Analysis of Bacterial Fatty Acids by Flow Modulated Comprehensive Two

Dimensional Gas Chromatography with Parallel Flame Ionization Detector / Mass Spectrometry



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INTRODUCTION

A commercially available flow modulated GC imes GC system was tested and optimized for the analysis of bacterial fatty acids (as methyl esters). The system configuration included parallel MS and FID detection. The results are compared to data obtained on a thermal modulation system.

EXPERIMENTAL

A bacterial acid methyl ester (BAME) solution in methyl caproate obtained from Supelco was used as a reference sample. A Stenotrophomonas maltophilia bacteria sample was prepared using the Sherlock MIDI standard operating procedure (M. Sasser, MIDI Technical Note 101, 1990, see www.midi-inc.com).

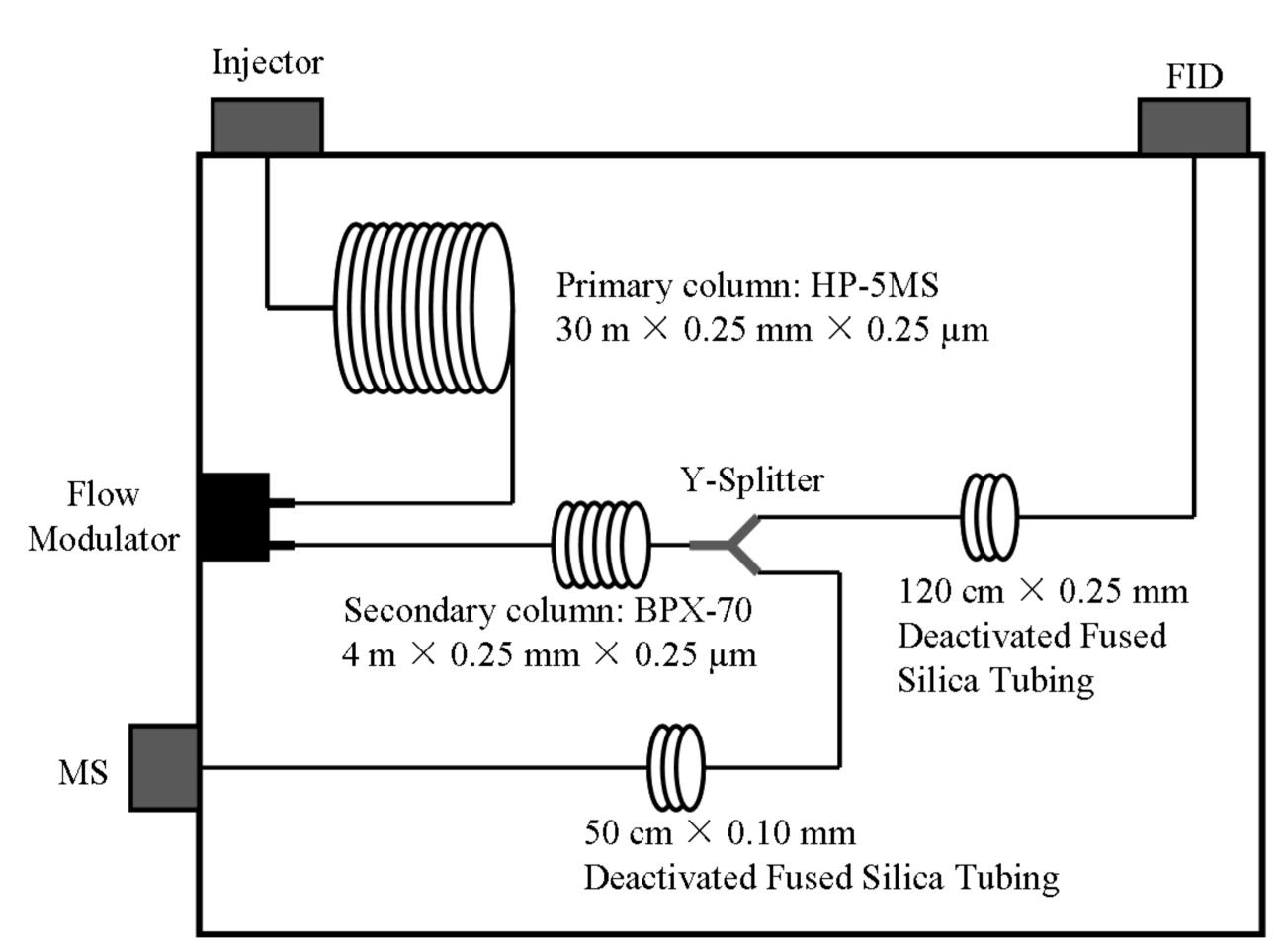


Fig.1 A schematic of the flow modulated GC×GC with FID and MS

Analytical parameters

Instrument: Agilent 7890A GC & 5975 MSD *Inlet*: SSL at 250 °C, 1 μL, split ratio 10:1 Carrier gas: Hydrogen, constant flow

First dimension column: HP-5MS 30 m x 0.25 mm x 0.25 μm

¹D gas flow: 0.6 mL/min

Second dimension column: BPX-70 4 m x 0.25 mm x 0.25 μm

²D gas flow: 25 mL/min

Modulation time: 2 s; Sample time: 1.9 s

Oven: 100 °C (2 min) - 2 °C /min - 240 °C (10 min) Detection: FID and MS (scan range m/z 40 - 430; 20 scans/s)

OPTIMIZATION

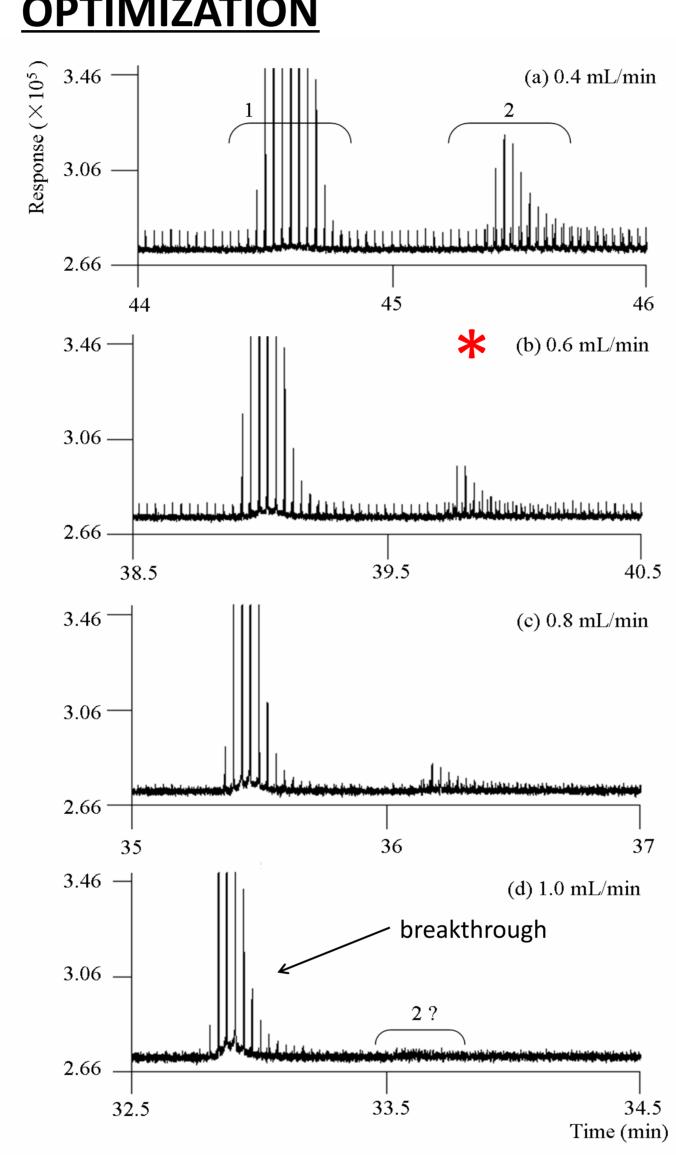


Fig. 2 The modulated chromatograms at different ¹D flow rates. 1. 13:0; 2. 12:0 2OH.

