Risk factors associated with acute respiratory illnesses in athletes: A systematic review by a subgroup of the IOC consensus on "Acute respiratory illness in the athlete"

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ABSTRACT

Objective: To review risk factors associated with Acute Respiratory Illness (ARill) in athletes, including non-infectious ARill and suspected or confirmed Acute Respiratory Infections (ARinf).

Design: Systematic review.

Data sources: Electronic databases: PubMed-Medline, EbscoHost and Web of Science.

Eligibility criteria: Original research articles published between January 1990 and July 2020 in English were searched for prospective and retrospective full text studies that reported quantitative data on risk factors associated with ARill/ARinf in athletes, at any level of performance (elite/non-elite), aged 15-65 years.

Results: 48 studies (n=19390 athletes) were included in the study. Risk factors associated with ARill/ARinf were: increased training monotony, endurance training programs, lack of tapering, training during winter or at altitude, international travel and vitamin D deficits. Low tear-[SIgA] and salivary-[IgA] were immune biomarkers associated with ARill/ARinf.

Conclusions: Modifiable training and environmental risk factors could be considered by sports coaches and athletes to reduce the risk of ARill/ARinf. Clinicians working with athletes can consider assessing and treating specific nutritional deficiencies such as vitamin D. More research regarding the role and clinical application of measuring immune biomarkers in athletes at high risk of ARill/ARinf is warranted.

PROSPERO registration number: CRD42020160928

Keywords: Respiratory tract disease/infection, acute respiratory illness, upper respiratory tract infection, athlete, risk factors

INTRODUCTION

Acute respiratory illnesses (ARill), especially respiratory tract infections (ARinf), are the most common illnesses affecting athletes.^{1,2} At major events, such as Olympic and Paralympic Games, ARinf have been reported to be common among elite athletes, and can cause absence from both training and competition.^{3–5} Exercise during ARinf may increase the risk of serious health complications, such as myocarditis.⁶ In the general population, adults typically experience 2-4 ARill per year.^{7,8}

Few studies have addressed the risk factors for ARill and ARinf in athletic cohorts. To date studies have not attempted to differentiate between ARill that can include both non-infective or infective causes, and suspected or confirmed ARinf. Non-infective causes of ARill can mimic symptoms of infections. These may be due to allergies or airway inflammation caused by factors such as pollution, chemical irritants and exposure to cold or dry air. As ARill and ARinf are common medical complaints in athletes, it is important for clinicians and training staff to understand the types and magnitude of risk factors predisposing athletes to ARill and/or ARinf.

Risk factors associated with ARill and ARinf can be categorised broadly into individual athlete factors (age, gender, medical history and co-morbidities), sport (type and level of participation), training and competition factors, nutritional factors, environmental factors (season, air temperature, pollution, altitude), exposure factors (international travel, household exposure, personal hygiene, physical distancing, crowded and indoor environments), and immune / haematological risk factors and biomarkers. Cross-sectional studies of athletes indicate that individuals with high training loads have a greater frequency of ARill.^{9,10} Longitudinal studies of athletes report an increased incidence of ARill during periods of increased physical and mental stressors which may suppress both innate and adaptive immunity.^{14–18} Individual studies have reported that strenuous exercise-induced immunosuppression, mental stress, nutritional restrictions, air travel, human crowding, housing with other athletes, low temperature with low humidity, and competition all potentially increase the risk for ARinf, especially during the winter season when respiratory viruses are more prevalent.^{4,12,18} No previous systematic review has been conducted that highlights important risk factors for ARill and ARinf in athletes.

The aim of this study was to conduct a systematic review of risk factors associated with general (undiagnosed) ARill and ARinf (suspected or confirmed by laboratory identification of the pathogen) in athletes.

METHODS

Protocol and registration

A protocol was developed according to guidelines outlined in the 2020 Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) statement¹⁹ and registered with the international prospective register of systematic reviews (PROSPERO) in 2019 (registration number CRD42020160928). The PRISMA checklist is presented in Online Supplementary S1.

Study selection and eligibility criteria

Eligibility criteria were established and agreed upon by all authors based on the concepts of population and outcome (PO). Studies that met the following criteria were considered eligible for inclusion in this systematic review:

- 1. Participants, male and female, who are athletes at any level (recreational to elite) or military populations engaged in training, aged 15-65 years old,
- 2. Reporting on self-reported and/or physician diagnosed ARill, as well as clinically diagnosed and laboratory confirmed ARinf,
- 3. Reporting ARill during training, events/tournaments, multi-stage events and directly after an event,
- 4. Prior to the search strategy implementation it was agreed across all IOC consensus groups that journal articles with full-text original prospective and/or retrospective studies published in English between 1 January 1990 and 31 July 2020 will be included,
- 5. Studies reporting factor/s predisposing athletes to ARill.

Exclusion criteria were set as studies:

- 1. Conducted with a heterogeneous sample (i.e., mixed sample of athletic and non-athletic populations) without reporting individual group findings separately,
- 2. Available as an abstract only (i.e., conference presentations), qualitative or case series, discussion paper, commentary, or literature review, and
- 3. Not available in English.

While asthma and allergy can be independent risk factors associated with ARill, it should be noted they were not included in this review, which has a focus on infections as a cause of ARill.

Search strategy

Researchers systematically searched three electronic databases: PubMed-Medline, EBSCOhost and Web of Science. Medical subject heading (MeSH) terms included: upper respiratory tract infection*

OR upper respiratory illness* OR upper respiratory symptom* AND athlet* AND risk factors, and relevant exclusions (see Online Supplementary S1). A secondary search of the reference lists of included articles and hand searching in Google Scholar were performed. Further articles the authors were aware of relating to the topic were added to the search results. Duplicate articles were removed from the combined searches. Article screening and selection utilised the online tool CADIMA.²⁰ The articles were then screened independently by three reviewers (LK, JG, KM). Full texts of articles were retrieved, and a second independent screening was undertaken by four independent reviewers (LK, JG, KM, MG). Any conflicts were resolved through discussion and consensus between reviewers.

Data Extraction

Data were extracted for each study independently and agreed by consensus (WD, MM, MG, JF, KM, MS, MB, JG, ME, LK). Extracted data included: Participant details (number, age, gender), study design, level of sport performance (elite/professional to non-elite/amateur), sport type, tournament or non-tournament and statistical measures of significance for risk ratios, prevalence ratios. Data related to risk factor/s and biomarkers associated with ARill and ARinf were grouped into the following main categories: 1) demographics (age, gender), 2) sport (type and level of participation), 3) training and competition factors, 4) nutritional factors, 5) environmental / exposure factors (season, altitude, international travel, household exposure), and 6) immune / haematological risk factors and biomarkers.

Criteria and definitions

The criteria and definitions of risk factor, odds ratio, risk, risk ratio/relative risk and level of athletic performance are outlined in Online Supplementary S2.

Definitions and classification of subgroups of ARill

The methods used to diagnose ARill/ARinf in each study were classified as follows: (1) self-reported symptoms of ARill only, (2) self-reported symptoms but with an algorithm validated for ARinf, (3) self-reported symptoms of an ARinf reviewed by a physician, but without clinical or laboratory evaluation, (4) clinical diagnosis of an ARinf by a physician, based on history and clinical examination, (5) diagnosis of ARinf by a physician that was confirmed by laboratory investigation to identify a specific pathogen. Studies were classified by the five methods of diagnosis and included in one of the main and subgroups of ARill, based on a pathological classification (Online Supplementary S2).

ARill, including ARinf, frequently present with both upper and lower respiratory tract symptoms/signs and it is not always possible to clearly distinguish between these anatomical regions when classifying ARill. A limitation of this anatomical classification is that several pathogens that

cause predominantly upper ARinf can, in some cases, present with lower respiratory and/or systemic symptoms. A clear distinction was made in many studies, hence the anatomical classification was assessed in this review according to the following classifications:

- Upper (ARill or ARinf): Studies where the predominant symptoms, signs, or confirmed pathology was mainly related to the upper respiratory tract (i.e., above the larynx), or if the study specifically referred to athletes with upper ARill or ARinf. A few studies referred to ARinf with non-specific terms such as "flu", "flu symptoms", "common cold", "symptoms suggestive of influenza", "influenza symptoms" or "influenza like". Studies referring to these clinical syndromes were also included in this broad anatomical classification because they are caused by pathogens that all present with predominantly upper respiratory tract symptoms.^{7,21–}
 ²³ Notably, this includes the influenza viruses, which predominantly present with upper respiratory tract symptoms.^{7,21–}
- *Lower (ARill or ARinf)*: Studies where the predominant symptoms were below the larynx (including chest symptoms i.e., cough, chest pain), or if the diagnosis specifically referred to lower respiratory illness (tracheal, bronchial or lung pathology e.g., pneumonia).
- *General (upper/lower) (ARill or ARinf)*: Studies where there were no data to distinguish between upper or lower respiratory tract ARill or ARinf, and could include upper, lower or both.

Measures of outcome and determination of strength of association

Risk factors and biomarkers reported in the studies for undiagnosed ARill and suspected and confirmed ARinf, were listed by category of risk and strength of each association evaluated. There was *significant* heterogeneity in outcome variables reported (e.g. relative risk or % athletes affected, single or confounders analysis). As a result, a 4 level metric was developed to classify the type and strength of an association between a risk factor and ARill or ARinf as follows: no association (0, 00 or 000), some association (+), good association (++) or strong association (+++). A risk factor association was rated as weak evidence "no association" when a simple analysis was performed, such as any of the following statistical tests: descriptive analysis, Pearson's correlation analysis or grouping t-student's analysis (0). Good evidence for "no association" was rated as (00) when the study performed a multivariable analysis without mentioning the confounding variables that were taken into account, while stronger evidence for "no association" was reported as (000) i.e. when the study documented a multivariable model analysis taking confounding factors into account (e.g., sex, age, season and level of performance). A risk factor association was rated as "some" association (+) if a study documented some form of single statistical analysis. "Good" association (++) was attributed if

the study used a statistical analysis which accounted for confounding factors. A risk-factor association was rated as "strong" (+++) if the study documented a multivariable model analysis taking confounding factors into account.

Quality Assessment and risk of bias

Studies were reviewed for the quality assessment and risk of bias using a modified Downs and Black tool.²⁵ This was conducted by seven reviewers (LK, JG, MB, WD, MB, KM, MG) independently scoring the articles and then discussing differences to reach a consensus score for each article. The same reviewers determined the level of evidence using the Oxford Centre for Evidence Based Medicine (OCEBM, 2009).²⁶ The articles fell into two main categories: Observational studies of the prevalence of symptoms of ARill; or Interventional studies where the incidence of ARill was determined in response to the intervention, with or without control groups. The OCEBM level of evidence was graded using the criteria for a Symptom Prevalence Study for the observational studies based on the degree of follow up for prospective studies as level 1b for good follow-up, level 2b for retrospective studies and level 3b for non-consecutive cohort studies. The intervention studies were graded using the Therapy/Prevention studies criteria of level 1b for randomised control trials (RCTs) with narrow confidence intervals and level 2b for the non-RCT studies.

RESULTS

Study selection

Four hundred sixty-one (461) studies were identified in the search. The study selection process and reasons for excluding studies is summarised in Figure 1. Eighty four (84) full text articles were assessed for eligibility, 36 were excluded and 48 were included. The characteristics of the 48 studies are presented in online Supplementary S3, and the quality assessment in online Supplementary S4. The 48 studies had a total of 19390 (range: 9 to 12594) participants. Studies were conducted across 17 sports and five performance levels: only elite/professional athletes (n=26; 54.2%); only recreational/trained/competitive athletes (n=16; 33.3%); mixed levels (n=6; 12.5 %).

Number of studies by pathological and anatomical classification of ARill

The pathological and anatomical classifications of ARill for each study are provided in Table 1. Of the 48 studies, 40 (83.3%) reported upper ARill, eight (16.7%) reported general ARill, with no studies reporting lower ARill only. Seventeen (35.5%) studies reported undiagnosed ARill. Of the 31(64.5%) studies classified as ARinf, 26 (54%) were suspected infections and five (10.4%) were confirmed ARinf.



Figure 1. Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) flow diagram visualizing the selection process of identified, screened and included articles following assessment of the eligibility criteria.

| Table 1. Number of studies (II, 70 of tot | al studies) by | pathological | and anatomic | al classifications. |
|---|----------------|--------------|--------------|---------------------------------------|
| | Upper | General | Total | References |
| All studies | 40 (83.3%) | 8 (16.7%) | 48 | |
| All general (undiagnosed) ARill | 16 (33.3%) | 1 (2.1%) | 17 (35.5%) | |
| Self-reported symptoms only | 6 (12.5.6%) | 1 (2.1%) | 7 (14.6%) | 29,31,42,49,57,67,70 |
| Self-reported symptoms with an algorithm | 7 (14.6%) | - | 7 (14.6%) | 32,33,41,43,58,61,68 |
| Symptoms reviewed by a physician, but | 1 (2.1%) | - | 1 (2.1%) | 46 |
| without clinical or laboratory evaluation | | | | |
| Clinical diagnosis by a physician, based on | 2 (4.2%) | - | 2 (4.2%) | 62,69 |
| history and examination | | | | |
| All infective (ARinf) | 24 (50.0%) | 7 (14.5%) | 31 (64.5%) | |
| Suspected infective | 19 (39.5%) | 7 (14.5%) | 26 (54%) | |
| Self-reported symptoms with an algorithm | 12 (25%) | 1 (2.1%) | 13 (27.1%) | 9,34,35,37,39,40,50,54,56,59,60,64,65 |
| Symptoms reviewed by a physician, but | 1 (2.1%) | - | 1 (2.1%) | 66 |
| without clinical or laboratory evaluation | | | | |
| Clinical diagnosis by a physician, based on | 6 (12.5%) | 6 (12.5%) | 12 (25%) | 13,27,28,30,36,38,44,45,47,48,55,63 |
| history and examination | | | | |
| Confirmed infective | 5 (10.4%) | - | 5 (10.4%) | 10,51–53,71 |
| Diagnosis by a physician and confirmed by | | | | |
| laboratory investigation to identify a | | | | |
| specific pathogen | | | | |
| | | | | |

Table 1. Number of studies (n; % of total studies) by pathological and anatomical classifications.

