

MYCN and HDAC5 transcriptionally repress CD9 to trigger invasion and metastasis in neuroblastoma

Johannes Fabian^{1,2}, Desirée Opitz¹, Kristina Althoff³, Marco Lodrini^{1,4}, Barbara Hero⁵, Ruth Volland⁵, Anneleen Beckers^{6,7}, Katleen de Preter^{6,7}, Anneleen Decock^{6,7}, Nitin Patil⁸, Mohammed Abba⁸, Annette Kopp-Schneider⁹, Kathy Astrahantseff⁴, Jasmin Wünschel^{1,4}, Sebastian Pfeil⁴, Maria Ercu⁴, Annette Künkele⁴, Jamie Hu^{1,10}, Theresa Thole^{1,4}, Leonille Schweizer¹¹, Gunhild Mechtersheimer¹², Daniel Carter¹³, Belamy B. Cheung¹³, Odilia Popanda¹⁴, Andreas v. Deimling^{11,15}, Jan Koster¹⁶, Rogier Versteeg¹⁶, Manfred Schwab¹⁷, Glenn M. Marshall^{13,18}, Frank Speleman^{6,7}, Ulrike Erb¹⁹, Margot Zoeller¹⁹, Heike Allgayer⁸, Thorsten Simon⁵, Matthias Fischer^{5,20,21}, Andreas E. Kulozik²², Angelika Eggert⁴, Olaf Witt^{1,22}, Johannes H. Schulte⁴ and Hedwig E. Deubzer^{1,4,22,23}

¹ Clinical Cooperation Unit Pediatric Oncology, German Cancer Research Center (DKFZ) and German Consortium for Translational Cancer Research (DKTK), INF, Heidelberg, Germany

² Phenex Pharmaceuticals AG, Waldhofer Straße, Heidelberg, Germany

³ Department of Pediatric Hematology and Oncology, University Children's Hospital Essen, Essen, Germany

⁴ Department of Pediatric Hematology, Oncology and SCT, Charité - University Hospital Berlin, Campus Virchow-Klinikum, Berlin, Germany

⁵ Department of Pediatric Hematology and Oncology, University of Cologne, Cologne, Germany

⁶ Center for Medical Genetics Ghent, Ghent University, De Pintelaan, Ghent, Belgium

⁷ Cancer Research Institute Ghent, De Pintelaan, Ghent, Belgium

⁸ Department of Experimental Surgery, Medical Faculty Mannheim, University of Heidelberg, Centre for Biomedicine and Medical Technology, Mannheim, Germany

⁹ Department of Biostatistics, German Cancer Research Center (DKFZ), INF, Heidelberg, Germany

¹⁰ Yale University, New Haven, CT

¹¹ Department of Neuropathology, University of Heidelberg, INF, Heidelberg, Germany

¹² Department of Pathology, University of Heidelberg, INF, Heidelberg, Germany

¹³ Children's Cancer Institute, UNSW, Randwick, NSW, Australia

¹⁴ Division of Epigenomics and Cancer Risk Factors, DKFZ, INF, Heidelberg, Germany

¹⁵ Clinical Cooperation Unit Neuropathology, DKFZ, INF, Heidelberg, Germany

¹⁶ Department of Oncogenomics and Emma Children's Hospital, Academic Medical Center, University of Amsterdam, Meibergdreef, Amsterdam, the Netherlands

¹⁷ Neuroblastoma Genetics, DKFZ, INF, Heidelberg, Germany

¹⁸ Kids Cancer Centre, Sydney Children's Hospital Randwick, Randwick, NSW, Australia

¹⁹ Experimental Surgery and Tumor Cell Biology, University of Heidelberg, INF, Heidelberg, Germany

²⁰ Center for Molecular Medicine Cologne, University of Cologne, Cologne, Germany

²¹ Max Plank Institute for Metabolism Research, Cologne, Germany

²² Department of Pediatric Hematology and Oncology, Heidelberg University, INF, Heidelberg, Germany

²³ Junior Neuroblastoma Research Group, Experimental and Clinical Research Center of the Max-Delbrück Center for Molecular Medicine in the Helmholtz Community and the Charité - University Medicine Berlin, Lindenberger Weg, Berlin, Germany

Correspondence to: Hedwig E. Deubzer, **email:** hedwig.deubzer@charite.de

Keywords: antimetastatic therapy, chromatin modulation, histone deacetylases, grainyhead-like transcription factor family, tetraspanin family

Received: April 29, 2016

Accepted: August 24, 2016

Published: August 27, 2016

ABSTRACT

The systemic and resistant nature of metastatic neuroblastoma renders it largely incurable with current multimodal treatment. Clinical progression stems mainly from the increasing burden of metastatic colonization. Therapeutically inhibiting the migration-invasion-metastasis cascade would be of great benefit, but the mechanisms driving this cycle are as yet poorly understood. In-depth transcriptome analyses and ChIP-qPCR identified the cell surface glycoprotein, CD9, as a major downstream player and direct target of the recently described *GRHL1* tumor suppressor. CD9 is known to block or facilitate cancer cell motility and metastasis dependent upon entity. High-level *CD9* expression in primary neuroblastomas correlated with patient survival and established markers for favorable disease. Low-level *CD9* expression was an independent risk factor for adverse outcome. *MYCN* and *HDAC5* colocalized to the *CD9* promoter and repressed transcription. *CD9* expression diminished with progressive tumor development in the *TH-MYCN* transgenic mouse model for neuroblastoma, and *CD9* expression in neuroblastic tumors was far below that in ganglia from wildtype mice. Primary neuroblastomas lacking *MYCN* amplifications displayed differential *CD9* promoter methylation in methyl-CpG-binding domain sequencing analyses, and high-level methylation was associated with advanced stage disease, supporting epigenetic regulation. Inducing CD9 expression in a SH-EP cell model inhibited migration and invasion in Boyden chamber assays. Enforced CD9 expression in neuroblastoma cells transplanted onto chicken chorioallantoic membranes strongly reduced metastasis to embryonic bone marrow. Combined treatment of neuroblastoma cells with HDAC/DNA methyltransferase inhibitors synergistically induced CD9 expression despite hypoxic, metabolic or cytotoxic stress. Our results show CD9 is a critical and indirectly druggable suppressor of the invasion-metastasis cycle in neuroblastoma.

INTRODUCTION

Neuroblastoma, an embryonic tumor of neuroectodermal origin, accounts for 11% of all cancer-related deaths in children [1]. Molecular aspects create the extreme heterogeneity of this disease, spanning spontaneous regression to rapid metastasizing progression [1, 2]. Treatment scenarios range between observation only and multimodal concepts including high-dose chemotherapy with autologous stem cell rescue, surgery, radiotherapy and immunotherapy [3]. Despite decades of considerable international efforts to improve outcome, long-term survival of high-risk disease remains as low as 20% [3]. Managing resistance to induction therapy, causing tumor progression and early death in ultrahigh-risk patients, and managing chemotherapy-resistant relapses, which can occur years after initial diagnosis, remain major obstacles. *MYCN* amplifications [4, 5], telomerase activation by genomic rearrangements [6, 7], *ATRX* loss-of-function mutations or deletions [8-10] and germline or somatically acquired activating *ALK* mutations [11-14] define patient subgroups with highly aggressive and frequently therapy-resistant neuroblastomas. A commonality among these aggressive subgroups is that tumors undergo an as yet poorly understood invasion-metastasis cascade, which may be druggable to prevent

increasing metastatic colonization.

Tetraspanin proteins all have four membrane-spanning domains, and form clusters between themselves and a large variety of molecules, predominantly including integrins, proteases and gangliosides. Tetraspanins can become engaged in signal transduction by recruiting cytoplasmic kinases to the glycolipid- and cholesterol-enriched membrane microdomains to which they are localized [15-17]. CD9 tetraspanin expression strongly varies among different organs, with the highest expression found in endocrine tissues and the gastrointestinal tract, and the lowest expression reported both in liver and gall bladder [18]. Here, we assessed the significance of CD9 expression in primary neuroblastomas using clinical and molecular tumor data, unraveled the upstream activating and repressive control including epigenetic events and analyzed the functional role of CD9 using targeted CD9 re-expression in cell and animal models of neuroblastoma.

RESULTS

GRHL1 is a transcriptional activator of *CD9*

High-level GRHL1 expression favorably influences neuroblastoma biology at the molecular and phenotypic

levels [19]. Time-resolved whole-genome expression analyses on BE(2)-C cells transfected with empty- or *GRHL1* vectors resulted in differential regulation over time of 170 genes, including *CD9* [19]. Increasing *CD9* expression following enforced *GRHL1* expression in BE(2)-C cells was confirmed both on the mRNA level by qRT-PCR and on the protein level by western blot analysis (Figure 1A). Transient knockdown of *GRHL1* in SH-EP cells by two different siRNAs to control for unspecific and off-target effects reduced *CD9* expression by approximately 50% (Figure 1B). We next conducted ChIP-qPCR using an antibody against the FLAG tag of FLAG-*GRHL1* in overexpressing BE(2)-C cells to assess whether *GRHL1* is recruited to the *CD9* promoter region (Figure 1C). *GRHL1* was significantly enriched (Figure 1D) above the control (ChIP for IgG) in the *CD9* transcriptional start site distal to a previously described SP1 binding site [20] and proximal to a MYC binding site (<http://genome.ucsc.edu>). We conclude that *GRHL1* activates *CD9* transcription in neuroblastoma cells.