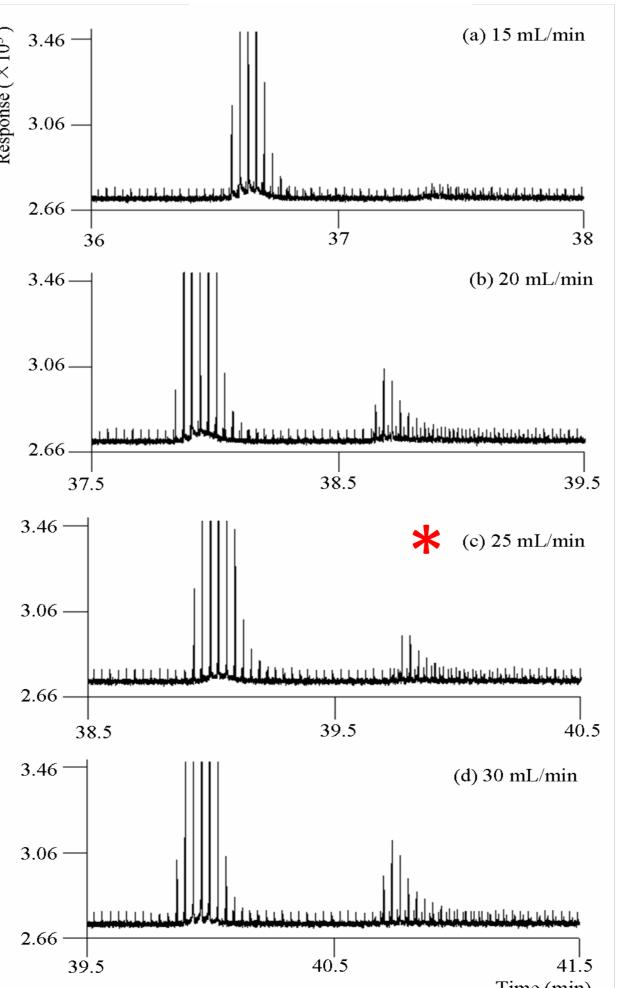


Fig. 4 The influence of the ²D flow

(b) 0.6 mL/min 18:2 w9,12cc (c) 0.4 mL/mi

Fig. 3 The $GC \times GC$ plots of the C18 fatty acid elution area at different ¹D flow rates

***** Optimal conditions:

¹D gas flow: 0.6 mL/min ²D gas flow: 25 mL/min *Modulation time:* 2 s Sample time: 1.9 s