Risk factors and biomarkers associated with ARill and ARinf

Risk factors and biomarkers associated with general (undiagnosed) ARill

The main categories of risk factors and biomarkers associated with general (undiagnosed) ARill, by category of risk and strength of each association are presented in Table 2a. Risk factors that showed a strong association (+++) with general (undiagnosed) ARill were: being a less competitive athlete, elevated white blood cell and neutrophil counts, and a lower serum Vitamin D concentration. Risk factors for which there was strong evidence for no association (000) with general (undiagnosed) ARill were intensified phase of training, competition phase, detection of IgE antibodies to aero-allergens, and a reduction in salivary flow rate. Of interest is that there was both strong evidence for a positive association (+++) and no association (000) between ARill and increased training load.

Risk factors and biomarkers associated with suspected ARinf

The main categories of risk factors and biomarkers associated with suspected ARinf, by category of risk and strength of each association are presented in Table 2b. Risk factors that showed a strong association (+++) with suspected ARinf were: increments in training load, endurance training, training monotony, training at altitude, winter season, post international travel, less competitive athletes, having reduced serum Vitamin D concentration, and experiencing prior episodes of respiratory infection. A strong association (+++) was found between lower risk of suspected ARinf and autumn season, as well as the tapering phase of training and increased training intensity. Risk factors for which there was strong evidence for no association (000) with suspected ARinf were: age, gender and household family exposure. Of interest is that there was both strong evidence for a positive association (+++) and no association (000) between suspected ARinf and increased training load, increased speed and strength training, and the competition period.

Risk factors and biomarkers associated with confirmed ARinf

The main categories of risk factors and biomarkers associated with confirmed ARinf, by category of risk and strength of each association are presented in Table 2c. Risk factors and biomarkers that showed a strong association (+++) with confirmed ARinf were: increasing training intensity, lower salivary-[IgA] (pre-season, pre-training and across a season) and reduced tear salivary-[IgA] and secretion rates. The only risk factor where there was strong evidence for no association (000) with suspected ARinf was post-season training salivary-[IgA].

DISCUSSION

The aim of this study was to conduct a systematic review of risk factors associated with general (undiagnosed) ARill and ARinf (suspected or confirmed) in athletes. The 48 studies meeting the

Table 2a. Main categories of risk factors and biomarkers associated with general (undiagnosed)ARill, by category of risk and strength of each association.

| Main Categories of Risk Factors and Biomarkers Assessed | Association Identified | Strength | Confounders | Variables adjusted for (Confounders/ Multivariable model) | Study |
|--|---------------------------|----------|--|---|------------------------------------|
| Demographics factors | , | | 1 | | |
| Age | No | 0 | - | No/No | Blume et al. ⁶⁸ |
| Gender | No | 0 | - | No/No | Blume et al. ⁶⁸ |
| Sport (type and level of participation |) | | | | |
| Being at a less competitive level | Yes | +++ | Age, age of menarche, number of inhabitants living in the same house | Yes/Yes | Novas et al. ³⁸ |
| | | + | - | No/No | Matthews et al. ²⁹ |
| Greater in runners | Yes | + | - | No/No | Ihalainen et al. ³² |
| Training & competition factors | | | | | |
| Increased training load | Yes | +++ | Subject (pre- exercise values), subject and session- intensity. | Yes/Yes | Novas et al. ⁵⁷ |
| | Yes | + | - | No/No | Novas et al. ⁵⁸ |
| | No | 000 | Training weeks and number of players | Yes/Yes | Tiernan et al. ⁴⁶ |
| | | 0 | - | No/No | Matthews et al. ²⁹ |
| Intensified phase of training | Yes | + | - | No/No | Novas et al. ⁵⁷ |
| | No | 000 | Training macrocycle, internal-TL values, wellbeing, muscle soreness ratings and age. | Yes/Yes | Thornton et al. ⁴² |
| Competition phase | No | 000 | Training macrocycle, internal-TL values, wellbeing, muscle soreness ratings and age. | Yes/Yes | Thornton et al. ⁴² |
| Nutritional factors | 1 1 | | | | G 160 |
| Lower [Vitamin-D] | Yes | +++ | Age, gender and years of training | Yes/Yes | Cox et al. ⁰⁹ |
| Serum [Vitamin-D] in both winter and summer | No | 0 | - | No/No | Scullion et al. ⁶⁷ |
| Environmental / exposure factors | | | | | |
| Winter | Yes | + | - | No/No | Scullion et al. ⁶⁷ |
| Longer International traveling | Yes | + | - | No/No | <i>Fowler et al.</i> ⁴¹ |
| Household size | No | 00 | Age, age of menarche, number of inhabitants living in the | Yes/No | Novas et al. ⁵⁷ |

| | | | same house | | |
|--|---------------|-----|---|---------|---------------------------------------|
| Immune / haematological risk factors | s and biomark | ers | | | |
| Elevated WBC counts | Yes | +++ | Age, gender and years of training | Yes/Yes | Cox et al. ⁶⁹ |
| Elevated neutrophil count | Yes | +++ | Age, gender and years of training | Yes/Yes | Cox et al. ⁶⁹ |
| Detection of IgE antibodies to aero- | Yes | + | - | No/No | Reid et al. ⁶² |
| allergens | No | 000 | Age, gender and years of training | Yes/Yes | Cox et al. ⁶⁹ |
| Higher atopic AQUA scores | Yes | + | - | No/No | Ansley et al. ³³ |
| Higher rates of expression of EBV- DNA in saliva (viral reactivation) | Yes | + | - | No/No | <i>Reid et al.</i> ⁶² |
| Lower resting and post-exercise [IL- 1ra] in illness-prone athletes | Yes | + | - | No/No | $Cox \ et \ al.^{31}$ |
| High expression IL-2 genotype (GG)– lower incidence | Yes | + | - | No/No | Cox et al. ⁷⁰ |
| Low expression IL-4 genotype (CC) | Yes | + | - | No/No | Cox et al. ⁷⁰ |
| High expression IL-6 genotype (GG) | Yes | + | - | No/No | Cox et al. ⁷⁰ |
| Low expression IL6 genotype (CC) | Yes | + | - | No/No | Zehsaz et al. ⁶¹ |
| Expression of IL-1ra, IL-8, IL-10, IFNy genotypes | No | 0 | - | No/No | Cox et al. ⁷⁰ |
| Higher post-exercise [IL-6] | Yes | + | - | No/No | Cox et al. ³¹ |
| Lower resting [IL8] in illness-prone athletes | Yes | + | - | No/No | Cox et al. ⁷⁰ |
| High expression IL-10 genotype (GG) | Yes | + | - | No/No | Zehsaz et al. ⁶¹ |
| Lower resting and post-exercise [IL- 10] in illness-prone athletes | Yes | + | - | No/No | $Cox \ et \ al.^{31}$ |
| Changes in post-exercise [IL-2], [IL-4], [IL-12] | No | 0 | - | No/No | Cox et al. ⁷⁰ |
| Reduction in salivary-AA, and IgM to total protein ratio | Yes | + | - | No/No | <i>Ihalainen et al.</i> ³² |
| Lower serum [IgG3] | Yes | + | - | No/No | Reid et al. ⁶² |
| Reduction in salivary-[Lysozyme] | No | 0 | - | No/No | Cunniffe et al.43 |
| Reduction in salivary flow rate | No | 000 | Training weeks and number of players | Yes/Yes | Tiernan et al. ⁴⁶ |
| | | 0 | - | No/No | Nakamura et al. ⁴⁹ |

Note: Strength of association, 0; no association with multiple models and/or correction for confounders, (00/000); some association, +; good association, ++; strong association, +++ **Abbreviations:** AA, Alpha amylase; EBV, Epstein Barr Virus; IgA, Immunoglobulin A; IgG3, Immunoglobulin G3; WBC, White blood cell; AQUA, Automated quantitative analysis.

Table 2b. Main categories of risk factors and biomarkers associated with suspected acute respiratory infection (ARinf) by category of risk and strength of each association.

| Main Categories of Risk Factors and Biomarkers Assessed | Association Identified | Strength | Confounders | Variables adjusted for (Confounders/ Multivariable model) | Study |
|--|---------------------------|----------|---|---|--------------------------------------|
| Demographic factors | | | | | |
| Younger athletes in the illness prone group | Yes | + | - | No/No | <i>Gleeson et al.</i> ⁶⁵ |
| Age | No | 000 | Performance level, sex, training phase and load, season | Yes/Yes | Svendsen et al. ³⁵ |
| Gender | No | 000 | Time, age, sex, training centre, and competition level | Yes/Yes | Hellard et al. ⁵⁵ |
| | No | 0 | - | No/No | He et al. ⁶⁴ |
| Sport (type and level of participa | tion) | | | | |
| Participating in endurance sports | Yes | ++ | Sex, type of sport | Yes/No | Edouard et al. ²⁸ |
| Participating in athletics compared to other Paralympic sports | Yes | + | - | No/No | Schwellnus et al. ³⁵ |
| Completing a marathon | No | 0 | - | No/No | <i>Furusawa et al.</i> ⁴⁰ |
| Being at a less competitive level | Var | +++ | Performance level, sex, training phase and load, season | Yes/Yes | Svendsen et al. ³⁵ |
| | Yes | +++ | Age, sex, competition level, season | Yes/Yes | Hellard et al. ¹³ |
| Training & competition factors | • | | | | · |
| Increased training load | | +++ | Age, sex, competition level, season | Yes/Yes | Hellard et al. ¹³ |
| | | +++ | Time, age, sex, training centre, and competition level | Yes/Yes | Hellard et al. ⁵⁵ |
| | | + | - | No/No | Moreira et al. ³⁴ |
| | Ves | + | - | No/No | Rama et al. ⁵⁴ |
| | 103 | + | - | No/No | Gleeson et al. ⁶⁵ |
| | | + | - | No/No | Fricker et al. ³⁰ |
| | | + | - | No/No No/No | Leicht et al. ³⁷ |
| | | + | - | No/No | Hausswirthet al ⁶⁰ |
| | | + | - | No/No | Ikonen et al. ⁶⁶ |
| | | + | - | No/No | Milanez et al. ⁵⁰ |
| | | + | - | No/No | Gleeson et al. ⁶⁵ |
| | No | 000 | Performance level, sex, training phase and load, season | Yes/Yes | Svendsen et al. ³⁵ |
| | | 0 | - | No/No | Dressendorfer et al. ³⁶ |
| | | 0 | - | No/No | Neville et al. ⁹ |
| Increased training intensity | Yes | + | - | No/No | Brisola et al. ³⁹ |
| T 14 ' ' ' 4 | No | 0 | - - | No/No | Dressendorfer et al. ³⁰ |
| Lower incidence | Yes | +++ | sex, training phase and load, season | Yes/Yes | Svenasen et al. ²⁵ |
| Increased strength & speed training | Yes | +++ | Age, sex, competition level, season | Yes/Yes | Hellard et al. ¹³ |
| | No | 000 | Performance level, sex, training phase and load, season | Yes/Yes | Svendsen et al. ³⁵ |
| Increased training monotony | Yes | +++ | Performance level, sex, training phase | Yes/Yes | Svendsen et al. ³⁵ |

| | | | and load, season | | |
|------------------------------------|---------------|--------|-----------------------|--------------|-------------------------------------|
| Endurance preparation phase | | | Performance level, | | Svendsen et al. ³⁵ |
| | Yes | +++ | sex, training phase, | Yes/Yes | |
| | | | and load, season | | ~ |
| Tapering phase – lower incidence | | | Performance level, | 37 /37 | Svendsen et al. ⁵⁵ |
| | | +++ | sex, training phase | Yes/Yes | |
| | Yes | | and load, season | | |
| | | | Age, sex, | Vaa/Vaa | Hellara et al. ¹⁵ |
| | | | season | I es/ I es | |
| Competition period | | | Gender age sport | | Schwellnus et al ⁴⁴ |
| competition period | Yes | +++ | types | Yes/Yes | Serweitnus et ut. |
| | 100 | + | - | No/No | Brisola et al. ⁵⁶ |
| | | | Age, sex, | | Hellard et al. ¹³ |
| | | 000 | competition level, | Yes/Yes | |
| | No | | season | | |
| | INO | | Time, age, sex, | | Hellard et al. ⁵⁵ |
| | | 000 | training centre, and | Yes/Yes | |
| | | | competition level | | |
| Nutritional factors | I | 1 | 1 1 | | 1 |
| Reduced serum [Vitamin-D] | Yes | +++ | Baseline values, | Yes/Yes | Hanstock et al. ³⁷ |
| | 105 | | time effect, exercise | 105,105 | |
| Vitamin D supplementation – | Yes | + | _ | No/No | He et al. 64 |
| lower incidence | | | | | 166 |
| Lower intake of arginine and | V | | | | <i>Ikonen et al.</i> ⁶⁶ |
| alanine amino acids during the | Yes | + | - | INO/INO | |
| Environmental (exposure factors | | | | | |
| Environmental / exposure factors | | 1 | Derformance level | | Swandson at al 35 |
| white | | +++ | sex training phase | Ves/Ves | svenusen et ut. |
| | | | and load, season | 103/103 | |
| | Yes | | Age, sex. | | Hellard et al ¹³ |
| | | +++ | competition level. | Yes/Yes | |
| | | | season | | |
| | | + | - | No/No | Fahlman and Engels ²⁷ |
| Autumn – Lower incidence | | | Team home | | Schwellnus et al.45 |
| | | | country, season, | | |
| | | | intercontinental | | |
| | Yes | +++ | travel and duration | Yes/Yes | |
| | | | in a specific travel | | |
| | | | stage of the | | |
| E (1'1) | | | tournament. | | <u> </u> |
| eltitude >1500 meel | Vas | | Performance level, | Vac/Vac | Svenasen et al. ³⁵ |
| attitude ~1500 masi | 1 05 | | sex, training phase | 1 05/ 1 05 | |
| Longer International traveling | | | Derformance level | | Swandson at al 35 |
| Longer mornational travening | | +++ | sex. training nhase | Yes/Yes | svenusen et ut. |
| | | | and load. season | 100, 100 | |
| | | | Team home | | Schwellnus et al.45 |
| | V- | | country, season, | | |
| | res | | intercontinental | | |
| | | +++ | travel and duration | Yes/Yes | |
| | | | in a specific travel | | |
| | | | stage of the | | |
| | | | tournament. | ST 5- | 1.0 1. 1.24 |
| Increased psychological stress | Yes | + | - | No/No | Moreira et al. ⁵⁴ |
| Description of 11 | No | 0 | - | No/No | Milanez et al. ⁵⁰ |
| Poor sleep quality | Yes | + | Doutours 1 1 | INO/INO | Hausswirthet al. ⁰⁰ |
| nousenoia iamily exposure | No | 000 | Performance level, | Vac/Vac | svenasen et al. ³⁵ |
| | INO | 000 | sex, utaining phase | 1 CS/ 1 CS | |
| Immune / haematological risk for | tors and biom | arkers | anu ioau, season | | |
| Prior respiratory tract infections | | | Age sex | | Hellard et al ¹³ |
| The respiratory fract infections | Yes | +++ | competition level. | Yes/Yes | 1101101101111 |
| | | | season | 1 - 5/ 1 - 6 | |
| Higher rates of expression of | Yes | + | - | No/No | <i>Gleeson et al.</i> ⁶³ |

