shown in Figure 2. The details of a single infection are shown in Tables 4, 5 for details of a mixed infection.

Detection rate of different types of pathogens

In this study, 220 stool samples of calves with diarrhea and 95 normal samples of calves were detected. Among bacterial pathogens, *E. coli* K99 and *C. perfringens* were detected, and *Proteus mirabilis* and *Salmonella* were not detected. The primers reported by Jiang (21) were used to identify *C. perfringens*. The *C. perfringens* detected in this study were all type A and had no pathogenicity.

TABLE 5	The details	of calf	diarrhea	mixed	infection.
---------	-------------	---------	----------	-------	------------

Pathogens	Number	Percent (%)
E. coli K99 & Cryptosporidium	13	5.91
BRV & Cryptosporidium	12	5.45
<i>E. coli</i> K99 & BRV	9	4.09
Cryptosporidium & Giardia	9	4.09
BCoV & Cryptosporidium	7	3.18
BoAstV & Cryptosporidium	5	2.27
BRV & BCoV	3	1.36
BCoV & Cryptosporidium & Giardia	2	0.91
BRV & Cryptosporidium & Giardia	2	0.91
E. coli K99 & Cryptosporidium & Giardia	2	0.91
<i>E. coli</i> K99 & BCoV	1	0.45
BToV & BoAstV	1	0.45
BToV & Giardia	1	0.45
BoAstV & Giardia	1	0.45
BRV & Giardia	1	0.45
BToV & Coccidia	1	0.45
BKoV & Cryptosporidium	1	0.45
BRV & BCoV & BKoV	1	0.45
BRV & BToV & BKoV	1	0.45
BRV & BToV & BoAstV	1	0.45
BCoV & BKoV & Coccidia	1	0.45
E. coli K99 & BCoV & Giardia	1	0.45
E. coli K99 & BRV & Cryptosporidium	1	0.45
E. coli K99 & BoAstV & Cryptosporidium	1	0.45
BRV & BCoV& BoAstV& Giardia	1	0.45
BRV & BToV& BKoV& Cryptosporidium	1	0.45
E. coli K99 & BToV & BKoV& Cryptosporidium	1	0.45
BCoV & BKoV & Coccidia & Cryptosporidium	1	0.45
E. coli K99 & BRV & BToV & BoAstV & Giardia	1	0.45
Total	83	37.73

The detection rates of all pathogens from high to low are *Cryptosporidium* (50.46%), BRV (23.18%), *E. coli* K99 (20.00%), BCoV (11.82%), *Giardia* (13.64%), BoAstV (5.46%), BToV (4.09%), BKoV (3.18%). Comparison of detection details between diarrhea stool samples and normal stool samples by chi-square test, among them, the detection rates of *Cryptosporidium* (p < 0.01), BRV (p < 0.01), *E. coli* K99 (p < 0.01), and BCoV (p < 0.01) were significantly different between diarrhea stool samples and normal stool samples and normal stool samples, while no significant differences were found for the other four pathogens including *Giardia* (p = 0.859), BoAstV (p = 0.118), BToV (p = 0.062), BKoV (p = 0.107).

Among the four diarrhea-related pathogens, BRV, *E. coli* K99, and BCoV were not detected in normal stool samples, but

Cryptosporidium (28.26%) was detected in normal stool samples. The results showed that *Cryptosporidium* had a certain content in normal stool samples and diarrhea stool samples. No clinical diarrhea symptoms in normal stool samples were due to the low content of *Cryptosporidium* in calves. Among other pathogens, *Giardia* (12.63%) and BoAstV (1.05%) were also detected in normal samples and were present in the same situation as *Cryptosporidium*. In contrast, BToV and BKoV were not detected in normal samples, but the results were not significantly different from BToV (4.09%) and BKoV (3.18%) detection rates of diarrhea samples.

The detection of pathogen in different cities

A total of 68 stool samples with diarrhea were detected in Yinchuan: *E. coli* K99 was detected in 19 samples (27.94%); BRV in 14 samples (20.59%); BCoV in 6 samples (8.82%); BToV in 2 samples (2.94%); BoAstV in 1 sample (1.47%); BKoV in 2 samples (2.94%); *Coccidia* in 2 samples (2.94%); *Cryptosporidium* in 28 samples (41.18%); *Giardia* in 6 (8.82%) samples.

