

Functional thermal limits are determined by rate of warming during simulated marine heatwaves

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Marine Ecology Progress Series

Manuscript:	MEPS-2021-06-001/R1 RESUBMISSION
Title:	Functional thermal limits are determined by rate of warming during simulated marine heatwaves
Authors(s):	Rebecca De Leij (Corresponding Author), Laura Grange (Co-author), Lloyd Peck (Co-author)
Keywords:	Climate change, Echinoderm , Extreme warming events, Polar, Segmented regression, Sub-lethal limits, Thermal tolerance
Type:	Research Article

Dear Dr. Lauzon-Guay,

Thank you for the opportunity to submit a revised draft of the manuscript titled: 'Functional thermal limits are determined by rate of warming during simulated marine heatwaves' to Marine Ecology Progress Series. We appreciated the detailed and valuable feedback that was provided by the managing editor, Christine Paetzold, and all three referees. In particular, the suggestion from reviewer 1 to revise the statistical analysis has enabled an important error to be amended. In addition to this, we also value the comment by reviewer 3 to further explore relationship between size and thermal thresholds. This additional analysis has led to some interesting insight into the interactive effect of urchin size and temperature on the time taken to right. The authors have been able to incorporate most of the suggested changes in the manuscript, all of which have been highlighted as tracked changes in the pdf uploaded as 'Manuscript showing edits'. For comments that have not resulted in changes to the manuscript, we have provided further explanation in the responses below.

Managing Editor Christine Paetzold comments:

Please also address the following points in your revision:

(i) Please carefully check your manuscript for correct spelling (e.g. 'Absract').

This has been amended

(ii) P4 L22-25: are all these references essential to be listed? If possible, please avoid 'strings' of citations (more than 3 cites in a row). Such strings make the text very tedious to read and give the impression of a literature review rather than the selectivity expected in a scientific article. Try to be more selective and just list 3-4 references preceded by 'e.g.'.

This has been amended throughout

(iii) When you report the first result from a statistical test, please provide the name of the test. If the following results are from the same test, there is no need to repeat the name.

This has been amended

(iv) P15L20-22: if these results are from post-hoc tests, the p-value suffices. Otherwise, please change the df-value (after the t) to subscript font and remove the brackets. Please clarify what the d-value you report is (if this was not made clear previously).

This has been amended and clarified

(v) Table 1: is it intentional that for 'Faeces produced, 1o C day-1', there is no result for Slope_2?

Yes this is intentional. There was no breakpoint identified in this regression and hence no second slope. The statistics are therefore reported for Slope_1 only. Further explanation has been added as a footnote to the table.

(vi) Tables and figures: please only capitalize the first word in table headers and figure labels. E.g. "Individuals feeding (%)"

This has been amended

Reviewer 1 report:

The manuscript explores the impact of sublethal warming during heatwaves on the physiology of an Antarctic sea urchin. The authors performed an experiment involving temperature ramps with different rates of temperature increase. The authors focus on a combination of feeding physiology, activity levels and metabolism is an interesting a much needed approach to fully understand the impact of this events.

I think the manuscript deserves publication. However I have some comments that I think need to be taken care off.

- in the introduction and discussion, warming and heatwaves are often mixed, while they are related their impact will be different and while the experiments may apply well to current cases of heatwaves, we have seen already these experiments don't relate well to long term warming. Also since up the experiments were performed from just one base temperature, these results are difficult to discuss under the context of heatwaves on a warmer scenario.

This is a valid comment and we agree that the distinction between temperature increase from MHWs and temperature increase by gradual climate warming was not made clear. We also acknowledge that the terms were confused in the original manuscript and as such we have now made amendments to the text to adjust the focus on MHWs in the current climate with only cautious interpretation in the context of the future.

- the methods need to be clarified. ANOVA is not adequate for this type of data and experiment (repeated measurements). The use of cumulative degree days needs to be justified better, also the resulting scale is confusing, lower cumulative effect results in larger impact, there is nothing wrong there but the justification of their use (a common scale to compare time points) is a little bit weak. Other analyses may have been considered, such as nonlinear decaying models, survival models, etc that consider dosage and time and kinetics of processes which are specially affected by temperature.

Firstly, thank you for bringing this statistical error to our attention. Instead of a traditional one-way ANOVA, we have now re-analysed the differences between treatments during the experiment using a 'one-way repeated measures ANOVA'. This analysis accounts for related and non-independent groups as is appropriate for our data. Treatment group variance are compared when treatments reach the same temperature increments and are referred to as t1, t2, t3 etc. A figure of the results of this analysis is now included in the supplementary materials.

With regards to cumulative intensity, on revising the manuscript we have decided that this metric creates unnecessary confusion. Although cumulative intensity allows temperature and exposure to be translated into a metric directly comparable between treatments, we think this is achieved by reporting only temperature and observing the rate at which this temperature was reached (i.e. 0.3oC/day, 0.5oC/day or 1.0oC/day). We hope this change will also make the paper and the data easier to reference in other studies since most previous studies report on temperature limits.

I also have a series of minor comments in the attached doc file

**These have all been carried out in the amended manuscript
In particular:**

Comments in the introduction: the use of the term ‘permanent heatwave’ has been removed to avoid confusion and the wording and sentence structure has been changed where appropriate (page 4 and 5).

Comments on page 6, line 23 and page 7, line 4: Further information regarding rationale behind non-feeding period and standardisation of metabolic activity has been added on page 8, line 21-24 and page 9 lines 1-12.

Comment page 11, line 1: A repeated measures ANOVA has now been used and results have been amended throughout.

Comments on page 12: The use of cumulative intensity has now been replaced by temperature to avoid confusion.

Comment page 13, line 16: We were not necessarily looking for the simplest model, but rather we were looking for any change in the regressions gradient which then indicated that the functions response to temperature increase had changed. A linear regression may be sufficient to explain the relationship, however it may mask the subtle change in the rate of degradation experienced when a species hits a thermal threshold. We have therefore removed any reference to model fit (R^2) that was used to justify the segmented regression and have instead emphasised the reason behind using a segmented regression as explained above in the manuscript.

Comment page 19, line 9: This is an interesting point and something that had not been thought of when interpreting the results. As a result, we have added a section to the discussion relating to the faecal production rates and the reasons driving these on page 26, lines 22-25 and page 27, lines 1-10.

Comment on page 20: Discussion on the effects of nutritional status has been added on page 29, lines 12-25 and page 30, lines 1-6.

Comment page 21, line 4: This paragraph has been amended considering this comment.

The following linked document contains further information from this reviewer:
https://www.ManuscriptManager.net/sLib/v4/marked_docs/mm_meps~1997~ce9b5128585e~1~965.ReviewerMarkUp.docx

Reviewer 2 report:

Functional thermal limits are determined by rate of warming

General comments

This is an important and excellent study, nicely executed and described. This is because while there are a number of studies examining the outcomes of warming in Antarctic species, the potential outcomes of marine heatwaves are less well understood in the regions ecosystems.

Research here is one of the few studies to directly address polar MHW, using the common Antarctic sea urchin *Sterechinus neumayeri*, to explore the outcomes of warming rate/time on key physiological processes. Treatments (rates of warming) are set at levels that are realistic based on observed temperatures in the Antarctic Peninsula level.

In line with previous research on Antarctic invertebrates, the rate of warming is important in determining upper thermal limit and maintenance of biological function. Such previous uni-directional experiments (temperature always increasing) have been used to understand the outcomes of long-term temperature increases, yet MHW are different in that they represent short-term warming periods (cyclic). These might need another experimental approach that mimics observed heat wave patterns (i.e. warming for a time then returning to ambient temperatures: as 0°C, up to +7°C for 10 days, then back to 10°C etc based on S2). Perhaps some discussion on the potential outcomes for animals on shorter term exposures and the responses that could be seen following returning animals back to ambient conditions. How might the present experiments be interpreted in terms of this type of cyclic treatment?

This is an interesting comment and something we have explored in a subsequent experiment (in prep). We have now added a section in the manuscript discussing the scope for recovery following MHWs (page 31, lines 17-25 and page 32, lines 1-4).

Minor comments

(1) Table 1 summarizes figures 2, 4, 6 and 8 so suggest these could go in the supplementary material.

After consideration, we have decided to only present the regression plots with the raw data within a single figure. This now includes the control data. It was apparent that the line plots did not provide any additional useful information and the raw data points plus the regression was enough to summarise the results. We also felt that the differing scales on the line plots and regressions (cumulative intensity vs temperature) was confusing and now therefore present temperature only.

(2) S1: Can you please include a plot over the 20 year period, indicating periods when the 90% percentile is exceeded. This would be more useful in addition to the table in understanding the magnitude and frequency of the HW in the region.

We have now presented a plot with this information in the manuscript (Figure 1).

(3) Page 19, line 8, and elsewhere: suggest changing 'different functions' to 'key biological functions'.

This has been amended

(4) Is fig 12 needed?

The referee is correct that fig 12 is not needed. The CTmax values are mentioned in-text and figure 12 has now been removed.

Reviewer 3 report:

Overview

The impacts of climate change are being felt in every marine system, but understanding the impacts on some of the most at risk (i.e. Antarctic habitats) remains a priority. Whilst it may be possible for species to adapt to the gradual increases in base temperature, the effects of more frequent and more intensive extreme climate events are already being shown to be a major issue. Using *S. neumayeri*, a common (but keystone species), this study aims to understand the connection and relationship between functional sub-lethal limits and critical thresholds. Monitoring basic functions of the sea urchin under different warming regimes enables the authors to assess the ability of this species to tolerate elevated temperatures. I believe that the premise of this paper is valid and the scientific questions underpinning it are important and need addressing within a journal such as MEPS. However, it does require some revision before it can be accepted and I have highlighted the areas for improvement below.

Abstract

A lot more results are required. Currently, only lines 18-21 present the outputs from the experiments. I would recommend reducing the introduction and expanding the results accordingly.

This has been amended as suggested.

Page 2, Line 12. It would be better to specify that it was respiration (or even oxygen consumption) that was measured rather than metabolism. Metabolism means some substances are broken down to produce energy and some other substances are synthesised. In contrast, cellular respiration means oxidation of organic compounds in cytoplasm to produce energy in the form of ATP. This energy helps the cell to perform all metabolic activities.

This has been amended as suggested.

Introduction

The Introduction is a little long and should be shortened to reflect the focussed nature of the experimental procedure and outputs.

The introduction has now been reduced by 300 words

Page 4, line 3. A very long sentence that needs to be re-organised.

This sentence and the whole paragraph has now been re-worked.

Page 5, line 25. It is important to remain scientific with the writing so I would remove the reference to a 'catholic diet'.

'Catholic' has now been replaced with 'varied'

Page 6, line 1. Although the authors give a number of examples of slow maturity for Antarctic species, according to Pearse and Giese (1966) '...oocytes take from 18 to 24 months to reach maturity after beginning growth, and spawning occurs sometime in the winter or spring between May and December' for *S. neumayeri*. It is important to contextualise this species in the methods in more detail.

We have added a paragraph in the methods to contextualise the species as the reviewer suggests (page 7, lines 15-25 and page 8, lines 1-8).

Material and Methods

Page 6, line 10. Size ranges should be included and it would be important to look at using size as cofactor that may explain some of the variability in the results. Including survivorship and growth data would also underpin the 'quality' of the experimental system used.

Size ranges have now been included on page 7, line 12. We could not include size as a cofactor in the analysis since data were pooled within replicate tanks, so the relationship had to be explored separately. As suggested, scatter plots of size vs function have also been explored to identify whether this metric caused any variability in the results (Included in the supplementary materials, Figure S5 and Table S3). Both feeding and oxygen consumption were standardised by animal weight to account for size variability and as such, no relationship was found between size and these functions. For the function of righting, the data showed that the larger individuals had the longer righting times at high temperatures. Clearly there is an effect of size here and as such the analysis and results have now been included in the manuscript (page 16, lines 16-22 and page 21, lines 7-9)

There were no mortalities in the ambient control conditions and mortalities in treatment conditions only occurred following CTmax. Growth data was not collected in the experiment since the growth rate of *Sterechinus neumayeri* is slow, with individuals reaching a maximum diameter of 70mm after 40 years (Brey et al., 1995) and the size or weight increase across two months (experiment duration) would be lost within the variability encountered from measurement error.

Page 6, line 16. Here and elsewhere, the analysis of the timeseries data for heatwaves using the R package is an important part of the story and results. I would, therefore, like to see this as a more than just supplementary as it seems to be novel. For example, a figure showing the temperatures with the identified heatwaves could then be linked back to the results, especially re-purposing/replacing Figure 12. It would also be good to link to the cumulative intensity (e.g. Tables 2 and S2) so that this explicitly links this measure to the 'real world'.

We have now included a plot (Figure 1) covering the 20-year period which indicates periods when the 90th percentile is exceeded for ≥ 5 days (i.e. MHW). We have included reference to this figure throughout the manuscript. With regard to the cumulative intensity comment, on revising the manuscript we have decided that this metric creates unnecessary confusion. Although cumulative intensity allows temperature and exposure to be translated into a metric directly comparable between treatments, we think this is achieved by reporting only temperature and observing the rate at which this temperature was reached (i.e. 0.30C/day, 0.50C/day or 1.00C/day). We hope this change will also make the paper and the data easier to reference in other studies since most previous studies report on temperature limits.

Page 6, line 22. I would change the replace 'starved' with 'not fed' as it is possible that the sea urchins were grazing the biofilms of the flow-through aquaria and gaining significant nutritional benefits during this time.

This has now been amended

Additional details of the these holding tanks e.g. capacity, light conditions, flow rates, physiochemical parameters are also needed.

Capacity and light regime are now included in-text. Reporting flow rate would not be useful for the experimental set-up. Urchin tanks were not operated on a flow-through system (urchin tanks were water changed every 48 hrs) but were instead floated in water baths which were flow-through. Flow rate in these tanks was variable and used for temperature maintenance.

Page 7, lines 6-16. The details of the experimental design (e.g. if the tank [water baths] were replicated themselves) need to be expanded. There is a hint of pseudoreplication, i.e. the temperature treatments were not replicated and this needs to be clarified and then considered for the statistical analyses performed. This also extends to the 6 urchins per tank, which are definitely pseudoreplicates and need to be accounted for in the data structure for the analyses (if not already).

Thank you for highlighting this error with regards to pseudo replication with floating tanks. We have now ensured that data are pooled prior to analysis and all results and data figures have been reconstructed considering this. With regards to potential pseudo replication within temperature treatments, we have addressed this comment in the manuscript and provided more detail on the experimental design (page 9, lines 13 – 24, page 10, lines 1-2).

Page 7, lines 12-14. I would contest that the parameters stated do not reflect good water quality conditions. Specifically, the large fluctuations in pH, and the presence of even low nitrite, (ammonia/ammonium needs to be clarified) and elevated nitrate are sub-optimum for invertebrates from pristine conditions. Physiochemical parameters: means, ranges etc should be presented in the supplementary. This will also be important as warming will have an impact on the level of dissolved oxygen (see Peck et al., 2007) and in addition to temperature will, therefore, influence the respiratory processes and the interpretation of the data in the discussion (page 20).

The ranges originally reported were those described on the water quality test kits, however the actual values/concentrations measured were much lower and we believe demonstrated good, if not, pristine conditions since they were comparable to the water pumped directly from Ryder Bay. Concentrations/values did not deviate outside the ranges stated in-text and the test kits used (JBL) did not provide further accuracy between these ranges. As such, we do not see the need to included additional information on these parameters in the supplementary.

We have, however, included a figure of daily temperature changes in each treatment in the supplementary information (Figure S4). Further to this we have added a section in the discussion regarding the relationship between temperature and dissolved oxygen, and the implications of this for our results (page 27, lines 16-25 and page28, lines 1-4).

Page 7, lines 21. It is not clear what was used as a control (no increase in temperature). In the figures ambient conditions are mentioned, but is this from urchins maintained in the same setup, but with ambient water/light levels etc. or is it from another system/field?

A sentence giving further explanation to the control system has been added on page 10, lines 24– 25 and page 11, lines 1-2) and the addition of the control data has now been included in the graphs (Figure 2)

Page 8, line 9. It is odd that this species was fed limpets when the authors highlight its ‘Catholic’ diet. Some discussion around the rationale and also the significant difference between an algal/biofilm grazing and essentially carnivorous diet is required as this will have an impact on the nutrient storage capacity (in the gonads) and ability to tolerate stress due to improved body condition.

It was understood prior to starting the experiment that a diet of limpets was not necessarily representative of a natural diet, however the rationale behind the decision has now been explained on page 11, lines 13-23. We also now state in this paragraph that body condition may be altered as a result with the inclusion of references which provide evidence of this. We originally considered in the discussion that food quality and quantity are factors which will likely have influence on temperature thresholds in the real world (page 32, lines 8-12).

Page 9, lines 3 and 16. Good examples of the potential replication issue ($n = 5$ rather than 10 as measurements of urchins from the same tank would have to be combined). Also, how were urchins selected and were they sampled more than once over the experimental period?

Further explanation on the sampling strategy has been added on page 12, lines 15-20 and again clarification for which urchins were sampled has been added on page 13, lines 8 and 24.

Page 10, line 4. Righting has been used as a biomarker for stress in many species, but it is known that some invertebrates can go in to a heat coma and then recover (see Sandison et al., 1966; Hamby, 1975; Watson et al., 2012). This also links in to other endpoints that were measured.

This is an interesting comment, especially in terms of how we defined CTmax for this species. However, if the individuals were to enter a heat coma from the experimental temperature, due to the continued ramping of temperature, we can expect that it would be unlikely that the animals would then recover (Sandison et al., 1966; Hamby, 1975). If temperatures resumed ambient levels, it would undoubtedly be possible that the urchins could recover, not only in righting ability but also the other key biological functions measured. Ability to recover following warming stress was not the aim of the experiment and to assess this would require further experimentation outside the scope of this paper. We have added a section on recovery post-MHWs in the manuscript (page 31, lines 17-25 and page 32, lines 1-4).

Page 11, line 11. The Wilcoxon test is a non-parametric equivalent of a T-test and so is not equivalent to multiple comparison tests (e.g. Tukey’s). More detail is required for this.

The original analysis using ANOVA and equivalent non-parametric tests were not deemed appropriate for this type of experiment following reviewer 1 comments. As such, analyses have been replaced with a) a repeated measures ANOVA, transforming data initially to achieve normality of data distribution, followed by a post-hoc t-test, or b) a one-way ANOVA followed by a post-hoc Tukey test (as per ctmx data). All data

are now pooled by tank before any analysis.

Results

It is important to have the basic physiochemical, survivorship and control data presented. It should be in the supplementary but is essential and also needs to be discussed.

We have amended the graphs presented in the manuscript in light of some of the reviewer's other comments. These data figures also now include control data which we feel has improved the visual interpretation of the results.

Twelve figures/tables are excessive for this type of paper and the results presented. The authors need to reduce the number. I would have thought half this would be appropriate.

As mentioned, we have reduced the number of data figures and tables to two and one, respectively, which we feel more clearly summarise the data.

Discussion

Page 18, lines 3-12. I had difficulty interpreting this section. It reads as though the data presented follow the failure-rate model, but in line eight the authors state 'However, contrary to this...'. Some work on this ambiguity is required.

The CTmax data presented follow the failure rate model, however the functional thresholds do not. We have changed the wording slightly to try and make this message clearer.

Page 18, line 17. Figure 12 is not very informative and so it would be much better to include the link to the ambient conditions and heatwaves data as discussed above.

Figure 12 has been removed and data on ambient control conditions and heatwaves incorporated as suggested in Figure 2.

Page 19, lines 1-6. The physiological processes stated as being important in lethal limits seem to be working at different scales. For example, nervous and circulatory failure are dependent on the higher functioning of an organisms, whilst enzyme tolerances and chaperone proteins are at the cellular/biochemical levels. These do not seem to sit together and, therefore, should be discussed separately.

This section has been amended so that these processes are discussed separately, and the distinction has been made clearer (page 25, lines 6–19).

Page 21, lines 1-15. This is a good example of where including the actual data from the environment would give added strength to the data presented. However, some discussion about the date 2100 and the number of generations that could enable phenotypic and genotypic adaptations would also be important.

We have added to the discussion (page 31 lines 1–22) to elaborate on the number of generations associated with 2100 for this species and the implications for adaptations. As mentioned before, we have now presented data on the MHWs from the environment in Figure 1 and Figure S2, and given reference to this in-text (page 8, line 14 and 16, and page 30, line 11).

Page 21, lines 16-20. This must link to the conditions provided in the experimental system, which I attest are suboptimum. The authors must also explore this in the context of the 'ambient control' and the potential food supply differences.

This comment has now been addressed through inclusion of temperature graphs, water quality ranges and discussion of diet and nutritional status. We have also included all ambient control data in the figures.

References cited

Hamby R.J. 1975. Heat effects on a marine snail. *The Biological Bulletin* 149: 331–347.

Peck, L.S., Morley, S.A., Pörtner, H.O. and Clark, M.S., 2007. Thermal limits of burrowing capacity are linked to oxygen availability and size in the Antarctic clam *Laternula elliptica*. *Oecologia*, 154(3), pp.479-484.

Sandison E.E. 1967. Respiratory responses to temperature and temperature tolerance of some inter-tidal gastropods. *Journal of Experimental Marine Biology and Ecology* 1: 271–281.

Watson, G.J., Bonner, A., Murray, J.M. and Hebblethwaite, Z., 2012. Offsetting the impact of the marine ornamental trade: a case study of two temperate top shells (*Osilinus lineatus* and *Gibbula umbilicalis*) as potential clean-up crew. *Aquatic Conservation: Marine and Freshwater Ecosystems*, 22(6), pp.731-742.

Brey, T., Pearse, J., Basch, L., McClintock, J., & Slattery, M. (1995). Growth and production of *Sterechinus neumayeri* (Echinoidea: Echinodermata) in McMurdo Sound, Antarctica. *Marine Biology*, 124(2), 279–292. <https://doi.org/10.1007/BF00347132>

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1 Article to Marine Ecology Press Series

2

3 **Functional thermal limits are determined by rate of**
4 **warming during simulated marine heatwaves**

5

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13 Running head: Functional thermal limits during marine heatwaves

1 **ABSTRACT**

2 Marine heatwaves (MHWs) are increasing in both intensity and frequency
3 against a backdrop of gradual warming associated with climate change. In the
4 context of MHWs, animals are likely to experience sub-lethal, rather than lethal
5 effects, defining long-term limits to survival and/or impacting individual and
6 population fitness. This study investigated how functional sub-lethal limits track
7 critical thresholds and how this relationship changes with warming rate. To this
8 end we monitored basic functioning, specifically the ability to right, feed and
9 assimilate energy, as well as oxygen consumption rate in the common Antarctic
10 sea urchin, *Sterechinus neumayeri*. Water temperature in experimental
11 systems was increased at rates of 1°C day⁻¹, 0.5°C day⁻¹ and 0.3°C day⁻¹, in
12 line with the characteristics of MHW events previously experienced at the site
13 where the study urchins were collected on the Antarctica Peninsula.
14 Functioning was assessed during the simulation of MHWs and sub-lethal limits
15 determined when the rate of functional degradation changed as temperature
16 increased. Results suggest that thermal sensitivity varies between the key
17 biological functions measured, with the ability to right having the highest
18 thermal threshold. Arguably, the most interesting result was that functions
19 deteriorated at lower temperatures when warming was more rapid (1°C day⁻¹),
20 contrary to lethal critical thresholds, which were reached at lower temperatures
21 when warming was slower (0.3°C day⁻¹). MHWs and their impacts extend far
22 beyond Antarctica and in this context, our analyses indicate that the onset rate
23 of MHWs is critical in determining an organism's ability to tolerate short-term
24 elevated temperatures.

1 Key words: Extreme warming events, sub-lethal limits, thermal tolerance,
2 climate change, polar, segmented regression, echinoderm

3 **1. INTRODUCTION**

4 Historical temperature records have now detected positive temperature trends
5 for the majority of the Earth's surface (Myrvoll-Nilsen et al. 2019), with the
6 oceans being key to the regulation and capture of much of the excess heat
7 present in the atmosphere (Marshall et al. 2015). As a result, marine
8 environments are changing both physically and biochemically (Bopp et al.
9 2013). Included in these changes is the occurrence of marine heat waves
10 (MHWs), which are increasing in duration, magnitude and frequency, with
11 alarming ecological consequences (Garrabou et al. 2009, Rubio-Portillo et al.
12 2016, Oliver et al. 2018).

13 Physiological flexibility of species is crucial to survival during MHW events
14 (Peck 2011) and species at low latitudes may be able to acclimate and adapt
15 across generations to altered environments (Donelson et al. 2012, Salinas &
16 Munch 2012, Clark et al. 2019a). As a result, predicting effects of MHWs on
17 lower latitude species may need to consider shifting thermal ranges as these
18 species adapt to climate change. It is unlikely that the same will apply to
19 Antarctic species, since many are physiologically limited by their capacity to
20 acclimate and adapt to new temperatures because of their long generation
21 times and delayed reproductive maturity (Peck et al. 2014, Peck 2018). For
22 example, several invertebrate species such as the Antarctic scallop
23 *Adamussium colbecki*, the limpet *Nacella concinna*, and the bivalves, *Laternula*
24 *elliptica* and *Adacnarca nitens*, take 4 – 7 years to mature. The Antarctic
25 bivalve, *Aequiyoldia eightsi*, starts reproducing at around 12 years (Peck &

1 Bullough 1993) and the brachiopod *Liothyrella uva*, can take up to 18 years
2 before brooding young (Peck 2005, 2018, Oliver et al. 2019).

3 Predicting species and ecosystem responses to MHWs is challenging, owed to
4 the past infrequency and variability of each event (Oliver et al. 2018). However,
5 if we can track the functional deterioration of organisms when temperatures
6 exceed their typical thermal range, this can inform our understanding of the
7 relationships between the sub-lethal and lethal limits likely to be encountered
8 during MHW events.

9 For organisms with slow growth and development and long generation times,
10 like many of those found in Antarctica, thermal stress caused by MHWs is likely
11 to trigger other mechanisms for survival such as biochemical and cellular stress
12 responses (e.g. Clark & Peck 2009, Payton et al. 2016). Biochemical and
13 genetic mechanisms, including a range of chaperone proteins, provide a short-
14 term buffer that allow functioning to continue temporarily at temperatures
15 outside an organism's thermal niche (Deschaseaux et al. 2010, Clark et al.
16 2019b). Once animals are no longer able to maintain basic functions by these
17 mechanisms, the sub-lethal limit to survival is reached.

18 Data on the functional thermal limits of species and MHW characteristics (i.e.
19 rate, magnitude and duration) at which these thresholds are reached are rare,
20 especially in fluctuating environments (Janecki et al. 2010, Peck et al. 2014,
21 Ardor Bellucci & Smith 2019). Little is known about functional deterioration as
22 a species approaches its critical thermal limit, and in the context of MHWs,
23 animals are likely to experience temperatures that cause sub-lethal, rather than
24 lethal effects, defining long-term limits to survival and/or inhibiting population
25 health (Pörtner et al. 2007).

1 This study aims to understand how functional (sub-lethal) limits track critical
2 (lethal) limits and how this relationship changes with warming rate during a
3 simulated MHW. To this purpose, we monitored the ability to right, feed and
4 assimilate energy as well as oxygen consumption rate, in the common Antarctic
5 sea urchin, *Sterechinus neumayeri*.

6 **2. MATERIALS AND METHODS**

7 2.1 Sample site and animal collections

8 *Sterechinus neumayeri* were sampled from South Cove, Rothera Point
9 (67°34'09.1"S 68°07'52.7"W), from sites near the British Antarctic Survey's
10 Rothera Research Station on the Western Antarctic Peninsula (WAP) during
11 December 2019 (Figure S1). 120 adult urchins (test diameter range, 28 mm –
12 49 mm) were SCUBA-diver collected at depths of 10-20 m and returned to the
13 Rothera aquarium facility within two hours of collection.

14 *Sterechinus neumayeri* is one of the most common and locally abundant
15 members of the Antarctic marine shallow benthos, forming a significant
16 component of the benthic community (Brockington 2001, Pierrat et al. 2012),
17 with reported densities up to 600 m² (Barnes & Brockington 2003). It is a major
18 scavenger of dead organisms and in iceberg scours on the shallow Antarctic
19 seabed (Dunlop et al. 2014), and is a significant grazer and bioturbator of
20 sediments (Lenihan et al. 2018). Because of this *S. neumayeri* is an important
21 carbon transformer in Antarctic shallow seas. Further to this, due to its
22 abundance and ease of maintenance in laboratory culture systems, *S.*
23 *neumayeri* has been the subject of extensive study of its embryonic and larval
24 development, which is highly extended, and up to in excess of 100 days (Bosch
25 et al. 1987). It has also been the subject of studies of the effects of temperature

1 on embryonic and larval development (Stanwell-Smith & Peck 1998), the
2 impact of ocean acidification on reproduction (Suckling et al. 2014) and energy
3 budgets (Morley et al. 2016). Furthermore, it has been shown that there are
4 long-term cycles in its reproduction (De Leij et al. 2021). These factors all make
5 *S. neumayeri* one of the most important members of the Antarctic shallow
6 benthic ecosystem and key to investigating responses to MHWs.