High-level *CD9* expression in neuroblastomas predicts favorable survival as an independent prognostic marker

Having shown that *CD9* is downstream of *GRHL1* in neuroblastoma cells, we assessed whether *CD9* is differentially expressed in primary tumors. We reanalyzed microarray expression data from a cohort of 476 neuroblastomas [21]. Kaplan-Meier analysis showed that high-level *CD9* expression in tumors, regardless of International Neuroblastoma Staging System (INSS) stage or patient age, correlated both with favorable event-free and overall patient survival (Figure 2A-2F and Supplementary Figure S1A-S1D). These correlations were confirmed in microarray expression data from an independent cohort of 122 neuroblastomas [22] (Supplementary Figure S2A-S2D). High-level *CD9* expression in neuroblastomas from the 476-tumor cohort also significantly correlated with established clinical and molecular markers for favorable tumor biology, including INSS localized or 4S disease stages, age at diagnosis ≤ 18 months, favorable Shimada/International Neuroblastoma Pathology Classification (INPC) tumor histology, lack of *MYCN* amplifications or 1p aberrations and a low-

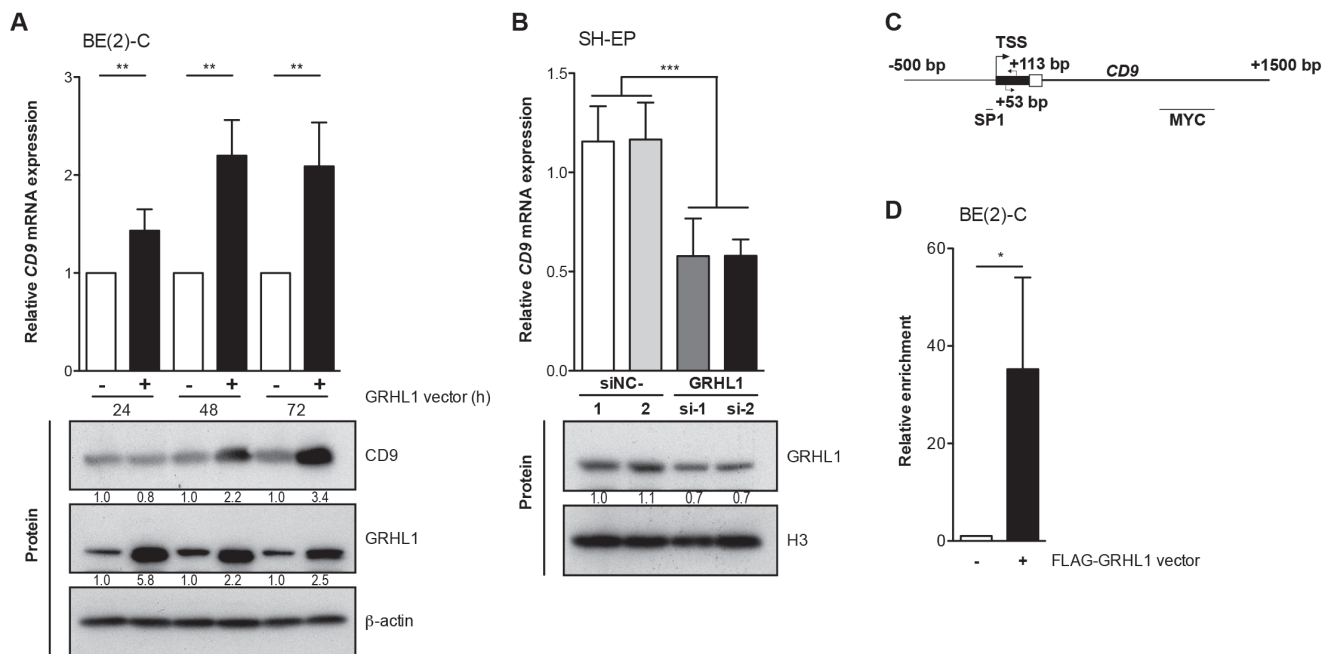


Figure 1: GRHL1 triggers *CD9* expression. **A.**, *CD9* expression in BE(2)-C cells 24h-72h after *GRHL1* plasmid or empty-vector transfection on mRNA (upper panel; qRT-PCR; mean \pm SD; $n = 3$) and protein level (lower panel; western blot). **B.**, *CD9* expression in SH-EP cells 96h after *GRHL1* knockdown with small interfering RNAs (si-1, si-2) (upper panel; qRT-PCR; mean fold-change over mock \pm SD; $n = 3$). Controls were transfected with two different negative control siRNAs (siNC-1, siNC-2). *GRHL1* knockdown efficacy was measured by western blot at 96h (lower panel) while *CD9* protein expression in SH-EP was below detection. Histone H3 served as loading control. **C.**, schematic representation of the *CD9* promoter indicating the primer positions (arrows), the transcriptional start site (TSS) and the localization of previously described binding sites for SP1 and MYC. **D.**, the mean enrichment (\pm SD, $n = 3$) of *CD9* promoter DNA associated with *GRHL1* is shown from ChIP-qPCR experiments. BE(2)-C cells from *GRHL1* transfected cells were immunoprecipitated with antibodies against FLAG-*GRHL1* or IgG, as negative control, and 0.1% input lysate served as normalization for comparing ChIPs (mean \pm SD, $n = 3$). * $P < 0.05$, ** $P \leq 0.01$, *** $P \leq 0.001$.

risk tumor transcriptional profile previously defined by PAM analysis [23] (Figure 2G-2L). Correlations between elevated *CD9* tumor expression and all available clinical and molecular parameters for favorable tumor biology were confirmed in the 122-tumor cohort (Supplementary Figure S2E-S2H). These included localized INSS localized or 4S disease stages, age at diagnosis ≤ 18 months, lack of *MYCN* amplifications or 1p aberrations. Multivariate survival analysis was performed in the 476-tumor cohort to determine whether *CD9* expression provides additional predictive power over established clinical and molecular prognostic markers. Established risk factors were associated both with favorable event-free and overall survival in univariate survival analyses (Supplementary Table S3) and the prognostic value of *CD9* expression was independent of these established risk markers in multivariate analyses testing all factors together (event-free survival, $P = 0.001$; overall survival, $P = 0.005$; Supplementary Table S3). Taken together, high-level *CD9* expression in primary neuroblastomas signals a favorable tumor biology and patient prognosis independent of established clinical and molecular risk factors.

MYCN is a transcriptional repressor of *CD9*

A major factor influencing neuroblastoma biology is the *MYCN* status in the tumor, which inversely correlated with *CD9* expression in primary neuroblastomas (Figure 2J, Supplementary Figure S2G). Kaplan-Meier analyses comparing the predictive power of *CD9* expression in tumors lacking or harboring *MYCN* amplifications produced only a significant correlation between high *CD9* expression and favorable event-free and overall survival in the patient cohort with tumors lacking *MYCN* amplifications (Figure 3A-3D), suggesting that *MYCN* may be upstream of *CD9*. To assess whether differential *CD9* expression in primary neuroblastomas with and without *MYCN* amplifications (Figure 2J, Supplementary Figure S2G) is mirrored at the protein level, we semiquantified *CD9* expression immunohistochemically in 20 selected samples from tumors with the most divergent characteristics. Tumors lacking *MYCN* amplifications expressed *CD9* in a variably strong manner, and expression was localized to the cell membrane, whereas no *CD9* expression was detected in any of the *MYCN*-amplified tumors (Figure 3E-3F). We investigated *CD9* gene methylation status by reanalyzing existing methyl-CpG-binding domain sequencing data from 60 neuroblastomas [24, 25]. The *CD9* promoter was differentially methylated within the group of tumors lacking *MYCN* amplifications ($n = 43$), and high-level promoter methylation was associated with advanced stage disease as defined by INSS stages 3 and 4 (Figure 3G). These results confirm the inverse correlation between *MYCN* status and *CD9* expression at the protein level

and point towards a functional role of *CD9* promoter methylation in tumors lacking *MYCN* amplifications. We next used several different neuroblastoma cell and mouse models to investigate the nature of this relationship and the influence of *MYCN* on *CD9* expression. We analyzed changes in *CD9* expression occurring between 4h to 48h in the synthetic *MYCN*-inducible system, SH-EP Tet-21/N. Conditional *MYCN* expression downregulated *CD9* expression by approximately 15% after 4h and by 50% after 24h (Figure 4A). Endogenous *MYCN* expression was depleted in BE(2)-C cells by transient expression of a short hairpin RNA (shRNA) plasmid directed against *MYCN*. *CD9* expression increased between 24h and 72h time-dependently, reaching a 2-fold increase in *CD9* mRNA expression and a 2.4-fold increase in *CD9* protein expression (Figure 4B). Results from these two models suggest that *MYCN* suppresses *CD9* expression. We performed ChIP-qPCR experiments to test whether *MYCN* directly associates with the *CD9* promoter. Not only was *MYCN* enriched in the *CD9* promoter (compare Figure 4C with Figure 1C), but this enrichment was confined to the region enriched for *GRHL1* (Figure 1D). We have previously shown that *MYCN* is a transcriptional repressor of *GRHL1* [19]. To test whether *GRHL1* is required for *CD9* induction mediated by *MYCN* depletion, we transiently knocked down both *GRHL1* and *MYCN* in BE(2)-C. Transient *GRHL1* knockdown diminished induction of the *CD9* transcript by *MYCN* depletion by 35%, indicating that *GRHL1* contributes to *CD9* transcriptional activating when silencing *MYCN* (Figure 4D). We next used the TH-*MYCN* neuroblastoma progression model to analyze the temporal *CD9* expression pattern. *CD9* expression was strongly reduced during progressive development of neuroblastic tumors in TH-*MYCN*^{+/+} mice and remained far below *CD9* expression in ganglia from wildtype TH-*MYCN*^{-/-} mice (Figure 4E). Collectively, our data argue for a role of *MYCN* in the transcriptional repression of *CD9* in neuroblastoma cells.

HDAC5 negatively regulates *CD9* and combined inhibition of HDAC5 and DNA MTases synergistically induces *CD9*

A model for transcriptional repression *via* *MYCN* involves histone deacetylase (HDAC) recruitment to promoter regions. To identify HDACs potentially involved in *CD9* transcriptional repression, we transiently knocked down each of the eleven HDACs (using two different siRNAs for each to control for unspecific and off-target effects, Supplementary Table S2) belonging to classes I, II a/b and IV in BE(2)-C cells, then assessed *CD9* expression. Only HDAC5 depletion induced *CD9* expression, which reached 1.7- to 2.3-fold of the control 96h after knockdown (Figure 5A). Depleting HDACs 1, 4, 6, 7, 8, 9 and 11 did not affect *CD9* expression,