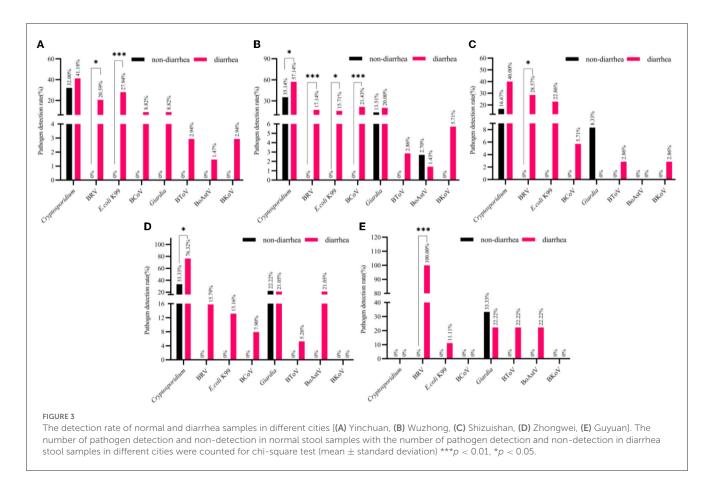
The Chi-square test showed that the main epidemic cause of diarrhea happened in Yinchuan was *E. coli* K99 (p < 0.01), followed by BRV (p < 0.05). Although the detection rate of *Cryptosporidium* (41.18%) was the highest in diarrhea stool samples in Yinchuan, it was also the highest in normal stool samples, and the difference was not significant (32.00%). Other pathogens were detected in normal stool samples. The specific results are illustrated in Figure 3A.

Wuzhong detected *E. coli* K99 in 11 (15.71%) of 70 diarrhea stool samples; BRV in 12 (17.14%) samples; BCoV in 15 (21.43%) samples; BToV in 2 (2.86%) samples; BoAstV in 1 (1.43%) sample; BKoV in 4 (5.71%) samples; *Coccidia* in 2 (2.68%) samples; *Cryptosporidium* in 40 (57.14%) samples; *Giardia* in 14 (20.00%) samples.

The Chi-square test showed that the main epidemic pathogen causing diarrhea in Wuzhong calves was BCoV (p < 0.01), followed by BRV (p < 0.01), E. coli K99 (p < 0.05), Cryptosporidium (p< 0.05). Only Cryptosporidium was detected in both diarrhea and normal stool samples, and the other three pathogens were not detected in normal stool samples. Although the detection rate of Giardia in Wuzhong diarrhea stool samples was higher (20.00%), and it (13.51%) was second only to Cryptosporidium (35.14%) in normal stool samples, and the detection rate of Giardia in normal stool samples and diarrhea stool samples showed no difference. BToV and BKoV were not detected in normal samples, the detection rates in diarrhea stool samples were low (2.86%, 5.71%), and the difference was not significant. The detection rate of BoAstV in normal stool samples (2.70%) was even higher than that in diarrhea stool samples (1.43%). The results are detailed in Figure 3B.

In Shizuishan, detected 35 diarrhea stool samples including eight samples (22.86%) of *E. coli* K99; 10 samples of BRV (28.57%); two samples of BCoV (5.71%); one sample of BToV (2.86%); one sample of BKoV (2.86%); 14 samples of *Cryptosporidium* (40.00%).

The Chi-square test showed that the main epidemic pathogen causing diarrhea in Shizuishan calves was BRV (p < 0.05). Although *Cryptosporidium* (40.00%) and *E. coli* K99 (22.86%) had higher detection rates in diarrhea stool samples, there was no significant difference between them and normal stool samples. In



particular, *Giardia* was not detected in diarrhea stool samples, but its detection rate in normal stool samples (8.33%) was second only to *Cryptosporidium* (16.67%). The results are detailed in Figure 3C.

A total of 38 stool samples with diarrhea were detected in Zhongwei: five samples (13.16%) of *E. coli* K99; four samples of BRV (15.79%); three samples of BCoV (7.90%); two samples of BToV (5.26%); eight samples of BOAstV (21.05%); *Cryptosporidium* 29 (76.32%) samples; *Giardia* 8 (21.05%) samples; BKoV and *Coccidia* were not detected.

The Chi-square test showed that the main epidemic pathogen causing diarrhea in Zhongwei calves was *Cryptosporidium* (p < 0.05), with a detection rate of 76.32%. The detection rates of other pathogens between diarrhea and normal stool samples were showed no significant difference. The detection rate of *Giardia* in normal stool samples (22.22%) was even higher than that in diarrhea stool samples (21.05%). The results are detailed in Figure 3D.

In Guyuan, a total of nine diarrhea stool samples were detected in *E. coli* K99 in 1 (11.11%); BRV in 9 (100%); BToV in 2 (22.22%); BoAstV in 2 (22.22%); *Giardia* in 2 (22.22%).

