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1 ***Freshwater Biology***

2 **Spatially variable hydrological and biological processes shape diverse post-flood aquatic**
3 **communities**

4
5 **Running head: Flood shape diverse aquatic communities**

6
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27
28 **Keywords**

29 floodplain; snowmelt recession; spatial heterogeneity; hydrological dynamics; biodiversity

30
31
32 **Abstract**

33 1. Diverse aquatic environments in floodplains support high biodiversity, including plankton,
34 benthos, nekton (fish), and amphibians. Variation in aquatic communities among waterbodies
35 should be explained not only by the spatial variation in the environment at low flow but also by
36 the hydrological dynamics and biological responses over flood and recession periods. However,

37 very few studies have examined the formation processes of floodplain aquatic communities over
38 flood periods.

39 2. In this study, we aimed to obtain a mechanistic understanding of the assembly of aquatic
40 communities on a temperate floodplain by conducting intensive fieldwork during and after
41 seasonal flooding. The study was conducted in the Butokamabetsu River watershed in northern
42 Hokkaido, where snowmelt causes annual spring floods. At peak flow, floodwater not only flow
43 through main stream river and the extant side channels, but also flow through some palaeo-side
44 channels, which are usually isolated from the mainstream river and harbour stagnant water. As
45 the floods recede, palaeo-side channels become isolated from the mainstream river and their
46 flow cease, but the timing of the flow cessation varied among palaeo-side channels. We
47 hypothesized that this spatial asynchrony in the timing of flow cessations shapes diverse
48 floodplain aquatic communities. Therefore, we conducted field surveys of four faunal groups
49 (plankton, benthos, nekton (fish), and amphibian) during and after the spring flood in 2019.

50 3. The post-flood aquatic communities varied spatially, according to the flow at peak flood, the
51 timing of flow cessation, and the flow at low flow. Plankton composition was influenced by the
52 water flow at peak flood, and they were more abundant in waterbodies that were never flushed
53 by floodwaters. Fish composition was also influenced by the water flow at peak flood; in
54 particular, salmonids were more abundant in waterbodies that were hydrologically connected to
55 the main stream at peak flood. The presence of amphibian eggs was influenced by the timing of
56 flow cessation; eggs were laid in waterbodies with stagnant water during breeding season. The
57 benthic composition was influenced by flow at low flow. After the flood had receded, these
58 different distribution patterns of the four biological groups led to gradual variation of aquatic
59 communities among waterbodies on a floodplain.

60 4. Overall, this study showed that hydrological dynamics during flood recession shapes the post-
61 flood aquatic communities, and the spatial variability in the hydrological dynamics and the
62 different responses of the four faunal groups support diverse aquatic communities on a
63 floodplain. Importance of the natural geomorphological complexities, where paleo-side
64 channels with wide range of hydrological connectivity to the river channel co-occur, as well as
65 the natural hydrological dynamics of snowmelt recession, where the flooding discharge
66 gradually decrease over certain period in spring for floodplain aquatic communities are
67 highlighted.

68

69 **1 INTRODUCTION**

70 Floodplains are one of the most dynamic landscape ecosystems. Many waterbodies on
71 floodplains are formed by fluvial processes such as channel migration and abandonment. Extant
72 and palaeo-side-channels, a representative waterbody type on floodplains, have diverse degrees

73 of hydrological connectivity to a river's main stream: some are permanently connected to the
74 river, some are connected to the river only at times of high flow, and others are never connected
75 to the river (Amoros & Roux, 1988). In many temperate floodplains, spring snowmelt causes
76 long-lasting floods that seasonally expand and gradually contract river channels, thus supporting
77 spatially and temporally dynamic aquatic habitats for various faunal groups, including plankton,
78 benthos, nekton (fish), and amphibians (Tockner, Malard & Ward, 2000).

79

80 Although an increasing number of studies have investigated floodplain ecosystems at low flow,
81 few have addressed the spatial and temporal dynamism of floodplain ecosystems during periods
82 of flood (Amoros & Bornette, 2002; Hauer et al., 2016). Studies conducted at low flow have
83 shown that the composition of aquatic communities on a floodplain varies among waterbodies
84 (Tockner et al., 2000; Amoros & Bornette, 2002), and this variation is often attributed to
85 differences in hydrological connectivity to the river. However, most of the floodplains are
86 isolated from river mainstream at low flow when most studies are conducted, and the concept of
87 the connectivity is not well defined. In fact, aquatic communities in floodplain waterbodies are
88 not only influenced by the environmental variation that is observed at low flow; they are also
89 strongly influenced by the hydrological and biological processes during flood and flood
90 recession periods (Thomaz, Bini & Bozelli., 2007; Larsen et al., 2019). For example, flow
91 pulses may differently modify the physical and chemical environment of each waterbody,
92 thereby altering the suitability of habitats for aquatic organisms over time and space (Ward,
93 Tockner & Schiemer, 1999; Tockner et al., 2000). Isolation of aquatic organisms from the river
94 limits their dispersal and migration and thus constrains their distributions (Ward et al., 1999;
95 Crook et al., 2020). Geomorphological and hydrological characteristics may also influence
96 breeding site selection by amphibious organisms such as frogs (Kupferberg, 1996) and aquatic
97 insects (Encalada & Peckarsky, 2006). Therefore, to gain a mechanistic understanding of aquatic
98 community assembly on a floodplain, it is necessary to closely examine the spatial and temporal
99 components of the hydrological dynamics over flood periods and investigate the respective
100 responses of diverse faunal groups.

101

102 Different responses of different faunal groups to the hydrological dynamics over flood periods
103 may drive the diversity of aquatic communities in floodplain waterbodies. In floodplain
104 waterbodies, we observe at least four biological groups, namely, plankton, benthos, nekton
105 (fish), and amphibians, whose responses to the hydrological dynamics should differ according to
106 their ecological characteristics. For example, limitations to dispersal and migration may be
107 critical for mobile organisms such as fish (Bolland et al., 2012; Crook et al., 2020). Flow pulses
108 during flood periods may have a large influence on plankton, which might be flushed

109 downstream by floodwaters (Van den Brink, Van Katwijk & Van der Velde, 1994), and on
110 benthos, through mobilization of substrate materials (Arscott, Tockner & Ward, 2005). A
111 growing number of studies have addressed the use of floodplain habitats by different taxonomic
112 or faunal groups (Van den Brink et al., 1994; Morand & Joly, 1995; Bolland et al., 2012;
113 Gallardo et al., 2014), but only a few studies have simultaneously examined the distributions of
114 the distinct faunal groups that inhabit a river-floodplain system (Tockner, Schiemer & Ward,
115 1998). However, given that hydrological dynamics influences each faunal group differently, by
116 focusing on a single faunal group, the critical environmental gradient that is required to support
117 diverse aquatic communities may be underestimated. Variation in responses to hydrological
118 dynamics among faunal groups is one factor that supports diverse aquatic communities on a
119 floodplain, and gradients that include a wider range of hydrological environments may support
120 more diverse aquatic communities.

121

122 In this study, we aimed to obtain a mechanistic understanding of the assembly of aquatic
123 communities on a floodplain that would enable us to predict the distribution of aquatic biota on
124 a floodplain from the hydrological dynamics. We hypothesized that spatial asynchrony in flow
125 cessations associated with side channel isolation from the main stream during flood recession
126 shapes diverse floodplain aquatic communities, which is driven by the differential responses of
127 various faunal groups to the hydrological dynamics. First, to examine the link between
128 hydrological dynamics and biological responses, we collected biological data at multiple times
129 during the flood recession period from four selected waterbodies with different hydrological
130 connectivity to the river over the flood recession period (hereafter, the flood-recession survey).
131 Second, we evaluated the residency of fish in palaeo-side channels by using an isoscape
132 approach, because some fish may be mobile whereas other faunal groups unlikely move
133 between sites. Finally, after the flood had receded, we surveyed aquatic communities in 26
134 waterbodies where we had recorded different hydrological patterns over the flood recession
135 period (hereafter, the post-flood survey). Considering the results of both surveys, we examined
136 how the hydrological dynamics of snowmelt recession shaped the post-flood aquatic community
137 and showed how a range of hydrological connectivity to the river supported co-existing diverse
138 aquatic communities on a floodplain.

