

1 Pollinator selection against toxic nectar as a key facilitator of a plant invasion

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13

14 Abstract

15 Plant compounds associated with herbivore defence occur widely in floral nectar and can impact
16 pollinator health. We showed previously that *Rhododendron ponticum* nectar contains grayanotoxin
17 I (GTX I) at concentrations that are lethal or sublethal to honeybees and a solitary bee in the plant's
18 non-native range in Ireland. Here we further examined this conflict and tested the hypotheses that
19 nectar GTX I is subject to negative pollinator-mediated selection in the non-native range– but that
20 phenotypic linkage between GTX I levels in nectar and leaves acts as a constraint on independent
21 evolution. We found that nectar GTX I experienced negative directional selection in the non-native
22 range, in contrast to the native Iberian range, and that the magnitude and frequency of pollinator
23 limitation indicated that selection was pollinator mediated. Surprisingly, nectar GTX I levels were
24 decoupled from those of leaves in the non-native range, which may have assisted post-invasion
25 evolution of nectar without compromising the anti-herbivore function of GTX I (here demonstrated
26 in bioassays with an ecologically relevant herbivore). Our study emphasizes the centrality of
27 pollinator health as a concept linked to the invasion process, and how post-invasion evolution can be
28 targeted towards minimising lethal or sub-lethal effects on pollinators.

29

30 Introduction

31 The occurrence of toxins in nectar may appear paradoxical since this is the primary reward for
32 pollinators [1, 2], but it is none-the-less widespread across many plant families [3-5]. While the
33 occurrence of toxic or deterrent phytochemicals in nectar could be maladaptive for plant fitness if
34 they impact pollinator health, they may also increase exclusivity or fidelity of pollinator visitation and
35 thereby the efficiency of pollen transfer [3, 6]. However, whether nectar phytochemicals are beneficial
36 or detrimental to plant fitness can also be context dependent [7]. Yet despite both these potential
37 beneficial and detrimental effects, pollinator-mediated selection either for or against so-called 'toxic
38 nectar' has still yet to be empirically demonstrated in natural plant populations.

39 As a model system, we examined natural selection on toxic nectar in native and non-native
40 populations of *Rhododendron ponticum* L. (Ericaceae). Species in this genus constitutively express
41 diterpene grayanotoxins (GTXs) throughout most plant parts that are toxic to a wide range of insect
42 and other animal species [8-11]. We previously reported that GTXs occur in *R. ponticum* nectar [12],
43 and that while some pollinators such the buff-tailed bumblebee (*Bombus terrestris* L.) can consume
44 GTX at naturally occurring nectar concentrations without adverse effects, other bee species, including
45 honeybees and the solitary mining bee (*Andrena scotica*, Pérez), cannot [13]. Observed lethal and sub-
46 lethal effects resulted from exposure to grayanotoxin I (GTX I) at ecologically relevant nectar
47 concentrations, whereas the deacetyl derivative, grayanotoxin III (GTX III), was non-toxic. Our past
48 work in this system also showed that GTX I, but not GTX III, was notably absent or significantly reduced
49 in non-native populations in Ireland [12]. Given that pollen limitation may be common in non-native
50 *R. ponticum* populations [14], this suggests that pollinator-mediated selection and loss of nectar toxins
51 could have played a central role in facilitating invasion. Further analysis of this model system thus has
52 the potential to reveal a better understanding of the benefits, trade-offs, and ecological significance
53 of toxic nectar for plants, especially in terms of how plants evolve to optimise interactions with
54 mutualists and antagonists. In this study, we consider the consequences of toxic nectar for pollinator
55 health, and if and how invasive plants can evolve to maximise the services of pollinators while
56 maintaining their defence against herbivores.

57 Beyond their potential influence on pollinator health, GTXs also serve as highly effective herbivore
58 antifeedants in *Rhododendron* species [8-11]. Phenotypic correlation between defence-related
59 compounds in nectar and other plant parts (such as leaves and phloem) appears to be a common
60 phenomenon across plant families [5, 15-17]. Thus, the potential for conflicting pressure on GTXs from
61 pollinators and herbivores exists across all plant parts. To investigate this possibility, we employed a
62 path analysis framework to assess the direction and magnitude of phenotypic selection on leaf, flower,

63 and nectar GTXs in native and non-native *R. ponticum* populations. This approach allowed us to devise
64 a realistic path model – reflecting a foliar biogenesis of grayanotoxins [18, 19] leading to linked
65 expression in flowers and nectar – to quantify the extent to which phenotypic selection on a certain
66 plant part was imposed directly, and indirectly (i.e., arising from phenotypic linkage with other plant
67 parts). Complementary to phenotypic selection analysis, we also undertook manipulative experiments
68 with pollinators and herbivores to examine their roles as potential selective agents.

