

Mississippi University for Women

## ATHENA COMMONS

---

Undergraduate Research Conference

2023 Undergraduate Research Conference

---

Mar 31st, 10:45 AM

### Triazole compounds – Potentials in the treatment of cystic fibrosis

Maggie Taylor

Follow this and additional works at: <https://athenacommons.muw.edu/urc>

 Part of the [Cells Commons](#), [Diseases Commons](#), and the [Heterocyclic Compounds Commons](#)

---

#### Recommended Citation

Taylor, Maggie, "Triazole compounds – Potentials in the treatment of cystic fibrosis" (2023).  
*Undergraduate Research Conference*. 6.  
<https://athenacommons.muw.edu/urc/2023/poster/6>

This Poster is brought to you for free and open access by the Conferences and Events at ATHENA COMMONS. It has been accepted for inclusion in Undergraduate Research Conference by an authorized administrator of ATHENA COMMONS. For more information, please contact [acpowers@muw.edu](mailto:acpowers@muw.edu).

## AIMS

1. To determine the influence of certain triazole compounds on  $\Delta F508$ -CFTR protein and mRNA expression.

2. To determine the effects of triazole compounds on chloride ion channel activation in  $\Delta F508$ -CFTR expressing cell lines.

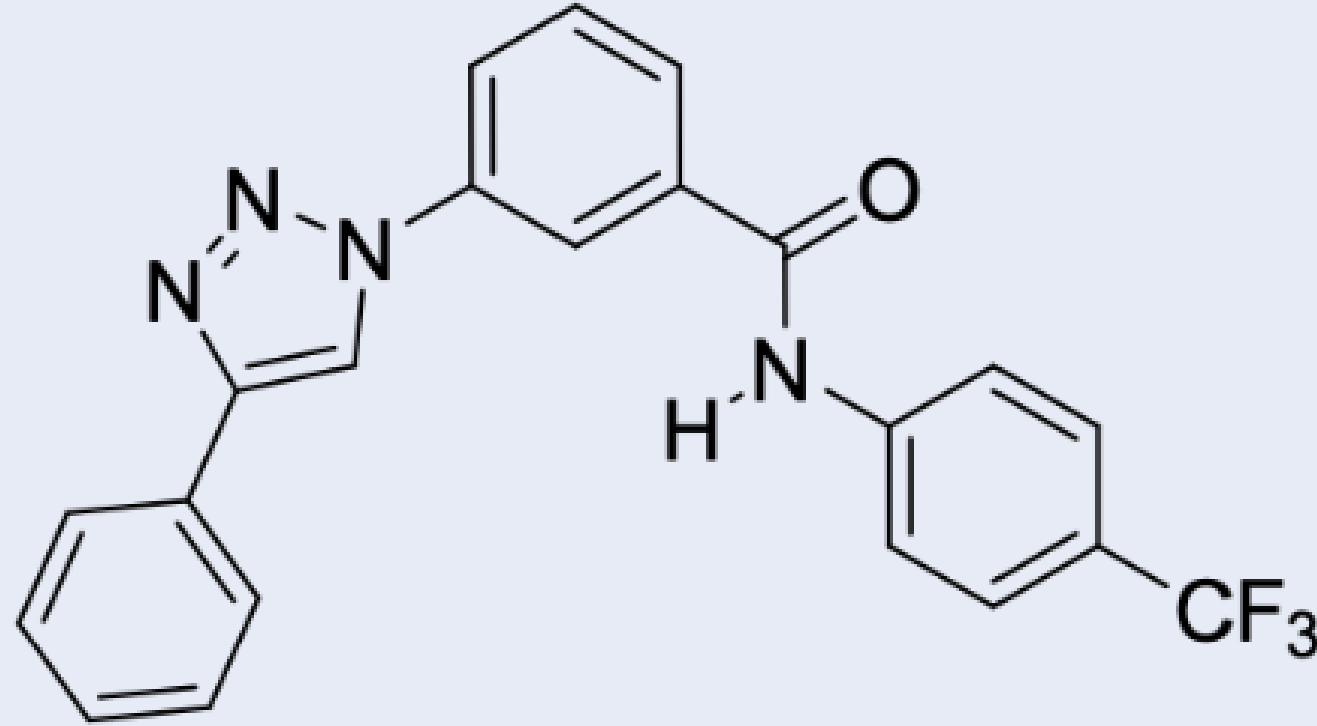
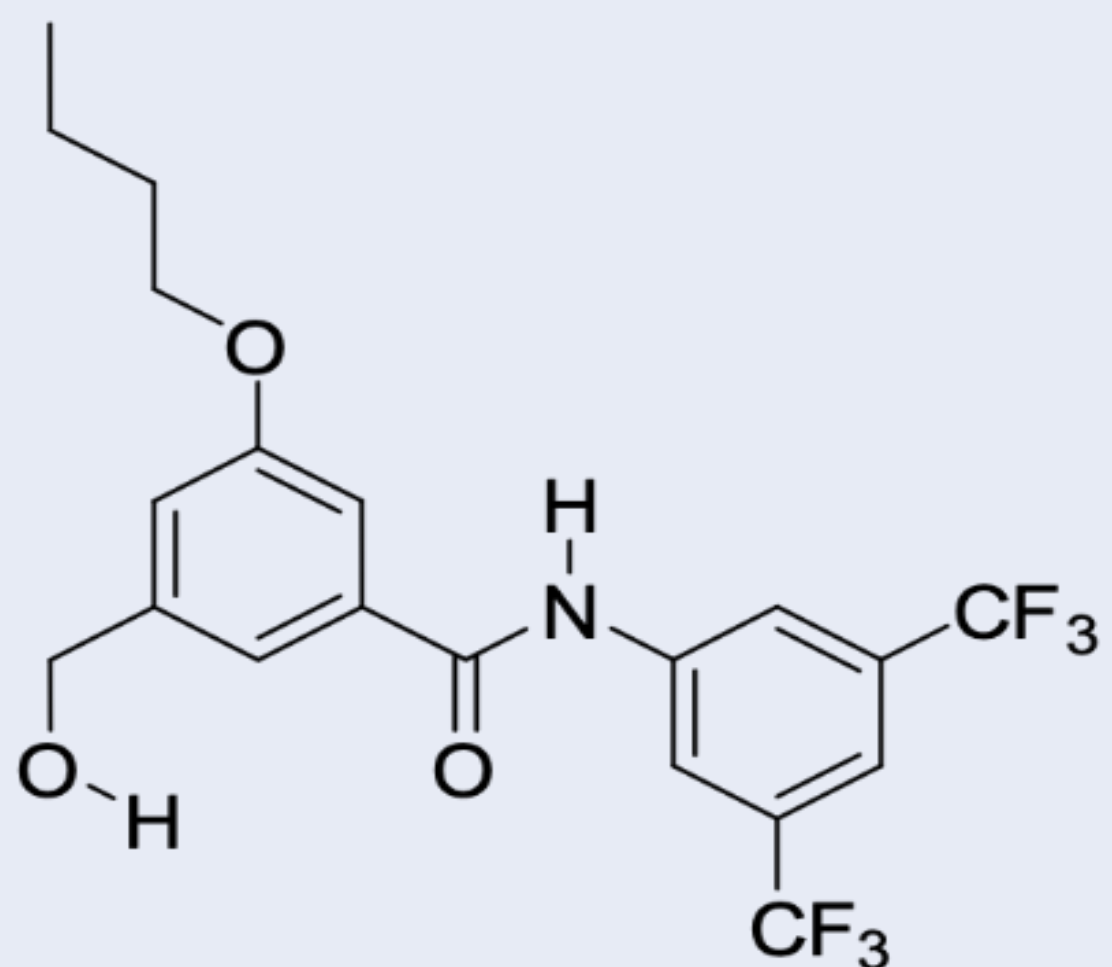
## INTRODUCTION

