Toxin-neutralizing antibodies protect against *Clostridium perfringens* challenge in an intestinal loop model for bovine enterotoxaemia

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Introduction

Objectives

CORE



Bovine enterotoxaemia

- Caused by Clostridium perfringens
 - Alpha toxin and perfringolysin O involved in the pathogenesis (Verherstraeten et al. 2013)
- Sudden death \rightarrow treatment \rightarrow preventive measures needed, such as vaccination
- Lesions of hemorrhagic enteritis in the small intestine
- Veal calves
 - produce less anitbodies against *C. perfringens* toxins than beef calves
 - more vulnerable to enterotoxaemia than beef calves (Valgaeren et al. 2015)



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- Can antibodies against C. perfringens toxins protect agianst the develoment of necrotic lesions in the intestine?
- Can we remove the undesired toxin activity, but conserve the immune-protective potential, from the toxin preparations?

Vaccination with *C. perfringens* toxins resulted in strong antibody responses

Two calves were immunized for each antigen

- Native *C. perfringens* toxins
- L-lysine protected, formaldehyde inactivated *C. perfringens* toxins
- Commercial formaldehyde inactivated clostridial vaccine

In all calves a strong antibody response against both alpha toxin and perfringolysin O was detected.

Table 1: antibody response towards alpha toxin and perfingolysin O measured by ELISA

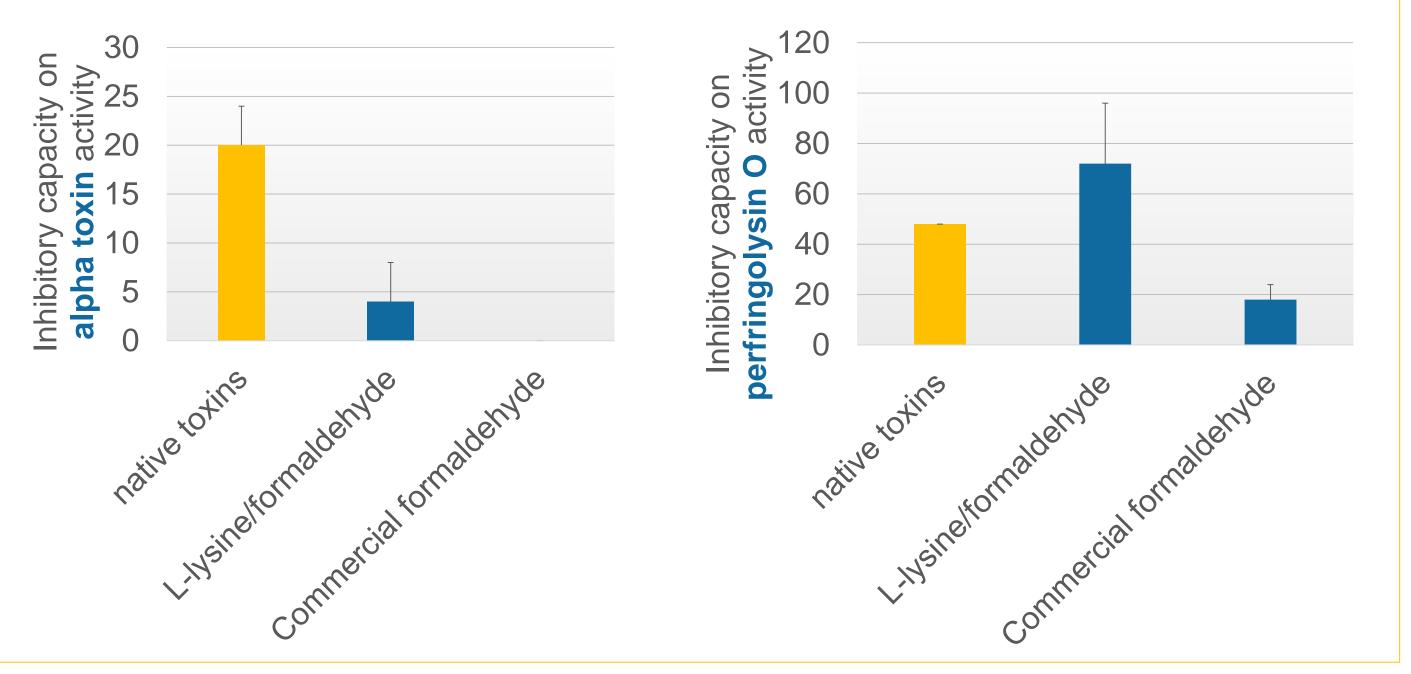
Vaccine	Alpha toxin titer	Perfringolysin O titer
Native toxins	64.44 ± 0.2227	25600 ± 0
L-lysine/formaldehyde toxoid	24.26 ± 2.960	16000 ± 9600
Commercial formaldehyde inactivated vaccine	45.14 ± 20.42	4800 ± 1600

