Check for updates

#### **OPEN ACCESS**

EDITED BY Yonghao Xu, Institute of Advanced Research in Artificial Intelligence (IARAI), Austria

REVIEWED BY Zhijie Wang, Guizhou University, China Ion Catalin Petritan, Transilvania University of Braşov, Romania

\*CORRESPONDENCE Meiqi Jiao ⊠ mqjiao@126.com Zhongke Feng ⊠ fengzhongke@bjfu.edu.cn Zhichao Wang ⊠ Zhichao@bjfu.edu.cn

SPECIALTY SECTION This article was submitted to Environmental Informatics and Remote Sensing, a section of the journal Frontiers in Ecology and Evolution

RECEIVED 28 November 2022 ACCEPTED 28 February 2023 PUBLISHED 31 March 2023

CITATION

Lu D, Qiu L, Jiao M, Feng Z and Wang Z (2023) Quantitative analysis of the spatial diversity of Moraceae in China. *Front. Ecol. Evol.* 11:1110018. doi: 10.3389/fevo.2023.1110018

#### COPYRIGHT

© 2023 Lu, Qiu, Jiao, Feng and Wang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Quantitative analysis of the spatial diversity of Moraceae in China

Dangui Lu<sup>1</sup>, Lichuan Qiu<sup>2</sup>, Meiqi Jiao<sup>3</sup>\*, Zhongke Feng<sup>1,4</sup>\* and Zhichao Wang<sup>1</sup>\*

<sup>1</sup>Precision Forestry Key Laboratory of Beijing, Forestry College, Beijing Forestry University, Beijing, China, <sup>2</sup>School of Geographical Sciences, Taiyuan Normal University, Jinzhong, China, <sup>3</sup>Department of Hospitality Management, Shanghai Business School, Shanghai, China, <sup>4</sup>College of Forestry, Hainan University, Haikou, China

Changes in distribution patterns of economically essential forest species under global change are urgently needed in the scientific forecast, and large-scale spatial modeling is a crucial tool. Using diversity pattern indicators and other data obtained through geographic information systems (GIS) and spatial data on Moraceae species obtained from published data, we quantitatively studied the spatial diversity patterns of genera in the Moraceae in China. The results revealed that the patch richness, diversity index, and total shape index of the genera with multiple species were significantly higher than those of the monotypic genera. Monotypic genera had no spatial diversity and no distribution in patterns of spatial diversity. Maclura had the most concentrated spatial distribution and the lowest distribution area among the Moraceae in China. The number of patches and the total area were the smallest, while the most significant patch index was the highest. Maclura had no spatial diversity. Streblus had the highest patch abundance compared to other genera with fewer species. Streblus had the smallest number of patches and total area of distribution, the lowest spatial distribution, and a small total shape index, indicating its concentrated distribution. The values of the Shannon's Diversity Index (SHDI) and Simpson's Diversity Index (SIDI) were the highest, and the spatial distribution was the most diverse among the genera with fewer species. The patch type of Streblus had a more considerable value than other genera, but the number of patches was small, and the total shape index was low. Streblus was primarily distributed in the south of Yunnan, western Guangxi, the west and central parts of Hainan, and southern Guangdong. Most of these areas were mountainous. The temperature decreased with elevation, providing diverse environmental conditions for the narrow-stem genus. Among the Moraceae in China, the spatial distribution of Ficus was the most diverse, with the highest number of patches, patch types, total shape index, SHDI, and SIDI values. The spatial diversity of Ficus could be used as a protected area for Moraceae in China.

KEYWORDS

Moraceae, spatial diversity, China, geographic information systems, genus

## 1. Introduction

Large-scale patterns of species diversity are not only one of the central issues in macroecological and biogeographical studies (Colwell and Lees, 2000). In modern China, Moraceae includes 12 genera: *Antiaris, Artocarpus, Broussonetia, Cannabis, Cudrania, Fatoua, Ficus, Humulus, Maclura, Malaisia, Morus*, and *Streblus*. The diversity of Moraceae has been

studied in terms of genetic diversity (Elhawary et al., 2018; Marcotuli et al., 2019; Martins et al., 2021), species diversity (Gardner et al., 2017; Machado et al., 2018; Shi et al., 2018; Pederneiras et al., 2020), and ecosystem diversity (Kong et al., 2020; Dong et al., 2022; Wang et al., 2022). Most studies on diversity in the Moraceae have been conducted for limited species of the genus Morus (Chen et al., 2016; Islam and Rahman, 2019; Hashemi and Khadivi, 2020) and Ficus species (Teixeira et al., 2018; Zhang et al., 2018; Chakraborty et al., 2022). On the contrary, in this work, all genera in the Chinese Moraceae were applied to investigate species diversity. Concerning research methodology for the study of the Moraceae in China, tabular statistics are frequently applied in studies to document plant species diversity (Chen et al., 2010), which has the disadvantage of requiring a great deal of work in discovering interprovincial relationships among species diversity. With the use of geographic information systems (GIS) technology, it is feasible to observe species diversity, i.e., spatial diversity, among regions. For now, the limited spatial distribution of some genera and species of Moraceae has been studied by researchers utilizing GIS (Berg, 1989). However, studies on the spatial distribution and diversity of all Moraceae across China are insufficient. Furthermore, studies on the species diversity of Moraceae are usually conducted using qualitative methods, and quantitative studies are insufficient (Zerega et al., 2005; Kumar et al., 2011). In this paper, we used GIS to study the species diversity of all Moraceae species in all of China. This was done because of these problems.