| EBV-DNA in saliva (viral reactivation) | No | 0 | - | No/No | Yamauchi et al. ⁴⁷ |
|---|-----|---|---|-------|-------------------------------------|
| Lower CD56+ cell counts (neutrophil cell marker) | Yes | + | - | No/No | Rama et al. ⁵⁴ |
| Higher CD56bright:CD56 dim ratio | Yes | + | - | No/No | Rama et al. ⁵⁴ |
| Lower salivary-IgA concentration | 37 | + | - | No/No | <i>Gleeson et al.</i> ⁶³ |
| | Yes | + | - | No/No | Fahlman and Engels ²⁷ |
| | Na | 0 | - | No/No | <i>Gleeson et al.</i> ⁶⁵ |
| | INO | 0 | - | No/No | Leicht et al. ³⁹ |
| Larger decrease in salivary [IgA] across training weeks | Yes | + | - | No/No | <i>Milanez et al.</i> ⁵⁰ |
| Reduced pre-training salivary | V | + | - | No/No | Neville et al. ⁴⁸ |
| [IgA] | Yes | + | - | No/No | Milanez et al. ⁵⁰ |
| Reduced salivary-IgA secretion | | + | - | No/No | <i>Gleeson et al.</i> ⁶⁵ |
| rate | | + | - | No/No | Fahlman and Engels ²⁷ |
| | Yes | + | - | No/No | Yamauchi et al.47 |
| | | + | - | No/No | Neville et al. ⁴⁸ |
| | | + | - | No/No | Milanez et al. ⁵⁰ |
| | No | 0 | - | No/No | Leicht et al. ³⁹ |
| Genetic risk score for predisposition to pro- inflammatory cytokine responses | Yes | + | - | No/No | <i>Gleeson et al.</i> ⁶³ |
| High expression IFN-y genotype | Yes | + | - | No/No | Gleeson et al. ⁶³ |
| Higher IFN-y production in | Yes | + | - | No/No | Gleeson et al. ⁶⁵ |
| illness prone athletes | No | 0 | - | No/No | Gleeson et al. ⁹ |
| TNF-a production | No | 0 | - | No/No | Gleeson et al. ⁶⁵ |
| IL-1ra cytokine genotypes | No | 0 | - | No/No | Gleeson et al. ⁶³ |
| IL-1ß production | No | 0 | - | No/No | <i>Gleeson et al.</i> ⁶⁵ |
| Higher IL-2 production | | + | - | No/No | Gleeson et al. ⁹ |
| 8 1 | Yes | + | - | No/No | <i>Gleeson et al.</i> ⁶⁵ |
| Higher IL-4 cytokine production | | + | - | No/No | Gleeson et al. ⁹ |
| | Yes | + | - | No/No | Gleeson et al. ⁶⁵ |
| | No | 0 | - | No/No | Gleeson et al. ⁶³ |
| High expression IL-6 (CC) and IFN _X (AA) genotypes | Yes | + | - | No/No | <i>Gleeson et al.</i> ⁶³ |
| Higher IL-6 production | No | 0 | - | No/No | Gleeson et al. ⁶⁵ |
| Higher IL-8 production | No | 0 | - | No/No | Gleeson et al. ⁶⁵ |
| Higher expression IL-8 cytokine genotypes | No | 0 | - | No/No | <i>Gleeson et al.</i> ⁶³ |
| Higher post exercise IL-10 | | + | - | No/No | <i>Gleeson et al.</i> ⁶⁵ |
| production | Yes | + | - | No/No | <i>Gleeson et al.</i> ⁹ |
| Higher expression IL-10 cytokine | No | 0 | - | No/No | <i>Gleeson et al.</i> ⁶³ |
| High expression IL-17 cytokine | No | 0 | - | No/No | <i>Gleeson et al.</i> ⁶³ |
| Higher plasma [IgM] | Yes | + | - | No/No | Gleeson et al.65 |
| Plasma [IgA] | No | 0 | - | No/No | Gleeson et al 65 |
| Plasma [IgG] | No | 0 | - | No/No | <i>Gleeson et al.</i> ⁶⁵ |

Note: Strength of association, no association (0/00/000), + some association, ++ good association, +++ strong association **Abbreviations**: EBV, Epstein Barr Virus; IgA/G/M, Immunoglobulin A/G/M; IL, Interleukin; INF, Interferon; TNF, Tumour necrosis factor; masl, meter above sea level. **Table 2c.** Main categories of risk factors and biomarkers associated with confirmed acute respiratory infection (ARinf) by category of risk and strength of each association.

| Main Categories of Risk Factors and Biomarkers Assessed | Association Identified | Strength | Confounders | Variables adjusted for (Confounders/ Multivariable model) | Study |
|---|---------------------------|----------|---|---|-------------------------------------|
| Sport & level of athlete | • | • | • | · | |
| Being a higher level athlete | Yes | + | - | No/No | Spence et al. ¹⁰ |
| Training & competition factors | | | | | |
| Increased training intensity | Yes | +++ | Baseline measures | Yes/Yes | Hanstock et al ⁷¹ |
| Immune / haematological risk factors a | nd biomarkers | - | 1 | 1 | - |
| Decline in Salivary [IgA] across a training season (slope) | Yes | +++ | Gender, age, training intensity and volume, psychological stress | Yes/Yes | Gleeson et al. ⁵¹ |
| Lower pre-season salivary-[IgA] | Yes | +++ | Gender, age, training intensity and volume, psychological stress | Yes/Yes | Gleeson et al. ⁵¹ |
| Pre or late season salivary-[IgA] | Yes | + | - | No/No | Gleeson et al. ⁵³ |
| Lower pre-training salivary-[IgA] | V | +++ | | Yes/Yes | <i>Gleeson et al.</i> ⁵¹ |
| | Y es | + | - | No/No | <i>Gleeson et al.</i> ⁵³ |
| Post-season training salivary-[IgA] | No | 000 | Gender, age, training intensity and volume psychological stress | Yes/Yes | Gleeson et al. ⁵¹ |
| Higher salivary-IgA secretion rate | Yes | + | - | No/No | Hanstock et al ⁷¹ |
| Lower pre-season salivary-[IgA1] | Yes | + | - | No/No | Gleeson et al. ⁵² |
| Pre or late season salivary-[IgA2] or ratios of salivary [IgA1]:[IgA2] | No | 0 | - | No/No | Gleeson et al. ⁵² |
| Reduced tear-[SIgA] | Yes | +++ | Baseline measures | Yes/Yes | Hanstock et al ⁷¹ |
| Reduced tear-SIgA secretion rate | Yes | +++ | Baseline measures | Yes/Yes | Hanstock et al ⁷¹ |

Note: Strength of association: no association (0/00/000), + some association, ++ good association, +++ strong association. **Abbreviations**: IgA: Immunoglobulin A; SIgA: secretory IgA;

eligibility criteria were graded as good or excellent, providing confidence in the quality of the studies. However, the small number of studies assessing each risk factor or biomarker made it difficult to draw firm conclusions for most risk factors. In addition, the differences in the methodologies for classifying respiratory illnesses/infections further impaired comparisons. Further discussion of the review findings now focuses on the evidence for associations between increased risk for ARill/ARinf in athletic populations and risk factors in six main categories.

Demographic factors

The findings of this review show that age and gender were not associated with increased risk of any ARill or ARinf (suspected or confirmed).

Sport type and level of participation

In general, sport type was not strongly associated with increased risk of ARill or ARinf (suspected or confirmed). There was some evidence of increased risk of ARill and ARinf in endurance athletes and runners specifically. There was a lower risk of prolonged ARill (symptom days) for elite athletes.^{37,68,72} One study hypothesized that the individual training load threshold, above which the risk of illness increases,⁷³ is lower in national level athletes than in international athletes. Other studies concluded that the differences may relate to underlying genetic predispositions for better resistance to infections^{61,70} or lower pro-inflammatory responses to infection that present as a reduced incidence of ARill.^{63,70,74,75} Previous research has suggested that higher-level athletes (top professional or elite) are linked to a better athletic lifestyle (personal, academic or professional schedules; better recovery, sleep quality or nutrition) that reduces the risk of ARill.^{14,76,77} One possibility, not examined, was that the differences are related to the type of sport rather than the level of performance.

Training and competition risk factors

While each risk factor had studies with conflicting results, the review findings for training factors indicated ARill/ARinf, irrespective of classifications, were mostly associated with increased training intensity, endurance phase training and competition periods, but there was a potential lower risk in the tapering phase of training. There was a higher risk for ARill/ARinf in less competitive level athletes, endurance sports and younger athletes. Training monotony, training in winter, at altitude and after international travel across time zones all increased the risk of ARill/ARinf.

Although the assessment of training intensity/load alone gave mixed results, the review indicates that high intensity training is a significant risk factor in athletes who experience recurrent episodes of ARinf/ARill and altered immune status.^{10,62,69} Increments in high intensity training, including speed and strength training, were associated with a higher risk of ARinf/ARill in these athletes.^{62,71} Intense exercise, particularly in endurance sports, can induce significant immune system disturbances.^{78,79} This review confirms findings of individual studies reporting an increase in ARinf/ARill symptoms during training periods characterized by high loads imposed continuously over several weeks or

months.^{53,80,81} The accumulation of elevated training loads without adequate recovery may be associated with a chronic depletion of cellular and mucosal immune parameters, which may lower resistance to potential viral^{51,53} and non-viral pathogens,¹⁴ or allow viral reactivation,^{51,53} thereby partially explaining the higher incidence of ARinf/ARill symptoms.^{9,54,81,82}

Nutritional factors

Vitamin D is an important component for effective immunity.^{83,84} The review confirmed previous research showing a vitamin D deficit predisposes athletes to longer and more severe ARill, compared to non-deficit athletes.^{59,69,71} He et al.,⁵⁹ found that vitamin D supplementation reduced the incidence of ARill. Scullion et al.,⁶⁷ found that multi-vitamin supplementation in an athlete's diet did not result in fewer ARill in winter compared to summer, and also found that an overload of vitamin D did not reduce the prevalence of ARill in athletes.

Environmental and exposure factors

Seasonality

Seasonal factors are important parameters to consider, as external factors can influence and increase the risk of ARinf/ARill.^{85–87} This review showed a consistent association of increased ARill with the winter months, supporting the previously established relationship of cold environments with a higher incidence of ARinf/ARill episodes and symptoms.^{14,59} The exposure to respiratory pathogens is highest in winter, but also significant in autumn and spring.^{35,55,67} Spring is associated with higher pollen counts that can cause symptoms of ARill in susceptible athletes, causing eosinophilic airway inflammation that is often confused with the symptoms of ARinf.^{35,62,69,88–91} Cold air can also damage the respiratory epithelium due to airway drying causing airway inflammation.^{89,92} These findings mirror the seasonal patterns for acute ARill and infections in the general population, as winter is characterized by a surge in viral acute respiratory infections.⁹³

Furthermore, during the colder months selected hormones that regulate immune function and vitamin D concentrations are at their lowest. Recent research indicates that a vitamin D deficit is a predictor of infections,⁵⁹ but supraphysiological doses of vitamin D do not protect against respiratory infections.⁹⁴ In the Northern Hemispheres, winter-time is usually characterized by increments in load in certain sports such as skiing, skating and ice hockey, and the intense competition period coincides with the winter season^{12,13,54,95} which potentially accentuates immune-suppression and increases the risk of infection. A similar pattern is evident in the Southern Hemisphere with swimmers preparing in winter months for major international competitions typically held in the Northern Hemisphere summer.^{10,30} However, time of year (season) appears to influence infection risk to a lower degree than the impact of training phase/type of sport.

International travel

International travel was shown to be a significant risk factor for ARill/ARinf³⁵ when athletes travelled across $>5^{45}$ and $>6^{41}$ time zones. Svendsen et al.³⁵ noted athletes were five times more likely to report symptoms the day following international air travel. Studies have reported that medical illness (most commonly affecting the respiratory system) affects elite athletes while travelling to international competitions.^{12,41,44,45} The reasons for a higher incidence of illness/infection/symptoms during international travel include: drying of respiratory epithelium, close contact with air travellers and exposure to re-circulated air (infections), time-zone changes associated with sleep/circadian rhythm disruption, variation of diet. Other travel factors that can augment the risk of ARill include: exposure to different environmental conditions (temperature, humidity, atmospheric pollution, aeroallergens) or exposure to different strains of pathogenic organisms, and high population density at competition venues.