139

140 **2 METHODS**

141 **2.1 Study system**

142 The study was conducted on the Butokamabetsu River floodplain, located in the Hokkaido
143 University Uryu Experimental Forest, northern Japan (44°24'N, 142°07'E) (Fig. 1). The
144 floodplain is covered predominantly by 1–2 m high broad-leaf bamboo (*Sasa senanensis*) and a

145 sporadic mix of deciduous trees such as alder (*Alnus japonica*) and Japanese white birch (*Betula*
146 *platyphylla*). Some wetlands and some areas of humic soils are distributed on the floodplain
147 (Shibata et al., 2004; Ogawa et al., 2006). The main stream of the Butokamabetsu River, which
148 is about 10 m wide under summertime low-flow conditions, has many extant and palaeo-side
149 channels. Extant side channels remain connected to the main stream even during periods of low
150 flow and the water remains flowing all year around. Palaeo-side channels are old side channels
151 that have become isolated from the main stream through the loss of their upstream connection
152 with the river as a result of fluvial sediment and debris accumulation and flood dynamics. Many
153 palaeo-side channels, including our study sites, harbour stagnant water during low flow (also
154 called oxbow lakes). Floodwater flows into some of the palaeo-side channels at high flow, and
155 the water keeps flowing through palaeo-side channels until the river water level drops to certain
156 levels for respective paleo-side channel depending on their geomorphology. Therefore, a
157 gradient of hydrological connectivity to the river exists among waterbodies on the floodplain:
158 extant side channels, seasonally connected palaeo-side channels, and permanently isolated
159 palaeo-side channels.

160

161 The study area receives mean annual precipitation of 1236 mm, largely as snow, and snow
162 cover usually lasts from late November to early May, with a maximum depth of ~3 m (Aoyama
163 et al., 2011). Snowmelt typically occurs from April to May, and the river discharge is elevated
164 over this period (Fig. 2). Amphibians, including the Ezo brown frog (*Rana pirica*) and the Ezo
165 salamander (*Hynobius retardatus*), breed and their larvae mature during the snowmelt period on
166 the floodplain. Fish fry of various species develop in the floodplain waterbodies. We conducted
167 a field study in 2019 from the peak of the snowmelt season through the flood recession to the
168 low-flow period (10 May to 17 June).

169

170 **2.2 Flood-recession survey**

171 We conducted a flood-recession survey to characterize the temporal dynamics of hydrology and
172 water chemistry, as well as the biological responses of the four faunal groups to the
173 environmental changes as floodwater receded. As focal study sites for the flood-recession
174 survey, we selected four representative waterbodies with different degrees of connectivity to the
175 mainstream river (Fig. 2): “No flow” sites A and B, a “Early” site C, and a “Late” site D, as
176 determined during the post-flood survey (see section 2.4). At these four focal sites and at one
177 mainstream site (site E), we recorded environmental data approximately every five days from 14
178 May through 17 June 2019. At each site, we measured pH and conductivity with a portable
179 pH/COND meter (D-74, HORIBA, Kyoto, Japan) and dissolved oxygen and water temperature
180 with a portable multimeter (HQ-30d, HACH, Loveland, Colorado, USA). We collected a 200-ml

181 water sample in prewashed polyethylene bottles for nutrient analysis. Water samples were
182 filtered through a GF/F filter within 24 h, and both the GF/F filter and the filtered water were
183 frozen for later chlorophyll-*a* analysis (as an indicator of phytoplankton density) and nutrient
184 analysis in the laboratory. Furthermore, 20 L of water was filtered through a 70- μ m-mesh
185 plankton net and preserved in Lugol solution for zooplankton analysis.

186

187 To examine the distributional shift of fish from the peak flooding season to the end of the flood,
188 we conducted fish surveys on 22 May and 12 June 2019. Fish were captured with a backpack
189 electrofishing unit (Model LR-24, Smith-Root Inc., Vancouver, Washington, USA) using a
190 pulsed direct current setting (300–400V, 30-45Hz, and 25% duty cycle). A crew of three or four
191 study participants sampled in an upstream direction. The entire area of small waterbodies or the
192 first 20–120 m of palaeo-side channels at the site longer than 120 m were sampled by the single-
193 pass method. All fish collected in the survey on 22 May were identified to species and released
194 back to the same site alive, but some individuals collected on 12 June were sacrificed for the
195 stable isotope analysis (described below). The fish catch per unit effort was calculated by
196 dividing the fish count by the total habitat area sampled.

197

198 To examine the timing of amphibian breeding, we recorded the presence/absence of amphibian
199 eggs by species on 15 and 29 May and 11 June 2019. This survey was conducted at all 26 sites,
200 including the four focal sites, where the post-flood survey was conducted (see section 2.4).

201

202 **2.3 Isoscape analysis of the stable carbon isotope ratio**

203 To infer the residency of fish within each palaeo-channel, we conducted a spatial analysis of
204 stable carbon isotopes (isoscape analysis: see Bowen, 2010) in fishes and benthic invertebrates,
205 which are the main prey of the fishes. We focused on the natural spatial variation of stable
206 carbon isotope ratios among the waterbodies, which in anoxic ponds reflects the contribution of
207 methane-derived carbon (Kohzu et al., 2004). The stable carbon isotope ratio of consumer
208 species reflect that of their diet (DeNiro & Epstein 1978). Therefore, fish carbon isotope ratios
209 should reflect the carbon isotope ratios of items that they had consumed some days before they
210 were captured; the duration of the prey signals reflected in the fish tissue can vary from a few
211 days to a few weeks, depending on the fish species and conditions and environmental factors
212 (Weidel et al., 2011). Therefore, the stable carbon isotope ratios of fish that have inhabited a
213 local waterbody for a few weeks before the sampling date should match those of their local prey
214 (i.e., benthic macroinvertebrates). In contrast, the stable carbon isotope ratios fish that have
215 moved between waterbodies with different environments should be more spatially homogeneous

216 than those of their prey items. Further, if only some fish have moved between waterbodies, then
217 stable carbon isotope ratios should exhibit large variation among individuals.

218

219 We collected fish and their potential prey items at the four focal sites at the end of the recession
220 period. We collected representative benthic invertebrates with dip nets on 17 June 2019. At each
221 site, we collected 7 to 13 individuals belonging to 12 taxa. We collected most fish samples
222 during the second fish survey on 12 June 2019 (see section 2.2). We collected two
223 supplementary fish samples from site B on 19 June 2020, together with benthic
224 macroinvertebrate specimens. We confirmed that the stable carbon and nitrogen isotope ratios
225 did not differ significantly between sampling years; thus, we pooled the data from 2019 and
226 2020 for the isoscape analysis. We collected 2 to 10 individuals of the dominant fish species at
227 each site for stable isotope analysis. All fish and benthic invertebrate samples were immediately
228 frozen and transported to Kyoto University for later analysis.

229

230 **2.4 Post-flood Survey**

231 To examine how the biological responses during the flood, observed in the flood-recession
232 survey, shaped the spatial distribution of the aquatic biota at low flow (post-flood), we
233 conducted a post-flood survey of waterbodies on 12–13 June 2019, after snowmelt had ceased.
234 A total of 26 waterbodies (including both palaeo- and extant side channels) with different
235 degrees of connectivity to the main stream in five spatial blocks along a 10-km segment of the
236 Butokamabetsu River were selected (Fig. 1). We confirmed that there was no significant bias in
237 the distribution of connectivity among the blocks (one-way analysis of variance [ANOVA]; $P =$
238 0.95). Before the post-flood survey, during and after the flood recession from early May to mid-
239 June, we visited each site every five days to record its connectivity to the river's main stream.
240 During these visits, we visually examined the surface water connection to the main stream at
241 both the upstream and downstream ends of each waterbody. When there were surface water
242 connection upstream, mainstream river water flew-in from the upstream end, flew through the
243 waterbodies, and flew-out from the downstream end. Therefore, in all cases when we observed
244 upstream connection in this study, we also observed downstream connection, and the
245 waterbodies were flushed by flow. In contrast, downstream connection did not make as much
246 influence on the physical environment of the entire waterbodies as the upstream connection.
247 Yet, we assumed the downstream connection could enable the immigration and emigration of
248 fishes. Based on these observations, we categorized the study sites into six connectivity
249 categories (Figs. 1, 2). "Isolated" sites were never connected to the river, even at peak discharge.
250 "No flow" sites were never flushed by floodwaters, because the upstream end of the palaeo-side
251 channel was always closed, but at flood peak they were hydrologically connected to the main

252 stream at their downstream end. At the "Early", "Middle", and "Late" sites, each waterbody was
253 flushed by flow at peak discharge and isolated from the main stream at low flow, but differed by
254 the timing of flow cessation. At "Early", "Middle", and "Late" sites, flow cessation occurred on
255 23 May, 28 May, and 6 June 2019, respectively. At "Flowing" sites, flow was continuous
256 throughout the study period. Among the 26 sites, 2 were categorized as "Isolated", 7 as "No
257 flow", 6 as "Early", 4 as "Middle", 3 as "Late", and 4 as "Flowing" sites.

