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Micro-haemodynamics at the maternal–fetal interface: experimental, theoretical and clinical perspectives

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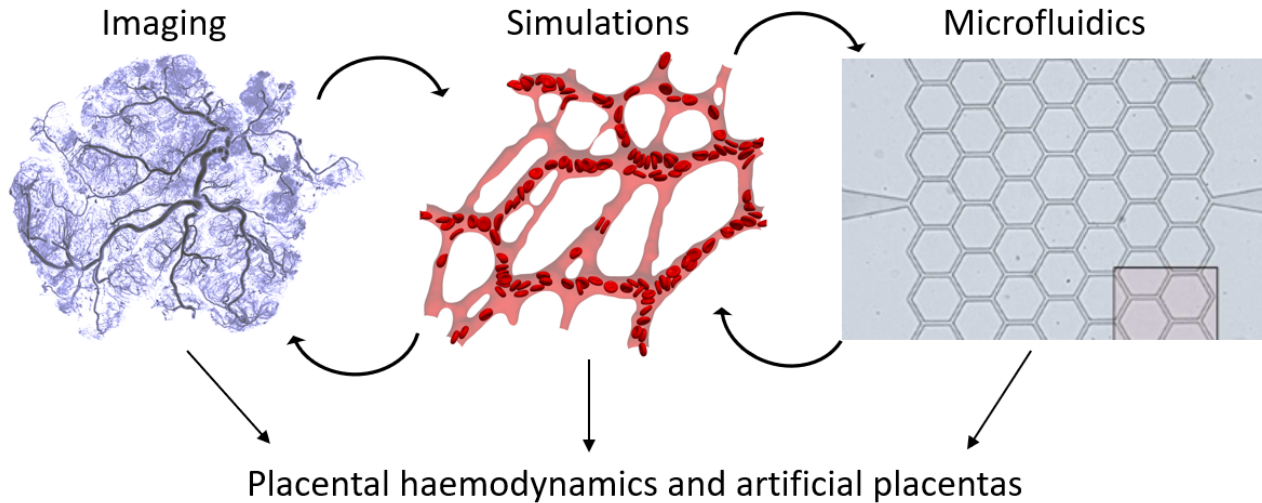
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The placenta is a vital interface between the mother and her developing fetus. Micro-haemodynamics of the placenta, where the particulate nature of blood flow cannot be ignored, mediates the relationship between the organ's structure and its function. However, the placenta's complex architecture and its relation to pregnancy pathologies remain poorly understood. This review covers current challenges in characterising placental micro-haemodynamics. Recent progress in three-dimensional multiscale imaging has stimulated development of image-based theoretical models, but existing approaches do not fully harness the available data, and new tools are needed for assimilation of complex imaging datasets. Although the placenta at term is available for *in vivo* imaging or *ex vivo* experimentation, insight into placental micro-rheology is limited, necessitating the use of biomimetic models. Microfluidic approaches offer opportunities for well-controlled characterisation of micro-rheology in complex geometries, but challenges remain in robust fabrication of these systems. Recent advances in high-performance simulations for suspension flows enable parametrisation of key physical processes at the micro-scale. Future progress can be made by optimising computational architecture and integrating micro-haemodynamics with solute transport. Both experimental and computational approaches require translation to the organ scale. New upscaling approaches will need to accommodate non-local interactions in microvascular network flows and address the lack of clear scale-separation across the placental architecture. Together, recent advances in cross-disciplinary imaging and modelling over the last ten years have opened a pathway for an *in silico* human placenta, accelerating the development of precision obstetrics medicine in the next decade.

Keywords: human placenta, artificial placenta, imaging, haemodynamics, microfluidics, computer simulations, upscaling.

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1 **A. Micro-haemodynamics in normal and complicated pregnancy**

2 The healthy development of a fetus critically depends on the maternal–fetal interface provided by the human
 3 placenta [3]. During gestation, the placenta rapidly develops into a densely packed solute-exchange system with
 4 a large surface-area-to-volume ratio [4] (Fig. 1). The maternal uterine circulation delivers nutrients and removes
 5 waste products via a heterogeneous porous placental space (also known as the *intervillous space, IVS*; Fig. 2a)
 6 interfaced with the feto-placental vascular tree (Fig. 1a,b), which is itself linked to the fetus via the umbilical cord.

7 Maternal and fetal components of the human placenta need to work synergistically to balance its multiple
 8 functions, such as robust exchange of a diverse range of solutes. In particular, the transport of oxygen and carbon
 9 dioxide is strongly facilitated by the haemoglobin of the red blood cells (RBCs) and thus depends on adequate
 10 local *haematocrit* (the volume fraction of RBCs) distribution [5]. Micro-haemodynamics of the placenta, where the
 11 particulate nature of blood flow cannot be ignored (Fig. 1c), mediates the relationship between the organ’s structure
 12 and its function. However, the role of placental architecture in pregnancy pathologies remains poorly understood,
 13 and multiple biological factors, such as oxidative- and mechanical-stress-induced damage [6], are associated with
 14 inadequate micro-haemodynamics.