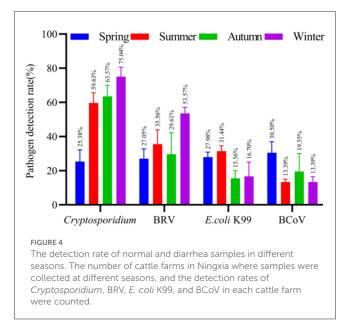
The Chi-square test showed that the main epidemic pathogen causing diarrhea in Guyuan calves was BRV (p < 0.01). Other pathogens were not significantly different. The detection rate of Giardia in normal stool samples (33.33%) was higher than that in diarrhea stool samples (22.22%), which was similar to the detection of Giardia in Zhongwei and Shizuishan. The results are detailed in Figure 3E.

The detection of pathogen in different seasons

The Chi-square test was performed on the number of cattle farms collected in different seasons and the pathogen detection rate of diarrhea fecal samples in each cattle farm. The correlation between the four main pathogens with the significant difference in detection rate in each season and diarrheal calves was analyzed.

The results showed that the dominant pathogens of diarrhea in spring in Ningxia were BCoV (30.50%), *E. coli* K99 (27.98%), BRV (27.05%) and *Cryptosporidium* (25.38%). In summer, the dominant pathogens of diarrhea were *Cryptosporidium* (59.63%), BRV (35.56%), *E. coli* K99 (31.44%) and BCoV (13.39%). In autumn, the dominant pathogens of diarrhea were *Cryptosporidium* (63.57%), BRV (29.61%), BCoV (19.55%) and *E. coli* K99 (15.56%). In winter, the dominant pathogens of diarrhea were *Cryptosporidium* (75.04%), BRV (53.57%), *E. coli* K99 (16.70%) and BCoV (13.39%). The detail results are illustrated in Figure 4.

After the chi-square test of the entire Ningxia, the pathogens of calf diarrhea that were prevalent in each season have been obtained. Yinchuan and Wuzhong are the concentrated breeding areas of cattle in Ningxia. Analyzing the correlation between the significant pathogens in Yinchuan and Wuzhong in each season and diarrheal calves is more important. Based on the detection rate of different pathogens in the cattle farms of Yinchuan and Wuzhong in different seasons, the average detection rate of pathogens in each cattle farm was calculated, and the epidemic diarrhea pathogens in Yinchuan and Wuzhong in different seasons were obtained.



In Yinchuan, the dominant pathogens of diarrhea in spring were BRV (39.55%), *E. coli* K99 (36.82%), *Cryptosporidium* (19.55%) and BCoV (4.55%). In summer, the dominant diarrhea pathogens were *Cryptosporidium* (53.46%), *E. coli* K99 (30.39%), BRV (10.00%), and BCoV was not detected. In autumn, the dominant diarrhea pathogens were *Cryptosporidium* (18.18%), BRV (9.09%), BCoV (9.09%), and *E. coli* K99 was not detected. In winter, the dominant diarrhea pathogens were *Cryptosporidium* (18.18%), BRV (9.09%), BCoV (9.09%), and *E. coli* K99 was not detected. In winter, the dominant diarrhea pathogens were *Cryptosporidium* (92.86%), BRV (28.57%), *E. coli* K99 (16.67%), BCoV (7.15%). The results are detailed in Figure 5A.

In Wuzhong, the dominant pathogens of diarrhea in spring were *Cryptosporidium* (45.00%), BCoV (36.67%), BRV (8.34%), and *E. coli* K99 was not detected. In summer, the dominant diarrhea pathogens were *Cryptosporidium* (81.62%), *E. coli* K99 (21.51%), BCoV (16.45%), BRV (6.03%). In autumn, the dominant diarrhea pathogens were *Cryptosporidium* (70.00%), BCoV (30.00%), BRV (20.00%), *E. coli* K99 (20.00%). In winter, the dominant diarrhea pathogens were BRV (38.33%), BCoV (33.33%), *Cryptosporidium* and *E. coli* K99 were not detected. The results are detailed in Figure 5B.

Distribution of different pathogens in different ages

The earliest onset time and the common age of nine pathogens were illustrated in Figure 6. The earliest onset age of *Cryptosporidium* was 4 days, and the frequent onset age was 5–18 days. The earliest onset age of BRV was 4 days, and the frequent onset age was 7–30 days. The earliest onset age of *E. coli* K99 was 1 day and the common onset age was 8–15 days The earliest onset age of *Giardia* was 7 days, and the most frequent age was 11–30 days. The earliest onset age of BCoV was 2 days, and the most frequent age was 9–26 days. The earliest onset age of BoAstV was 8 days, and the most frequent age was 8–30 days. The earliest onset age of BToV was 8 days, and the most frequent age was 8–44 days. The

earliest onset age of BKoV was 10 days, and the most frequent age was 10–26 days. The earliest onset age of *Coccidia* was 27 days.