7 2.2 Experimental set-up and warming system

8 A decade of temperature data (1997-2017) from Ryder Bay on the WAP
9 (sourced from the Rothera Time-Series (RaTS) environmental monitoring
10 programme (Clarke et al. 2008, Venables et al. 2013)) was used in the R
11 package “heatwaveR” (Schlegel & Smit 2018), to detect past warming events
12 (Figure 1) (see details of warming event analysis methodology and
13 characteristics summary in the Supplementary Materials, Text S1, Table S1,
14 Figure S2). Studying the characteristics of these past warming events, including
15 onset rate and magnitude, allowed us to set realistic warming rates for the
16 experimental systems.

17 Urchins were held in flow-through aquaria (170 L) at ambient temperatures
18 typical for December and January (-1.5°C to +0.5°C) for six weeks on a
19 continuous light regime. During this time, animals were not fed to allow any
20 ingested food to be processed and the production of faeces to cease. The
21 cessation of faeces production is an indicator that metabolic rates had reached
22 a “standard” level at the start of the experiment. Previous research suggests
23 that these urchins are able to sustain and experience natural periods of
24 starvation for up six months during winter (Brockington 2001), and hence six

1 weeks without feeding was unlikely to be detrimental to the physiological
2 metrics measured in this study. Previous studies of oxygen consumption in
3 Antarctic marine invertebrates has demonstrated that standard levels are
4 reached in less, and often significantly less, than this time in the brachiopod
5 *Liothyrella uva* and the limpet *Nacella concinna* (Peck 1989), in the amphipod
6 *Waldeckia obesa* (Chapelle et al. 1994), in the isopod *Glyptonotus antarcticus*
7 (Robertson et al. 2001), and in the sea star *Odontaster Validus* (Peck et al.
8 2008).

9 After urchins were maintained in the flow-through aquarium (170 L) at ambient
10 temperatures, 30 urchins were distributed to four main aquarium tanks to
11 represent each warming treatment as well as the ambient control treatment.
12 Urchins were distributed at random. Replication within each of these treatments
13 was achieved by floating five separate 6-litre tanks, each containing six urchins
14 in each main aquarium tank (170 L). Each main aquarium tank functioned as a
15 temperature bath (Figure S3; 30 urchins per treatment, 5 replicates per
16 treatment where data from urchins in the same replicate floating tank were
17 pooled). Temperature treatments were not replicated due to space restrictions.
18 The same treatment conditions (i.e., temperature) was translated to all replicate
19 urchins, and as such, temperature was closely monitored to note and control
20 variability (Figure S4).

21 The water in each floating tank was aerated using air stones and refreshed by
22 50% water change every other day. Water changes not only ensured that
23 overall water quality was maintained, but also meant any metabolic products,
24 especially potentially toxic nitrogenous chemical species, were maintained at
25 very low levels. Tank water samples were periodically analysed for pH (ranging

1 7.5 - 8.0), NO₂ (ranging 0.05 mg l⁻¹ – 0.1 mg l⁻¹), NO₃ (ranging 0.5 mg l⁻¹ - 1.0
2 mg l⁻¹) and NH₄ (stable at 0.1 mg l⁻¹) to ensure good water quality. Throughout
3 the experiment, concentrations of the aforementioned compounds remained
4 within the ranges stated.

5 Urchins within each replicate tank were separated by aquaria egg crates and
6 fine mesh partitions to ensure individuals were isolated and any faeces
7 produced was retained within compartments (Figure S3). During warming trials
8 experimental temperatures in the aquaria water baths were raised by 1°C,
9 0.5°C or 0.3°C each evening, depending on treatment. Temperatures in the
10 floating tanks increased more gradually than the water baths, allowing urchins
11 to adjust slowly to each new temperature. Temperatures were checked every
12 30 minutes after each temperature change to ensure required temperatures
13 were achieved and kept constant. Initially, temperatures fluctuated by up to ±
14 0.3°C before stabilising after 1-2 hrs. Temperatures were subsequently
15 monitored throughout the following day and held within ± 0.1°C of the target
16 experimental temperature (Figure S4). For ambient controls, urchins were held
17 in the aquarium with the set-up and light conditions identical to the warming
18 treatment conditions. Temperatures were maintained at those experienced in
19 Ryder Bay which naturally fluctuated between 0.9 °C and 1.9°C.

20 2.3 Feeding trials

21 Urchins were fed pre-portioned amounts of food every 48 hrs. Previous studies
22 fed *S. neumayeri* high protein diets, such as fish fillets, *Polachius virens*
23 (Suckling et al. 2014, Morley et al. 2016). In the current study, urchins were fed
24 the foot of the common Antarctic limpet, *Nacella concinna*, which has a
25 comparable protein content to that of *P. virens* muscle. Based on feeding

1 protocols in Morley et al. (2016b), urchins were fed ~4% of their mean body
2 mass every three weeks, but this was spread across 48 hr feeding increments
3 in order to keep feeding activity constant and reduce the variability in daily
4 metabolic activity.

5 Limpets were chosen as a food source since nutrient content could be
6 controlled and pre-portioned. A more representative diet would be a varied one
7 with algal biofilm, animal tissues and/ or detritus (McClintock 1994). However,
8 administering a varied diet would make it difficult to assess the amount of food
9 consumed per urchin at the same time as standardising the nutritional content.

10 There is evidence that diet, especially protein levels, can affect development
11 and gonad growth (Liu et al. 2007, Zupo et al. 2019) as well as ingestion and
12 assimilation rates in sea urchins (Azad et al. 2011). As such, by feeding a diet
13 of limpets it is possible that body condition may be altered and the ability to
14 tolerate stress may be improved as a result.

15 Feeding was initiated two days before the beginning of the experiment to start
16 the digestion process. Each urchin was allowed to feed for 48 hrs before any
17 remaining food was removed and refreshed. After 48 hrs, each urchin was
18 recorded as feeding or not feeding. Infrequently, urchins may have only partially
19 consumed the food piece, which was recorded.

20 2.4 Faecal collection

21 Faecal production began four days into the experiment, 6-days after feeding
22 was initiated. The presence of faeces was recorded for all urchins every 48 hrs.
23 To measure faecal production, faeces were collected every 48 hrs by pipette
24 and transferred to falcon tubes from 10 urchins per treatment, where at least

1 one sample was taken from each replicate tank within the treatment. The same
2 urchins were targeted for faecal collection to minimise subconscious
3 preferences towards urchins producing more faeces. This was not always
4 possible since sometimes urchins did not produce any faeces or else CT_{max}
5 was reached, and these urchins were removed. In these cases, a different
6 urchin was chosen at random to sample from. For all other urchins, any
7 remaining faecal matter was removed.

8 Collected faecal matter was centrifuged and the supernatant seawater
9 decanted. Faeces were then rinsed with RO (Reverse Osmosis purified) water
10 by agitating and centrifuging to remove any seawater salt. Washed faeces were
11 pipetted into pre-ashed and pre-weighed foil boats and dried at 60°C for 24 hrs.
12 Dry foil boats and faeces were placed in a desiccator to cool and then weighed
13 (± 1 mg). Dry faeces were subsequently ignited in a muffle furnace at 475°C for
14 6 hrs. Foil boats and ashed faeces were cooled in a desiccator and weighed (\pm
15 1 mg). Dry mass (DM) and Ash-Free Dry Mass (AFDM) (i.e., organic content)
16 were obtained by subtraction.

17 2.5 Respirometry

18 Oxygen consumption was recorded for 10 urchins per treatment, sampling two
19 individuals from each replicate tank within each treatment. Oxygen
20 consumption was recorded for the same urchins for every 2°C rise in
21 temperature from ambient in each treatment. Methods for measuring oxygen
22 consumption followed those described by Suckling et al., (2015), using 200 -
23 250 ml volume chambers. For each urchin, live wet mass (± 0.01 g) was
24 recorded where O_2 consumption was measured. AFDM was determined from
25 live wet mass vs AFDM regressions determined from a subsample of urchins

1 (n = 40) collected from the same site. To obtain the ash mass of urchins,
2 individuals were weighed live before freezing in liquid nitrogen and storing at –
3 40°C. Frozen urchins were then placed in pre-ashed and pre-weighed ceramic
4 crucibles and dried at 60°C until constant mass was obtained (± 0.01 g). Once
5 dried, urchins were ignited in a muffle furnace at 475°C for 6 hrs and
6 subsequently weighed to obtain ash mass after cooling in a desiccator (± 1 mg).

7 2.6 Righting

8 The time taken for urchins to right themselves was recorded for 10 urchins per
9 treatment, sampling two urchins from each replicate tank within each treatment.
10 The time taken to right was recorded for the same urchins every 2°C rise in
11 temperature from ambient in each treatment. Ten individuals were removed
12 from their experimental tanks and placed in individual containers. These
13 containers were previously filled and floated in water already at the
14 experimental target temperature. Urchins were immediately inverted following
15 transfer from experimental tanks to the floating containers and timed until the
16 individual was fully upright. Urchins could not reach the sides of containers to
17 aid in righting. Once righted, urchins were returned to their experimental tanks.

18 2.7 Critical temperature limits (CT_{max})

19 The critical thermal limit (CT_{max}) was recorded for all experimental urchins in
20 the warming treatments, where the limit was defined as the point at which the
21 individual was unable to right itself within 12 hrs, had stopped eating and
22 producing faeces. When an urchin began to show signs of reaching the CT_{max}
23 (not feeding or producing faeces), they were inverted in the tank and left for 12

1 hrs. If the urchin had not righted itself after this period, they were removed and
2 weighed suspended in water to obtain live wet volumes (± 0.01 mL).

3 2.8 Statistical Analysis

4 Where multiple urchins were sampled within the same floating tank,
5 measurements of feeding, faecal production, righting, and oxygen consumption
6 were pooled so that $n = 5$, and the standard errors were calculated from these
7 five replicate tanks.

8 To determine differences in functional responses between treatments, a one-
9 way repeat measures analysis of variance (ANOVA) was carried out in R (v.
10 4.0.5). This analysis was considered appropriate for this experiment due to the
11 related and non-independent groups at each temperature timepoint. For this
12 analysis, treatment group variances were compared when treatments reached
13 the same temperature increments. For ambient controls, temperature
14 timepoints were aligned with measurements taken at similar dates to treatment
15 sampling. Variances were compared between groups and within timepoints for
16 righting and oxygen consumption rates and the resultant p-value was adjusted
17 using the Bonferroni correction method. Significant differences ($p < 0.05$) were
18 followed up with a paired t-test and again, p-values were adjusted using the
19 Bonferroni correction method. Data were initially log transformed to ensure
20 assumptions of normal distribution were met.

21 Segmented linear regression models were fitted in the R package 'segmented'
22 (Muggeo 2008) to identify breakpoints in the linear relationships between
23 functional process and temperature. Breakpoints were identified where the
24 gradient of the relationship changed (McWhorter et al. 2018). The change in

1 gradient was used to define the functional threshold of the process measured.
2 It was especially important to use a method such as segmented regression to
3 identify breakpoints in process rates. Segmented regressions were used to
4 model these relationships not necessarily for the purpose of fitting the simplest
5 model, but rather to identify any change in the regressions gradient which then
6 indicated that the functions response to temperature increase had changed. In
7 some cases, a linear regression would be sufficient to explain the relationship,
8 however a linear model could mask the subtle change in the rate of degradation
9 experienced when a species hits a thermal threshold. Alternatives would be to
10 fit curves and identify changes in slope (e.g. Pörtner et al. 2006), but curves
11 were not appropriate here. A Davies test was also conducted to determine
12 significant ($p < 0.05$) differences in the gradients of the segmented slopes.

13 Size effects on functional response were explored through scatter plots. Where
14 relationships were observed, the effect of size (as test diameter) and
15 temperature on the functional response, was assessed with a linear mixed
16 effects model using the package 'lme4' and the function 'lmer' in R (v. 4.0.5).
17 Test diameter and temperature were added as interacting fixed terms and
18 replicate tank ID was added as a random effect. Prior to any modelling, function
19 responses were transformed to achieve normality in the distribution.

20 **3. RESULTS**

21 3.1 Feeding and faecal egestion

22 On average, $80\% \pm 19\%$ of animals fed in ambient conditions for the duration
23 of the experiment. For the first four days of the experiment, in treatments where
24 $T \uparrow 1^\circ\text{C day}^{-1}$, the proportion of animals feeding exceeded all other treatments
25 ($97\% \pm 4\%$), including ambient conditions ($87\% \pm 10\%$). Fifty percent of animals

1 stopped feeding in treatments when temperatures exceeded 7.2°C, 8.2°C, and
2 9.2°C, where T↑ by 1°C, 0.5°C and 0.3°C day⁻¹, respectively (Figure 1).

3 A breakpoint (where the slope of the regression changed) for the % individuals
4 feeding was identified at 4.0°C and 6.2°C in treatments where T↑ 1°C day⁻¹
5 and 0.5°C day⁻¹, respectively (Table 1). However, changes in the segmented
6 slope gradients were not significantly different from linear regressions for these
7 two treatments (Davies p-value = 0.329 and 0.301, respectively). A breakpoint
8 for the % feeding in T↑ 0.3°C day⁻¹ was identified at 8.2°C (Table 1), from which
9 point the % individuals feeding declined rapidly and the relationship between
10 temperature and the proportion of individuals feeding became significant (p
11 <0.001). The mean temperature breakpoint for the function of % feeding was
12 6.1°C ± 1.2°C, averaged across all treatments.

13 The percentage of animals producing faeces tracked the proportion of animals
14 feeding after the first four days (Figure 1). Following each breakpoint, the
15 relationship between temperature and % individuals producing faeces became
16 significant (Table 1). For the fastest rate of warming where T↑ 1°C day⁻¹, a
17 breakpoint was identified at 5.2°C, above which the % individuals producing
18 faeces rapidly declined from 100% to 10.3% within 6 days. Where T↑ 0.3°C
19 day⁻¹ and 0.5°C day⁻¹, the regression breakpoint for faecal production was
20 8.3°C and 4.5°C respectively (Table 1). The mean temperature breakpoint for
21 the function of % producing faeces was 6.0°C ± 2.0°C, averaged across all
22 treatments.

23 The mean mass of faeces produced in treatments where T↑ 0.3°C day⁻¹, was
24 significantly greater than the faecal mass produced in ambient control

1 conditions and treatments where $T \uparrow 1^\circ \text{C day}^{-1}$, until temperatures exceeded
2 2.1°C ($t_{(4)} = 8.74$, $p = 0.006$ and $t_{(4)} = 5.02$, $p = 0.044$, respectively). Where $T \uparrow$
3 $0.5^\circ \text{C day}^{-1}$, the mass of faeces produced was significantly greater than
4 treatments where $T \uparrow 1^\circ \text{C day}^{-1}$, until temperatures exceeded 2.1°C ($t_{(4)} = 5.31$,
5 $p = 0.036$). Despite this observation, no additional food was consumed in these
6 treatments. There was no significant difference between the treatments or
7 control as temperatures increased beyond 2.1°C .

8 Breakpoints in regressions were identified at 5.0°C and 3.1°C for treatments
9 where $T \uparrow 0.5^\circ \text{C day}^{-1}$ and $0.3^\circ \text{C day}^{-1}$, respectively (Table 1). The breakpoints
10 for these regressions marked a reduction in the gradient of the 2nd slope,
11 whereby faeces produced $\text{day}^{-1} \text{ mgAFDM}^{-1}$ as a function of temperature
12 decreased at a slower rate as temperatures increased. The mean temperature
13 breakpoint for faeces produced was $4.1^\circ \text{C} \pm 0.95^\circ \text{C}$, averaged across the
14 slowest ($T \uparrow 0.3^\circ \text{C day}^{-1}$) and intermediate ($T \uparrow 0.5^\circ \text{C day}^{-1}$) rates of warming.

15 3.2 Righting

16 In treatments where $T \uparrow 1.0^\circ \text{C day}^{-1}$, time taken to right became significantly
17 longer than ambient controls when temperatures reached 9.2°C ($t_{(4)} = 6.06$, $p <$
18 0.022). For treatments where $T \uparrow 0.3^\circ \text{C day}^{-1}$, time taken to right only became
19 significantly longer than ambient controls just before CT_{max} was reached, when
20 temperatures reached 11.2°C ($t_{(4)} = 6.04$, $p < 0.023$). For treatments where $T \uparrow$
21 $0.5^\circ \text{C day}^{-1}$, time taken to right never exceeded ambient controls significantly,
22 however mean righting times were consistently higher than control conditions
23 throughout the warming period.

1 A breakpoint in the linear regression was identified at 8.7°C in treatments where
2 temperature was raised at 0.3°C day⁻¹ (Table 1). The relationship between
3 temperature and the time taken to right became significant above this
4 breakpoint temperature ($p < 0.001$). For the other treatments righting time
5 increased linearly without a breakpoint in the regression.

6 The interactive effect of urchin size and temperature on the time taken to right
7 was significant ($t_{(204)} = 2.11$, $p = 0.034$), where larger urchins took longer to right
8 at higher temperatures (Figure S5, Table S3).

9 3.3 Oxygen consumption

10 Oxygen consumption rates were significantly higher in heatwave treatments
11 compared to ambient controls when temperatures reached 7.2°C for all
12 treatments. However, oxygen consumption rates were significantly higher than
13 ambient controls from lower temperatures of 3.2°C in treatments where $T \uparrow$
14 0.3°C day⁻¹ ($t_{(4)} = 5.62$, $p = 0.030$) and 5.2°C in treatments where $T \uparrow$ 1.0°C
15 day⁻¹ ($t_{(4)} = 4.98$, $p = 0.045$). Overall, there was a positive linear trend between
16 oxygen consumption and temperature for all treatments. However, where $T \uparrow$
17 1°C day⁻¹, a drop in O₂ consumption occurred at 9.2°C, and where $T \uparrow$ 0.3°C
18 day⁻¹, a drop occurred just before the CT_{max} at 11.2°C.

19 O₂ consumption increased at a faster rate per increase in temperature where
20 warming rates were fastest at 1°C day⁻¹ (slope gradient = 1.50) and increased
21 at the slowest rate when warming rates were slowest at 0.3°C day⁻¹ (slope
22 gradient = 0.96) (Table 1). No breakpoint was identified in any treatment.

23 3.4 CT_{max}

1 The CT_{max} for urchins in treatments where $T \uparrow 0.3^\circ\text{C day}^{-1}$, $T \uparrow 0.5^\circ\text{C day}^{-1}$ and
2 $T \uparrow 1^\circ\text{C day}^{-1}$ ranged from $10.6^\circ\text{C} - 13.8^\circ\text{C}$, $11.2^\circ\text{C} - 13.7^\circ\text{C}$, and $12.2^\circ\text{C} -$
3 14.2°C , respectively. The effect of warming rate on the CT_{max} was significant
4 ($F_{(2, 12)} = 7.29$, $p = 0.008$), with post-hoc analysis identifying that for treatments
5 where temperature increased at the fastest rate ($T \uparrow 1^\circ\text{C day}^{-1}$), the CT_{max} was
6 significantly higher compared to treatments where temperature increased at a
7 slower rate ($T \uparrow 0.3^\circ\text{C day}^{-1}$) ($t_{(8)} = -6.02$, $p = 0.001$).

8 Across all functions where breakpoints were identified, the slowest rate of
9 warming ($T \uparrow 0.3^\circ\text{C day}^{-1}$) had a mean temperature breakpoint of $8.3^\circ\text{C} \pm 1.3^\circ\text{C}$.
10 In comparison, the mean temperature breakpoint was $5.4^\circ\text{C} \pm 0.5^\circ\text{C}$, and 4.6°C
11 $\pm 0.6^\circ\text{C}$ for intermediate ($T \uparrow 0.5^\circ\text{C day}^{-1}$) and fast ($T \uparrow 1^\circ\text{C day}^{-1}$) warming
12 rates, respectively.

13 **4. DISCUSSION**

14 MHWs are predicted to increase in frequency, intensity, and duration in the
15 coming decades. Deterioration of basic animal functioning, critical for long-term
16 survival, will likely be a more frequent consequence of the short-term warming
17 (i.e., weeks-months) caused by MHWs, rather than mortality. However, little is
18 known about functional impacts, especially thresholds and how these limits
19 deteriorate with respect to CT_{max} . By understanding how key biological
20 functions are affected by short term temperature elevations and different
21 warming rates, we can better understand how extreme climate events, typified
22 by short-term warming, may impact individuals and populations, and hence
23 communities.

1 In this study, we investigated the effect of warming rates typical of those
2 expected during Antarctic MHW events on the functioning of the Antarctic sea
3 urchin, *S. neumayeri*. Functional thresholds were identified using segmented
4 regressions, where a breakpoint indicated a gradient change in the response
5 trend with temperature. The identification of regression breakpoints, or slope
6 changes has been used previously to define ecological thresholds, and is
7 considered a more flexible and realistic approach when interpreting complex,
8 often non-linear, ecological relationships (Piepho & Ogutu 2003, Ferrarini 2011,
9 Morley et al. 2014).

10 Several studies have shown that faster warming rates result in higher CT_{max} in
11 terrestrial (e.g. Terblanche et al. 2007, Allen et al. 2016) and marine (Peck et
12 al. 2009) species. These observations, along with the CT_{max} data in this study,
13 follow the failure rate model proposed by Kingsolver & Umbanhowar (2018),
14 who showed that critical limits are reached at lower temperatures when
15 warming accumulates over extended periods. However, our results for
16 functional thermal limits follow the opposite trend to the CT_{max} , where functions
17 are impacted negatively at lower temperatures when warming is rapid. Overall,
18 in this study higher functional thresholds were reached when temperatures
19 were raised slowly (thresholds averaging $8.3^{\circ}C \pm 1.3^{\circ}C$). At the faster warming
20 rates functional thresholds were lower ($5.4^{\circ}C \pm 0.5^{\circ}C$ or $4.6^{\circ}C \pm 0.6^{\circ}C$). There
21 was even evidence that some functions declined linearly, with significant
22 deterioration from temperatures $+2.8^{\circ}C$ above ambient when warmed at the
23 fastest rate. Thus, short-term exposure to more extreme temperatures has
24 more impact on functioning than longer, chronic exposure to more slowly
25 elevated temperatures.

1 Although metabolic acclimation is unlikely over such short time periods
2 (apparent from the oxygen consumption data here, and also previous research
3 on long-term acclimation of *S. neumayeri* (Peck et al. 2014, Suckling et al.
4 2015)), short-term acclimation for some functions might be possible after an
5 initial shock response when temperatures are increased slowly. In our study,
6 the shock response did not appear to subside at faster rates of warming, and
7 instead mean functional thresholds were lower as warming rate increased.
8 These results suggest that functional and lethal limits are likely driven and
9 determined by different mechanisms. Previous studies have shown that lethal
10 limits are likely set by one or both of physiological processes or cellular and
11 biochemical mechanisms. At very rapid rates of warming, such as 1°C h^{-1} or
12 $1^{\circ}\text{C day}^{-1}$, physiological mechanisms such as nervous and circulatory failure
13 appear to be the limiting factors (Young et al. 2006, Pörtner et al. 2007, Bilyk &
14 DeVries 2011). At slower rates of warming ($1^{\circ}\text{C 3 days}^{-1}$ to $1^{\circ}\text{C month}^{-1}$) cellular
15 and biochemical mechanisms such as accumulation of toxic products, e.g.
16 protein carbonyls, enzyme tolerances or insufficiency of chaperone protein
17 capacity appear to be limiting (Peck et al. 2009, Clark et al. 2017, 2018).
18 Recently the factors setting thermal limits and responses to warming have been
19 shown to be highly species specific (Clark et al. 2021, Collins et al. 2021).

20 Our results also indicate that thermal sensitivity varies among key biological
21 functions. For example, the function of righting in urchins was similar between
22 treatments and ambient control conditions until temperatures reached 9.2°C for
23 the fastest rates of warming, and the highest breakpoint of 8.7°C was identified
24 in the slowest rates of warming. However, lower thresholds were identified for
25 the other functions related to digestion such as % feeding or producing faeces.

1 Variation between functional thresholds could be related to function complexity,
2 where a function involving multiple processes would be more likely to fail
3 (Pörtner et al. 2007, Stevens et al. 2010, Peck 2011). Another explanation could
4 be related to the extent to which functions limit survival and fitness, where an
5 organism's energy reserves allow for short periods of negative energy balance.
6 In Antarctic marine species such periods of negative energy balance can be
7 very long, extending to months or even years of low food supply or starvation,
8 because of the extreme environmental seasonality and the very low metabolic
9 energy use characteristic of this fauna (Brockington et al. 2001, Harper & Peck
10 2003, Obermüller et al. 2010). However, being able to right provides immediate
11 protection from predation, equivalent to mechanisms such as the ability to stay
12 attached to the substratum in limpets (Morley et al. 2012b) or reburying in
13 infaunal clams when disturbed and removed from the sediment by, for example,
14 iceberg scour (Peck et al. 2004). Finally, where a function has a higher
15 metabolic energy demand, it is more likely to be limited by food availability and
16 energy delivery capacity (van der Meer 2006, Morley et al. 2012a, Peck 2018).
17 The breakpoints identified for the mass of faeces produced might not indicate
18 a functional threshold. Instead, the initial high faecal production in the slowest
19 and intermediate warming rates is likely a result of the initial increase in
20 temperature causing food to move faster through the urchin, as also seen in
21 the Antarctic plunderfish *Harpagifer antarcticus* (Boyce et al. 2000). This
22 elevation in faecal production was only observed when temperatures increased
23 initially, after which faecal production reduced to rates similar to ambient control
24 conditions. This effect was not observed in treatments with the fastest rates of
25 warming since these slight increases in temperature of 1°C – 2°C were likely

1 not maintained long enough for gut passage rate to increase. Therefore, our
2 results indicate that the breakpoints for faecal production may not have any
3 direct implications on functionality and instead give evidence for the relationship
4 between temperature and gut evacuation rate (GER).

5 In thermally stressed environments, animals usually increase their oxygen
6 uptake in order to meet increasing demands of functional processes (Gillooly
7 et al. 2001). However, when oxygen uptake is increased, yet functioning
8 deteriorates, it is hypothesised that this indicates a threshold where uptake,
9 transport, and delivery of oxygen can no longer meet the animal's functional
10 demands. This theory has been termed the oxygen and capacity limited thermal
11 tolerance hypothesis (OCLTT) (Pörtner et al. 2017). This theory focuses on the
12 limitations set by the animal's physiology. However, as temperature increases
13 the concentration of oxygen diminishes, further reducing the availability of
14 oxygen to the animal and potentially amplifying the effects of OCLTT. Reducing
15 the concentration of oxygen in the water can limit functioning (Peck et al. 2007,
16 Pörtner et al. 2007) and as such, the functional thresholds identified in this study
17 may not only indicate thermal limits but may also be influenced by the reduced
18 oxygen content as temperatures increased. If oxygen concentration was
19 controlled and elevated throughout warming, the functional thresholds identified
20 would likely be higher (Pörtner et al. 2006). However, warmer oceans will be
21 accompanied by lower oxygen concentrations (Oschlies et al. 2018, Spicer et
22 al. 2019) and as such the functional thresholds determined in this study will be
23 more representative of a natural system than if oxygen were controlled.

24 Food availability and quality can also be a significant factor in determining
25 functional scope (Welch et al. 1998, Lemoine & Burkepille 2012, Cheng et al.

1 2018), whereby the nutritional status and condition of the animal could affect
2 energy delivery capacity similarly to OCLTT. For example, feeding and
3 digestive capacity limited the thermal tolerance of juvenile spiny lobsters,
4 *Sagmariasus verreauxi* (Fitzgibbon et al. 2017) and digestive capacity and food
5 intake of individuals at high temperatures related to depressed mitochondrial
6 respiratory capacity in brown trout *Salmo trutta* (Salin et al. 2016). The capacity
7 to assimilate energy would also play a role in determining energy delivery to
8 tissues and is determined by physiological processes including consumption
9 rate, absorption of food and GER (Boyce et al. 2000, Angilletta 2001). Hence,
10 assimilation itself is energetically demanding and may limit functional thermal
11 thresholds (Sandersfeld et al. 2015, Salin et al. 2016).