69 The main objective of this study therefore was to examine if pollinators potentially drive post-invasion
70 evolution of nectar and an important plant defence trait, and thereby act as key facilitators of invasion
71 by the entomophilous species *Rhododendron ponticum*. In particular, we tested the hypotheses that: 1)
72 the direction and magnitude of natural selection on nectar GTX I varies across the native and non-
73 native range of *R. ponticum*, consistent with the reduced levels observed in non-native populations;
74 2) pollinators are important drivers of this selection in the non-native range (i.e., pollinator limitation
75 of plant fitness is correlated with nectar GTX I levels); 3) GTX I levels in nectar are phenotypically
76 correlated with those in leaves and flowers, which should therefore also show reduced levels in the
77 non-native range; and 4) that any reduction in leaf GTX I represents a trade-off owing to its adaptive
78 value against herbivory (determined in a feeding bioassay with an ecologically important insect
79 herbivore).

80 **Methods**

81 **Location, traits and abiotic variables measured**

82 The study was conducted in nine native populations of *R. ponticum* subsp. *baeticum* in southern Spain
83 and northern and southern Portugal, and four populations in the species' non-native range in Ireland
84 (Table S1). Between six and ten plants were sampled per population, which typically numbered about
85 20-30 flowering individuals. A minimum distance of 20 m was kept between individuals so as to reduce
86 the chance of sampling ramets. For each plant individual, nectar, leaf and corolla material was
87 collected, and floral morphological and abiotic variables were quantified. Nectar was collected from
88 between 8-15 unbagged flowers using microcapillary tubes, and was pooled until ca. 1 μ l was obtained
89 per plant. This volume was more than sufficient to obtain large quantifiable peaks for GTX I in LC-MS
90 analysis (see below). So as to standardise the time point of collection across individuals, nectar was
91 sampled from flowers in their beta-phase of phenology around the time of stigma receptivity [20].
92 From each flower that nectar was sampled, the corolla and nearest sub-tending leaf were also
93 removed and immersed in silica gel in snap-seal bags in a composite sample for each plant. The
94 appropriateness of this sampling technique was supported by the fact that grayanotoxins are known
95 to function as constitutive defences in plants (as opposed to being specifically induced by damage) [9]

96 and are comparatively stable in dried tissues and in solution [21, 22]. Nonetheless, care was taken not
97 to damage any plant tissue until after nectar samples were collected. In the lab, water was removed
98 by freeze drying nectar and oven drying leaf and flower samples at <50°C. Dried flower and leaf
99 samples (30 mg) were ground to a homogenous powder and extracted (3 X 20 ml) in MeOH, from
100 which a 200 µl aliquot was transferred to analysis vials. Dried nectar was re-suspended in 200 µl MeOH
101 for analysis. Quantification of GTX I was carried out by LC-MS analysis as previously reported [12].
102 Final values of GTX I were expressed as a concentration of dry weight of tissue (µg/mg dw). Mean
103 corolla width (measured as the widest horizontal distance between the tips of petal wings) and corolla
104 tube width (measured as the internal diameter of the corolla tube at its base) were recorded with dial
105 callipers from five flowers per plant [23]. We previously reported that several microhabitat factors
106 (canopy cover, aspect, elevation and irradiance) explained a significant amount of variation in nectar
107 toxin levels in *R. ponticum* [12]. Where appropriate, we utilised these variables to control for the
108 confounding effect of environmental heterogeneity in models featuring toxin levels as an explanatory
109 variable.

110 **Relative fitness and pollinator limitation**

111 Maternal fitness was measured as total seed set per plant. Calculation of seed set in tall, profusely
112 flowering shrub or tree species can prove challenging, and hence a sub-sampling approach is often
113 employed [24, 25]. To obtain estimates of total seed set in *R. ponticum* plants, we first calculated mean
114 seed set from 8-40 capsules (depending on flower abundance per plant). Established regression
115 equations from native and non-native populations [20, 26] were used to estimate viable seed number
116 based on mature capsule length. Viable and non-viable seeds are easily discerned in this species due
117 to miniscule size and weight of the latter. To then estimate the total number of flowers per plant, we
118 counted the number of flower trusses (racemes consisting of a pseudo-whorl of usually 9-12 flowers)
119 per individual and multiplied this by the mean flower number (inclusive of those at pre and post-
120 anthesis stage) obtained from 15-20 trusses. Although not all flowers mature into fruiting capsules,
121 these measures are none-the-less highly correlated in *R. ponticum* [20]. Finally, we multiplied total
122 number of flowers per plant by the mean seed set per capsule to afford total seed set. Relative fitness
123 was calculated by dividing individual seed set by the native or non-native range mean.

