

# THE UNIVERSITY of EDINBURGH

# Edinburgh Research Explorer

# Receptive field estimation in large visual neuron assemblies using a super-resolution approach

#### Citation for published version:

Pamplona, D, Hilgen, G, Hennig, MH, Cessac, B, Sernagor, E & Kornprobst, P 2022, 'Receptive field estimation in large visual neuron assemblies using a super-resolution approach', *Journal of Neurophysiology*, vol. 127, no. 5, pp. 1334-1347. https://doi.org/10.1152/jn.00076.2021

### **Digital Object Identifier (DOI):**

10.1152/jn.00076.2021

Link: Link to publication record in Edinburgh Research Explorer

**Document Version:** Peer reviewed version

Published In: Journal of Neurophysiology

#### General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

#### Take down policy

The University of Édinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.



# Receptive field estimation in large visual neuron assemblies using a super-resolution approach \*

Daniela Pamplona <sup>1,2</sup>	<sup>†</sup> Gerrit Hilgen <sup>3,4</sup>	Matthias H $\mathrm{Hennig}^5$
Bruno $Cessac^2$	Evelyne Sernagor <sup>3</sup>	Pierre Kornprobst <sup>2‡</sup>

<sup>1</sup>Ecole Nationale Supérieure de Techniques Avancées, Institut Polytechnique de Paris, U2IS, 828 Boulevard des Marchaux, 91120 Palaiseau, France <sup>2</sup>Université Côte d'Azur, Inria, France

 <sup>3</sup>Biosciences Institute, Faculty of Medical Sciences, Newcastle University, Framlington Place, Newcastle upon Tyne, NE2 4HH, United Kingdom
 <sup>4</sup>Applied Sciences, Health and Life Sciences, Northumbria University, Elison Building, Newcastle upon Tyne, NE1 8ST, United Kingdom
 <sup>5</sup>Institute for Adaptive and Neural Computation, School of Informatics, University of Edinburgh, 10 Crichton Street, Edinburgh, EH8 9AB, United Kingdom

#### Abstract

Computing the spike-triggered average (STA) is a simple method to estimate linear receptive fields (RFs) in sensory neurons. For random, uncorrelated stimuli the STA provides an unbiased RF estimate, but in practice, white noise at high resolution is not an optimal stimulus choice as it usually evokes only weak responses. Therefore, for a visual stimulus, images of randomly modulated blocks of pixels are often used. This solution naturally limits the resolution at which an RF can be measured. Here we present a simple super-resolution technique that can be overcome these limitations. We define a novel stimulus type, the shifted white noise (SWN), by introducing random spatial shifts in the usual stimulus in order to increase the resolution of the measurements. In simulated data we show that the average error using the SWN was 1.7 times smaller than when using the classical stimulus, with successful mapping of 2.3 times more neurons, covering a broader range of RF sizes. Moreover, successful RF mapping was achieved with brief recordings of light responses, lasting only about one minute of activity, which is more than 10 times more efficient than the classical white noise stimulus. In recordings from mouse retinal ganglion cells with large scale multi-electrode arrays, we successfully mapped 21 times more RFs than when using the traditional white noise stimuli. In summary, randomly shifting the usual white noise stimulus significantly improves RFs estimation, and requires only short recordings.

<sup>\*</sup>This work was partially supported by the EC IP project FP7-ICT-2011-9 no. 600847 (RENVISION) <sup>†</sup>Corresponding author. Email address: daniela.pamplona@ensta-paris.fr

<sup>&</sup>lt;sup>‡</sup>Co-corresponding author. Email address: pierre.kornprobst@inria.fr

# 1 New & Noteworthy

We present a novel approach to measure receptive fields in large and heterogeneous populations of sensory neurons recorded with large-scale, high-density multielectrode arrays. Our approach leverages super-resolution principles to improve the yield of the spike-triggered average method. By simply designing a new stimulus, we provide experimentalists with a new and fast technique to simultaneously detect more receptive fields at higher resolution in population of hundreds to thousands of neurons.

# 2 Introduction

Sensory neurons are characterised by their receptive field (RF), which is the area of the sensory space they respond to upon stimulation. In visual neurons it is the area of the visual field these cells respond to when light intensity changes. Estimating the position, size and shape of an RF with high accuracy requires measurements at sufficiently high spatial resolution. Ideally, RF measurements should consist of sampling at very high resolution, which means stimulating small subunits (pixels) of the RF in sequence. However, when these pixels are too small, detectable neural response are unlikely to occur because cells usually respond to simultaneous stimulation of many pixels in their RF. On the other hand, when the pixel size is too large, responses do not reflect RF position, sizes and shapes faithfully. This problem is exacerbated by the fact that RF positions and sizes are not homogeneous across the neuronal population. For example, many retinal ganglion cells (RGCs) types have smaller RFs in the centre of the retina than in the periphery, hence the optimal pixel size to determine central RFs is smaller than for measurements in the periphery. Owing to new technological developments in recording approaches consisting of large-scale, high-density multi-electrode arrays (MEAs), it is now possible to record responses to light from hundreds to thousands of neurons simultaneously [3], encompassing both central and peripheral cells. Such an experimental scenario requires designing new stimuli that can yield high-resolution measurements for all cells across the neural population, regardless of their size. In this study, we present a novel approach to measure RFs at high fidelity from large and heterogeneous neural populations recorded simultaneously. The classical way of measuring RFs is to estimate the spike-triggered average (STA) from evoked neural recordings. In short, STA calculates the average stimulus before a spike. If the stimulus is white noise and the recording sufficiently long, then this average corresponds to the neuron's RF[21, 10]. The practical implementation of a "white noise" stimulus consists of a series of non-overlapping binary images shown successively in time, with individual images showing a black or white pixel of similar size presented in random order but with equal probability. This stimulus, here termed basic white noise (BWN), has the size of the blocks as parameter.