12 Thus, OCLTT may be a possible mechanism for determining functional limits
13 observed in our experiments. However, there is no empirical support in our data
14 for this theory. In both experiments and in natural MHWs, other factors are likely
15 to be important, and obtaining sufficient energy from food may be important for
16 successful functioning. Impacts on animal condition from warming may be
17 especially important in highly seasonal polar environments where warming in
18 winter, when food supplies are scarce, would increase energy use with little or
19 no opportunity to mitigate the cost (Peck 2018). Species such as *S. neumayeri*
20 that have been shown to spend periods in winter up to seven months without
21 feeding (Brockington 2001) may be particularly vulnerable to such impacts.

22 Our experiment included a period of six weeks without feeding to allow
23 metabolic activity to stabilise and be comparable between individuals.
24 However, a caveat to this initial standardisation of condition could influence the
25 urchin's physiological response to the warming in treatments. Nutritional status

1 has been shown to affect the reproductive state of *S. neumayeri*, with a
2 reduction in gonad index and maturation of gametes following six weeks without
3 food, comparative to animals foraging naturally in the environment (De Leij
4 2021). Functional capacity has also been affected in other invertebrates under
5 low food coupled with environmental stress, for example the blue mussel
6 *Mytilus edulis* had a reduced ability to repair shells when high CO₂ was coupled
7 with low food (Melzner et al. 2011) and the green sea urchin *Strongylo-*
8 *centrotus droebachiensis*, exhibited severe metabolic acidosis when exposed
9 to elevated CO₂ with empty digestive tracts (Stumpp et al. 2012). Hence, we
10 might consider that the elevated temperatures coupled with the suboptimal
11 nutritional status at the start of the experiment, may have impacted the thermal
12 limits of certain functions. This would likely have resulted from a mismatch
13 between a limited energy supply and stores, and an increased energy demand
14 of the animal. However, the data in this study shows a reduction in the number
15 of urchins feeding as temperatures increase, suggesting that food was not the
16 limiting factor when this species approached its functional thermal limits.

17 From our analysis of the RaTS environmental data, previous MHW events
18 reached maximum temperatures of $2.3^{\circ}\text{C} \pm 0.36^{\circ}\text{C}$, with onset rates of 0.3°C
19 day^{-1} . Days at heatwave status have extended up to 95 days, and cumulative
20 intensities (a combination of temperature intensity and heatwave duration) have
21 reached maxima of $54^{\circ}\text{C} \times \text{day}$ (Figure S2). Mean climate temperatures are
22 predicted to shift by $+2^{\circ}\text{C}$ by 2100, and with that, climate extremes such as
23 MHWs will increase in magnitude relative to this (IPCC 2014, 2019). Our results
24 suggest that functions such as feeding and faecal egestion are likely to be
25 affected by MHW events occurring in 2100, if not before, and this will include

1 increased metabolic demands with consequent impacts on annual energy
2 budgets.

3 For a long-lived (>40 year (Brey et al. 1995)) and slow to mature (8-9 years
4 (Peck 2018)) species such as *S. neumayeri*, there will be less scope for
5 phenotypic and genotypic adaptations to a warming climate as might be
6 possible for short-lived and rapidly maturing species (Peck 2011, Donelson et
7 al. 2012, Salinas & Munch 2012). However, there may still be opportunity for *S.*
8 *neumayeri* to adapt to a warmer world. Within 80 years (2020 - 2100), eight
9 generations of *S. neumayeri* will have succeeded the present population, and
10 in the year 2100, the 5th, 6th and 7th generation could be present and
11 reproducing in populations around Antarctica. If we consider the evidence of *S.*
12 *neumayeri*'s capacity to acclimate, it may be possible for this species to
13 acclimate and adapt successfully to function in a +2°C warmer world (Morley et
14 al. 2016). It is still uncertain, however, how this species will respond to acute
15 warming, like that experienced during MHWs, in this warmer climate. The data
16 in this study cannot predict the implications of acclimation and adaptation on
17 the subsequent tolerance to MHWs for *S. neumayeri*. Instead, the data provides
18 insight into the effect of onset rate of acute warming, the thermal vulnerability
19 of key biological functions, and the difference between critical thermal limits and
20 functional thermal limits. Thus, according to our data we could see reduced
21 energy availability for *S. neumayeri* from changes in feeding and food
22 processing rates during MHWs in warmer oceans, which would very likely
23 reduce survival in marginal environments.

1 Following the results from this study, it would be important to explore recovery
2 following MHW events. Our data indicate reduced functioning as temperatures
3 are raised across all rates of warming. However, the ability and rate of *S.*
4 *neumayeri* to resume 'normal' functioning if returned to ambient temperatures
5 is uncertain. It has been shown that the marine snail, *Littorina littorea*, loses
6 motility under thermal stress, however if temperatures are lowered again, this
7 function returns (Hamby 1975). To resume a single function may not indicate
8 full recovery, and our study shows that different biological functions have
9 varying thermal tolerances. As such, performance of all functions, including
10 metabolic activity, would need to return to baseline levels for an animal to
11 recover completely (Walter et al. 2013). Developing our understanding of
12 recovery following acute warming and even the effects of repeat MHW events,
13 could better predict the long-term implications of MHWs for this species.

14 It is important to note that the functional and critical limits measured in this study
15 are likely an example of a 'best case scenario'. Experiments such as these can
16 only predict the isolated effects of one variable. However, the additional
17 energetic costs associated with physical factors such as salinity change and
18 biological factors including varying food quality and quantity, species
19 interactions, diseases and scavenging for food, need to be included before we
20 can obtain dependable predictions for 'real world' scenarios that give
21 information relevant to the variable conditions experienced across a species
22 distribution range. What is limiting at the range margins for a species will differ
23 from core areas (Kolzenburg et al. 2021).

24 Our data highlight that the deterioration of functioning when temperatures are
25 raised, especially during MHWs, has implications for long term survival, and

1 physiological functions. Therefore, functioning should be considered when
2 determining organism thermal limits, rather than traditional critical thermal
3 limits. Our findings show that fitness cannot be determined from a single
4 function and instead functions vary in thermal sensitivity. A whole organism
5 approach to functional fitness is therefore necessary, considering functional
6 complexity, importance, and energetic demand. Our results suggest that
7 contrary to the relationship between critical thermal limits and onset rate,
8 functional degradation occurs at lower temperatures when exposed to rapid
9 warming ($1^{\circ}\text{C day}^{-1}$). Therefore, when investigating the impact of MHWs on
10 organisms and populations, it is important to consider the key features of the
11 heatwave event, including the onset rate, exposure duration, and how these
12 characteristics act together to determine functional thresholds.

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22 Antarctic season.

23 **AUTHOR CONTRIBUTIONS**

1 L.S.P and R.D conceived and designed the study. R.D carried out the
2 practical work and data processing. R.D, L.J.G and L.S.P analysed the data,
3 drafted the manuscript and approved its publication.

4 **COMPETING INTERESTS**

5 The authors declare no competing interests.

6 **LITERATURE CITED**

- 7 Allen JL, Chown SL, Janion-Scheepers C, Clusella-Trullas S (2016)
8 Interactions between rates of temperature change and acclimation affect
9 latitudinal patterns of warming tolerance. *Conserv Physiol* 4:1–14.
- 10 Ardor Bellucci LM, Smith NF (2019) Crawling and righting behavior of the
11 subtropical sea star *Echinaster (Othilia) graminicola*: effects of elevated
12 temperature. *Mar Biol* 166:1–9.
- 13 Azad AK, Pearce CM, McKinley RS (2011) Effects of diet and temperature on
14 ingestion, absorption, assimilation, gonad yield, and gonad quality of the
15 purple sea urchin (*Strongylocentrotus purpuratus*). *Aquaculture* 317:187–
16 196.
- 17 Barnes DKA, Brockington S (2003) Zoobenthic biodiversity, biomass and
18 abundance at Adelaide Island, Antarctica. *Mar Ecol Prog Ser* 249:145–
19 155.
- 20 Bilyk KT, DeVries AL (2011) Heat tolerance and its plasticity in Antarctic
21 fishes. *Comp Biochem Physiol - A Mol Integr Physiol* 158:382–390.
- 22 Bopp L, Resplandy L, Orr JC, Doney SC, Dunne JP, Gehlen M, Halloran P,
23 Heinze C, Ilyina T, Séférian R, Tjiputra J, Vichi M (2013) Multiple
24 stressors of ocean ecosystems in the 21st century: Projections with
25 CMIP5 models. *Biogeosciences* 10:6225–6245.
- 26 Bosch I, Beauchamp KA, Steele ME, Pearse JS (1987) Development,
27 metamorphosis, and seasonal abundance of embryos and larvae of the
28 Antarctic sea urchin *Sterechinus neumayeri*. *Biol Bull* 173:126–135.

- 1 Boyce SJ, Murray AWA, Peck LS (2000) Digestion rate, gut passage time and
2 absorption efficiency in the Antarctic spiny plunderfish. *J Fish Biol*
3 57:908–929.
- 4 Brey T, Pearse J, Basch L, McClintock J, Slattery M (1995) Growth and
5 production of *Sterechinus neumayeri* (Echinoidea: Echinodermata) in
6 McMurdo Sound, Antarctica. *Mar Biol* 124:279–292.
- 7 Brockington S (2001) The seasonal ecology and physiology of *Sterechinus*
8 *neumayeri* (Echinodermata; Echinoidea) at Adelaide Island, Antarctica.
9 PhD thesis The Open University.
- 10 Brockington S, Clarke A, Chapman ALG (2001) Seasonality of feeding and
11 nutritional status during the austral winter in the Antarctic sea urchin
12 *Sterechinus neumayeri*. *Mar Biol* 139:127–138.
- 13 Chappelle G, Peck LS, Clarke A (1994) Effects of feeding and starvation on the
14 metabolic rate of the necrophagous Antarctic amphipod *Waldeckia obesa*
15 (Chevreux, 1905). *J Exp Mar Bio Ecol* 183:63–76.
- 16 Clark MS, Peck LS (2009) Triggers of the HSP70 stress response:
17 environmental responses and laboratory manipulation in an Antarctic
18 marine invertebrate (*Nacella concinna*). *Cell Stress Chaperones* 14:649–
19 660.
- 20 Clark MS, Peck LS, Thyrring J (2021) Resilience in Greenland intertidal
21 *Mytilus*: The hidden stress defense. *Sci Total Environ* 767:144366.
- 22 Clark MS, Sommer U, Sihra JK, Thorne MAS, Morley SA, King M, Viant MR,
23 Peck LS (2017) Biodiversity in marine invertebrate responses to acute
24 warming revealed by a comparative multi-omics approach. *Glob Chang*
25 *Biol* 23:318–330.
- 26 Clark MS, Suckling CC, Cavallo A, Mackenzie CL, Thorne MAS, Davies AJ,
27 Peck LS (2019a) Molecular mechanisms underpinning transgenerational
28 plasticity in the green sea urchin *Psammechinus miliaris*. *Sci Rep* 9:1–12.
- 29 Clark MS, Thorne MAS, King M, Hipperson H, Hoffman JI, Peck LS (2018)
30 Life in the intertidal: Cellular responses, methylation and epigenetics.
31 *Funct Ecol* 32:1982–1994.

- 1 Clark MS, Villota Nieva L, Hoffman JI, Davies AJ, Trivedi UH, Turner F,
2 Ashton G V, Peck LS (2019b) Lack of long-term acclimation in Antarctic
3 encrusting species suggests vulnerability to warming. *Nat Commun* 10:1–
4 10.
- 5 Clarke A, Meredith MP, Wallace MI, Brandon MA, Thomas DN (2008)
6 Seasonal and interannual variability in temperature, chlorophyll and
7 macronutrients in northern Marguerite Bay, Antarctica. *Deep Res Part II*
8 *Top Stud Oceanogr* 55:1988–2006.
- 9 Collins M, Peck LS, Clark MS (2021) Large within, and between, species
10 differences in marine cellular responses: Unpredictability in a changing
11 environment. *Sci Total Environ* 794:148594.
- 12 De Leij R (2021) Functional response of the Antarctic sea urchin, *Sterechinus*
13 *neumayeri*, to environmental change and extreme events in the context of
14 a warming climate (In Press). PhD Thesis, University of Southampton
- 15 De Leij R, Peck LS, Grange LJ (2021) Multiyear trend in reproduction
16 underpins interannual variation in gametogenic development of an
17 Antarctic urchin. *Sci Rep* 11:1–13.
- 18 Deschaseaux ESM, Taylor AM, Maher WA, Davis AR (2010) Cellular
19 responses of encapsulated gastropod embryos to multiple stressors
20 associated with climate change. *J Exp Mar Bio Ecol* 383:130–136.
- 21 Donelson JM, Munday PL, McCormick MI, Pitcher CR (2012) Rapid
22 transgenerational acclimation of a tropical reef fish to climate change. *Nat*
23 *Clim Chang* 2:30–32.
- 24 Dunlop KM, Barnes DKA, Bailey DM (2014) Variation of scavenger richness
25 and abundance between sites of high and low iceberg scour frequency in
26 Ryder Bay, West Antarctic Peninsula. *Polar Biol* 37:1741–1754.
- 27 Ferrarini A (2011) Detecting ecological breakpoints: a new tool for piecewise
28 regression. *Comput Ecol Softw* 1:121–124.
- 29 Fitzgibbon QP, Simon CJ, Smith GG, Carter CG, Battaglione SC (2017)
30 Temperature dependent growth , feeding , nutritional condition and
31 aerobic metabolism of juvenile spiny lobster, *Sagmariasus verreauxi*.

1 Comp Biochem Physiol Part A 207:13–20.

2 Garrabou J, Coma R, Bensoussan N, Bally M, Chevaldonné P, Cigliano M,
3 Diaz D, Harmelin JG, Gambi MC, Kersting DK, Ledoux JB, Lejeusne C,
4 Linares C, Marschal C, Pérez T, Ribes M, Romano JC, Serrano E,
5 Teixido N, Torrents O, Zabala M, Zuberer F, Cerrano C (2009) Mass
6 mortality in Northwestern Mediterranean rocky benthic communities:
7 Effects of the 2003 heat wave. *Glob Chang Biol* 15:1090–1103.

8 Gillooly JF, Brown JH, West GB, Savage VM, Charnov EL (2001) Effects of
9 size and temperature on metabolic rate. *Science* (80-) 293:2248–2251.

10 Hamby RJAY (1975) Heat Effects on a Marine Snail. *Biol Bull* 149:331–347.

11 Harper EM, Peck LS (2003) Predatory behaviour and metabolic costs in the
12 Antarctic muricid gastropod *Trophon longstaffi*. *Polar Biol* 26:208–217.

13 IPCC (2014) Climate Change 2014: Synthesis Report. Contribution of
14 Working Groups I, II and III to the Fifth Assessment Report of the
15 Intergovernmental Panel on Climate Change. Core Writing Team
16 Pachauri RK and Meyer LA (ed) IPCC, Geneva, Switzerland, 151pp.

17 IPCC (2019) Summary for Policymakers. In: Climate Change and Land: an
18 IPCC special report on climate change, desertification, land degradation,
19 sustainable land management, food security, and greenhouse gas fluxes
20 in terrestrial ecosystems. Shukla PR, Skea J, Calvo Buendia E, Masson-
21 Delmotte V, Pörtner HO, Roberts DC, Zhai P, Slade R, Connors S, van
22 Diemen R, Ferrat M, Haughey E, Luz S, Neogi S, Pathak M, Petzold J,
23 Portugal Pereira J, Vyas P, Huntley E, Kissick K, Malley J (ed) In press.

24 Janecki T, Kidawa A, Potocka M (2010) The effects of temperature and
25 salinity on vital biological functions of the Antarctic crustacean *Serolis*
26 *polita*. *Polar Biol* 33:1013–1020.

27 Kingsolver JG, Umbanhowar J (2018) The analysis and interpretation of
28 critical temperatures. *J Exp Biol* 221.

29 Kolzenburg R, D’Amore F, McCoy SJ, Ragazzola F (2021) Marginal
30 populations show physiological adaptations and resilience to future
31 climatic changes across a North Atlantic distribution. *Environ Exp Bot*

1 188:104522.

2 Lenihan HS, Peterson CH, Miller RJ, Kayal M, Potoski M (2018) Biotic
3 disturbance mitigates effects of multiple stressors in a marine benthic
4 community. *Ecosphere* 9.

5 Liu H, Kelly MS, Cook EJ, Black K, Orr H, Zhu JX, Dong SL (2007) The effect
6 of diet type on growth and fatty-acid composition of sea urchin larvae, I.
7 *Paracentrotus lividus* (Lamarck, 1816) (Echinodermata). *Aquaculture*
8 264:247–262.

9 Marshall J, Scott JR, Armour KC, Campin JM, Kelley M, Romanou A (2015)
10 The ocean's role in the transient response of climate to abrupt
11 greenhouse gas forcing. *Clim Dyn* 44:2287–2299.

12 McClintock J (1994) Trophic biology of Antarctic shallow-water echinoderms.
13 *Mar Ecol Prog Ser* 111:191–202.

14 van der Meer J (2006) An introduction to Dynamic Energy Budget (DEB)
15 models with special emphasis on parameter estimation. *J Sea Res*
16 56:85–102.

17 Melzner F, Stange P, Trubenbach K, Thomsen J, Casties I, Panknin U, Gorb
18 SN, Gutowska MA (2011) Food Supply and Seawater pCO₂ Impact
19 Calcification and Internal Shell Dissolution in the Blue Mussel *Mytilus*
20 *edulis*. *PLoS One* 6:e24223.

21 Morley SA, Lai CH, Clarke A, Tan KS, Thorne MAS, Peck LS (2014) Limpet
22 feeding rate and the consistency of physiological response to
23 temperature. *J Comp Physiol B Biochem Syst Environ Physiol* 184:563–
24 570.

25 Morley SA, Martin SM, Bates AE, Clark MS, Ericson J, Lamare M, Peck LS
26 (2012a) Spatial and temporal variation in the heat tolerance limits of two
27 abundant Southern Ocean invertebrates. *Mar Ecol Prog Ser* 450:81–92.

28 Morley SA, Martin SM, Day RW, Ericson J, Lai CH, Lamare M, Tan KS,
29 Thorne MAS, Peck LS (2012b) Thermal Reaction Norms and the Scale of
30 Temperature Variation: Latitudinal Vulnerability of Intertidal Nacellid
31 Limpets to Climate Change. *PLoS One* 7:7–10.

- 1 Morley SA, Suckling CC, Clark MS, Cross EL, Peck LS (2016) Long-term
2 effects of altered pH and temperature on the feeding energetics of the
3 Antarctic sea urchin, *Sterechinus neumayeri*. *Biodiversity* 17:34–45.
- 4 Muggeo VMR (2008) Segmented: An R Package to Fit Regression Models
5 with Broken-Line Relationships. *R News* 3:343–4.
- 6 Myrvoll-Nilsen E, Fredriksen HB, Sørbye SH, Rypdal M (2019) Warming
7 Trends and Long-Range Dependent Climate Variability Since Year 1900:
8 A Bayesian Approach. *Front Earth Sci* 7:1–8.
- 9 Obermüller BE, Morley SA, Barnes DKA, Peck LS (2010) Seasonal
10 physiology and ecology of Antarctic marine benthic predators and
11 scavengers. *Mar Ecol Prog Ser* 415:109–126.
- 12 Oliver ECJ, Burrows MT, Donat MG, Sen Gupta A, Alexander L V., Perkins-
13 Kirkpatrick SE, Benthuyesen JA, Hobday AJ, Holbrook NJ, Moore PJ,
14 Thomsen MS, Wernberg T, Smale DA (2019) Projected Marine
15 Heatwaves in the 21st Century and the Potential for Ecological Impact.
16 *Front Mar Sci* 6:1–12.
- 17 Oliver ECJ, Donat MG, Burrows MT, Moore PJ, Smale DA, Alexander L V.,
18 Benthuyesen JA, Feng M, Sen Gupta A, Hobday AJ, Holbrook NJ,
19 Perkins-Kirkpatrick SE, Scannell HA, Straub SC, Wernberg T (2018)
20 Longer and more frequent marine heatwaves over the past century. *Nat*
21 *Commun* 9:1–12.
- 22 Oschlies A, Brandt P, Stramma L, Schmidtko S (2018) Drivers and
23 mechanisms of ocean deoxygenation. *Nat Geosci* 11:467–473.
- 24 Payton SL, Johnson PD, Jenny MJ (2016) Comparative physiological,
25 biochemical and molecular thermal stress response profiles for two
26 unionid freshwater mussel species. *J Exp Biol* 219:3562–3574.
- 27 Peck LS (2018) Antarctic Marine Biodiversity: Adaptations, Environments and
28 Responses to Change. *Oceanogr Mar Biol An Annu Rev* 56:105–236.
- 29 Peck LS (2011) Organisms and responses to environmental change. *Mar*
30 *Genomics* 4:237–243.
- 31 Peck LS (2005) Prospects for survival in the Southern Ocean: Vulnerability of

- 1 benthic species to temperature change. *Antarct Sci* 17:497–507.
- 2 Peck LS (1989) Temperature and basal metabolism in two Antarctic marine
3 herbivores. *J Exp Mar Bio Ecol* 127:1–12.
- 4 Peck LS, Bullough LW (1993) Growth and population structure in the infaunal
5 bivalve *Yoldia eightsi* in relation to iceberg activity at Signy Island,
6 Antarctica. *Mar Biol* 117:235–241.
- 7 Peck LS, Clark MS, Morley SA, Massey A, Rossetti H (2009) Animal
8 temperature limits and ecological relevance: Effects of size, activity and
9 rates of change. *Funct Ecol* 23:248–256.
- 10 Peck LS, Morley SA, Pörtner H-O, Clark MS (2007) Thermal limits of
11 burrowing capacity are linked to oxygen availability and size in the
12 Antarctic clam *Laternula elliptica*. *Oecologia* 154:479–484.
- 13 Peck LS, Morley SA, Richard J, Clark MS (2014) Acclimation and thermal
14 tolerance in Antarctic marine ectotherms. *J Exp Biol* 217:16–22.
- 15 Peck LS, Webb KE, Bailey DM (2004) Extreme sensitivity of biological
16 function to temperature. *Funct Ecol* 18:625–630.
- 17 Peck LS, Webb KE, Miller A, Clark MS, Hill T (2008) Temperature limits to
18 activity, feeding and metabolism in the Antarctic starfish *Odontaster*
19 *validus*. *Mar Ecol Prog Ser* 358:181–189.
- 20 Piepho HP, Ogutu JO (2003) Inference for the break point in segmented
21 regression with application to longitudinal data. *Biometrical J* 45:591–601.
- 22 Pierrat B, Saucède T, Laffont R, De Ridder C, Festeau A, David B (2012)
23 Large-scale distribution analysis of Antarctic echinoids using ecological
24 niche modelling. *Mar Ecol Prog Ser* 463:215–230.
- 25 Pörtner H-O, Bock C, Mark FC (2017) Oxygen- & capacity-limited thermal
26 tolerance: Bridging ecology & physiology. *J Exp Biol* 220:2685–2696.
- 27 Pörtner H-O, Peck LS, Hirse T (2006) Hyperoxia alleviates thermal stress in
28 the Antarctic bivalve, *Laternula elliptica*: Evidence for oxygen limited
29 thermal tolerance. *Polar Biol* 29:688–693.
- 30 Pörtner H-O, Peck LS, Somero G (2007) Thermal limits and adaptation in

- 1 marine Antarctic ectotherms: An integrative view. *Philos Trans R Soc B*
2 *Biol Sci* 362:2233–2258.
- 3 Robertson R, El-Haj AJ, Clarke A, Peck LS, Taylor E (2001) The effects of
4 temperature on metabolic rate and protein synthesis following a meal in
5 the isopod *Glyptonotus antarcticus* Eights (1852). *Polar Biol* 24:677–686.
- 6 Rubio-Portillo E, Izquierdo-Muñoz A, Gago JF, Rosselló-Mora R, Antón J,
7 Ramos-Esplá AA (2016) Effects of the 2015 heat wave on benthic
8 invertebrates in the Tabarca Marine Protected Area (southeast Spain).
9 *Mar Environ Res* 122:135–142.
- 10 Salinas S, Munch SB (2012) Thermal legacies: Transgenerational effects of
11 temperature on growth in a vertebrate. *Ecol Lett* 15:159–163.
- 12 Schlegel RW, Smit AJ (2018) HeatwaveR: A central algorithm for the
13 detection of heatwaves and cold-spells. *J Open Source Softw* 3:821.
- 14 Spicer JI, Morley SA, Bozinovic F (2019) Physiological diversity, biodiversity
15 patterns and global climate change: Testing key hypotheses involving
16 temperature and oxygen. *Philos Trans R Soc B Biol Sci* 374:8–11.
- 17 Stanwell-Smith D, Peck LS (1998) Temperature and embryonic development
18 in relation to spawning and field occurrence of larvae of three Antarctic
19 echinoderms. *Biol Bull* 194:44–52.
- 20 Stevens MM, Jackson S, Bester SA, Terblanche JS, Chown SL (2010)
21 Oxygen limitation and thermal tolerance in two terrestrial arthropod
22 species. *J Exp Biol* 213:2209–2218.
- 23 Suckling CC, Clark MS, Beveridge C, Brunner L, Hughes AD, Harper EM,
24 Cook EJ, Davies AJ, Peck LS (2014) Experimental influence of pH on the
25 early life-stages of sea urchins II: Increasing parental exposure times
26 gives rise to different responses. *Invertebr Reprod Dev* 58:161–175.
- 27 Suckling CC, Clark MS, Richard J, Morley SA, Thorne MAS, Harper EM, Peck
28 LS (2015) Adult acclimation to combined temperature and pH stressors
29 significantly enhances reproductive outcomes compared to short-term
30 exposures. *J Anim Ecol* 84:773–784.
- 31 Terblanche JS, Deere JA, Clusella-Trullas S, Janion C, Chown SL (2007)

1 Critical thermal limits depend on methodological context. Proc R Soc B
2 Biol Sci 274:2935–2942.

3 Venables HJ, Clarke A, Meredith MP (2013) Wintertime controls on summer
4 stratification and productivity at the western Antarctic Peninsula. Limnol
5 Oceanogr 58:1035–1047.

6 Walter J, Jentsch A, Beierkuhnlein C, Kreyling J (2013) Ecological stress
7 memory and cross stress tolerance in plants in the face of climate
8 extremes. Environ Exp Bot 94:3–8.

9 Young JS, Peck LS, Matheson T (2006) The effects of temperature on
10 walking and righting in temperate and Antarctic crustaceans. Polar Biol
11 29:978–987.

12 Zupo V, Glaviano F, Paolucci M, Ruocco N, Polese G, Di Cosmo A, Costantini
13 M, Mutalipassi M (2019) Roe enhancement of *Paracentrotus lividus*:
14 Nutritional effects of fresh and formulated diets. Aquac Nutr 25:26–38.

15

16 **TABLES**

17 Table 1: Summary statistics for linear regression relationships between the
18 measured functions of *Sterechinus neumayeri* and temperature. β indicates the
19 slope of the linear regression lines before the breakpoint (Slope_1) and after
20 the breakpoint (Slope_2); SE_a indicates standard error for the intercept and
21 slopes; df = degrees of freedom; bold p-values indicate significant relationships
22 ($p < 0.05$) between temperature and the variable measured and bold Davies p-
23 values represent a significant change ($p < 0.05$) in the gradient of the slope of
24 segmented regressions. Values in the column BP indicate the localisation of
25 the breakpoint or else NA indicates a single linear regression; SE_b (standard
26 error) and R^2 refers to the goodness of fit for the entire model.