RESULTS

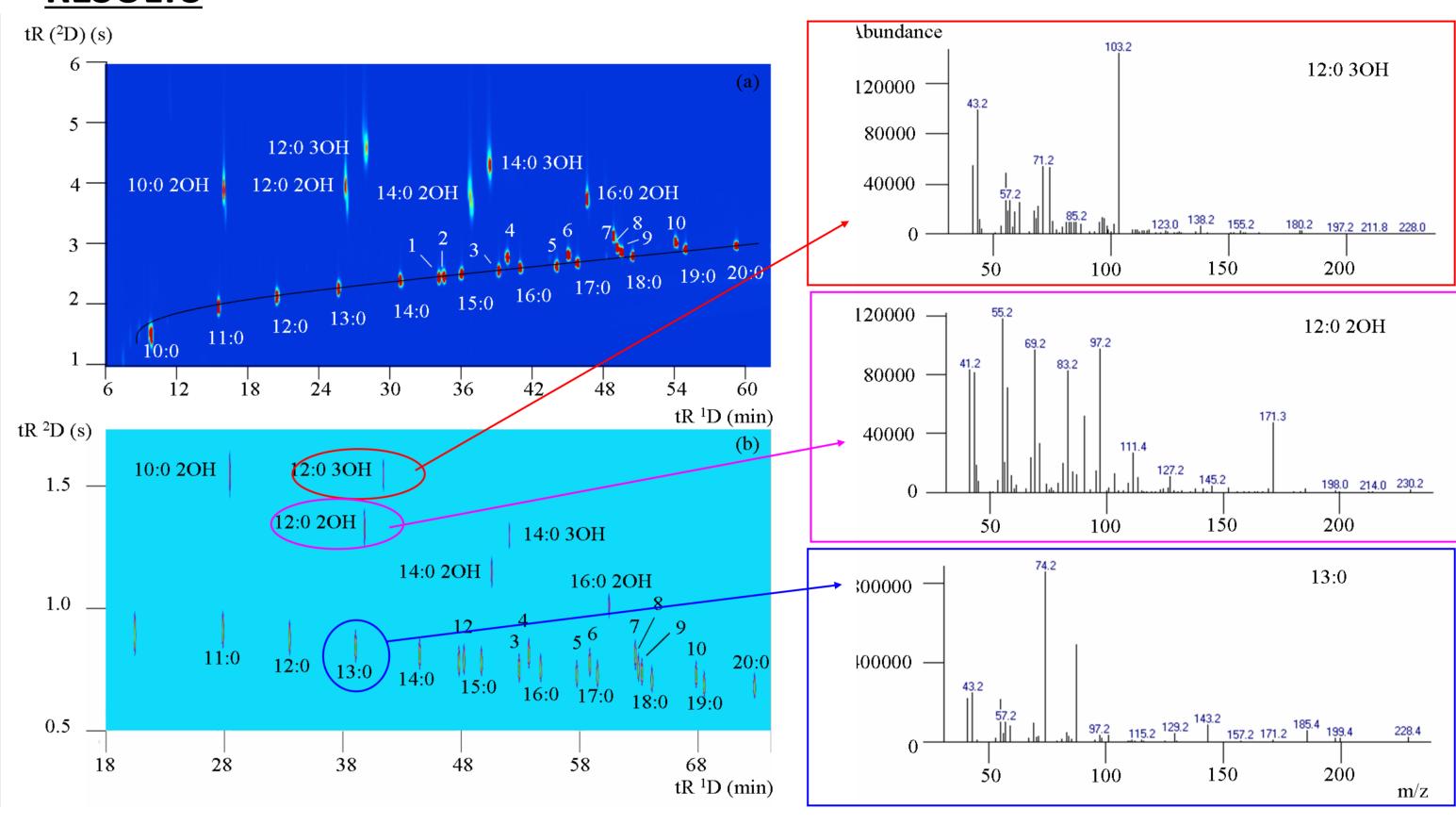


Fig. 5 GC×GC plots of a BAME reference sample obtained by thermal modulation (a) and flow modulation (b) and mass spectra of typical solutes in BAMEs standards