Toxin-neutralizing antibodies protect against *C. perfringens-*induced necrotic lesions

In vitro neutralization of alpha toxin and perfringolysin O

Alpha toxin activity \rightarrow lecithinase effect on egg yolk lipoproteins. Perfringolysin activity \rightarrow hemolysis of horse erythrocytes. Toxin neutralization by pre-incubation of toxins with a dilution series of the antibodies

- Antibodies from calves vaccinated with native C. perfingens toxins were able to neutralize both toxin activities
- Antibodies from calves vaccinated with L-lysine protected, formaldehyde inactivated toxins were able to inhibit the perfringolysin O activity, but had less effect on the alpha toxin activity
- Antibodies from calves vaccinated with a commercial formaldehyde inactivated vaccine were less capable to inhibit both toxin activities



The potential of the antisera to inhibit *C. perfringens*-induced necrosis, was evaluated by neutralizing the development of necrotic lesions using the antisera in an intestinal loop assay (Figure 1).





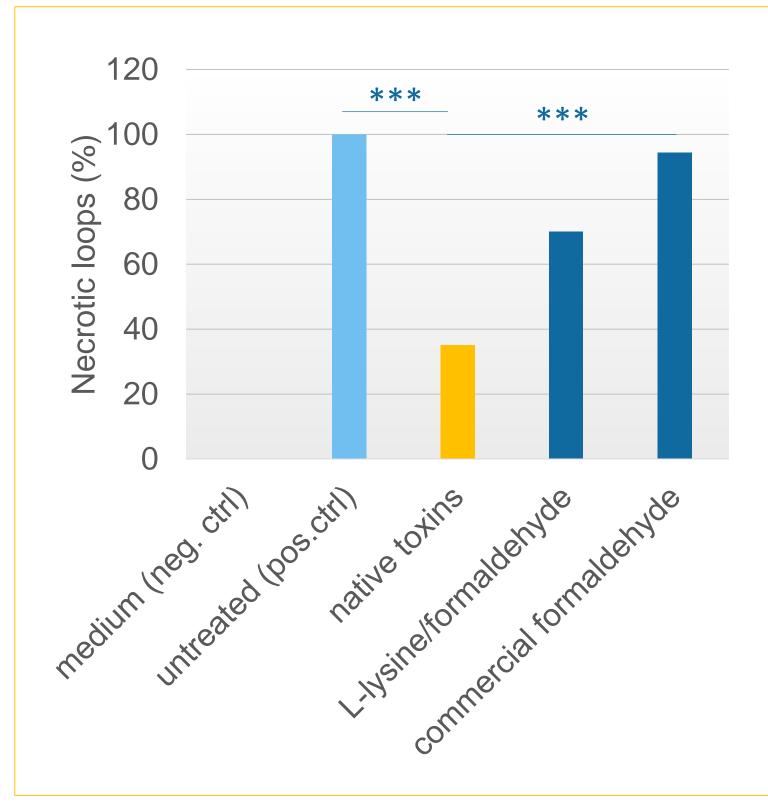


Induction of ana Ligation of loops			Sacrification
Preparation		Incubation (5h)	Sampling
	Inoculatio	n	