258

259 In the post-flood survey, in addition to measuring the water chemistry, we surveyed four faunal
260 groups: plankton, benthos, nekton (fishes), and amphibians. Water chemistry measurements and
261 water, plankton, and fish sampling were conducted following the same methods as for the flood-
262 recession survey. Benthic invertebrates were sampled with core samplers (13 cm diameter; five
263 times) at most sites, but at 12 sites where the substrate materials were larger than the core size,
264 Surber net samplers (30 cm × 30 cm) were used, and two samples were collected at each site on
265 each sampling date and combined. Benthic samples were immediately sieved through a 0.5 mm
266 mesh and preserved in 99% ethanol for later sorting. Fish were sampled by the same method as
267 for the flood-recession survey, and the presence or absence of amphibian eggs or larvae was also
268 recorded during the post-flood survey.

269

270 **2.5 Laboratory analysis**

271 All analyses were conducted at the Center for Ecological Research, Kyoto University, Otsu,
272 Japan. Nitrate (NO_3^-) nitrogen was quantified by first reducing nitrate to nitrite using a cadmium
273 coil followed by diazotization with sulfanilamide and coupling with *N*-(1-naphthyl)
274 ethylenediamine dihydrochloride. Ammonium (NH_4^+) nitrogen was quantified by the
275 indophenol blue method. Soluble reactive phosphorous (SRP) was quantified by the
276 molybdenum blue method. A continuous flow auto-analyser (QuAatro 2-HR, BLTEC, Osaka,
277 Japan) was used for these analyses.

278

279 Chlorophyll-*a* was extracted with dimethylformamide, and the mean chlorophyll-*a* content per
280 unit of water was measured by spectro-fluorometry (RF-5300PC, Shimadzu, Kyoto, Japan).
281 Zooplankton individuals > 0.5 mm in size were counted at the sub-order level, and their density
282 per water volume was estimated. Benthic macroinvertebrates were identified to family, and the
283 density of each family per unit area was estimated.

284

285 Stable carbon and nitrogen isotope analyses were also conducted at the Center for Ecological
286 Research, Kyoto University. Benthic insect samples were freeze-dried, and their whole bodies
287 were individually ground prior to the analysis. For fish samples, a sample of muscle tissue was

288 dissected out, then freeze-dried and ground for the analysis. A subsample of fish muscle tissues
289 was immersed in chloroform/methanol (2:1) solution for 24 h and then rinsed with methanol to
290 remove lipids. Stable nitrogen isotope ratios of samples without lipid removal and stable carbon
291 isotope ratios of samples after lipid removal were used as the representative values of fish
292 muscle, as described by Sotiropoulos, Tonn & Wassenaar (2004). Stable carbon and nitrogen
293 isotopes were measured with an elemental analyzer/isotope ratio mass spectrometer (EA/IRMS;
294 DELTA-V plus, Thermo Fisher Scientific, United States).. Carbon ($\delta^{13}\text{C}$) and nitrogen ($\delta^{15}\text{N}$)
295 isotope ratios are expressed as the per mil (‰). $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ of standard samples and the
296 analytical methods described above were referenced from Tayasu et al. (2011).

297

298 **2.6 Data analysis**

299 To examine spatial and temporal variations in water chemistry, we conducted a principal
300 component analysis (PCA) of pH, conductivity, DO, NO_3^- , NH_4^+ and SRP data collected at all
301 sites in all seasons. To examine the spatial variation in water chemistry during the post-flood
302 survey, scores on the first PCA axis (PCA1) were compared with the connectivity of each site,
303 where spatial block was included as a random factor in a linear mixed model, by using the lmer
304 function in the lme4 package (Bates et al., 2021) and the anova function in the lmerTest package
305 (Kuznetsova et al. 2020) in the R software environment (R Core Team, 2018). To examine
306 differences in the temporal variation in water chemistry at among the four focal sites with
307 different degrees of connectivity to the main stream, we compared the PCA1 score of the focal
308 sites with the Julian date by one-way ANOVA, including sampling site as a random factor.
309 Additionally, the same analyses were conducted for pH, conductivity, DO, NO_3^- , NH_4^+ , and
310 SRP, as well as chlorophyll *a* and zooplankton density. To compare the temporal shift in fish
311 species composition from the peak flood to the end of the flood season and the spatial variation
312 of fish species composition across waterbodies, we performed permutational multivariate
313 analysis of variance (PERMANOVA) using the adonis function in the vegan package (Oksanen
314 et al., 2020). Plankton and fish density data were log-transformed prior to all analysis.

315

316 Mean $\delta^{13}\text{C}$ of benthic macroinvertebrates and fish were estimated at each site. The relationship
317 between hydrological connectivity and the mean $\delta^{13}\text{C}$ of benthic macroinvertebrates was
318 examined by Spearman's rank correlation test. We conducted a regression analysis between the
319 mean $\delta^{13}\text{C}$ of benthic macroinvertebrates and the mean $\delta^{13}\text{C}$ of fish at five sites.

320

321 Using the post-flood survey data, we examined how the temporal hydrological dynamics
322 influenced each faunal group by comparing the density and diversity (taxonomic richness) of
323 each faunal group with the hydrological changes over time at each site. We fit five linear mixed

324 models to the data assuming five different distribution threshold (Fig. 2c), and compared their
325 goodness of fit based on their Akaike information criterion (AIC) values. All models included a
326 connectivity factor, described below, and habitat area as fixed effects, and block as a random
327 effect, to control for inherent longitudinal habitat heterogeneity. The first model assumed a
328 hydrological connection at flood peak (river connection on 15 May) influenced the biota, and
329 included as binary factors Isolated sites versus all other sites. The second model assumed that
330 the presence of flow at peak flood (on 15 May) influenced the biota, and included the binary
331 factors Isolated+No_flow sites versus other sites. The third and fourth models assumed that the
332 timing of the flow cessation at each site influenced the biota. The third model assumed that the
333 presence of flow on 23 May influenced the biota and included Isolated+No_flow+Early sites
334 versus Middle+Late+Flowing sites as binary factors. The fourth model assumed that the
335 presence of flow on 28 May influenced the biota, and included
336 Isolated+No_flow+Early+Middle” sites versus Late+Flowing sites as binary factor. The final
337 model assumed that the aquatic communities were structured primarily by the flow at the time
338 of sampling, and included Flowing sites versus all other sites as binary factors. We fit these five
339 models to the data on the total density of each respective faunal group and calculated the AIC
340 value of each model. We then selected the model with the lowest AIC value as the best model
341 for that faunal group. We analysed the diversity (taxonomic richness) of the four faunal groups
342 similarly.

343

344 We used multivariate techniques to examine the influence of hydrological connectivity on the
345 aquatic community, including the four faunal groups. Density data of all faunal groups were
346 standardized by using the decostand function in the vegan package (Oksanen et al., 2020) in R
347 prior to analysis. Then we analysed the data of all faunal groups by nonmetric multidimensional
348 scaling (NMDS) using Bray-Curtis dissimilarity index as pairwise beta-diversity values. We
349 applied PERMANOVA procedures to the β -diversity values to test for statistical differences in
350 the biota along the connectivity gradient. To account for spatial patterns, we included “block” as
351 a random effect in this analysis. Finally, we applied a similarity percentage analysis (SIMPER)
352 to examine the contribution of each taxon to the differences in the whole biota. The contribution
353 of each faunal group was inferred by summing the contributions of all taxa belonging to each
354 faunal group.

