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Article

Synthesis of Extended Uridine Phosphonates Derived from an Allosteric P2Y₂ Receptor Ligand

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Abstract: In this study we report the synthesis of C5/C6-fused uridine phosphonates that are structurally related to earlier reported allosteric $P2Y_2$ receptor ligands. A silyl-Hilbert-Johnson reaction of six quinazoline-2,4-(1*H*,3*H*)-dione-like base moieties with a suitable ribofuranosephosphonate afforded the desired analogues after full deprotection. In contrast to the parent 5-(4-fluoropheny)uridine phosphonate, the present extended-base uridine phosphonates essentially failed to modulate the $P2Y_2$ receptor.

Keywords: nucleoside phosphonates; extended uridine; P2Y2 receptor

1. Introduction

Amongst the many physiological functions of nucleotides, their capacity to mediate signal transmission via cell-surface receptors was discovered in 1929 [1]. Purinergic receptors are subdivided into two families: metabotropic P2Y receptors (P2YRs) and fast acting P2X ion channels [2]. The P2X receptors generally respond to the native agonist ATP, whereas P2YRs can be activated by ATP, UTP, UDP and UDP-glucose. The P2YRs family consist of eight members, which can be divided in two

subgroups based on receptor structure and second messenger system: P2Y_{1,2,4,6,11} and P2Y_{12,13,14} [3]. Ligand preferences of the eight P2YRs have been well established: P2Y₁ (ADP), P2Y₂ (UTP=ATP), P2Y₄ (UTP), P2Y₆ (UDP), P2Y₁₁ (ATP, NAD⁺), P2Y₁₂ (ADP), P2Y₁₃ (ADP), and P2Y₁₄ (UDP, UDP nucleotide sugars) [4].

The P2Y₂ receptor (P2Y₂R) is responsible for many physiological functions and therefore attracts considerable attention for developing therapeutics. For example, the P2Y₂R agonist diquafosol, which stimulates the secretion of chloride and water by epithelial cells, is approved in Japan for treatment of dry eye disease [5]. P2Y₂R activation protects cardiomyocytes from hypoxia *in vitro* and reduces post-ischemic myocardial damage *in vivo* [2]. Furthermore, the P2Y₂R is a promising target for the treatment of neurodegenerative diseases, including Alzheimer's disease [6]. Recently, ATP activation of the P2Y₂R was also shown to retard bone mineralization [1].

The development of selective P2Y₂R agonists and antagonists is a challenging task. Introduction of a 2'-amino and a 2-thio modification in UTP afforded analogue **1** (Figure 1) with enhanced potency and selectivity (versus P2Y₄R) [7]. Unfortunately, most P2Y₂R agonists contain a polyphosphate group, which is easily hydrolyzed by ecto-nucleotidases [4]. We recently reported a series of stable 5-substituted uridine phosphonate analogues, some of which (e.g., **2**, Figure 1) showed promising allosteric partial agonistic activity at P2Y₂R [8]. Given the interesting profile of the 5-aryluridine analogues we envisaged modifying this nucleobase to a more rigid structure as a way to potentially enhance affinity for the P2Y₂R. In this light we opted for a ring transformation from a 5-aryluracil to a quinazoline-2,4-dione motif, analogous to the biphenyl-naphthalene ring transformations often encountered in medicinal chemistry [9]. The first series of 5,6-annulated uridine analogues were based on the aromatic Topliss-scheme [6-H (**3a**), Cl (**3c**), OMe (**3e**) and Me (**3d**)] [10]. Additionally the 6-fluorine derivative **3b** was chosen based on the notable potency of parent compound **2** which bears a fluorine atom. Finally two nucleobases involving substitution of the benzene ring for a thiophene (**3f**) or a naphthalene core (**3g**) were included to gauge the effect of bioisosteric replacement or ring expansion of the nucleobase on the affinity for the P2Y₂R.

Figure 1. Structure of UTP, the selective $P2Y_2$ agonist 1, the allosteric partial agonist 2 and the target nucleoside phosphonates 3.



2. Results and Discussion

2.1. Synthesis

To gain access to the target C_5/C_6 -fused uridine phosphonates we first prepared the required "base" moieties **4a–g** by condensation of the appropriate *ortho*-amino aromatic carboxylic acids and urea at high temperature [11–13].

The synthesis of the appropriate glycosyl donor started from the known diol 6, which was obtained in two steps from the commercial 1,2:5,6-O-diisopropylidene- α -D-allofuranose (5) following a known procedure (Scheme 1) [14]. Cleavage of the resulting vicinal diol with sodium periodate gave the aldehyde [15], which was used without purification in the subsequent Wittig-type reaction. During formation of the diethylvinylphosphonate 7, C-4 may be prone to epimerization, which could be limited by performing the reaction at lower temperature. The yield for the conversion of 6 to 7 was significantly increased by using dichloromethane instead of ethanol for the periodate reaction. Hydrogenation with 10% Pd/C at atmospheric pressure allowed to simultaneous reduction of the double bond and removal of the benzyl group. One-pot deprotection of the isopropylidene group and acetylation of 8 afforded 9 (with an anomeric ratio of *ca*. 4:1 in favor of the α anomer), which was used in a silyl-Hilbert-Johnson reaction with the silylated benzo- or thieno[3,2-d]pyrimidine-2,4diones to give compounds **10a–g** [16]. When HMDS was used as the silvlating agent, careful removal of the silvlating agents prior to the addition of the base and trimethylsilvltrifluoromethane sulfonate was required. Hence, the use of BSTFA for in situ silvlation was preferred, despite the fact that it generally gave lower yields. Final deacetylation, followed by treatment with TMSBr/DCM afforded the desired nucleoside phosphonate analogues **3a**–g.

Scheme 1. Synthesis of the Target Extended Uridine-5'-methylenephosphonates 3a-g.



Reagents and conditions: (a) (i) NaIO₄, DCM, H₂O, rt; (ii) tetraethyl methylenediphosphonate, NaH, THF, 0 °C \rightarrow rt; (b) H₂, 10% Pd/C, rt; (c) (i) 60% AcOH, 85 °C, overnight; (ii) Ac₂O, pyridine, overnight; (d) (i) BSTFA, ACN, 65 °C, 2 h; (ii) TMSOTf, CH₃CN; (iii) NaOMe/MeOH, rt; (e) TMSBr, DCM, rt.

2.2. Pharmacological Evaluation

were determined measuring PLC-dependent phosphoinositide hydrolysis in 1321N1 human astrocytoma cells stably expressing the human $P2Y_2$ receptor (1321N1-P2Y₂ cells) [17,18] (Figure 2).