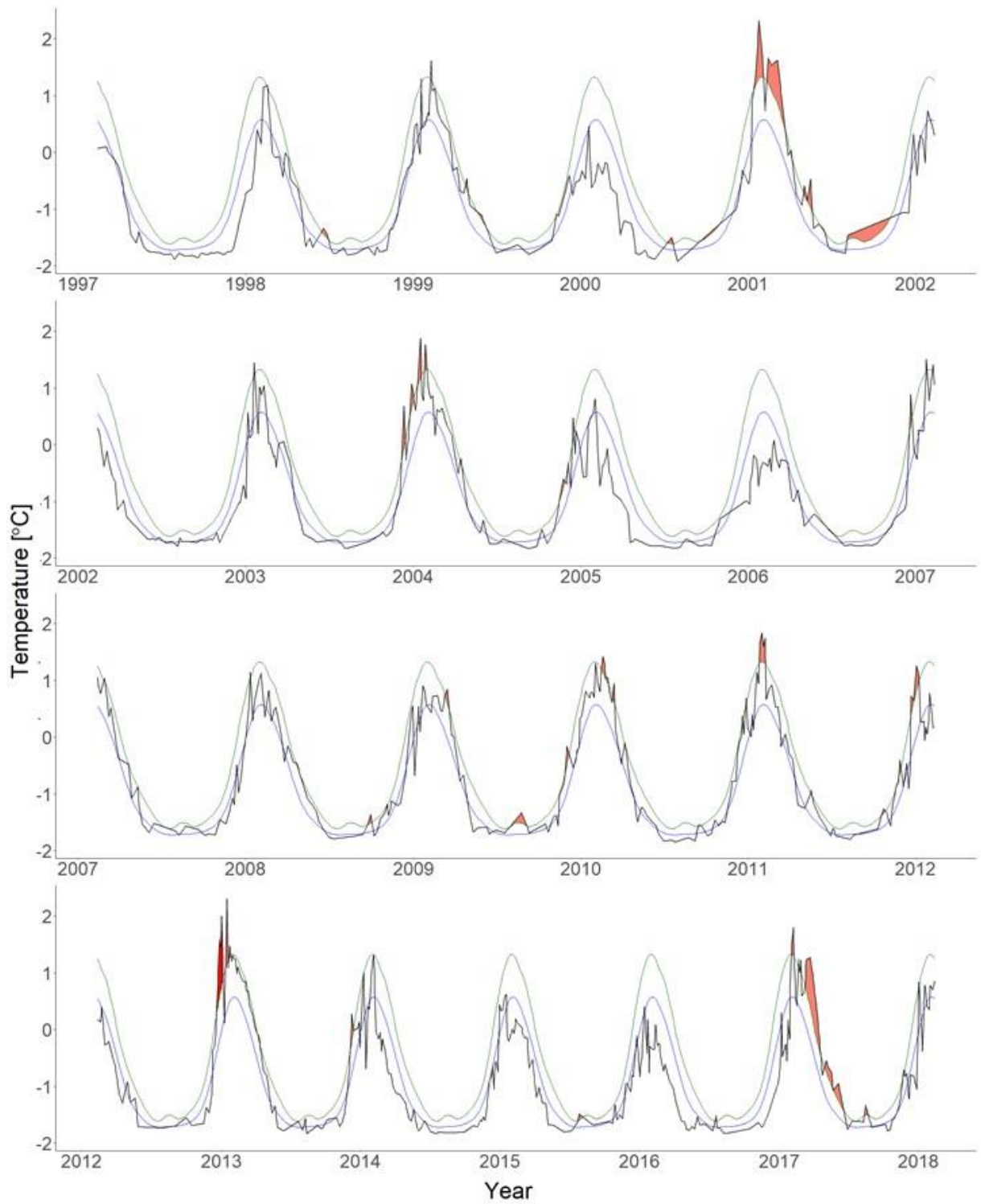
Function	β	SE_a	p-value	BP	SE_b	R^2	Davies p-value
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Individuals feeding, 1°C day ⁻¹ (Intercept) Slope_1 Slope_2	89.0 3.45 -12.9	25.4 10.5 2.35	df=3 0.039 0.764 0.012	4.0	14.9	0.894	0.329
Individuals feeding, 0.5°C day ⁻¹ (Intercept) Slope_1 Slope_2	110.3 -6.34 -11.5	12.7 3.14 1.05	df=7 <0.001 0.083 <0.001	6.2	6.78	0.964	0.301
Individuals feeding, 0.3°C day ⁻¹ (Intercept) Slope_1 Slope_2	95.3 -2.73 -20.3	7.53 1.38 2.92	df=12 <0.001 0.071 <0.001	8.2	8.48	0.922	0.001
Individuals producing faeces, 1°C day ⁻¹ (Intercept) Slope_1 Slope_2	-29.0 24.1 -13.3	23.1 9.54 2.13	df=3 0.298 0.085 0.008	5.2	13.5	0.881	0.019
Individuals producing faeces, 0.5°C day ⁻¹ (Intercept) Slope_1 Slope_2	34.0 13.3 -10.3	28.6 8.54 8.68	df=7 0.274 0.162 <0.001	4.5	12.1	0.844	0.039
Individuals producing faeces, 0.3°C day ⁻¹ (Intercept) Slope_1 Slope_2	77.9 -0.306 -18.6	11.1 2.02 4.29	df=12 <0.001 0.882 <0.001	8.3	12.5	0.762	0.006
Faeces produced, 1°C day ⁻¹ (Intercept) Slope_1	0.645 -0.040	0.137 0.027	df=14 <0.001 0.165	NA	0.216	0.071	0.858
Faeces produced, 0.5°C day ⁻¹ (Intercept) Slope_1 Slope_2	1.52 -0.23 -0.06	0.214 0.072 0.025	df=31 <0.001 0.007 0.016	4.9	1.11	0.664	0.043
Faeces produced, 0.3°C day ⁻¹ (Intercept) Slope_1 Slope_2	3.54 -0.718 -0.051	0.509 0.202 0.020	df=34 <0.001 0.001 0.012	3.3	0.294	0.729	<0.001
Time taken to right, 1°C day ⁻¹ (Intercept) Slope_1 ¹	-8.60 6.83	9.04 1.35	df=26 0.350 <0.001	NA	23.3	0.476	NA
Time taken to right, 0.5°C day ⁻¹ (Intercept) Slope_1 ¹	8.88 2.61	5.03 0.731	df=26 0.089 0.001	NA	13.1	0.302	NA
Time taken to right, 0.3°C day ⁻¹ (Intercept) Slope_1 Slope_2	14.6 0.384 55.7	20.1 3.66 13.8	df=25 0.237 0.459 <0.001	8.7	0.556	0.588	<0.001
Oxygen consumption, 1°C day ⁻¹ (Intercept) Slope_1 ¹	1.64 1.50	1.76 0.248	df=28 0.358 <0.001	NA	4.64	0.551	NA
Oxygen consumption, 0.5°C day ⁻¹ (Intercept) Slope_1 ¹	4.29 0.611	1.10 0.134	df=33 <0.001 <0.001	NA	3.17	0.368	NA

Oxygen consumption, 0.3°C day ⁻¹ (Intercept) Slope_1 ¹	3.30 0.957	1.36 0.185	df=28 0.022 <0.001	NA	3.49	0.471	NA
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¹ Reporting only a single slope (Slope_1) indicates that no breakpoint was detected in the regression and statistics for a single linear regression model is reported for the data instead.

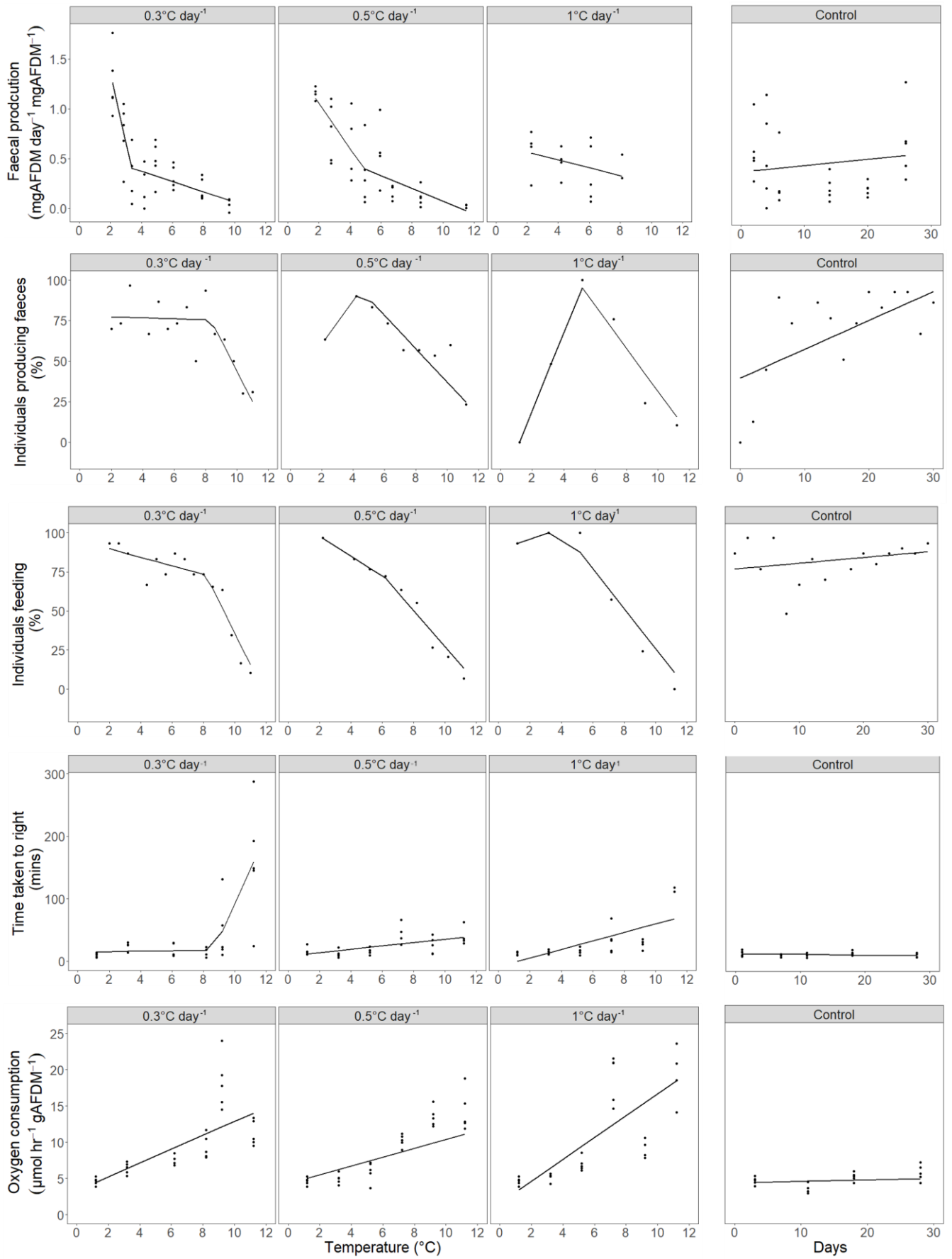
1 FIGURES



2

3 Figure 1: Times-series of temperatures (°C) experienced in Ryder Bay,
4 Antarctica, at depths of 15 m, represented by the black lines. The data are split
5 into panels to cover the entire span of the time-series, where the x-axis

1 represents time in years. Blue lines represent the seasonal climatology of the
2 region based on the full time-series of daily temperatures (1997 – 2018). Green
3 lines represent the seasonally varying threshold for a marine heatwave (90th
4 percentile). Temperatures exceeding the threshold for ≥ 5 days are highlighted
5 in red and indicate the occurrence of a marine heatwave.



1 Figure 2: *Stereochinus neumayeri*. Biological functions measured in *Stereochinus*

1 *neumayeri* in experimental conditions where temperatures were increased daily
2 by 0.3°C, 0.5°C and 1°C. Functions in warming conditions are plotted against
3 increasing temperature and ambient control treatments are plotted against the
4 number of days in the experiment. Data points represent the pooled data within
5 replicate floating tanks (n=5). Regressions are either segmented where
6 appropriate for treatment conditions or linear for controls and treatment data
7 where breakpoints were not identified.

1 Article to Marine Ecology Press Series

2

3 Functional thermal limits are determined by rate of
4 warming during simulated marine heatwaves

5

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13 Running head: Functional thermal limits during marine heatwaves

1 **ABSTRACT**

2 Marine heatwaves (MHWs) are increasing in both intensity and frequency
3 against a backdrop of gradual warming associated with climate change.

4 ~~Temperatures experienced during these warming events are likely to cause~~
5 ~~sub-lethal effects to animals as organism functioning deteriorates. Little is~~
6 ~~known about functional deterioration as critical thermal limits are approached,~~

7 ~~and i~~n the context of ~~marine heat waves (MHWs)~~, animals are likely to
8 experience ~~temperatures producing~~ sub-lethal, rather than lethal effects,
9 defining long-term limits to survival and/or inhibiting impacting individual and

10 population healthfitness. This study ~~aims to understand~~investigated how
11 functional sub-lethal limits track critical thresholds and how this relationship

12 changes with ~~the~~ warming rate. To this ~~end~~purpose we monitored basic
13 functioning, specifically the ability to right, feed, and assimilate energy, as well

14 as oxygen consumption rate~~and metabolise energy~~, in the common Antarctic
15 sea urchin, *Sterechinus neumayeri*. Water temperature in experimental

16 systems was increased at ~~warming~~ rates of 1°C day⁻¹, 0.5°C day⁻¹ and 0.3°C
17 day⁻¹, in line with the characteristics of ~~warming~~ MHW events previously

18 experienced at the site where the study urchins were collected on the ~~in~~
19 Antarctica Peninsula. Functioning was assessed during the simulation of

20 MHWs warming and sub-lethal limits determined when the rate of functional
21 degradation changed as temperature increased. Results suggest that ~~the rate~~

22 ~~of functional degradation, and ultimately~~ thermal sensitivity, varies across
23 warming rates and the specific between the key biological functions measured,

24 with the ability to right having the highest thermal threshold. ~~Arguably, the most~~
25 interesting result was that functions deteriorated at lower temperatures when

1 warming was more rapid (1°C day⁻¹), contrary to lethal critical thresholds, which
2 were reached at lower temperatures when warming was slower (0.3°C day⁻¹).

3 MHWs and their impacts extend far beyond Antarctica. and i-In this context, our
4 analyses indicate that the onset rate of a warming MHWs is critical in
5 determining an organism's ability to tolerate short-term elevated temperatures
6 with global relevance.

7 Key words: Extreme warming events, sub-lethal limits, thermal tolerance,
8 climate change, polar, segmented regression, echinoderm

9 1. INTRODUCTION

10 ~~Paleoclimate records have identified the onset of industrial-era warming as~~
11 ~~early as the mid-19th century (Abram et al. 2016) and h~~Historical temperature
12 records have now detected positive temperature trends for the majority of the
13 Earth's surface ~~(Myrvoll-Nilsen et al. 2019).~~ (Myrvoll-Nilsen et al. 2019), with
14 ~~T~~the oceans are being key to the regulation and capture of much of the excess
15 heat present in the atmosphere (Marshall et al. 2015), ~~and as~~As a result,
16 marine environments are changing both physically and biochemically (Bopp et
17 al. 2013). Included in these changes is the occurrence of marine heat waves
18 (MHWs), which are increasing in duration, magnitude and frequency, frequency
19 ~~and intensity~~ with alarming ecological consequences (Garrabou et al. 2009,
20 Rubio-Portillo et al. 2016, Oliver et al. 2018). ~~Not only are the duration,~~
21 ~~magnitude and frequency of MHW's expected to increase (Hobday et al. 2016,~~
22 ~~Oliver et al. 2018), but models also predict that by the end of the 21st century~~
23 ~~50% of the world's oceans will be in a state of 'permanent MHW' (sensu Oliver~~
24 ~~et al. 2019) under RCP4.5 scenarios, and >90% under RCP8.5 scenarios~~
25 ~~(Oliver et al. 2019).~~

1 Physiological flexibility of species is crucial to survival during MHW events
2 (Peck 2011) and species at low latitudes may be able to acclimate and adapt
3 through multipleacross generations to altered environments (Donelson et al.
4 2012, Salinas & Munch 2012, Clark et al. 2019a). As a result, predicting effects
5 of MHWs on lower latitude species may need to consider shifting thermal
6 ranges as these species adapt to climate change. It is unlikely that the same
7 will apply to Antarctic species, since many are physiologically limited by their
8 capacity to acclimate and evolveadapt to new temperatures Benthic marine
9 invertebrates in Antarctica are characterised by a limited capacity to acclimate
10 and evolve to new temperatures (Peck et al. 2014, Peck 2018). Therefore, the
11 physiological flexibility of these species becomes crucial to their survival during
12 extreme warming events (Peck 2011). The Antarctic has been subject to
13 warming of 0.61 ± 0.34 °C per decade since 1990, more than three times the
14 global average (Clem et al. 2020), with rapid warming in some marine areas
15 such as the Western Antarctic Peninsula (Meredith & King 2005, Turner et al.
16 2014, Spence et al. 2017). Despite predictions that permanent heat wave status
17 in Antarctica will be reached at a slower rate than lower latitudes (Oliver et al.
18 2019), the species that live in the Southern Ocean are less likely to be able to
19 adapt quickly to changing temperatures. Species at lower latitudes may be able
20 to acclimate and adapt through multiple generations to gradual warming. As a
21 result, Ppredicting effects of MHWs on lower latitude species may need to
22 consider a shift in species thermal thresholds over time.involve a shifting
23 baseline as species adapt to warming temperatures because of their long
24 generation times and delayed reproductive maturity (Peck et al. 2014, Peck
25 2018). For example, several invertebrate species such as the Antarctic scallop

1 *Adamussium colbecki*, the limpet *Nacella concinna*, and the bivalves, *Laternula*
2 *elliptica* and *Adacnarca nitens*, take 4 – 7 years to mature. The Antarctic
3 bivalve, *Aequiyoldia eightsi*, starts reproducing at around 12 years (Peck &
4 Bullough 1993) and the brachiopod *Liothyrella uva*, can take up to 18 years
5 before brooding young (Peck 2005, 2018, Oliver et al. 2019).

6 ~~. However, it is unlikely that the same will be possible for many Antarctic species~~
7 ~~owed to their long generation times and delayed reproductive maturity, with~~
8 ~~several invertebrate species, including the Antarctic scallop *Adamussium*~~
9 ~~*colbecki*, the limpet *Nacella concinna*, and the bivalves, *Laternula elliptica* and~~
10 ~~*Adacnarca nitens*, taking 4 – 7 years to mature, and others such as the~~
11 ~~brachiopod *Liothyrella uva*, taking up to 18 years before brooding young (Peck~~
12 ~~2005, 2018, Oliver et al. 2019).~~

13 Predicting species and ecosystem responses to MHWs is challenging, owed to
14 the past infrequency and variability of each event (Oliver et al. 2018). However,
15 if we can track the functional deterioration of organisms, when temperatures
16 exceed their typical thermal range, this can inform our understanding of the
17 relationships between the sub-lethal and lethal limits likely to be encountered
18 during MHW events.

19 For organisms with slow growth and development and long generation times,
20 like many of those found in Antarctica, thermal stress caused by MHWs is likely
21 to trigger other mechanisms for survival such as biochemical and cellular stress
22 responses (e.g. Clark & Peck 2009, Payton et al. 2016). ~~a range of biological~~
23 ~~responses and ultimately survival mechanisms compared to those required for~~
24 ~~gradual warming (Somero 2010, Peck 2011). Where evolution or even~~
25 ~~acclimation is not possible due to the rate of temperature change, other~~

1 ~~mechanisms for survival must come into play, such as biochemical and cellular~~
2 ~~stress responses (Clark & Peck 2009, Payton et al. 2016). Thus, if species are~~
3 ~~able to acclimate rapidly then a small temperature change might have little~~
4 ~~effect. However, for species that acclimate very slowly, increases in~~
5 ~~temperature not usually considered significant elsewhere, might put animals~~
6 ~~out of physiological balance with detrimental consequences.~~

7 Biochemical and genetic mechanisms, including a range of chaperone
8 proteins, provide a short-term buffer that allow functioning to continue
9 temporarily at temperatures outside an organism's thermal niche
10 (Deschaseaux et al. 2010, Clark et al. 2019b). Once animals are no longer able
11 to maintain basic functions by these mechanisms, the sub-lethal limit to survival
12 is reached.

13 Data on the functional thermal limits of species and ~~the warming~~ MHW
14 characteristics (i.e. rate, magnitude and duration) at which these thresholds are
15 reached are rare, especially in fluctuating environments (Janecki et al. 2010,
16 Peck et al. 2014, Ardor Bellucci & Smith 2019). Little is known about functional
17 deterioration as a species approaches its critical thermal limit, and in the context
18 of MHWs, animals are likely to experience temperatures that cause sub-lethal,
19 rather than lethal effects, defining long-term limits to survival and/or inhibiting
20 population health (Pörtner et al. 2007).

21 This study aims to understand how functional ~~limits~~-(sub-lethal) limits track
22 critical ~~limits~~-(lethal) limits and how this relationship changes with warming rate
23 during a simulated MHW. To this purpose, we monitored the ability to right, feed
24 and, assimilate energy as well as oxygen consumption rate ~~and metabolise~~
25 energy, in the common Antarctic sea urchin, *Sterechinus neumayeri*.

1 ~~*Sterechinus neumayeri* is an abundant Antarctic species that forms a significant~~
2 ~~component of the benthic community (Brockington 2001, Pierrat et al. 2012),~~
3 ~~with a circumpolar distribution (Kroh 2010). It is a grazing urchin with a catholic~~
4 ~~diet, and is a broadcast spawning species that releases gametes into the water~~
5 ~~column during the austral summer (Pearse & Giese 1966, Stanwell-Smith &~~
6 ~~Peck 1998, Brockington et al. 2007).~~

7 **2. MATERIALS AND METHODS**

8 2.1 Sample site and animal collections

9 *Sterechinus neumayeri* were sampled from South Cove, Rothera Point
10 (67°34'09.1"S 68°07'52.7"W), from sites near the British Antarctic Survey's
11 Rothera Research Station on the Western Antarctic Peninsula (WAP) during
12 December 2019 (Figure S1). 120 adult urchins (test diameter range, 28 mm
13 – 49 mm) were SCUBA-diver collected at depths of 10-20 m and returned to
14 the Rothera aquarium facility within two hours of collection.

15 *Sterechinus neumayeri* is one of the most common and locally abundant
16 members of the Antarctic marine shallow benthos, forming a significant
17 component of the benthic community (Brockington 2001, Pierrat et al. 2012),
18 with reported densities up to 600 m² (Barnes & Brockington 2003). It is a major
19 scavenger of dead organisms and in iceberg scours on the shallow Antarctic
20 seabed (Dunlop et al. 2014), and it is a significant grazer and bioturbator of
21 sediments (Lenihan et al. 2018). Because of this *S. neumayeri* is an important
22 carbon transformer in Antarctic shallow seas. Further to this, ~~because of~~ due to
23 its abundance and ease of maintenance in laboratory culture systems, *S.*
24 *neumayeri* has been the subject of extensive study of its embryonic and larval
25 development, which is highly extended, and up to in excess of 100 days (Bosch

1 et al. 1987). It has also been the subject of studies of the effects of temperature
2 on embryonic and larval development (Stanwell-Smith & Peck 1998), the
3 impact of ocean acidification on reproduction (Suckling et al. 2014) and energy
4 budgets (Morley et al. 2016). Furthermore, it has been shown that there are
5 long-term cycles in its reproduction (De Leij et al. 2021). These factors all
6 makeOverall *S. neumayeri* is one of the most important members of the
7 Antarctic shallow benthic ecosystem and key to investigating responses to
8 MHWs.

9 2.2 Experimental set-up and warming system

10 A decade of temperature data (1997-2017) from Ryder Bay on the WAP
11 (sourced from the Rothera Time-Series (RaTS) environmental monitoring
12 programme (Clarke et al. 2008, Venables et al. 2013)) was used in the R
13 package “heatwaveR” (Schlegel & Smit 2018), to detect past warming events
14 (Figure 1) (see details of warming event analysis methodology and
15 characteristics summary in the Supplementary Materials, Text S1-~~&~~, Table S1,
16 Figure S2). Studying the characteristics of these past warming events,
17 including onset rate and magnitude, allowed us to set realistic warming rates
18 for the experimental systems.

19 Urchins were held in flow-through aquaria (170 L) at ambient temperatures
20 typical for December and January (-1.5°C to +0.5°C) for six weeks on a
21 continuous light regime. During this time, animals were ~~starved-not fed~~ to allow
22 any ingested food to be processed and the production of faeces to cease. Since
23 feeding and faeces production was not occurring, it was assumedThe cessation
24 of production of faeces production is an indicator that metabolic rates that This

1 ~~step ensured all individuals were at the same~~ had reached a “standard”
2 ~~metabolic state at the level at the~~ start of the experiment. Previous research
3 suggests that these urchins are able to sustain, and experience natural periods
4 of starvation for up ~~six~~6 months during winter (Brockington 2001), and hence
5 six weeks ~~starvation without feeding~~ was unlikely to ~~impact be detrimental to~~
6 their physiological ~~metrics measured in this study capabilities.~~ Previous
7 studies of oxygen consumption in other Antarctic marine invertebrates has
8 demonstrated that standard levels are reached in less, and often significantly
9 less, than this time –in the brachiopod *Liothyrella uva* and the limpet *Nacella*
10 *concinna* (Peck 1989), in the amphipod *Waldeckia obesa* (Chapelle et al.
11 1994), in the isopod *Glyptonotus antarcticus* (Robertson et al. 2001), and in the
12 starfish/sea star *Odontaster Validus* (Peck et al. 2008).

13 After urchins were maintained in the flow-through aquarium (170 L) at ambient
14 temperatures, 30 urchins were distributed to four main aquarium tanks to
15 represent each warming treatment as well as the ambient control treatment.
16 Urchins were distributed at random. Replication within each of these treatments
17 was achieved by floating five separate 6-litre tanks, each containing six urchins
18 ~~(30 urchins per treatment)~~ in each main aquarium tank (170 L). Each main
19 aquarium tank functioned, which functioned as a temperature baths (Figure S3;
20) (30 urchins per treatment, 5 replicates per treatment where data from urchins
21 in the same replicate floating- tank were pooled). (Figure S2). Temperature
22 treatments were not replicated due to space restrictions in the aquarium. It is
23 acknowledged that t Treatment temperature The same treatment conditions
24 (i.e., temperature) would be was translated to all replicate urchins, and as such,

1 temperature was closely monitored to note and control any unintentional
2 variability (Figure S4).

3 The water in each floating tank was aerated using air stones and refreshed by
4 50% water change every other day. Water changes not only ensured that
5 overall water quality was maintained, but also meant any metabolic products,
6 especially potentially toxic nitrogenous chemical species, were maintained at
7 very low levels. Tank water samples were periodically analysed for pH
8 (~~aim: ranging~~ 7.9-5 - 8.50), NO₂ (~~aim: ranging <0.2~~ 0.05 mg l⁻¹ – 0.1 mg l⁻¹), NO₃
9 (~~aim: <20~~ ranging 0.5 mg l⁻¹ – 1.0 mg l⁻¹) and NH₄ (~~aim: stable at <1.20~~ 1 mg l⁻¹)
10 to ensure good water quality. Throughout the experiment, concentrations of the
11 aforementioned compounds remained within ~~the advised the~~ ranges stated.

12 Urchins within each replicate tank were separated by aquaria egg crates and
13 fine mesh partitions to ensure individuals were isolated and any faeces
14 produced was retained within compartments (Figure ~~S2S3~~). During warming
15 trials, ~~we aimed to increase~~ experimental temperatures ~~in~~ the aquaria water
16 baths were raised by 1°C, 0.5°C or 0.3°C each evening, depending on
17 treatment. Temperatures in the floating tanks increased more gradually than
18 the water baths, allowing urchins to adjust ~~more~~ slowly to ~~the each~~ new
19 temperature. Temperatures were checked every 30 minutes after each
20 temperature change to ensure required temperatures were achieved and kept
21 constant. Initially, temperatures fluctuated by up to ± 0.3°C before stabilising
22 after 1-2 hrs. Temperatures were subsequently monitored throughout the
23 following day and held within ± 0.1°C of the target experimental temperature
24 (Figure S4). For ambient controls, urchins were held in the aquarium with the
25 set-up and light conditions identical to the warming treatment conditions.

1 Temperatures were maintained at those experienced in Ryder Bay which
2 naturally fluctuated between 0.9 °C and 1.9°C.

3 2.3 Feeding trials

4 Urchins were fed pre-portioned amounts of food every 48 hrs. Previous studies
5 fed *S. neumayeri* high protein diets, such as fish fillets, *Polachius virens*
6 (Suckling et al. 2014, Morley et al. 2016). In the current study, urchins were fed
7 the foot of the common Antarctic limpet, *Nacella concinna*, which has a
8 comparable protein content to that of *P. virens* muscle. Based on feeding
9 protocols in Morley et al. (2016b), urchins were fed ~4% of their mean body
10 mass every three weeks, but this was spread across 48 hr feeding increments
11 in order to keep feeding activity constant and reduce the variability in daily
12 metabolic activity.