355

356 **3 RESULTS**

357 **3.1 Physical environment**

358 The all 26 studied waterbodies ranged in width from 0.7 m to 2.9 m, in length from 4.7 m to 160
359 m, and in depth from 0.08 m to 0.54 m at the time of the post-flood survey (i.e., after the

360 snowmelt flooding). Bottom sediment composition differed among the waterbodies depending
361 on connectivity to the main stream; more connected sites had a lower proportion of organic
362 sediments and contained larger rocks. Width, depth, and water velocity at the four focal sites
363 gradually decreased as the river discharge decreased over the study period.

364

365 **3.2 Water chemistry**

366 The multivariate analysis of the water chemistry data collected during flood-recession and post-
367 flood surveys revealed a link between the spatial and temporal water chemistry patterns. PCA1
368 explained 42% of the total variation in the chemistry data (Fig. 3) and was inferred to represent
369 temporal changes in habitat characteristics (ANOVA, $F_{1,29} = 70.3$, $P < 0.05$). Sites with different
370 degrees of connectivity were arranged along PCA1 such that more isolated sites exhibited lower
371 scores (ANOVA, $F_{5,13.5} = 4.6$, $P < 0.05$). Over the course of the flood-recession survey,
372 conductivity and NH_4^+ increased, and pH, DO, and NO_3^- decreased, but temporal changes in
373 SRP were not significant. The spatial data showed that pH and DO were lower at more isolated
374 sites, but connectivity was not significantly related to conductivity, NO_3^- , NH_4^+ , or SRP.

375

376 **3.3 Flood-recession survey**

377 Quantities of both phytoplankton and zooplankton increased as the flood receded (Fig. 3).
378 Chlorophyll-*a*, an indicator of phytoplankton density, significantly increased as the flood
379 receded (ANOVA, $F_{1,23} = 8.16$, $P < 0.05$). During the flood-recession zooplankton survey, a
380 total of 3407 copepods, 47 other crustaceans, and 105 mosquito larvae were counted. Total
381 zooplankton density also increased significantly as the flood receded (ANOVA, $F_{1,23} = 4.38$, $P <$
382 0.05).

383

384 In the fish survey, a total of 89 and 159 individuals were collected on 22 May and on 12 June
385 2019, respectively, at the four focal sites (Fig. 4). Young-of-the-year charr and trout were
386 recruited to the sampling sites over time; they were found only on the 12 June survey and only
387 at sites C and D, where flow had occurred at peak discharge but had ceased by 6 June. Overall,
388 fish species composition at the four sites did not differ significantly from before to after the
389 flood, and variation in fish species among sites persisted. The PERMANOVA analysis showed
390 that the spatial variation was greater than the temporal variation; the temporal change in fish
391 species composition was not statistically significant (Table S1).

392

393 No amphibian eggs were found on 15 May 2019. On 29 May, among the 26 sites, *Rana pirica*
394 eggs were found at 18 sites and *Hynobius retardatus* eggs were found at 14 sites. No additional
395 amphibian eggs were laid between 29 May and 11 June.

396

397 **3.4 Stable carbon isoscape**

398 $\delta^{13}\text{C}$ values of benthic macroinvertebrates and fish varied among the sites (Fig. 5; Fig. S1). $\delta^{13}\text{C}$
399 values of benthic macroinvertebrates were lower at sites with low connectivity (Spearman's rank
400 correlation test; $P < 0.05$, $\rho = 1$). At the No flow sites A and B, they ranged from -42‰ to -33‰
401 and from -43‰ to -33‰ , respectively, and at sites C (Middle) and D (Late), they ranged from $-$
402 35‰ to -31‰ and from -31‰ to -27‰ , respectively; at site E (on the main stream), they ranged
403 from -30‰ to -25‰ . $\delta^{13}\text{C}$ values of fish were positively correlated with those of benthic
404 macroinvertebrates at each site ($Y = 0.81x - 1.86$, $R^2 = 0.95$, $P < 0.01$). They ranged from -36‰
405 to -32‰ at site A, from -34‰ to -29‰ at site B, from -39‰ to -25‰ at site C, from -25‰ to
406 -24‰ at site D, and from -25‰ to -25‰ at the site E.

407

408 **3.5 Post-flood survey**

409 A total of 4626 copepods, 129 other crustaceans, and 350 mosquito larvae (Culicidae) were
410 collected during the post-flood survey (Fig. 6a). Copepods were more abundant at more isolated
411 sites. Other crustaceans and mosquito larvae were found only at "Isolated" and "No flow" sites.
412 The model comparison showed that flow on 15 May (peak flow) best explained zooplankton
413 density, and flow cessation on 23 May best explained zooplankton diversity (Table 1).

414

415 A total of 3789 individual benthic macroinvertebrates belonging to 37 families (32 aquatic
416 insect families and 5 others) were counted (Fig. 6b). Chironomidae were the most common. The
417 number of families was highest at "Flowing" sites, and 15 families were found only at
418 "Flowing" sites. The model comparison showed that flow on 15 May (peak flow) best explained
419 the density of benthic macroinvertebrates, and flow on 11 June (low flow, time at sampling) best
420 explained their diversity (Table 1).

421

422 A total of 559 individual fish belonging to nine species were found during the post-flood survey
423 (Fig. 4). *Salvelinus leucomaenis* (Salmonidae) occurred at the highest density, 50.2% of all
424 captured individuals, followed by stone loach (*Noemacheilus barbatulus*), Asiatic brook
425 lamprey (*Lethenteron reissneri*), and Masu salmon (*Oncorhynchus masou*). Fish density was
426 negatively associated with habitat area, possibly owing to higher competition (Nakano, 1995) or
427 lower capturing efficiency in large habitats. Importantly, the model comparison showed that
428 flow on 15 May (peak flow) best explained both fish density and diversity (Table 1).

429

430 Only two species of amphibians, *Rana pirica* and *Hynobius retardatus*, were observed during
431 the present study. The model comparison showed that the presence of amphibian eggs was best

432 explain by flow cessation on 23 May (and secondarily by flow on 28 May) (Table 1); thus, the
433 timing of flow cessation determined whether eggs of these amphibian species were present.

434

435 When all of the biological data, including those for the four faunal groups, were considered
436 altogether, the NMDS biplot showed clear differences in the biota along the gradient of
437 connectivity to the main stream (Fig. 7a; Fig. S2), and the PERMANOVA results also showed
438 significant variation of the biota in relation to the connectivity gradient (Table S2). The
439 SIMPER results showed different contributions of the four faunal groups to the variation of the
440 biota along the connectivity gradient (Fig. 7b; Table S3): The contribution of plankton to the
441 separation of Isolated and No flow sites from other sites was high, whereas the contribution of
442 amphibians to the separation of Early, Middle, and Late sites was high. Benthic
443 macroinvertebrates contributed greatly to the separation of Flowing sites from other sites,
444 whereas fish contributed equally to the variations among the sites except at the Flowing sites.

445

446 **4 DISCUSSION**

447 Environmental shifts occurred asynchronously among the waterbodies of the floodplain, and the
448 hydrological dynamics of each waterbody influenced the local community composition. The
449 flood-recession survey showed that the four aquatic faunal groups (plankton, benthos, fish, and
450 amphibians) responded dynamically but differently to the hydrological changes over the flood
451 recession period. The post-flood survey showed that the distribution patterns of the four faunal
452 groups differed according to the hydrological dynamics over the flood period, as was predicted
453 from the different biological responses of each faunal group to the hydrological conditions. Our
454 results indicate that the distinct responses of each biological group to conditions during the
455 spring flood were responsible for their respective post-flood distribution patterns in the
456 waterbodies. Our overall results support our hypothesis that the spatial asynchrony in channel
457 isolation from the main stream during flood recession results in diverse aquatic communities on
458 the floodplain, and that the different responses of the various faunal groups to the hydrological
459 dynamics drives that diversity.