15 1. Pathologies of the human placenta and the role of micro-rheology

16 Many pregnancy complications are associated with inadequate placental micro-haemodynamics and associated
 17 pathophysiology [3, 6, 7, 8]. In normal pregnancy, the shear stress at the materno-placental interface (trophoblast
 18 syncytium) is estimated to be $\lesssim 1$ Pa [4], which illustrates that in the healthy placenta, the materno-placental IVS
 19 pore space operates as a low-resistance and low-pressure flow system [3]. The endothelium in feto-placental villous
 20 capillaries is predicted to face shear stresses of similar order of magnitude [5]. Notably, both placental interfaces
 21 are characterised by considerable spatial heterogeneity [4, 5]. *Pre-eclampsia* and *fetal growth restriction (FGR)*,
 22 highly prevalent pregnancy disorders, are often accompanied by elevated blood pressure and flow velocities in the

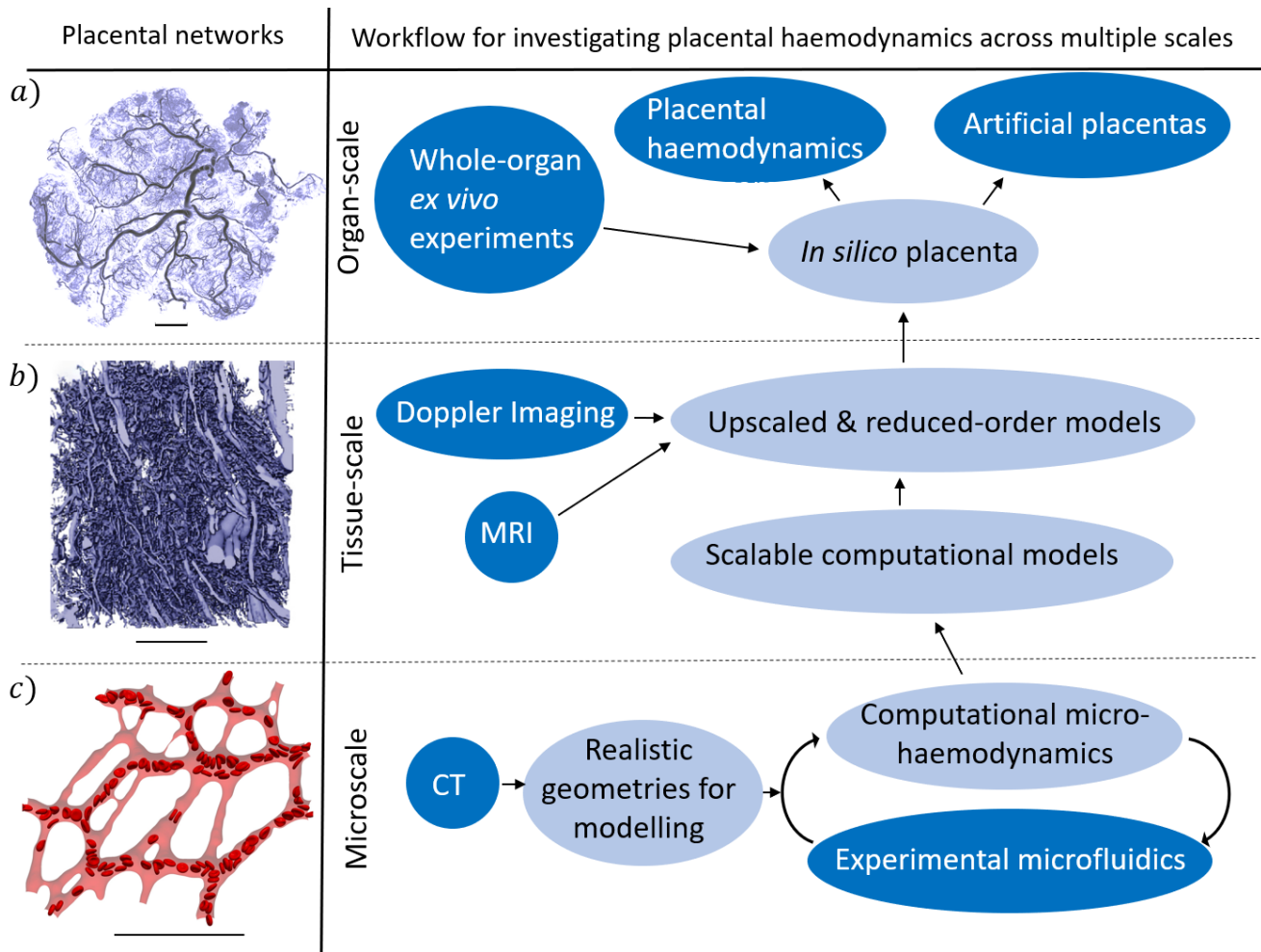


FIGURE 1. WORKFLOW FOR BIOMIMETIC MODELLING OF THE MICRO-HAEMODYNAMICS IN THE HUMAN PLACENTA. Dark blue bubbles refer to experimental methods and light blue refer to theoretical methods. (a) Whole-organ synchrotron micro-CT showing the fetal **placental** vasculature (scale bar: 25 mm; reproduced from [1]). (b) Segmented **feto-placental** vascular network from synchrotron micro-CT image (scale bar: 500 μm ; reproduced from [1]). (c) Computational simulation of red blood cells flowing through realistic microvascular networks (scale bar: 100 μm ; reproduced from [2]). **All images reproduced under CC BY 4.0.**