124 A cohort of five individuals per population was selected for application of a supplementary pollination
125 treatment to measure pollinator limitation. Although *R. ponticum* is self-fertile, optimal seed set
126 occurs under out-crossing [26], and in particular due to intrapopulation cross-pollination [20]. The
127 supplemental treatment thus consisted of application of recently dehisced anthers from neighbouring
128 plants (≥ 35 m distance away) to receptive stigmas of target flowers, ensuring deposition of the long

129 viscin pollen threads. The treatment was implemented at the start of the flowering period (late April
130 in the native range; early June in the non-native range) when the activity of important pollinators (e.g.
131 bumblebees, and solitary bees) was apparent [27]. Both treated flowers, and non-treated control
132 flowers at the same phenological stage, were tagged and collected just preceding capsule dehiscence
133 (mid-October in the native range; late January in the non-native range), with an overall retrieval rate
134 of 88 % (due to wind damage, natural excision etc.). Pollinator limitation was therefore assumed in
135 plants where supplementally treated flowers exhibited significantly higher seed set than open-
136 pollinated control flowers, according to one-tailed Welch's *t*-tests. The resultant *t*-value of this test
137 was taken as a continuous, quantitative measure of the magnitude of pollinator limitation per plant.
138 While the ability to differentially allocate resources to out-crossed flowers has been noted in some
139 species [28, 29], we did not believe this to be a confounding issue in our measure of pollinator
140 limitation given the large gradients and spatially consistent patterns which were subsequently
141 observed.

142 **Field and experimental assessment of resistance to herbivory**

143 All plants from which traits were measured were also surveyed for herbivore damage at the same time
144 as when pollinator treatments were initiated in the native range (see above) and in early to mid-July
145 in the non-native range. These time points hence permitted sufficient current-season herbivore
146 damage to accumulate, in addition to previous years' damage evident on older leaves. Due to the
147 typically large size of shrubs, we assessed herbivore damage in 1 m³ areas at the edge of individuals
148 from ground level upwards. The total number of young and old leaves within this area was counted,
149 and the number of leaves exhibiting herbivore damage were recorded for each age class. If present,
150 the area of damage on leaves was usually consistent (ca. 10-15 % area removed). A generalist species
151 of broad-nosed weevil (Coleoptera: Curculionidae: Entiminae) known to feed on *R. ponticum* in the
152 non-native range [30, 31], the Black vine weevil (*Otiorhynchus sulcatus* Fabricius), was selected for
153 bioassays and reared from larval stage in a glasshouse on strawberry plants. Bioassays with black vine
154 weevils (BVWs) were conducted using late instar adults in pre-oviposition period; a phase lasting 3-6
155 weeks during which time they consume the most plant foliage. Thirty adults were placed into
156 individual arenas (20 X 10 X 6 cm) and randomly allocated to three treatments: 1) a control artificial
157 diet; 2) an artificial diet with GTX I incorporated at natural leaf concentrations; and 3) an artificial diet
158 in which ten times the natural concentrations of GTX I was incorporated. Artificial diets for BVWs were
159 constructed following established techniques [32, 33], which consisted of cellulose acetate disks (0.45
160 µm pore size) treated with water-dissolved sucrose and β-sitosterol phagostimulants at
161 concentrations known to solicit high feeding rates [34]. Sample sizes (the number of BVWs per
162 treatment) were constrained by the limited quantity of GTX I we were able to isolate from several kg

163 of dried *R. ponticum* flowers, as per methods previously reported [35]. However, since there is
164 typically low between-individual variation in BVWs due to obligate parthenogenesis [36], we
165 considered these sample sizes adequate. Experiments were conducted for a total of 11 days (with a
166 single change of cellulose disks at day 5.5) in conditions maintained at ca. 21 °C and 85 % relative
167 humidity [37, 38]. The cumulative area eaten (mm²) from disks was quantified per weevil from digital
168 scans using ImageJ analysis software (National Institutes of Health, Bethesda, Maryland, USA). For
169 both field and laboratory assessments, results are reported in terms of resistance (i.e. 1 minus %
170 herbivore damage).

171 **Data analysis**

172 **Comparison of GTX I across ranges** – Geographic variation in nectar, leaf, and flower GTX I levels was
173 analysed in separate linear mixed models (LMMs) fit by restricted maximum likelihood estimation
174 using the R package nlme [39]. As three separate LMMs were conducted, we employed Benjamini-
175 Hochberg adjustment of *p*-values to reduce the familywise error rate. Non-native plants are known
176 to have originated from Spanish as opposed to Portuguese populations [40], and for this reason we
177 restricted range comparisons to the former only. Nectar, leaf, and flower GTX I levels were square
178 root transformed (to improve normality) and fit in LMMs as dependant variables against range (native
179 and non-native) as a fixed effect and population as a nested random effect – with microhabitat
180 variables included as covariates. For model validation, standardised residuals were examined for
181 normality, homogeneity and independence, including spatial autocorrelation [41]. Non-equal variance
182 of residuals between populations was accounted for in the leaf GTX I model by incorporation of a
183 variance correlation structure (based on population identity), which significantly improved model AIC
184 (Likelihood-ratio test; $L = 20.3$, $p=0.016$).