460

461 The temporal dynamics of nutrient concentrations and phyto- and zooplankton over the flood
462 recession period showed that plankton bloomed in isolated waterbodies never flushed by
463 flowing floodwaters, and that the nutrients became depleted in such isolated waterbodies over
464 time. The lower nutrient concentrations (NO_3^- and SRP) in waterbodies with high densities of
465 phyto- and zooplankton indicate that plankton density was limited by factors other than nutrient
466 availability, and that in isolated waterbodies nutrients were depleted by plankton (Van Den
467 Brink et al., 1992). Plankton communities are known to be vulnerable to flow (Chaparro et al.,

468 2018; Van den Brink et al., 1994); thus, in many of the waterbodies, phyto- and zooplankton
469 were likely flushed by the floodwaters. However, the emergence of plankton in waterbodies that
470 had been flushed indicates that some plankton somehow remained in these waterbodies during
471 the flood period (Frisch, 2002) or that they colonized them after the flood (Jenkins & Boulton,
472 2003). Although in each waterbody the plankton density started to increase once the flow
473 stopped, and it is possible that later in the low-flow season (i.e., after the study period) their
474 density would become higher (Baranyi et al., 2002; Grosholz & Gallo, 2006; Schagerl et al.,
475 2009), their recovery was not fast enough to compensate for the impacts of the flow at flood
476 peak by the time of sampling.

477

478 In contrast to plankton, fish were more abundant and diverse in waterbodies that were flushed
479 by the peak flow. The spatial match between the stable carbon isotope ratios of fish fry and
480 benthos, together with the similarity of the fish species composition at each site between the
481 flood-recession and post-flood surveys, indicates that the residency of fish fry in each palaeo-
482 side channel was high. Salmonids spawn in habitats with gravel and pebble substrates
483 (Montgomery et al., 1999), which are common in the main stream and side channels of the
484 studied system. Furthermore, most of the fish in the floodplain waterbodies (palaeo-side
485 channels) were juveniles; adults were found only in the main stream. Fitzgerald et al. (2021)
486 estimated that the dominant species in the studied system, *Oncorhynchus masou* and *Salvelinus*
487 *leucomaenis*, hatch primarily in February and March. Therefore, it is assumed that salmonid fry
488 that hatched in extant side channels or the main stream of the river moved to the palaeo-side
489 channels during the flood to continue their development as described by Rosenfeld et al.,
490 (2008), whereas those hatched in extant side channels during the summer low-flow period
491 remained there during their development. Floodplain waterbodies are important nurseries for
492 many fishes in the river (Jeffres, Opperman & Moyle, 2008; Bellmore et al., 2013), and they
493 out-migrate from side channels to the river and sometimes to downstream lakes as they grow
494 (Tamate & Maekawa, 2004; Kanno et al., 2020). Although fish diversity was lower in
495 waterbodies with lower connectivity, we observed some fish species in those waterbodies that
496 were not found in the more connected waterbodies. Species that occurred in isolated
497 waterbodies included Cyprinidae and Cobitidae, which are known to tolerate anoxic conditions
498 (Fagernes et al., 2017). The spatial variation in hydrological dynamics over flood recession
499 period thus contributed to the diversity of fish fauna on the floodplain.

500

501 Interestingly, the presence of amphibians was determined by the timing of flow cessation in
502 each waterbody. Both amphibian species reproduced in the floodplain waterbodies soon after
503 the flood peak, and their eggs hatched and larvae developed in the waterbodies during the low-

504 flow period, a common pattern in amphibians (Griffiths, 1997; Tockner et al., 2006; Holgerson
505 et al., 2019). Our post-flood survey showed that amphibian eggs were laid and larvae were
506 observed in waterbodies where the flow had stopped in the early flood recession period, before
507 their reproductive season. Waterbodies on the floodplain provide important habitats for
508 amphibian larvae, yet use of this temporary habitat exposes them to various risks, including
509 desiccation (Gervasi & Foufopoulos, 2008) and being flushed by flow (Indermaur et al., 2010).
510 Therefore, the timing of environmental shifts such as flood recession may be particularly
511 important for amphibians, which use the floodplain waterbodies only seasonally.

512
513 Finally, benthos distributions were greatly influenced by the occurrence of flow at the time of
514 sampling. We observed diverse aquatic insect taxa, including Ephemeroptera, Plecoptera, and
515 Trichoptera, in flowing waterbodies; most of these taxa require flowing water for respiration
516 (Merritt, Cummins & Berg, 2008), and they were not found in more isolated floodplain
517 waterbodies. In contrast, Chironomidae dominated most of the floodplain waterbodies in this
518 study. Relatively low Chironomidae density in isolated waterbodies, which were not flushed by
519 flow at peak flood, in contrast to other floodplain waterbodies may be explained by the
520 extremely anoxic conditions of benthic habitats in those waterbodies. The low $\delta^{13}\text{C}$ (-40%) of
521 benthos in isolated waterbodies indicates that benthic conditions were so anoxic that methane-
522 derived carbon was consumed by the benthos (Kohzu et al., 2004). In this study, we sorted
523 benthos at the family level, but species level identification could have shown spatial variation of
524 Chironomidae according to the hydrological dynamics over flood.

525
526 Overall, the observed unique responses of the four faunal groups to the hydrological dynamics
527 over the snowmelt flood recession period well explained the spatial distributions of the four
528 faunal groups on a floodplain. This indicates spatial asynchrony in channel isolation from the
529 main stream during flood recession and responses of the four biological groups led their distinct
530 distributions on a floodplain. We however acknowledge the difficulties in clearly teasing out the
531 effect of spatial variation in hydrological dynamics over the flood recession period we focused
532 in this study from the effect of spatial variations in local environmental conditions (such as
533 dissolved oxygen, pH, and substrate type). Future experimental studies would be required to
534 clearly distinguish among these factors. Furthermore, the four faunal groups that are
535 individually presented in this study interact with each other through food webs. Diverse aquatic
536 communities on the floodplain observed in this study indicate that food web structures are also
537 variable. Further studies of community structures and species interactions in relation to
538 hydrological dynamics would provide insights into the dynamic meta-community (Leibold et
539 al., 2004) and meta-ecosystem structures (Gounand et al., 2018) on a floodplain.

540

541 Our results supported the findings of previous studies that showed the link of hydrological
542 connectivity to the river and the floodplain aquatic communities (Tockner et al., 2000; Amoros
543 & Bornette, 2002). Moreover, the results of our studies during the flood period showed how
544 hydrological dynamics during the flood shaped the distribution of each faunal group: plankton
545 and fishes were primarily influenced by whether the waterbody was flushed by peak flow,
546 whereas amphibians were influenced by the timing of flow cessation and continuing flow during
547 low-flow periods was important for benthos. Advanced technology for surveying the
548 environment on broad spatial and temporal scales is now readily available, including time-lapse
549 photography (Parajka et al., 2012), drones (Woodget et al., 2017), and remote sensing (Arnesen
550 et al., 2013). By using these techniques to monitor the hydrological dynamics on floodplains
551 over the flood period, together with our mechanistic understanding of the biological responses
552 to hydrological dynamics, it should be possible to predict the spatial distribution of aquatic
553 communities on floodplains.

554

555 Spatially variable hydrological dynamics and the biological responses on the floodplain were
556 supported by a combination of the natural geomorphic complexity of the floodplain and the
557 natural seasonal dynamics of river discharge over the snowmelt recession period. This study
558 showed that three components of the seasonal river flow dynamics played particularly important
559 roles in supporting diverse aquatic communities: the peak flood stage, flow at low stage, and the
560 timing of flow recession. The magnitude and timing of the snowmelt flood shows substantial
561 year-to-year variations (Yarnell et al., 2010). Long-term monitoring of the floodplain biota may
562 reveal interannual habitat shifts of the aquatic community on a floodplain caused by variation of
563 the hydrological dynamics of the snowmelt recession. Factors such as climate change and
564 regulation by dam construction can also change the seasonal dynamics of river discharge (Poff
565 & Schmidt, 2016; Yarnell et al., 2010). The results of this study highlight the importance of the
566 natural hydrological dynamics of snowmelt recession on floodplain communities.

567

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582 Hokkaido (ID 610).

583

584 **Conflicts of Interest Statement**

585 The authors declare no conflict of interest.

586

587 **Data Availability Statement**

588 The data that support the findings of this study are available from the corresponding author upon
589 reasonable request.

590

591 **Legends**

592 Table 1: Model selection to reveal the threshold of connectivity to the river that influences the
593 density and diversity of each faunal group. Bold numbers indicate the lowest AIC of each row.