13 Limpets were chosen as a food source since nutrient content could be
14 controlled and pre-portioned. A more representative diet would be a varied one
15 with algal biofilm, animal tissues and/ or detritus (McClintock 1994). ~~However,~~
16 ~~with these sources it administering a varied diet would be very difficult to assess~~
17 ~~themake it difficult to assess the amount of food consumed per urchin as well~~
18 ~~asat the same time as standardisinge the nutritional content. There is evidence~~
19 ~~that diet, especially protein levels, can affect development and gonad growth~~
20 (Liu et al. 2007, Zupo et al. 2019) as well as ingestion and assimilation rates in
21 sea urchins (Azad et al. 2011). As such, by feeding a diet of limpets it should
22 be acknowledgedis possible that body condition may be altered and the ability
23 to tolerate stress may be improved as a result.

1 Feeding was initiated two days before the start beginning of the experiment to
2 start the digestion process. ~~Urchins were offered food by placing it directly onto~~
3 ~~the test. This technique had been used previously in experimental trials and~~
4 ~~allows the urchin to detect the food and move it to the mouth without disturbing~~
5 ~~the animal (references needed).~~ Each urchin was allowed to feed for 48 hrs
6 before any remaining food was removed and refreshed. After 48 hrs, each
7 urchin was recorded as feeding or not feeding. Infrequently, urchins may have
8 ~~not only partially~~ consumed ~~all the the~~ entire food piece, which was recorded.

9 2.4 Faecal collection

10 Faecal production began four days into the experiment, 6-days after feeding
11 was initiated. The presence of faeces was recorded for all urchins every 48 hrs.
12 To measure faecal production, faeces were collected every 48 hrs by pipette
13 and transferred to falcon tubes from 10 urchins per treatment, and where at
14 least one sample was taken from each replicate tank within the treatment. The
15 same urchins were targeted for faecal collection in order to minimise
16 subconscious preferences towards urchins producing more faeces. This was
17 not always possible since sometimes urchins did not produce any faeces or
18 else CT_{max} was reached, and these urchins were removed. In these cases, a
19 different urchin was chosen at random to sample from. -For all other urchins,
20 any remaining faecal matter was removed ~~and disposed of~~.

21 Collected faecal matter was centrifuged and the supernatant seawater
22 decanted. Faeces were then rinsed with RO (Reverse Osmosis purified) water
23 by agitating and centrifuging to remove any seawater salt. Washed faeces were
24 pipetted into pre-ashed and pre-weighed foil boats and dried at 60°C for 24 hrs.
25 Dry foil boats and faeces were placed in a desiccator to cool and then weighed

1 (± 1 mg). Dry faeces were subsequently ignited in a muffle furnace at 475°C for
2 6 hrs. Foil boats and ashed faeces were cooled in a desiccator and weighed (\pm
3 1 mg). Dry mass (DM) and Ash-Free Dry Mass (AFDM) (i.e., organic content)
4 were obtained by subtraction.

5 2.5 Respirometry

6 Oxygen consumption was recorded for 10 urchins per treatment, sampling two
7 individuals from each replicate tank within each treatment. Oxygen
8 consumption was recorded for the same urchins for every 2°C rise in
9 temperature from ambient in each treatment. Methods for measuring oxygen
10 consumption followed those described by Suckling et al., (2015), using 200 -
11 250 ml volume chambers. For each urchin, live wet mass (± 0.01 g) was
12 recorded where O₂ consumption was measured. AFDM was determined from
13 live wet mass vs AFDM regressions determined from a subsample of urchins
14 (n = 40) collected from the same site. ~~For the purpose of To obtaining~~ the ash
15 mass of urchins, individuals were weighed live before freezing in liquid nitrogen
16 and storing at -40°C. Frozen urchins were then placed in pre-ashed and pre-
17 weighed, ceramic crucibles, and dried at 60°C until constant mass was obtained
18 (± 0.01 g). Once dried, urchins were ~~then~~ ignited in a muffle furnace at 475°C
19 for 6 hrs and subsequently weighed to obtain ash mass ~~once-after~~ cooling in a
20 desiccator (± 1 mg).

21 2.6 Righting

22 The time taken for urchins to right themselves was recorded for 10 urchins per
23 treatment, sampling two urchins from each replicate tank within each treatment.
24 The time taken to right was recorded for the same urchins every 2°C rise in

1 temperature from ambient in each treatment. Ten individuals were removed
2 from their experimental tanks and placed in individual containers. These
3 containers were previously filled and floated in water already at the
4 experimental target temperature. Urchins were immediately inverted following
5 transfer from experimental tanks to the floating containers and timed until the
6 individual was fully upright. Urchins could not reach the sides of containers to
7 aid in righting. Once righted, urchins were returned to their experimental tanks.

8 2.7 Critical temperature limits (CT_{max})

9 The critical thermal limit (CT_{max}) was recorded for ~~each urchin~~all experimental
10 urchins in the warming treatments, where the limit was defined as the point at
11 which the individual was unable to right itself within 12 hrs, had stopped eating
12 and ~~had stopped~~ producing faeces. When an urchin began to show signs of
13 reaching the CT_{max} (not feeding or producing faeces), they were inverted in the
14 tank and left for 12 hrs. If the urchin had not righted itself after this period, they
15 were removed and weighed suspended in water to obtain live wet volumes (\pm
16 0.01 mL).

17 2.8 Statistical Analysis

18 Where multiple urchins were sampled within the same floating tank,
19 measurements of feeding, faecal production, ~~righting~~righting, and oxygen
20 consumption were pooled so that $n = 5$, and the standard errors were calculated
21 from these five replicate tanks.

22 To determine differences in functional responses between treatments, a one-
23 way repeat measures ~~n~~-analysis of variance (ANOVA) was carried out in R (v.
24 4.0.5). This analysis was considered appropriate for this experiment due to the

1 related and non-independent groups at each temperature timepoint. For this
2 analysis, treatment group variances were compared when treatments reached
3 the same temperature increments. For ambient controls, temperature
4 timepoints were aligned with measurements taken at similar dates to treatment
5 sampling. Variances were compared between groups and within timepoints for
6 righting and oxygen consumption rates and the resultant p-value was adjusted
7 using the Bonferroni correction method. Significant differences ($p < 0.05$) were
8 followed up with a paired t-test and again, p-values were adjusted using the
9 Bonferroni correction method. Data were initially log transformed to ensure
10 assumptions of normal distribution were met.

11 ~~When a significant difference was observed ($p < 0.05$), a post-hoc Tukey test~~
12 ~~was undertaken to test all pairwise comparisons among means. A Shapiro-Wilk~~
13 ~~and Levene's test were run on the residuals of the ANOVA for normality of~~
14 ~~distribution, and to test for homogeneity of variance, respectively. Where the~~
15 ~~results of these tests indicated either a non-normal distribution of residuals or~~
16 ~~unequal variances between treatment groups, the response data were~~
17 ~~logarithmically transformed and all tests repeated. Where normality and/or~~
18 ~~homogeneity of variance could not be achieved by transforming the data, the~~
19 ~~non-parametric Kruskal-Wallis and pair-wise Wilcoxon tests were used to~~
20 ~~determine differences in functional response between treatments.~~

21 Segmented linear regression models were fitted in the R package 'segmented'
22 (Muggeo 2008) to identify breakpoints in the linear relationships between
23 functional process and temperature. Breakpoints ~~_points~~ were identified where
24 the gradient of the relationship changed (McWhorter et al. 2018). The change
25 in gradient was used to define the functional threshold of the process

1 measured. It was especially important to use a method such as segmented
2 regression to identify breakpoints in process rates. Segmented regressions
3 were used to model these relationships not necessarily for the purpose of fitting
4 the simplest model, but rather to identify any change in the regressions gradient
5 which then indicated that the functions response to temperature increase had
6 changed. In some cases, a linear regression would be sufficient to explain the
7 relationship, however a linear model could mask the subtle change in the rate
8 of degradation experienced when a species hits a thermal threshold. Where the
9 R² and standard error of the segmented regression model was improved
10 compared to better than a linear regression, a segmented regression was used
11 to model the data. It was especially important to use a method such as
12 segmented regression to identify breakpoints in process rates. Alternatives
13 would be to fit curves and identify changes in slope (e.g. Pörtner et al. 2006),
14 but curves were not appropriate here. A Davies test was also conducted to
15 determine significant ($p < 0.05$) differences in the gradients of the segmented
16 slopes.

17 Size effects on functional response were explored through scatter plots. Where
18 relationships were observed, the effect of size (as test diameter) and
19 temperature on the functional response, was assessed with a linear mixed
20 effects model using the package 'lme4' and the function 'lmer' in R (v. 4.0.5).
21 Test diameter and temperature were added as interacting fixed terms and
22 replicate tank ID was added as a random effect. Prior to any modelling, function
23 responses were transformed to achieve normality in the distribution.

24

1 ~~For time points to be comparable between treatments, the mean temperature~~
2 ~~and time exposed to temperatures above ambient were combined to estimate~~
3 ~~cumulative intensity (°C x day). Other variations on cumulative temperature~~
4 ~~have been used to combine temperature intensity and duration in heatwave~~
5 ~~studies (Perkins-Kirkpatrick & Lewis 2020, Domínguez et al. 2021), however~~
6 ~~cumulative intensity was used to detect past warming events in the~~
7 ~~“heatwaveR” package (Text S1), and so this metric was used to facilitate~~
8 ~~comparison between our experimental conditions and ‘real world’ events.~~
9 ~~(Comparisons between cumulative temperature and cumulative intensity is~~
10 ~~provided in the Supplementary Materials, Table S2).~~

11 ~~The following equation was used to calculate cumulative intensity for a~~
12 ~~temperature in each treatment:~~

$$\text{Exp}(\bar{T} - \text{Amb } \bar{T}) = \text{Cumulative intensity (°C x day)}$$

14 ~~Where: Exp = Exposure in days, above ambient temperatures; \bar{T} = Mean~~
15 ~~temperature experienced °C; Amb \bar{T} = Mean ambient temperature.~~

16 ~~Warming rate was not consistent across treatments averaging 0.32 ± 0.13~~
17 ~~°C day⁻¹ for the slowest warming rate, 0.49 ± 0.17 °C day⁻¹ for intermediate~~
18 ~~warming rates, and 0.97 ± 0.31 °C day⁻¹ for the fastest warming rate. Therefore,~~
19 ~~it was not possible to calculate cumulative intensity directly using the equation.~~
20 ~~Instead, polynomial regressions that took account of the varying rate of~~
21 ~~temperature increase were obtained for each treatment and used to estimate~~
22 ~~cumulative intensity (Figure S3) ($T\uparrow$ = temperature increase):~~

$$T\uparrow \text{ of } 1^\circ\text{C day}^{-1}: (0.49T^2) - (0.99T) = \text{Cumulative intensity (°C x day)}$$

$$T\uparrow \text{ of } 0.5^\circ\text{C day}^{-1}: (0.96T^2) - (1.52T) = \text{Cumulative intensity (°C x day)}$$

1 ~~$T\uparrow$ of $0.3^\circ\text{C day}^{-1}$: $(1.47T^2) - 2.22T$ = Cumulative intensity ($^\circ\text{C} \times \text{day}$)~~

2 ~~Cumulative intensity as an explanatory variable was used for comparing~~
3 ~~functional responses between treatments (Table 2).~~

4 **3. RESULTS**

5 3.1 Feeding and faecal egestion

6 On average, $80\% \pm 19\%$ of animals fed in ambient conditions for the duration
7 of the experiment. For the first four days of the experiment, in treatments where
8 $T\uparrow 1^\circ\text{C day}^{-1}$, the proportion of animals feeding exceeded all other treatments
9 ($97\% \pm 4\%$), including ambient conditions ($87\% \pm 10\%$). ~~50%~~Fifty percent of
10 animals stopped feeding in treatments when ~~cumulative intensity~~temperatures
11 exceeded ~~$18^\circ\text{C} \times \text{day}$~~ 7.2°C , ~~$52^\circ\text{C} \times \text{day}$~~ 8.2°C , and ~~$104^\circ\text{C} \times \text{day}$~~ 9.2°C , where
12 $T\uparrow$ by 1°C , 0.5°C and $0.3^\circ\text{C day}^{-1}$, respectively (Figure 1).

13 A breakpoint (where the slope of the regression changed) for the % individuals
14 feeding was identified at 4.0°C and 6.2°C in treatments where $T\uparrow 1^\circ\text{C day}^{-1}$
15 and $0.5^\circ\text{C day}^{-1}$, respectively (~~Figure 2A & 2B~~, Table 1). However, changes in
16 the segmented slope gradients were not significantly different from linear
17 regressions for these two treatments (Davies p-value = 0.329 and 0.301,
18 respectively). A breakpoint for the % feeding in $T\uparrow 0.3^\circ\text{C day}^{-1}$ was identified
19 at 8.2°C (~~Figure 2C~~, Table 1), from which point the % individuals feeding
20 declined rapidly and the relationship between temperature and the proportion
21 of individuals feeding became significant ($p < 0.001$). The mean temperature
22 breakpoint for the function of % feeding was $6.1^\circ\text{C} \pm 1.2^\circ\text{C}$, averaged across all
23 treatments.

1 The percentage of animals producing faeces tracked the proportion of animals
2 feeding after the first four days (Figure 31). Following each breakpoint, the
3 relationship between temperature and % individuals producing faeces became
4 significant (Table 1). For the fastest rate of warming where $T \uparrow 1^\circ\text{C day}^{-1}$, a
5 breakpoint was identified at 5.2°C , whereby above which the % individuals
6 producing faeces rapidly declined from 100% to 10.3% ~~of individuals~~, within 6
7 days ~~(Figure 4A)~~. Where $T \uparrow 0.3^\circ\text{C day}^{-1}$ and $0.5^\circ\text{C day}^{-1}$, the regression
8 breakpoint for faecal production was 8.3°C and 4.5°C respectively (Table 1).
9 The mean temperature breakpoint for the function of % producing faeces was
10 $6.0^\circ\text{C} \pm 2.0^\circ\text{C}$, averaged across all treatments.

11 The mean mass of faeces produced ~~per day~~ in treatments where $T \uparrow 0.5^\circ\text{C}$
12 ~~day⁻¹ and $0.3^\circ\text{C day}^{-1}$~~ , was significantly greater than exceeded the faecal mass
13 produced in ambient ambient control conditions (~~mean = $2.11 \text{ mg day}^{-1} \pm 0.23$~~
14 ~~mg day⁻¹) and treatments also in treatments~~ where $T \uparrow 1^\circ\text{C day}^{-1}$, until
15 cumulative intensity temperatures exceeded -2.1°C ($t_{(4)} = 8.74$, $p = 0.006$ and
16 $t_{(4)} = 5.02$, $p = 0.044$, respectively) ~~reached $6^\circ\text{C} \times \text{day}$, and $7^\circ\text{C} \times \text{day}$,~~
17 respectively (Figure 5). Where $T \uparrow 0.5^\circ\text{C day}^{-1}$, the mass of faeces produced
18 was significantly greater than treatments where $T \uparrow 1^\circ\text{C day}^{-1}$, until
19 temperatures exceeded 2.1°C ($t_{(4)} = 5.31$, $p = 0.036$). Despite this observation,
20 no additional food was consumed in these treatments. There was no significant
21 difference between the treatments or control as temperatures increased
22 beyond 2.1°C urchins in those in. 5

23 Breakpoints in regressions were identified at $5.06.5^\circ\text{C}$ and 3.13°C for
24 treatments where $T \uparrow 0.5^\circ\text{C day}^{-1}$ and $0.3^\circ\text{C day}^{-1}$, respectively (~~Figure 6B &~~
25 ~~6C~~, Table 1). The breakpoints for these regressions marked a reduction in the

1 gradient of the 2nd slope, whereby faeces produced day⁻¹ mgAFDM⁻¹ as a
2 function of temperature decreased at a slower rate as temperatures
3 increased (Table 1). The mean temperature breakpoint for the function of faeces
4 produced was 4.19°C ± 1.60.95°C, averaged across the slowest ($T \uparrow 0.3^\circ\text{C day}^{-1}$)
5 and intermediate ($T \uparrow 0.5^\circ\text{C day}^{-1}$) rates of warming.

6 3.2 Righting

7 ~~After 6 days, righting time was significantly longer in treatments where $T \uparrow$~~
8 ~~$0.3^\circ\text{C day}^{-1}$ compared to ambient conditions ($W = 396, p = .003$). However,~~
9 ~~beyond 6 days, righting time reduced as cumulative intensity increased until~~
10 ~~$194^\circ\text{C} \times \text{days}$. From here, righting time increased linearly until CT_{max} was~~
11 ~~reached (Figure 7). In treatments where $T \uparrow 0.5^\circ\text{C day}^{-1}$, In treatments where~~
12 ~~$T \uparrow 1.0^\circ\text{C day}^{-1}$, time taken to right became significantly longer than ambient~~
13 ~~controls when temperatures reached 9.2°C ($t_{(4)} = 6.06, p < 0.022$).~~ 6For
14 ~~treatments where $T \uparrow 0.3^\circ\text{C day}^{-1}$, time taken to right only became significantly~~
15 ~~longer than ambient controls just before CT_{max} was reached, when~~
16 ~~temperatures reached 11.2°C ($t_{(4)} = 6.04, p < 0.023$).~~ For treatments where $T \uparrow$
17 ~~$0.5^\circ\text{C day}^{-1}$, time taken to right never exceeded ambient controls significantly,~~
18 ~~however mean righting times were consistently higher than control conditions~~
19 ~~throughout the warming period.~~

20 ~~and $T \uparrow 1^\circ\text{C day}^{-1}$, righting time was significantly greater than in ambient~~
21 ~~conditions when exposed for $39^\circ\text{C} \times \text{day}$ ($W = 440, p = 0.003$) and $18^\circ\text{C} \times \text{day}$~~
22 ~~($W = 357, p = 0.003$), respectively. From here, the time taken to right fluctuated,~~
23 ~~but was significantly longer than ambient values until CT_{max} was reached.~~

1 A breakpoint in the linear regression was identified at ~~6.8°C and~~ 8.7°C in
2 treatments where temperature was raised at $T \uparrow 1^\circ\text{C day}^{-1}$ and $0.3^\circ\text{C day}^{-1}$,
3 respectively (Figure 8A & 8C, Table 1). ~~The relationship between temperature~~
4 ~~and the time taken to right became significant above these~~ this breakpoint
5 temperatures (6.8°C ; $p = 0.001$ and 8.7°C ; $p < 0.001$). For the other treatments
6 righting time increased linearly without a breakpoint in the regression. ~~The~~
7 ~~mean temperature breakpoint for the function of righting was $7.8^\circ\text{C} \pm 1.0^\circ\text{C}$,~~
8 ~~averaged across the fastest ($T \uparrow 1^\circ\text{C day}^{-1}$) and slowest ($T \uparrow 0.3^\circ\text{C day}^{-1}$) rates~~
9 ~~of warming.~~

10 The interactive effect of urchin size and temperature on the time taken to right
11 was significant ($t_{(204)} = 2.11$, $p = 0.034$), where larger urchins took longer to right
12 at higher temperatures (Figure S5, Table S3).

13 3.3 Oxygen consumption

14 Oxygen consumption rates were significantly higher in heatwave treatments
15 compared to ambient controls ~~after cumulative intensities of $2^\circ\text{C} \times \text{day}$ when~~
16 temperatures reached 7.2°C for all treatments. ~~H,~~ however, oxygen
17 consumption rates were significantly higher than ambient controls from lower
18 temperatures of 3.2°C in treatments where $T \uparrow 0.3^\circ\text{C day}^{-1}$ ($t_{(4)} = 5.62$, $p =$
19 0.030) and 5.2°C in treatments where $T \uparrow 1.0^\circ\text{C day}^{-1}$ ($t_{(4)} = 4.98$, $p = 0.045$).
20 where $T \uparrow 0.3^\circ\text{C day}^{-1}$ ($t_{(57)} = 4.69$, $p < 0.001$, $d = 1.63$), of $18^\circ\text{C} \times \text{day}$, where
21 $T \uparrow 0.5^\circ\text{C day}^{-1}$ ($t_{(56)} = 3.79$, $p < 0.001$, $d = 1.38$), and of $8^\circ\text{C} \times \text{day}$, where $T \uparrow$
22 1°C day^{-1} ($t_{(57)} = 6.28$, $p < 0.001$, $d = 2.18$) (Figure 9). Overall, there was a
23 positive linear trend between oxygen consumption and temperature for all
24 treatments. However, where $T \uparrow 1^\circ\text{C day}^{-1}$, a drop in O_2 consumption occurred

1 at ~~32°C x day~~9.2°C, and where $T \uparrow 0.3^\circ\text{C day}^{-1}$, a ~~peak~~drop occurred just
2 before the CT_{\max} at ~~40°C x day~~11.2°C.

3 O_2 consumption increased ~~more per cumulative intensity~~at a faster rate per
4 increase in temperature where warming rates were fastest at 1°C day^{-1} (slope
5 gradient = 1.50) and increased at the slowest rate when warming rates were
6 slowest at $0.3^\circ\text{C day}^{-1}$ (slope gradient = 0.96) (T_a , compared to the other two
7 slower warming rates (Figure ~~ble~~ 19). No breakpoint was identified in any
8 treatmentTo this effect, the linear relationship between O_2 consumption rate
9 and cumulative intensity was significantly different for treatments where $T \uparrow$
10 1°C day^{-1} , compared to $0.3^\circ\text{C day}^{-1}$ and $0.5^\circ\text{C day}^{-1}$ ($F_{(2,143)} = 16.86$, $p < 0.001$).

11 Owed to the variability observed in treatments where $T \uparrow 1.0 \text{ day}^{-1}$ and 0.3°C
12 day^{-1} , a breakpoint could only be identified in the treatment where $T \uparrow 0.5^\circ\text{C}$
13 day^{-1} (Figure 10B). This breakpoint occurred at 7.0°C , after which the
14 relationship between temperature and O_2 consumption became significant ($p <$
15 0.001), increasing at a faster rate as the temperature was increased, However,
16 the gradient of the two slopes was not significantly different (Davies p -value =
17 0.260).

18 3.4 CT_{\max}

19 The CT_{\max} for urchins in treatments where $T \uparrow 0.3^\circ\text{C day}^{-1}$, $T \uparrow 0.5^\circ\text{C day}^{-1}$ and
20 $T \uparrow 1^\circ\text{C day}^{-1}$ ranged from $10.6^\circ\text{C} - 13.8^\circ\text{C}$, $11.2^\circ\text{C} - 13.7^\circ\text{C}$, and $12.2^\circ\text{C} -$
21 14.2°C , respectively. ~~(Figure 11)~~. The effect of warming rate on the CT_{\max} was
22 significant ($F_{(2, 12)} = 7.29$, $p = 0.008$ chi square = 16.9, $p = < 0.001$, $df = 2$), with
23 post-hoc analysis identifying that for treatments where temperature increased
24 at the fastest rate ($T \uparrow 1^\circ\text{C day}^{-1}$), the CT_{\max} was significantly higher compared

1 to treatments where temperature increased at a slower rate ($T \uparrow 0.5^\circ\text{C day}^{-1}$)
2 (Wilcoxon rank sum test, $p = 0.020$) and where ($T \uparrow 0.3^\circ\text{C day}^{-1}$) (Wilcoxon rank
3 sum test, $p < 0.001$, $t_{(8)} = -6.02$, $p = 0.001$).

4 Across all functions where breakpoints were identified, the slowest rate of
5 warming ($T \uparrow 0.3^\circ\text{C day}^{-1}$) had a mean temperature breakpoint of $78.34^\circ\text{C} \pm$
6 1.3°C . In comparison, the mean temperature breakpoint was $6.15.4^\circ\text{C} \pm 0.5^\circ\text{C}$,
7 and $4.65.3^\circ\text{C} \pm 1.40.6^\circ\text{C}$ for intermediate ($T \uparrow 0.35^\circ\text{C day}^{-1}$) and fast ($T \uparrow 1^\circ\text{C}$
8 day^{-1}) warming rates, respectively.

9 **4. DISCUSSION**

10 ~~Marine heat waves~~ MHWs are predicted to increase in frequency,
11 ~~intensity~~ intensity, and duration in the coming decades. Deterioration of basic
12 animal functioning, critical for long-term survival, will likely be a more frequent
13 consequence of the short-term warming (i.e., weeks-months) caused by
14 MHWs, rather than mortality. However, little is known about functional impacts,
15 especially thresholds and how these limits deteriorate with respect to CT_{max} . By
16 understanding how different functions key biological functions are affected by
17 increasing short term temperature elevations and different warming rates, we
18 can better predict understand how extreme climate events, typified by short-
19 term warming, may impact individuals and populations, and hence
20 communities.

21 In this study, we investigated the effect of warming rates typical of those
22 expected during Antarctic MHW events on the functioning of the Antarctic
23 sea Antarctic sea urchin, *S. neumayeri*. Functional thresholds were identified
24 using segmented regressions, where a breakpoint indicated a gradient change

1 in the response trend with temperature. The identification of regression
2 breakpoints, or slope changes has been used previously to define ecological
3 thresholds, and is considered a more flexible and realistic approach when
4 interpreting complex, often non-linear, ecological relationships (Piepho & Ogutu
5 2003, Ferrarini 2011, Morley et al. 2014).

6 Several studies have shown that faster warming rates result in higher CT_{max} [in](#)
7 [terrestrial](#) (e.g. Terblanche et al. 2007, Allen et al. 2016) [and marine](#) (Peck et
8 al. 2009) [species](#). These observations, along with ~~the findings here~~ [the \$CT_{max}\$](#)
9 [data in this study](#), follow the failure rate model proposed by Kingsolver &
10 Umbanhowar (2018), who showed that critical limits are reached at lower
11 temperatures when warming accumulates over extended periods. However,
12 ~~contrary to this,~~ our results ~~indicate that~~ [for](#) functional thermal limits follow the
13 opposite trend to the CT_{max} , where functions are impacted [detrimentally](#)
14 [negatively](#) at lower temperatures when warming is rapid. Overall, [in this study](#)
15 higher functional thresholds were reached when temperatures were raised
16 slowly (thresholds averaging $87.31^{\circ}\text{C} \pm 1.3^{\circ}\text{C}$). ~~At compared to~~ the faster
17 warming rates, ~~where~~ functional thresholds were ~~either~~ lower ($5.44.9^{\circ}\text{C} \pm 0.5^{\circ}\text{C}$
18 [or \$4.5.36^{\circ}\text{C} \pm 1.40.6^{\circ}\text{C}\$](#)). ~~There was even evidence that or else~~ [some functions](#)
19 declined linearly, with significant ~~functional~~-deterioration [from temperatures](#)
20 ~~from~~ $+2.84^{\circ}\text{C}$ [above ambient when warmed at the fastest rate](#)ing. Thus, short-
21 term exposure to more extreme temperatures has more impact on functioning
22 than longer, chronic exposure to [more slowly](#) elevated temperatures. ~~(Figure~~
23 ~~12).~~

24 Although metabolic acclimation is unlikely over such short time periods
25 (apparent from the oxygen consumption data here, and also previous research

1 on long-term acclimation of *S. neumayeri* (Peck et al. 2014, Suckling et al.
2 2015)), short-term acclimation for some functions might be possible after an
3 initial shock response when temperatures are increased slowly. In our study,
4 the shock response did not appear to subside at faster rates of warming, and
5 instead mean functional thresholds were lower as warming rate increased.
6 These results suggest that functional and lethal limits are likely driven and
7 determined by different mechanisms. ~~Our data suggest, with limits to~~
8 ~~functioning are likely restricted related to by energy availability (as seen in this~~
9 ~~study) and .~~ Previous studies have shown that lethal limits are likely restricted
10 set by one or both of physiological processes or cellular and biochemical
11 mechanisms. At very rapid rates of warming, such as 1°C h⁻¹ or 1°C day⁻¹,
12 physiological mechanisms such as nervous ~~or~~ and circulatory failure appear to
13 to be the limiting factors (Young et al. 2006, Pörtner et al. 2007, Bilyk & DeVries
14 2011). At slower rates of warming (1°C per 3 days⁻¹ to 1°C month⁻¹) ~~or,~~ ~~or~~
15 ~~ecellular~~ and biochemical mechanisms such ~~as~~ as accumulation of toxic
16 products, e.g. protein carbonyls, enzyme tolerances or insufficiency of
17 chaperone proteins capacity appear to be limiting (Peck et al. 2009, Clark et al.
18 2017, 2018). Recently the factors setting thermal limits and responses to
19 warming have been shown to be highly species specific ~~(Clark et al. 2021,~~
20 ~~Collins et al. 2021~~ (Clark et al. 2021, Collins et al. 2021).