Compared to compound **2**, the current extended uridine phosphonates were found to stimulate the $P2Y_2$ receptor to a lesser extent, indicating that rigidifying the base moiety is unfavorable for binding to a presumed allosteric binding site. It can be concluded that structural flexibility of the 5-substituent of the earlier discovered series of allosteric P2Y₂ agonists is crucial for allosteric agonist activity.

Figure 2. Inositol phosphate production in 1321N1 cells stably expressing the human P2Y₂ receptor. After labeling with [³H]myo-inositol overnight, the cells were treated with agonists for 30 min at 37 °C in the presence of 10 mM LiCl, and inositol phosphate accumulation was quantified. The concentration of all phosphonate analagues was 30 μ M and of UTP was 3 μ M. The data are presented as the mean ± S.E.M. and are representative of results obtained in three separate experiments.



The possibility that the modified uridine 5'-phosphonate analogues could inhibit the effect of UTP by displacing it from the orthosteric site was examined by assessing their capacity at high concentrations to inhibit the effect of a near-maximal concentration of UTP (Figure 3). None of the phosphonate analogues gave an inhibitory effect on the action of UTP.

Figure 3. Effect of high concentrations of phosphonate analogues 3a-g on the P2Y₂ receptor-mediated activity of UTP. The effects of 30 μ M concentrations of the indicated phosphonates were tested in the presence of 100 μ M UTP in 1321N1-P2Y₂ cells.



3. Experimental

3.1. General Methods and Materials

All reagents were from standard commercial sources and of analytic grade. All reaction were performed under argon atmosphere except specified otherwise. Dry solvents were directly acquired from commercial sources. Precoated Merck Silica Gel F254 plates were used for TLC. Spots were examined under ultraviolet light at 254 nm and further visualized by sulfuric acid-anisaldehyde spray. Column chromatography was performed on silica gel (200–400 mm, 60 Å). ¹H, ¹³C, and ³¹P-NMR spectra were recorded in CDCl₃, DMSO-*d*₆, or D₂O on a Varian Mercury 300 MHz spectrometer (Varian, Palo Alto, CA, USA). Chemical shifts are given in parts per million (ppm δ), δ relative to residual solvent peak or TMS for ¹H and ¹³C and to external D₃PO₄ for ³¹P. Structural assignment was confirmed with COSY, HSQC and HMBC. Exact mass measurements were performed on a LCT Premier XE orthogonal time-of flight spectrometer with API-ES source (Waters, Zellik, Belgium) Waters LCT Premier XETM Time of flight (TOF) instrument. Samples were infused in a CH₃CN/H₂O (1:1) mixture at 10 mL/min.

Diethyl ((E)-2-((3aR, 5R, 6R, 6aR)-6-(benzyloxy)-2, 2-dimethyltetrahydrofuro[2, 3-d][1,3]dioxol-5-yl)vinyl)phosphonate (7): NaIO₄ (0.52 g, 2.22 mmol) was added to a stirred solution of compound**6**inCH₂Cl₂ (15 mL) and H₂O (10 mL) and the reaction mixture was stirred for 0.5 h at rt. The mixture wasthen filtrated and the filtrate was extracted with CH₂Cl₂. The combined organic layers were concentrated*in vacuo*to give the crude aldehyde, which was used in the next step without further purification. To astirred suspension of NaH (0.31 g, 7.81 mmol) in dry THF (10 mL) was added tetraethyl methylenebisphosphonate (1.41 g, 4.88 mmol) under argon. After 0.5 h the mixture was cooled in an ice bath anda solution of the crude aldehyde in THF (10 mL) was added. The reaction mixture was allowed towarm to rt and stirred overnight. The crude mixture was diluted with water (20 mL) and extracted withethyl acetate (3 × 20 mL). The combined organic layers were dried (Na₂SO₄) and concentrated *in vacuo*. Silica gel column chromatography (CH₂Cl₂/MeOH 97:3) afforded compound 7 as a colorless oil (0.77 g, 84.2%). ¹H-NMR (300 MHz, CDCl₃) δ : 1.28–1.35 (m, 6H, OCH₂CH₃), 1.36, 1.60 (2s, 6H, C(CH₃)₂), 3.53 (dd, J = 9.2, 4.2 Hz, 1H, H-3'), 3.98–4.16 (m, 4H, OCH₂CH₃), 4.55–4.66 (m, 3H, H-2',H-4', OCH₂Ph), 4.70–4.77 (m, 1H, OCH₂Ph), 5.77 (d, J = 3.8 Hz, 1H, H-1'), 5.96–6.12 (m, 1H, CH=CH), 6.74–6.90 (m, 1H, CH=CH), 7.28–7.42 (m, 5H, Ph). ¹³C-NMR (75 MHz, CDCl₃) δ : 16.47, 16.56 (OCH₂CH₃), 26.63, 26.93 (C(CH₃)₂), 61.97 (t, J = 4.7 Hz, OCH₂CH₃), 72.57 (OCH₂Ph), 77.68 (d, J = 9.4 Hz, C-4'), 78.04 (C-2'), 81.93 (d, J = 1.7 Hz, C-3'), 104.15 (C-1'), 113.48 (C(CH₃)₂), 118.45 (d, J = 186.0 Hz, C-6'), 128.18 , 128.30, 128.68, 137.19 (Ph), 148.00 (d, J = 6.1 Hz, C-5'). ³¹P-NMR (CDCl₃) δ : 17.49. HRMS (ESI): Calculated for C₂₀H₃₀O₇P [(M+H)⁺], 413.1724; found, 413.1754.