594

595 Figure 1: Locations of the flood-recession and post-flood survey sites with different
596 connectivity to the main stream of the Butokamabetsu River, Hokkaido, Japan. Spatial sampling
597 was conducted at 26 palaeo- or extant side channels of the Butokamabetsu River (coloured
598 circles) distributed along a 10 km segment of the river and divided into five blocks. The flood-
599 recession survey was conducted at four focal sites (sites A–D; bold circles).

600

601 Figure 2. Seasonal hydrological changes of the Butokamabetsu River. (a) Total discharge of the
602 Butokamabetsu River from 1 April to 1 July 2019. (b) Examples of spatial changes in
603 hydrological connectivity and flow of various palaeo- and extant side channels during the spring
604 snowmelt and flood recession. (c) The six categories of hydrological connectivity used in this
605 research. Sampling sites were assigned to a hydrological connectivity category based on
606 repeated observations of hydrological connectivity and flow at each site. (d) Changes in flow
607 speed at the four focal sites where the flood-recession survey was conducted.

608

609 Figure 3: Spatial variation (left column; post-flood survey) and temporal variation (right
610 column; flood-recession survey) of water chemistry and plankton density. In the graphs in the
611 left column, each symbol (or bar for Zooplankton) shows the average value of multiple sites

612 with the same connectivity index, and the error bars show ± 1 standard error among the sites.
613 The right panels show the temporal variation of the data at the four focal sites and one main
614 stream site measured during the flood-recession survey.

615

616 Figure 4: Fish species composition at the focal sites during (22 May) and after (12 June) the
617 spring snowmelt flood. Each colour represents a different species, as indicated in the legend.
618 For salmonids, young of the year individuals (YOY) were counted separately, and their densities
619 are indicated hatching.

620

621 Figure 5: Stable carbon isotope ratios of benthos and fish at the focal sites and the mainstream
622 site. Each data point shows the average $\delta^{13}\text{C}$ of benthos (x -axis) and $\delta^{13}\text{C}$ of fish (y -axis) at one
623 site. The error bars indicate ± 1 standard error of individual variations within each site. The
624 dashed line shows the fitted regression line along the connectivity gradient.

625

626 Figure 6: Spatial variation of density and diversity of the four faunal groups in the post-flood
627 sampling. Each bar indicates the average density of each faunal group at multiple sites with the
628 same connectivity to the river. Circles indicate the average taxa richness of each faunal group at
629 multiple sites with the same connectivity. The error bars indicate ± 1 standard error among sites.
630 Black inverted triangles indicate primary and secondary ($\Delta\text{AIC} < 4$) thresholds of faunal
631 density, and white inverted triangles indicate the thresholds of faunal diversity based on the
632 model selection results shown in Table 1.

633

634 Figure 7: (a) Nonmetric multidimensional scaling (NMDS) ordination of the whole aquatic
635 biota (including the four faunal group) at each of the 26 sites sampled during the post-flood
636 survey. Sites are coloured according to their connectivity to the river. Each polygon represents a
637 convex hull created by connecting the outermost site scores for each of the six connectivity
638 levels. Data for the four faunal groups were combined after standardization. (b) Results of
639 similarity percentage analysis (SIMPER). Vertical axis shows the total contribution of each
640 faunal group to the variation in the aquatic biota between connectivity thresholds.

641

642

643

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| Faunal group | Factor | Unit | 15 May | 15 May | 23 May | 28 May | 11 June | Best fit model | Estimate | SE | t-value | p-value | Note |
|--------------|---------------|---------------------|---------------------------|---------------|---------------|---------------|---------------|----------------|----------|-------|---------|---------|------|
| | | | River connection (AIC) | Flow (AIC) | Flow (AIC) | Flow (AIC) | Flow (AIC) | | | | | | |
| Zooplankton | Density | ind L ⁻¹ | 257.6 | 251.9 | 256.3 | 258.4 | 258.5 | 15 May Flow | -40.1 | 13.4 | -3 | <0.01 | |
| | Diversity | # of families | 95.6 | 81 | 75.6 | 89.4 | 91.9 | 23 May Flow | -1.8 | 0.29 | -6.1 | <0.001 | |
| Benthos | Density | ind m ⁻² | 385.6 | 384.9 | 389 | 389 | 389 | 15 May Flow | 550.7 | 225.6 | 2.4 | <0.05 | |
| | Diversity | # of families | 141.5 | 136.5 | 140.1 | 140 | 124.8 | 11 June Flow | 7.24 | 1.37 | 5.29 | <0.001 | |
| Fish | Density | ind m ⁻² | 35.3 | 31.7 | 34.1 | 35.7 | 37.9 | 15 May Flow | 0.32 | 0.11 | 2.9 | <0.01 | * |
| | Diversity | # of species | 112.4 | 109.7 | 116.1 | 117.1 | 116.6 | 15 May Flow | 1.9 | 0.57 | 3.31 | <0.01 | |
| Amphibian | Presence rate | | 84.1 | 79.5 | 69.8 | 71.5 | 79.6 | 23 May Flow | -1.24 | 0.25 | -5 | <0.001 | |
| | Diversity | # of species | 84.1 | 79.5 | 69.8 | 71.5 | 79.6 | 23 May Flow | -1.24 | 0.25 | -5 | <0.001 | |

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* significant effect of sampling area: Estimate=-0.00093, SE=0.00042, t-value=-2.2, p-value<0.05