A quantitative study of species diversity in communities can utilize the Shannon-wiener index, which integrates species richness and evenness, combined with the Simpson index (Nagendra, 2002; Ganivet et al., 2020; Shukla et al., 2020), the Pielou evenness index (Pielou, 1966; Ricotta and Avena, 2003; He et al., 2019), and the Jaccard similarity index (Oluyinka Christopher, 2020). The index formula of spatial diversity patterns, such as the number of patches and total area, was applied to compute the index of spatial diversity patterns of the genus Moraceae in China. However, all of them lacked the ability to link with geoinformation to provide national-scale knowledge. In this research, GIS was applied to map the spatial distribution of 12 genera of Moraceae in China, with the county as the fundamental unit of spatial data and the Moraceae as the research object.

Large-scale vegetation distribution data can be obtained using remote sensing images, and landscape diversity indicators can analyze the spatial distribution pattern of vegetation. The Moraceae are primarily dominated by trees and shrubs, with a small number of vines. When the Moraceae building species are evident, the spatial pattern of the dominant species in the primary layer can be obtained using remote sensing technology. When the prevalent phenomenon of Moraceae is not evident, the spatial pattern does not contain species. However, species are the basis of plant community composition, and the workload of a species survey is immense. The published botanical histories of various places in China have the distribution sites of Moraceae, which lay the foundation for the study of the spatial distribution of Moraceae. Studying how species are spread out in space can help protect species and give a guide for figuring out what information about plants is in high-resolution remote sensing images.

In this study, GIS was used to create a map of the spatial diversity of 12 Moraceae genera in Chinese, using counties as the basic spatial data unit and Moraceae as the research object, from which not only the spatial distribution characteristics of the genera, but also further indicators of spatial diversity calculation, such as the number of patches and patch richness, can be obtained, providing a foundation for its quantitative study of spatial divergence. The quantitative study of the spatial diversity of Moraceae can provide a more accurate basis for species diversity, conservation, use, and restoration of Moraceae in China.

## 2. Materials and methods

#### 2.1. Data collection and processing

In the first step, based on the contents of the Flora Reipublicae Popularis Sinicae [Vol. 23(1)], the species of Moraceae in China and their Latin names were collated and reviewed by plant taxonomy experts to determine a list of Moraceae species in China. Secondly, the database of Moraceae in China was established by compiling all kinds of data sources to record the geographical distribution of Moraceae. The primary data sources for geographical distribution were the fulltext FRPS website<sup>1</sup>, the National Plant Specimen Resource Center<sup>2</sup>, and the National Specimen Information Infrastructure website<sup>3</sup>. Up to the end of 2021, they issued papers and journals on the geographical distribution of Moraceae in China (Wu Zhengyi, 1989; Arimoto et al., 2020), and regional flora was issued by some provinces and regions. Determine the particular distribution of each plant in the county. The data of provinces and regions and the range of distribution that the counties cannot determine will not be adopted. In the final step, we tested all species names for synonyms and merged all synonymized species records.

We also excluded cultivated species and hybrids, and all distribution data were natural species distributions. The attribute data in the database includes genus name, species name, Latin name, province of distribution, county of issuance, and data source, with a total of 28,537 county-level distribution records. The attribute data in the database contained genus name, species name, Latin name, distribution province, distribution county, and data source. It was eventually determined that there were 12 genera and 149 species of Moraceae in China, and a total of 28,537 county-level distribution records were compiled.

## 2.2. Methods

All of the species' spatial data, which includes both graphic and attribute data, was made in ArcGIS 10.2. The graphic data used a map of China with the county as the basic unit (a scale of 1:1,000,000). The attribute data was the distribution of species. Then, the number of genera was obtained, and the spatial diversity map was created. In the spatial distribution of the genus, an isolated county was a patch, and adjacent counties jointly formed a patch. The number of genera in the patch was different, forming different patch types. The spatial diversity and its pattern for the genus could be obtained by calculating the

<sup>1</sup> http://www.iplant.cn/frps2019/

<sup>2</sup> https://www.cvh.ac.cn/index.php

<sup>3</sup> http://www.nsii.org.cn/2017/home.php

index of spatial diversity pattern. The applicable data was obtained from the spatial data and studied according to the spatial diversity pattern indicators (Table 1). Complete all raster data calculations in Fragstats 3.3.