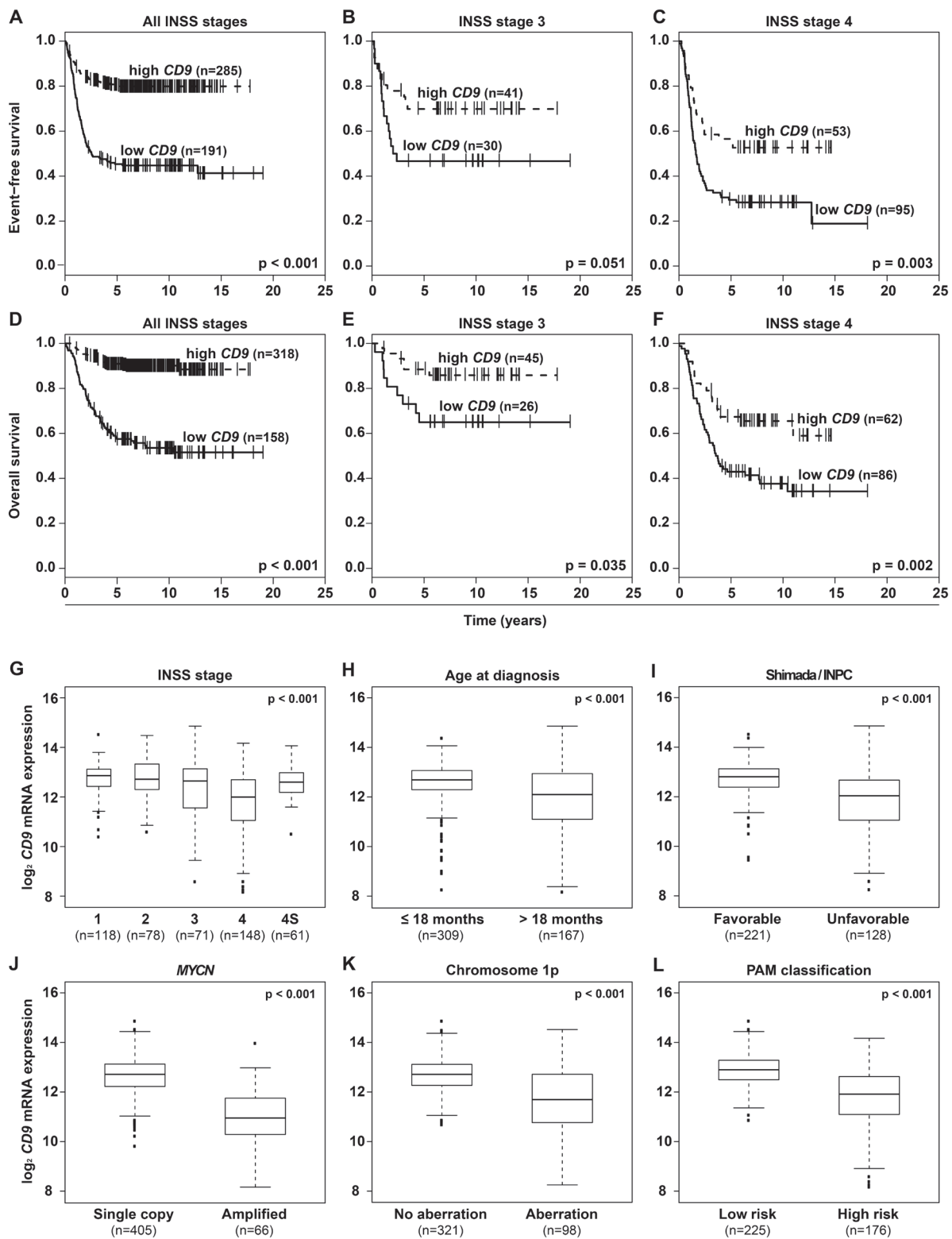


Figure 2: High-level *CD9* expression in primary neuroblastomas predicts favorable patient survival in a cohort of 476 tumors and correlates with favorable established clinical and molecular markers. Kaplan-Meier analysis of event-free **A.-C.** and overall patient survival **D.-F.** irregardless of INSS stage **A., D.** and in INSS stage 3 **B., E.** and stage 4 patients **C., F.** Box-plots compare *CD9* expression in tumors with varying stages according to INSS **G.**, in tumors from patients with age at diagnosis under or over 18 months **H.**, in tumors with favorable or unfavorable classification according to Shimada/INPC **I.**, in tumors either lacking or harboring *MYCN* amplifications **J.**, in tumors either lacking or harboring 1p aberrations **K.**, and in tumors classified either as low or high risk by a PAM classifier using microarray expression profiles **L.**

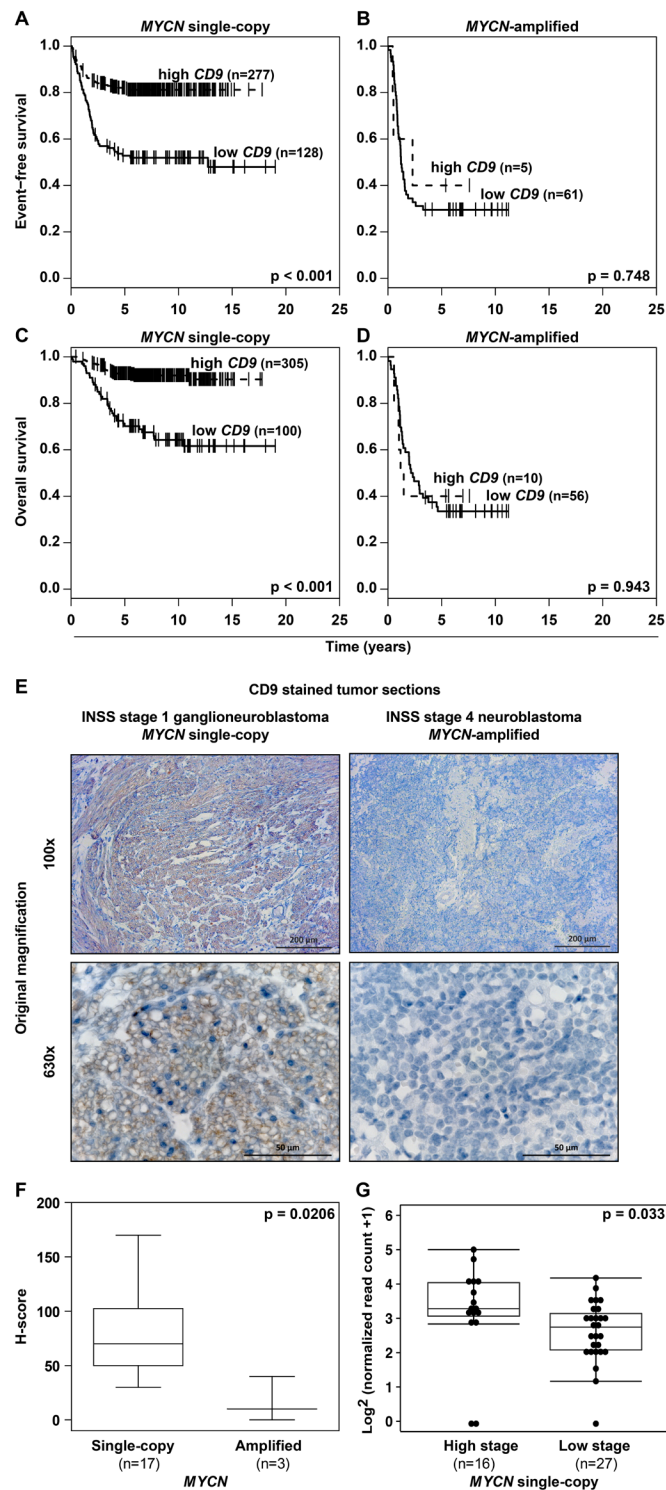


Figure 3: *MYCN* status and *CD9* expression inversely correlate in primary neuroblastomas, and *CD9* promoter hypermethylation is associated with advanced stage disease in tumors lacking *MYCN* amplifications. A.-D., high-level *CD9* expression in *MYCN* single-copy but not in *MYCN*-amplified tumors predicts favorable patient survival in a cohort of 476 tumors. Kaplan-Meier analysis of event-free A.-B. and overall C.-D. patient survival. E.-F., the differential pattern of *CD9* expression is translated to the protein level in neuroblastomas having the most divergent tumor biologies. *CD9* immunohistochemical staining (brown) is exemplarily shown E. for a low-risk INSS stage 1 ganglioneuroblastoma lacking *MYCN* amplification and an INSS stage 4 poorly differentiated high-risk neuroblastoma harboring a *MYCN* amplification. *CD9* expression was semi-quantitatively analyzed, and the data for all 20 tumors were presented as box-plots comparing the *MYCN* single-copy and *MYCN*-amplified tumor groups F. G., Box-plots compare *CD9* promoter (chr12: 6308102-6310102; Human Genome Issue HG-19) normalized read counts of methyl-CpG-binding domain (MBD) sequencing data in high- (INSS stages 3 and 4) versus low stage *MYCN* single copy primary neuroblastomas (INSS stages 1, 2 and 4S).

while depleting HDACs 2, 3 and 10 decreased *CD9* levels (Figure 5A), suggesting HDACs 2, 3 and 10 may counteract HDAC5 influence on *CD9* expression. To test whether HDAC5 is recruited to the *CD9* promoter site, we performed ChIP-qPCRs using anti-FLAG M2-conjugated agarose to capture immunocomplexes after HDAC5-FLAG enforced expression in BE(2)-C cells. HDAC5 was enriched by ~3.5-fold at the *CD9* promoter (compare

Figure 5B with Figure 1C), and localized to the region where GRHL1 was also enriched as a transcriptional activator and MYCN was enriched as a transcriptional repressor.

As an alternative approach to HDAC5 knockdown, we treated BE(2)-C cells with a variety of small molecule HDAC inhibitors, then assessed *CD9* expression. *CD9* induction by the clinically approved pan-HDAC inhibitor,