185 **Natural selection on plant toxin levels** – Before implementing selection analyses, we first: A.)
186 controlled traits for potential confounding effects of environmental heterogeneity, as strong abiotic-
187 mediated covariance between traits and fitness can bias estimates of selection gradients [42, 43]; and
188 B.) affirmed the legitimacy of pooling population data [44] in order to assess selection at the range
189 level. Details of these steps are provided (see Supplementary Methods). Subsequently, estimates of
190 directional selection were obtained for each range through multiple regression of relative fitness on
191 standardized traits [45] within a path analytical framework – following terminology of Scheiner et al.
192 [44]. For path models, a hypothesized causal structure between leaf, nectar and flower GTX I levels
193 and relative fitness was assessed. In addition, we tested for non-linear selection on traits, including
194 quadratic (disruptive/stabilizing) and correlational selection [45]. However, as no significant non-
195 linear selection was detected (data not shown), we focussed on directional selection only. We

196 employed mean-standardization of traits to allow output of mean-standardized selection gradients
197 (β_{μ}) from analyses, as a measure of intensity of selection. These are deemed superior where
198 comparisons of the strength of natural selection are desired, for instance between traits, or across
199 geographic space [46] – with the added advantage of their interpretation as fitness elasticities [47,
200 48]; the resultant change in relative fitness from doubling trait values.

201 Path and mediation analyses were carried out using the R package ‘lavaan’ [49] for structural equation
202 modelling. Data for both ranges were assessed for multivariate normality by Mardia's test in the R
203 package ‘MVN’ [50]. As neither dataset met this requirement, we opted for robust maximum
204 likelihood estimation of path coefficients as a non-parametric alternative. Path model goodness-of-fit
205 is reported as the Satorra-Bentler adjusted Chi-squared (χ^2), which can provide better approximation
206 of p -values under non-normality. Following the estimation of path coefficients, mediation analysis was
207 employed to test the significance of three parameters in path models: 1) direct selection gradients
208 (β_{μ}) (assessed along forward-connected paths from a trait to fitness, inclusive of any mediation
209 through intermediate traits); 2) indirect selection (assessed as paths which lead forward to fitness first
210 through a backwards step); and 3) total selection differentials (denoted s ; the sum of direct and
211 indirect selection) [44, 51]. Selection differentials estimated within a path model are also referred to
212 as the ‘predicted covariance’, as values will usually differ from as typically measured (i.e. through
213 simple trait-fitness correlations) in the absence of causal structure [44, 52]. As fitness measurements
214 were not taken and/or could not be retrieved on all plants, missing values were casewise deleted.
215 Final sample sizes for path and mediation analyses were $n = 68$ (i.e., $n = 38$ for the native range and $n =$
216 30 for the non-native range). These sample sizes are ca. 45-60 % of the median sample size reported
217 for plants in a systematic review on selection [53], and are at least ten times the number of model
218 explanatory variables, as per standard guidelines [54].

219 **Biotic selection pressures on plant toxin levels** – Differences in the frequency and intensity of
220 pollinator limitation between ranges were assessed through Pearson's Chi-square Test for
221 Independence and by t-test, respectively. Following this, multiple regression analyses were conducted
222 for each range, to examine potential biotic and abiotic determinants of pollinator limitation. In
223 addition to nectar toxins, a range of floral morphological (corolla and tube width) and microhabitat
224 variables (canopy cover, aspect, elevation and irradiance) were considered for inclusion in models as
225 potential co-determinants. Multicollinearity was monitored using variance inflation factors (VIFs).
226 Final regression models contained explanatory variables significant after Benjamini-Hochberg
227 adjustment of p -values. A Generalised Linear Model (GLM) with quasi-binomial errors (to account for
228 overdispersed proportional data) was used to determine if there were differences in resistance to

229 BVW among GTX I treatments (control, normal, x10). Post-hoc Tukey pairwise comparisons were used
230 to determine which treatments were significantly different from one another, using the R package
231 multcomp [55]), and corrected for multiple comparisons by Benjamini-Hochberg adjustment. The
232 frequency of herbivore damage on plants in the field was analysed according to the factors of leaf age
233 and range of provenance (i.e. native or non-native) using Pearson's Chi-square Test for Independence.
234 To investigate whether observed levels of plant resistance (i.e. 1 minus % herbivore damage) in the
235 field could be explained by leaf GTX I and other microhabitat variables (as listed above) we fitted GLMs
236 for each range with a quasi-binomial distribution (to account for over-dispersion). The overall
237 significance of GLMs was assessed through comparison with a null model, and McFadden's pseudo- R^2
238 were generated to assess model fit.