Relationship between main diarrhea pathogens and river distribution in Ningxia

Ningxia is a province through which the Yellow River flows, with a length of about 397 km. There are two other tributaries, the Qingshui River and the Kushui River. Among the 23 large-scale cattle farms in this study, 18 cattle farms were close to the river, and the average number of *Cryptosporidium* detected per farm was 7.22, of which 5 cattle farms detected *Cryptosporidium* number \geq 10. Among the five cattle farms where no *Cryptosporidium* were detected and where *E. coli* K99, BRV, and BCoV were the main diarrhea pathogens, three cattle farms were not surrounded by a river and one cattle farm was relatively far from a river.

This suggests *Cryptosporidium* is the main diarrhea pathogen in cattle farms, <500 m from the water source. However, the detection rate of *Cryptosporidium* was positively correlated with the distance from cattle farms to rivers, but not significant (r = 0.1941), while the detection rates of *E.coli* K99, BRV, and BCoV were not correlated with the distance from cattle farms to rivers. The specific analysis results are detailed in Figure 7.

Discussion

Wuzhong is the city with the most types of pathogens and the highest average detection rate, followed by Yinchuan. Because the etiology of calf diarrhea is more complex, in addition to other environmental factors, it is predominantly caused by pathogens [viruses (31), bacteria (1), parasites (32)], especially Cryptosporidium, which is principally transmitted by fecal-oral transmission (33). Therefore, calf density is one of the important factors affecting its transmission rate, and Wuzhong, Yinchuan, and some counties in Shizuishan and Zhongwei are the location of cattle breeding areas in Ningxia, and the density of calf herds is extremely high more than other cities. Consistently, like the results reported in other studies, Cryptosporidium is an important cause of diarrhea in Ningxia calves (33-35). In this study, a total of 315 stool samples were collected from all five cities in Ningxia, and 137 stool samples (43.49%) were positive for Cryptosporidium, including diarrhea samples (50.46%) and normal samples (27.37%). In 2015, researchers reported on Cryptosporidium infection in Ningxia and Gansu (35), 150 positive samples (5.09%) were detected in 2,945 stools in both diarrhea and normal calves. The detection rate of our study is significantly higher than 5.09%, which suggests that the infection rate of *Cryptosporidium* in Ningxia is rising year by year. Since December 2011, the detection rate of Cryptosporidium in Ningxia has shown a significant increase, from 1.68% (23/1,366) (33) to 50.46% (111/220). The infection of Cryptosporidium in calves with diarrhea and normal calves also coexist in this study, which is consistent with the results of the above studies.

At present, the treatment measures for *Cryptosporidium* are only preventive, and there is no effective commercial vaccine on the market to prevent long-term infection in cattle. The increased prevalence is one of the serious problems faced by researchers.

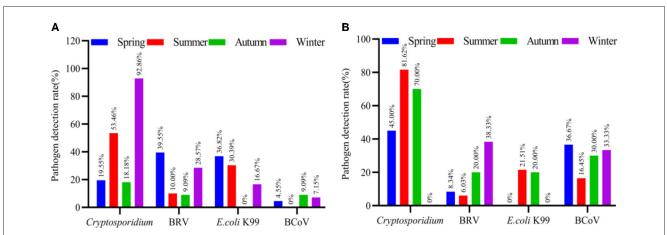
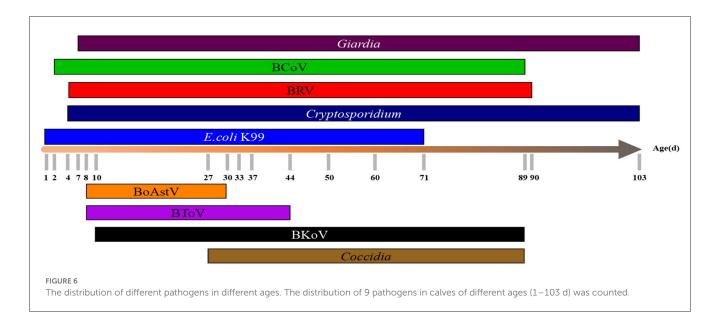


FIGURE 5

The detection rate of normal and diarrhea samples in different seasons [(A) Yinchuan, (B) Wuzhong]. The number of cattle farms in Yinchuan and Wuzhong, where samples were collected at different seasons, and the detection rates of *Cryptosporidium*, BRV, *E. coli* K99, and BCoV in each cattle farm were counted.