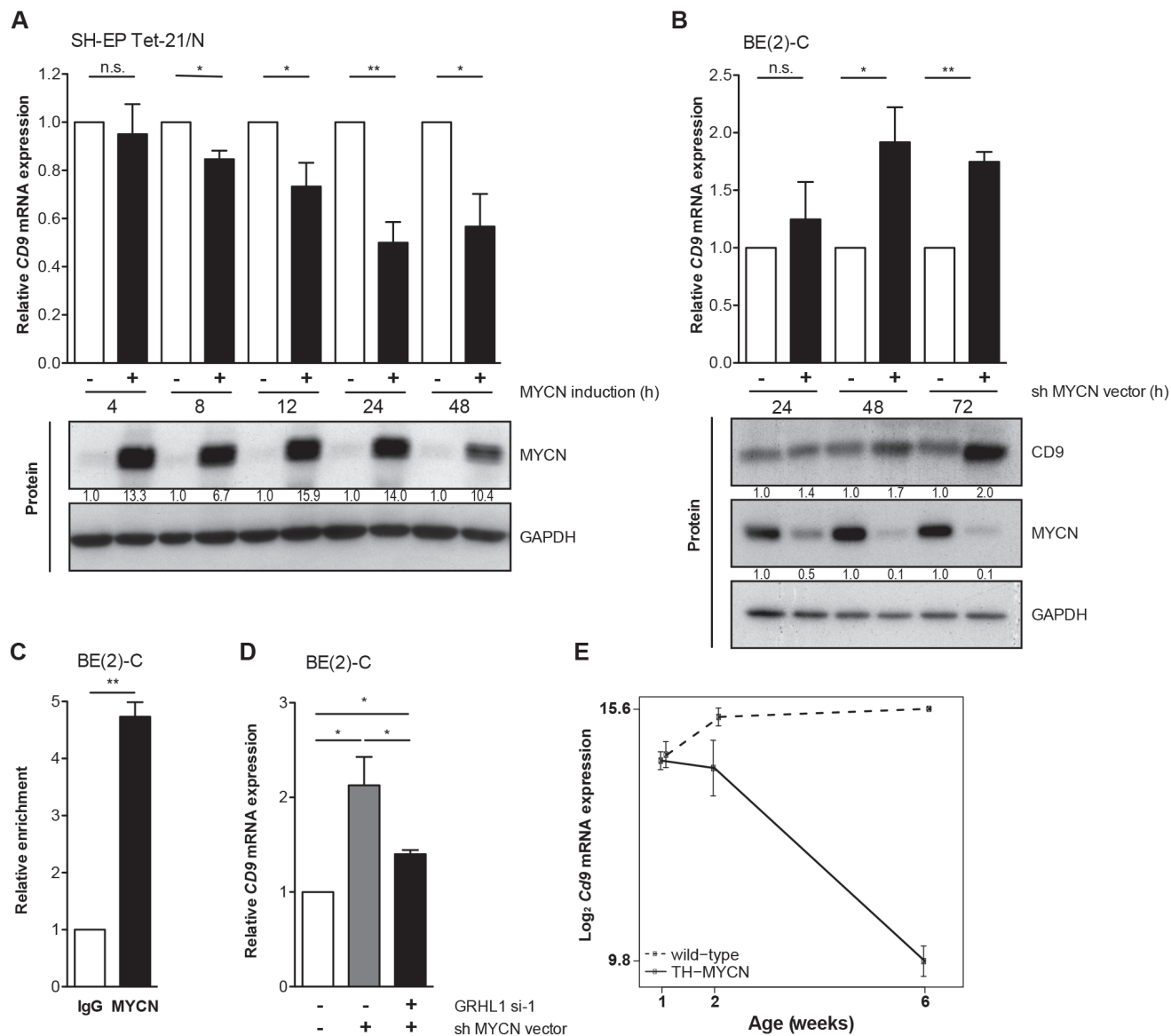


Figure 4: MYCN inhibits CD9 expression. A.-B., *CD9* expression was assessed on the mRNA (qRT-PCR; mean \pm SD, $n = 3$) and protein level (western blots) in the synthetic MYCN-inducible SH-EP Tet-21/N cell model with/without MYCN induction in time-course A. and in BE(2)-C at designated time points after MYCN depletion B. *CD9* protein expression in SH-EP Tet-21/N was below detection. GAPDH served as loading control. C., ChIP-qPCR showing an enrichment of *CD9* promoter DNA associated with MYCN. BE(2)-C lysates were immunoprecipitated with antibodies against MYCN or IgG, as negative control, and 0.1% of the input served for normalization (mean \pm SD, $n = 3$). D., *CD9* expression was assessed on the mRNA level (qRT-PCR; mean \pm SD, $n = 3$) in BE(2)-C 96h after GRHL1 knockdown using a small interfering RNA (si-1) and 72h after MYCN depletion using a short hairpin RNA (shRNA) directed against *MYCN*. Controls were transfected with respective negative control siRNA and empty vector. E., the expression of *Cd9* during neuroblastoma progression from tumor-prone ganglia to tumors in transgenic mice ($n = 4$; full line) and in comparison to *Cd9* expression in wild-type ganglia ($n = 4$; dashed line). Linear regression analysis; $P = 4.7 \times 10^{-11}$; delta slope = -1.1. * $P < 0.05$, ** $P \leq 0.01$.

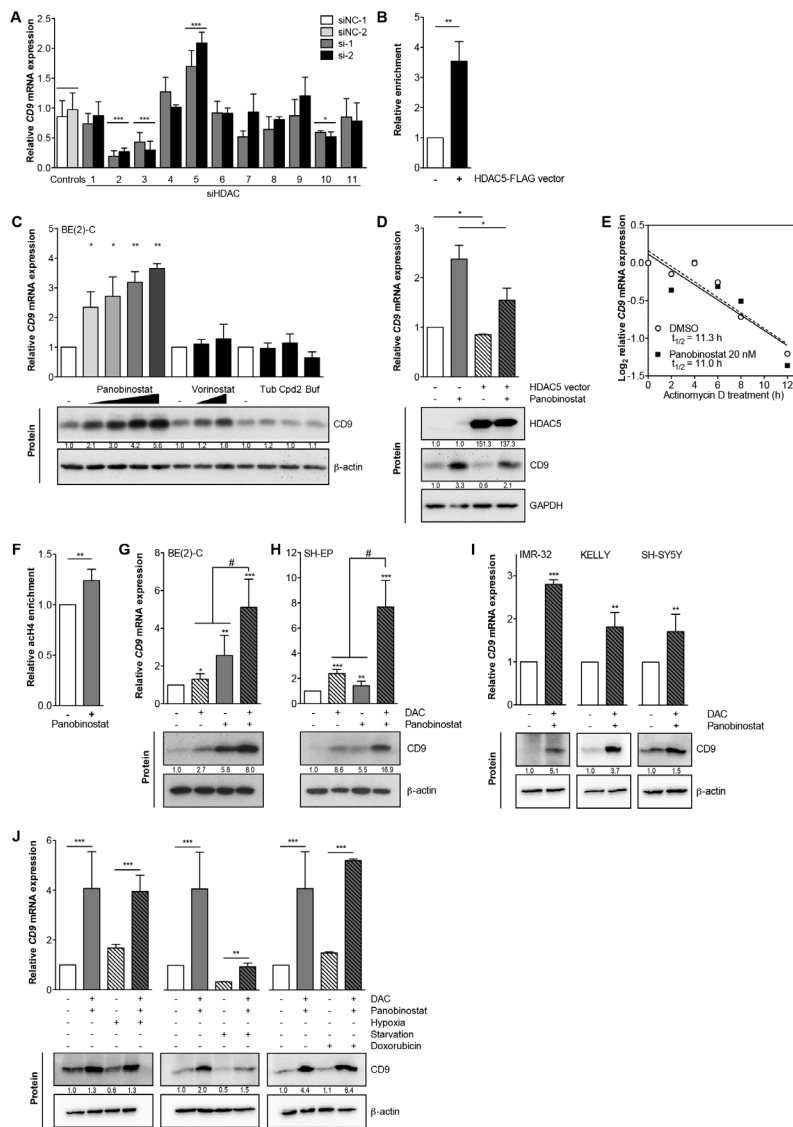


Figure 5: HDAC5 negatively regulates *CD9*, and combined DAC/HDAC inhibitor synergistically triggers *CD9* expression. **A.**, RNAi targeting HDACs 1-11 singly in BE(2)-C cells with siRNAs (si-1, si-2). Control were transfected with two different negative control siRNAs (siNC-1, siNC-2). *CD9* expression was measured by qRT-PCR 96 hours after transfection and is represented as mean fold-change over mock control \pm SD ($n \geq 2$). **B.**, ChIP-qPCR (mean \pm SD, $n = 3$) showing an enrichment of *CD9* promoter DNA associated with HDAC5. BE(2)-C lysates were immunoprecipitated with an antibody against HDAC5 or IgG as negative control. **C.**, *CD9* expression (assessed by qRT-PCR and Western blotting) after treating BE(2)-C cells with panobinostat (5 nM, 10 nM, 15 nM, 20 nM), vorinostat (0.5 μ M, 1 μ M), 5 μ M tubacin (Tub), 20 μ M compound 2 (Cpd2), 30 μ M bufexamac (Buf) or respective solvent controls for 48 hours (mean fold-change over control \pm SD is shown, $n \geq 3$). **D.**, *CD9* expression (assessed by qRT-PCR and Western blotting) in BE(2)-C cells transfected with HDAC5 or empty vector for 72h and treated with 15 nM panobinostat or solvent control for 48h. GAPDH served as loading control. **E.**, *CD9* expression (assessed by qRT-PCR) in BE(2)-C cells treated up to 12 hours with 1 μ g/ml Actinomycin D, 20 nM panobinostat or solvent control ($n = 3$). **F.**, the mean enrichment (\pm SD, $n = 3$) of *CD9* promoter DNA associated with pan-acetylated histone H4 after 5 hours of 20 nM panobinostat treatment is shown from ChIP-qPCR experiments. BE(2)-C lysates were immunoprecipitated with an antibody against pan-acetylated histone H4, and 0.1% input lysate was used as a loading control for comparing ChIPs. **G.-H.**, *CD9* expression was assessed on the mRNA (qRT-PCR; mean \pm SD, $n = 3$) and protein level (western blotting) in the neuroblastoma cell lines BE(2)-C **G.** and SH-EP **H.** 96 hours after treatment with 3 μ M DAC, 10 nM panobinostat or combined 3 μ M DAC/10 nM panobinostat treatment. β -actin served as loading control. **I.**, *CD9* expression in the neuroblastoma cell lines IMR-32, Kelly and SH-SY5Y 48 hours after combined treatment with 3 μ M DAC and 5 (IMR-32) - 10 nM panobinostat (Kelly, SH-SY5Y) or solvent controls. **J.**, RNA and protein were isolated from BE(2)-C cells 48 hours after combined treatment with 3 μ M DAC/10 nM panobinostat or solvent controls under normoxia (21% O_2) versus hypoxia (3% O_2), serum substitution (10% FCS) versus starvation (0.1% FCS) and normal condition versus genotoxic stress (0.1 μ g/ml doxorubicin). *CD9* expression was measured at the mRNA and protein levels using qRT-PCR (mean fold-change over solvent \pm SD, $n = 3$) and western blotting. DAC, 5-aza-2'-deoxycytidine. * $P < 0.05$; ** $P \leq 0.01$; *** $P \leq 0.001$; # $P = 0.0131$ for BE(2)-C and $P \leq 0.0001$ for SH-EP.