Diethyl (2-((3aR,5R,6R,6aR)-6-hydroxy-2,2-dimethyltetrahydrofuro[2,3-d][1,3]dioxol-5-yl)ethyl) phosphonate (8): To a solution of compound 7 (0.91 g, 2.22 mmol) in MeOH was added 10% Pd/C (0.5 g). The reaction mixture was stirred for 2 h under hydrogen atmosphere at rt and filtered. The filtrate was evaporated under vacuum and the residue purified by silica gel column chromatography (CH₂Cl₂/MeOH 97:3) to yield compound 8 as a colorless oil (0.58 g, 81.5%). ¹H-NMR (300 MHz, CDCl₃) δ : 1.29–1.35 (m, 6H, OCH₂CH₃), 1.36, 1.56 (2s, 6H, OC(CH₃)₂), 1.79–2.12 (m, 4H, H-5'a, H-5'b, H-6'a and H-6'b), 3.62 (ddd, *J* = 10.3, 8.7, 5.3 Hz, 1H, H-3'), 3.77 (ddd, *J* = 8.6, 6.7, 4.2 Hz, 1H, H-4'), 4.0–4.23 (m, 4H, OCH₂CH₃), 4.56 (dd, *J* = 5.0, 4.1 Hz, 1H, H-2'), 5.77 (d, *J* = 4.1 Hz, 1H, H-1); ¹³C-NMR (75 MHz, CDCl₃) δ : 16.43, 16.50 (OCH₂CH₃), 20.61 (d, *J* = 141.7 Hz, C-6'), 24.99 (d, *J* = 5.0 Hz, C-5'), 26.40, 26.52 (C(CH₃)₂), 61.67 (d, *J* = 6.63 Hz, OCH₂CH₃), 75.55 (C-3'), 78.76 (C-2'), 79.49 (d, *J* = 15.8 Hz, C-4'), 103.65 (C-1'), 112.52 (*C*(CH₃)₂); ³¹P-NMR (CDCl₃) δ : 31.66. HRMS (ESI): calculated for C₁₃H₂₆O₇P [(M+H)⁺], 325.1411; found, 325.1429.

(3R,4R,5R)-5-(2-(Diethoxyphosphoryl)ethyl)tetrahydrofuran-2,3,4-trivl triacetate (9): A solution of compound 8 (0.45 g, 1.38 mmol) and 60% AcOH (10 mL) was heated at 80 °C under argon for 16 h. When the starting material was consumed completely, the reaction mixture was evaporated *in vacuo* and co-evaporated twice with AcCN. The residue was dissolved in dry pyridine (12 mL) at 0 °C under argon followed by dropwise addition of acetic anhydride (1.56 mL, 16.52 mmol). The reaction mixture was slowly warmed to rt and stirred overnight. Then it was concentrated and co-evaporated with toluene. The residue was purified by silica gel column chromatography (CH₂Cl₂/MeOH 95:5) to give 9 (0.47 g, 83.2%) as a low melting solid (α : β anomeric ratio = 41:9). α anomer: ¹H-NMR (300 MHz. CDCl₃) δ: 1.22–1.30 (m, 6H, OCH₂CH₃), 1.63–1.94 (m, 4H, H-5'a, H-5'b, H-6'a and H-6'b), 1.99, 2.02, 2.04 (s, 9H, 3COCH₃), 3.96–4.08 (m, 4H, OCH₂CH₃), 4.08–4.17 (m, 1H, H-4'), 5.09 (dd, J = 7.0, 4.9 Hz, 1H, H-3'), 5.25 (d, J = 4.9 Hz, 1H, H-2'), 6.06 (s, 1H, H-1'). ¹³C-NMR (75 MHz, CDCl₃) δ : 16.49, 16.57 (OCH₂CH₃), 20.58, 20.60 (COCH₃), 21.80 (d, J = 143.3 Hz, C-6'), 21.15 (COCH₃), 27.27 (d, J = 4.4 Hz, C-5'), 61.73, 61.82 (OCH₂CH₃), 73.27 (C-3'), 74.65 (C-2'), 81.27 (d, J = 18.2 Hz, C-4'), 98.24 (C-1'), 169.19, 169.52, 169.87 (CO). ³¹P-NMR (CDCl₃) δ: 30.82. β anomer: ¹H-NMR (300 MHz, CDCl₃) δ : 1.32 (t, J = 7.0 Hz, 6H, OCH₂CH₃), 1.72–1.98 (m, 4H, H-5'a, H-5'b, H-6'a and H-6'b), 2.06, 2.10, 2.11 (s, 9H, 3 COCH₃), 4.01-4.17 (m, 4H, OCH₂CH₃), 4.20-4.29 (m, 1H, H-4'), 5.02 (dd, J = 6.8, 3.7 Hz, 1H, H-3'), 5.19 (dd, J = 6.8, 4.6 Hz, 1H, H-2'), 6.36 (d, J = 4.6 Hz, 1H, H-1'). ¹³C-NMR (75 MHz, CDCl₃) δ: 16.54, 16.62 (OCH₂CH₃), 20.43, 20.76, 21.17 (COCH₃), 21.81 (d, J = 141.83 Hz, C-6'), 26.72 (d, J = 4.4 Hz, C-5'), 61.81, 61.88 (OCH₂CH₃), 69.95 (C-2'), 72.02 (C-3'),

83.06 (d, J = 17.1 Hz, C-4'), 93.95 (C-1'), 169.46, 169.80, 170.17 (CH₃CO). ³¹P-NMR (CDCl₃) δ : 30.57. ESI-HRMS for $[C_{16}H_{27}O_{10}P+H]^+$ calcd, 411.1420; found, 411.1436. HRMS (ESI): calculated for $C_{16}H_{28}O_{10}P$ [(M+H)⁺], 411.1415; found 411.1436.

3.2. General Procedures for the Silyl-Hilbert-Johnson Reaction to Synthesize Products 10a to 10g

Method 1: To a suspension of the appropriate quinazoline-2,4-(1*H*,3*H*)-dione (1.5 eq.) in dry acetonitrile (50 eq.) was added BSTFA (4 eq.) under argon. The solution was heated at 65 °C for 2 h. After cooling, a solution of triacetate 9 (1 eq.) in dry acetonitrile (50 eq.) and trimethylsilyl triflate (1.5 eq.) were added. The solution was stirred for 2 h at rt. The reaction was quenched with saturated aqueous NaHCO₃ (50 mL for 1 mmol triacetate 9) and extracted with dichloromethane (50 mL for 1 mmol triacetate 9). The combined organic layer was washed with saturated aqueous NaHCO₃ (3 × 50 mL for 1 mmol triacetate 9), dried over anhydrous MgSO₄ and evaporated. The crude mixture was dissolved in MeOH (50 eq.) and 5.4 N sodium methoxide (8 eq.) in MeOH was added dropwise. After 1 h, the mixture was neutralized with acetic acid (10 eq.) and evaporated. Purification of the residue by silica-gel column chromatography (CH₂Cl₂/MeOH 97:3 \rightarrow 95:5) gave the pure product as a white foam.

Method 2: To a suspension of appropriate quinazoline-2,4-(1*H*,3*H*)-dione (1.5 eq.) in hexamethyldisilazane (50 eq.) was added trimethylsilyl chloride (0.7 eq.) and pyridine (10 eq.) under argon. The mixture was stirred at 130 °C overnight. The reaction mixture was evaporated to dryness under high vacuum. To the obtained residue a solution of triacetate **9** (1 eq.) in dry acetonitrile (50 eq.) was added under nitrogen followed by trimethylsilyl triflate (1.5 eq.). The solution was stirred for 2 h at rt. and the work up was similar as described in method 1.