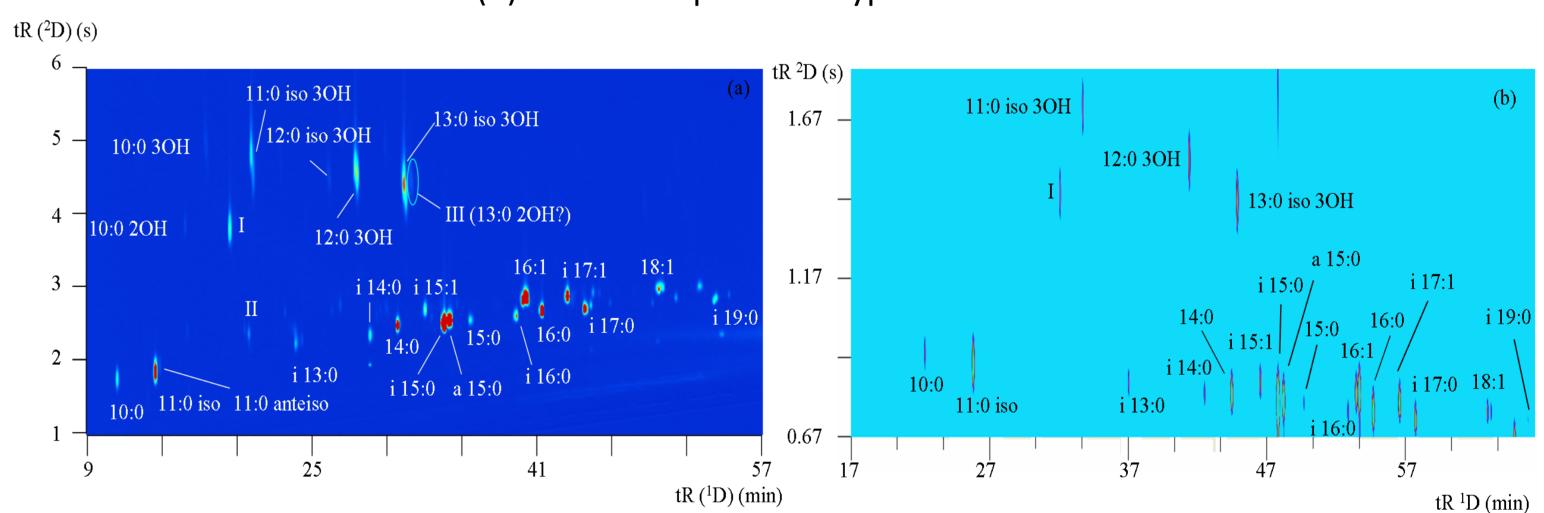


Fig. 6 GC \times GC plots of BAMEs from *S. maltophilia* by thermal (a) and flow modulated GC \times GC (b)

CONCLUSIONS

The GC×GC plots obtained for a reference sample of bacterial fatty acid esters and a bacteria sample (S. maltophilia) were very similar to those obtained by thermal modulated $GC \times GC$. The $GC \times GC$ approach is especially interesting in detecting the presence of hydroxy fatty acids. The parallel FID/MS set-up is useful since the MS allows identification and confirmation, while the FID allows comparison of the relative fatty acid composition with existing databases (Table 1).

Table 1. Relative composition of BAMEs in *S. maltophilia*

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| Compound | Peak I (min) | Peak II (sec) | volume (%) | MIDI data (%) |
|--------------|--------------|---------------|------------|---------------|
| 10:0 | 22.30 | 0.94 | 0.78 | 0.76 |
| 11:0 iso | 25.80 | 0.90 | 4.82 | 4.37 |
| I (unknown) | 32.07 | 1.44 | 1.55 | 2.02 |
| 11:0 iso 30H | 33.70 | 1.71 | 1.74 | 2.22 |
| i 13:0 | 37.00 | 0.85 | 0.78 | 0.50 |
| 12:0 3OH | 41.40 | 1.54 | 3.04 | 3.85 |
| i 14:0 | 42.50 | 0.81 | 0.58 | 0.62 |
| 14:0 | 44.47 | 0.80 | 4.10 | 3.04 |
| 13:0 iso 30H | 44.83 | 1.42 | 3.38 | 4.93 |
| i 15:1 | 46.50 | 0.84 | 1.33 | 0.91 |
| i 15:0 | 47.77 | 0.77 | 44.15 | 35.24 |
| a 15:0 | 48.20 | 0.78 | 6.12 | 9.29 |
| 15:0 | 49.67 | 0.78 | 0.35 | 0.45 |
| i 16:0 | 52.83 | 0.75 | 0.57 | 1.05 |
| 16:1 w9c | 53.43 | 0.81 | 2.82 | 2.79 |
| 16:1 w7c | 53.67 | 0.81 | 11.58 | 10.74 |
| 16:0 | 54.67 | 0.75 | 5.03 | 6.35 |
| i 17:1 w 9c | 56.57 | 0.78 | 3.76 | 4.18 |
| i 17:0 | 57.70 | 0.72 | 2.22 | 3.22 |
| 18:1 w9c | 62.90 | 0.75 | 0.83 | 1.14 |
| 18:1 w7c | 63.17 | 0.75 | 0.47 | 0.63 |
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