panobinostat, was dose-dependent, and ranged between 2.3- and 3.8-fold on the mRNA level and between 2- and 5.3-fold on the protein level (Figure 5C). Another clinically approved HDAC-inhibitor, vorinostat, weakly inhibited HDAC5, and only induced *CD9* expression by 1.4-fold and *CD9* protein levels by 2-fold (Figure 5C). *CD9* mRNA or protein expression was not induced by the HDAC6-selective inhibitor, tubacin; the HDAC8-selective

inhibitor, compound 2; or by bufexamac, which inhibits both HDAC6 and HDAC10 (Figure 5C). To test whether enforced HDAC5 expression could at least partially counteract *CD9* induction by HDAC inhibitor treatment, BE(2)-C, Kelly and SH-EP cells were transiently transfected with a HDAC5 expression vector then treated with panobinostat. Enforced HDAC5 expression decreased the *CD9* induction triggered by panobinostat

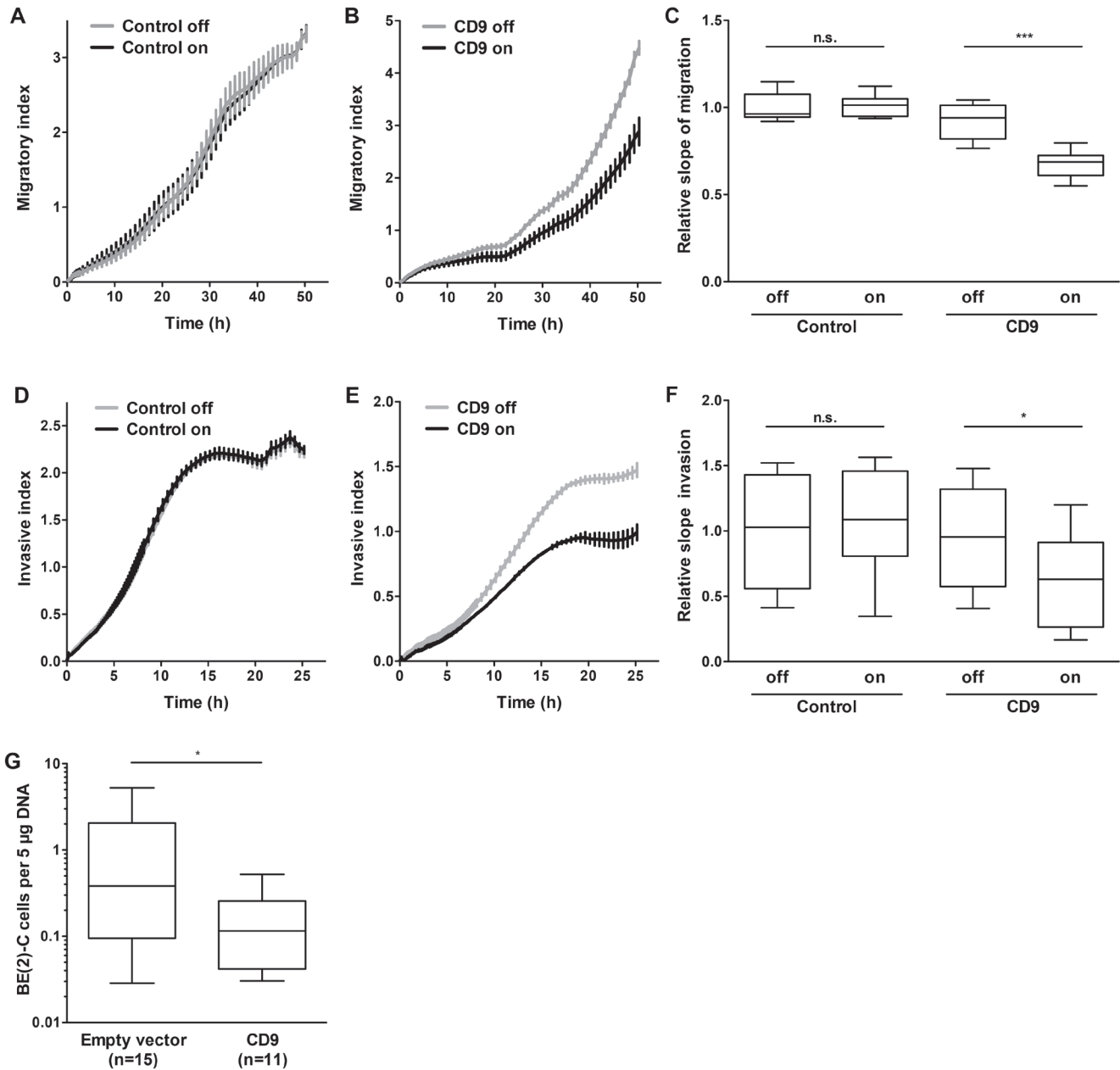


Figure 6: High-level CD9 expression inhibits migration and invasion of neuroblastoma cells in Boyden chambers and metastatic disease to the chicken embryo bone marrow. The stable CD9-inducible SH-EP cell model and the respective stable control cells were used to determine the effect of CD9 expression on migration **A.-C.** and invasion **D.-F.** Migration/invasion was measured with the xCELLigence Real-Time Cell Analyzer ($n = 3$ in quadruplicate). **E.**, number of metastasizing BE(2)-C cells with and without enforced CD9 expression in the bone marrow chicken embryos. Cells were transplanted onto the chorioallantoic membranes of fertilized pathogen-free leghorn eggs ($n \geq 11$ per study group). On day 17, the embryos were sacrificed, and the bone marrow was isolated. Human *ALU* sequences were amplified and quantified by qRT-PCR. n.s., not significant; * $P < 0.05$; *** $P \leq 0.001$.

up to 15% on both transcript and protein levels (Figure 5D, Supplementary Figure S3A-S3B), demonstrating that the effect of pan-HDAC inhibition could be partially outcompeted by HDAC5 overexpression. To differentiate between *de novo* transcript synthesis and increased transcript stability, we cotreated BE(2)-C cells with actinomycin D and panobinostat or solvent control for up to 12h. Blocking *de novo* RNA synthesis with actinomycin D resulted in equivalent *CD9* expression in cells treated with HDAC inhibitor and untreated control cells, confirming that control is enacted *via* transcriptional activation (Figure 5E). To examine epigenetic changes prior to *CD9* transcriptional activation, we performed chromatin immunoprecipitation (ChIP) in BE(2)-C cells treated for 5h with panobinostat or solvent control using an antibody against pan-acetylated histone H4, and looked for changes in the *CD9* promoter. We detected an increase in pan-acetylated histone H4 associated with the transcriptional start site of the *CD9* gene after HDAC inhibitor treatment, indicating that epigenetic changes preceded transcriptional activation (compare Figure 5F with Figure 1C). Taken together, *CD9* is epigenetically and transcriptionally repressed by the chromatin-modifying enzyme, HDAC5, in association with MYCN, and *CD9* expression can be triggered in neuroblastoma cells by

inhibition of HDAC5.

Next, we assessed whether co-treatment with a nucleoside inhibitor of DNA methylation, 5-Aza-2'-deoxycytidine (DAC), could enhance transcriptional activation. Combined treatment of BE(2)-C and SH-EP neuroblastoma cells with panobinostat and DAC synergistically induced *CD9* up to 8-fold on the transcript and 6.5-fold on the protein levels (Figure 5G-5H). We also achieved similar results using the IMR-32, Kelly and SH-SY5Y neuroblastoma cell lines (Figure 5I). We next analyzed the effectiveness of *CD9* induction in BE(2)-C cells by combined panobinostat/DAC treatment under conditions mimicking those in tumors such as hypoxia, starvation and stress induced by cytotoxic drugs such as doxorubicin. Starvation caused by culture for 72h in serum-free medium strongly reduced endogenous *CD9* levels, while both hypoxia and genotoxic stress had no major effect on *CD9* expression (Figure 5J). Drug treatment triggered *CD9* expression by 4- to 5-fold compared to controls in all culture conditions (Figure 5J), suggesting transcription can still be robustly activated. Our data argue a role for HDAC5 in transcriptionally repressing *CD9* in neuroblastoma cells beyond the repression achieved by MYCN and DNA methyltransferases, and offers an avenue for pharmacological intervention.

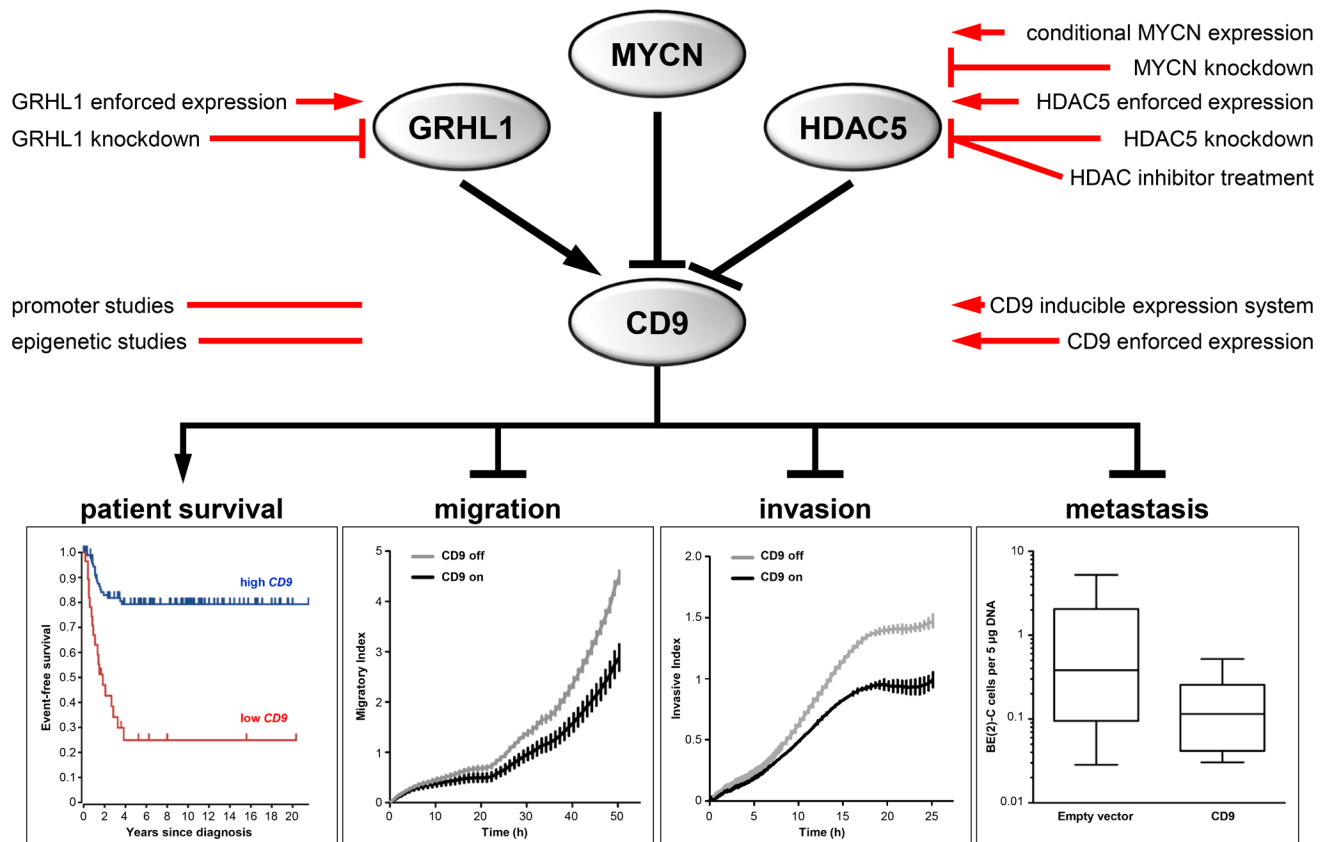


Figure 7: Schematic model summarizing the expression, regulation and function of *CD9* in neuroblastoma and indicating the experimental strategies applied.