Diethyl-(2-((2R,3S,4R,5R)-5-(2,4-dioxo-3,4-dihydroquinazolin-1(2H)-yl)-3,4-dihydroxy tetrahydrofuran-2yl)ethyl)phosphonate (**10a**): Following Method 1 reaction between **9** (0.47 g, 1.14 mmol) and **4a** (0.28 g, 1.71 mmol) yielded compound **10a** (0.22 g, 50%). ¹H-NMR (300 MHz, CDCl₃) δ : 1.27 (dt, J = 18.3, 7.1 Hz, 6H, OCH₂CH₃), 1.79–2.15 (m, 4H, H-5'a, H-5'b, H-6'a and H-6'b), 2.15–2.36 (m), 4.45 (br s), 4.97 (br s, 2OH, NH), 3.88–3.97 (m, 1H, H-4'), 3.97–4.15 (m, 4H, OCH₂CH₃), 4.59 (t, J = 6.7 Hz, 1H, H-3'), 4.80–4.89 (m, 1H, H-2'), 5.96 (d, J = 2.0 Hz, 1H, H-1'), 7.19–7.27 (m, 1H), 7.45 (d, J = 8.7 Hz, 1H), 7.59–7.67 (m, 1H), 8.14 (d, J = 7.7 Hz, 1H, Ph). ¹³C-NMR (75 MHz, CDCl₃) δ : 15.99–16.20 (m, OCH₂CH₃), 71.95 (C-3'), 72.50 (C-2'), 82.32 (d, J = 17.7 Hz, C-4'), 91.68 (C-1'), 114.27,115.92, 123.41, 128.37, 135.12 and 140.98 (Ph), 149.74 (C-2), 162.04 (C-4). ³¹P-NMR (CDCl₃) δ : 32.08. HRMS (ESI): calculated for C₁₈H₂₄N₂O₈P [(M–H)⁻], 427.1276; found, 427.1284.

Diethyl-(2-((2R,3S,4R,5R)-5-(6-fluoro-2,4-dioxo-3,4-dihydroquinazolin-1(2H)-yl)-3,4-dihydroxytetrahydrofuran-2-yl)ethyl)phosphonate (**10b**): Following Method 2 reaction between **9** (0.34 g, 0.84 mmol) and **4b** (0.18 g, 1.01 mmol) yielded compound **10b** (133.3 mg, 61.3%). ¹H-NMR (300 MHz, CDCl₃) δ : 1.29 (dt, J = 17.9, 7.1 Hz, 6H, OCH₂CH₃), 1.80–2.15 (m, 4H, H-5'a, H-5'b, H-6'a and H-6'b), 3.89–3.98 (m, 1H, H-4'), 3.98–4.16 (m, 4H, OCH₂CH₃), 4.29, 5.01, 10.31 (br s, 2OH, NH), 4.60 (t, J = 6.2 Hz, 1H, H-3'), 4.83 (dd, J = 6.2, 2.3 Hz, 1H, H-2'), 5.90 (d, J = 2.6 Hz, 1H, H-1'), 7.33–7.42 (m, 1H, Ph), 7.43–7.50 (m, 1H, Ph), 7.78–7.85 (m,1H, Ph). ¹³C-NMR (75 MHz, CDCl₃) δ : 16.57 (d, J = 5.53 Hz, OCH₂CH₃), 21.14 (d, J = 140.1 Hz, C-6'), 25.52 (t, J = 5.3 Hz, C-5'), 62.14 (dd, J = 13.82, 6.6 Hz, OCH₂CH₃), 72.36 (C-3'), 73.01 (C-2'), 82.87 (d, J = 17.1 Hz, C-4'), 92.40 (C-1'), 114.10–114.57 (m, Ph), 116.86 (d, J = 7.2 Hz, Ph), 117.68–117.98 (m, Ph), 122.86–123.38 (m, Ph), 137.76 (Ph), 149.83 (C-2), 156.99–160.41 (m, Ph), 161.39 (C-4). ³¹P-NMR (CDCl₃) δ : 32.71. HRMS (ESI): Calculated for C₁₈H₂₃FN₂O₈P [(M–H)⁻], 445.1182; found, 445.1188.

Diethyl-(2-((2R,3S,4R,5R)-5-(6-chloro-2,4-dioxo-3,4-dihydroquinazolin-1(2H)-yl)-3,4-dihydroxytetra-hydrofuran-2-yl)ethyl)phosphonate (**10c**): The reaction between **9** (0.34 g, 0.84 mmol) and **4c** (0.25 g, 1.26 mmol) as described for method 1 to yield compound **10c** (0.18 g, 45.1%). ¹H-NMR (300 MHz, CDCl₃) δ :1.28 (dt, *J* = 16.69, 7.03 Hz, 6H, OCH₂CH₃), 1.79–2.14 (m, 4H, H-5'a, H-5'b, H-6'a and H-6'b), 3.92 (q, *J* = 5.8 Hz, 1H, H-4'), 3.97–4.15 (m, 4H, OCH₂CH₃), 4.55 (t, *J* = 6.7 Hz, 1H, H-3'), 4.85 (dd, *J* = 6.15, 2.3 Hz, 1H, H-2'), 4.96, 10.49 (br s OH, NH), 5.89 (d, *J* = 2.6 Hz, 1H, H-1'), 7.40 (d, *J* = 9.4 Hz, 1H, Ph), 7.57 (dd, *J* = 9.1, 2.6 Hz, 1H, Ph), 8.04 (d, *J* = 2.3 Hz, 1H, Ph). ¹³C-NMR (75 MHz, CDCl₃) δ : 16.44–16.63 (m, OCH₂CH₃), 21.13 (d, *J* = 141.0 Hz, C-6'), 25.48 (d, *J* = 4.4 Hz, C-5'), 62.13 (dd, *J* = 11.6, 6.6 Hz, OCH₂CH₃), 72.42 (C-3'), 72.80 (C-2'), 82.84 (d, *J* = 17.1 Hz, C-4'), 92.25 (C-1'), 116.51, 117.55, 128.08, 129.76, 135.46 and 139.87 (Ph), 149.85 (C-2), 161.33 (C-4). ³¹P-NMR (CDCl₃) δ : 32.68. HRMS (ESI): calculated for C₁₈H₂₃ClN₂O₈P [(M–H)⁻], 461.0886; found, 461.0899.