1 materno-placental **IVS** [3, 9]. On the other hand, the feto-placental tree and corresponding vascular network are
 2 often smaller, **with** less developed **branching structure**, in severe FGR and pre-eclampsia than in normal pregnancy,
 3 **and therefore** increasing the resistance to fetal blood flow [6]. Both structural alterations result in an environment
 4 of high mechanical stress that increases the chance of RBC *lysis* (disintegration, **associated with** the release of
 5 toxic cell-free haemoglobin [8, 10]) **and damage to the endothelial or syncytial trophoblast cellular linings** in FGR
 6 placentas (**as observed in other micro-haemodynamical systems** [11]).

7 In normal pregnancy, the growing demand of the fetus has to be balanced with maternal circulatory capacity
 8 and protection against adverse haemodynamical events for the mother and her placenta. While the production
 9 of maternal RBCs is increased during gestation, the maternal haematocrit and overall haemoglobin content per
 10 blood volume are reduced [8, 12]. At the same time, the volume of each RBC and its haemoglobin content are
 11 slightly elevated, resulting in a more spherical shape. This makes maternal RBCs potentially more susceptible

1 to osmotic stress damage, further evidenced by *microcytosis* (reduced RBC volume and haemoglobin content) in
2 pre-eclampsia as a potential adaptive response [8]. Similarly, reduced placental oxygen supply at abnormally low
3 maternal haematocrit and/or haemoglobin levels, such as in the case of maternal *anemia* (iron deficiency), could
4 be partly compensated by placental hypertrophy and increased feto-placental vascularisation [12].

5 A related set of conditions known as *haemoglobinopathies*, for example *sickle-cell* disease, also strongly impact
6 the micro-rheology of the placental blood flow. The altered shape and mechanical properties of RBCs increase the
7 incidence of IVS occlusions, haemolysis, hypoxia and associated pathophysiology [13].

8 The structure of placenta in *diabetic* patients (including gestational, Type I and Type II diabetes mellitus) can
9 often be altered in a way that is radically different from pre-eclampsia, while still leading to FGR. In particular,
10 the materno-placental IVS is less sparse in diabetes than in normal (and much less than in pre-eclamptic) placental
11 tissue, while the feto-placental network is less mature but more hyper-vascularised at the exchange interface [7].
12 Future studies should further evaluate the relative contributions of altered placental micro-architecture, RBC shape
13 and mechanical properties to the pathophysiology of placental haemodynamics and solute transport.

14 2. The need for biomimetic *in vitro* and *in silico* models

15 To facilitate early diagnosis of developing placental dysfunction during pregnancy, a mechanistic understanding of
16 the relationship between placental structure and function is required, which is mediated by micro-haemodynamics
17 at the intricate maternal–fetal interface [14]. Despite recent progress in advanced 3D microscopy of placental
18 architecture (Fig. 1b), current clinical *in vivo* imaging and physiological *ex vivo* perfusion experiments lack the
19 resolution needed for functional assessment of the placental micro-haemodynamical environment [15]. The high
20 evolutionary divergence of placental anatomy and physiology makes common animal models (*e.g.* mouse, rat and
21 sheep) unsuitable for reliable inferences about the human placenta [16], and the use of non-human primates is
22 largely inaccessible due to ethical and cost considerations. However, emerging rich imaging datasets pave the way
23 to advanced biomimetic modelling, either *in vitro* or *in silico*, which allows for exhaustive testing of hypotheses
24 regarding the interplay of placental microstructure and micro-haemodynamics [15].

25 **B. Assimilating complex heterogeneous geometries into models**

26 To develop biomimetic models of placental micro-haemodynamics, we must first establish robust and efficient
27 workflows to register and characterise representative tissue geometries.

28 1. Robust segmentation and characterisation of micro-geometry for image-based modelling

29 The human placenta presents a unique challenge as a highly heterogeneous and dynamic organ, requiring mas-
30 sively multiscale and multi-modal imaging to quantify its function [17, 18]. Additionally, the feto-placental vascular
31 space, materno-placental IVS and the villous tissue barrier comprise three distinct domains which must be distin-
32 guished within imaging modalities for accurate reconstruction of placental micro-architecture. Progress has been