CD9 attenuates neuroblastoma cell migration, invasion and metastasis formation

Data obtained from primary tumors suggested that high-level *CD9* expression is associated with favorable tumor biology. To investigate whether high-level *CD9* expression exerts its antitumorigenic effect *via* influencing proliferation and viability, SH-EP cells stably transfected with an inducible *CD9* construct or a negative control expression system (Supplementary Figure S4A-S4B) were analyzed in proliferation assays. *CD9* induction did not significantly alter cell proliferation or viability in the SH-EP cell model (Supplementary Figure S5A-S5B). Similarly, transient enforced *CD9* expression did not alter BE(2)-C cell proliferation or viability compared to control cells transfected with empty or LacZ vectors (Supplementary Figure S4C-S4D and Supplementary Figure S5C-S5D). Xenograft tumor take or growth rate was also not influenced by prior transient transfection with either *CD9* or LacZ control vector in BE(2)-C cells subcutaneously implanted into CB17-SCID mice (Supplementary Figure S5E). These experiments strongly indicate that *CD9* exerts its antitumoral effects *via* other avenues than influencing tumor cell proliferation. To investigate whether migratory or invasive capacity was affected, we functionally analyzed the phenotypic consequences of inducing *CD9* expression in the SH-EP cell model during Boyden chamber migration and invasion assays. Inducing *CD9* expression reduced cell migration by 40%, while inducing the negative control expression system produced no difference in cell migration (Figure 6A-6C). Similarly, *CD9* induction inhibited invasion capacity by 40%, while inducing the negative control expression system again had no effect (Figure 6D-6F). We next evaluated whether our *in vitro* findings were also applicable in the *in vivo* chicken embryo chorioallantoic membrane metastasis assay. The number of BE(2)-C cells that metastasized from the chorioallantoic membrane to the chicken embryonic bone marrow was strongly reduced by enforced *CD9* expression compared with empty vector-transfected control cells (Figure 6G). SH-EP cells, which do not metastasize in the chicken embryo chorioallantoic membrane metastasis assay, were not investigated. Transient enforced expression was applied in BE(2)-C cells as inducible gene expression by tetracycline addition is technically not feasible in the chicken embryo chorioallantoic membrane metastasis assay. Taken together, our data supports a mode of action by *CD9*, in which high-level expression in neuroblastoma cells exerts an antitumoral effect by inhibiting several different steps in the invasion-metastasis cascade (schematic model shown in Figure 7).

DISCUSSION

This study identified *CD9* as a critical and indirectly druggable node in the neuroblastoma invasion-metastasis cascade. This is of particular interest since widespread and extensive metastatic burden causes most neuroblastoma-related deaths, mirroring the perception that activating invasion and metastasis belongs to the cancer hallmarks acquired during the multistep development of human tumors [26]. Suppressing neuroblastoma metastasis would, taken in this light, be a top priority to improve survival of high-risk disease with targeted therapeutic approaches.

Neuroblastoma originates from aberrant neural crest progenitor cells, which physiologically give rise both to sympathetic neurons and chromaffin cells of the adrenal glands [1, 2]. A review of the literature indicates that *CD9* is expressed in early neural crest-derived embryonic rat sympathetic neurons and adrenal chromaffin cells. *CD9* expression is also sustained throughout adulthood, localized to the surface of nerve cell bodies, axons and growth cones [27]. Yet, so far, no role for *CD9* has been described in neuroblastoma pathogenesis. We detected low-level *CD9* expression in primary neuroblastomas that correlated with metastasized high-risk disease, low probability of patient survival and established clinical and molecular markers for unfavorable disease including *MYCN* amplification in the tumor. High-level *CD9* expression in primary neuroblastomas served as an independent favorable prognostic marker. Patients stratified into subgroups based on different *CD9* expression levels in the tumors also segregated known prognostic markers such as age at diagnosis (\leq or $>$ 18 months), and INSS tumor stages 3 or 4 *versus* stages 1, 2 or 4S. The major strength of *CD9* tumor expression as a novel prognostic marker may be its potential for predicting disease progression in neuroblastoma patient subgroups.

Using functional assays, we demonstrated that *CD9* suppresses neuroblastoma cell migration and invasion. This is in line with evidence from studies with ovarian [28] and bladder carcinoma cell lines [29] that low-level *CD9* expression triggers tumor cell motility and invasiveness. On a molecular level, *CD9* is known to inhibit integrin-mediated motility and associate with the platelet aggregation-inducing factor podoplanin, thereby preventing platelet aggregation and, consequently, a tumor-cell protective microenvironment (reviewed in [30]). We moved beyond *in vitro* assays for invasion and metastasis, to an *in vivo* model where human neuroblastoma cells explanted to the chicken chorioallantoic membrane invade the chicken embryo bone marrow, thus, reflecting human neuroblastoma metastatic behavior. Enforced *CD9* expression strongly inhibited the number of neuroblastoma cells metastasized to the chicken bone marrow demonstrating that *CD9* acts to inhibit the invasion-metastasis cascade in an *in vivo* neuroblastoma model. These data indicate that pharmacological strategies

to raise CD9 expression in the tumor could potentially prevent an increasing burden of metastatic colonization in high-risk neuroblastoma patients.

Here we identified GRHL1 as a transcriptional activator of the *CD9* gene and showed that MYCN and HDAC5 transcriptionally repress *CD9* in neuroblastoma. Our experiments with histone deacetylase inhibitors demonstrated the potential indirect druggability of *CD9*. Statistical models identified the synergistic effect of combining an HDAC inhibitor and the DNA methyltransferase inhibitor, 5'-azadeoxycytidine, supporting the observation that epigenetic events can be remodeled by pharmacological intervention. Previous investigations by us and others [31-35] have delineated and deciphered specific functions of single HDACs in neuroblastomas with defined molecular profiles. We showed that HDAC2 cooperates with MYCN to suppress apoptosis mediated by miR-183 [36], and that HDAC3 interacts with MYCN to transcriptionally repress the *GRHL1* transcription factor, which exerts tumor suppressive effects in neuroblastoma [19]. We demonstrated that HDAC8 inhibits neuroblastoma cell differentiation [37] and HDAC10 promotes autophagy-mediated neuroblastoma cell survival [38]. We detected elevated levels of HDAC11 expression in neuroblastomas and several carcinomas compared to corresponding healthy tissues, and showed that HDAC11 depletion was sufficient to cause apoptosis and inhibit metabolic activity in different cancer cell lines, representing major oncological indications for industrial drug development programs [39]. Beyond the previously described specific and non-redundant oncogenic functions of HDAC2, HDAC3, HDAC8, HDAC10 and HDAC11 in neuroblastoma pathophysiology, this study defines a role for HDAC5 in triggering the invasion-metastasis cascade in neuroblastoma.

As we are only at the beginning of understanding the potential therapeutic use of HDAC inhibition strategies and their combination within complex rationales utilizing several epigenetic-cancer therapies, further studies are required to define how to best construct drug strategies targeting epigenetic players such as HDACs and bromodomain and extra-terminal domain (BET) protein family members. Taken together, the present study (i) identifies low-level *CD9* expression as a novel prognostic marker in high-risk neuroblastoma patient subgroups, (ii) exposes *CD9* transcriptional activation by GRHL1 and its repression by MYCN and HDAC5, (iii) defines the potential of high-level CD9 expression to interrupt the neuroblastoma invasion-metastatic cascade and (iv) demonstrates the indirect druggability of CD9 by combined inhibition of HDAC5 and DNA methyltransferase inhibitors.

MATERIALS AND METHODS

Tumor samples and cell culture

Formalin-fixed, paraffin-embedded samples from 20 primary neuroblastomas (13 localized or stage 4S and 7 stage 4, 3 of which harboring *MYCN* amplifications) available from patients in the NB2004 trial (informed consent available) were selected for CD9 immunohistochemistry. Authenticated neuroblastoma cell lines and models were maintained under standard cell culture conditions. Details are supplied in Supplementary Materials and Methods.

Quantitative reverse-transcriptase real-time PCR (qRT-PCR)

Total RNA was isolated from cell lines using the RNeasy Mini Kit (Qiagen). The Thermo Scientific First Strand cDNA Synthesis Kit was used to transcribe cDNAs for qRT-PCR analysis. Relative gene expression was measured in qRT-PCR using SYBR Green Dye (Eurogentec) and primers (Supplementary Table S1) on an ABI Prism 7500 thermal cycler (Perkin-Elmer Applied Biosystems), and normalized to the averaged expression of *ACTB* and *18S* rRNA or the averaged expression of *SDHA* and *HPRT1*, a gene pair that is consistently expressed in stage 4 and 4S neuroblastomas [40]. Data were analyzed using Applied Biosystems 7500 software v2.0.5, and changes in expression were calculated using the $\Delta\Delta C_t$ method [41].

In vitro cell migration and invasion

Migration and invasion of cells was measured using the CIM-Plate 16 Boyden chamber system (ACEA Biosciences) and the xCELLigence Real-Time Cell Analyzer (ACEA Biosciences) [42, 43]. Boyden chambers were prepared for invasion assays by adding 160 μ l FCS-containing medium to the lower chamber and successively adding 50 μ l serum-free medium, 20 μ l Matrigel diluted 1:40 in serum-free medium followed by 4h at 37°C, 5% CO₂ and 30 μ l serum-free medium above the Matrigel to the upper chamber. After 24h in serum-free medium, 6x10⁴ cells were seeded into the upper chamber, and migrating/invading cells were monitored by serial impedance measurements [42, 43]. Assays were performed in quadruplicate and repeated three times. Cell migration and invasion were calculated using RTCA Software version 2.0 (ACEA Biosciences).