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Table 1

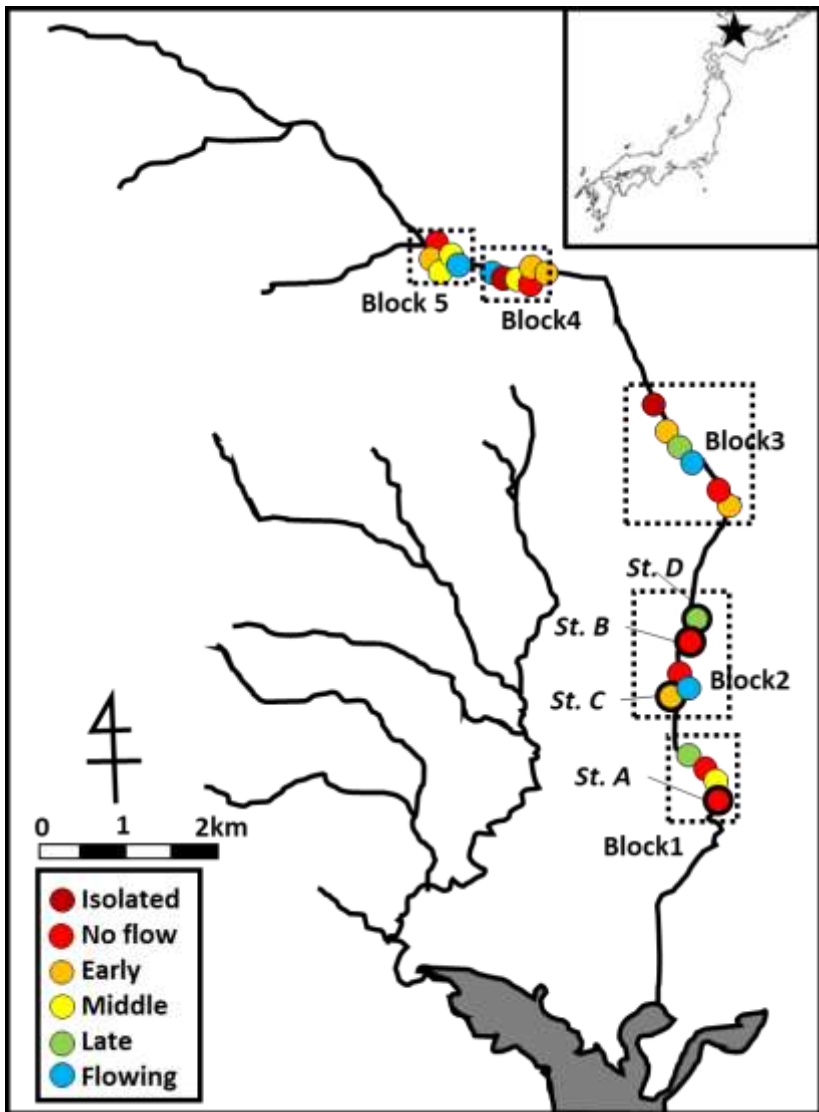
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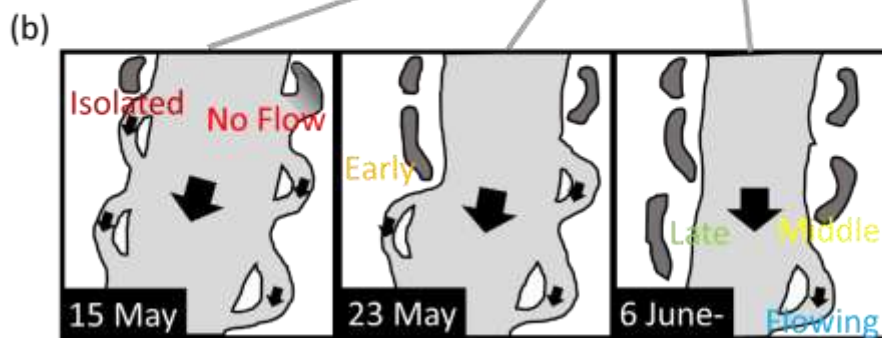
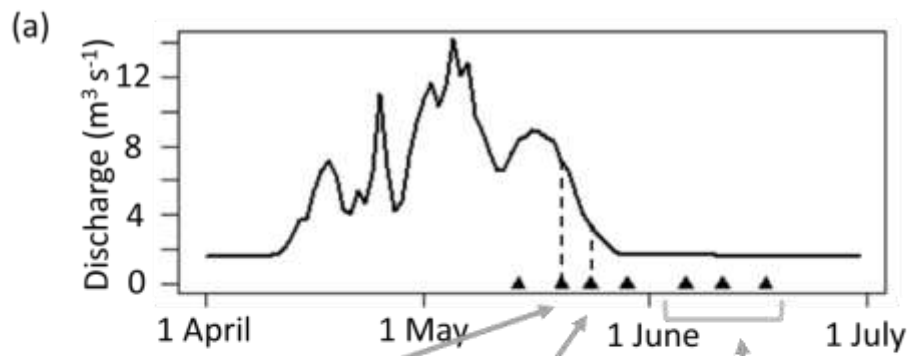
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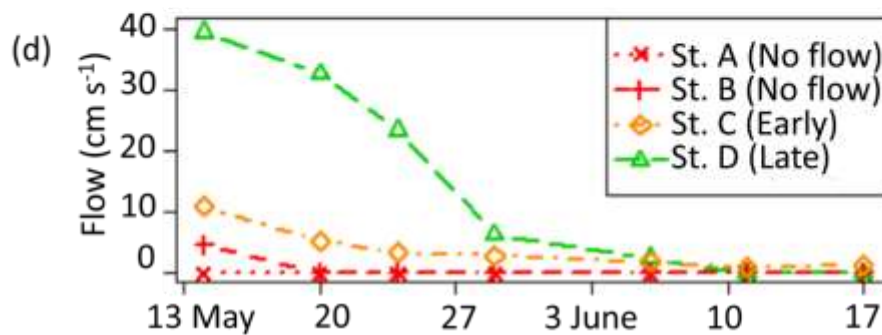
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815 Figure1



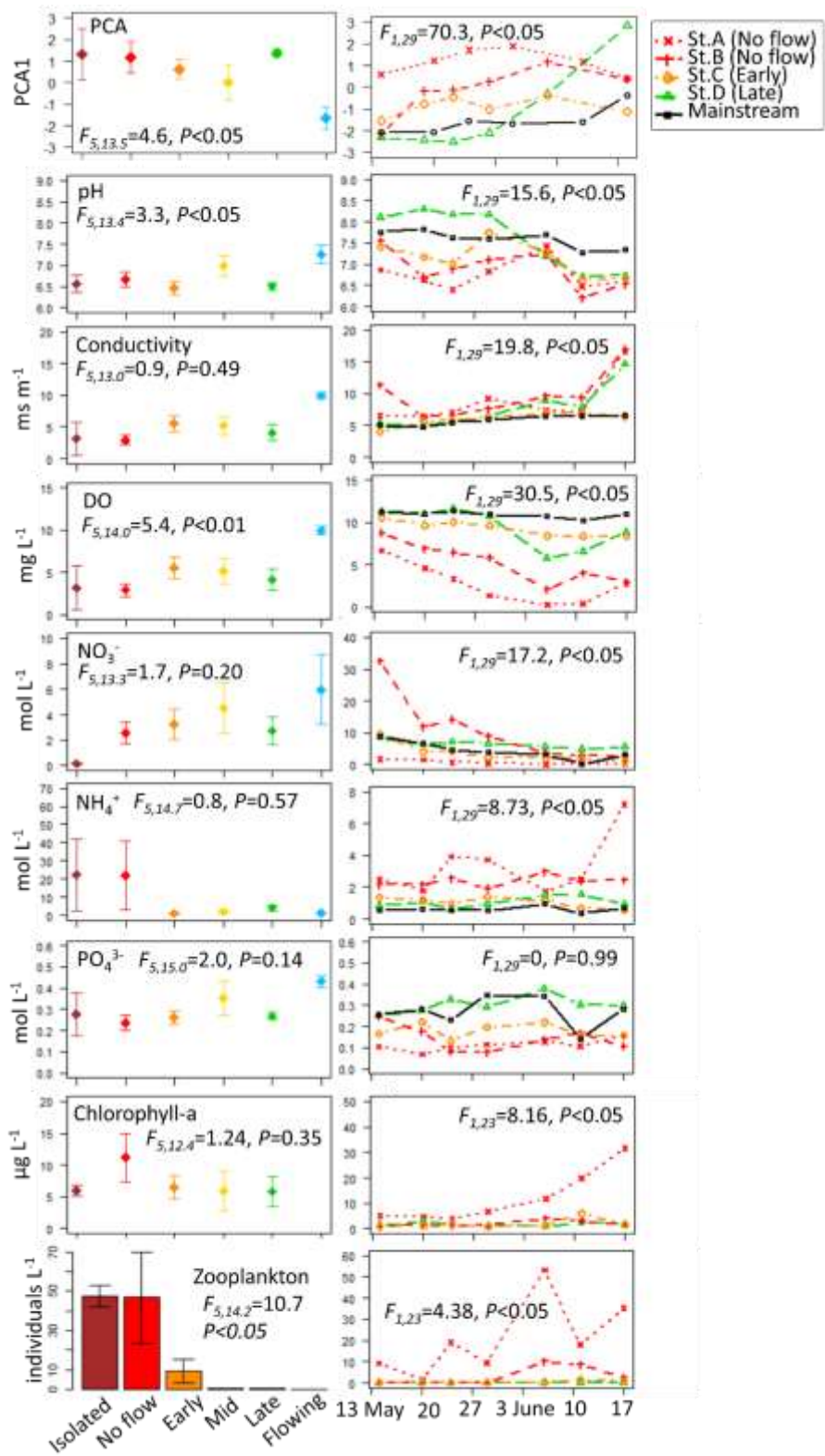
(c)

| Connectivity category | n | 15 May River connection | 15 May Flow | 23 May Flow | 28 May Flow | 6 June Flow | 11 June Flow |
|-----------------------|---|-------------------------|-------------|-------------|-------------|-------------|--------------|
| Flowing | 4 | ○ | ○ | ○ | ○ | ○ | ○ |
| Late | 3 | ○ | ○ | ○ | ○ | × | × |
| Middle | 4 | ○ | ○ | ○ | × | × | × |
| Early | 6 | ○ | ○ | × | × | × | × |
| No flow | 7 | ○ | × | × | × | × | × |
| Isolated | 2 | × | × | × | × | × | × |