- *Microstructure*: Robust and efficient **experimental and computational** pipelines are needed for sample preparation, imaging, segmentation and statistical characterisation.
- *Boundaries*: Physiological boundary conditions must be extracted at the tissue level, accounting for RBC–RBC and RBC–surface (in particular, the syncytial microvillous surface) interactions.
- *Microfluidics*: New **experimental** methods are needed for micro-fabrication of biomimetic capsules and complex three-dimensional micro-architectures.
- *Simulations*: Cell-scale flow and transport simulators must be integrated and optimised, with respect to spatio-temporal decomposition and hybrid GPU/CPU parallelisation.
- *Reduced-order modelling*: New mathematical models are needed for nonlinear and non-local transport in disordered networks and porous media.
- *Clinical imaging*: Tissue-scale haemodynamics must be linked to MRI and Doppler ultrasound physics, accounting for local haematocrit heterogeneities.
- *Artificial placenta*: Design of robust low flow-resistance and high flux oxygenators needs to be optimised with the help of biomimetic models.

made recently using *ex vivo* synchrotron X-ray tomography with novel contrast agents or *in vivo* MRI techniques, in combination with **machine-learning-based** segmentation algorithms, such as **U-net** [1, 19]. Multiple challenges however remain in the robust preparation of samples that preserve their 3D morphology and in quality-assured and efficient structural image analysis **that accurately captures geometrically complex maternal and fetal placental domains**.

Characterisation of placental microstructure, once obtained, is another challenge shared with other complex heterogeneous disordered media. A combination of topological data analysis, spatial probability and statistical physics approaches enable increasingly deep insights into the role of microstructural fluctuations for flow and transport in disordered porous media [20] or networks [21]. Nevertheless, much remains unknown about robust and optimal strategies for identifying representative volumes of placental tissue, given its heterogeneity and markedly different spatial scales of the maternal and fetal placental domains [1]. Future progress in computational modelling and biomimetic microfluidics will be enabled by generating more accurate synthetic porous media and vascular networks that match statistically the placental geometry.

2. Tackling boundary condition uncertainty in micro-haemodynamical models

Even with accurate characterisation of the placental microstructure, micro-haemodynamical models depend strongly on tissue-scale boundary conditions, which are often uncertain. Furthermore, the two principal circulatory domains in the human placenta, the maternal IVS and the fetal villous capillary networks, require different modelling approaches. For the former, blood flow in a heterogeneous IVS can be modelled by assuming a tissue-scale pressure gradient [1] (Fig. 2a). For the latter, boundary conditions at network inlets and outlets need to be prescribed

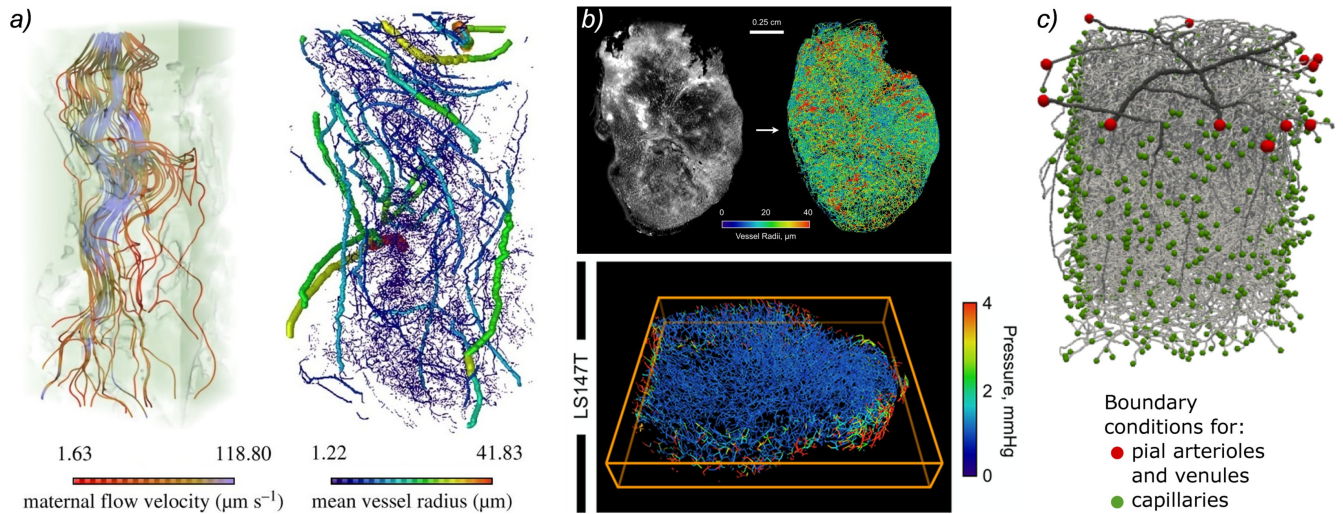


FIGURE 2. ASSIMILATION OF COMPLEX BIOLOGICAL GEOMETRIES INTO COMPUTATIONAL MODELS. (a) Three-dimensional flow through the maternal intervillous porous space (IVS; left) embedded in the fetal vascular network (right) of human placental tissue (reproduced from [1]). (b) Image segmentation (top) and intravascular pressure analysis (bottom) of human colorectal carcinoma xenograft (reproduced from [22]). (c) Hierarchical boundary conditions applied to the microvascular networks of mouse parietal cerebral cortex (reproduced from [23]). All images reproduced under CC BY 4.0.

iteratively to match physiological ranges for blood pressure, haematocrit and wall shear stress, borrowing modelling approaches from other biological networks, such as vascular tumours [22] and cerebral cortex [23] (Fig. 2b,c).