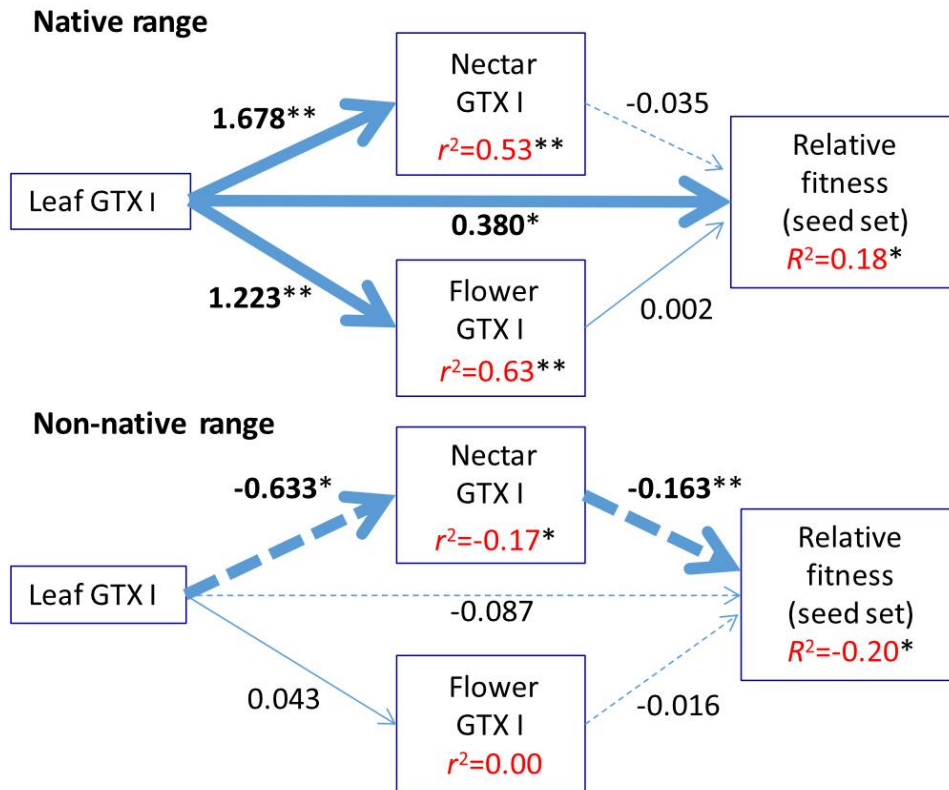
239 **Results**

240 **Natural selection on plant toxin levels**

241 Directional selection on plant toxin levels was apparent in *R. ponticum* (Fig. 1; Table 1), with our *a*
242 *priori* hypothesis of causal linkage between leaf, flower, and nectar GTX I and fitness deemed
243 adequately representative of the observed data in path models for the native ($\chi^2 = 0.86$, $df = 1$, $p =$
244 0.347) and non-native range ($\chi^2 = 2.21$, $df = 1$, $p = 0.137$). However, the intensity and direction of
245 phenotypic selection on traits was not consistent among regions; with strong positive total selection
246 on leaf, flower, and nectar GTX I observed for plants in the native range, in contrast to significant
247 negative total selection on nectar GTX I in the non-native range (Table 1). This discrepancy is indicative
248 of divergent selection acting on nectar toxin levels, and is consistent with the pattern of phenotypic
249 differentiation in nectar toxin levels found between ranges (Fig. 2). In contrast, leaf and flower GTX I
250 which were selectively neutral in the non-native range did not differ in their phenotypic expression
251 between ranges (Fig. 2). Linkage in toxin levels across leaves and nectar, and leaves and flowers, also
252 appeared altered between ranges (Fig. 1), in which a breakdown in phenotypic correlation was
253 indicated in the non-native range.

254 Decomposition of total selection on traits into direct and indirect components revealed that total
255 selection on nectar and flower toxin levels in the native range is the result of large indirect selection
256 acting through leaves (Table 1). While in the non-native range, only direct selection on nectar toxin
257 levels was observed. Within ranges, no instances of conflicting selection on traits were observed
258 (Table 1).

259



260

261 **Figure 1.** Solved path diagrams for directional selection on traits in the native (top) and non-native (bottom)
 262 range of *Rhododendron ponticum*. Mean-standardized path coefficients are presented, with dashed lines
 263 representing negative coefficients, and arrow width indicative of the strength of effect (bold values sig. at:
 264 * $P \leq 0.05$; ** $P \leq 0.001$). Direct selection is assessed along forward-connected paths to fitness, inclusive of
 265 any mediation through intermediate variables, and indirect selection as paths which lead forward to fitness
 266 first through a backwards step. The confounding influence of abiotic environment on traits was controlled
 267 for. Path analyses and multiple regressions are based on $N=38$ and $N=30$ for the native and invasive range
 268 respectively; and single correlations between tissues on $N=53$ and $N=30$.

269

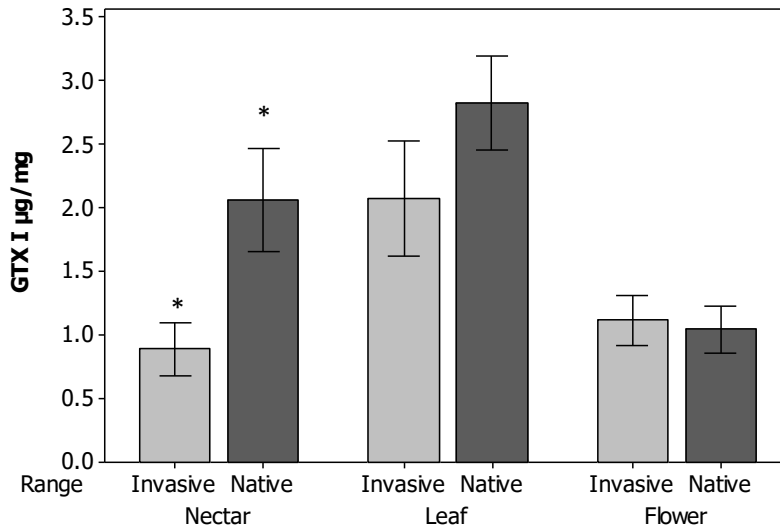
270 **Table 1.** Mediation analysis of total selection on toxin levels in the native and invasive range, partitioned
 271 into direct and indirect components. Total selection (s) on a trait is the sum of all direct selection gradients
 272 (β) and indirect selection. Units are mean-standardized selection coefficients (\pm robust SE).
 273