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817 Figure 2

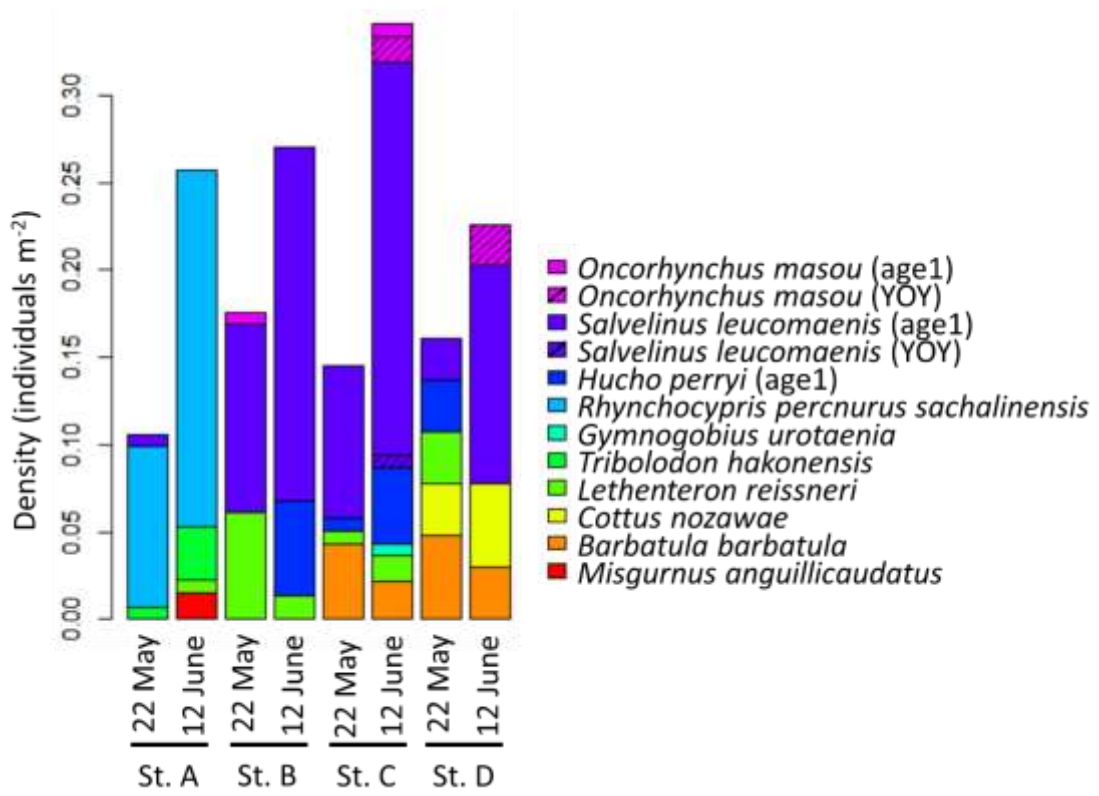


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819 Figure 3

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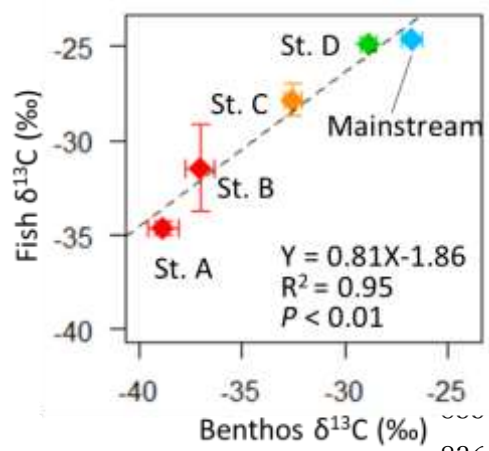
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824 Figure 4

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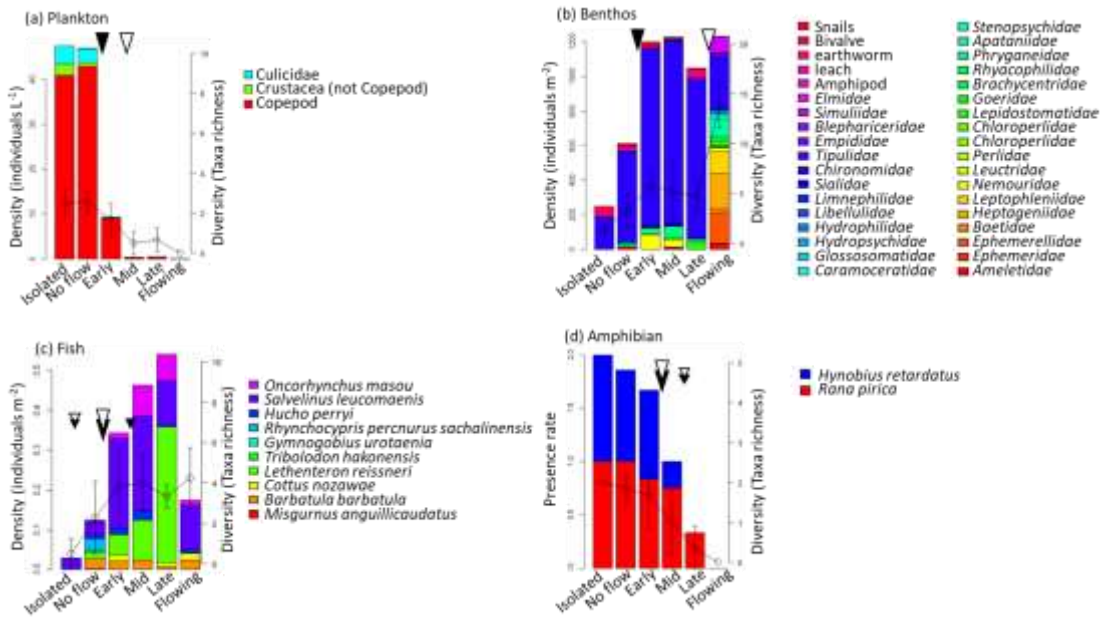
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838 Figure 5

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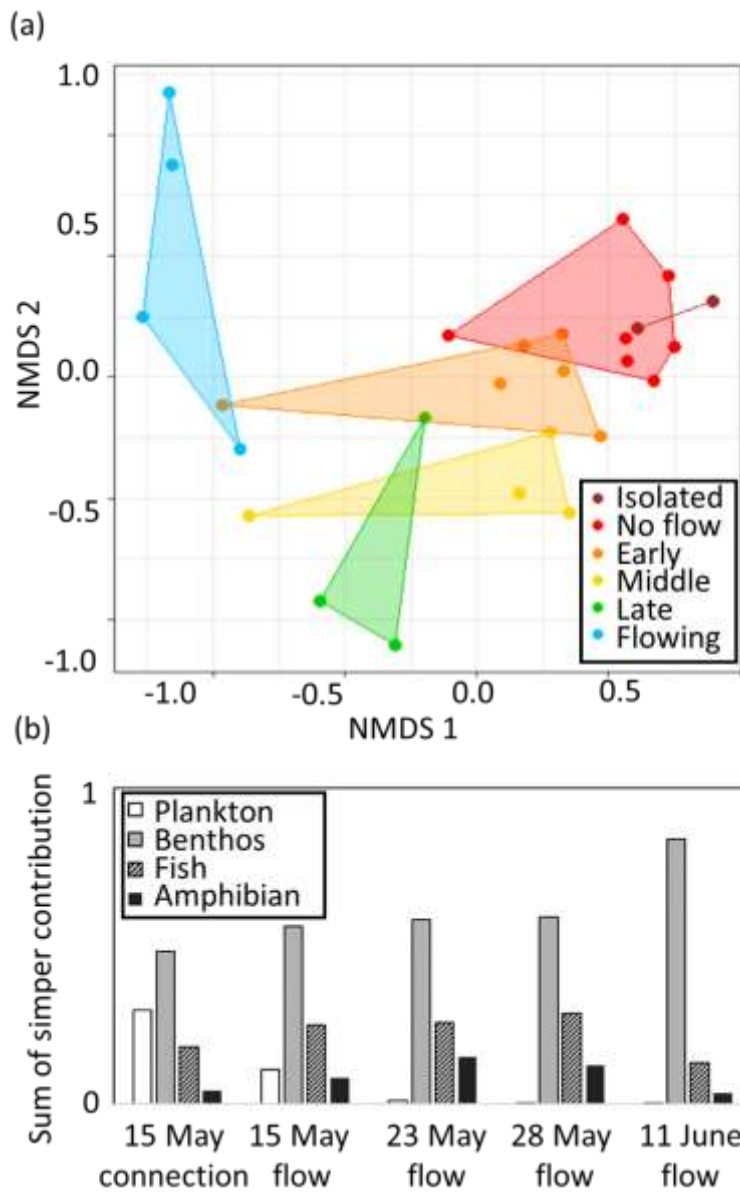


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843 Figure 6

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847 Figure 7

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