In both domains, the model boundary conditions and physiological target ranges rely on *in vivo* or *ex vivo* measurements, with their associated uncertainties, arising from individual and regional variability due to the inherent heterogeneity of placental microstructure and the resolution limit of measurement protocols [15, 24]. Coupling the two circulatory domains across a complex placental barrier also presents many open challenges. In particular, future models will need to address capillary- and pore-scale boundary conditions due to passive, facilitated and active transport of solutes at the villous syncytial and endothelial sides of the barrier, which are likely to be spatially varying and solute-specific [4].

C. Biomimetic *in vitro* and *in silico* models of the placental microcirculations

Placental haemodynamics shares important features with circulation in other microvascular networks [25]. Because placental capillaries and pores are of similar size to individual RBCs, cell-scale blood flow needs to be resolved for a haemodynamical model to account for microcirculatory phenomena, such as geometry-induced haematocrit bias and oxygenation heterogeneity [26]. Placental haemodynamics is akin to a suspension flow in a porous medium (i.e. the flow of a heterogeneous mixture of RBCs and other blood constituents), an emerging research topic with many open fundamental questions. On the other hand, there is growing interest in biomimetic micro-engineered ‘placenta-on-a-chip’ systems that could provide more accurate placental drug transport and toxicology models [16] (Fig. 3a). However, there is a paucity of *in vitro* and *in silico* studies of cellular blood flow in placenta-specific geometries [4, 14].

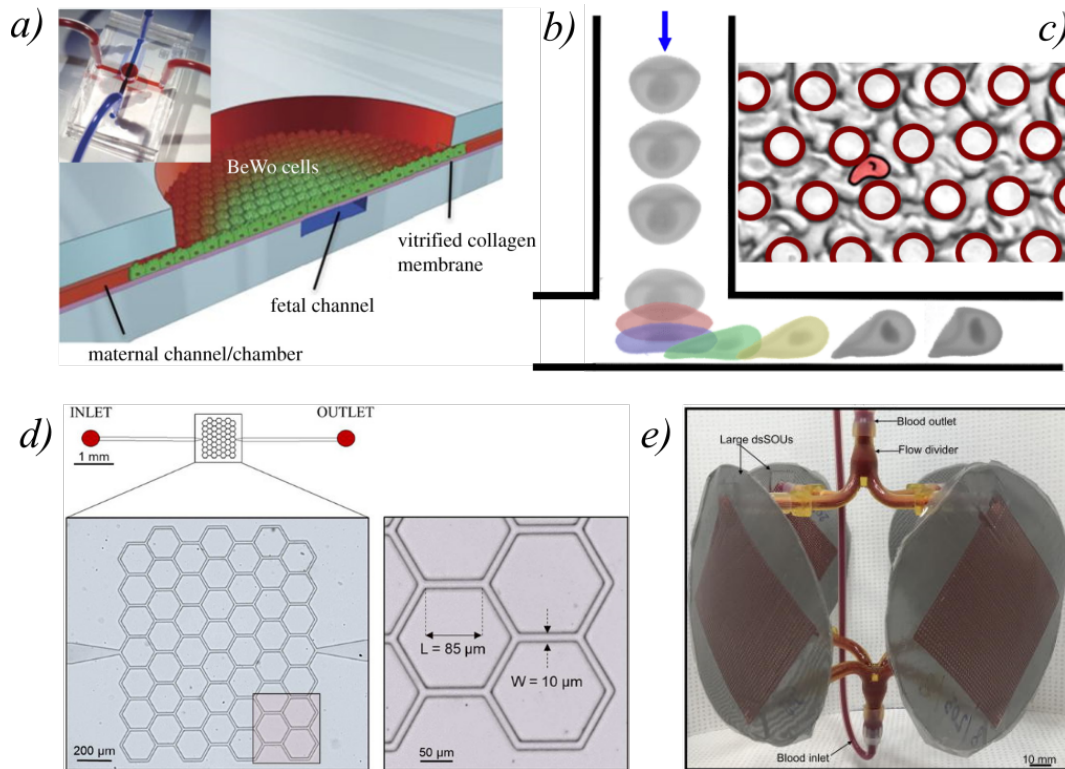


FIGURE 3. BIOMIMETIC EXPERIMENTAL MICROFLUIDICS AND ITS APPLICATIONS. (a) A ‘placenta-on-a-chip’ micro-engineered device for placental transfer analysis (reproduced from [27], CC BY 4.0). (b) Time-lapse image of an alginate capsule deforming at a T-junction (reproduced from [28], with the permission of Cambridge University Press). (c) A microfluidic model of blood flow in a regular porous medium, with one labelled deformed RBC (red) and marked posts (red circles; adapted from [29], with the permission of AIP publishing). (d) A microfluidic model of blood flow in a biomimetic capillary network (reproduced from [30], with the permission of AIP publishing). (e) Prototype neonatal life-support ‘artificial placenta’ device (reproduced from [31], with the permission of AIP publishing).