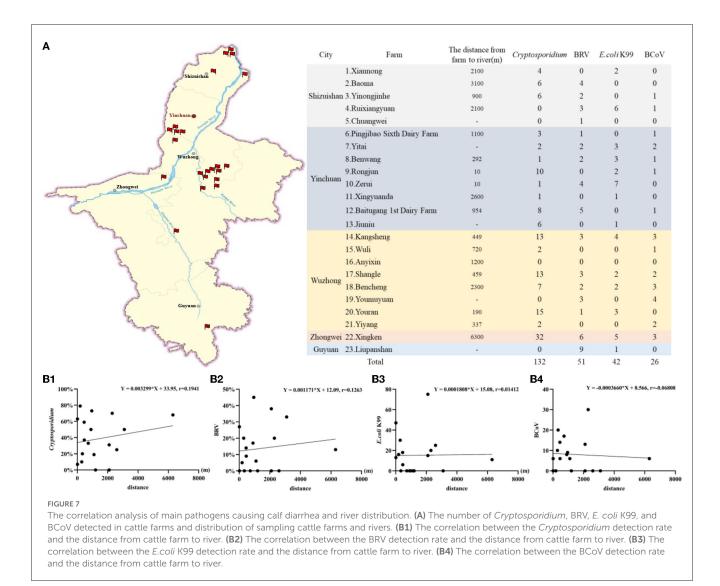


Therefore, it is important to take care of deworming cattle in all growth stages and pay attention to biological safety measures.

BRV is the main pathogen that causes calf diarrhea worldwide. It has been also reported in many regions of China. Rotaviruses are a major causative pathogen of diarrhea in humans and animals, involving the deaths of 200,000 children in developing countries and causing economic losses in the livestock industry globally. In this study, the detection rate of BRV in Ningxia from 2021 to 2022 (23.18 %) was lower than the average detection rate of Ningxia over the years (32%), which was lower than the pooled prevalence of BRV in China 46% (6,635/10,677) (36). This is greatly related to the fact that the Ningxia agricultural department pays more attention to the impact of viruses on the cattle industry.

Compared with *Cryptosporidium* and BRV, the infection rates of *E. coli* K99 in Ningxia were relatively low. However, compared with other pathogens, *E. coli* K99 and other pathogenic *Escherichia coli* are still important pathogens causing calf diarrhea. The detection rate of BoAstV in Zhongwei (21.05%) was significantly higher than that in other cities, but it was not the main cause of diarrhea in Zhongwei calves, the reason may be that the BoAstV detected in this study was neurotype rather than diarrhea type. Evolutionary analyses showed that astrovirus strains from bovine brain tissue were closely related to astrovirus strains from humans, pigs, sheep and other animals with neurological symptoms, indicating that cross-species transmission may occur.

To date, *Cryptosporidium*, *E. coli* K99, BRV and BCoV have been identified as important pathogens prevalent in calf diarrhea in China. In addition, previous studies have demonstrated that BRV can be transmitted to humans directly or through recombination during the evolution of the strain and *Cryptosporidium* and *E. coli* K99, and is typical zoonosis (37). Thus, the in-depth investigation of the above calf diarrhea pathogens is the basis for the prevention and treatment of calf diarrhea, and how to avoid the mixed infection caused by multiple pathogens is of clinical significance. Thus, more efforts should be taken to block the spread of these pathogens in cattle farms and reduce the external factors leading



to calf diarrhea. In total, it is possible to reduce the incidence of calf diarrhea.

The area around the reiver is a high-frequency area for parasite reproduction and transmission, and many parasites, including *Cryptosporidium*, can be transmitted through water (38, 39). *Cryptosporidium* in its oocyst stage can remain infectious for many months under cool, moist conditions such as rivers, lakes and ponds (40), and in a relatively dry environment, it is more suitable for the growth of viruses and bacteria (24, 41). The distribution of calf diarrhea pathogens in Ningxia also showed similar characteristics in this study, and how to prevent the spread of the pathogen due to geographic environmental factors is one of the issues the researchers have been facing.

Conclusion

In this study, *Cryptosporidium* can be detected in both diarrheal calves and normal calves, and other pathogens are a mixed infection of two or more pathogens in the same or different calves. Together, *Cryptosporidium*, BRV, *E. coli* K99 and BCoV are the

main pathogens causing calf diarrhea in Ningxia, the remaining four pathogens are mainly infected in the form of mixed infection.