Diethyl-(2-((2R,3S,4R,5R)-5-(6-methyl-2,4-dioxo-3,4-dihydroquinazolin-1(2H)-yl)-3,4-dihydroxytetrahydrofuran-2-yl)ethyl)phosphonate (10d): The reaction between 9 (0.25 g, 0.61 mmol) and 4d (0.16 g, 0.91 mmol) as described for method 1 to yield compound 10d (125.2 mg, 46.5%). ¹H-NMR (300 MHz, CDCl₃) δ :1.28 (dt, *J* = 19.0, 7.1 Hz, 6H, OCH₂CH₃), 1.81–2.18 (m, 4H, H-5'a, H-5'b, H-6'a and H-6'b), 2.37 (s, 3H, CH₃), 3.88–3.97 (m, 1H, H-4'), 3.97–4.16 (m, 4H, OCH₂CH₃), 4.28 (br s, 1H, OH), 4.56–4.68 (m, 1H, H-3'), 4.80–4.88 (m, 1H, H-2'), 4.98 (d, *J* = 5.0 Hz, 1H, OH), 5.94 (d, *J* = 2.3 Hz, 1H, H-1'), 7.34 (d, *J* = 8.8 Hz, 1H, Ph), 7.40–7.48 (m, 1H, Ph), 7.94 (s, 1H, Ph), 10.28 (s, 1H, NH). ¹³C-NMR (75 MHz, CDCl₃) δ : 15.78–16.15 (m, OCH₂CH₃), 20.51 (d, *J* = 141.0 Hz, C-6'), 19.95 (CH₃), 24.83 (d, *J* = 4.4 Hz, C-5'), 61.26–61.77 (m, OCH₂CH₃), 71.76 (C-3'), 72.55 (C-2'), 82.18 (d, *J* = 17.1 Hz, C-4'), 91.52 (C-1'), 113.98, 115.56, 127.99, 133.17, 135.95 and 138.69 (Ph), 149.56 (C-2), 161.96 (C-4). ³¹P-NMR (CDCl₃) δ : 32.84. HRMS (ESI): calculated for C₁₉H₂₆N₂O₈P [(M–H)⁻], 441.1432; found, 441.1427.

Diethyl-(2-((2R,3S,4R,5R)-5-(6-methoxy-2,4-dioxo-3,4-dihydroquinazolin-1(2H)-yl)-3,4-dihydroxytetrahydrofuran-2-yl)ethyl)phosphonate (**10e**): The reaction between **9** (0.25 g, 0.61 mmol) and **4e** (0.18 g, 0.91 mmol) as described for method 1 to yield compound **10e** (122.7 mg, 44.0%). ¹H-NMR (300 MHz, CDCl₃) δ :1.21–1.35 (m, 6H, OCH₂CH₃), 1.79–2.17 (m, 4H, H-5'a, H-5'b, H-6'a and H-6'b), 3.86 (s, 3H, OCH₃), 3.89–3.97 (m, 1H, H-4'), 3.98–4.15 (m, 4H, OCH₂CH₃), 4.63 (t, J = 6.7 Hz, 1H, H-3'), 4.85 (dd, J = 6.0, 2.5 Hz, 1H, H-2'), 5.04 (br s, OH), 5.92 (d, J = 2.3 Hz, 1H, H-1'), 7.20–7.25 (m, 1H, Ph), 7.39 (d, J = 9.7 Hz, 1H, Ph), 7.59 (d, J = 3.2 Hz, 1H, Ph), 10.27 (br s, 1H, NH). ¹³C-NMR (75 MHz, CDCl₃) δ : 16.42–16.64 (m, OCH₂CH₃), 21.09 (d, J = 141.0 Hz, C-6'), 25.42 (d, J = 4.4 Hz, C-5'), 56.02 (OCH₃), 62.08 (dd, J = 14.9, 6.6 Hz, OCH₂CH₃), 72.33 (C-3'), 73.08 (C-2'), 82.78 (d, J = 17.1 Hz, C-4'), 92.19 (C-1'), 109.63, 116.27, 117.12, 124.35 and 135.41 (Ph), 149.94 (C-2), 155.95 (Ph), 162.30 (C-4). ³¹P-NMR (CDCl₃) δ : 32.82. HRMS (ESI): calculated for C₁₉H₂₆N₂O₉P [(M–H)⁻], 457.1381; found, 457.1390.

Diethyl-(2-((2R,3S,4R,5R)-5-(2,4-dioxo-3,4-dihydrothieno[3,2-d]pyrimidin-1(2H)-yl)-3,4-dihydroxytetrahydrofuran-2-yl)ethyl)phosphonate (**10f**): The reaction between **9** (0.25 g, 0.61 mmol) and **4f** (0.15 g, 0.91 mmol) as described for method 1 to yield compound **10f** (198.8 mg, 75.2%). ¹H-NMR (300 MHz, CDCl₃) δ :1.20–1.35 (m, 6H, OCH₂CH₃), 1.79–2.15 (m, 4H, H-5'a, H-5'b, H-6'a and H-6'b), 3.92 (q, J = 6.1 Hz, 1H, H-4'), 3.97–4.14 (m, 4H, OCH₂CH₃), 4.50 (t, J = 6.3 Hz, 2H, H-3', OH), 4.80 (dd, J = 5.9, 2.9 Hz, 1H, H-2'), 4.95 (br s, 1H, OH), 5.81 (d, J = 3.2 Hz, 1H, H-1'), 7.13 (d, J = 5.3 Hz, 1H, H-6), 7.73 (d, J = 5.6 Hz, 1H, H-7), 10.33 (br s, 1H, NH). ¹³C-NMR (75 MHz, CDCl₃) δ : 16.43–16.62 (m, OCH₂CH₃), 21.15 (d, J = 141.0 Hz, C-6'), 25.54 (d, J = 4.4 Hz, C-5'), 62.12 (dd, J = 14.4, 6.6 Hz, OCH₂CH₃), 72.39 (C-3'), 72.71 (C-2'), 83.14 (d, J = 17.1 Hz, C-4'), 93.87 (C-1'), 114.38 (C-4a), 116.94 (C-6), 135.47 (C-7), 146.87 (C-7a), 151.09 (C-2), 158.53 (C-4). ³¹P-NMR (CDCl₃) δ : 32.63. HRMS (ESI): calculated for C₁₆H₂₂N₂O₈PS [(M-H)⁻], 433.0840; found, 433.0838.