1. Experimental microfluidics of suspension flow in complex geometries

Multiple research fields can inform micro-scale blood flow in the human placenta. Other microvascular systems, such as lung capillary networks [29], share common micro-haemodynamical phenomena [25, 30]. Microfluidic sorting devices [32] use complex geometries to direct the flow of cells and particles. Two-phase flows in porous media in the oil-recovery context, while often relating to imbibition/drainage problems, also include emulsion flows [33]. Pore-scale models motivated by subsurface flow problems address the influence of pore-scale geometry on macro-scale flow parameters [34].

There are two experimental avenues for the exploration of haemodynamics in complex geometries like the placenta. Firstly, whole blood or diluted RBC suspensions can be transported through biomimetic porous-medium-like [29] or capillary-network-based [30] artificial structures, thus enabling measurements under well-controlled flow conditions (Fig. 3c,d). These measurements have the advantage of retaining biological features related to RBC variability and interaction, but flow conditions need to be carefully monitored to keep the RBCs in a physiological state. Secondly, control over particle properties in addition to flow conditions requires RBC analogues. This

1 approach can reveal the underlying physics of the suspension flow but largely bypasses biological effects. The main
2 challenge is to create physical objects that have biomimetic properties that closely reproduce the mechanics of RBCs
3 in suspension. Potential candidates are elastic beads [35], droplets, vesicles and capsules [36]. While elastic beads
4 offer limited deformability under laboratory flow conditions, droplets and vesicles can exhibit large deformations
5 but they lack the RBCs' shear elasticity. Liquid droplets encapsulated by an elastic membrane offer the closest
6 analogue to RBCs, but properties like membrane thickness, material properties and inflation need to be finely tuned
7 to access the deformations observed in RBCs (Fig. 3b).

8 The micro-fabrication of complex porous media presents additional challenges. In microfluidics, polydimethyl-
9 siloxane (PDMS) is widely used for moulding complex geometries from negatives created by photolithography or
10 micro-milling [37]. These techniques can produce versatile planar and layered geometries but are less suitable for
11 three-dimensional placental geometries. Future prospects of rapid prototyping of 3D architectures are provided by
12 advances in 3D printing technology [38].

13 2. Computational models of micro-haemodynamics

14 Cell-resolved computational models of microscopic blood flow in sparse and complex geometries [2, 39, 40]
15 have recently provided access to high-resolution flow features (e.g. local shear stresses and pressure gradients in
16 microvasculature), which are very difficult to measure reliably in conventional experiments. These computational
17 models are readily translatable to human placental micro-haemodynamics. Such models rely on efficient spatial and
18 temporal decomposition of complex domains into subsystems for parallel computing and have achieved physiological
19 haematocrit levels (40–60% [39, 40]) in large-scale microvasculature, capturing key RBC features, such as cell
20 deformation and dynamics.

21 Nevertheless, the uncertainty in prescribing inflow/outflow boundary conditions in micro-haemodynamical mod-
22 els (see Section B) requires additional steps, such as network-scale simulations with simplified blood rheology, to
23 provide input data [2, 23] (Fig. 2c). For cell-scale models to faithfully reproduce physiological transport processes
24 in the human placenta, multi-scale biophysical and biochemical processes (including blood coagulation, cell–cell
25 and cell–surface interactions) need to be incorporated without compromising computational efficiency. Also, longer
26 simulations and larger computational domains are needed to match the realistic temporal and spatial scales over
27 which biological processes occur [39].

28 3. Consideration of interfacial features

29 Thus far, key interfacial micro-/nano-structures at the maternal–fetal interface have been mostly neglected in
30 *in vitro* or *in silico* models. These include the microvilli on the materno-placental trophoblast syncytium layer [24]
31 and the glycocalyx on the endothelium of fetal capillaries [25]. The influence of glycocalyx on the flow of RBCs
32 [41] and the transport of substances across the placental endothelium and trophoblast syncytium remains poorly
33 understood [16]. Recent progress in growing three-dimensional ‘organoids’ from placental trophoblast cells *in vitro*
34 [42] opens further opportunities for characterising the maternal–fetal interface. With the increasing availability

1 of high-resolution imaging, e.g. transmission electron microscopy, for the placenta [24], a future challenge is to
2 incorporate these interfacial features into blood flow models. This goal may be achieved experimentally by coating
3 channels with cultured trophoblast cells [27], endothelial cells [43] or polymer brushes [44]. However, the disparity
4 in scales, ranging from nanometres for glycoproteins to micrometres for RBCs, hinders physiologically realistic
5 representation of these features in numerical models.

6 **D. Scaling up models and simulations**

7 Due to strong spatial heterogeneity arising from both topological and micro-haemodynamical variations, any
8 microscopic model is likely to be computationally prohibitive at the placental tissue scale. Tractable testing of
9 physiologically relevant hypotheses could be enabled through a combination of optimised computational strategies
10 and mathematical model-reduction approaches.