# 3. Results and analysis

## 3.1. Spatial diversity of monotypic genera

Monotypic genera are genera that comprise just one species. Moraceae includes *Maclura*, *Antiaris*, *Malaisia*, and *Cannabis*. Among Moraceae in China, these genera had the smallest patch richness, with a value of 1 (Table 2), indicating merely one patch type. However, the different genera had different numbers of patches. Shannon's diversity index (SHDI), Simpson's diversity index (SIDI), Shannon's evenness index (SHEI), and Simpson's evenness index (SIEI) of monotypic genera were all 0 (Table 2), implying that there was no diversity. The monotypic genus exhibited a high degree of aggregation. Additionally, it had a maximum plaque index but a low total shape index (Table 2).

The total number of all patches (NAP) on *Maclura* was six (Table 2), indicating six patches. Among the 12 genera of Moraceae, the spatial distribution of *Maclura* was the lowest, predominantly distributed in Medog County in Tibet and the eastern, central,

	Index	Formula	Meaning	Unit, Range
1	Patch richness	PR = m	Number of patch types in the genus	$PR \ge 1$
2	Shannon's diversity index	$SIDI = -\sum_{i=1}^{m} (P_i \times \ln P_i)$	Diversity of spatial distribution in the genus	$0 \leq SIDI < 1$
3	Simpson's diversity index	$SHDI = 1 - \sum_{i=1}^{m} P_i^2$	Diversity of spatial distribution in the genus	$SHDI \ge 0$
4	Shannon's evenness index	$SHEI = \frac{-\sum_{i=1}^{m} (P_i \times \ln P_i)}{\ln m}$	Evenness of spatial distribution in the genus	0 ≤ <i>SHEI</i> ≤ 1
5	Simpson's evenness index	$SIEI = \frac{1 - \sum_{i=1}^{m} P_i^2}{1 - \left(\frac{1}{m}\right)}$	Evenness of spatial distribution in the genus	$0 \le SIEI \le 1$
6	Area of patch type	$APT = \sum_{j=1}^{m} a_{ij} \times \frac{1}{10000} \times \frac{1}{10000000}$	Area of patch type <i>i</i>	10 <sup>7</sup> hm <sup>2</sup> APT>0
7	Percentage of the genus	$PG = \frac{\sum_{j=1}^{n} a_{ij}}{A} \times 100$	Proportional abundance of patch type <i>i</i> in the genus	%, 0< <i>PG</i> ≤100
8	Number of patches	$NP = n_i$	Number of patches in patch type <i>i</i>	$NP \ge 1$
9	Largest patch index	$LPI = \frac{\prod_{j=1}^{n} (a_{ij})}{A} \times 100$	Percentage of the genus area comprwased by the largest patch in patch type <i>i</i>	%, 0< <i>LPI</i> ≤100
10	Shape index	$SI = \frac{e_i}{\min e_i}$	Measure of aggregation in patch type <i>i</i>	$SI \ge 1$
11	Total area	$TAP = \sum APT$	Area of genus	$10^7 hm^2$
12	Number of all patches	$NAP = \sum NP$	Number of all patches in genus	$NAP \ge 1$
13	Total largest patch index	$TLPI = \sum LPI$	Sum of largest patch index of every patch type in genus	%, 0< <i>TLPI</i> ≤100
14	Total shape index	$TSI = \sum SI$	Sum of shape index of every patch type in genus	$TSI \ge 1$

m = number of patch types in the genus;  $P_i =$  proportion of the genus occupied by patch type i;  $a_{ij} =$  area of patch j in patch type i; A = area of genus;  $n_i =$  number of patches in patch type i;  $e_i =$  total length of patch type i.

TABLE 1 Indices of spatial diversity patterns.

Genera	Index										
	PR	SHDI	SIDI	SHEI	SIEI	TA	NAP	TLPI	TSI		
Maclura	1	0	0	0	0	0.534	6	59.081	3.492		
Antiaris	1	0	0	0	0	0.934	14	27.985	6.702		
Malaisia	1	0	0	0	0	1.338	20	37.607	7.605		
Cannabis	1	0	0	0	0	17.973	139	26.131	16.262		
Fatoua	2	0.411	0.246	0.594	0.492	4.513	114	13.340	22.312		
Humulus	3	0.034	0.01	0.031	0.015	11.661	158	12.488	20.836		
Broussonetia	3	1.033	0.624	0.941	0.937	17.917	295	11.717	44.713		
Cudrania	4	0.864	0.499	0.623	0.665	15.982	227	16.056	41.419		
Streblus	5	1.302	0.683	0.809	0.854	1.964	42	32.517	23.884		
Morus	7	1.184	0.628	0.609	0.733	31.357	428	18.856	61.714		
Artocarpus	8	1.294	0.654	0.622	0.747	6.675	104	23.872	41.345		
Ficus	41	3.017	0.931	0.813	0.954	23.664	742	15.975	200.219		