Altitude

It is well established that ascent to high altitude alters physiological and metabolic function and can influence immune function.⁹⁶ In this review, training at altitude was shown to increase the risk of ARill but not when findings were adjusted for sex, performance level, training phases and season.³⁵ Tiollier et al.⁹⁷ found no significant differences in mucosal immunity between elite cross-country skiers sleeping at 2500–3500m above sea level and training at 1200m for 18 days, compared with a control group living and training at 1200m. However, the typical cold and dry conditions of training at altitude may present with upper respiratory symptoms due to airway drying and inflammation and be considered a risk factor for non-infective ARill through this effect on respiratory mucosal membranes.^{98,99}

Immune / haematological biomarkers and risk factors

Changes in systemic and mucosal immune parameters have been extensively studied in response to exercise training and competitions at all levels of sports and in many different types of sports. This systematic review of associations between immune parameters with ARinf/ARill revealed only a limited number of studies combining both ARill and exercise/training measures. The major factor affecting the immune response that appears to be associated with a higher risk of upper ARinf/ARill in athletes is a reduction in tear or salivary-[IgA]. Salivary-[IgA] is the most studied immune parameter and represents a biomarker for altered mucosal immunity in the respiratory tract. It is well established that low concentrations of salivary-[IgA] at mucosal surfaces is a risk for mucosal infections in the general population.¹⁰⁰ Salivary-[IgA] plays a major role in immune defence not only at mucosal surfaces but also in responding to and eliminating pathogens that cross mucosal surfaces.^{101,102} The studies in this review revealed an association between the appearance of EBV-DNA (viral reactivation) in saliva and the incidence of ARinf/ARill and the time-frame for

association with low concentrations of salivary-IgA. These biomarkers reflect immune parameters that are known risk factors associated with respiratory illness in the general population and have therefore been evaluated as tools to monitor ARill/ARinf risks in athletic populations known to have exercise-induced alterations in immune function and parameters.^{4,14,78,79,82,103–106}

Salivary & Tear IgA

Regardless of the methodology used for characterising ARinf/ARill, this review found a consistent association between lower concentrations of salivary-[IgA] and tear salivary-[IgA] with an increased incidence of ARinf/ARill, with 83% of studies reporting this association. The majority, but not all studies that assessed secretions rates, also found an increased incidence of ARinf/ARill with reduced secretion rates of salivary-IgA or tear-IgA. It is possible that the one study with the reverse finding of higher salivary-IgA concentrations with increased ARill was sampled during the infective period⁷¹ when salivary-IgA would be expected to increase in response to an infection in subjects with a fully functioning immune system.

The cumulative effects of long term training at high loads and intensity were observed in a decline in immune protection over time. Pre-training or pre-season lower salivary-IgA concentrations were shown to be associated with the increase in episodes of ARill/ARinf, symptom duration^{74,75} and severity in elite swimmers.^{90,107} A 65% reduction in salivary-IgA concentration was reported 1–2 weeks before the appearance of a suspected ARinf in rugby union players.⁴⁶ Similarly, in a cohort of elite yacht racing sailors, low individual relative salivary-IgA values (<40 % drop) suggested a 48% chance of an ARill within 3 weeks.⁴⁸ In elite swimmers, an additional infection was observed for each 10 % drop (slope) of pre-training salivary-IgA level over time (per month).⁵¹ In recreationally active individuals (various sports), low salivary-IgA (<5.5 µg/mL) and reduced secretion rate (>30 %) was associated with ARill in the week following a competition.⁷¹ The reductions in salivary-IgA concentration and secretion rates may have been the result of increments in training load^{48,50} or inadequate recovery time between training sessions.^{106,108}

Strengths and limitations

The quality of the studies included in this review and the variables explored as risk factors for ARill/ARinf provides some direction on the topic, specifically for elite/high performance athletes. A strength of this review is that it followed a systematic approach for inclusion and although a metaanalysis could not be performed, studies were reviewed for the quality assessment and risk of bias using a modified Downs and Black tool.²⁵

However, this review has some limitations. First, while a consensus of the research group was used to reduce inclusion/exclusion bias, we acknowledge that the selected criteria may have (to a certain

extent) led to selection bias. For example, the inclusion of studies in the English Language might have resulted in language restriction bias. There are other possible biases not considered by the selected appraisal tool in this study, which have the potential to affect study outcomes. For example, measurement bias could result from selected studies reporting self-reported symptoms only, without clinical verification by a physician. Additionally, residual confounding bias could result from studies which did not adequately consider adjustments of the confounders when reporting the strength of association. Further, sparse data bias,¹⁰⁹ may have arisen in studies which had fewer participants, subsequently influencing the odds ratio and relative risk outcomes, with considerable upward biases when there were minimal athletes at key combinations of the outcome, exposure and covariates.

Second, the focus on statistically positive findings (p<0.05) may result in researchers losing results reporting some evidence that could be a clinically relevant factor associated with ARill. Third, the differences in methodological design, definitions of ARill/ARinf, outcome measures within diagnostic methodologies and heterogeneity of athletes' levels of performance and sports codes made it difficult to interpret the magnitude of each risk factor. Also, the approach we adopted might be considered "reductionist' in the identification and stratification of risk factors. Indeed, there is considerable complexity of these identified risk factors and their interaction with other risk factors for example, the interactions of training variations and dietary changes on immune function. Fourth, only a few studies identified the infections by clinical assessment and confirmed with laboratory diagnosis. Fifth, asthma, atopy and allergy were excluded as a risk factor for ARill. Sixth, this review considered research published only in the English language, such that relevant studies conducted in non-English languages were overlooked.

The broad search strategy provided a degree of confidence that, within the inclusion criteria of risk factors for respiratory infections/illnesses, the studies were of a high level of quality. Interpretation of findings should consider that there are potentially other influences on the risks for ARill/ARinf than those examined. Future studies would need to standardise diagnostic methods, and outcome measurements to allow comparisons between studies, variables and to enable a future meta-analysis.

SUMMARY AND CONCLUSIONS

The review identified several modifiable risk factors that could be considered by sports coaches when preparing training programs, particularly for athletes who experience recurrent episodes of ARill/ARinf and those at a less competitive level (Table 3). Risk factors included increased training monotony, endurance training programs, lack of tapering, training during winter and at altitude, and international travel. It is also important for clinicians working with athletes to consider vitamin D deficits, particularly those prone to repeated ARill/ARinf. Biomarkers for monitoring athletes at a

| Pathological classification | Strong evidence supporting a positive association | Strong evidence supporting no association | Strong evidence supporting both a positive association and no association |
|--|---|---|--|
| General (undiagnosed) acute respiratory illness (ARill) (Table 3a) | Less competitive athletes Elevated neutrophil and WBC counts Low serum [Vitamin D] | Intensified training phase Competition Reduction in salivary flow rate Detection of IgE antibodies to aero-allergens | • Increased training load |
| Suspected acute respiratory tract infection (ARinf) (Table 3b) | Less competitive athletes Decreased training intensity Increased training monotony Endurance preparation phase No tapering phase Winter Exposure to high altitude International travel Previous respiratory infections Low serum [Vitamin-D] | Age Gender Household family exposure | Increased training load Increased strength and speed training Competition period |
| Confirmed acute respiratory tract infection (ARinf) (Table 3c) | Increased training intensity Lower salivary-[IgA] preseason and pre-training and decline across a training program Reduced tear-[SIgA] and secretion rate | Post-season salivary-[IgA] | |

Table 3. A summary of risk factors and biomarkers associated with ARill and ARinf (suspected and confirmed) for which there is strong evidence of a positive association, no association or both.

higher risk of ARill/ARinf included: low tear-SIgA concentration and low salivary-IgA concentrations. Whilst other possible risk factors for ARill/ARinf were identified in this review, conflicting evidence limits conclusions to be draw. Further research in these areas is therefore warranted.

What is already known?

- Acute respiratory illnesses (ARill), especially respiratory tract infections (ARinf), are the most common acute illnesses affecting athletes.
- ARill can result in time loss from training and competition.
- Individual studies have reported that strenuous exercise-induced immunosuppression, mental stress, nutritional restrictions, air travel, human crowding, housing with other athletes, low temperature with low humidity, and competition all potentially increase the risk for ARill.

What are the new findings?

- Increased training load, monotony, endurance training programs, lack of tapering, training during winter and at altitude, and international travel were reported to increase the risk of ARinf.
- It is important for clinicians working with athletes to consider vitamin D deficits, particularly those prone to repeated ARill/ARinf.
- Biomarkers for monitoring athletes at a higher risk of ARill/ARinf include low tear-[SIgA] and low salivary-[IgA].

Multiple choice questions (MCQs) – Risk factors SR

What is the most common competition and training risk factor associated with suspected acute respiratory tract infection in athletes?

- a) A change in training load
- b) The tapering period
- c) Increased strength and speed training
- d) International travel associated with the competition
- e) The competition period

Which methodology is considered the "gold standard" to diagnose acute respiratory infection?

- a) Self-reported symptoms with a validated questionnaire or checklist
- b) Symptoms reviewed by a physician but without clinical or laboratory evaluation
- c) Clinical diagnosis by a physician based on history and clinical examination
- d) Diagnosis by a physician and confirmed by laboratory investigation to identify a specific pathogen
- e) All the above

Which immune biomarker might be the best choice to monitor the risk of acute respiratory infection in athletes?

- a) Leucocyte concentration
- b) IgM concentration
- c) IL-6 production

d) Salivary IgA concentration

e) Anti-inflammatory biomarkers

Which season is associated with increased risk of a suspected infective respiratory infection in athletes?

- a) Winter
- b) Summer
- c) Autumn
- d) Spring
- e) No season was identified

Which nutritional factor is associated with an increased risk of acute respiratory infection in athletes?

- a) Low serum vitamin B-12 concentrations
- b) Low serum vitamin B-6 concentrations
- c) Low serum niacin concentrations
- d) Low carbohydrate intake
- e) Low serum vitamin D concentrations

Contributions: All authors contributed towards the generation of key search terms used to identify relevant articles for this systematic review. Furthermore, (LK, JG, KM & MG) were involved in the data extraction and secondary search for articles missed by the search strategy. KM and MG performed the categorisation of clinical diagnoses of ARinf, ARill and URS which were verified by WD and MS. Critical appraisal and OCEBM levels of evidence were performed by LK, JG & MG. All authors were involved in the analysis, interpretation and writing of the manuscript.

Funding: Authors received no funding for the design of the study, analysis, and interpretation of data and in writing of the manuscript

Competing interests: Authors declare no conflict of interest. Authors declare that the results of this study are presented clearly, honestly, and without fabrication, falsification, or inappropriate data manipulation.

Patient consent for publication: Not required.

Provenance and peer review: Not commissioned; externally peer reviewed.

REFERENCES

1. Gałązka-Franta A, Jura-Szołtys E, Smółka W, Gawlik R. Upper Respiratory Tract Diseases in Athletes in Different Sports Disciplines. *J Hum Kinet*. 2016;53(1):99--106. doi:10.1515/hukin-2016-0014

2. Nieman DC. Risk of upper respiratory tract infection in athletes: an epidemiologic and immunologic perspective. *J Athl Training*. 1997;32(4):344-349.

3. Valtonen M, Waris M, Vuorinen T, et al. Common cold in Team Finland during 2018 Winter Olympic Games (PyeongChang): epidemiology, diagnosis including molecular point-of-care testing (POCT) and treatment. *Brit J Sport Med.* 2019;53(17):1093. doi:10.1136/bjsports-2018-100487

4. Gleeson M. Mucosal Immunity and Respiratory Illness in Elite Athletes. *Int J Sports Med.* 2000;21(Supplement 1):33-43. doi:10.1055/s-2000-1450

5. Schwellnus M, Soligard T, Alonso J-M, et al. How much is too much? (Part 2) International Olympic Committee consensus statement on load in sport and risk of illness. *Brit J Sport Med.* 2016;50(17):1043. doi:10.1136/bjsports-2016-096572

6. Maron BJ, Udelson JE, Bonow RO, et al. Eligibility and disqualification recommendations for competitive athletes with cardiovascular abnormalities: task force 3: Hypertrophic cardiomyopathy, Arrhythmogenic right ventricular cardiomyopathy and other cardiomyopathies, and myocarditis A Scientific Statement From the American Heart Association and American College of Cardiology. *J Am Coll Cardiol.* 2015;66(21):2362-2371. doi:10.1016/j.jacc.2015.09.035

7. Heikkinen T, Järvinen A. The common cold. *Lancet*. 2003;361(9351):51-59. doi:10.1016/s0140-6736(03)12162-9

8. Monto AS. Epidemiology of viral respiratory infections. *Am J Medicine*. 2002;112(6):4-12. doi:10.1016/s0002-9343(01)01058-0

9. Gleeson M, Bishop N, Oliveira M, Tauler P. Influence of training load on upper respiratory tract infection incidence and antigen-stimulated cytokine production. *Scand J Med Sci Spor*. 2013;23(4):451-457. doi:10.1111/j.1600-0838.2011.01422.x

10. Spence L, Brown WJ, Pyne DB, et al. Incidence, Etiology, and Symptomatology of Upper Respiratory Illness in Elite Athletes. *Medicine Sci Sports Exerc*. 2007;39(4):577--586. doi:10.1249/mss.0b013e31802e851a

11. Nieman DC. Is infection risk linked to exercise workload? *Medicine Sci Sports Exerc*. 2000;32(7):S406-S411. doi:10.1097/00005768-200007001-00005

12. Svendsen IS, Taylor IM, Tønnessen E, Bahr R, Gleeson M. Training-related and competitionrelated risk factors for respiratory tract and gastrointestinal infections in elite cross-country skiers. *Brit J Sport Med.* 2016;50(13):809. doi:10.1136/bjsports-2015-095398

13. Hellard P, Avalos M, Guimaraes F, Toussaint J-F, Pyne DB. Training-Related Risk of Common Illnesses in Elite Swimmers over a 4-yr Period. *Medicine Sci Sports Exerc*. 2015;47(4):698-707. doi:10.1249/mss.000000000000461

14. Walsh NP, Gleeson M, Pyne DB, et al. Position statement. Part two: Maintaining immune health. *Exerc Immunol Rev.* 2011;17:64-103.