**CFTR and Cystic Fibrosis:** Cystic fibrosis (CF) is a genetic disease that is most common among Caucasians of Northern European origin. CF is caused by mutations in a membrane protein CFTR (Cystic Fibrosis Transmembrane-conductance Regulator) impairing its chloride ion channel function.  $\Delta F508$  is the most common CFTR mutation affecting over 70% of CF cases. Our laboratory has shown that  $\Delta F508$ -CFTR mutation can be partially reversed by physical-chemical means [1]. CFTR can be visualized in 3-electrophoretic forms in western blotting. Band-C ( $M_r \sim 160$  kDa) is a plasma membrane bound fully glycosylated form; band-B ( $M_r \sim 130$  kDa) is an endoplasmic reticulum bound unglycosylated form, and band-A ( $M_r \sim 120$  kDa) is a newly synthesized apoprotein form. Immature CFTR band-B is prominently visible in  $\Delta F508$ -CFTR expressing cell lines, whereas mature CFTR band-C is prominently visible in wild type-CFTR expressing cell lines. Apoprotein CFTR band-A is rarely visible in either cell types.

**Triazole Compounds:** Synthetic anion carriers have shown to augment the chloride ion channel function in cell lines expressing  $\Delta F508$ -CFTR [2]. Triazole, a heterocyclic type of compounds, synthesized in Talukdar lab (IISER, Pune, India), known for their ability to bind and facilitate chloride influx in cultured cell lines are used in this study to determine their effects on  $\Delta F508$ -CFTR upregulation. PJ and ABS series compounds with different chemistries can form  $Cl^-$  ion channels in cellular vesicles [3].