#### TABLE 2 Spatial diversity of genera in the Moraceae in China.



northern, and southern parts of Yunnan. *Maclura* in the five counties had a significantly lower distribution range than the other genera. Consistent with the data shown in Figure 1A, the number of all patches (NAP) and total area (TA) of *Maclura* were the smallest (Table 2). Among monotypic genera, the *Maclura* spatial distribution was the most concentrated (Figure 1A), and the most extensive patch index (TLPI) was the highest among the 12 genera of Moraceae (Table 2).

The NAP value of *Antiaris* was 14 (Table 2), indicating that the number of patches was 14. The number of all patches (NAP) of Antiaris was slightly higher than that of Maclura, but Antiaris' distribution range was significantly larger. The numbers of all patches (NAP), total area (TA), and full shape index (TSI) of *Antiaris* in Table 2 were consistent with those of *Maclura*.

The number of all patches (NAP) of *Malaisia* was 20 (Table 2), which indicates that the number of patches in this genus was 20, and the number of patches was slightly larger than that of *Antiaris*. The distribution range was somewhat more extensive than that of *Antiaris*. The number of all patches (NAP) and total area (TA) of the three genera in Table 2 were the smallest among the Moraceae in China, and the total area (TA) of the three genera was comparable.

Among the monotypic genera, the number of all patches (NAP), total area (TA), and complete shape index (TSI) of *Cannabis* were extensive (Table 2). *Cannabis* was distributed in other parts of the county, though not in regions including Hainan, Taiwan, Hong Kong, and Macau. *Cannabis* had the most comprehensive distribution, and the sum of the most extensive patch index of every patch type in the genus (TLPI) was the lowest among the monotypic genera (Table 2).

#### 3.2. Spatial diversity of lesser genera

The lesser genera (2–7 species) included *Fatoua, Humulus, Cudrania, Broussonetia*, and *Streblus*. Patch richness (PR) values of the five species were more significant than 1 (Table 2), and the patch richness increased as patch type gradually increased from 2 to 5. The species in these genera were diverse, and their SHDI, SIDI, SHEI, and SIEI index values were all greater than 0 (Table 2) and slightly higher than those of monotypic genera. Table 1 shows that the correlation index of the lesser genus species (except for *Cannabis*) was higher than monotypic genera for patch richness (PR), total area (TA), and the number of all patches (NAP). The species were more widely distributed and more dispersed than monotypic genera. The total shape index (TSI) was significantly higher than that of monotypic genera (Table 2).

The total area (TA) of *Fatoua* was close to that of *Artocarpus* (Table 2). The difference can be easily observed by comparing the plots of these two genera. The plaque richness (PR) value of *Fatoua* is 2 (Table 2). There are two types of patches with low patch richness. One plaque type had one species, and the other had two species, corresponding to 1 and 2 in the legend of Figure 1A, higher than *Fatoua*. In Figure 1G, the patch types are classified into five classes. *Fatoua* is more widely distributed, and it was concentrated in tropical and southern subtropical regions (Figure 1A). *Artocarpus* is concentrated in tropical and a slightly larger number of patches, *Artocarpus* had more patch types and a slightly higher total area (TA).

The spatial distribution of Humulus (Figure 1B) was analogous to that of monotypic Cannabis. The PR of Humulus was 3 (Table 2), indicating that its patch richness was higher than that of Cannabis. Humulus has three patches: patches with species, patches with two species, and patches with three species, which correspond to 1, 2, and 3 in the legend of Figure 1B, respectively. According to the total shape index (TSI), Humulus species had greater spatial dispersion and uniform spatial distribution than Cannabis (Table 2). However, the total most extensive patch index (TLPI) was less than that of Cannabis. Compared with Cannabis, Humulus had a slightly smaller number of patches, but the total area was slightly larger (Table 2). The SHDI, SIDI, SHEI, and SIEI index values of Humulus were more significant than 0, but the value was lower because the first patch type was more evenly distributed. In contrast, the second and third patches were distributed in the southwest in small quantities (Figure 1B).

Compared with the lesser genera in the previous section, the total area of *Broussonetia* was the largest, as was the number of patches. The PR of *Broussonetia* was 3 (Table 2), and patch richness was low. The spatial distribution of *Broussonetia* (Figure 1C) was similar to that of *Cudrania* (Figure 1D), and its total area (TA) and several patches (NP) were slightly higher than those of *Cudrania* (Table 2), with distribution in the south and north. The SHDI and SIDI diversity index values and the SHEI and SIEI meanness index values of *Broussonetia* (Table 2), so we determined that the spatial distribution of *Broussonetia* was diverse.