Diethyl-(2-((2R,3S,4R,5R)-5-(2,4-dioxo-3,4-dihydrobenzo[g]quinazolin-1(2H)-yl)-3,4-dihydroxytetrahydrofuran-2-yl)ethyl)phosphonate (**10g**): The reaction between **9** (0.25 g, 0.61 mmol) and **4g** (0.19 g, 0.91 mmol) as described for method 1 to yield compound **10g** (125.0 mg, 43.1%). ¹H-NMR (300 MHz, CDCl₃) δ :1.22 (t, J = 7.0 Hz, 6H, OCH₂CH₃), 1.77–2.07 (m, 4H, H-5'a, H-5'b, H-6'a and H-6'b), 3.81 (d, J = 4.4 Hz, 1H, H-4'), 3.98 (quin, J = 7.1 Hz, 4H, 2 × OCH₂CH₃), 4.13–4.26 (m, 1H, H-3'), 4.65 (br s, 1H, H-2'), 5.05 (d, J = 6.2 Hz, 1H, OH), 5.21–5.32 (m, 1H, OH), 6.11 (d, J = 4.1 Hz, 1H, H-1'), 7.54 (t, J = 7.3 Hz, 1H, Ph), 7.66 (t, J = 7.3 Hz, 1H, Ph), 7.91 (s, 1H, Ph), 8.06 (d, J = 8.2 Hz, 1H, Ph), 8.14 (d, J = 8.2 Hz, 1H, Ph), 8.75 (s, 1H, Ph), 11.68 (br s, 1H, NH). ¹³C-NMR (75 MHz, CDCl₃) δ : 16.24, 16.31 (OCH₂CH₃), 20.74 (d, J = 140.1 Hz, C-6'), 25.63 (d, J = 3.9 Hz, C-5'), 60.92 (dd, J = 6.1, 2.8 Hz, OCH₂CH₃), 70.12 (C-2'), 72.01 (C-3'), 82.07 (d, J = 17.1 Hz, C-4'), 91.20 (C-1'), 111.46, 116.35, 125.85, 127.59, 128.34, 129.05, 129.36, 129.52, 135.99 and 136.04 (Ph), 149.80 (C-2), 161.65 (C-4). ³¹P-NMR (CDCl₃) δ : 32.08. HRMS (ESI): calculated for C₂₂H₂₆N₂O₈P [(M-H)⁻], 477.1432; found, 477.1419.

3.3. General Procedure for the Deprotection of 10a–g to 3a–g

The appropriate phosphonic ester **10** (1 eq.) was dissolved in dry CH_2Cl_2 (50 eq.) and treated with TMSBr (20 eq.) at 0 °C under argon. After stirring overnight at rt, the mixture was quenched with 7 N NH₃/MeOH (20 eq.) and evaporated. The residue was dissolved in H₂O (15 mL for 0.1 mmol phosphonic ester) and washed with CH_2Cl_2 (3 × 10 mL for 0.1 mmol phosphonic ester). The aqueous layer was evaporated and purified with RP high-performance liquid chromatography (HPLC, Phenomenex Luna C-18, H₂O/0.1% HCOOH in CH₃CN, 90:10 \rightarrow 0:100 in 23 min, flow 17.5 mL/min) to give the desired product after lyophilization of the appropriate fractions.

(2-((2R,3S,4R,5R)-5-(2,4-Dioxo-3,4-dihydroquinazolin-1(2H)-yl)-3,4-dihydroxytetrahydrofuran-2-yl)ethyl) phosphonic acid (**3a**): Deprotection of compound **10a** (0.13 g, 0.30 mmol) with TMSBr (0.91 g, 5.95 mmol) according to the general procedure yielded compound **3a** as a white power (32.2 mg, 29.1%). ¹H-NMR (300 MHz, D₂O) δ : 1.79–2.21 (m, 4H, H-5'a, H-5'b, H-6'a and H-6'b), 3.93–4.06 (m, 1H, H-4'), 4.34 (t, *J* = 6.9 Hz, 1H, H-3'), 4.86 (dd, *J* = 6.2, 4.4 Hz, 1H, H-2'), 6.09 (d, *J* = 3.8 Hz, 1H, H-1'), 7.35 (t, *J* = 7.5 Hz, 1H, Ph), 7.48 (d, *J* = 8.5 Hz, 1H, Ph), 7.71–7.81 (m, 1H, Ph), 8.00 (d, *J* = 7.9 Hz, 1H, Ph). ¹³C-NMR (75 MHz, D₂O) δ : 22.89 (d, *J* = 135.8 Hz, C-6'), 25.82 (d, *J* = 3.9 Hz, C-5'), 71.04 (C-2'), 72.22 (C-3'), 82.47 (d, *J* = 17.7 Hz, C-4'), 90.85 (C-1'), 115.11, 115.72, 124.53, 128.05, 136.31 and 140.37 (Ph) 150.95 (C-2), 164.12 (C-4). ³¹P-NMR (D₂O) δ : 29.02. HRMS (ESI): calculated for C₁₄H₁₆N₂O₈P [(M–H)⁻], 371.0650; found, 371.0651.

(2-((2R,3S,4R,5R)-5-(6-fluoro-2,4-dioxo-3,4-dihydroquinazolin-1(2H)-yl)-3,4-dihydroxytetrahydro-1(2H)-3,4-dihydroxytetrahydro-1(2H)-3,4-dihydroxytetrahydro-1(2H)-3,4-dihydroxytetrahydro-1(2H)-3,4-dihydroxytetrahydro-1(2H)-3,4-dihydroxytetrahydro-1(2H)-3,4-dihydroxytetrahydro-1(2H)-3,4-dihydroxytetrahydro-1(2H)-3,4-dihydroxytetrahydro-1(2H)-3,4-dihydroxytetrahydro-1(2H)-3,4-dihydroxytetrahydro-1(2H)-3,4-dihydroxytetrahydro-1(2H)-3,4-dihydroxytetrahydro-1(2H)-3,4-dihydroxytetrahydro-1(2H)-3,4-dihydroxytetrahydro-1(2H)-3,4-dihydroxytetrahydro-1(2H)-3,4-dihydroxytetrahydro-1(2H)-3,4-dihydroxytetrahydro-1(2H)-3,4-dihydroxytetrahydroxyt