21 Our results also indicate that thermal sensitivity varies among different
22 ~~functions~~ key biological functions ~~have different thermal sensitivities. Mean~~
23 ~~thresholds were lowest for faecal production (mg day⁻¹) (4.9°C ± 1.6°C), and~~
24 ~~highest for righting (7.8°C ± 1.0°C).~~ For example, for the function of righting in
25 urchins, this function was maintained similar between treatments and to

1 ambient control conditions until temperatures reached 9.2°C for the fastest
2 rates of warming, and the highest breakpoint of 8.7°C was identified in the
3 slowest rates of warming ~~for this function.~~ Comparatively ~~However,~~ lower
4 thresholds were identified for the other functions related to digestion such as %
5 feeding or producing faeces. Variation between functional thresholds could be
6 related to function complexity, where a function involving multiple processes
7 would be more likely to fail (Pörtner et al. 2007, Stevens et al. 2010, Peck 2011).

8 Another explanation could be related to the extent to which fundamental
9 functions limit survival and fitness, where an organism's energy reserves allow
10 for short periods of negative energy balance. In Antarctic marine species such
11 periods of negative energy balance can be very long, extending to months or
12 even years of low food supply or starvation, because of the extreme
13 environmental seasonality and the very low metabolic energy use characteristic
14 of this fauna (Brockington et al. 2001, Harper & Peck 2003, Obermüller et al.
15 2010). However, being able to right provides immediate protection from
16 predation, equivalent to mechanisms such as the ability to stay attached to the
17 substratum in limpets (Morley et al. 2012b) or reburying in infaunal clams when
18 disturbed and removed from the sediment by ~~e.g., for example,~~ iceberg scour
19 (Peck et al. 2004). Finally, where a function has a higher metabolic energy
20 demand, it is more likely to be limited by food availability and energy delivery
21 capacity (van der Meer 2006, Morley et al. 2012a, Peck 2018).

22 The breakpoints identified for the mass of faeces produced might not indicate
23 a functional threshold. Instead, the initial high faecal production in the slowest
24 and intermediate warming rates is likely a result of the initial increase in
25 temperature causing food to move faster through the urchin, as also seen in

1 [the Antarctic plunderfish *Harpagifer antarcticus*](#) (Boyce et al. 2000). [This](#)
2 [elevation in faecal production was only observed when temperatures increased](#)
3 [initially, after which faecal production reduced to rates ~~comparative~~similar to](#)
4 [ambient control conditions. This effect was not observed in treatments with the](#)
5 [fastest rates of warming since these slight increases in temperature of 1°C –](#)
6 [2°C were likely not maintained long enough for gut passage rate to increase~~as~~](#)
7 [a result. Therefore, our results indicate that the breakpoints for faecal](#)
8 [production may not have any direct implications on functionality and instead](#)
9 [give evidence for the relationship between temperature and gut evacuation rate](#)
10 [\(GER\).](#)

11 In thermally stressed environments, animals usually increase their oxygen
12 uptake in order to meet increasing demands of functional processes (Gillooly
13 et al. 2001). However, when oxygen uptake is increased, yet functioning
14 deteriorates, it is hypothesised that this indicates a threshold where uptake,
15 ~~transport~~[transport](#), and delivery of O_2 [oxygen](#) can no longer meet the animal's
16 functional demands. This theory has been termed the oxygen and capacity
17 limited thermal tolerance [hypothesis](#) (OCLTT) (Pörtner et al. 2017). [This theory](#)
18 [focuses on the limitations set by the animal's physiology. ~~H,~~ however, as](#)
19 [temperature increases the concentration of oxygen diminishes, further reducing](#)
20 [the availability of oxygen to the animal and potentially amplifying the effects of](#)
21 [OCLTT. Reducing the concentration of oxygen in the water can limit functioning](#)
22 (Peck et al. 2007, Pörtner et al. 2007) [and as such, the functional thresholds](#)
23 [identified in this study may not only indicate thermal limits but may also be](#)
24 [influenced by the reduced oxygen content as temperatures increased. If oxygen](#)
25 [concentration was controlled and elevated throughout warming, the functional](#)

1 thresholds identified ~~may~~would likely be higher (Pörtner et al. 2006). However,
2 warmer oceans will be ~~conjuisive~~toaccompanied by lowerd oxygen
3 concentrations (Oschlies et al. 2018, Spicer et al. 2019) and as such the
4 functional thresholds determined in this study ~~may~~will be more
5 representative of a natural system than if oxygen were controlled.

6 ~~However, this theory is the subject of much debate and there are concerns over~~
7 ~~its use (Clark et al. 2013b, Clark & Mark 2017, Jutfelt et al. 2018).~~ In particular,
8 the theory makes the assumption that functioning is limited by energy derived
9 from respiration only and does not consider effects from the variation in the
10 energetic value of food ~~is the subject of much debate and there are concerns~~
11 ~~over its use (Clark et al. 2013b, Clark & Mark 2017, Jutfelt et al. 2018).~~

12 Evidence suggests that fFood availability and quality can ~~be~~ also be a

13 significant factor in determining functional scope (Welch et al. 1998, Lemoine
14 & Burkepile 2012, Cheng et al. 2018), whereby the ~~nutritional status and~~
15 condition of the animal could ~~effect~~affect energy delivery capacity similarly to
16 OCLTT. For example, feeding and digestive capacity limited the thermal
17 tolerance of juvenile spiny lobsters, *Sagmariasus verreauxi* (Fitzgibbon et al.
18 2017) and digestive capacity and food intake of individuals at high
19 temperatures related to depressed mitochondrial respiratory capacity in brown
20 trout *Salmo trutta* (Salin et al. 2016). The capacity to assimilate energy would
21 also play a role in determining ~~the energy delivery to tissues~~ and is determined
22 by physiological processes including consumption rate, absorption of food and
23 GER (Boyce et al. 2000, Angilletta 2001). Hence, assimilation itself is
24 energetically demanding and may limit functional thermal thresholds
25 (Sandersfeld et al. 2015, Salin et al. 2016).

1 Thus, OCLTT may be a possible mechanism for determining functional limits
2 observed in our experiments. However, there is no empirical support in our data
3 for this theory. In rapid warming both experiments and in natural heatwaves
4 MHWs, other factors are likely to be important, and obtaining sufficient energy
5 from food may be important for successful functioning. Impacts on animal
6 condition from warming may be especially important in highly seasonal polar
7 environments where warming in winter, when food supplies are scarce, would
8 increase energy use with little or no opportunity to mitigate the cost (Peck
9 2018). Species such as *S. neumayeri* that have been shown to spend periods
10 in winter up to 7 months without feeding (Brockington 2001) may be particularly
11 vulnerable to such impacts.

12 Our experiment included a period of 6 weeks without feeding to allow metabolic
13 activity to stabilise and be comparable between individuals. However, a caveat
14 to this initial standardisation of condition could influence the urchin's
15 physiological response to the warming in treatments. Nutritional status has
16 been shown to affect the reproductive state of *S. neumayeri*, with a reduction
17 in gonad index and maturation of gametes following 6 weeks without food,
18 comparative to animals foraging naturally in the environment (De Leij 2021).
19 Functional capacity has also been affected in other invertebrates under low
20 food coupled with environmental stress, for example the blue mussel *Mytilus*
21 *edulis* had a reduced ability to repair shells when high CO₂ was coupled with
22 low food (Melzner et al. 2011) and the green sea urchin *Strongylo-centrotus*
23 *droebachiensis*, exhibited severe metabolic acidosis when exposed to elevated
24 CO₂ with empty digestive tracts (Stumpff et al. 2012). Hence, we might consider
25 that the elevated temperatures coupled with the suboptimal nutritional status at

1 the start of the experiment, may have impacted the thermal limits of certain
2 functions. This would likely have resulted from a mismatch between a limited
3 energy supply and stores, and an increased energy demand of the animal.
4 However, the data in this study shows a reduction in the number of urchins
5 feeding as temperatures increase, suggesting that food was not the limiting
6 factor when this species approached its functional thermal limits.

7 From our analysis of the ~~Rothera environmental monitor (RaTS)~~ environmental
8 data, previous MHW events reached maximum temperatures of $2.3^{\circ}\text{C} \pm 0.36^{\circ}\text{C}$,
9 with onset rates of $0.3^{\circ}\text{C day}^{-1}$. Days at heatwave status have extended up to
10 95 days, and cumulative intensities (a combination of temperature intensity and
11 heatwave duration) have reached maxima of $54^{\circ}\text{C} \times \text{day}$ (Figure S2). ~~If we~~
12 ~~consider the latest climate change predictions (IPCC 2014, 2019)~~ Mean climate
13 temperatures are predicted to shift by $+2^{\circ}\text{C}$ by 2100, and with that, climate
14 extremes such as MHWs will increase in magnitude relative to this (IPCC 2014,
15 2019). ~~of the most likely scenario of $+2^{\circ}\text{C}$ ocean warming, then o~~ Our results
16 suggest that functions such as feeding and faecal egestion are likely to be
17 affected by MHW events by occurring in 2100, if not before, and this will include
18 increased metabolic demands with consequent impacts on annual energy
19 budgets. ~~Thus we would predict reduced energy availability for *S. neumayeri*~~
20 ~~from reduced feeding rates and food processing rates in warmer oceans, which~~
21 ~~is very likely to reduce survival in marginal environments. In our experiments,~~
22 ~~functional limits were reached within the cumulative intensities already~~
23 ~~experienced in the environment. However, this observation was limited to the~~
24 ~~most rapid warming rate of $1^{\circ}\text{C day}^{-1}$, a rate not yet reported from the Southern~~
25 ~~Ocean.~~

1 For a long-lived (>40 year (Brey et al. 1995)) and slow to mature (8-9 years
2 (Peck 2018)) species such as *S. neumayeri*, there will be less scope for
3 phenotypic and genotypic adaptations to a warming climate as might be
4 possible for short-lived and rapidly maturing species (Peck 2011, Donelson et
5 al. 2012, Salinas & Munch 2012). However, there may still be opportunity for *S.*
6 *neumayeri* to ~~acclimate~~adapt to a warmer world. Within 80 years (2020 - 2100),
7 eight generations of *S. neumayeri* will have succeeded the present population,
8 and in the year 2100, the 5th, 6th and 7th generation could be present and
9 reproducing in populations around Antarctica. If we consider the evidence of *S.*
10 *neumayeri*'s capacity to acclimate, it may be possible for this species to
11 acclimate and adapt successfully to ~~and~~function atin a +2°C warmer world
12 (Morley et al. 2016).; ~~however~~it is still uncertain, ~~however~~, how this species
13 will respond to acute warming, like that experienced during MHWs, in this
14 warmer climate. The data in this study cannot predict the implications of
15 acclimation and adaptation ~~and~~on the subsequent tolerance to MHWs for *S.*
16 *neumayeri*. Instead, the data provides insight into the effect of onset rate of
17 acute warming, the thermal vulnerability of key biological functions, and the
18 difference between critical thermal limits and functional thermal limits. Thus,
19 according to our data we ~~could predict~~see reduced energy availability for *S.*
20 *neumayeri* ~~from reduced~~from changes in feeding ~~rates~~and food processing
21 rates ~~during MHWs~~ in warmer oceans, which ~~is~~would very likely ~~to~~reduce
22 survival in marginal environments.

23 Following the results from this study, it would be important to explore recovery
24 following MHW events. Our data indicate reduced functioning as temperatures
25 are raised across all rates of warming. ~~H~~, however, the ability and rate of *S.*

1 neumayeri to resume 'normal' functioning if returned to ambient temperatures
2 is uncertain. It has been shown that the marine snail, *Littorina littorea*, loses
3 motility under thermal stress, however if temperatures are lowered again, this
4 function returns (Hamby 1975). To resume a single function may not indicate
5 full recovery, and our study shows that different biological functions have
6 varying thermal tolerances. As such, performance of all functions, including
7 metabolic activity, would need to return to baseline levels for an animal to
8 recover completely (Walter et al. 2013). Developing our understanding of
9 recovery following acute warming and even the effects of repeat MHW events,
10 could better predict the long-term implications of MHWs for this species.

11 It is important to acknowledge ~~note~~ that the functional and critical limits
12 measured in this study are likely an example of a 'best case scenario'.
13 Experiments such as these can only predict the isolated effects of one variable,
14 However, the additional energetic costs associated with physical factors such
15 as salinity change and biological factors including varying food quality and
16 quantity, species interactions, diseases and scavenging for food, must need to
17 be considered ~~included~~ before we can obtain dependable predictions for 'real
18 world' scenarios that give information relevant to the ~~wide range of~~ variable
19 conditions experienced across a species distribution range. What is limiting at
20 the range margins for a species will differ from core areas (Kolzenburg et al.
21 2021). ~~The results reported here are best-case scenarios for this species~~
22 ~~*S. neumayeri*, with a high protein food supply, which likely resulted in functional~~
23 ~~limits at the high end for this species.~~

24 ~~This study~~ Our data highlights that the deterioration of functioning with
25 warming ~~when temperatures are raised~~, especially heat waves ~~during MHWs~~,

1 has implications for long term survival, and physiological functions. Therefore,
2 functioning ~~at processes~~ should be considered when determining organism
3 thermal limits, rather than traditional critical thermal limits. Our findings show
4 that fitness cannot be determined from a single function and instead functions
5 vary in thermal sensitivity. A whole organism approach to functional fitness is
6 therefore necessary, considering functional complexity, ~~importance~~ importance,
7 and energetic demand. Our results suggest ~~that, that~~ contrary to the relationship
8 between critical thermal limits and onset rate, functional degradation occurs at
9 lower temperatures when exposed to rapid warming (1°C day⁻¹). Therefore,
10 when investigating the impact of ~~marine heatwaves~~ MHWs on organisms and
11 populations, it is important to consider the key features of the heatwave event,
12 including the onset rate, exposure duration, and how these characteristics act
13 together to determine functional thresholds.

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24 **AUTHOR CONTRIBUTIONS**

1 L.S.P and R.D conceived and designed the study. R.D carried out the
2 practical work and data processing. R.D, L.J.G and L.S.P analysed the data,
3 drafted the manuscript and approved its publication.

4 **COMPETING INTERESTS**

5 The authors declare no competing interests.

6 **LITERATURE CITED**

- 7 Allen JL, Chown SL, Janion-Scheepers C, Clusella-Trullas S (2016)
8 Interactions between rates of temperature change and acclimation affect
9 latitudinal patterns of warming tolerance. *Conserv Physiol* 4:1–14.
- 10 Ardor Bellucci LM, Smith NF (2019) Crawling and righting behavior of the
11 subtropical sea star *Echinaster (Othilia) graminicola*: effects of elevated
12 temperature. *Mar Biol* 166:1–9.
- 13 Azad AK, Pearce CM, McKinley RS (2011) Effects of diet and temperature on
14 ingestion, absorption, assimilation, gonad yield, and gonad quality of the
15 purple sea urchin (*Strongylocentrotus purpuratus*). *Aquaculture* 317:187–
16 196.
- 17 Barnes DKA, Brockington S (2003) Zoobenthic biodiversity, biomass and
18 abundance at Adelaide Island, Antarctica. *Mar Ecol Prog Ser* 249:145–
19 155.
- 20 Bilyk KT, DeVries AL (2011) Heat tolerance and its plasticity in Antarctic
21 fishes. *Comp Biochem Physiol - A Mol Integr Physiol* 158:382–390.
- 22 Bopp L, Resplandy L, Orr JC, Doney SC, Dunne JP, Gehlen M, Halloran P,
23 Heinze C, Ilyina T, Séférian R, Tjiputra J, Vichi M (2013) Multiple
24 stressors of ocean ecosystems in the 21st century: Projections with
25 CMIP5 models. *Biogeosciences* 10:6225–6245.
- 26 Bosch I, Beauchamp KA, Steele ME, Pearse JS (1987) Development,
27 metamorphosis, and seasonal abundance of embryos and larvae of the
28 Antarctic sea urchin *Sterechinus neumayeri*. *Biol Bull* 173:126–135.

- 1 Boyce SJ, Murray AWA, Peck LS (2000) Digestion rate, gut passage time and
2 absorption efficiency in the Antarctic spiny plunderfish. *J Fish Biol*
3 57:908–929.
- 4 Brey T, Pearse J, Basch L, McClintock J, Slattery M (1995) Growth and
5 production of *Sterechinus neumayeri* (Echinoidea: Echinodermata) in
6 McMurdo Sound, Antarctica. *Mar Biol* 124:279–292.
- 7 Brockington S (2001) The seasonal ecology and physiology of *Sterechinus*
8 *neumayeri* (Echinodermata; Echinoidea) at Adelaide Island, Antarctica.
9 PhD thesis The Open University.
- 10 Brockington S, Clarke A, Chapman ALG (2001) Seasonality of feeding and
11 nutritional status during the austral winter in the Antarctic sea urchin
12 *Sterechinus neumayeri*. *Mar Biol* 139:127–138.
- 13 Chapelle G, Peck LS, Clarke A (1994) Effects of feeding and starvation on the
14 metabolic rate of the necrophagous Antarctic amphipod *Waldeckia obesa*
15 (Chevreux, 1905). *J Exp Mar Bio Ecol* 183:63–76.
- 16 Clark MS, Peck LS (2009) Triggers of the HSP70 stress response:
17 environmental responses and laboratory manipulation in an Antarctic
18 marine invertebrate (*Nacella concinna*). *Cell Stress Chaperones* 14:649–
19 660.
- 20 Clark MS, Peck LS, Thyrring J (2021) Resilience in Greenland intertidal
21 *Mytilus*: The hidden stress defense. *Sci Total Environ* 767:144366.
- 22 Clark MS, Sommer U, Sihra JK, Thorne MAS, Morley SA, King M, Viant MR,
23 Peck LS (2017) Biodiversity in marine invertebrate responses to acute
24 warming revealed by a comparative multi-omics approach. *Glob Chang*
25 *Biol* 23:318–330.
- 26 Clark MS, Suckling CC, Cavallo A, Mackenzie CL, Thorne MAS, Davies AJ,
27 Peck LS (2019a) Molecular mechanisms underpinning transgenerational
28 plasticity in the green sea urchin *Psammechinus miliaris*. *Sci Rep* 9:1–12.
- 29 Clark MS, Thorne MAS, King M, Hipperson H, Hoffman JI, Peck LS (2018)
30 Life in the intertidal: Cellular responses, methylation and epigenetics.
31 *Funct Ecol* 32:1982–1994.

- 1 Clark MS, Villota Nieva L, Hoffman JI, Davies AJ, Trivedi UH, Turner F,
2 Ashton G V, Peck LS (2019b) Lack of long-term acclimation in Antarctic
3 encrusting species suggests vulnerability to warming. *Nat Commun* 10:1–
4 10.
- 5 Clarke A, Meredith MP, Wallace MI, Brandon MA, Thomas DN (2008)
6 Seasonal and interannual variability in temperature, chlorophyll and
7 macronutrients in northern Marguerite Bay, Antarctica. *Deep Res Part II*
8 *Top Stud Oceanogr* 55:1988–2006.
- 9 Collins M, Peck LS, Clark MS (2021) Large within, and between, species
10 differences in marine cellular responses: Unpredictability in a changing
11 environment. *Sci Total Environ* 794:148594.
- 12 De Leij R (2021) Functional response of the Antarctic sea urchin, *Sterechinus*
13 *neumayeri*, to environmental change and extreme events in the context of
14 a warming climate (In Press). PhD Thesis, University of Southampton
- 15 De Leij R, Peck LS, Grange LJ (2021) Multiyear trend in reproduction
16 underpins interannual variation in gametogenic development of an
17 Antarctic urchin. *Sci Rep* 11:1–13.
- 18 Deschaseaux ESM, Taylor AM, Maher WA, Davis AR (2010) Cellular
19 responses of encapsulated gastropod embryos to multiple stressors
20 associated with climate change. *J Exp Mar Bio Ecol* 383:130–136.
- 21 Donelson JM, Munday PL, McCormick MI, Pitcher CR (2012) Rapid
22 transgenerational acclimation of a tropical reef fish to climate change. *Nat*
23 *Clim Chang* 2:30–32.
- 24 Dunlop KM, Barnes DKA, Bailey DM (2014) Variation of scavenger richness
25 and abundance between sites of high and low iceberg scour frequency in
26 Ryder Bay, West Antarctic Peninsula. *Polar Biol* 37:1741–1754.
- 27 Ferrarini A (2011) Detecting ecological breakpoints: a new tool for piecewise
28 regression. *Comput Ecol Softw* 1:121–124.
- 29 Fitzgibbon QP, Simon CJ, Smith GG, Carter CG, Battaglione SC (2017)
30 Temperature dependent growth , feeding , nutritional condition and
31 aerobic metabolism of juvenile spiny lobster, *Sagmariasus verreauxi*.

1 Comp Biochem Physiol Part A 207:13–20.

2 Garrabou J, Coma R, Bensoussan N, Bally M, Chevaldonné P, Cigliano M,
3 Diaz D, Harmelin JG, Gambi MC, Kersting DK, Ledoux JB, Lejeune C,
4 Linares C, Marschal C, Pérez T, Ribes M, Romano JC, Serrano E,
5 Teixido N, Torrents O, Zabala M, Zuberer F, Cerrano C (2009) Mass
6 mortality in Northwestern Mediterranean rocky benthic communities:
7 Effects of the 2003 heat wave. *Glob Chang Biol* 15:1090–1103.

8 Gillooly JF, Brown JH, West GB, Savage VM, Charnov EL (2001) Effects of
9 size and temperature on metabolic rate. *Science* (80-) 293:2248–2251.

10 Hamby RJAY (1975) Heat Effects on a Marine Snail. *Biol Bull* 149:331–347.

11 Harper EM, Peck LS (2003) Predatory behaviour and metabolic costs in the
12 Antarctic muricid gastropod *Trophon longstaffi*. *Polar Biol* 26:208–217.

13 IPCC (2014) Climate Change 2014: Synthesis Report. Contribution of
14 Working Groups I, II and III to the Fifth Assessment Report of the
15 Intergovernmental Panel on Climate Change. Core Writing Team
16 Pachauri RK and Meyer LA (ed) IPCC, Geneva, Switzerland, 151pp.

17 IPCC (2019) Summary for Policymakers. In: Climate Change and Land: an
18 IPCC special report on climate change, desertification, land degradation,
19 sustainable land management, food security, and greenhouse gas fluxes
20 in terrestrial ecosystems. Shukla PR, Skea J, Calvo Buendia E, Masson-
21 Delmotte V, Pörtner HO, Roberts DC, Zhai P, Slade R, Connors S, van
22 Diemen R, Ferrat M, Haughey E, Luz S, Neogi S, Pathak M, Petzold J,
23 Portugal Pereira J, Vyas P, Huntley E, Kissick K, Malley J (ed) In press.

24 Janecki T, Kidawa A, Potocka M (2010) The effects of temperature and
25 salinity on vital biological functions of the Antarctic crustacean *Serolis*
26 *polita*. *Polar Biol* 33:1013–1020.

27 Kingsolver JG, Umbanhowar J (2018) The analysis and interpretation of
28 critical temperatures. *J Exp Biol* 221.

29 Kolzenburg R, D’Amore F, McCoy SJ, Ragazzola F (2021) Marginal
30 populations show physiological adaptations and resilience to future
31 climatic changes across a North Atlantic distribution. *Environ Exp Bot*

1 188:104522.

2 Lenihan HS, Peterson CH, Miller RJ, Kayal M, Potoski M (2018) Biotic
3 disturbance mitigates effects of multiple stressors in a marine benthic
4 community. *Ecosphere* 9.

5 Liu H, Kelly MS, Cook EJ, Black K, Orr H, Zhu JX, Dong SL (2007) The effect
6 of diet type on growth and fatty-acid composition of sea urchin larvae, I.
7 *Paracentrotus lividus* (Lamarck, 1816) (Echinodermata). *Aquaculture*
8 264:247–262.

9 Marshall J, Scott JR, Armour KC, Campin JM, Kelley M, Romanou A (2015)
10 The ocean's role in the transient response of climate to abrupt
11 greenhouse gas forcing. *Clim Dyn* 44:2287–2299.

12 McClintock J (1994) Trophic biology of Antarctic shallow-water echinoderms.
13 *Mar Ecol Prog Ser* 111:191–202.

14 van der Meer J (2006) An introduction to Dynamic Energy Budget (DEB)
15 models with special emphasis on parameter estimation. *J Sea Res*
16 56:85–102.

17 Melzner F, Stange P, Trubenbach K, Thomsen J, Casties I, Panknin U, Gorb
18 SN, Gutowska MA (2011) Food Supply and Seawater pCO₂ Impact
19 Calcification and Internal Shell Dissolution in the Blue Mussel *Mytilus*
20 *edulis*. *PLoS One* 6:e24223.

21 Morley SA, Lai CH, Clarke A, Tan KS, Thorne MAS, Peck LS (2014) Limpet
22 feeding rate and the consistency of physiological response to
23 temperature. *J Comp Physiol B Biochem Syst Environ Physiol* 184:563–
24 570.

25 Morley SA, Martin SM, Bates AE, Clark MS, Ericson J, Lamare M, Peck LS
26 (2012a) Spatial and temporal variation in the heat tolerance limits of two
27 abundant Southern Ocean invertebrates. *Mar Ecol Prog Ser* 450:81–92.

28 Morley SA, Martin SM, Day RW, Ericson J, Lai CH, Lamare M, Tan KS,
29 Thorne MAS, Peck LS (2012b) Thermal Reaction Norms and the Scale of
30 Temperature Variation: Latitudinal Vulnerability of Intertidal Nacellid
31 Limpets to Climate Change. *PLoS One* 7:7–10.