The total area of distribution of *Streblus* was similar to *Malaisia* (Table 2). The number of patches (NP) of *Streblus* was twice that of *Malaisia*, and the total shape index (TSI) was three times that of the latter. The spatial distribution of *Streblus* was western. The distribution was more dispersed (Figure 1E). The PR of *Streblus* was 5 (Table 2),

indicating that its patch richness was the highest among the lesser genera. However, the total area (TA) and number of all patches (NAP) of Streblus were the smallest (Table 2). The smallest spatial distribution range was found (Figure 1E). Its total shape index (TSI) was moderately small and relatively concentrated. The SHDI and SIDI diversity indices of Streblus were the highest among the lesser genera, and the SHEI and SIEI values were higher than average. The spatial distribution was the most diverse among the more inferior generation. Streblus had the most considerable number of patch types, but the number of patches (NP) was small, and the total shape index (TSI) was weak. Because Streblus was predominantly distributed in the south of Yunnan, western Guangxi, the west and central parts of Hainan, and southern Guangdong (Figure 1E), most were in mountainous areas. The temperature decreases with height, providing different and challenging environmental conditions for Streblus, which grows in a limited range of suitable conditions.

## 3.3. Spatial diversity of multiple genera

The multiple genera (more than seven species) include *Morus*, *Artocarpus*, and *Ficus*. Among the 12 genera of Moraceae in China, the genera with more than 10 species had the most outstanding patch richness (PR). The number of patches (NP), total shape index (TSI), SHDI, SIDI, SHEI, and SIEI values of these genera were higher than those of smaller genera, but the most extensive patch index (LPI) values were lower (Table 2).

*Morus* had a PR of 7 (Table 2), with seven types of patches. Patch types 1, 2, 3, 4, 5, 6, and 7 represented patches comprising 1, 2, 3, 4, 5, 6, and 7 species, respectively. In Figure 1F, the patch types were divided into five levels. The number of patches (NP) and total shape index (TSI) values of *Morus* were high (Table 2), which was consistent with the number and distribution of patches in Figure 1F. Among genera with more than 10 species, *Morus* had the largest total area (TA) and was the most evenly distributed across the county. *Morus's* SHDI, SIDI, SHEI, and SIEI values were higher than those of other large genera, and the spatial distribution had higher diversity.

The PR of *Artocarpus* was 8 (Table 2), and patch richness (PR) was slightly higher than *Morus*'s. Patch types 1, 2, 3, and 4 represented patches containing 1, 2, 3, and 4 species, respectively, and patch type 5 represented patches containing 5–8 species. In Figure 1G, the patch types were divided into five levels. Among genera with more than 10 species, the patch richness of *Artocarpus* was relatively high. However, the number of patches (NP), total area (TA), and total shape index (TSI) values were the smallest (Table 2). The distribution range of *Artocarpus* was the smallest, and the spatial distribution was the most concentrated (Figure 1G). In Table 2, the SHDI, SIDI, SHEI, and SIEI values of *Artocarpus* were shown to be between the corresponding indicators for *Morus* and *Ficus*, with high spatial diversity.

The PR of *Ficus* was 41 (Table 2), the highest among the 12 genera of Moraceae in China. The patch types were species comprised of 1 to 37, 38, 40, 43, 49, and 50 (Figure 1H). The spatial distribution of *Ficus* was close to that of *Broussonetia*, though farther to the south (Figure 1H), and the spatial distribution was centered. However, the number of *Ficus* patches was 2.5 times that of *Broussonetia*, and the total shape index (TSI) was 4.5 times that

of *Broussonetia*. The most extensive patch index (LPI), SHDI, and SIDI values of *Ficus* were higher than those of *Broussonetia* (Table 2). Among the Moraceae in China, the spatial distribution of *Ficus* was the most diverse and widely distributed, with the highest number of all patches (NAP), number of patch types, total shape index (TSI), SHDI, and SIDI values (Table 2). Therefore, areas with a substantial number of *Ficus* could be used as protected areas to protect the Moraceae resources in China.

## 4. Discussion

Our research applied spatial variety pattern indicators using spatial calculations based on the number of patches, genus and species types, and distribution areas. Spatial diversity patterns were calculated using spatial arrows based on the number of patches, genus or species types, and their distribution areas to create Moraceae's genus and species diversity in each county. Based on the patches, the analysis of the kinds of genera or species and their distribution areas in the study enables the species of mulberry genera and species in each county to be derived, and the spatial variety index can be computed. The computation of the spatial diversity index can quantify spatial diversity, which is unique to this research.