furan-2-yl)ethyl)phosphonic acid (**3b**): The reaction between compound **10b** (286.4 mg, 0.54 mmol) and TMSBr (1.65 g, 10.79 mmol) as described above, yielded compound **3b** as a white powder (15.7 mg, 14.8%). ¹H-NMR (300 MHz, DMSO-*d*₆) δ :1.50–2.04 (m, 4H, H-5'a, H-5'b, H-6'a and H-6'b), 3.65–3.76 (m, 1H, H-4'), 4.04 (t, *J* = 6.7 Hz, 1H, H-3'), 4.49 (dd, *J* = 6.4, 4.7 Hz, 1H, H-2'), 5.98 (d, *J* = 4.4 Hz, 1H, H-1'), 6.66 (br s, 4H, 4OH), 7.47–7.55 (m, 1H, Ph), 7.55–7.64 (m, 1H, Ph), 7.71 (dd, *J* = 8.2, 2.9 Hz, 1H, Ph), 11.79 (br s, 1H, NH). ¹³C-NMR (75 MHz, DMSO-*d*₆) δ : 23.81 (d, *J* = 137.3 Hz, C-6'), 26.41 (C-5'), 69.88 (C-2'), 72.11 (C-3'), 82.49 (d, *J* = 17.7 Hz, C-4'), 90.77 (C-1''), 112.63–113.19 (m, C-5), 117.91 (d, *J* = 7.7 Hz, C-4a), 118.12 (d, *J* = 7.7 Hz, C-8), 122.31–122.85 (m, C-7), 136.87 (d, *J* = 1.7 Hz, C-8a), 149.73 (C-2), 156.06–159.37 (m, C-6), 160.85 (d, *J* = 2.2 Hz, C-4). ³¹P-NMR (CDCl₃) δ : 26.92. HRMS (ESI): calculated for C₁₄H₁₅FN₂O₈P [(M–H)⁻], 389.0556; found, 389.0560.

(2-((2R,3S,4R,5R)-5-(6-chloro-2,4-dioxo-3,4-dihydroquinazolin-1(2H)-yl)-3,4-dihydroxytetrahydrofuran-2-yl)ethyl)phosphonic acid (**3c**): The reaction between compound **10c** (0.14 g, 0.31 mmol) and TMSBr (0.94 g, 6.11mmol) as described above, yielded compound **3c** as a white powder (49.0 mg, 39.5%). ¹H-NMR (300 MHz, D₂O) δ :1.80–2.17 (m, 4H, H-5'a, H-5'b, H-6'a and H-6'b), 3.87–3.98 (m, 1H, H-4'), 4.28 (t, *J* = 7.0 Hz, 1H, H-3'), 4.76 (d, *J* = 3.8 Hz, 1H, H-2'), 5.88 (d, *J* = 3.8 Hz, 1H, H-1'), 7.25 (d, *J* = 9.4 Hz, 1H, Ph), 7.50 (dd, *J* = 9.1, 2.3 Hz, 1H, Ph), 7.56–7.61 (m, 1H, Ph). ¹³C-NMR (75 MHz, D₂O) δ : 22.69 (d, *J* = 135.8 Hz, C-6'), 25.63 (d, *J* = 3.3 Hz, C-5'), 71.29 (C-2'), 72.21 (C-3'), 82.29 (d, *J* = 18.3 Hz, C-4'), 90.98 (C-1'), 116.62, 116.96, 126.91, 129.32, 135.69 and 138.75 (Ph), 150.20 (C-2), 162.11 (C-4). ³¹P-NMR (D₂O) δ : 30.04. HRMS (ESI): calculated for [C₁₄H₁₅ClN₂O₈P [(M–H)⁻], 405.0260; found, 405.0266.

(2-((2R, 3S, 4R, 5R)-3, 4-dihydroxy-5-(6-methyl-2, 4-dioxo-3, 4-dihydroquinazolin-1(2H)-yl)tetrahydrofuran-2-yl)ethyl)phosphonic acid (**3d**): The reaction between compound **10d** (0.11 g, 0.25 mmol) and TMSBr (0.77 g, 5.01mmol) as described above, yielded compound **3d** as a white powder (39.6 mg, 41.0%). ¹H-NMR (300 MHz, D₂O) δ :1.82–2.13 (m, 4H, H-5'a, H-5'b, H-6'a and H-6'b), 2.20 (s, 3H, CH₃), 3.86–3.97 (m, 1H, H-4'), 4.28 (t, *J* = 7.0 Hz, 1H, H-3'), 4.73 (m, 1H, H-2'), 5.86 (d, *J* = 3.2 Hz, 1H, H-1'), 7.12 (d, *J* = 8.5 Hz, 1H, Ph), 7.31–7.44 (m, 2H, Ph). ¹³C-NMR (75 MHz, D₂O) δ : 19.62 (CH₃), 22.76 (d, *J* = 133.5 Hz, C-6'), 25.66 (d, *J* = 2.8 Hz, C-5'), 71.32 (C-2'), 72.26 (C-3'), 82.13 (d, *J* = 17.7 Hz, C-4'), 90.82 (C-1'), 114.83, 114.96, 127.30, 134.51, 136.96 and 137.79 (Ph), 150.46 (C-2), 163.38 (C-4). ³¹P-NMR (D₂O) δ : 29.86. HRMS (ESI): calculated for C₁₅H₁₈N₂O₈P [(M–H)[–]], 385.0806; found, 385.0819. (2-((2R, 3S, 4R, 5R)-3, 4-dihydroxy-5-(6-methoxy-2, 4-dioxo-3, 4-dihydroquinazolin-1(2H)-yl)tetrahydrofuran-2-yl)ethyl)phosphonic acid (**3e**): The reaction between compound **10e** (0.11 g, 0.23 mmol) and TMSBr (0.70 g, 4.58 mmol) as described above, yielded compound **3e** as a white powder (48.9 mg, 53.1%). ¹H-NMR (300 MHz, D₂O) δ:1.78–2.18 (m, 4H, H-5'a, H-5'b, H-6'a and H-6'b), 3.86 (s, 3H, CH₃), 3.93–4.01 (m, 1H, H-4'), 4.32 (t, J = 6.8 Hz, H-3'), 4.84 (dd, J = 6.6, 4.2 Hz, 1H, H-2'), 6.04 (d, J = 4.0 Hz, 1H, H-1'), 7.29–7.35 (m, 1H, Ph), 7.37–7.43 (m, 2H, Ph). ¹³C-NMR (75 MHz, D₂O) δ: 22.99 (d, J = 126.1 Hz, C-6'), 25.51 (C-5'), 55.90 (CH₃), 70.98 (C-2'), 72.21 (C-3'), 82.46 (d, J = 18.2 Hz, C-4'), 90.83 (C-1'), 109.87, 116.65, 116.92, 123.80, 134.38 and 155.46 (Ph), 150.68 (C-2), 163.60 (C-4). ³¹P-NMR (D₂O) δ: 28.81. HRMS (ESI): calculated for C₁₅H₁₈N₂O₉P [(M–H)⁻], 401.0755; found, 401.0755.