- 1 Morley SA, Suckling CC, Clark MS, Cross EL, Peck LS (2016) Long-term
2 effects of altered pH and temperature on the feeding energetics of the
3 Antarctic sea urchin, *Sterechinus neumayeri*. *Biodiversity* 17:34–45.
- 4 Muggeo VMR (2008) Segmented: An R Package to Fit Regression Models
5 with Broken-Line Relationships. *R News* 3:343–4.
- 6 Myrvoll-Nilsen E, Fredriksen HB, Sørbye SH, Rypdal M (2019) Warming
7 Trends and Long-Range Dependent Climate Variability Since Year 1900:
8 A Bayesian Approach. *Front Earth Sci* 7:1–8.
- 9 Obermüller BE, Morley SA, Barnes DKA, Peck LS (2010) Seasonal
10 physiology and ecology of Antarctic marine benthic predators and
11 scavengers. *Mar Ecol Prog Ser* 415:109–126.
- 12 Oliver ECJ, Burrows MT, Donat MG, Sen Gupta A, Alexander L V., Perkins-
13 Kirkpatrick SE, Benthuyesen JA, Hobday AJ, Holbrook NJ, Moore PJ,
14 Thomsen MS, Wernberg T, Smale DA (2019) Projected Marine
15 Heatwaves in the 21st Century and the Potential for Ecological Impact.
16 *Front Mar Sci* 6:1–12.
- 17 Oliver ECJ, Donat MG, Burrows MT, Moore PJ, Smale DA, Alexander L V.,
18 Benthuyesen JA, Feng M, Sen Gupta A, Hobday AJ, Holbrook NJ,
19 Perkins-Kirkpatrick SE, Scannell HA, Straub SC, Wernberg T (2018)
20 Longer and more frequent marine heatwaves over the past century. *Nat*
21 *Commun* 9:1–12.
- 22 Oschlies A, Brandt P, Stramma L, Schmidtko S (2018) Drivers and
23 mechanisms of ocean deoxygenation. *Nat Geosci* 11:467–473.
- 24 Payton SL, Johnson PD, Jenny MJ (2016) Comparative physiological,
25 biochemical and molecular thermal stress response profiles for two
26 unionid freshwater mussel species. *J Exp Biol* 219:3562–3574.
- 27 Peck LS (2018) Antarctic Marine Biodiversity: Adaptations, Environments and
28 Responses to Change. *Oceanogr Mar Biol An Annu Rev* 56:105–236.
- 29 Peck LS (2011) Organisms and responses to environmental change. *Mar*
30 *Genomics* 4:237–243.
- 31 Peck LS (2005) Prospects for survival in the Southern Ocean: Vulnerability of

- 1 benthic species to temperature change. *Antarct Sci* 17:497–507.
- 2 Peck LS (1989) Temperature and basal metabolism in two Antarctic marine
3 herbivores. *J Exp Mar Bio Ecol* 127:1–12.
- 4 Peck LS, Bullough LW (1993) Growth and population structure in the infaunal
5 bivalve *Yoldia eightsi* in relation to iceberg activity at Signy Island,
6 Antarctica. *Mar Biol* 117:235–241.
- 7 Peck LS, Clark MS, Morley SA, Massey A, Rossetti H (2009) Animal
8 temperature limits and ecological relevance: Effects of size, activity and
9 rates of change. *Funct Ecol* 23:248–256.
- 10 Peck LS, Morley SA, Pörtner H-O, Clark MS (2007) Thermal limits of
11 burrowing capacity are linked to oxygen availability and size in the
12 Antarctic clam *Laternula elliptica*. *Oecologia* 154:479–484.
- 13 Peck LS, Morley SA, Richard J, Clark MS (2014) Acclimation and thermal
14 tolerance in Antarctic marine ectotherms. *J Exp Biol* 217:16–22.
- 15 Peck LS, Webb KE, Bailey DM (2004) Extreme sensitivity of biological
16 function to temperature. *Funct Ecol* 18:625–630.
- 17 Peck LS, Webb KE, Miller A, Clark MS, Hill T (2008) Temperature limits to
18 activity, feeding and metabolism in the Antarctic starfish *Odontaster*
19 *validus*. *Mar Ecol Prog Ser* 358:181–189.
- 20 Piepho HP, Ogutu JO (2003) Inference for the break point in segmented
21 regression with application to longitudinal data. *Biometrical J* 45:591–601.
- 22 Pierrat B, Saucède T, Laffont R, De Ridder C, Festeau A, David B (2012)
23 Large-scale distribution analysis of Antarctic echinoids using ecological
24 niche modelling. *Mar Ecol Prog Ser* 463:215–230.
- 25 Pörtner H-O, Bock C, Mark FC (2017) Oxygen- & capacity-limited thermal
26 tolerance: Bridging ecology & physiology. *J Exp Biol* 220:2685–2696.
- 27 Pörtner H-O, Peck LS, Hirse T (2006) Hyperoxia alleviates thermal stress in
28 the Antarctic bivalve, *Laternula elliptica*: Evidence for oxygen limited
29 thermal tolerance. *Polar Biol* 29:688–693.
- 30 Pörtner H-O, Peck LS, Somero G (2007) Thermal limits and adaptation in

- 1 marine Antarctic ectotherms: An integrative view. *Philos Trans R Soc B*
2 *Biol Sci* 362:2233–2258.
- 3 Robertson R, El-Haj AJ, Clarke A, Peck LS, Taylor E (2001) The effects of
4 temperature on metabolic rate and protein synthesis following a meal in
5 the isopod *Glyptonotus antarcticus* Eights (1852). *Polar Biol* 24:677–686.
- 6 Rubio-Portillo E, Izquierdo-Muñoz A, Gago JF, Rosselló-Mora R, Antón J,
7 Ramos-Esplá AA (2016) Effects of the 2015 heat wave on benthic
8 invertebrates in the Tabarca Marine Protected Area (southeast Spain).
9 *Mar Environ Res* 122:135–142.
- 10 Salinas S, Munch SB (2012) Thermal legacies: Transgenerational effects of
11 temperature on growth in a vertebrate. *Ecol Lett* 15:159–163.
- 12 Schlegel RW, Smit AJ (2018) HeatwaveR: A central algorithm for the
13 detection of heatwaves and cold-spells. *J Open Source Softw* 3:821.
- 14 Spicer JI, Morley SA, Bozinovic F (2019) Physiological diversity, biodiversity
15 patterns and global climate change: Testing key hypotheses involving
16 temperature and oxygen. *Philos Trans R Soc B Biol Sci* 374:8–11.
- 17 Stanwell-Smith D, Peck LS (1998) Temperature and embryonic development
18 in relation to spawning and field occurrence of larvae of three Antarctic
19 echinoderms. *Biol Bull* 194:44–52.
- 20 Stevens MM, Jackson S, Bester SA, Terblanche JS, Chown SL (2010)
21 Oxygen limitation and thermal tolerance in two terrestrial arthropod
22 species. *J Exp Biol* 213:2209–2218.
- 23 Suckling CC, Clark MS, Beveridge C, Brunner L, Hughes AD, Harper EM,
24 Cook EJ, Davies AJ, Peck LS (2014) Experimental influence of pH on the
25 early life-stages of sea urchins II: Increasing parental exposure times
26 gives rise to different responses. *Invertebr Reprod Dev* 58:161–175.
- 27 Suckling CC, Clark MS, Richard J, Morley SA, Thorne MAS, Harper EM, Peck
28 LS (2015) Adult acclimation to combined temperature and pH stressors
29 significantly enhances reproductive outcomes compared to short-term
30 exposures. *J Anim Ecol* 84:773–784.
- 31 Terblanche JS, Deere JA, Clusella-Trullas S, Janion C, Chown SL (2007)

1 Critical thermal limits depend on methodological context. Proc R Soc B
 2 Biol Sci 274:2935–2942.

3 Venables HJ, Clarke A, Meredith MP (2013) Wintertime controls on summer
 4 stratification and productivity at the western Antarctic Peninsula. Limnol
 5 Oceanogr 58:1035–1047.

6 Walter J, Jentsch A, Beierkuhnlein C, Kreyling J (2013) Ecological stress
 7 memory and cross stress tolerance in plants in the face of climate
 8 extremes. Environ Exp Bot 94:3–8.

9 Young JS, Peck LS, Matheson T (2006) The effects of temperature on
 10 walking and righting in temperate and Antarctic crustaceans. Polar Biol
 11 29:978–987.

12 Zupo V, Glaviano F, Paolucci M, Ruocco N, Polese G, Di Cosmo A, Costantini
 13 M, Mutalipassi M (2019) Roe enhancement of *Paracentrotus lividus*:
 14 Nutritional effects of fresh and formulated diets. Aquac Nutr 25:26–38.

15

16 **TABLES**

17 Table 1: Summary statistics for linear regression relationships between the
 18 measured functions of *Sterechinus neumayeri* and temperature. β indicates the
 19 slope of the linear regression lines before the breakpoint (Slope_1) and after
 20 the breakpoint (Slope_2); SE_a indicates standard error for the intercept and
 21 slopes; df = degrees of freedom; bold p-values indicate significant relationships
 22 ($p < 0.05$) between temperature and the variable measured and bold Davies p-
 23 values represent a significant change ($p < 0.05$) in the gradient of the slope of
 24 segmented regressions. Values in the column BP indicate the localisation of
 25 the breakpoint or else NA indicates a single linear regression; SE_b (standard
 26 error) and R^2 refers to the goodness of fit for the entire model.

<u>Function-Function</u>	β	$\frac{SE_a S}{E_a}$	<u>P-value</u> p-value	$\frac{BP}{P}$	$\frac{SE_b S}{E_b}$	R^2	<u>Davies</u>
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							p-value Davies p-value
<u>Individuals feeding, 1°C day⁻¹</u> (Intercept) Slope_1 Slope_2	89.0 3.45 -12.9	25.4 10.5 2.35	df=3 0.039 0.764 0.012	df=3 4.04 0	14.94 4.9	0.894 0.894	0.329 0.329
<u>Individuals feeding, 0.5°C day⁻¹</u> (Intercept) Slope_1 Slope_2	110.3 -6.34 -11.5	12.7 3.14 1.05	df=7 <0.001 0.083 <0.001	df=7 6.26 2	6.786 -78	0.964 0.964	0.301 0.301
<u>Individuals feeding, 0.3°C day⁻¹</u> (Intercept) Slope_1 Slope_2	95.3 -2.73 -20.3	7.53 1.38 2.92	df=12 <0.001 0.071 <0.001	df=12 8.28 2	8.488 -48	0.922 0.922	0.001 0.001
<u>Individuals producing faeces, 1°C day⁻¹</u> (Intercept) Slope_1 Slope_2	-29.0 24.1 -13.3	23.1 9.54 2.13	df=3 0.298 0.085 0.008	df=3 5.25 2	13.54 3.5	0.881 0.882	0.019 0.019
<u>Individuals producing faeces, 0.5°C day⁻¹</u> (Intercept) Slope_1 Slope_2	34.0 13.3 -10.3	28.6 8.54 8.68	df=7 0.274 0.162 <0.001	df=7 4.54 5	12.14 2.4	0.844 0.844	0.039 0.039
<u>Individuals producing faeces, 0.3°C day⁻¹</u> (Intercept) Slope_1 Slope_2	-0.306 -18.6 -19.0	2.02 4.29 4.29	df=12 <0.001 0.882 <0.001	df=12 8.38 3	12.54 2.5	0.762 0.762	0.006 0.006
<u>Faeces produced, 1°C day⁻¹</u> (Intercept)	0.645 -0.040	0.137 0.027	df=14 <0.001	NAN A	0.216 4.35	0.071 0.364	0.858 NAN

<u>Slope_1</u> Faeces produced, 1°C day ⁻¹ (Intercept) Slope_1	3.63 -0.31	0.422 0.057	0.165-df =47 <0.001 <0.001				
<u>Faeces produced, 0.5°C day⁻¹</u> (Intercept) Slope_1 <u>Slope_2</u> Faeces produced, 0.5°C day ⁻¹ (Intercept) Slope_1 Slope_2	1.52 -0.23 -0.06 9.53 -1.33 -0.118	0.214 0.072 0.025 0.774 0.184 0.185	df=31 <0.001 0.007 0.016-d f=74 <0.001 <0.001 0.526	4.65 9	1.114 -92	0.664 0.611	0.043<0.001
<u>Faeces produced, 0.3°C day⁻¹</u> (Intercept) Slope_1 <u>Slope_2</u> Faeces produced, 0.3°C day ⁻¹ (Intercept) Slope_1 Slope_2	3.54 -0.718 -0.051 18.4 -4.86 -0.232	0.509 0.202 0.020 2.54 1.04 0.065	df=34 <0.001 0.001 0.012-d f=87 <0.001 <0.001	3.33 8	0.294 4.57	0.729 0.673	<0.001<0.001
<u>Time taken to right, 1°C day⁻¹</u> (Intercept) <u>Slope_1</u> Time taken to right, 1°C day ⁻¹ (Intercept) Slope_1 Slope_2	-8.60 6.83 11.7 0.897 15.2	9.04 1.35 7.33 2.03 2.86	df=26 0.350 <0.001-d f=44 0.119 0.664 <0.001	NA6 8	23.34 8.19	0.476 0.553	NA <0.001
<u>Time taken to right, 0.5°C day⁻¹</u> (Intercept) <u>Slope_1</u> Time taken to right, 0.5°C day ⁻¹ (Intercept) Slope_1	8.88 2.61 9.54 2.42	5.03 0.731 4.46 0.649	df=26 0.089 0.001 df =51 0.037 <0.001	NA A	13.14 6.2	0.302 0.198	NA NA
<u>Time taken to right, 0.3°C day⁻¹</u> (Intercept) Slope_1 <u>Slope_2</u> Time taken to right, 0.3°C day ⁻¹ (Intercept) Slope_1 Slope_2	14.6 0.384 55.7 14.2 0.537 64.1	20.1 3.66 13.8 15.0 2.77 10.9	df=25 0.237 0.459 <0.001 - df=54 0.350 0.847 <0.001	8.78 7	0.556 45.8	0.588 0.589	<0.001<0.001
<u>Oxygen consumption, 1°C day⁻¹</u> (Intercept) <u>Slope_1</u> Oxygen consumption, 1°C day ⁻¹ (Intercept) Slope_1	1.64 1.50 -0.042 0.177	1.76 0.248 0.227 0.030	df=28 0.358 <0.001 d f=45 0.856 <0.001	NA A	4.640 -581	0.551 0.425	NA NA
<u>Oxygen consumption, 0.5°C day⁻¹</u> (Intercept) Slope_1	4.29 0.611	1.10 0.134	df=33 <0.001 <0.001 df=52	NA7 0	3.170 -169	0.368 0.804	NA0.260

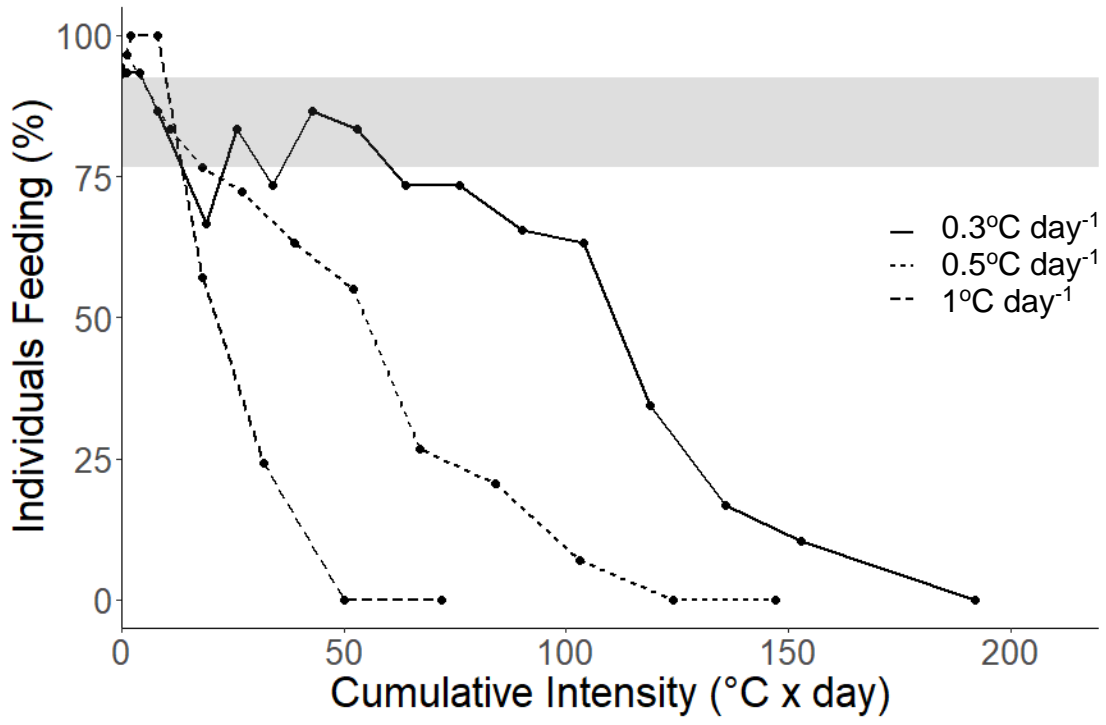
¹ Reporting only a single slope (Slope_1) indicates that no breakpoint was detected in the regression and statistics for a single linear regression model is reported for the data instead.

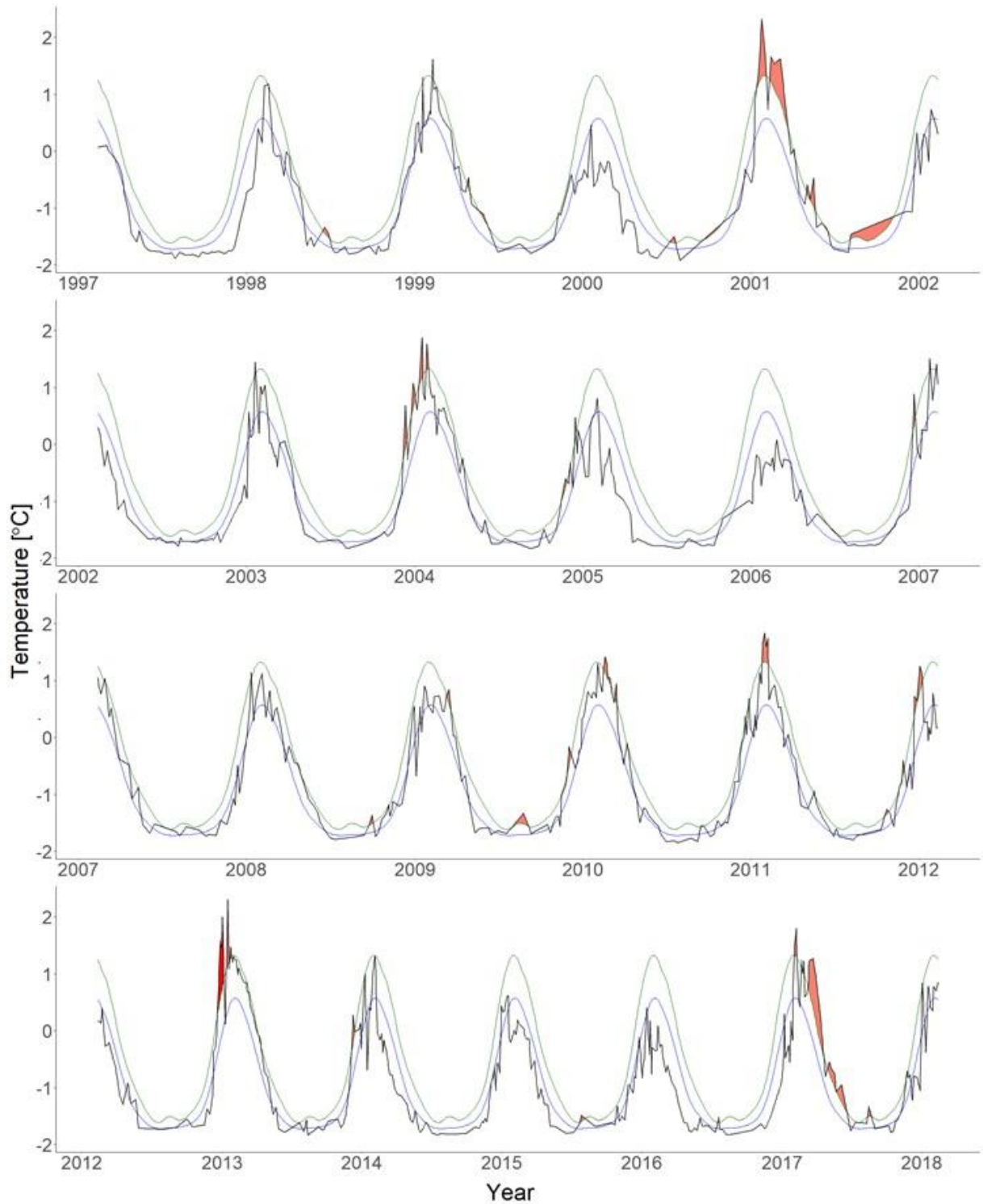
Oxygen consumption, 0.5°C day ⁻¹	0.264	0.172	0.135				
(Intercept)	0.069	0.040	0.088				
Slope_1	0.120	0.013	<0.001				
Slope_2							
Oxygen consumption, 0.3°C day ⁻¹			df=28				
(Intercept)			0.022				
Slope_1	3.30	1.36	<0.001	NAN	3.490	0.471	NANA
Oxygen consumption, 0.3°C day ⁻¹	0.957	0.185	df=44	A	.394	0.339	
(Intercept)	0.327	0.155	0.040				
Slope_1	0.100	0.020	<0.001				

1 Table 2: Cumulative intensity (°C x day) in relation to temperature for each
2 treatment. Colours represent cumulative intensity magnitude, where green
3 indicates low magnitude and red indicates high magnitude, relative to those
4 experienced in the experiment. Cumulative intensity calculated from following
5 regressions: 1°C day⁻¹: (0.49T²) - (0.99T), 0.5°C day⁻¹: (0.96T²) - (1.52T) and
6 0.3°C day⁻¹: (1.47T²) - (2.22T), where T=Temperature (Supplementary
7 Materials, Figure S3).

Temperature (°C)	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15
Treatment	Cumulative Intensity (°C x day)														
1°C day ⁻¹	4	3	6	10	15	21	28	36	45	55	66	78	91	105	120
0.5°C day ⁻¹	2	7	13	21	31	43	57	72	90	110	131	155	180	207	237
0.3°C day ⁻¹	3	9	18	30	45	63	84	108	135	165	198	234	273	315	361

1 FIGURES

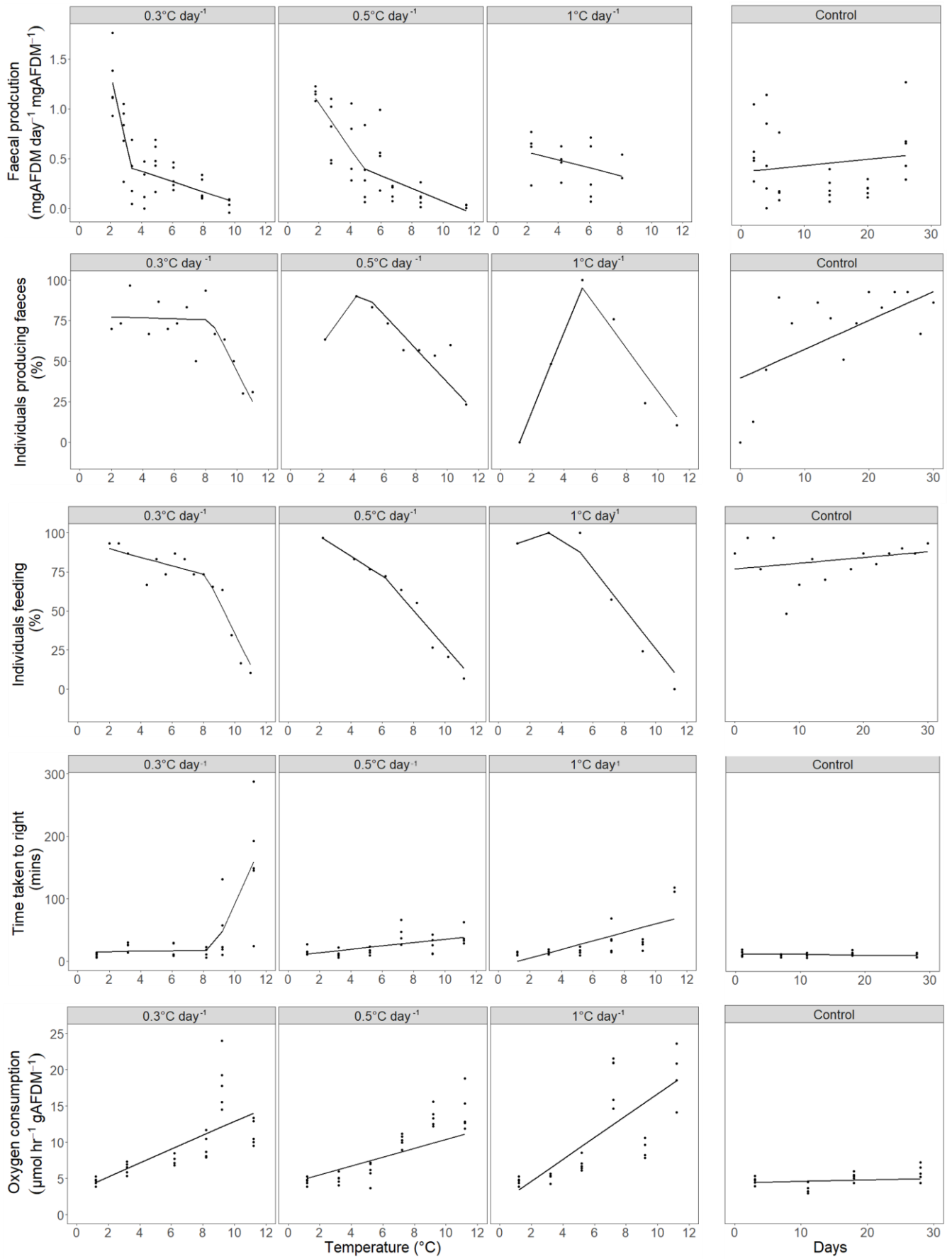




1

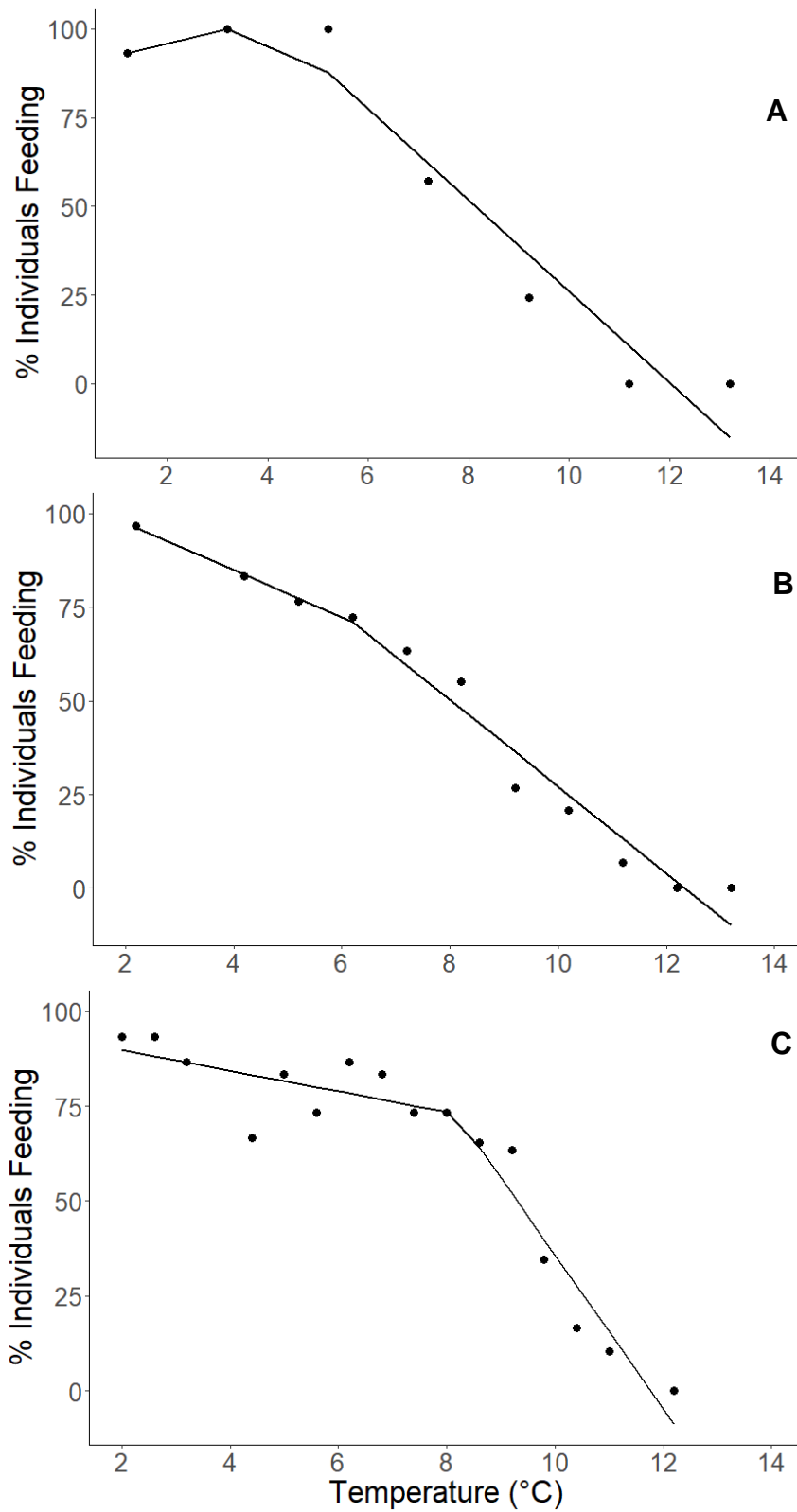
2 Figure 1: Times-series of temperatures (°C) experienced in Ryder Bay,
 3 Antarctica, at depths of 15 m, represented by the black lines. The data are split
 4 into panels to cover the entire span of the time-series, where the x-axis
 5 represents time in years. Blue lines represent the seasonal climatology of the

1 region based on the full time-series of daily temperatures (1997 – 2018). Green
2 lines represent the seasonally varying threshold for a marine heatwave (90th
3 percentile). Temperatures exceeding the threshold for ≥ 5 days are highlighted
4 in red and indicate the occurrence of a marine heatwave.