| Range/Trait | Direct selection β (\pm SE) | Indirect selection (\pm SE) | Total selection [†] s (\pm SE) |
|-----------------|---|-----------------------------------|---|
| Native | | | |
| Leaf | 0.324* (± 0.103) | n/a [^] | 0.324* (± 0.103) |
| Flower | 0.002 (± 0.090) | 0.393* (± 0.178) | 0.396* (± 0.140) |
| Nectar | -0.035 (± 0.059) | 0.642 (± 0.339) | 0.607* (± 0.299) |
| Invasive | | | |
| Leaf | 0.015 (± 0.079) | n/a [^] | 0.015 (± 0.079) |
| Flower | -0.016 (± 0.113) | 0.001 (± 0.004) | -0.015 (± 0.113) |
| Nectar | -0.163** (± 0.040) | 0.055 (± 0.052) | -0.107* (± 0.049) |

274 [†] Also referred to as 'predicted covariance' within context of a path model (see methods)

275 [^] n/a due to the implied causal structure of path models

276 Bold values sig. at: * $P \leq 0.05$; ** $P \leq 0.001$



277
 278 **Figure 2.** Mean toxin levels (GTX I µg/mg ± 95% CI) per dried sample type in the native and non-native
 279 range of *Rhododendron ponticum*. For nectar, leaf, and flowers, linear mixed models were fitted with
 280 'range' as a fixed effect and 'population' as a nested random effect, and were controlled for abiotic
 281 environment. After adjustment for multiple comparisons, significant differences were detected between
 282 ranges for nectar ($t = 3.82$, $N_{[pops]} = 13$, $N_{[plants]} = 87$, $p = 0.008$), but not for leaves ($t = 1.81$, $N_{[pops]} = 10$, $N_{[plants]} =$
 283 66 , $p = 0.162$) or flowers ($t = -0.07$, $N_{[pops]} = 10$, $N_{[plants]} = 66$, $p = 0.949$).

284

285 Pollinators as drivers of selection on plant toxin levels

286 Plants in the native and non-native range differed significantly in the frequency of pollinator limitation
 287 experienced ($\chi^2 = 17.6$, $df = 1$, $p = \leq 0.001$); with seed set in 76 % of plants in the non-native range found
 288 to be pollen limited, compared to only 15 % of plants in the native range. There was also a difference
 289 in the intensity of pollinator limitation in plants between the native (mean = 0.95 ± 0.34) and non-native
 290 range (mean = 4.24 ± 0.57) (t-test: $t = 5.22$, $df = 44$, $p = \leq 0.001$). Subsequently, we examined a range of
 291 biotic and abiotic factors to determine potential causes of pollination limitation in each range. The
 292 same pattern was observed within both ranges, in that plants which were more highly pollen limited
 293 possessed higher levels of nectar toxins and also wider flower corollas (Table 3). However, the strength
 294 of association between nectar GTX I and pollen limitation was more than three times greater in the
 295 non-native than the native range.

296

297 **Table 3.** Multiple regression analysis of determinants of pollination limitation. In addition to nectar toxins,
 298 a range of floral morphological and microhabitat variables (as listed in Methods) were considered for
 299 inclusion in models. Adjusted p -values are reported.
 300

| Range | Coefficient (± SE) | t -value | p -value |
|----------------------|--------------------|------------|------------|
| Native [†] | | | |
| Nectar GTX I (µg/mg) | 0.58 (±0.23) | 2.52 | 0.020 |

| | | | |
|--|---------------------|------|-------|
| Flower corolla width (mm) | 0.22 (± 0.07) | 3.03 | 0.012 |
| Non-native* | | | |
| Nectar GTX I ($\mu\text{g}/\text{mg}$) | 1.83 (± 0.76) | 2.41 | 0.027 |
| Flower corolla width (mm) | 0.32 (± 0.14) | 2.24 | 0.039 |

301 [†] R^2 (adj) = 0.35 ($F= 7.18, n= 26, p=0.004$)

302 * R^2 (adj) = 0.27 ($F= 4.43, n= 20, p=0.028$)