TaqMan[®]-based chicken embryo metastasis assay

Fertilized pathogen-free leghorn eggs (Charles River) were incubated at 37°C in 60% humidity with rotation in a Marsh Incubator (Lyon Electric). The allantoic vein was localized at day 10, over which a window was cut in the shell using a miniaturized rotating electric saw (Dremel) after removing air from the natural air space under mild vacuum pressure. Single-cell suspensions of empty vector- or CD9 plasmid-transfected BE(2)-C cells were prepared in serum-free medium, 2×10^6 cells inoculated onto each chorioallantoic membrane and the windows sealed with cellotape. Embryos were sacrificed on day 17 ($n = 18$ per study group), and bone marrow was isolated and snap frozen for preparation of genomic DNA. The TaqMan-based ALU assay takes advantage of the abundantly present *ALU* family of short interspersed repeated DNA elements in the human genome to identify human DNA [44]. PCR reactions were carried out as described [45] using 2x qRT-PCR MasterMix (Eurogentec) and primers for YB8-ALU-S68, YB8-ALU-AS244 and YB8-ALU-167 (Supplementary Table 1). A standard curve was generated from DNA samples from 10,000, 1000, 100, 10 and 0 BE(2)-C cells mixed with 1 μ g of chicken liver genomic DNA each to quantify the BE(2)-C cells metastasized to the chicken bone marrow.

TH-MYCN neuroblastoma progression model

TH-MYCN^{+/+} mice [46] were sacrificed at day 7 ($n = 4$) and day 14 ($n = 4$) of life to harvest sympathetic ganglia containing hyperplastic neuroblast foci, and at week 6 of life to harvest advanced neuroblastic tumors ($n = 4$). Sympathetic ganglia were also dissected from TH-MYCN^{-/-} mice at day 7 ($n = 4$), day 14 ($n = 4$) and week 6 ($n = 4$) of life to assess gene expression changes during normal development. Total RNA was isolated using the miRNeasy Mini Kit, and samples profiled on Agilent SurePrint G3 Gene Expression Microarrays according to the manufacturers' protocols. Data were summarized and normalized with the vsn method [47] in the R statistical programming language [48] using the *limma* package [49]. Linear regression analysis was performed to evaluate differential temporal expression in ganglia from wildtype mice and ganglia and tumors from transgenic mice.

Subcutaneous BE(2)-C xenograft tumor model in CB17-SCID mice

BE(2)-C cells were transfected for 24h with LacZ- or CD9-vectors in culture, then 2×10^6 viable cells suspended in 200 μ l Matrigel (BD Biosciences) were subcutaneously implanted in the flanks of 6-week old female CB17-SCID mice ($n = 12$ per study group). Animals were sacrificed 13 days after grafting. Tumor size was measured daily with a caliper, and volume

calculated by $\pi/6(w_1 \times w_2 \times w_2)$, where w_1 equals largest tumor diameter, and w_2 equals smallest tumor diameter. Experiments conformed to regulatory standards and were approved by the local ethics committee.

Data analysis

CD9 mRNA expression values were calculated using existing whole-genome expression profiles from 476 primary neuroblastomas [21], and validated using whole-genome expression profiles from 122 further neuroblastomas [22]. Kaplan-Meier curves were generated for the 476 neuroblastoma cohort using the R Survival package and compared using log-rank testing. The *CD9* probesets with the highest average signal were selected for analysis. Relapse, progression and death from disease were considered events for calculation of event-free survival. Both overall and event-free survival were calculated from the time of diagnosis to death, event or last examination. Optimal cut-offs for overall and event-free survival analyses were calculated using the R Maxstat package according to the method defined by Lausen and Schumacher [50] determining the expression value best separating outcome into two groups using a maximally selected two-sample log-rank statistic. Optimal cut-offs were 4774.47 (overall survival) and 5330.71 (event-free survival). Results were corrected for multiple testing. *CD9* expression between two patient subgroups were compared using the Mann-Whitney U test. Univariate and multivariate analyses were performed for overall and event-free survival using Cox proportional hazards regression models using SPSS (IBM, 20.0.0 release) or R (version 3.0.1). The R2 platform (<http://r2.amc.nl>) was used to generate Kaplan-Meier survival curves and box plots of *CD9* expression comparisons for the 122-neuroblastoma cohort [22]. *CD9* gene methylation status was derived from existing methyl-CpG-binding domain sequencing data from 60 internationally collected primary neuroblastoma samples [24, 25]. Single values from cell culture experiments were compared using the one-sample *t*-test in GraphPad Prism version 5.0 (GraphPad Software Inc., La Jolla, CA). *CD9* expression in cell groups transfected with either of two negative controls or either of two HDAC-specific siRNAs were compared using a mixed linear model with fixed-factor treatment and random intercept for each siRNA using SAS PROC MIXED, Version 9.2 (SAS Institute Inc., Cary, NC, USA). Delta-Ct values were evaluated by a two-factorial mixed linear model with interaction and random effect for each experimental run to assess potential synergistic effects of combined HDAC inhibitor/5-aza-cytidine treatment on *CD9* expression. *P* values below 0.05 were considered significant.

ACKNOWLEDGMENTS

We thank the German Neuroblastoma Tumor Bank (Cologne) for providing tumor samples, and the Imaging and Cytometry Core Facility of the DKFZ for their valuable services.

CONFLICTS OF INTEREST

The authors disclosed no potential conflicts of interest.

FINANCIAL SUPPORT

This work was supported by the BMBF through e:MED SYSMED-NB (M. Fischer, A. Eggert, J.H. Schulte, H.E. Deubzer) and e:MED SMOOSE (M. Fischer, J.H. Schulte), by the Berlin Institute of Health (BIH) through TERMINATE-NB (A. Eggert, J.H. Schulte, H.E. Deubzer) and a translational PhD project grant (H.E. Deubzer), by the Deutsche Krebshilfe (M. Lodrini, H. Allgayer, M. Fischer, H.E. Deubzer), by the Wilhelm Sander-Stiftung (H. Allgayer, M. Zoeller), by the Kind-Philipp-Stiftung through doctoral scholarships (D. Opitz, T.M. Thole) and by the DAAD through a graduate scholarship (J. Hu).

REFERENCES

1. Maris JM. Recent advances in neuroblastoma. *New Engl J Med.* 2010; 362:2202-2211.
2. Brodeur GM and Bagatell R. Mechanisms of neuroblastoma regression. *Nat Rev Clin Oncol.* 2014; 11:704-713.
3. Pinto NR, Applebaum MA, Volchenboum SL, Matthay KK, London WB, Ambros PF, Nakagawara A, Berthold F, Schleiermacher G, Park JR, Valteau-Couanet D, Pearson AD and Cohn SL. Advances in risk classification and treatment strategies for neuroblastoma. *J Clin Oncol* 2015; 33:3008-3017.
4. Schwab M, Alitalo K, Klempnauer KH, Varmus HE, Bishop JM, Gilbert F, Brodeur G, Goldstein M and Trent J. Amplified DNA with limited homology to myc cellular oncogene is shared by human neuroblastoma cell lines and a neuroblastoma tumour. *Nature.* 1983; 305:245-248.
5. Brodeur GM, Seeger RC, Schwab M, Varmus HE and Bishop JM. Amplification of N-myc in untreated human neuroblastomas correlates with advanced disease stage. *Science.* 1984; 224:1121-1124.
6. Peifer M, Hertwig F, Roels F, Drexler D, Gartlgruber M, Menon R, Kramer A, Roncaioli JL, Sand F, Heuckmann JM, Ikram F, Schmidt R, Ackermann S, Engesser A, Kahlert Y, Vogel W, et al. Telomerase activation by genomic rearrangements in high-risk neuroblastoma. *Nature.* 2015; 526:700-704.

7. Valentijn LJ, Koster J, Zwijnenburg DA, Hasselt NE, van Sluis P, Volckmann R, van Noesel MM, George RE, Tytgat GA, Molenaar JJ and Versteeg R. TERT rearrangements are frequent in neuroblastoma and identify aggressive tumors. *Nat Genet* 2015; 47:1411-4.
8. Cheung NK, Zhang J, Lu C, Parker M, Bahrami A, Tickoo SK, Heguy A, Pappo AS, Federico S, Dalton J, Cheung IY, Ding L, Fulton R, Wang J, Chen X, Becksfors J, et al. Association of age at diagnosis and genetic mutations in patients with neuroblastoma. *JAMA* 2012; 307:1062-1071.
9. Pugh TJ, Morozova O, Attiyeh EF, Asgharzadeh S, Wei JS, Auclair D, Carter SL, Cibulskis K, Hanna M, Kiezun A, Kim J, Lawrence MS, Lichtenstein L, McKenna A, Pedamallu CS, Ramos AH, et al. The genetic landscape of high-risk neuroblastoma. *Nature Genet* 2013; 45:279-284.
10. Ambros IM, Brunner C, Abbasi R, Frech C and Ambros PF. Ultra-High Density SNParray in Neuroblastoma Molecular Diagnostics. *Front Oncol* 2014; 4:202.
11. Mosse YP, Laudenslager M, Longo L, Cole KA, Wood A, Attiyeh EF, Laquaglia MJ, Sennett R, Lynch JE, Perri P, Laureys G, Speleman F, Kim C, Hou C, Hakonarson H, Torkamani A, et al. Identification of ALK as a major familial neuroblastoma predisposition gene. *Nature* 2008; 455:930-935.
12. Chen Y, Takita J, Choi YL, Kato M, Ohira M, Sanada M, Wang L, Soda M, Kikuchi A, Igarashi T, Nakagawara A, Hayashi Y, Mano H and Ogawa S. Oncogenic mutations of ALK kinase in neuroblastoma. *Nature* 2008; 455:971-974.
13. Janoueix-Lerosey I, Lequin D, Brugieres L, Ribeiro A, de Pontual L, Combaret V, Raynal V, Puisieux A, Schleiermacher G, Pierron G, Valteau-Couanet D, Frebourg T, Michon J, Lyonnet S, Amiel J and Delattre O. Somatic and germline activating mutations of the ALK kinase receptor in neuroblastoma. *Nature* 2008; 455:967-970.
14. George RE, Sanda T, Hanna M, Frohling S, Luther W, 2nd, Zhang J, Ahn Y, Zhou W, London WB, McGrady P, Xue L, Zozulya S, Gregor VE, Webb TR, Gray NS, Gilliland DG, et al. Activating mutations in ALK provide a therapeutic target in neuroblastoma. *Nature* 2008; 455:975-978.
15. Reyes R, Monjas A, Yanez-Mo M, Cardenas B, Morlino G, Gilsanz A, Machado-Pineda Y, Lafuente E, Monk P, Sanchez-Madrid F and Cabanas C. Different states of integrin LFA-1 aggregation are controlled through its association with tetraspanin CD9. *Biochim Biophys Acta* 2015; 1853:2464-2480.
16. Odintsova E, Butters TD, Monti E, Sprong H, van Meer G and Berdichevski F. Gangliosides play an important role in the organization of CD82-enriched microdomains. *Biochem J* 2006; 400:315-325.
17. Schroder HM, Hoffmann SC, Hecker M, Korff T and Ludwig T. The tetraspanin network modulates MT1-MMP cell surface trafficking. *Int J Biochem Cell Biol* 2013; 45:1133-1144.