One of the most important features of species spatial distribution was the large-scale pattern of species diversity (Tittensor et al., 2010). The large-scale pattern of species diversity and its formation mechanism is one of the core issues in ecology and the basis of biodiversity conservation planning (Stuart-Smith et al., 2013). The most significant feature of this study was that the distribution was precise to the county level, quantitatively studying the spatial diversity of the Moraceae in China, and the diversity types were divided based on the number of species. Using GIS to draw distribution maps enabled us to carry out a spatially simple and intuitive observation of the species diversity of Moraceae in China and obtain spatial diversity computation indicators from patch maps. We obtained data on the spatial diversity, evenness index, and several patches using the spatial diversity model indicators. And then, we can quantitatively analyze the spatial diversity of Moraceae in China. Carrying out a study at this large scale not only enabled us to discover the spatial diversity center of Moraceae in China more accurately but could also aid in protecting and utilizing it.

The diversity analysis of genera can provide evidence and indications for investigating the evolution of flora (Yue, 2001). Among the Moraceae in China, the patch indicators of monotypic genera were consistent with the species, and spatial diversity did not differ between the species and genus. The number of all patches (NAP), total area (TA), and spatial distribution range of *Maclura* were the lowest among all 12 genera. *Maclura*'s most extensive patch index (LPI) was the highest among the 12 genera, and the spatial distribution was the most concentrated. Among monotypic genera, *Cannabis* had a moderately substantial total shape index (TSI), the most substantial number of patches (NP) and entire area (TA), and the most comprehensive distribution range. Among monotypic genera, the patch richness of *Streblus* was the highest, the total number of patches and total distribution area was the smallest, and the spatial distribution range was the smallest. The total shape index of *Streblus* was moderately small, and the distribution was relatively concentrated. We found that Streblus was mainly distributed in the south of Yunnan, the western part of Guangxi, the central and western parts of Hainan, and the southern part of Guangdong. This was because these areas are mostly mountainous, and the temperature decreases with elevation, providing different environmental conditions for Streblus, which can tolerate a narrow range of conditions.

The SHDI and SIDI diversity indices of *Streblus* were the highest, and the spatial distribution was the most diverse among the genera with few species. *Streblus* had more patch types, but the number of patches was small, and the total shape index (TSI) was low. *Streblus* was a non-monophyletic group, far from other genera. We found that *Streblus* was mainly distributed in the south of Yunnan, the western part of Guangxi, the central and western parts of Hainan, and the southern part of Guangdong. Because these areas were primarily mountainous, the temperature decreased with elevation, providing different environmental conditions for *Streblus*, which can tolerate a narrow range of conditions.

Among the genera with lesser species, the number of all patches (NAP), total area (TA), total shape index (TSI), SIDI, and SHEI values of *Broussonetia* were the highest. *Broussonetia* was widely distributed, and its distribution was the most diverse. The ecological adaptation of *Broussonetia papyrifera* was strong, and this species was widely distributed in China. The reason why is that the *B. papyrifera* population has high genetic diversity due to the fact that the tree was a cross-pollinated plant and geographical isolation due to features such as rivers, mountains, roads, and canals that block the flow of genes between *Broussonetia papyrifera* populations.

Among the Moraceae in China, the spatial distribution of *Ficus* was the most diverse, with the most significant number of all patches (NAP), number of patch types, total shape index (TSI), SHDI, and SIDI values. The diversified centers of *Ficus* were distributed in southeastern Gansu, northeastern Guizhou, southern Yunnan, southwestern Guangxi, southern Taiwan, and western Hainan. The spatial diversity of *Ficus* could be used when designating protected areas for the Moraceae in China. The spatial diversity center of the Moraceae in China was mainly distributed in tropical and subtropical regions. This conclusion was consistent with the view put forth by the Flora Reipublicae Popularis Sinicae that the Moraceae in China were prolific in tropical and subtropical areas, with a few distributed in the temperate zone.

# Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository and accession number(s) can be found in the article.

# Author contributions

DL: conceptualization, validation, formal analysis, data curation, and writing - original draft. LQ: conceptualization, resources, and acquisition of the financial support for the project leading to this publication. MJ: software, validation, formal analysis,

data curation, and acquisition of the financial support for the project leading to this publication. ZF: conceptualization and resources. ZW: provided guidance on article format and acquisition of the financial support for the project leading to this publication. All authors contributed to manuscript revision, read and approved the submitted version.

## Funding

This work was supported by grants from the National Natural Science Foundation of China (41071335, 41171423).