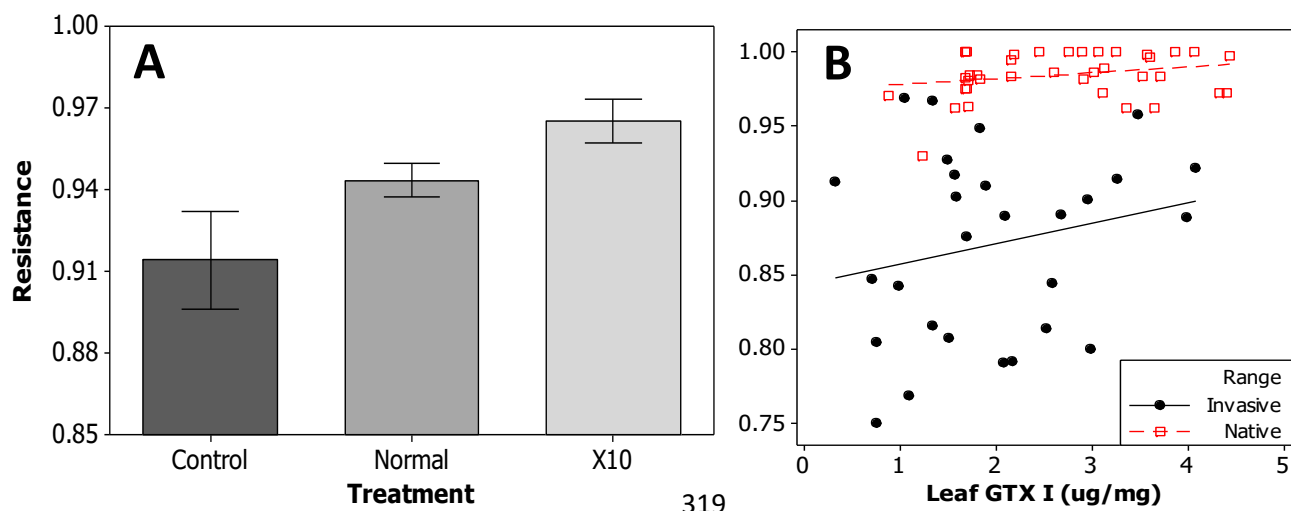
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305 Herbivores as drivers of selection on plant toxin levels

306 Evidence from controlled feeding experiments utilising an ecologically relevant herbivore of *R.*
307 *ponticum* indicated that leaf GTX I functions as an important chemical defence conferring resistance
308 to herbivory (Fig. 3a). Field observations in both the native and non-native range corroborated this
309 finding, in that herbivory levels varied according to leaf age ($\chi^2 = 1159.2, df = 1, p= \leq 0.001$), with
310 younger leaves exhibiting higher levels of damage and significantly less GTX I than older leaves (Paired
311 t-test: $t= 4.05, df= 36, p= \leq 0.001$). However, the apparent ecological value of leaf GTX I in conferring
312 resistance was not consistent across ranges (Fig 3b), with herbivore damage to plants much more
313 prevalent in the non-native rather than native range ($\chi^2 = 2181.8, df = 1, p= \leq 0.001$). Hence, in the non-
314 native range, a significant association was observed between resistance and leaf GTX I levels ($p= 0.008$)
315 together with canopy cover ($p= 0.014$) (quasi-binomial GLM: $F_{2,29}= 5.2, p= 0.014, \text{pseudo } R^2= 0.30$), while
316 neither of these variables were significant in the native range (quasi-binomial GLM: $F_{2,37}= 0.4, p= 0.700,$
317 $\text{pseudo } R^2= 0.02$).

318



319

320 **Figure 3.** (A.) Resistance to Black vine weevil (*Otiorynchus sulcatus*) feeding as conferred by grayanotoxin
321 I (GTX I mean \pm SE). Treatments represent artificial diets in which GTX I was absent (Control), or
322 incorporated at average leaf levels in *Rhododendron ponticum* (Normal), or ten times this amount (X10).
323 Each mean differed significantly from the other (at $p \leq 0.05$) according to one-tailed Tukey pairwise

324 contrasts (corrected for multiple comparisons); and (B.) the relationship between leaf GTX I levels and
325 resistance of *R. ponticum* plants to herbivory in wild populations. A non-significant 'Range X Leaf GTX I'
326 interaction revealed equivalency in this relationship across ranges (ANCOVA homogeneity of slopes: $df=$
327 $61, p= >0.05$).

328

329 **Discussion**

330 This study tested and confirmed the hypothesis that natural selection on a toxic plant chemical
331 defence varied in direction and magnitude across the native and non-native range of an invasive
332 species, given the expectation that mutualists and antagonists should exert conflicting selection
333 pressures on leaves, flowers, and nectar. In the native range, positive total selection on toxin levels in
334 flowers and nectar was as a result of an indirect selection on leaves; whereas in the non-native range
335 nectar toxin levels experienced negative total selection, while other traits were selectively neutral.