15. Simpson RJ, Kunz H, Agha N, Graff R. Chapter Fifteen Exercise and the Regulation of Immune Functions. *Prog Mol Biol Transl.* 2015;135:355-380. doi:10.1016/bs.pmbts.2015.08.001

16. Colbey C, Cox AJ, Pyne DB, Zhang P, Cripps AW, West NP. Upper Respiratory Symptoms, Gut Health and Mucosal Immunity in Athletes. *Sports Med.* 2018;48(Suppl 1):65-77. doi:10.1007/s40279-017-0846-4

17. Shaw DM, Merien F, Braakhuis A, Dulson D. T-cells and their cytokine production: The antiinflammatory and immunosuppressive effects of strenuous exercise. *Cytokine*. 2018;104:136-142. doi:10.1016/j.cyto.2017.10.001

18. Walsh NP. Recommendations to maintain immune health in athletes. *Eur J Sport Sci.* 2018;18(6):1-12. doi:10.1080/17461391.2018.1449895

19. Page MJ, McKenzie JE, Bossuyt PM, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. *Syst Rev.* 2021;10(1):89. doi:10.1186/s13643-021-01626-4

20. Kohl C, McIntosh EJ, Unger S, et al. Online tools supporting the conduct and reporting of systematic reviews and systematic maps: a case study on CADIMA and review of existing tools. *Environ Évid.* 2018;7(1):8. doi:10.1186/s13750-018-0115-5

21. Micah T, A B Paul. Upper respiratory tract infection. StatPearls [Internet]. 2020.

22. Mäkelä MJ, Puhakka T, Ruuskanen O, et al. Viruses and Bacteria in the Etiology of the Common Cold. *J Clin Microbiol*. 1998;36(2):539-542. doi:10.1128/jcm.36.2.539-542.1998

23. Bruce B. Viral upper respiratory infection. Integrative Medicine. 2018:170.

24. Carrat F, Vergu E, Ferguson NM, et al. Time Lines of Infection and Disease in Human Influenza: A Review of Volunteer Challenge Studies. *Am J Epidemiol*. 2008;167(7):775-785. doi:10.1093/aje/kwm375

25. Downs SH, Black N. The feasibility of creating a checklist for the assessment of the methodological quality both of randomised and non-randomised studies of health care interventions. *J Epidemiol Commun H.* 1998;52(6):377. doi:10.1136/jech.52.6.377

26. Phillips B, Ball C, Badenoch D, Straus S, Haynes B, Dawes M. Oxford Centre for Evidence-Based Medicine—Levels of Evidence. Centre for Evidence Based Medicine Web site. Published 2009. Accessed June 1, 2021.

27. Fahlman MM, Engels H-J. Mucosal IgA and URTI in American College Football Players & colon; A Year Longitudinal Study. *Medicine Sci Sports Exerc*. 2005;37(3):374-380. doi:10.1249/01.mss.0000155432.67020.88

28. Edouard P, Junge A, Sorg M, Timpka T, Branco P. Illnesses during 11 international athletics championships between 2009 and 2017: incidence, characteristics and sex-specific and discipline-specific differences. *Brit J Sport Med.* 2019;53(18):1174. doi:10.1136/bjsports-2018-100131

29. Matthews A, Pyne D, Saunders P, Fallon K, Fricker P. A self-reported questionnaire for quantifying illness symptoms in elite athletes. *Open Access J Sports Medicine*. 2010;Volume 1:15. doi:10.2147/oajsm.s7654

30. Fricker PA, Pyne DB, Saunders PU, Cox AJ, Gleeson M, Telford RD. Influence of Training Loads on Patterns of Illness in Elite Distance Runners. *Clin J Sport Med.* 2005;15(4):246-252. doi:10.1097/01.jsm.0000168075.66874.3e

31. Cox AJ, Pyne DB, Saunders PU, Callister R, Gleeson M. Cytokine Responses to Treadmill Running in Healthy and Illness-Prone Athletes. *Medicine Sci Sports Exerc*. 2007;39(11):1918-1926. doi:10.1249/mss.0b013e318149f2aa

32. Ihalainen JK, Schumann M, Häkkinen K, Mero AA. Mucosal immunity and upper respiratory tract symptoms in recreational endurance runners. *Appl Physiology Nutrition Metabolism*. 2016;41(1):96-102. doi:10.1139/apnm-2015-0242

33. Robson-Ansley P, Howatson G, Tallent J, et al. Prevalence of allergy and upper respiratory tract symptoms in runners of the London marathon. *Med Sci Sport Exer*. 2012;44(6):999--1004. doi:10.1249/mss.0b013e318243253d

34. Moreira A, Arsati F, Lima-Arsati YB de O, Simões AC, Araújo VC de. Monitoring stress tolerance and occurrences of upper respiratory illness in basketball players by means of psychometric tools and salivary biomarkers. *Stress Health*. 2010;27(3):e166--e172. doi:10.1002/smi.1354

35. Svendsen IS, Gleeson M, Haugen TA, Tønnessen E. Effect of an intense period of competition on race performance and self-reported illness in elite cross-country skiers. *Scand J Med Sci Spor*. 2015;25(6):846-853. doi:10.1111/sms.12452

36. Dressendorfer RH, Petersen SR, Lovshin SEM, Hannon JL, Lee SF, Bell GJ. Performance Enhancement With Maintenance of Resting Immune Status After Intensified Cycle Training. *Clin J Sport Med.* 2002;12(5):301. doi:10.1097/00042752-200209000-00008

37. Hanstock HG, Govus AD, Stenqvist TB, Melin AK, Sylta Ø, Torstveit MK. Influence of Immune and Nutritional Biomarkers on Illness Risk During Interval Training. *Int J Sport Physiol*. 2020;15(1):60-67. doi:10.1123/ijspp.2018-0527

38. Schwellnus M, Derman W, Jordaan E, et al. Factors associated with illness in athletes participating in the London 2012 Paralympic Games: a prospective cohort study involving 49,910 athlete-days. *Brit J Sport Med.* 2013;47(7):433--40. doi:10.1136/bjsports-2013-092371

39. Leicht CA, Bishop NC, Paulson TAW, Griggs KE, Goosey-Tolfrey VL. Salivary Immunoglobulin A and Upper Respiratory Symptoms During 5 Months of Training in Elite Tetraplegic Athletes. *Int J Sport Physiol.* 2012;7(3):210-217. doi:10.1123/ijspp.7.3.210

40. Furusawa K, Tajima F, Okawa H, Takahashi M, Ogata H. The incidence of post-race symptoms of upper respiratory tract infection in wheelchair marathon racers. *Spinal Cord.* 2007;45(7):513-517. doi:10.1038/sj.sc.3102028

41. Fowler PM, Duffield R, Lu D, Hickmans JA, Scott TJ. Effects of Long-Haul Transmeridian Travel on Subjective Jet-Lag and Self-Reported Sleep and Upper Respiratory Symptoms in Professional Rugby League Players. *Int J Sport Physiol.* 2016;11(7):876--884. doi:10.1123/ijspp.2015-0542

42. Thornton HR, Delaney JA, Duthie GM, et al. Predicting Self-Reported Illness for Professional Team-Sport Athletes. *Int J Sport Physiol*. 2016;11(4):543--550. doi:10.1123/ijspp.2015-0330

43. Cunniffe B, Griffiths H, Proctor W, Davies B, Baker JS, Jones KP. Mucosal Immunity and Illness Incidence in Elite Rugby Union Players across a Season. *Medicine Sci Sports Exerc*. 2011;43(3):388-397. doi:10.1249/mss.0b013e3181ef9d6b

44. Schwellnus M, Derman W, Page T, et al. Illness during the 2010 Super 14 Rugby Union tournament – a prospective study involving 22 676 player days. *Brit J Sport Med*. 2012;46(7):499--504. doi:10.1136/bjsports-2012-091046

45. Schwellnus MP, Derman WE, Jordaan E, et al. Elite athletes travelling to international destinations >5 time zone differences from their home country have a 2-3-fold increased risk of illness. *Brit J Sport Med*. 2012;46(11):816--21. doi:10.1136/bjsports-2012-091395

46. Tiernan C, Lyons M, Comyns T, Nevill AM, Warrington G. Salivary IgA as a Predictor of Upper Respiratory Tract Infections and Relationship to Training Load in Elite Rugby Union Players. *J Strength Cond Res.* 2020;34(3):782-790. doi:10.1519/jsc.000000000003019

47. Yamauchi R, Shimizu K, Kimura F, et al. Virus Activation and Immune Function During Intense Training in Rugby Football Players. *Int J Sports Med.* 2011;32(05):393-398. doi:10.1055/s-0031-1271674

48. Neville V, Gleeson M, Folland JP. Salivary IgA as a Risk Factor for Upper Respiratory Infections in Elite Professional Athletes. *Medicine Sci Sports Exerc*. 2008;40(7):1228-1236. doi:10.1249/mss.0b013e31816be9c3

49. Nakamura D, Akimoto T, Suzuki S, Kono I. Daily changes of salivary secretory immunoglobulin A and appearance of upper respiratory symptoms during physical training. *J Sports Medicine Phys Fit.* 2006;46(1):152-157.

50. Milanez VF, Ramos SP, Okuno NM, Boullosa DA, Nakamura FY. Evidence of a Non-Linear Dose-Response Relationship between Training Load and Stress Markers in Elite Female Futsal Players. *J Sports Sci Medicine*. 2014;13(1):22-29.

51. Gleeson M, McDonald WA, Pyne DB, et al. Salivary IgA levels and infection risk in elite swimmers. *Medicine Sci Sports Exerc.* 1999;31(1):67-73. doi:10.1097/00005768-199901000-00012

52. Gleeson M, Hall ST, McDonald WA, Flanagan AJ, Clancy RL. Salivary IgA subclasses and infection risk in elite swimmers. *Immunol Cell Biol.* 1999;77(4):351-355. doi:10.1046/j.1440-1711.1999.00839.x

53. Gleeson M, Pyne DB, Austin JP, et al. Epstein-Barr virus reactivation and upper-respiratory illness in elite swimmers. *Medicine Sci Sports Exerc*. 2002;34(3):411-417. doi:10.1097/00005768-200203000-00005

54. Rama L, Teixeira AM, Matos A, et al. Changes in natural killer cell subpopulations over a winter training season in elite swimmers. *Eur J Appl Physiol*. 2012;113(4):859--68. doi:10.1007/s00421-012-2490-x

55. Hellard P, Guimaraes F, Avalos M, Houel N, Hausswirth C, Toussaint JF. Modeling the Association between HR Variability and Illness in Elite Swimmers. *Medicine Sci Sports Exerc*. 2011;43(6):1063-1070. doi:10.1249/mss.0b013e318204de1c

56. Brisola GMP, Claus GM, Dutra YM, et al. Effects of Seasonal Training Load on Performance and Illness Symptoms in Water Polo. *J Strength Cond Res*. 2020;34(2):406-413. doi:10.1519/jsc.00000000003358

57. Novas AM, Rowbottom DG, Jenkins DG. Tennis, Incidence of URTI and Salivary IgA. *Int J Sports Med.* 2003;24(3):223--229. doi:10.1055/s-2003-39096

58. Novas A, Rowbottom D, Jenkins D. Total Daily Energy Expenditure and Incidence of Upper Respiratory Tract Infection Symptoms in Young Females. *Int J Sports Med.* 2002;23(7):465-470. doi:10.1055/s-2002-35075

59. He C-S, Handzlik M, Fraser WD, et al. Influence of vitamin D status on respiratory infection incidence and immune function during 4 months of winter training in endurance sport athletes. *Exerc Immunol Rev.* 2013;19:86-101.

60. Hausswirth C, Louis J, Aubry A, Bonnet G, Duffield R, Meur YL. Evidence of Disturbed Sleep and Increased Illness in Overreached Endurance Athletes. *Medicine Sci Sports Exerc*. 2014;46(5):1036-1045. doi:10.1249/mss.00000000000177

61. Zehsaz F, Farhangi N, Monfaredan A. Interleukin-6 G-174C gene polymorphism and susceptibility to upper respiratory tract infection among endurance athletes. *J Exerc Sci Fit.* 2014;12(1):15-19. doi:10.1016/j.jesf.2013.12.002

62. Reid VL, Gleeson M, Williams N, Clancy RL. Clinical investigation of athletes with persistent fatigue and/or recurrent infections. *Brit J Sport Med.* 2004;38(1):42--45. doi:10.1136/bjsm.2002.002634

63. Gleeson M, Pyne DB, Elkington LJ, et al. Developing a multi-component immune model for evaluating the risk of respiratory illness in athletes. *Exerc Immunol Rev.* 2017;23:52-64.

64. He C-S, Bishop NC, Handzlik MK, Muhamad AS, Gleeson M. Sex differences in upper respiratory symptoms prevalence and oral-respiratory mucosal immunity in endurance athletes. *Exerc Immunol Rev.* 2014;20:8-22.

65. Gleeson M, Bishop N, Oliveira M, McCauley T, Tauler P, Muhamad AS. Respiratory infection risk in athletes: association with antigen-stimulated IL-10 production and salivary IgA secretion. *Scand J Med Sci Spor*. 2012;22(3):410-417. doi:10.1111/j.1600-0838.2010.01272.x

66. Ikonen JN, Joro R, Uusitalo AL, et al. Effects of military training on plasma amino acid concentrations and their associations with overreaching. *Exp Biol Med.* 2020;245(12):1029-1038. doi:10.1177/1535370220923130

67. Scullion L, Baker D, Healey P, Edwards A, Love T, Black K. No Association between Vitamin D and Acute Respiratory Tract Infections Amongst Elite New Zealand Rugby Players and Rowers. *Int J Vitam Nutr Res.* 2018;88(1-2):8-15. doi:10.1024/0300-9831/a000285

68. Blume K, Körber N, Hoffmann D, Wolfarth B. Training Load, Immune Status, and Clinical Outcomes in Young Athletes: A Controlled, Prospective, Longitudinal Study. *Front Physiol.* 2018;9:120. doi:10.3389/fphys.2018.00120

69. Cox AJ, Gleeson M, Pyne DB, Callister R, Hopkins WG, Fricker PA. Clinical and Laboratory Evaluation of Upper Respiratory Symptoms in Elite Athletes. *Clin J Sport Med.* 2008;18(5):438-445. doi:10.1097/jsm.0b013e318181e501

70. Cox AJ, Gleeson M, Pyne DB, Callister R, Fricker PA, Scott RJ. Cytokine gene polymorphisms and risk for upper respiratory symptoms in highly-trained athletes. *Exerc Immunol Rev.* 2010;16:8-21.

71. Hanstock HG, Walsh NP, Edwards JP, et al. Tear Fluid SIgA as a Noninvasive Biomarker of Mucosal Immunity and Common Cold Risk. *Med Sci Sport Exer*. 2016;48(3):569--77. doi:10.1249/mss.00000000000000001

72. Schwellnus MP, Lichaba M, Derman WE. Respiratory tract symptoms in endurance athletes-a review of causes and consequences. *Current Allergy & Clinical Immunology*. 2010;23(2):52-57.