## MATERIALS

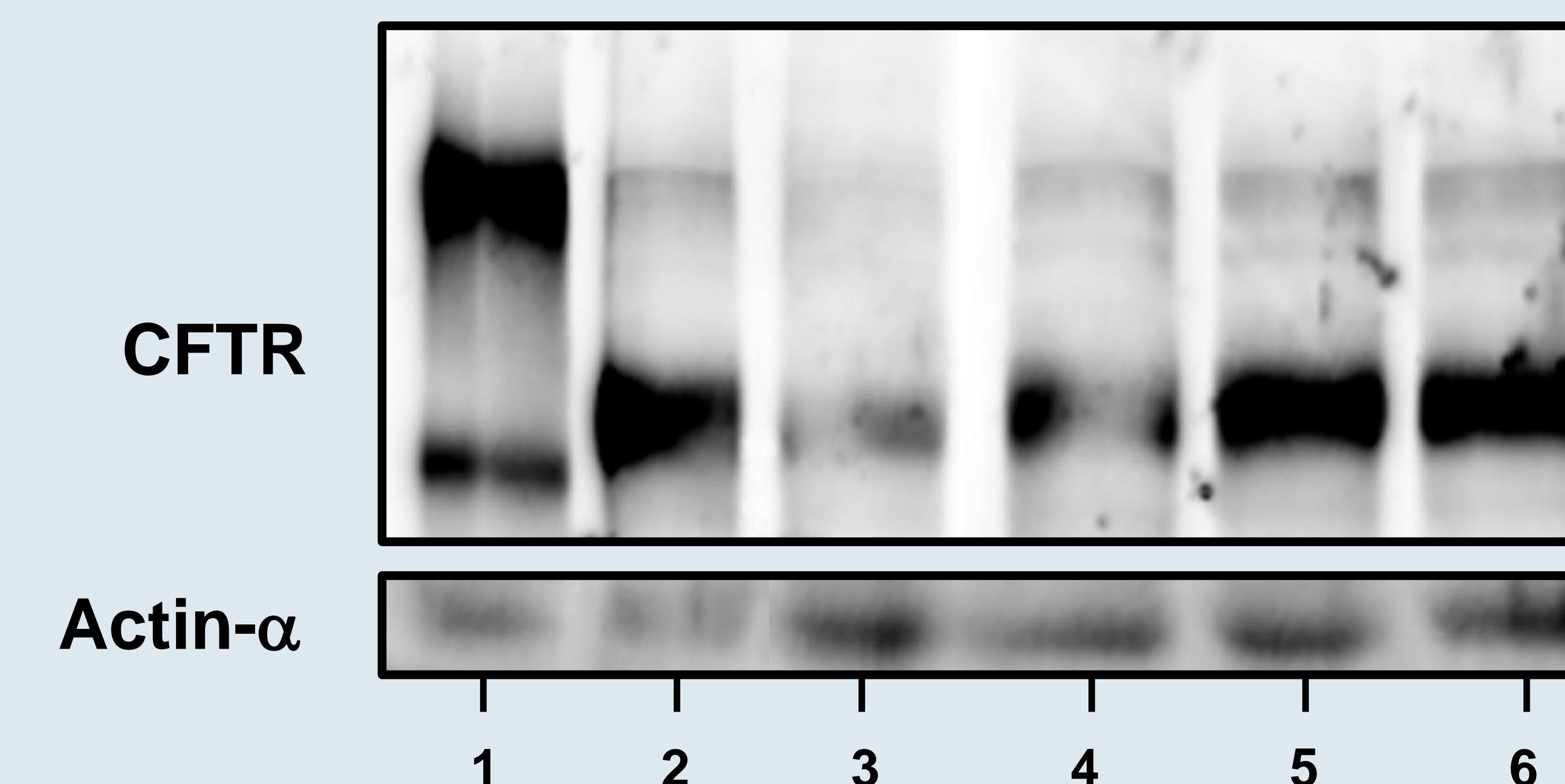
Table-I

Triazole Compounds	Properties
	<b>PJ-08</b> Triazole-((4-trifluoromethyl)phenyl)-benzamide Mr 410 Solubility: DMSO
	<b>ABS-094</b> ((3,5-bis (trifluoromethyl) phenyl) benzamide-alcohol Mr 463 Solubility: Ethanol