From June 2021 to May 2022, the main pathogens causing calf diarrhea in Yinchuan were *E. coli* K99 and BRV; the main pathogens causing calf diarrhea in Wuzhong are *Cryptosporidium*, BCoV, BRV and *E. coli* K99; BRV was the main pathogen causing calf diarrhea in Shizuishan; *Cryptosporidium* was the main pathogen causing calf diarrhea in Zhongwei; BRV was the main pathogen causing calf diarrhea in Guyuan.

Different seasons had a more obvious effect on the detection rate of calf diarrhea-related pathogens. In addition, the rivers had an effect on the detection rate of *Cryptosporidium*. In conclusion, the distribution of diarrhea pathogens in Ningxia calves is associated with geographical and environmental factors.

Data availability statement

The original contributions presented in the study are included in the article/supplementary

material, further inquiries can be directed to the corresponding authors.

Ethics statement

The animal study was reviewed and approved by Executive Committee of Laboratory Animal Management and Ethics Inspection of Northwest A&F University, Xianyang, China. Written informed consent was obtained from the owners for the participation of their animals in this study.

Author contributions

KG, XK, and HG designed the experiments. DW, LZ, and WD carried out the experiments. HG and CL collected samples. DW wrote the manuscript. XZ and YL contributed to data analysis and helped complete the experiments. All authors discussed the results and commented on the manuscript.

References

1. Ali A, Liaqat S, Tariq H, Abbas S, Arshad M, Li WJ, et al. Neonatal calf diarrhea: A potent reservoir of multi-drug resistant bacteria, environmental contamination and public health hazard in Pakistan. *Sci Total Environ.* (2021) 799:149450. doi: 10.1016/j.scitotenv.2021.149450

2. Hur J, Jeon BW, Kim YJ, Oh IG, Lee JH. Escherichia coli isolates from calf diarrhea in Korea and their virulent genetic characteristics. *J Vet Med Sci.* (2013) 75:519–22. doi: 10.1292/jvms.12-0378

3. Cho YI, Yoon KJ. An overview of calf diarrhea - infectious etiology, diagnosis, and intervention. *J Vet Sci.* (2014) 15:1–17. doi: 10.4142/jvs.2014.15.1.1

4. Gomez DE, Weese JS. Viral enteritis in calves. Can Vet J. (2017) 58:1267-74.

5. McGuirk SM. Disease management of dairy calves and heifers. The Veterinary clinics of North America. *Food Animal Pract.* (2008) 24:139–53. doi: 10.1016/j.cvfa.2007.10.003

6. Windeyer MC, Leslie KE, Godden SM, Hodgins DC, Lissemore KD, LeBlanc SJ. (2014). Factors associated with morbidity, mortality, and growth of dairy heifer calves up to 3 months of age. *Prevent Vet Med.* 113:231-240. doi: 10.1016/j.prevetmed.2013.10.019

 Caffarena RD, Casaux ML, Schild CO, Fraga M, Castells M, Colina R, et al. Causes of neonatal calf diarrhea and mortality in pasture-based dairy herds in Uruguay: a farm-matched case-control study. *Brazilian J Microbiol.* (2021) 52:977– 88. doi: 10.1007/s42770-021-00440-3

8. Chae JB, Kim HC, Kang JG, Choi KS, Chae JS Yu DH, et al. The prevalence of causative agents of calf diarrhea in Korean native calves. *J Animal Sci Technol.* (2021) 63:864–71. doi: 10.5187/jast.2021.e63

9. Dall Agnol AM, Lorenzetti E, Leme RA, Ladeia WA, Mainardi RM, Bernardi A, et al. Severe outbreak of bovine neonatal diarrhea in a dairy calf rearing unit with multifactorial etiology. *Brazilian J Microbiol.* (2021) 52:2547–53. doi: 10.1007/s42770-021-00565-5

10. Geng HL, Ni HB, Li JH, Jiang J, Wang W, Wei XY, et al. Prevalence of *Cryptosporidium* spp. in Yaks (Bos grunniens) in China: a systematic review and metaanalysis. Front Cell Infect Microbiol. (2021) 11:770612. doi: 10.3389/fcimb.2021.770612

11. Odagiri K, Yoshizawa N, Sakihara H, Umeda K, Rahman S, Nguyen SV, et al. Development of genotype-specific anti-bovine rotavirus a immunoglobulin yolk based on a current molecular epidemiological analysis of bovine rotaviruses a collected in Japan during 2017-2020. *Viruses.* (2020) 12:1386. doi: 10.3390/v12121386

12. Lombardelli JA, Tomazic ML, Schnittger L, Tiranti KI. Prevalence of Cryptosporidium parvum in dairy calves and GP60 subtyping of diarrheic calves in central Argentina. *Parasitol Res.* (2019) 118:2079–86. doi: 10.1007/s00436-019-06366-y

13. Burimuah V, Sylverken A, Owusu M, El-Duah P, Yeboah R, Lamptey J, et al. Molecular-based cross-species evaluation of bovine coronavirus

Funding

This work was supported by the Key Research and Development Program of Ningxia under Grant [Project No. 2021BEF03005].