73. Foster C. Monitoring training in athletes with reference to overtraining syndrome. *Medicine Sci Sports Exerc.* 1998;30(7):1164--1168. doi:10.1097/00005768-199807000-00023

74. Ekblom B, Ekblom Ö, Malm C. Infectious episodes before and after a marathon race. *Scand J Med Sci Spor*. 2006;16(4):287-293. doi:10.1111/j.1600-0838.2005.00490.x

75. Trammell RA, Toth LA. Genetic susceptibility and resistance to influenza infection and disease in humans and mice. *Expert Rev Mol Diagn.* 2014;8(4):515-529. doi:10.1586/14737159.8.4.515

76. Nieman DC. Immunonutrition support for athletes. *Nutr Rev.* 2008;66(6):310-320. doi:10.1111/j.1753-4887.2008.00038.x

77. Pyne DB, Gleeson, McDonald, Clancy, Jr P, Fricker. Training Strategies to Maintain Immunocompetence in Athletes. *Int J Sports Med.* 2000;21(Supplement 1):51--60. doi:10.1055/s-2000-1452

78. Nieman D, Henson D, Sampson C, et al. The Acute Immune Response to Exhaustive Resistance Exercise. *Int J Sports Med.* 1995;16(05):322-328. doi:10.1055/s-2007-973013

79. Pedersen BK, Hoffman-Goetz L. Exercise and the Immune System: Regulation, Integration, and Adaptation. *Physiol Rev.* 2000;80(3):1055-1081. doi:10.1152/physrev.2000.80.3.1055

80. Pyne DB, Ker MS, Fricker PA, McDonald WA, Telfo RD, Weidemann MJ. Effects of an intensive 12-wk training program by elite swimmers on neutrophil oxidative activity. *Medicine Sci Sports Exerc.* 1995;27(4):536. doi:10.1249/00005768-199504000-00011

81. Gleeson M, McDonald WA, Pyne DB, et al. Immune Status and Respiratory Illness for Elite Swimmers During a 12-Week Training Cycle. *Int J Sports Med.* 2000;21(4):302-307. doi:10.1055/s-2000-313

82. Gleeson M. Mucosal immune responses and risk of respiratory illness in elite athletes. *Exerc Immunol Rev.* 2000;6:5-42.

83. Martens P-J, Gysemans C, Verstuyf A, Mathieu C. Vitamin D's Effect on Immune Function. *Nutrients*. 2020;12(5):1248. doi:10.3390/nu12051248

84. Bishop EL, Ismailova A, Dimeloe S, Hewison M, White JH. Vitamin D and Immune Regulation: Antibacterial, Antiviral, Anti-Inflammatory. *Jbmr Plus*. 2021;5(1). doi:10.1002/jbm4.10405

85. Hanstock HG, Ainegren M, Stenfors N. Exercise in Sub-zero Temperatures and Airway Health: Implications for Athletes With Special Focus on Heat-and-Moisture-Exchanging Breathing Devices. *Frontiers Sports Active Living*. 2020;2:34. doi:10.3389/fspor.2020.00034

86. Zhao H, Chen S, Yang F, et al. Alternation of nasopharyngeal microbiota in healthy youth is associated with environmental factors : implication for respiratory diseases. *Int J Environ Heal R*. 2020:1-11. doi:10.1080/09603123.2020.1810209

87. Keaney LC, Kilding AE, Merien F, Dulson DK. The impact of sport related stressors on immunity and illness risk in team-sport athletes. *J Sci Med Sport*. 2018;21(12):1192-1199. doi:10.1016/j.jsams.2018.05.014

88. Bermon S. Airway inflammation and upper respiratory tract infection in athletes: is there a link? *Exerc Immunol Rev.* 2007;13:6-14.

89. Martin N, Lindley MR, Hargadon B, Monteiro WR, Pavord ID. Airway Dysfunction and Inflammation in Pool- and Non–Pool-Based Elite Athletes. *Medicine Sci Sports Exerc*. 2012;44(8):1433-1439. doi:10.1249/mss.0b013e31824c823c

90. Gleeson M, Pyne DB. Respiratory inflammation and infections in high-performance athletes. *Immunol Cell Biol.* 2015;94(2):124--31. doi:10.1038/icb.2015.100

91. Kennedy MD, Davidson WJ, Wong LE, Traves SL, Leigh R, Eves ND. Airway inflammation, symptoms, and skiing. *Scand J Med Sci Spor*. 2016;26(7):835-842. doi:10.1111/sms.12527

92. Passàli D, Damiani V, Passàli GC, Passàli FM, Bellussi L. Alterations in rhinosinusal homeostasis in a sportive population: our experience with 106 athletes. *European Archives Oto-rhino-laryngology Head Neck*. 2004;261(9):502-506. doi:10.1007/s00405-003-0723-7

93. Baker C, Kimberlin D, Long S. Infectious diseases. Epidemiology and control. In: *Red Book Atlas of Pediatric Infectious Diseases*. ; 2009:131-138.

94. Bergman P, Lindh ÅU, Björkhem-Bergman L, Lindh JD. Vitamin D and Respiratory Tract Infections: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *Plos One*. 2013;8(6):e65835. doi:10.1371/journal.pone.0065835

95. Dias R, Baganha RJ, Cieslak F, et al. Immunological parameters and upper respiratory tract infections in team sports athletes. *Revista Brasileira De Medicina Do Esporte*. 2017;23(1):66-72.

96. Mazzeo RS. Altitude, exercise and immune function. Exerc Immunol Rev. 2005;11:6-16.

97. Tiollier E, Schmitt L, Burnat P, et al. Living high-training low altitude training: effects on mucosal immunity. *Eur J Appl Physiol*. 2005;94(3):298-304. doi:10.1007/s00421-005-1317-4

98. Bergeron M, Bahr R, Bärtsch P, et al. International Olympic Committee consensus statement on thermoregulatory and altitude challenges for high-level athletes. *Brit J Sport Med.* 2012;46(11):770-779. doi:10.1136/bjsports-2012-091296

99. Dünnwald T, Gatterer H, Faulhaber M, Arvandi M, Schobersberger W. Body Composition and Body Weight Changes at Different Altitude Levels: A Systematic Review and Meta-Analysis. *Front Physiol.* 2019;10:430. doi:10.3389/fphys.2019.00430

100. Ludvigsson JF, Neovius M, Hammarström L. Risk of Infections Among 2100 Individuals with IgA Deficiency: a Nationwide Cohort Study. *J Clin Immunol*. 2016;36(2):134-140. doi:10.1007/s10875-015-0230-9

101. Mantis NJ, Rol N, Corthésy B. Secretory IgA's complex roles in immunity and mucosal homeostasis in the gut. *Mucosal Immunol*. 2011;4(6):603-611. doi:10.1038/mi.2011.41

102. Brandtzaeg P. Secretory IgA: Designed for Anti-Microbial Defense. *Front Immunol*. 2013;4:222. doi:10.3389/fimmu.2013.00222

103. Gabriel H, Kindermann W. The Acute Immune Response to Exercise: What Does It Mean? *Int J Sports Med.* 1997;18(S 1):S28-S45. doi:10.1055/s-2007-972698

104. Walsh NP, Gleeson M, Shephard RJ, et al. Position statement. Part one: Immune function and exercise. *Exerc Immunol Rev.* 2011;17:6-63.

105. Gleeson M. Immune function in sport and exercise. *J Appl Physiol*. 2007;103(2):693-699. doi:10.1152/japplphysiol.00008.2007

106. Gleeson M, Pyne DB. Exercise effects on mucosal immunity. *Immunol Cell Biol*. 2000;78(5):536-544. doi:10.1111/j.1440-1711.2000.t01-8-.x

107. Pyne DB, Gleeson M. Effects of Intensive Exercise Training on Immunity in Athletes. *Int J Sports Med.* 1998;19(S 3):S183-S194. doi:10.1055/s-2007-971991

108. Bishop NC, Gleeson M. Acute and chronic effects of exercise on markers of mucosal immunity. *Front Biosci.* 2009;Volume(14):4444. doi:10.2741/3540

109. Greenland S, Mansournia MA, Altman DG. Sparse data bias: a problem hiding in plain sight. *Br. Med. J.* 2016, 352.

110. Smith BM, Evans CT, Kurichi JE, Weaver FM, Patel N, Burns SP. Acute Respiratory Tract Infection Visits of Veterans With Spinal Cord Injuries and Disorders: Rates, Trends, and Risk Factors. *J Spinal Cord Medicine*. 2007;30(4):355-361. doi:10.1080/10790268.2007.11753951

111. Chen H, Cohen P, Chen S. How Big is a Big Odds Ratio? Interpreting the Magnitudes of Odds Ratios in Epidemiological Studies. *Commun Statistics - Simul Comput.* 2010;39(4):860-864. doi:10.1080/03610911003650383

See Checklist

| Section and Topic | ltem # | Checklist item | Location where item is reported |
|-------------------------------|-----------|--|---|
| TITLE | П | | |
| Title | 1 | Identify the report as a systematic review. | 1 |
| ABSTRACT | П | | |
| Abstract | 2 | See the PRISMA 2020 for Abstract's checklist. | 3, see below for abstract checklist |
| INTRODUCTION | | | |
| Rationale | 3 | Describe the rationale for the review in the context of existing knowledge. | 4 |
| Objectives | 4 | Provide an explicit statement of the objective(s) or question(s) the review addresses. | 4 |
| METHODS | | · | |
| Eligibility criteria | 5 | Specify the inclusion and exclusion criteria for the review and how studies were grouped for the syntheses. | 5 |
| Information sources | 6 | Specify all databases, registers, websites, organisations, reference lists and other sources searched or consulted to identify studies. Specify the date when each source was last searched or consulted. | 5 |
| Search strategy | 7 | Present the full search strategies for all databases, registers and websites, including any filters and limits used. | 5-6, Online supplementary S1 for generic, below for full string for each database |
| Selection process | 8 | Specify the methods used to decide whether a study met the inclusion criteria of the review, including how many reviewers screened each record and each report retrieved, whether they worked independently, and if applicable, details of automation tools used in the process. | 5-6 |
| Data collection process | 9 | Specify the methods used to collect data from reports, including how many reviewers collected data from each report, whether they worked independently, any processes for obtaining or confirming data from study investigators, and if applicable, details of automation tools used in the process. | 5-7 |
| Data items | 10a | List and define all outcomes for which data were sought. Specify whether all results that were compatible with each outcome domain in each study were sought (e.g. for all measures, time points, analyses), and if not, the methods used to decide which results to collect. | 6-7, see below for further details |
| | 10b | List and define all other variables for which data were sought (e.g. participant and intervention characteristics, funding sources). Describe any assumptions made about any missing or unclear information. | 6-7, further details below |
| Study risk of bias assessment | 11 | Specify the methods used to assess risk of bias in the included studies, including details of the tool(s) used, how many reviewers assessed each study and whether they worked independently, and if applicable, details of automation tools used in the process. | 7-8 |
| Synthesis methods | 13a | Describe the processes used to decide which studies were eligible for each synthesis (e.g. tabulating the study intervention characteristics and comparing against the planned groups for each synthesis (item #5)). | 6-8 (no sub-group analysis was performed) |
| | 13b | Describe any methods required to prepare the data for presentation or synthesis, such as handling of missing summary statistics, or data conversions. | 5-8, see below for further information |
| | 13c | Describe any methods used to tabulate or visually display results of individual studies and syntheses. | 5-8 |
| | 13d | Describe any methods used to synthesize results and provide a rationale for the choice(s). If meta-analysis was performed, describe the | 6-8 (no meta- |

Online Supplementary File:

| Section and Topic | ltem # | Checklist item | Location where item is reported |
|-------------------------------|-----------|--|---|
| | | model(s), method(s) to identify the presence and extent of statistical heterogeneity, and software package(s) used. | analysis was performed) |
| | 13e | Describe any methods used to explore possible causes of heterogeneity among study results (e.g. subgroup analysis, meta-regression). | - |
| | 13f | Describe any sensitivity analyses conducted to assess robustness of the synthesized results. | (no subgroup analysis was performed) |
| Reporting bias assessment | 14 | Describe any methods used to assess risk of bias due to missing results in a synthesis (arising from reporting biases). | 7-8, see below for further details |
| Certainty assessment | 15 | Describe any methods used to assess certainty (or confidence) in the body of evidence for an outcome. | 7-8 |
| RESULTS | | | |
| Study selection | 16a | Describe the results of the search and selection process, from the number of records identified in the search to the number of studies included in the review, ideally using a flow diagram. | 8, Figure 1 |
| | 16b | Cite studies that might appear to meet the inclusion criteria, but which were excluded, and explain why they were excluded. | See below for details. |
| Study characteristics | 17 | Cite each included study and present its characteristics. | 8-9, Table 2, Tables 3a-3c, & online supplementary S3. |
| Risk of bias in studies | 18 | Present assessments of risk of bias for each included study. | 37, Online supplementary S4 |
| Results of individual studies | 19 | For all outcomes, present, for each study: (a) summary statistics for each group (where appropriate) and (b) an effect estimate and its precision (e.g. confidence/credible interval), ideally using structured tables or plots. | Table 2, Tables 3a-c, online supplementary S4 |
| Results of syntheses | 20a | For each synthesis, briefly summarise the characteristics and risk of bias among contributing studies. | 8, see below for further details |
| | 20b | Present results of all statistical syntheses conducted. If meta-analysis was done, present for each the summary estimate and its precision (e.g. confidence/credible interval) and measures of statistical heterogeneity. If comparing groups, describe the direction of the effect. | 8, no subgroup analysis was performed |
| | 20c | Present results of all investigations of possible causes of heterogeneity among study results. | 8-9 |
| | 20d | Present results of all sensitivity analyses conducted to assess the robustness of the synthesized results. | No sub-group analysis was performed |
| Reporting biases | 21 | Present assessments of risk of bias due to missing results (arising from reporting biases) for each synthesis assessed. | No subgroup analysis performed |
| Certainty of evidence | 22 | Present assessments of certainty (or confidence) in the body of evidence for each outcome assessed. | 8-9 |

| Section and Topic | ltem # | Checklist item | Location where item is reported |
|--|-----------|--|---------------------------------------|
| DISCUSSION | | | |
| Discussion | 23a | Provide a general interpretation of the results in the context of other evidence. | 9-14 |
| | 23b | Discuss any limitations of the evidence included in the review. | 13-14 |
| | 23c | Discuss any limitations of the review processes used. | 13-14 |
| | 23d | Discuss implications of the results for practice, policy, and future research. | 14-15 |
| OTHER INFORMA | TION | | |
| Registration and protocol | 24a | Provide registration information for the review, including register name and registration number, or state that the review was not registered. | 3 & 5 |
| | 24b | Indicate where the review protocol can be accessed, or state that a protocol was not prepared. | 5 |
| | 24c | Describe and explain any amendments to information provided at registration or in the protocol. | See below for details |
| Support | 25 | Describe sources of financial or non-financial support for the review, and the role of the funders or sponsors in the review. | 17 |
| Competing interests | 26 | Declare any competing interests of review authors. | 17 |
| Availability of data, code and other materials | 27 | Report which of the following are publicly available and where they can be found: template data collection forms; data extracted from included studies; data used for all analyses; analytic code; any other materials used in the review. | Online supplementary S1- 4 |