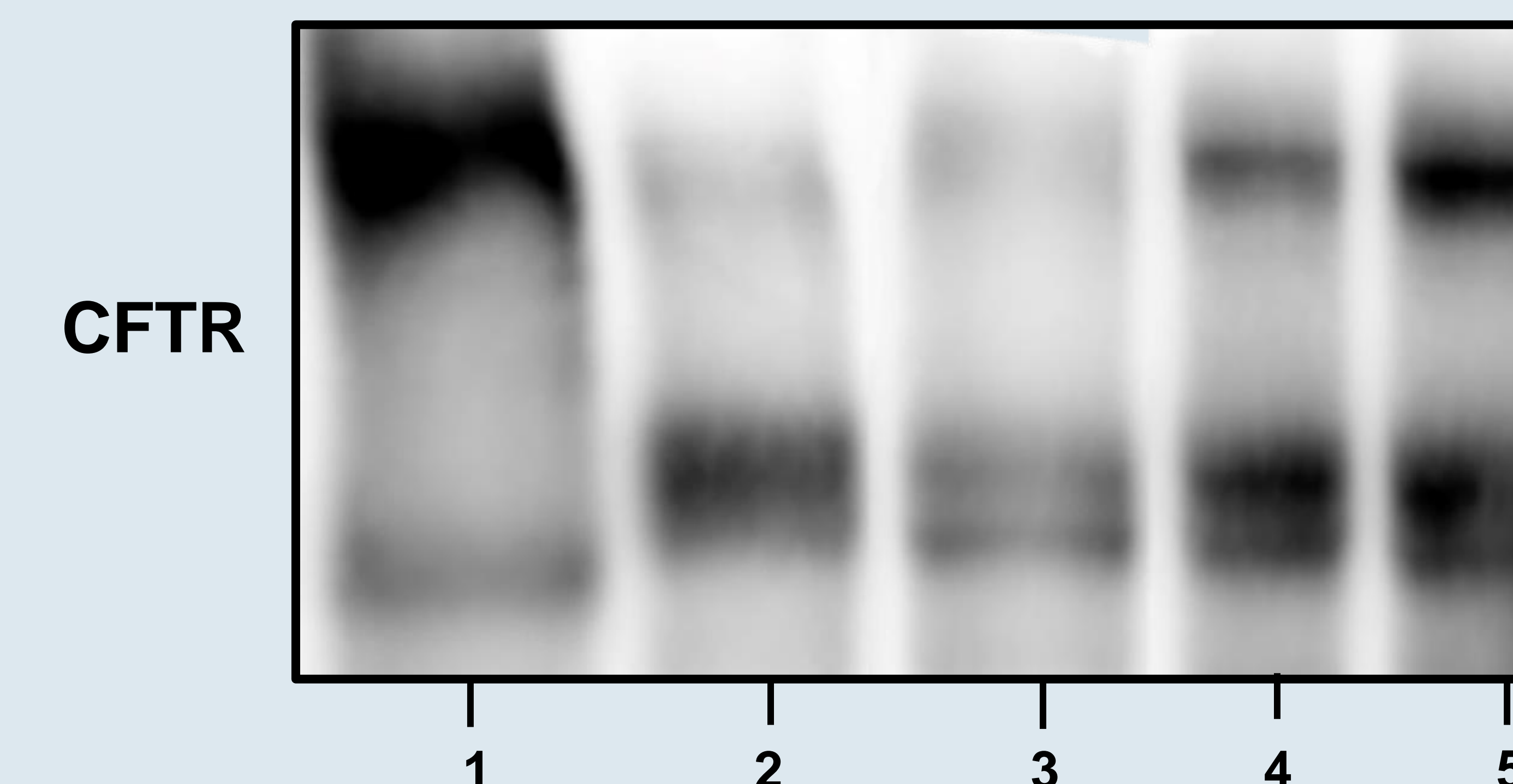
## METHODS

- CFBE cells** (a human lung epithelial cell line) transfected with  $\Delta F508$ -CFTR gene was treated with various concentrations of triazole compounds (see table-1 above).
- Positive control:** CFBE cells transfected with wild type or  $\Delta F508$  CFTR treated with 5 mM sodium butyrate [1].
- Negative control:** CFBE cells expressing  $\Delta F508$ -CFTR was treated with vehicle alone (ethanol or DMSO) to the optimal concentration.
- Treatment:** Cells were treated with the reagents when they reached  $\sim 75\%$  confluency followed by incubation at  $27^\circ C$  for 60 hrs.
- Immunoblotting:** Cell lysates were prepared, and assayed for protein concentrations. Known protein concentrations were electrophoresed on a 7.5% SDS-PAGE and transferred to nitrocellulose membranes. Expression levels of CFTR and loading control Actin- $\alpha$  was measured by chemiluminescence using c400 image analyzer (Azure Biosystems).

## RESULTS



**FIGURE 1: Effects of PJ-08 on  $\Delta F508$ -CFTR expression.**  $\Delta F508$  cells were treated with 2.5  $\mu M$  (lane 4), 5  $\mu M$  (lane 5), or 10  $\mu M$  (lane 6) of PJ-08 as described in methods. CFBE cells transfected with wild type (lane 1) or  $\Delta F508$  CFTR (lane 2 treated with 5mM sodium butyrate) were used as positive controls. Lane 3 treated with vehicle (DMSO) alone was used as negative control. Lane 4-6 are protein samples from positive controls (lanes 1,2) and 25  $\mu g$  protein samples from negative control (lane 3) and PJ-08 treated samples (lanes 4-6) were electrophoresed on a 7.5% SDS-PAGE and transferred to nitrocellulose membranes. Expression levels of CFTR and loading control Actin- $\alpha$  were measured by chemiluminescence using c400 image analyzer (Azure Biosystems). *PJ-08 treatment significantly increased immature CFTR band-B and increase in mature CFTR band-C as well when compared with the vehicle control.*



**FIGURE 2: Effects of ABS-094 on  $\Delta F508$ -CFTR expression.**  $\Delta F508$  cells were treated with ethanol alone (lane 3, as negative control), 2.5  $\mu M$  (lane 4) or 5  $\mu M$  (lane 5) ABS-094. CFBE cells expressing wild type (lane 1) or  $\Delta F508$  CFTR (lane 2 treated with 5mM sodium butyrate) were used as positive controls. Lane 3 is protein samples from positive control samples (lanes 1, 2) and 25  $\mu g$  of vehicle and ABS-094 treated samples (lanes 4, 5) were immunoblotted with anti-CFTR antibody as described in methods. *ABS-094 treatment has increased band-C and band-B CFTR in a concentration dependent manner.*

## REFERENCES

## FUTURE DIRECTIONS

## ACKNOWLEDGEMENTS