336 Two lines of evidence supported the second hypotheses tested, that pollinators are important drivers
337 of observed selection on toxic nectar. Firstly, the finding of negative selection on nectar GTX I in the
338 non-native range, coupled with observed phenotypic change in GTX I that is specific to nectar (that is
339 not seen for the comparatively less toxic GTX III [12]), provides evidence consistent with adaptive post-
340 invasion evolution driven by pollinators. Furthermore, direct investigation of biotic selection pressures
341 in both ranges revealed that plants that exhibited high nectar GTX I levels also experienced more
342 pollen limitation. The vast majority of individuals in the non-native range were pollen-limited
343 (compared to just 15 % in the native range). This high frequency and intensity of pollen limitation
344 supports that nectar toxin levels were here subject to negative pollinator-mediated selection, given
345 the importance of pollinators as selective-agents via seed production when plants are pollen limited
346 [42, 59, 60]. Furthermore, due to the demonstrated sublethal post-ingestive effects of GTX I [13],
347 established pollinators in the non-native range such as solitary bees may be differentially deterred by
348 plant individuals on the basis of nectar toxicity. This type of preferential foraging behaviour by
349 pollinators could be facilitated by the fact that high and low toxin producing plants tend to be spatially
350 aggregated at the patch-level within plant populations [12].

351 Our findings led us to partially reject our third hypothesis, that GTX I levels in nectar are phenotypically
352 correlated with those in leaves and flowers, which should therefore also show reduced levels in the
353 non-native range. While such phenotypic correlation was indeed evident in the native range, this was
354 not the case in the non-native range, where natural selection appears to have been able to act
355 independently on nectar. While theory predicts that phenotypic expression of secondary compounds
356 should become uncoupled across tissue types when these experience opposing selection pressures

357 [56, 57], such scenarios have seldom been tested or demonstrated [17]. Our study provides evidence
358 of such an uncoupling, which has seemingly permitted non-native plants to reduce nectar GTX I levels
359 without compromising the notable anti-herbivore function of GTX I in leaves and flowers. Far from a
360 mere up-loading of phloem constitutions, the production of floral nectar in plants follows a complex,
361 multi-stage process involving transport or *de novo* synthesis of components in various nectary
362 ultrastructures [1, 58]. Such processes could hence form targets for the adaptive modification of
363 nectar, which here may have permitted natural selection to act directly on nectar GTX I in the non-
364 native range, while not compromising chemical defence in other tissues. In contrast, due to the
365 observed linkage between tissue types in native plants, positive selection on leaf toxin levels resulted
366 in large indirect selection on nectar toxin levels.

367 In relation to leaf GTX I levels and biotic selection pressures imposed by herbivores, we accepted our
368 final hypothesis that that any reduction in leaf GTX I levels would represent a trade-off owing to its
369 adaptive value against herbivory. Here, positive directional selection was observed on leaf and flower
370 GTX I levels in the native range, consistent with the finding that GTX I conferred resistance against a
371 generalist herbivore of this species, and that young leaves with lower toxin levels showed more
372 herbivore damage throughout populations of both ranges. However, while there was a similarity
373 between ranges in the general form of the relationship between leaf GTX I levels and herbivore
374 resistance, this relationship was only significant in the non-native range, where *R. ponticum* has
375 evidentially experienced a notable gain in levels of herbivore damage. This scenario represents a
376 seemingly rare contradiction [61, 62] of the enemy-release hypothesis, which is often invoked to
377 explain the success of invasive species in their non-native range. Hence, explanation as to finding that
378 leaf and flower toxin levels were under positive selection in the native range, but not so in the invasive
379 range, may therefore relate to the existence of other unmeasured relevant sources of herbivory.

380 **Conclusions**

381 Where interactions involving mutualists and antagonists are mediated by the same trait in plants,
382 rarely are pollinators implicated as predominate selective agents [63]. This study therefore represents
383 the first evidence of pollinator-mediated selection acting on a defence-related compound in nectar.
384 These results also indicate how the possible microevolutionary adaptation of nectar by plants – which
385 is generally held as the most important mediator of interactions with mutualists [1] – may facilitate
386 colonization of exotic habitat, such as occurs in the invasion process. We conclude that pollinator-
387 mediated selection and the subsequent loss of nectar toxins are likely to have played a key role in
388 facilitating invasion by *R. ponticum*. These findings in addition emphasize the centrality of pollinator
389 health as a concept linked to the invasion process, and how post-invasion evolutionary pressures can

390 minimise lethal or sub-lethal effects on pollinators. However, beyond only plant invasions, the
391 generality of these findings on pollinator-mediated selection may in fact be broad – given the relatively
392 widespread occurrence of toxic nectar amongst plant families [3, 5], and where species are distributed
393 over large ranges throughout which conflicting selective pressures by pollinators and herbivores may
394 occur.

395

396 **Authors' contributions**

397 All authors contributed to the design of the study. J.S. secured funding for the project, P.E. and J.S.
398 conceived and designed the experiments, P.E. conducted the fieldwork and bioassay, and P.E. and
399 P.S. conducted the chemical analysis. P.E. undertook the statistical analysis and drafted the initial
400 manuscript. All authors contributed to the final article.

401 **Competing interests**

402 The Authors declare no competing interests.

403

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411

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