18. Uhlén M, Fagerberg L, Hallström BM, Lindskog C, Oksvold P, Mardinoglu A, Sivertsson A, Kampf C, Sjöstedt E, Asplund A, Olsson I, Edlund K, Lundberg E, Navani, Szgyarto CA, Odeberg J, et al. Proteomics. Tissue-based map of the human proteome. *Science* 2015; 347:1260419.
19. Fabian J, Lodrini M, Oehme I, Schier MC, Thole TM, Hielscher T, Kopp-Schneider A, Opitz L, Capper D, von Deimling A, Wiegand I, Milde T, Mählknecht U, Westermann F, Popanda O, Roels F, et al. GRHL1 acts as tumor suppressor in neuroblastoma and is negatively regulated by MYCN and HDAC3. *Cancer Res* 2014; 74:2604-2616.
20. Le Naour F, Prenant M, Francastel C, Rubinstein E, Uzan G and Boucheix C. Transcriptional regulation of the human CD9 gene: characterization of the 5'-flanking region. *Oncogene* 1996; 13:481-486.
21. Oberthuer A, Juraeva D, Li L, Kahlert Y, Westermann F, Eils R, Berthold F, Shi L, Wolfinger RD, Fischer M and Brors B. Comparison of performance of one-color and two-color gene-expression analyses in predicting clinical endpoints of neuroblastoma patients. *Pharmacogenomics J* 2010; 10:258-266.
22. Molenaar JJ, Koster J, Zwijnenburg DA, van Sluis P, Valentijn LJ, van der Ploeg I, Hamdi M, van Nes J, Westerman BA, van Arkel J, Ebus ME, Haneveld F, Lakeman A, Schild L, Molenaar P, Stroeken P, et al. Sequencing of neuroblastoma identifies chromothripsis and defects in neuritogenesis genes. *Nature* 2012; 483:589-593.
23. Oberthuer A, Hero B, Berthold F, Juraeva D, Faldum A, Kahlert Y, Asgharzadeh S, Seeger R, Scaruffi P, Tonini GP, Janoueix-Lerosey I, Delattre O, Schleiermacher G, Vandesompele J, Vermeulen J, Speleman F, et al. Prognostic impact of gene expression-based classification for neuroblastoma. *J Clin Oncol* 2010; 28:3506-3515.
24. Decock A, Ongenaert M, Cannoodt R, Verniers K, De Wilde B, Laureys G, Van Roy N, Berbegall AP, Bienertova-Vasku J, Bown N, Clement N, Combaret V, Haber M, Hoyoux C, Murray J, Noguera R, et al. Methyl-CpG-binding domain sequencing reveals a prognostic methylation signature in neuroblastoma. *Oncotarget* 2016; 7:1960-1972.
25. Decock A, Ongenaert M, Van Criekinge W, Speleman F and Vandesompele J. DNA methylation profiling of primary neuroblastoma tumors using methyl-CpG-binding domain sequencing. *Sci Data* 2016; 3:160004.
26. Hanahan D and Weinberg RA. Hallmarks of cancer: the next generation. *Cell* 2011; 144:646-674.
27. Tole S and Patterson PH. Distribution of CD9 in the developing and mature rat nervous system. *Dev Dyn*. 1993; 197:94-106.
28. Furuya M, Kato H, Nishimura N, Ishiwata I, Ikeda H, Ito R, Yoshiki T and Ishikura H. Down-regulation of CD9 in human ovarian carcinoma cell might contribute to peritoneal dissemination: morphologic alteration and reduced expression of beta1 integrin subsets. *Cancer Res* 2005; 65:2617-2625.
29. Mitsuzuka K, Handa K, Satoh M, Arai Y and Hakomori S. A specific microdomain ("glycosynapse 3") controls phenotypic conversion and reversion of bladder cancer cells through GM3-mediated interaction of alpha3beta1 integrin with CD9. *J Biol Chem* 2005; 280:35545-35553.
30. Zoller M. Tetraspanins: push and pull in suppressing and promoting metastasis. *Nat Rev Cancer*. 2009; 9:40-55.
31. Sun Y, Liu PY, Scarlett CJ, Malyukova A, Liu B, Marshall GM, MacKenzie KL, Biankin AV and Liu T. Histone deacetylase 5 blocks neuroblastoma cell differentiation by interacting with N-Myc. *Oncogene* 2014; 33:2987-2994.
32. Wang C, Liu Z, Woo CW, Li Z, Wang L, Wei JS, Marquez VE, Bates SE, Jin Q, Khan J, Ge K and Thiele CJ. EZH2 mediates epigenetic silencing of neuroblastoma suppressor genes CASZ1, CLU, RUNX3, and NGFR. *Cancer Res* 2012; 72:315-324.
33. Frumm SM, Fan ZP, Ross KN, Duvall JR, Gupta S, VerPlank L, Suh BC, Holson E, Wagner FF, Smith WB, Paranal RM, Bassil CF, Qi J, Roti G, Kung AL, Bradner JE, et al. Selective HDAC1/HDAC2 inhibitors induce neuroblastoma differentiation. *Chem Biol* 2013; 20:713-725.
34. Marshall GM, Gherardi S, Xu N, Neiron Z, Trahair T, Scarlett CJ, Chang DK, Liu PY, Jankowski K, Iraci N, Haber M, Norris MD, Keating J, Sekyere E, Jonquieres G, Stossi F, et al. Transcriptional upregulation of histone deacetylase 2 promotes Myc-induced oncogenic effects. *Oncogene* 2010; 29:5957-5968.
35. Iraci N, Diolaiti D, Papa A, Porro A, Valli E, Gherardi S, Herold S, Eilers M, Bernardoni R, Della Valle G and Perini G. A SP1/MIZ1/MYCN repression complex recruits HDAC1 at the TRKA and p75NTR promoters and affects neuroblastoma malignancy by inhibiting the cell response to NGF. *Cancer Res* 2011; 71:404-412.
36. Lodrini M, Oehme I, Schroeder C, Milde T, Schier MC, Kopp-Schneider A, Schulte JH, Fischer M, De Preter K, Pattyn F, Castoldi M, Muckenthaler MU, Kulozik AE, Westermann F, Witt O and Deubzer HE. MYCN and HDAC2 cooperate to repress miR-183 signaling in neuroblastoma. *Nucleic Acids Res* 2013; 41:6018-6033.
37. Oehme I, Deubzer HE, Wegener D, Pickert D, Linke JP, Hero B, Kopp-Schneider A, Westermann F, Ulrich SM, von Deimling A, Fischer M and Witt O. Histone deacetylase 8 in neuroblastoma tumorigenesis. *Clin Cancer Res* 2009; 15:91-99.
38. Oehme I, Linke JP, Bock BC, Milde T, Lodrini M, Hartenstein B, Wiegand I, Eckert C, Roth W, Kool M, Kaden S, Grone HJ, Schulte JH, Lindner S, Hamacher-Brady A, Brady NR, et al. Histone deacetylase 10 promotes autophagy-mediated cell survival. *Proc Natl Acad Sci U S A* 2013; 110:E2592-2601.
39. Deubzer HE, Schier MC, Oehme I, Lodrini M, Haendler B, Sommer A and Witt O. HDAC11 is a novel drug target in carcinomas. *Inter J Cancer*. 2013; 132:2200-2208.

40. Fischer M, Skowron M and Berthold F. Reliable transcript quantification by real-time reverse transcriptase-polymerase chain reaction in primary neuroblastoma using normalization to averaged expression levels of the control genes HPRT1 and SDHA. *J Mol Diagn* 2005; 7:89-96.
41. Deubzer HE, Ehemann V, Kulozik AE, Westermann F, Savelyeva L, Kopp-Schneider A, Riester D, Schwab M and Witt O. Anti-neuroblastoma activity of *Helminthosporium carbonum* (HC)-toxin is superior to that of other differentiating compounds in vitro. *Cancer Lett* 2008; 264:21-28.
42. Liang XJ, Choi Y, Sackett DL and Park JK. Nitrosoureas inhibit the stathmin-mediated migration and invasion of malignant glioma cells. *Cancer Res* 2008; 68:5267-5272.
43. Scrace S, O'Neill E, Hammond EM and Pires IM. Use of the xCELLigence system for real-time analysis of changes in cellular motility and adhesion in physiological conditions. *Methods Mol Biol* 2013; 1046:295-306.
44. Batzer MA and Deininger PL. A human-specific subfamily of Alu sequences. *Genomics* 1991; 9:481-487.
45. van der Horst EH, Leupold JH, Schubbert R, Ullrich A and Allgayer H. TaqMan-based quantification of invasive cells in the chick embryo metastasis assay. *Biotechniques* 2004; 37:940-942, 944, 946.
46. Weiss WA, Aldape K, Mohapatra G, Feuerstein BG and Bishop JM. Targeted expression of MYCN causes neuroblastoma in transgenic mice. *EMBO J* 1997; 16:2985-2995.
47. Wu Z, Irizarry RA, Gentleman R, F.M M and Spencer F. A model-based background adjustment for oligonucleotide expression arrays. *J Am Stat Assoc* 2004; 99:909-917.
48. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria 2013.
49. Smyth GK. Limma: linear models for microarray data. In: 'Bioinformatics and Computational Biology Solutions using R and Bioconductor'. Springer, New York; 2005:397-420.
50. Lausen B and Schumacher M. Maximally selected rank statistics. *Biometrics* 1992; 48:85.