11 **1. Code accessibility and scalability**

12 Large-scale parallel simulators of cellular haemodynamics in complex and sparse geometries are primarily based
13 on flow solvers using the lattice-Boltzmann method [2, 40] or dissipative particle dynamics [39, 45]. Several computa-
14 tional models are available as open-source research software, such as HemeLB [2], and Mirheo [45], which demonstrate
15 excellent scalability and can run on hundreds to thousands of distributed computational (CPU) nodes. Optimisation
16 of the load balancing of computational nodes improves code scalability in simulations of heterogeneous microvas-
17 cular networks [46]. Nevertheless, the computational efficiency of conventional spatial decomposition schemes is
18 limited by the CPU node communication rate, and efforts have been focused on developing a hybrid parallelism that
19 combines temporal decomposition (computed via GPU) with spatial decomposition [39]. However, even optimised
20 microscopic flow and transport simulators require extensive model parametrisation, uncertainty quantification and
21 validation against tissue- and organ-scale observations, which are not feasible without systematic model-reduction
22 approaches.

23 **2. Reduced-order modelling and upscaling**

24 A possible solution to harnessing the computational complexity of placental micro-haemodynamics is to imple-
25 ment reduced-order modelling or upscaling techniques. A classic example in which experimental data have been
26 used to inform a reduced-order model is the work of A. R. Pries and T. W. Secomb, who established a set of empirical
27 laws for haematocrit transport through capillary networks [25]. This work is widely used but may not be applicable
28 to maternal flow in the IVS. Upscaling techniques for continuum models over microstructured tissues have been
29 used extensively to investigate tissue properties, often drawing on theoretical developments driven by geophysical
30 applications. These techniques (reviewed by [47]) largely hinge upon identifying an RVE (*representative volume*
31 *element*). In a popular method of upscaling, asymptotic homogenisation, RVEs are often assumed to be organised

1 in a periodic array. For a highly heterogeneous and disordered tissue of the human placenta, the assumption of pe-
2 riodicity does not apply, and identifying the features of an RVE is an open and contentious question. Furthermore,
3 spatial scale separation is typically less strong in biological tissues (tens of microns to millimetres) compared to
4 geophysical subsurface applications (nanometres to kilometres). This ultimately restricts the applicability of many
5 approximation methods, meaning new approaches, such as stochastic homogenisation [48] or generalized multiscale
6 finite element methods [49], must be used to construct upscaled models of the placental flow and transport.

7 Alternatively, network models are commonly employed to investigate the effects of spatial disorder on flow
8 and transport in complex media [34]. Spectral graph theory (which decomposes complex networks according to
9 their topology and physical properties) has shown promise for effective model-reduction and characterisation of
10 heterogeneity in lung airways [50], an approach which could be adapted for the human placenta. However, any
11 possible network model for the placenta will be fundamentally different to those already constructed for other organs
12 (such as brain or tumour vasculature [22, 23]) due to differences in network topology. Specifically, developing a
13 coupled model for two distinct placental circulations, the maternal pore network and the fetal vascular network,
14 remains an open challenge. More work is also needed to understand fundamental mechanisms for suspension flows
15 in disordered geometries, such as non-local transport effects [49] and haematocrit heterogeneity [23], in order to
16 inform the development of tissue- and organ-scale models.

17 E. Emerging diagnostics and therapies for precision obstetrics medicine

18 Advances in multiscale *ex vivo* imaging, *in vitro* and *in silico* biomimetic micro-haemodynamical modelling
19 enable more mechanistic understanding and interpretation of clinical imaging (such as Doppler sonography and
20 MRI). Likewise, ‘reverse-engineering’ the fundamental building blocks of the human placental micro-circulation can
21 help devise new therapeutic strategies in pregnancy complications and optimise the design of adequate biomimetic
22 replicas for clinical applications.

23 1. *In vivo* imaging and management of placental haemodynamics

24 Doppler ultrasound has been used in maternity care for more than three decades following the recognition that
25 the umbilical artery Doppler waveform is different in pregnancies affected by placental dysfunction [14]. Until
26 recently all assessments have been made on the basis of the relative peak of velocity compared to diastolic velocity
27 to create either a *pulsatility index* or *resistance index* with gestationally-dependent reference ranges created from
28 low-risk pregnancies. Intra-placental Doppler may also have a role in delineating normal placentas from those
29 with dysfunction [14], but existing resolution constraints potentially limit the applicability of this technique in
30 the clinical assessment of placental micro-rheology. Furthermore, the impact of local haematocrit fluctuations in
31 a highly irregular placental IVS, spiral uterine and helical umbilical arteries on the Doppler signal remains to be
32 quantified. There has also been a growing recognition that improving the understanding of why and how Doppler
33 waveforms change will lead to improved risk stratification and more individualised clinical care [14, 51].

34 Placental magnetic resonance imaging (MRI) offers the potential to expand *in vivo* imaging of the placenta

beyond what Doppler ultrasound can offer by providing information on flow at a microstructural level [9] and oxygenation status, using functional MRI (e.g. blood-oxygenation-level-dependent (BOLD) T_2^* imaging sequence) [52]. However, the spatial resolution of these modalities, which is generally in the millimetric range [18], is still a limiting factor. Structural MRI T_1 and T_2 maps have shown some promise in the early diagnosis of placental dysfunction, but the relationship of data obtained to the placental micro-haemodynamics and postnatal histopathological findings has yet to be fully determined.