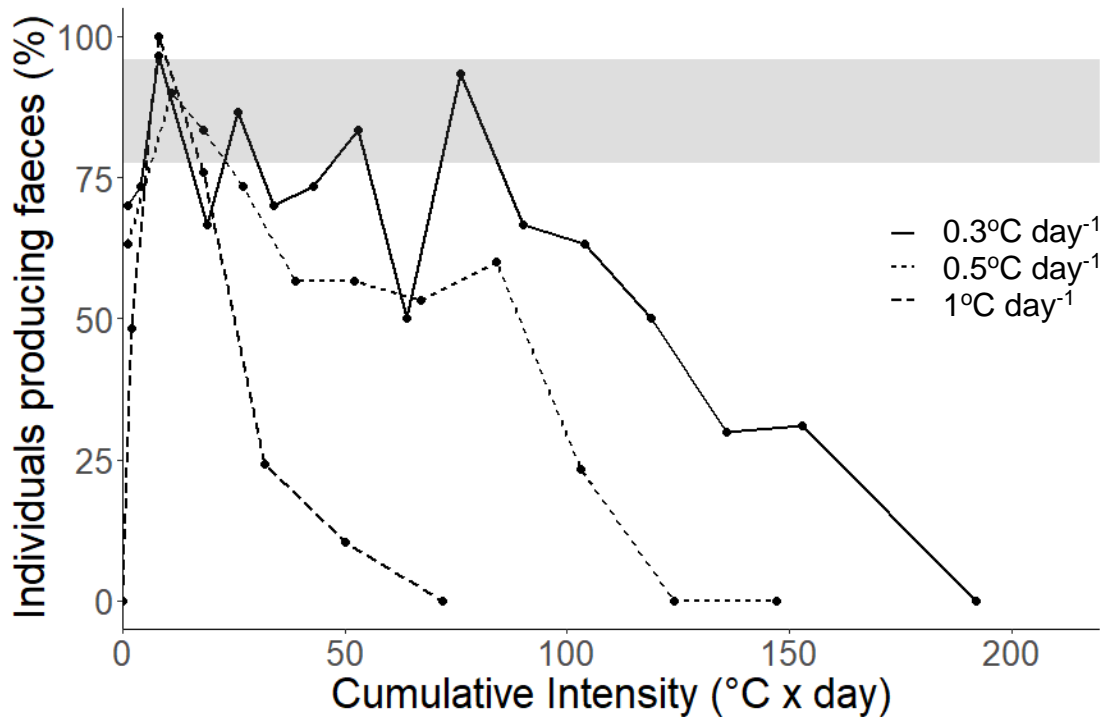
1 Figure 42: *Stereochinus neumayeri*. ~~Percentage of individuals recorded as~~

1 ~~feeding in experimental treatments (n=30)~~ Biological functions measured in
2 *Sterechinus neumayeri* in experimental conditions -where temperatures were
3 increased daily by 0.3°C, 0.5°C and 1°C. Functions in warming conditions are
4 plotted against ~~, relative to the increasing e in cumulative intensity~~ temperature
5 and ~~, ambient control treatments are plotted against the number of days in the~~
6 experiment. Data points represent the pooled data within replicate floating
7 tanks (n=5). Regressions are either segmented where appropriate for treatment
8 conditions or linear for controls and treatment data where breakpoints were not
9 identified. Grey area represents interquartile range of % individuals feeding in
10 ambient conditions throughout the experiment.

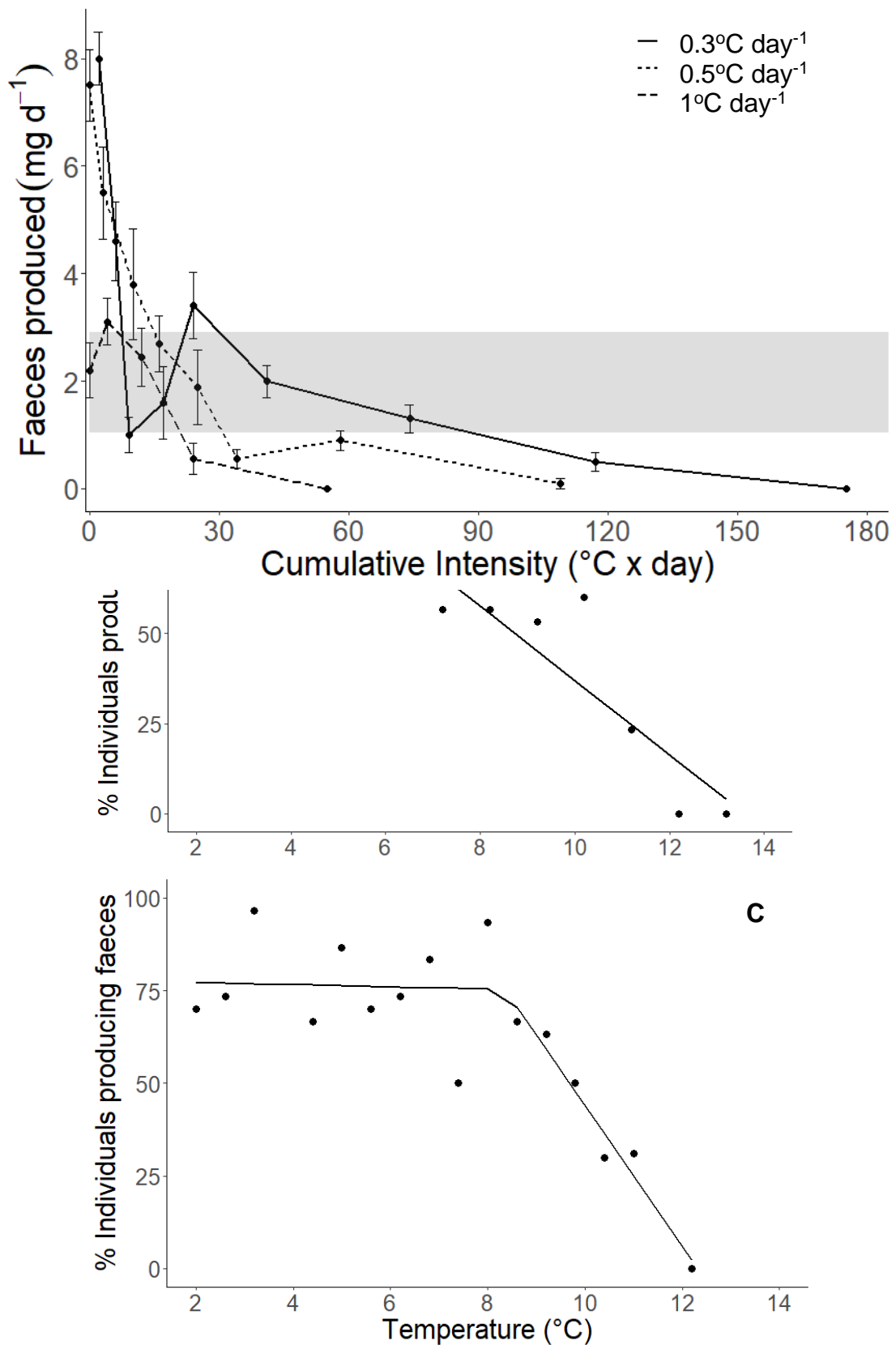


1 ~~Figure 2: *Stereochinus neumayeri*. Segmented linear regression models for the~~
 2 ~~percentage of animals feeding in experimental treatments (n=30) where~~
 3 ~~temperatures were increased daily by 0.3°C, 0.5°C and 1°C. A)~~

- 1 Temperatures increased by $1^{\circ}\text{C day}^{-1}$, breakpoint identified at 4°C ($R^2 = 0.894$).
- 2 B) Temperatures increased by $0.5^{\circ}\text{C day}^{-1}$, breakpoint identified at 6.2°C ($R^2 =$
- 3 0.964). Temperatures increased by $0.3^{\circ}\text{C day}^{-1}$, breakpoint identified at 8.2°C
- 4 ($R^2 = 0.922$).



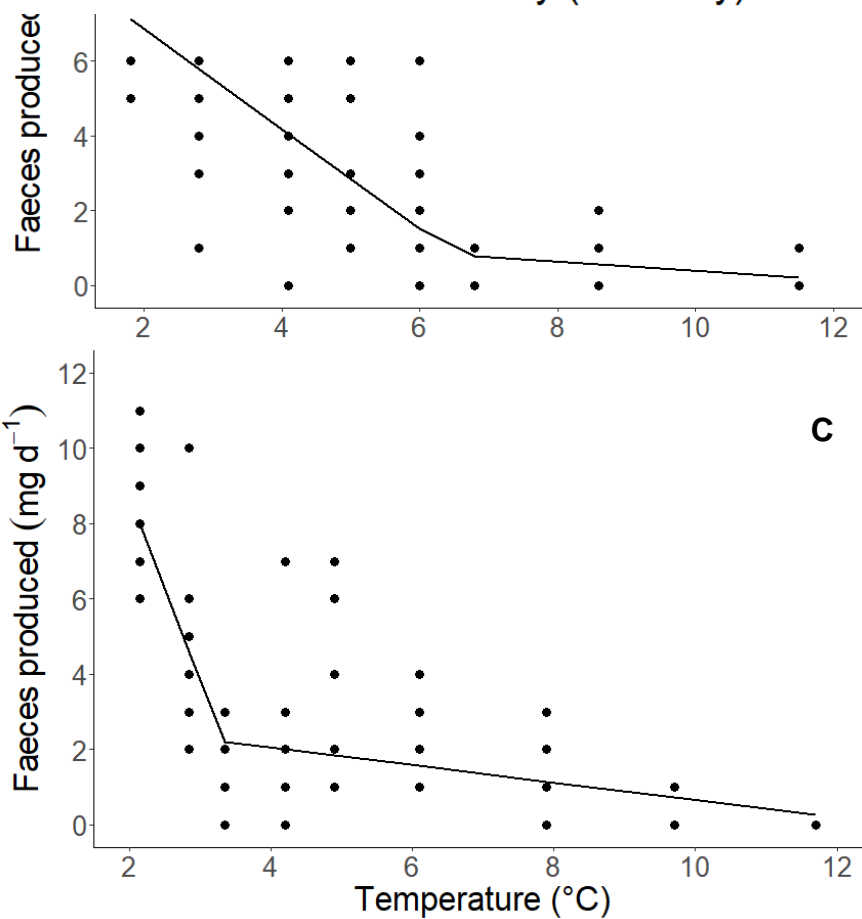
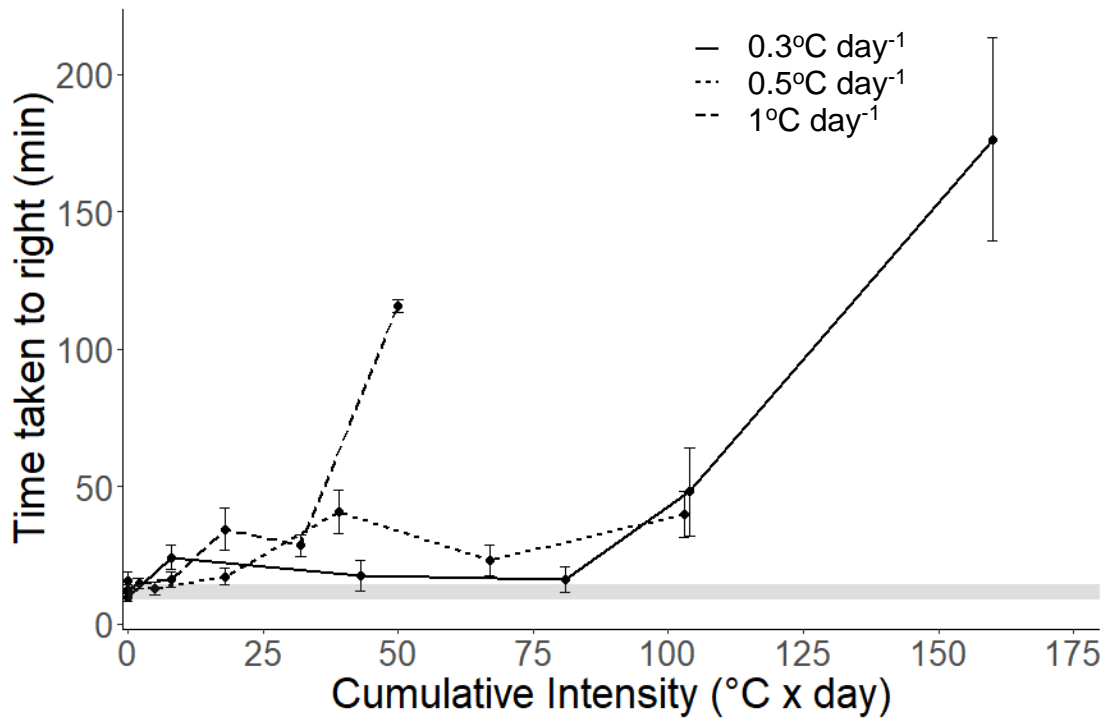
- 5 ~~Figure 3: *Stereochinus neumayeri*. Percentage of individuals recorded as~~
- 6 ~~producing faeces in experimental treatments (n=30) where temperatures were~~
- 7 ~~increased daily by 0.3°C , 0.5°C and 1°C , relative to the increase in cumulative~~
- 8 ~~intensity. Grey area represents interquartile range of % individuals producing~~
- 9 ~~faeces in ambient conditions throughout the experiment.~~



1 ~~Figure 4: *Stereochinus neumayeri*. Segmented linear regression models for the~~
 2 ~~percentage of animals producing faeces in experimental treatments (n=10)~~

1 where temperatures were increased daily by 0.3°C, 0.5°C and 1°C. A)
2 Temperatures increased by 1°C day⁻¹, breakpoint identified at 5.2°C (R² =
3 0.882). B) Temperatures increased by 0.5°C day⁻¹, breakpoint identified at
4 4.5°C (R² = 0.844). Temperatures increased by 0.3°C day⁻¹, breakpoint
5 identified at 8.3°C (R² = 0.762).

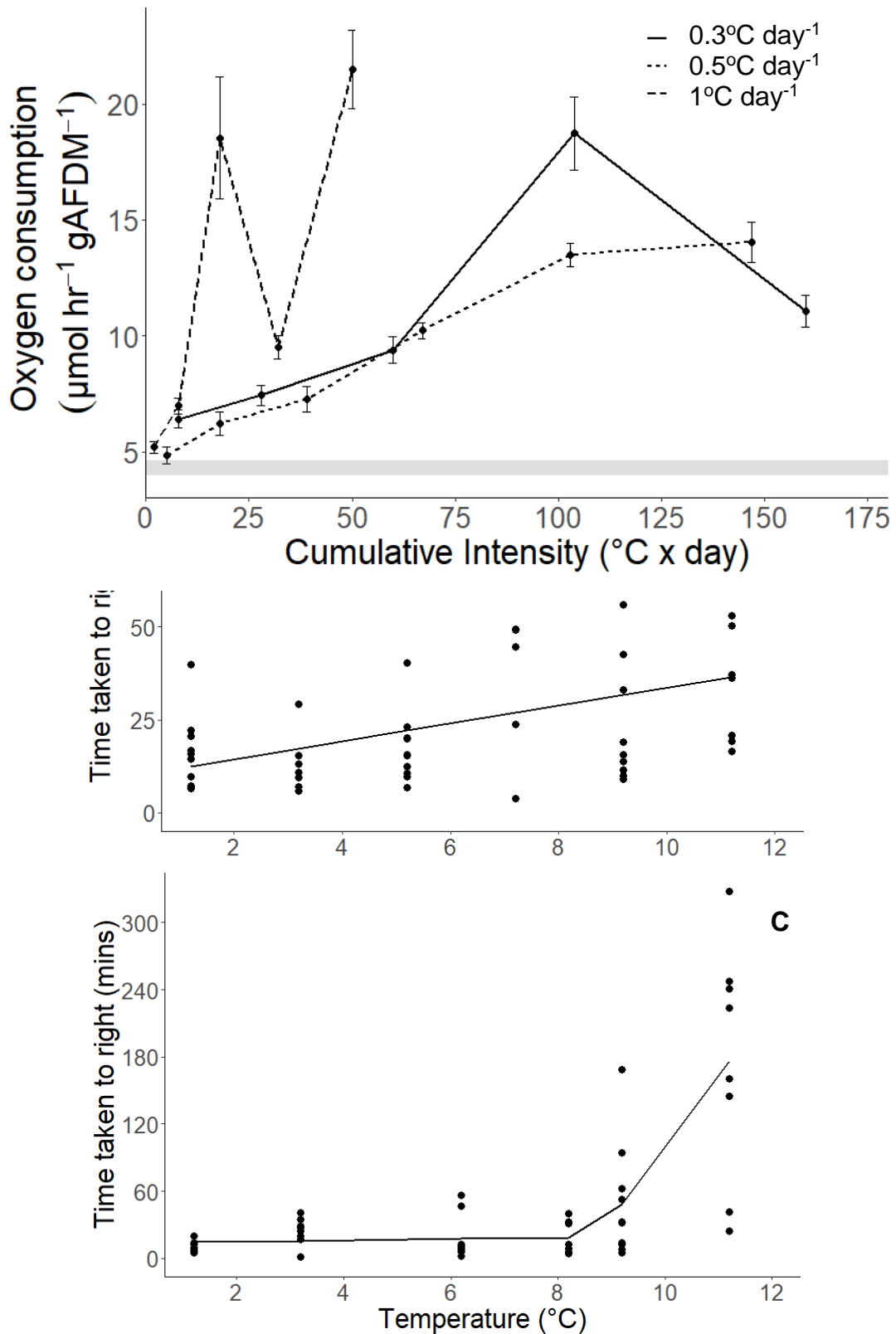
6 Figure 5: *Stereochinus neumayeri*. Faeces produced (mg day⁻¹) in experimental
7 treatments (n=10) where temperatures were increased daily by 0.3°C, 0.5°C
8 and 1°C, relative to the increase in cumulative intensity. Error bars = +/-
9 standard error. Grey area represents interquartile range of faeces produced
10 (mg day⁻¹) in ambient conditions throughout the experiment.



1 *Figure 6: *Stereochinus neumayeri*. Linear and segmented regression models for*
 2 *the rate of faecal production in experimental treatments (n=10) where*
 3 *temperatures were increased daily by 0.3°C, 0.5°C and 1°C. A)*

1 Temperatures increased by $1^{\circ}\text{C day}^{-1}$, no breakpoint identified ($R^2 = 0.364$). B)
2 Temperatures increased by $0.5^{\circ}\text{C day}^{-1}$, breakpoint identified at 6.5°C ($R^2 =$
3 0.611). Temperatures increased by $0.3^{\circ}\text{C day}^{-1}$, breakpoint identified at 3.3°C
4 ($R^2 = 0.553$).

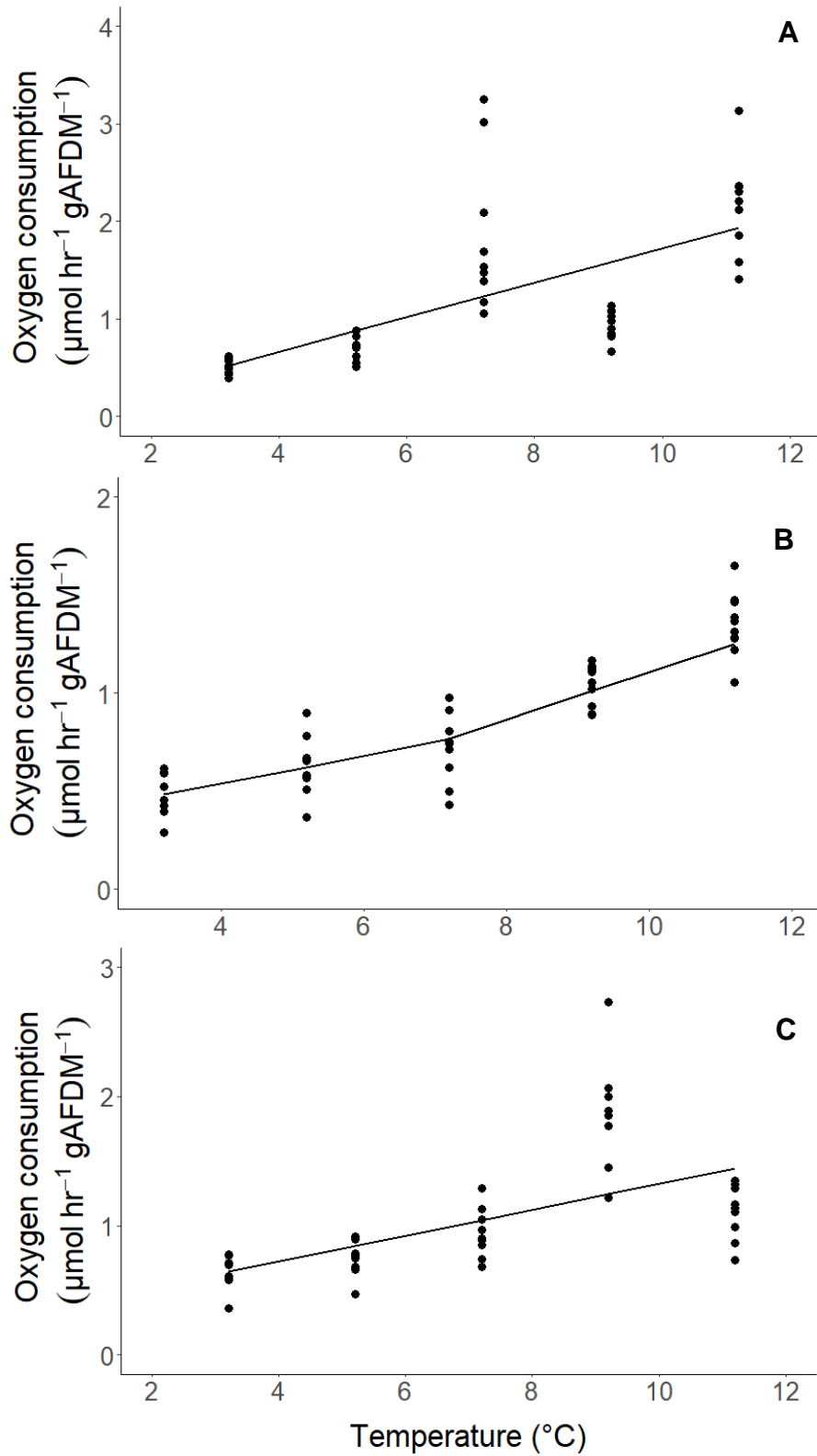
5 ~~Figure 7: *Stereochinus neumayeri*. Time taken to right in experimental~~
6 ~~treatments (n=10) where temperatures were increased daily by 0.3°C , 0.5°C~~
7 ~~and 1°C , relative to the increase in cumulative intensity. Error bars = +/-~~
8 ~~standard error. Grey area represents interquartile range of time taken to right~~
9 ~~in ambient conditions throughout the experiment.~~



1 ~~Figure 8: *Stereochinus neumayeri*. Linear and segmented regression models for~~
 2 ~~the time taken to right in the experimental treatments (n=10) where~~
 3 ~~temperatures were increased daily by 0.3°C , 0.5°C and 1°C . A) Temperatures~~

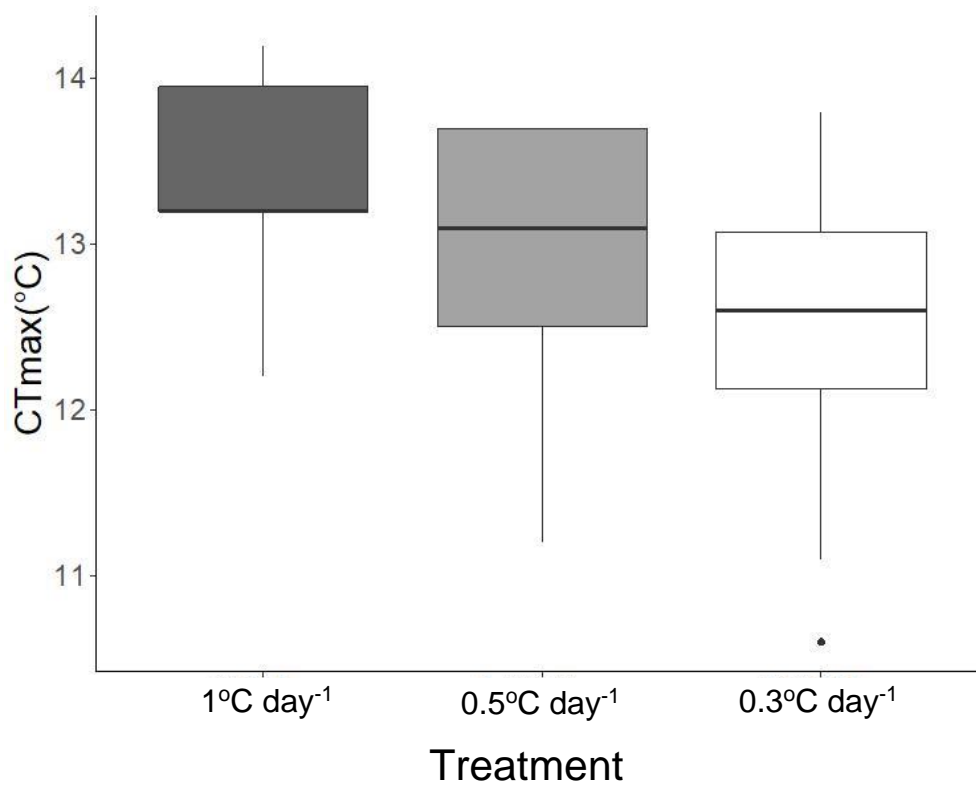
1 increased by $1.0^{\circ}\text{C day}^{-1}$, breakpoint identified at 6.8°C ($R^2 = 0.553$). B)
2 Temperatures increased by $0.5^{\circ}\text{C day}^{-1}$, no breakpoint identified ($R^2 = 0.198$).
3 Temperatures increased by $0.3^{\circ}\text{C day}^{-1}$, breakpoint identified at 8.7°C ($R^2 =$
4 0.589).

5 ~~Figure 9: *Sterechinus neumayeri*. Oxygen consumption rate of urchins in~~
6 ~~experimental treatments (n=10) where temperatures were increased daily by~~
7 ~~0.3°C , 0.5°C and 1°C , relative to the increase in cumulative intensity. Error bars~~
8 ~~\pm standard error. Grey area represents interquartile range of oxygen~~
9 ~~consumption rates in ambient conditions throughout the experiment.~~

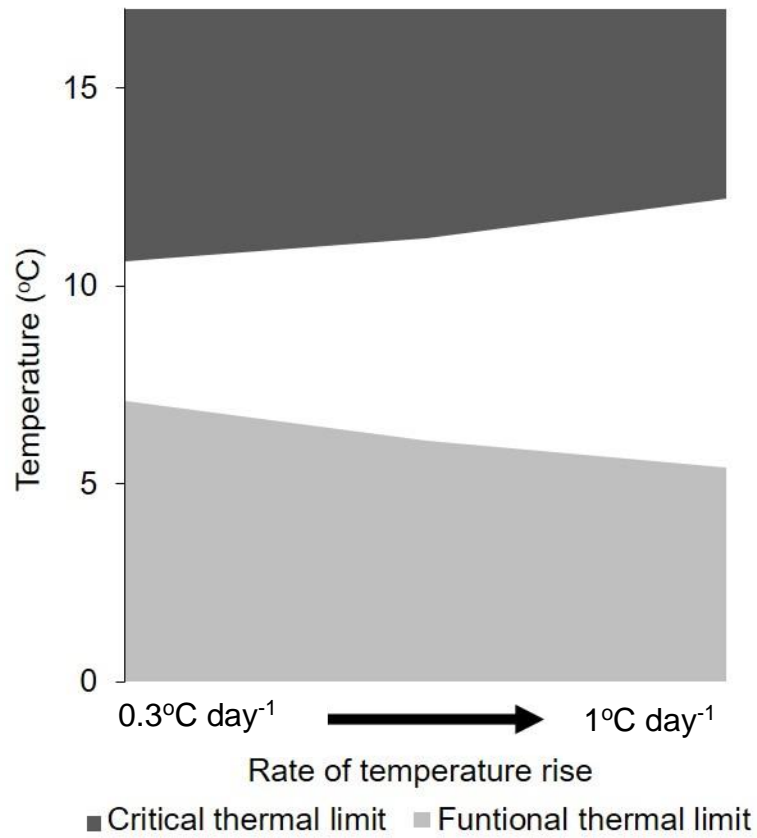


1 *Stereochinus neumayeri*. Segmented and linear regression models
 2 for the rate of oxygen consumption in the experimental treatment (n=10) where
 3 temperatures were increased daily by 0.5°C. A) Temperatures increased by

- 1 ~~1°C day⁻¹, no breakpoint identified ($R^2 = 0.425$). B) Temperatures increased by~~
 2 ~~0.5°C day⁻¹, breakpoint identified at 7.0°C ($R^2 = 0.804$). Temperatures~~
 3 ~~increased by 0.3°C day⁻¹, no breakpoint identified ($R^2 = 0.339$).~~



- 4 ~~Figure 11: Critical thermal maximum (CT_{max}) recorded in the experimental~~
 5 ~~treatments ($n=30$) where temperatures were increased daily by 0.3°C, 0.5°C~~
 6 ~~and 1°C. Data are displayed as box plots with the central line in the boxes~~
 7 ~~representing the median value, the upper and lower hinges representing the~~
 8 ~~25th and 75th percentiles, and the upper/lower whiskers representing the~~
 9 ~~largest/smallest value, no further than 1.5 times the interquartile range from the~~
 10 ~~hinge. Data outside these ranges are plotted as points.~~



- 1 ~~Figure 12: Schematic representation of the relationship between functional~~
- 2 ~~thermal limits and critical thermal limits for *Stereochinus neumayeri* as the rate~~
- 3 ~~of warming is increased from 0.3°C day⁻¹ to 1°C day⁻¹.~~