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

infection in cattle, sheep and goats in Ghana. BMC Vet Res. (2020) 16:405. doi: 10.1186/s12917-020-02606-x

14. Guo Z, He Q, Yue H, Zhang B, Tang C. First detection of Nebovirus and Norovirus from cattle in China. *Arch Virology.* (2018) 163:475–8. doi: 10.1007/s00705-017-3616-6

15. Li N, Wang R, Cai M, Jiang W, Feng Y, Xiao L. Outbreak of cryptosporidiosis due to *Cryptosporidium* parvum subtype IIdA19G1 in neonatal calves on a dairy farm in China. *Int J Parasitol.* (2019) 49:569–577. doi: 10.1016/j.ijpara.2019.02.006

16. Liu X, Yan N, Yue H, Wang Y, Zhang B, Tang C. Detection and molecular characteristics of bovine rotavirus A in dairy calves in China. *J Vet Sci.* 22:e69. doi: 10.4142/jvs.2021.22.e69

17. Wei X, Wang W, Dong Z, Cheng F, Zhou X, Li B, et al. Detection of infectious agents causing neonatal calf diarrhea on two large dairy farms in Yangxin County, Shandong Province, China. *Front Vet Sci.* (2021) 7:589126. doi: 10.3389/fvets.2020.589126

18. Keykhaei N, Salari S, Rashki A. Frequency of k99, stx1, and stx2 virulence factors in escherichia coli isolated from diarrheic and clinically healthy suckling calves in Sistan and Baluchistan Province, Iran. *Arch Razi Inst.* (2021) 76:283–91. doi: 10.22092/ari.2019.124040.1268

19. Zishiri OT, Mkhize N, Mukaratirwa S. Prevalence of virulence and antimicrobial resistance genes in Salmonella spp isolated from commercial chickens and human clinical isolates from South Africa and Brazil. *Onderstepoort J Vet Res.* (2016) 83:1067. doi: 10.4102/ojvr.v83i1.1067

20. Sanches MS, Rodrigues da Silva C, Silva LC, Montini VH, Lopes Barboza MG, Migliorini Guidone GH, et al. Proteus mirabilis from community-acquired urinary tract infections (UTI-CA) shares genetic similarity and virulence factors with isolates from chicken, beef and pork meat. *Microbial Pathogenesis*. (2021) 158:105098. doi: 10.1016/j.micpath.2021.105098

21. Jiang Y, Ma Y, Liu Q, Li T, Li Y, Guo K, et al. Tracing Clostridium perfringens strains from beef processing of slaughter house by pulsed-field gel electrophoresis, and the distribution and toxinotype of isolates in Shaanxi province, China. *Food Microbiol.* (2022) 101:103887. doi: 10.1016/j.fm.2021.103887

22. Park SI, Jeong C, Kim HH, Park SH, Park SJ, Hyun BH, et al. Molecular epidemiology of bovine noroviruses in South Korea. *Vet Microbiol.* (2007) 124:125–33. doi: 10.1016/j.vetmic.2007.03.010

23. Guo Z, He Q, Zhang B, Yue H, Tang C. Detection molecular characteristics of neboviruses in dairy cows in China. J Gen Virol. (2019) 100:35–45. doi: 10.1099/jgv.0.001172

24. Shi Z, Wang W, Chen C, Zhang X, Wang J, Xu Z, et al. First report and genetic characterization of bovine torovirus in diarrhoeic

calves in China. BMC Vet Res. (2020) 16:272. doi: 10.1186/s12917-020-02494-1

25. He H, Tang C, Chen X, Yue H, Ren Y, Liu Y, et al. Isolation and characterization of a new enterovirus F in yak feces in the Qinghai-Tibetan Plateau. *Arch Virol.* (2017) 162:523–7. doi: 10.1007/s00705-016-3119-x