From: Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. BMJ 2021;372:n71. doi: 10.1136/bmj.n71

For more information, visit: <u>http://www.prisma-statement.org/</u>

PRISMA abstract checklist:

| Section and Topic | Item # | Checklist item | Reported (Yes/No) |
|-------------------------|-----------|---|---------------------------|
| TITLE | | | |
| Title | 1 | Identify the report as a systematic review. | Y |
| BACKGROUND | | | |
| Objectives | 2 | Provide an explicit statement of the main objective(s) or question(s) the review addresses. | Y |
| METHODS | | | |
| Eligibility criteria | 3 | Specify the inclusion and exclusion criteria for the review. | Y (some) |
| | | | See paper for the rest |
| Information sources | 4 | Specify the information sources (e.g. databases, registers) used to identify studies and the date when each was last searched. | Y |
| Risk of bias | 5 | Specify the methods used to assess risk of bias in the included studies. | No, within the paper |
| Synthesis of results | 6 | Specify the methods used to present and synthesise results. | Y (some) |
| RESULTS | | | |
| Included studies | 7 | Give the total number of included studies and participants and summarise relevant characteristics of studies. | Y |
| Synthesis of results | 8 | Present results for main outcomes, preferably indicating the number of included studies and participants for each. If meta-analysis was done, report the summary estimate and confidence/credible interval. If comparing groups, indicate the direction of the effect (i.e. which group is favoured). | Y (no meta-analyses done) |
| DISCUSSION | | | |
| Limitations of evidence | 9 | Provide a brief summary of the limitations of the evidence included in the review (e.g. study risk of bias, inconsistency and imprecision). | Y (some) |
| Interpretation | 10 | Provide a general interpretation of the results and important implications. | Y |
| OTHER | | | |
| Funding | 11 | Specify the primary source of funding for the review. | No, within the paper |
| Registration | 12 | Provide the register name and registration number. | Y |

7. Present the full search strategies for all databases, registers and websites, including any filters and limits used.

PubMed: (Rhinovirus OR Parainfluenza OR Adenovirus OR coronavirus OR "human metapneumovirus" OR enterovirus OR "respiratory syncytial virus" OR "bordetella pertussis" OR "Chlamydophila pneumoniae" OR "mycoplasma pneumoniae" OR Rhinitis OR influenza OR "common cold" OR flu OR sinusitis OR "rhino sinusitis" OR "acute pharyngitis" OR tonsillitis OR pharyngitis OR pneumonia OR bronchitis OR "lung disease" OR "Respiratory tract disease*" OR "Respiratory tract infection*" OR "respiratory system disease*" OR "upper respiratory tract disease*" OR "Lower respiratory tract influenzes*" OR "Viral disease*" OR tuberculosis) AND (athlete* OR sport* OR exercis*) AND (risk factor*) NoT (asthma) NoT (COPD OR "chronic obstructive pulmonary disease" OR animal* OR HIV OR "human immunodeficiency virus" OR AIDS OR "acquired immunodeficiency syndrome" OR post-operative) Filters: Journal Article, Humans, English, MEDLINE, from 1990-July 2020

EbscoHost: (Rhinovirus OR Parainfluenza OR Adenovirus OR coronavirus OR "human metapneumovirus" OR enterovirus OR "respiratory syncytial virus" OR "bordetella pertussis" OR "Chlamydophila pneumoniae" OR "mycoplasma pneumoniae" OR Rhinitis OR influenza OR "common cold" OR flu OR sinusitis OR "rhino sinusitis" OR "acute pharyngitis" OR tonsillitis OR pharyngitis OR pneumonia OR bronchitis OR "lung disease" OR "Respiratory tract disease*" OR "Respiratory system disease*" OR "nespiratory system disease*" OR "respiratory system disease*" OR "upper respiratory tract disease*" OR "Lower respiratory tract disease*" OR "Viral disease*" OR tuberculosis) AND (athlete* OR sport* OR exercis*) AND (risk factor*) NoT (asthma) NoT (COPD OR "chronic obstructive pulmonary disease" OR animal* OR HIV OR "human immunodeficiency virus" OR AIDS OR "acquired immunodeficiency syndrome" OR post-operative) Scholarly (Peer Reviewed) Journals; Published Date: 19920101-20201231; Document Type: Article; Language: English, Species: Human

Web of Science: TOPIC: (Rhinovirus OR Parainfluenza OR Adenovirus OR coronavirus OR "human metapneumovirus" OR enterovirus OR "respiratory syncytial virus" OR "bordetella pertussis" OR "Chlamydophila pneumoniae" OR "mycoplasma pneumoniae" OR Rhinitis OR influenza OR "common cold" OR flu OR sinusitis OR "rhino sinusitis" OR "acute pharyngitis" OR tonsillitis OR pharyngitis OR epiglotitis OR laryngitis OR pneumonia OR bronchitis OR "lung disease" OR "Respiratory tract disease*" OR "Respiratory tract disease*" OR "Respiratory tract disease*" OR "Lower respiratory tract infection*" OR "respiratory tract disease*" OR "Urial disease*" OR tuberculosis) AND (athlete* OR sport* OR exercis*) AND (risk factor*) NoT (asthma) NoT (COPD OR "chronic obstructive pulmonary disease" OR animal* OR HIV OR "human immunodeficiency virus" OR "acquired immunodeficiency syndrome" OR post-operative): Refined by DOCUMENT TYPES: (ARTICLE), LANGUAGES: (ENGLISH), SPECIES: (HUMANS) Time span: 1990-2020. Indexes: SCI-EXPANDED, SSCI, A&HCI, CPCI-S, CPCI-SSH, BKCI-SSH, ESCI, CCR-EXPANDED, IC.

10a. List and define all outcomes for which data were sought. Specify whether all results that were compatible with each outcome domain in each study were sought (e.g. for all measures, time points, analyses), and if not, the methods used to decide which results to collect.

All studies included in the review reported the overall domain of risk factors for ARill and ARinf (undiagnosed and diagnosed). The small number of studies assessing each risk factor or biomarker made it difficult to draw consensus conclusions for most risks. Furthermore, the differences in the methodologies for classifying respiratory illnesses/infections further impaired comparisons.

10b. List and define all other variables for which data were sought (e.g. participant and intervention characteristics, funding sources). Describe any assumptions made about any missing or unclear information.

All details of variables are reported in the paper. No assumptions were made, however risk association was determined based on the types of statistical tests performed and whether this took confounders into account or not as well as whether the statistical test was a multi-variable analaysis or not.

13b. Describe any methods required to prepare the data for presentation or synthesis, such as handling of missing summary statistics, or data conversions.

Not all papers performed an analysis to determine risk association, risk association and strength of association was determined based on a 4 level metric to classify the type and strength of an association between a risk factor and ARill and ARinf as follows: no association (0, 00 or 000), some association (+), good association (++) or strong association (++). For more details please review paper.

14. Describe any methods used to assess risk of bias due to missing results in a synthesis (arising from reporting biases).

The domains from the modified Downs and Black tool that assessed risk of bias were (yes, no or unable to determine):

If any of the results of the study were based on "data dredging", was this made clear? Were the statistical tests used to assess the main outcomes appropriate? Were the main outcome measures used accurate (valid and reliable)? Were losses of patients to follow-up taken into account?

These 4 questions were part of the quality assessment. It must be noted that this review is not on RCTs, so the bias is not as clear as in reviews of RCTs, and therefore this was not specifically taken into consideration when performing the synthesis. The overall quality of article was assessed as per guidelines (including this risk of bias, however was used as an overall measure).

16b. Cite studies that might appear to meet the inclusion criteria, but which were excluded, and explain why they were excluded.

We cite studies on asthma and allergy that appeared to meet the inclusion criteria but were excluded after IOC consensus subgroup 1 meeting which resolved that asthma and allergy were being covered by another IOC sub-group as such they should be removed from this study.

20a. For each synthesis, briefly summarise the characteristics and risk of bias among contributing studies.

As mentioned above, that this review is not on RCTs, so the bias is not as clear as in reviews of RCTs, and therefore this was not specifically taken into consideration when performing the synthesis. The overall quality of article was assessed as per guidelines (including this risk of bias, however as an overall measure). Therefore for each synthesis the bias was not reported, this was further validated as no studies were rated as "poor" with all 48 studies rated as either excellent or good.

21. Present assessments of risk of bias due to missing results (arising from reporting biases) for each synthesis assessed.

There was no meta-analysis performed for this study and no studies with missing data were included in the review, therefore no assessment of risk of bias due to missing results (arising from reporting biases are presented in this review.

24c. Describe and explain any amendments to information provided at registration or in the protocol.

The protocol was amended by the following:

-The search period was extended by 7 months from 2019 to July 2020, due to the COVID-19-related delay of the IOC consensus meeting.

-The exclusion criteria were revised to exclude studies that only included non-infective acute respiratory illnesses such as asthma and allergy.

Risk factor: Variable associated with an increased risk of disease or infection.¹¹⁰

Odds ratio: An odds ratio (OR) is another measure of association that quantifies the relationship between exposure with two categories and health outcome.¹¹¹

Risk: The probability or chance, as measured by the occurrence of new cases of disease in a defined population over a defined period. Risk relates to the number of newly observed cases.

Risk ratio/relative risk: A risk ratio (RR), also called relative risk, compared the risk of a health event (disease/illness, injury, risk factor, or death) among one group with the risk among another group.

Level of athlete performance: Studies were categorized according to the level of performance of the athletes participating in the study and included: elite/professional, amateur, trained/competitive, recreational or a combination thereof.

Pathological classification (main and subgroups) of acute respiratory illness (ARill) and infections (ARinf) by diagnostic method.

| Pathological c | lassification | Methods to diagnose ARill | Description | | | |
|--|---|---|--|--|--|--|
| Main group | Subgroup | | | | | |
| General (undiagnosed) acute respiratory illness (ARill) | | Self-reported symptoms of ARill only Self-reported symptoms combined with an algorithm at least partially validated for ARill Self-reported symptoms of an ARill reviewed by a physician, but without clinical or laboratory evaluation Clinical diagnosis of an ARill by a physician, based on history and clinical examination | General symptoms of an ARill where the pathology could not be attributed specifically to an infection ARill studies could include illnesses that are due to either infective or non- infective causes but were not specified in the study design | | | |
| Acute respiratory infection (ARinf) | Suspected acute respiratory tract infection (ARinf) | Self-reported symptoms combined with an algorithm that has been validated for ARinf Self-reported symptoms of an ARinf reviewed by a physician, but without clinical or laboratory evaluation Clinical diagnosis of an ARinf by a physician, based on history and clinical examination | General symptoms and/or physical signs suggestive of an ARinf, but where the specific pathology of an infection was not confirmed The validated questionnaires that were used included the Wisconsin Upper Respiratory Symptom Survey (WURSS-21®); the Jackson Cold Scale (JCS); or other questionnaires in which the severity of the symptoms were scored to provide a quantitative assessment (AIS Symptom log).²¹ | | | |
| | Confirmed acute respiratory tract infection (ARinf) | Clinical diagnosis of ARinf by a physician that was confirmed by laboratory investigation to identify a specific pathogen utilising polymerase chain reaction (PCR) testing on specimen(s), culture of an organism from specimen(s), or serology (e.g. rise in antibody titres) | • In some studies, the identified pathogen was associated with a viral outbreak in a sporting team. The incidence rates in these studies may not reflect the rates of ARinf in general studies monitoring for ARinf in athletes. | | | |

Study characteristics (sorted alphabetically by sport): Study design, sport, level of training (category), number of participants, age (years) and gender (\bigcirc , female; \eth , male - reported separately where specified) and period of study (duration).