## References

Arimoto, K., MacGowan, I., and Su, Z.-H. (2020). New data on lance flies (Diptera, Lonchaeidae) associated with figs (Moraceae, *Ficus* spp.) in Japan and Taiwan, with descriptions of two new species of the genus Silba Macquart. *J. Asia Pac. Entomol.* 23, 364–370. doi: 10.1016/j.aspen.2019.11.007

Berg, C. (1989). Classification and distribution of *Ficus*. *Experientia* 45, 605–611. doi: 10.1007/BF01975677

Chakraborty, A., Mahajan, S., Bisht, M. S., and Sharma, V. K. (2022). Genome sequencing and comparative analysis of *Ficus bengalensis* and *Ficus religiosa* species reveal evolutionary mechanisms of longevity. *iScience* 25:105100. doi: 10.1016/j. isci.2022.105100

Chen, H., Chen, J., Yang, H., Chen, W., Gao, H., and Lu, W. (2016). Variation in total anthocyanin, phenolic contents, antioxidant enzyme and antioxidant capacity among different mulberry (*Morus* sp.) cultivars in China. *Sci. Hortic.* 213, 186–192. doi: 10.1016/j.scienta.2016.10.036

Chen, R.-F., Xu, L., Yu, M. D., Liu, X.-Q., and Chen, L.-Q. (2010). Determination of the Origin and Evolution of Morus (Moraceae) by Analyzing the Internal Transcribed Spacer (ITS) Sequences. In 2010 4th International Conference on Bioinformatics and Biomedical Engineering. IEEE.

Colwell, R. K., and Lees, D. C. (2000). The mid-domain effect: geometric constraints on the geography of species richness. *Trends Ecol. Evol.* 15, 70–76. doi: 10.1016/S0169-5347(99)01767-X

Dong, J.-L., Li, Z., Gao, J. X., Sun, B. N., and He, Y. L. (2022). Ficus leaves within the *Ficus* subgenus *Urostigma* (Moraceae) from the middle Miocene in South China and their biogeography implications. *Rev. Palaeobot. Palynol.* 302:104671. doi: 10.1016/j. revpalbo.2022.104671

Elhawary, S. S., Younis, I. Y., el Bishbishy, M. H., and Khattab, A. R. (2018). LC– MS/MS-based chemometric analysis of phytochemical diversity in 13 *Ficus* spp. (Moraceae): correlation to their *in vitro* antimicrobial and *in silico* quorum sensing inhibitory activities. *Ind. Crop. Prod.* 126, 261–271. doi: 10.1016/j.indcrop. 2018.10.017

Ganivet, E., Unggang, J., Bodos, V., Demies, M., Ling, C. Y., Sang, J., et al. (2020). Assessing tree species diversity and structure of mixed dipterocarp forest remnants in a fragmented landscape of North-Western Borneo, Sarawak, Malaysia. *Ecol. Indic.* 112:106117. doi: 10.1016/j.ecolind.2020.106117

Gardner, E. M., Sarraf, P., Williams, E. W., and Zerega, N. J. C. (2017). Phylogeny and biogeography of *Maclura* (Moraceae) and the origin of an anachronisticfruit. *Mol. Phylogenet. Evol.* 117, 49–59. doi: 10.1016/j.ympev.2017.06.021

Hashemi, S., and Khadivi, A. (2020). Morphological and pomological characteristics of white mulberry (*Morus alba* L.) accessions. *Sci. Hortic.* 259:108827. doi: 10.1016/j. scienta.2019.108827

He, Y.-H., Gao, P.-L., and Qiang, S. (2019). An investigation of weed seed banks reveals similar potential weed community diversity among three different farmland types in Anhui Province, China. *J. Integr. Agric.* 18, 927–937. doi: 10.1016/S2095-3119(18)62073-8

Islam, S. M. S., and Rahman, M. S. (2019). Genetic diversity analysis based on morphological characters in mulberry (*Morus* spp.). *J. Biosci.* 28, 111–119. doi: 10.3329/jbs.v28i0.44717

Kong, W., Wu, S. H., Wu, X. Q., Zheng, X. R., Sun, X. R., Ye, J. N., et al. (2020). First report of leaf spot disease caused by Colletotrichum tropical on *Ficus binnendijkii* var. *variegata* in China. *Plant Dis.* 104:585. doi: 10.1094/PDIS-04-19-0834-PDN

Kumar, A., Bajpai, O., Mishra, A. K., Sahu, N., Behera, S. K., and Chaudhary, L. B. (2011). Assessment of diversity in the genus *Ficus* L. (Moraceae) of Katerniaghat wildlife sanctuary, Uttar Pradesh, India. American. *J. Plant Sci.* 2, 78–92. doi: 10.4236/ajps.2011.21011

Machado, A. F. P., Rønsted, N., Bruun-Lund, S., Pereira, R. A. S., and Paganucci de Queiroz, L. (2018). Atlantic forests to the all Americas: biogeographical history and

## **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

#### Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

divergence times of neotropical *Ficus* (Moraceae). *Mol. Phylogenet. Evol.* 122, 46–58. doi: 10.1016/j.ympev.2018.01.015