26. Xiao L, Morgan UM, Limor J, Escalante A, Arrowood M, Shulaw W, et al. Genetic diversity within *Cryptosporidium* parvum and related *Cryptosporidium* species. *Appl Environ Microbiol.* (1999) 65:3386–91. doi: 10.1128/AEM.65.8.3386-3391.1999

27. Dong H, Li C, Zhao Q, Li J, Han H, Jiang L, et al. Prevalence of Eimeria infection in yaks on the Qinghai-Tibet Plateau of China. *J Parasitol.* (2012) 98:958–62. doi: 10.1645/GE-3079.1

28. Dong H, Zhao Q, Han H, Jiang L, Zhu S, Li T, et al. Prevalence of coccidial infection in dairy cattle in Shanghai, China. *J Parasitol.* (2012) 98:963–6. doi: 10.1645/GE-2966.1

29. Sulaiman IM, Fayer R, Bern C, Gilman RH, Trout JM, Schantz PM, et al. Triosephosphate isomerase gene characterization and potential zoonotic transmission of Giardia duodenalis. *Emerg Infect Dis.* (2003) 9:1444–52. doi: 10.3201/eid0911.030084

30. Shimoda T, Okubo T, Enoeda Y, Yano R, Nakamura S, Thapa J, et al. Effect of thermal control of dry fomites on regulating the survival of human pathogenic bacteria responsible for nosocomial infections. *PLoS ONE.* (2019) 14:e0226952. doi: 10.1371/journal.pone.0226952

31. Lotfollahzadeh S, Madadgar O, Reza Mohebbi M, Reza Mokhber Dezfouli M, George Watson D. Bovine coronavirus in neonatal calf diarrhoea in Iran. *Vet Med Sci.* (2020) 6:686–94. doi: 10.1002/vms3.277

32. Li N, Zhao W, Song S, Ye H, Chu W, Guo Y, et al. Diarrhoea outbreak caused by coinfections of *Cryptosporidium* parvum subtype IIdA20G1 and rotavirus in pre-weaned dairy calves. *Transbound Emerg Dis.* (2022) 69:e1606–17. doi: 10.1111/tbed.14496

33. Huang J, Yue D, Qi M, Wang R, Zhao J, Li J, et al. Prevalence and molecular characterization of *Cryptosporidium* spp. and Giardia duodenalis

in dairy cattle in Ningxia, northwestern China. BMC Vet Res. (2014) 10:292. doi: 10.1186/s12917-014-0292-6

34. Cui Z, Wang R, Huang J, Wang H, Zhao J, Luo N, et al. Cryptosporidiosis caused by *Cryptosporidium* parvum subtype IIdA15G1 at a dairy farm in Northwestern China. *Parasites Vectors.* (2014) 7:529. doi: 10.1186/s13071-014-0529-z

35. Zhang XX, Tan QD, Zhou DH, Ni XT, Liu GX, Yang YC, et al. Prevalence and molecular characterization of *Cryptosporidium* spp. in dairy cattle, northwest China. *Parasitology Res.* (2015) 114:2781–7. doi: 10.1007/s00436-015-4537-5

36. Chen S, Zhang W, Zhai J, Chen X, Qi Y. Prevalence of bovine rotavirus among cattle in mainland China: a meta-analysis. *Microb Pathog.* (2022) 170:105727. doi: 10.1016/j.micpath.2022.105727

37. Doan YH, Nakagomi T, Aboudy Y, Silberstein I, Behar-Novat E, Nakagomi O, et al. (2013). Identification by full-genome analysis of a bovine rotavirus transmitted directly to and causing diarrhea in a human child. *J Clini Microbiol.* 51, 182–189. doi: 10.1128/JCM.02062-12

38. Bajer A, Toczylowska B, Bednarska M, Sinski E. Effectiveness of water treatment for the removal of *Cryptosporidium* and Giardia spp. *Epidemiol Infect*. (2012) 140:2014–22. doi: 10.1017/S0950268811002780

39. Omarova A, Tussupova K, Berndtsson R, Kalishev M, Sharapatova K. Protozoan parasites in drinking water: a system approach for improved water, sanitation and hygiene in developing countries. *Int J Res Public Health.* (2018) 15:495. doi: 10.3390/ijerph15030495

40. Fayer R. (2004). *Cryptosporidium*: a water-borne zoonotic parasite. *Vet Parasitol.* 126:37–56. doi: 10.1016/j.vetpar.2004.09.004

41. Audi A, Allbrahim M, Kaddoura M, Hijazi G, Yassine HM, Zaraket H. Seasonality of Respiratory Viral Infections: Will COVID-19 Follow Suit? *Front Public Health.* (2020) 8:567184. doi: 10.3389/fpubh.2020.567184