hydrofuran-2-yl)ethyl)phosphonic acid (**3f**): The reaction between compound **10f** (0.18 g, 0.41 mmol) and TMSBr (1.24 g, 8.13 mmol) as described above, yielded compound **3f** as a white powder (75.6 mg, 49.2%). ¹H-NMR (300 MHz, DMSO-*d*₆) δ :1.48–2.04 (m, 4H, H-5'a, H-5'b, H-6'a and H-6'b), 3.67–3.77 (m, 1H, H-4'), 4.01 (t, *J* = 6.4 Hz, 1H, H-3'), 4.41 (dd, *J* = 6.4, 5.3 Hz, 1H, H-2'), 5.91 (d, *J* = 5.0 Hz, 1H, H-1'), 6.53 (br s, 2H, 2OH), 7.25 (d, *J* = 5.6 Hz, 1H, H-6), 8.12 (d, *J* = 5.3 Hz, 1H, H-7), 11.62 (s, 1H, NH). ¹³C-NMR (75 MHz, DMSO-*d*₆) δ : 23.78 (d, *J* = 135.8 Hz, C-6', 26.48 (C-5'), 70.41 (C-2'), 71.94 (C-3'), 83.00 (d, *J* = 17.7 Hz, C-4'), 90.78 (C-1'), 113.85 (C-4a), 118.02 (C-6), 135.77 (C-7), 145.17 (C-7a), 150.77 (C-2), 157.91 (C-4). ³¹P-NMR (DMSO-*d*₆) δ : 28.07. HRMS (ESI): calculated for C₁₂H₁₄N₂O₈PS [(M–H)⁻], 377.0214; found, 377.0197.

(2-((2R,3S,4R,5R)-5-(2,4-dioxo-3,4-dihydrobenzo[g]quinazolin-1(2H)-yl)-3,4-dihydroxytetrahydro-1(2H)-3,4-dihydroxytetrahydroxytetrahydro

furan-2-yl)ethyl)phosphonic acid (**3g**): The reaction between compound **10g** (0.32 g, 0.56 mmol) and TMSBr (1.73 g, 11.30 mmol) as described above. The working up as follows: After stirring overnight, the reaction mixture was evaporated and co-distilled with toluene. The residue was dissolved with 7 N NH₃/MeOH (10 mL) and stirred at room temperature for 3 h. The reaction mixture was concentrated and purified with RP high-performance liquid chromatography (HPLC, Phenomenex Luna C-18, H₂O/0.1% HCOOH in CH₃CN, 90:10 \rightarrow 0:100 in 23 min, flow 17.5 mL/min) to give the compound **3g** as a white power (50.0 mg, 21.0%). ¹H-NMR (300 MHz, DMSO-*d*₆) δ :1.52–2.14 (m, 4H, H-5'a, H-5'b, H-6'a and H-6'b), 3.75–3.85 (m, 1H, H-4'), 4.16 (t, *J* = 6.6 Hz, 1H, H-3'), 4.63 (dd, *J* = 6.4, 4.1 Hz, 1H, H-2'), 5.21 (br s, 2H, 2OH), 6.10 (d, *J* = 4.1 Hz, 1H, H-1'), 7.49–7.58 (m, 1H, Ph), 7.61–7.70 (m, 1H, Ph), 7.93 (s, 1H, Ph), 8.05 (d, *J* = 8.2 Hz, 1H, Ph), 8.13 (d, *J* = 8.2 Hz, 1H, Ph), 8.74 (s, 1H, Ph), 11.67 (s, 1H, NH). ¹³C-NMR (75 MHz, DMSO-*d*₆) δ : 2.354 (d, *J* = 137.1 Hz, C-6'), 26.14–26.32 (m, C-5'), 69.83 (C-2'), 71.82 (C-3'), 82.23 (d, *J* = 17.4 Hz, C-4'), 90.86 (C-1'), 111.25, 116.09, 125.56, 127.35, 128.06, 128.75, 129.11, 129.22, 135.70 and 135.74 (Ph), 149.54 (C-2), 161.38 (C-4). ³¹P-NMR (DMSO-*d*₆) δ : 26.39. HRMS (ESI): calculated for C₁₈H₁₈N₂O₉P [(M–H)⁻], 421.0806; found, 421.0795.

3.4. Procedures for Phospholipase C assay

Stable cell lines for study of the human (h) $P2Y_2$, were generated by retroviral expression of the receptor in 1321N1 human astrocytoma cells, which do not natively express P2YRs. Agonist-induced [³H]inositol phosphate production was measured in cells plated at 20,000 cells/well on 96-well plates

two days prior to assay. Sixteen hours before the assay, the inositol lipid pool of the cells was radiolabeled by incubation in 100 μ L of serum-free inositol-free Dulbecco's modified Eagle's medium, containing 1.0 μ Ci of [³H]myo-inositol. No changes of medium were made subsequent to the addition of [³H]inositol. On the day of the assay, cells were challenged with 25 μ L of a five-fold concentrated solution of receptor agonists in 100 mM Hepes (*N*-(2-hydroxyethyl)-piperazine-N"-2-ethanesulfonic acid), pH 7.3 in HBSS, containing 50 mM LiCl for 30 min at 37 °C. Incubations were terminated by aspiration of the drug-containing medium and addition of 90 μ L of ice-cold 50 mM formic acid. After 30 min, supernatants were neutralized with 30 μ L of 150 mM NH₄OH and applied to Dowex AG1-X8 anion exchange columns. Total inositol phosphates were conducted in a similar manner using a maximally efficatious dose of agonist with variable amount of antagonist.

4. Conclusions

Based on the earlier identification a 5-aryl-substituted uridine phosphonate analogue as a promising allosteric partial agonist of the $P2Y_2R$, we prepared a small collection of a novel C_5/C_6 -fused uridine phosphonates. Pharmacological evaluation revealed that the target compounds exhibit much weaker agonistic activity than the parent compound.

Author Contributions

L.S. synthesized the target phosphonates, which were conceptualized by M.D.P.R. M.O.B. and K.A.B. performed the pharmacological evaluation under the supervision of T.K.H. I.K. contributed to the characterization of the compounds. L.S. and S.V.C. wrote the manuscript. S.V.C. coordinated this study.

Conflicts of Interest

The authors declare no conflict of interest.

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Sample Availability: Samples of the compounds **3a–g** and **10a–g** are available from the authors.

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