Marcotuli, I., Mazzeo, A., Nigro, D., Giove, S. L., Giancaspro, A., Colasuonno, P., et al. (2019). Analysis of genetic diversity of *Ficus carica* L. (Moraceae) collection using simple sequence repeat (SSR) markers. *Acta Sci. Polon. Hortorum Cultus* 18, 93–109. doi: 10.24326/asphc.2019.4.9

Martins, L. A. R., Lorenzoni, R. M., Pereira, R. M., de Miranda, F. D., Fontes, M. M., Carrijo, T. T., et al. (2021). Genetic diversity and structure of *Dorstenia elata* (Moraceae) in an Atlantic Forest remnant. *Rodriguésia*:72.

Nagendra, H. (2002). Opposite trends in response for the Shannon and Simpson indices of landscape diversity. Appl. Geogr. 22, 175–186. doi: 10.1016/S0143-6228(02)00002-4

Oluyinka Christopher, A. (2020). Comparative analyses of diversity and similarity indices of West Bank Forest and block a Forest of the International Institute of Tropical Agriculture (IITA) Ibadan, Oyo state, Nigeria. *Int. J. Forest. Res.* 2020, 1–8. doi: 10.1155/2020/4865845

Pederneiras, L. C., da Costa, A. F., Medeiros, H., Rivera, N. M., Forzza, R. C., Romaniuc-Neto, S., et al. (2020). Species diversity of *Ficus* L. sect. Americanae (Moraceae) in acre, Brazil. *Brittonia* 72, 215–231. doi: 10.1007/s12228-020-09620-1

Pielou, E. C. (1966). The measurement of diversity in different types of biological collections. *J. Theor. Biol.* 13, 131–144. doi: 10.1016/0022-5193(66)90013-0

Ricotta, C., and Avena, G. (2003). On the relationship between Pielou's evenness and landscape dominance within the context of Hill's diversity profiles. *Ecol. Indic.* 2, 361–365. doi: 10.1016/S1470-160X(03)00005-0

Shi, Y., Mon, A. M., Fu, Y., Zhang, Y., Wang, C., Yang, X., et al. (2018). The genus *Ficus* (Moraceae) used in diet: its plant diversity, distribution, traditional uses and ethnopharmacological importance. *J. Ethnopharmacol.* 226, 185–196. doi: 10.1016/j. jep.2018.07.027

Shukla, G., Rai, P., Abha Manohar, K., and Chakravarty, S. (2020). Quantification of diversity, biomass and carbon storage of climber and liana community in a foothill forest of Indian Eastern Himalayas. *Acta Ecol. Sin.* 40, 478–482. doi: 10.1016/j. chnaes.2020.09.009

Stuart-Smith, R. D., Bates, A. E., Lefcheck, J. S., Duffy, J. E., Baker, S. C., Thomson, R. J., et al. (2013). Integrating abundance and functional traits reveals new global hotspots offish diversity. *Nature* 501, 539–542. doi: 10.1038/nature12529

Teixeira, S. P., Costa, M. F. B., Basso-Alves, J. P., Kjellberg, F., and Pereira, R. A. S. (2018). Morphological diversity and function of the stigma in *Ficus* species (Moraceae). *Acta Oecol.* 90, 117–131. doi: 10.1016/j.actao.2018.02.008

Tittensor, D. P., Mora, C., Jetz, W., Lotze, H. K., Ricard, D., Berghe, E. V., et al. (2010). Global patterns and predictors of marine biodiversity across taxa. *Nature* 466, 1098–1101. doi: 10.1038/nature09329

Wang, H.-F., Xu, X., Cheng, X. L., Liu, Y., Luo, A., Lyu, T., et al. (2022). Spatial patterns and determinants of Moraceae richness in China. *J. Plant Ecol.* 15, 1142–1153. doi: 10.1093/jpe/rtac025

Wu Zhengyi, Z. X. (1989). Taxa nova nonnulla moracearum sinensium. *Plant Divers*. 11, 1–3.

Yue, T. (2001). Studies and questions of biological diversity. Acta Ecol. Sin. 21, 462-467.

Zerega, N. J., Clement, W. L., Datwyler, S. L., and Weiblen, G. D. (2005). Biogeography and divergence times in the mulberry family (Moraceae). *Mol. Phylogenet. Evol.* 37, 402–416. doi: 10.1016/j.ympev.2005.07.004

Zhang, J., Zhu, W. F., Xu, J., Kitdamrongtham, W., Manosroi, A., Manosroi, J., et al. (2018). Potential cancer chemo preventive and anticancer constituents from the fruits of *Ficus hispida* L.f. (Moraceae). *J. Ethnopharmacol.* 214, 37–46. doi: 10.1016/j.jep.2017.11.016