- Log phase culture
- Antibodies against *C. perfringens* toxins

Figure 1: experimental set up

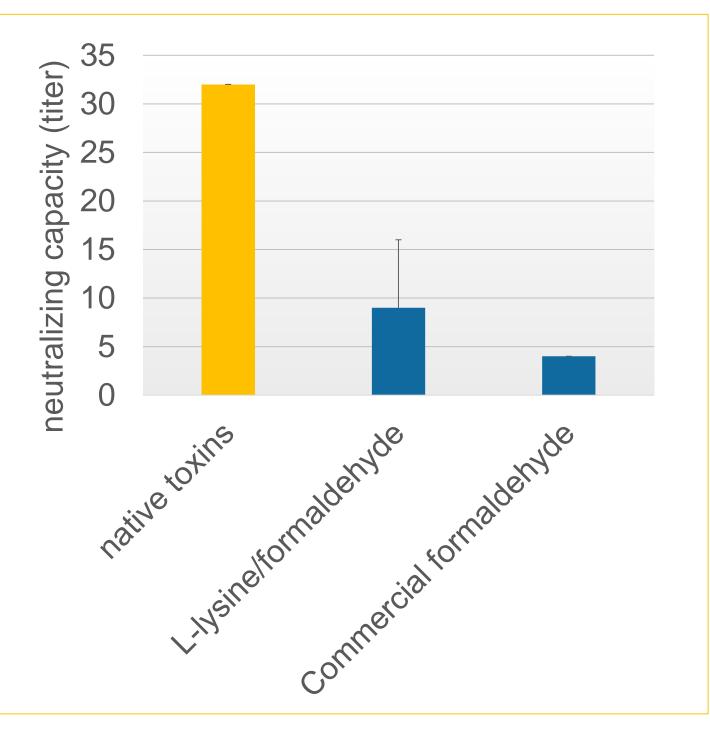
The antisera were tested in 4 calves, with a total of 20 loops for each vaccine (5 loops per animal)



All control loops inoculated with *C. perfringens* alone (untreated) developed necrosis.

Figure 3: in vitro neutralization of biological activities of alpha toxin and perfringolsyin

Toxin-neutralizing antibodies neutralize the *C. perfringens*induced cytotoxicity on bovine endothelial cells



Cytotoxicity assay: Bovine Umbilical Vein Endothelial Cells (BUVEC) exposed to filter-sterilized supernatant of *C. perfringens* Cytotoxicity quantified by a neutral red uptake (NRU) assay

Neutralization of cytotoxicity by preincubation with the antibodies.

Antibodies from calves vaccinated with native *C. perfingens* toxins were able to neutralize the cytotoxicity of *C. perfringens* on bovine endothelial cells.

Injection of *C. perfringens* together with antisera from calves vaccinated with native toxins resulted in significantly fewer necrotic loops.

Antisera from calves vaccinated with formaldehyde inactivated toxins (either L-lysine protected or the commercial inactivated vaccine) were unable to neutralize the lesion induction.

Figure 2: neutralization of the lesion-inducing potential of *C. perfringens.* *** p < 0.001 (Kruskall-Wallis analysis, followed by a Dunn's multiple comparison test) Figure 4: neutralization of *C. perfringens* cytotoxicity

Conclusion

- Toxin-neutralizing antibodies protected against C. perfringens challenge
- Prevention of endothelial damage may be the mechanisme underlying this protective effect
- Immunization of both native and formaldehyde inactivated C. perfringnes toxins
 resulted in a strong immune respons against alpha toxin and perfringolysin O
- Only antibodies raised against native toxins were protective

At least for alpha toxin and perfringolysin O mediated diseases, antibody titers detected by ELISA are not a guarantee for protection even if protection against the disease is antibody mediated

Verherstraeten S, Goossens E, Valgaeren B, et al. The synergistic necrohemorrhagic action of *Clostridium perfringens* perfringolysin and alpha toxin in the bovine intestine and against bovine endothelial cells. Veterinary research. 2013;44:45 Valgaeren B, Pardon B, Goossens E, et al. Veal Calves Produce Less Antibodies against *C. Perfringens* Alpha Toxin Compared to Beef Calves. Toxins. 2015;7:2586-97