| Study | Study design | Sport | Category | Participants | Age (years) | Period | |
|---------------------------------------|--|---|--------------------------|--------------|---|-------------------------------------|--|
| Fahlman ²⁷ | Prospective study Longitudinal cohort | American Football | Trained | 75 | ♂ 20.5 ± 1.5 | 12 months | |
| Edouard et al. ²⁸ | Prospective study | Athletics | Elite | 12594 | - | 11 competitions (59 days) | |
| Matthews et al. ²⁹ | Prospective study | Athletics (Endurance) | Elite | 12 | 31.8 ± 4.0 | 31 days | |
| Fricker et al. ³⁰ | Prospective study Longitudinal cohort | Athletics (Endurance) | Elite | 20 | ♂ 24.2 ± 3.1 | 4 months | |
| $Cox \ et \ al.^{31}$ | Prospective study | Athletics | Well trained | 18 | ♂ 31.2 ± 8.2 | 3 sessions | |
| <i>Ihalainen et al.</i> ³² | Prospective study | Athletics | Trained | 25 | ♂ 34.6 ± 1.3 | 12 weeks | |
| Ansley et al. ³³ | Prospective study | Athletics (Endurance) | Recreational | 201 | $37.4 \pm 9.6;$ 40.3 ± 10.9 | 1 day | |
| <i>Moreira et al.</i> ³⁴ | Prospective study Longitudinal cohort | Basketball | Elite | 15 | $3 19.0 \pm 0.6$ | 4 weeks | |
| Svendsen et al. ³⁵ | Retrospective study | Cross-country skiers | Elite | 39 | - | 8 seasons $(2007 - 2015)$ | |
| Dressendorfer et al. ³⁶ | Prospective study Longitudinal cohort | Cycling | Competitive | 9 | ♂ 24.7 ± 2.1 | 14 weeks | |
| Spence et al. ¹⁰ | Prospective study Longitudinal cohort | Cycling / Triathlon | Elite and Competitive | 63 | Elite: 22.5 ± 3.8 Recreational: 25.2 ± 3.6 | 4 months | |
| Hanstock et al. ³⁷ | Prospective study Randomised control trial | Cycling / Triathlon | Trained | 27 | ♂ 29.9 ± 9.1 | 4 weeks | |
| Schwellnus et al. ³⁸ | Prospective study | Paralympic athletes | Elite | 3565 | 30.9 ±9.2 (13 to 61) | 14 days | |
| Leicht et al. ³⁹ | Prospective study Longitudinal cohort | Paralympic athletes (tetraplegic) | Elite | 14 | 33 ± 5 | 4 months | |
| Furusawa et al. ⁴⁰ | Prospective study Longitudinal cohort | Paralympic athletes (Endurance) | Trained | 21 | 42.0 ± 1.7 | 6 weeks | |
| Fowler et al. ⁴¹ | Prospective study | Rugby League | Elite | 18 | ∂ 24.2 ± 3.3 | 10 days | |
| <i>Thornton et al.</i> ⁴² | Prospective study | Rugby League | Professional | 32 | 26.0 ± 4.8 | 29 weeks (14-18 competitions) | |
| <i>Cunniffe et al.</i> ⁴³ | Prospective study | Rugby Union | Elite | 31 | 326.8 ± 0.9 | 11 months | |
| Schwellnus et al.44 | Prospective study | Rugby Union | Elite | 259 | - | 16 weeks | |
| Schwellnus et al.45 | Prospective study | Rugby Union | Elite | 259 | - | 16 weeks | |
| Tiernan et al. ⁴⁶ | Prospective study | Rugby Union | Elite | 19 | ∂ 19.7 ± 1.1 | 10 weeks | |
| Yamauchi et al.47 | Prospective study | Rugby Union | Trained | 32 | ∂ 20.4 ± 1.4 | 7 weeks | |
| Neville et al. ⁴⁸ | Prospective study Longitudinal cohort | Sailors | Elite | 38 | ♂ 36 ± 7 | 18 months | |
| Nakamura et al. ⁴⁹ | Prospective study | Soccer | Well trained | 12 | ♂ 19 to 21 | 33 days | |
| Milanez et al. ⁵⁰ | Prospective study | Soccer (Futsal) | Elite and Competitive | 13 | ♀ 22.1 ± 4.2 | 5 weeks | |
| <i>Gleeson et al.</i> ⁵¹ | Prospective study Longitudinal cohort | Swimmers | Elite | 25 | 16 to 24 | 7 months | |
| Hellard et al. ¹³ | Prospective study Longitudinal cohort | Swimmers | Elite | 28 | 16 to 30 | 4 years | |
| <i>Gleeson et al.</i> ⁵² | Prospective study Longitudinal cohort | Swimmers | Elite | 25 | 16 to 24 | 7 months | |
| <i>Gleeson et al.</i> ⁵³ | Prospective study Longitudinal cohort | Swimmers | Elite | 14 | ♂ 21.4 ± 2.3 | 30 days | |
| Rama et al. ⁵⁴ | Prospective study Longitudinal cohort | Swimmers | Elite | 19 | ♂: 17.2 ± 1.8 ♀: 15,8 ± 0,8 | 13 weeks | |
| Hellard et al. ⁵⁵ | Prospective study Longitudinal cohort | Swimmers | Elite | 18 | 19 to 30 | 2 years | |
| Brisola et al. ⁵⁶ | Prospective study Longitudinal cohort | Swimmers (Water polo) | Elite | 25 | ♀ 15.7 ± 1.3 | 15 weeks | |
| Novas et al. ⁵⁷ | Prospective study Longitudinal cohort | Tennis | Elite | 17 | \bigcirc 14 to 21 | 12 weeks | |

| Novas et al. ⁵⁸ | Prospective study Longitudinal cohort | Tennis | Elite, Trained and Recreational | 31 | \bigcirc 16 ± 2 | 12 weeks | |
|-------------------------------------|--|---------------------------|---------------------------------------|--------------|---|------------------------------------|--|
| <i>He et al.</i> ⁵⁹ | Prospective study Longitudinal cohort | Triathlon | Recreational to Elite | 225 | ♂ 22 ± 3 | 4 months (winter) | |
| Hausswirth et al. ⁶⁰ | Prospective study Randomized control trial | Triathlon | Trained | 27 | 37±6 | 6 weeks | |
| Zehsaz et al. ⁶¹ | Retrospective study | Various (Endurance) | Elite | 100 | ♂ 24.0 ± 5.9 | 2 years | |
| <i>Reid et al.</i> ⁶² | Prospective study | Various (Endurance) | Elite and Competitive | 41 | 12 to 56 | 12 months URS (Clinical study) | |
| <i>Gleeson et al.</i> ⁶³ | Prospective study Longitudinal cohort | Various (Endurance) | Highly trained | 16 | ♂ 32.5 ± 8.1 | 9 months | |
| Gleeson et al. ⁹ | Prospective study Longitudinal cohort | Various (Endurance) | Trained | 75 | 22.5 ± 4.0 | 4 months | |
| <i>He et al.</i> ⁶⁴ | Prospective study Longitudinal cohort | Various (Endurance) | Recreational | 210 | 21 ± 3 | 16 weeks (winter) | |
| Gleeson et al. ⁶⁵ | Prospective study Longitudinal cohort | Various (Endurance) | Recreational to Elite | 80 | ♂ -22.5 ±4.0 | 4 months (winter) | |
| Ikonen et al. ⁶⁶ | Prospective study Longitudinal cohort | Various (Military) | Highly trained | 53 | ♂ 19.6 ± 0.3 | 8 weeks | |
| Scullion et al. ⁶⁷ | Prospective study Cross sectional | Various (Rugby/Rowing) | Elite | 53 | 22.9 ± 3.2 | 6 months | |
| Blume et al. ⁶⁸ | Prospective study Longitudinal cohort | Various (Youth) | Trained | 274 | 13.8 ± 1.5 | 4 years | |
| Cox et al. ⁶⁹ | Prospective study | Various | Elite | 70 | 19.3 ± 2.6 | Single session (Clinical study) | |
| Cox et al. ⁷⁰ | Retrospective study | Various | Elite | 170 | 25.4 ± 8.6 | 12 months URS | |
| Hanstock et al. ⁷¹ | Prospective study | Various | Recreational | 40 | $3: 22 \pm 4;$ | 3 weeks | |
| | Repeated measures | | | (sub-cohort: | $\begin{array}{c} \ \ \ \ \ \ \ \ \ \ \ \ \ $ | (winter) | |
| | crossover trial | | | n=13) | 3 sub-cohort: 24 ± 4 | | |

Modified Downs and Black²⁵ Quality assessment scores and 2009 OCEBM²⁶ classifications for studies included.

| | Downs and Black Question | | | | | | | | | | | | OCE BM | | | |
|---------------------------------------|--------------------------|---|---|---|---|---|---|---|---|----|----|----|-----------|-------|-----------|----------|
| Included studies | | | | | _ | | _ | | | | | | | Total | Quality | Study |
| | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | score | of study | level |
| Fahlman ²⁷ | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 12 | Excellent | 1b |
| Edouard et al. ²⁸ | 1 | 1 | 0 | 1 | 1 | 1 | 0 | 1 | 1 | 0 | 1 | 1 | 1 | 10 | Good | 2b |
| Matthews et al. ²⁹ | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 0 | 1 | 1 | 1 | 11 | Excellent | 1b |
| Fricker et al. ³⁰ | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 0 | 1 | 1 | 1 | 11 | Excellent | 1b |
| $Cox \ et \ al.^{31}$ | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 0 | 1 | 1 | 1 | 11 | Excellent | 1b |
| <i>Ihalainen et al.</i> ³² | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 12 | Excellent | 2b |
| Ansley et al. ³³ | 1 | 1 | 1 | 1 | 0 | 1 | 1 | 0 | 0 | 0 | 1 | 1 | 1 | 9 | Good | 3b |
| Moreira et al. ³⁴ | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 0 | 1 | 0 | 1 | 1 | 1 | 10 | Good | 3b |
| Svendsen et al. ³⁵ | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 0 | 1 | 0 | 1 | 1 | 1 | 10 | Good | 2b |
| Dressendorfer et al. ³⁶ | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 0 | 1 | 1 | 1 | 11 | Excellent | 3b |
| Spence et al. ¹⁰ | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 12 | Excellent | 1b |
| Hanstock et al. ³⁷ | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 0 | 1 | 1 | 1 | 11 | Excellent | 1b |
| Schwellnus et al. ³⁸ | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 1 | 0 | 1 | 1 | 1 | 11 | Excellent | 3b |
| Leicht et al. ³⁹ | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 12 | Excellent | 1b |
| Furusawa et al. ⁴⁰ | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 0 | 1 | 1 | 1 | 11 | Excellent | 3b |
| Fowler et al. ⁴¹ | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 0 | 1 | 0 | 1 | 1 | 1 | 10 | Good | 1b |
| <i>Thornton et al.</i> ⁴² | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 0 | 1 | 1 | 1 | 11 | Excellent | 1b |
| Cunniffe et al. ⁴³ | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 0 | 1 | 1 | 1 | 11 | Excellent | 1b |
| Schwellnus et al.44 | 1 | 1 | 0 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 11 | Excellent | 1b |
| Schwellnus et al. ⁴⁵ | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 1 | Õ | 1 | 1 | 1 | 11 | Excellent | 3b |
| <i>Tiernan et al.</i> ⁴⁶ | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 0 | Õ | 1 | 1 | 1 | 10 | Good | 1b |
| Yamauchi et al. 47 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | Õ | 1 | 1 | 1 | 12 | Excellent | 1b |
| Neville et al. 48 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 0 | 1 | Õ | 1 | 1 | 1 | 10 | Good | 1b |
| Nakamura et al ⁴⁹ | 1 | 1 | 1 | 1 | 1 | 1 | Õ | 1 | 1 | Õ | 1 | 1 | 1 | 11 | Excellent | 3b |
| Milanez et al ⁵⁰ | 1 | 1 | 1 | 1 | 1 | 1 | ĩ | 0 | 1 | Õ | 1 | 1 | 1 | 11 | Excellent | 2b |
| Gleeson et al 51 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | Õ | 1 | Ő | 1 | 1 | 1 | 11 | Excellent | 1b |
| Hellard et al. ¹³ | 1 | 1 | 1 | 1 | 1 | 1 | 1 | Ő | 1 | Ő | 1 | 1 | 1 | 11 | Excellent | 16 1b |
| Gleeson et al 52 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | Ő | 1 | 1 | 1 | 12 | Excellent | 1b |
| Gleeson et al ⁵³ | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | Ő | 1 | 1 | 1 | 11 | Excellent | 16 1b |
| Rama et al 54 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | Õ | 1 | Ő | 1 | 1 | 1 | 11 | Excellent | 3h |
| Hellard et al 55 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | Ő | 1 | 1 | 1 | 12 | Excellent | 1b |
| Brisola et al 56 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | Ő | 1 | 1 | 1 | 11 | Excellent | 10 1h |
| Novas et al 57 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | Ő | 1 | 1 | 1 | 12 | Excellent | 1b |
| Novas et al 58 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | Ő | 1 | 1 | 1 | 11 | Excellent | 10 1b |
| He et al 59 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | Ő | 1 | 1 | 1 | 12 | Excellent | 1b |
| Hausswirth et al 60 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | Ő | 1 | 1 | 1 | 12 | Excellent | 2h |
| Zehsaz et al ⁶¹ | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | Ő | 1 | 1 | 1 | 12 | Excellent | 1b |
| $Gleeson et al^{63}$ | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | Ő | 1 | 1 | 1 | 11 | Excellent | 10 1b |
| He et al 64 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | Ő | 1 | 1 | 1 | 12 | Excellent | 10 1h |
| $Gleeson et al^{65}$ | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | Ő | 1 | 1 | 1 | 12 | Excellent | 1b |
| Ikonen et al 66 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | Ő | 1 | 1 | 1 | 11 | Excellent | 3h |
| Scullion et al 67 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | Ő | 1 | 1 | 1 | 12 | Excellent | 1b |
| $Gleeson et al^9$ | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 1 | 0 | 1 | 1 | 1 | 12 | Excellent | 10 1h |
| Rlume et al 68 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 12 | Excellent | 1b |
| Cor $et al^{69}$ | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 0 | 1 | 0 | 1 | 1 | 1 | 10 | Good | 2h |
| $Cor et al^{70}$ | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 1 | 1 | 12 | Excellent | 20 3h |
| Reid et al 62 | 1 | 1 | 1 | 1 | 0 | 1 | 0 | 0 | 1 | 0 | 1 | 1 | 1 | Q | Good | 1h |
| Hanstock et al ⁷¹ | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 0 | 1 | 1 | 1 | 11 | Excellent | 16 1h |

Note: 1, Is the hypothesis/aim/objective of the study clearly described?; 2, Are the main outcomes to be measured clearly described in the Introduction or Methods section?; 3, Are the characteristics of the patients included in the study clearly described; 4, Are the main findings of the study clearly described?; 5, Does the

study provide estimates of the random variability in the data for the main outcomes?; 6, Have the characteristics of patients lost to follow-up been described?; 7, Have actual probability values been reported (e.g. 0.035 rather than <0.05) for the main outcomes except where the probability value is less than 0.001?; 8, Were the subjects asked to participate in the study representative of the entire population from which they were recruited?; 9, Were those subjects who were prepared to participate representative of the entire population from which they were recruited?; 10, If any of the results of the study were based on "data dredging", was this made clear?; 11, Were the statistical tests used to assess the main outcomes appropriate?; 12, Were the main outcome measures used accurate (valid and reliable)?; 13, Were losses of patients to follow-up taken into account?.