Little is known about placental micro-haemodynamics in early pregnancy. For example, one important open question is the interaction of blood flow with porous trophoblast plugs in the spiral arteries that limit the maternal placental circulation until the second trimester, and whose abnormal dynamics is associated with pre-eclampsia and other pathologies [14]. Likewise, placenta-associated bleeding and abruption are challenging to diagnose and characterise pathophysiologically [53].

Better understanding of how micro-haemodynamics and biochemistry interact with complex utero-placental geometries will also inform future therapies and facilitate precision-medicine interventions. In particular, early establishment of normal and abnormal placental circulation in pregnancy (aided by upscaled biomimetic *in silico* and microfluidic models) could enable differential therapies to be administered to reduce the severity of FGR and associated pregnancy pathologies [3].

2. Artificial placentas for neonatal support

With the number of pre-term births increasing, considerable attention has been paid to develop critical life-support technology for neonates. In the event of a pre-term birth, the lungs are only partially developed and cannot meet oxygenation needs provided by the placenta *in utero*, often leading to respiratory insufficiency and other conditions that increase neonatal morbidity and mortality.

Several technologies have been developed that aim to mimic the support received by the fetus inside the mother's body, either fully or partially. One class of technology, termed as *artificial womb*, aims to replicate the oxygenation, thermal-control and nutrient-supply functions of the human placenta [54]. An artificial womb is important for pre-term neonates bordering on viability who require a stable extra-uterine environment. Pre-term lamb fetuses (105–130 days gestational age, GA) were maintained in a biobag consisting of artificial amniotic fluid and catheters that connected the fetus to an external pumpless oxygenator, nutrient supply and waste removal device through the umbilical vessels. The device was able to sustain the fetuses physiologically for up to four weeks. Even smaller pre-term lambs (GA 95 days) were sustained for up to five days and shown to have a stable and normal stage of development [55].

The other class of technology known as *artificial placenta* is for more mature pre-term neonates where respiratory distress is common. In this scenario, oxygenation support in a biomimetic fashion is desired in order to avoid the complications associated with mechanical ventilation. Artificial placental devices consisting of hollow-fiber membrane oxygenators have maintained pre-term lambs (GA 118 days) through a venous–venous connection for over ten days, which enabled the lungs to develop in a normal manner and protected them from injuries associated with ventilation support. Here, a pump is often used to extract the blood and perfuse the oxygenator. In order

1 to extend this technology to smaller neonates, with smaller size and blood volume, alternative microfluidic designs
2 have been considered [31, 56] (Fig. 3e). The advantage of a microfluidic device is that it can be precisely designed to
3 operate not only in a pumpless manner, but also to avoid stagnation zones and high-shear regions where thrombotic
4 reactions can occur. Very small channel dimensions similar to blood capillaries in the lungs or the placenta, as
5 well as extremely thin gas perfusion membranes, are possible in the microfluidic format. Recently, this format has
6 rescued piglets of the size and blood volume of human neonates from respiratory distress [57], consistently increasing
7 oxygen saturation in the animal from 50–60% to above 80%. This proof of concept in animal models, combined
8 with recent progress in biomimetic *in vitro* and *in silico* models, shows promise for translation of artificial placenta
9 technology into human trials and for eventual approval and use in human pre-term neonates.

10 F. Concluding remarks

11 Modelling of flow and transport in the human placenta shares some common challenges with other complex
12 biological systems, which include robust extraction and characterisation of the microstructure and identifying ap-
13 propriate boundary conditions. However, the intertwining of the porous IVS with the irregular fetoplacental villous
14 capillary network distinguishes the human placenta not only from other exchange organs but also from the placentas
15 of other non-primate species.

16 Future progress in the field is expected by a combination of well-controlled biomimetic microfluidics, hybrid and
17 highly parallelised computational micro-haemodynamics and systematic upscaling of these high-resolution models
18 to the organ or device scale.

19 In addition to the important biomedical and clinical applications in fetal and neonatal medicine, deeper un-
20 derstanding of suspension flows in complex geometries will address many fundamental questions and stimulate
21 development in other areas of science and engineering.

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25 Author contributions

26 QZ, ED, KS & QC prepared visualisations; ILC prepared a list of challenges; QZ, ES, TK & ILC contributed
27 to the first draft of Section A (placental physiology); QZ, ED, MOB, TK & ILC contributed to the first draft of
28 Section B (geometry assimilation); QZ, KS, QC, AJ & TK contributed to the first draft of Section C (microfluidics);
29 QZ, ED, OEJ, TK & ILC contributed to the first draft of Section D (upscaling); EDJ, PRS & ILC contributed to
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31 responsible for overall coordination. All authors read and approved the final manuscript.

1 Competing interests

2 The authors declare that they have no competing interests.

3 Original papers of particular interest, published within the period of review, have been highlighted as:

4 * of special interest, ** of outstanding interest.

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Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.