In the case of single-cell recordings, the block size is defined according to the experimenter's expectations: it must be smaller than the expected RF size in order to yield high-resolution measurements, but it cannot be too small, in order to avoid weak neural responses. Various more or less heuristic approaches could, in principle, fulfill these conditions. For instance, one can start from a tiny block size and gradually increase it during the experiment in the direction of the larger stimulus-neural response correlation [9] or mutual information [15, 16]. However, in the case of large neuronal populations, one cannot merely apply the same procedure as for individual cells. Indeed, a optimal block size for one neuron will be sub-optimal for another one in the population. Similarly, a high stimulus-neural response correlation (or mutual information) for one neuron might be low for another one. As a consequence, experimenters must design stimuli that best suit the population as a whole, taking into account its heterogeneity. Here we use a novel white noise stimulus, the shifted white noise (SWN). The size of each block is large, ensur-

ing strong responses from all RGCs. However, the blocks are shifted only by a fraction of their size, yielding high-resolution sampling. In other words, large pixels ensure strong light responses while sub-pixel shifts yield super-resolution measurements, enabling us to reliably measure all RFs with great accuracy.

Super-resolution is a class of image processing methods used to estimate high-resolution images from a set of low-resolution ones [30]. It has been successfully implemented in a variety of applications such as, e.g., video [14], remote sensing [29, 18], and medical imagery [11, 23]. In a nutshell, to increase the resolution of RFs, we propose to use low resolution images, meaning with high block size, while introducing additional spatial variability in the stimulus to improve the accuracy of the responses. Our method ensures ensures that the majority of the cells rapidly respond to the low resolution stimulus, while at the same time, the resulting RFs have much higher resolution.

# A novel class of visual stimuli

The classical stimulus to estimate RFs, here called BWN, is illustrated in Fig. 1(A). It consists of a sequence of binary images showing equal-sized blocks, with colors, black or white, drawn randomly from a Bernoulli distribution with a probability 0.5. Each image is displayed for a fixed time. Since the STA relies on averaging stimuli within time windows, the resulting spatial precision (resolution) of the estimated RF is equal to the BWN's block size. In order to increase resolution of the measurements, the most obvious approach would be to decrease the block size, as shown in Fig. 1(B). However, this decreases the responsiveness of most neurons and computing the STA reliably would require much longer recordings.

The super-resolution approach used here preserves large block sizes to guarantee stronger responses, but it randomly shifts the binary images in space at each presentation to increase the stimulus spatial resolution. Two such examples are shown in Fig. 1(C–D). In isolation these shifted blocks will not change responses compared to BWN blocks of similar size. However, combining these low-resolution estimates with the shifts will yield significantly higher resolution RFs.

This allows using the following strategy. Let us denote by  $\beta$  the block size of the stimuli, and define the target resolution for the RF as  $\alpha = \beta/k$ , where k denotes the increase in resolution (e.g. double resolution: k = 2). The target resolution defines a baseline shift of the same value from which one can define a series of random spatial shifts  $s = n\alpha$  with  $n \in \{0, \ldots, k-1\}$ . Considering blocks of BWN images, two independent shifts are applied to each block, one vertical and one horizontal, yielding a succession of blocks characterized by the same shift size, but in random direction. Using the STA for each block provides one RF per block. These are then combined to get a high-resolution RF.

Instead of applying the STA for each block, we randomize the different shifts and apply the STA globally, yielding to what we call the SWN. It is illustrated in Fig. 1(E). The main advantage of SWN over BWN is that the high-resolution RF directly results from the STA rather than having to estimate intermediary low-resolution RFs and then combine them.

In summary, while the resolution of the RF is given by the block size with BWN, it is independent of block size with SWN. Instead, it is provided by the baseline shift. This provides the experimenter with the option of choosing a sufficiently large block size to increase activity levels in response to the stimulus. Since the resolution is given by the block shift size, the only remaining limitations may be technical such as the size of the projected pixels, something inherent to the experimental set up and light stimulation equipment. Another significant advantage of SWN is that it introduces more variability, favoring better light responses across the overall



Figure 1: Exploring variation across white noise stimuli: From BWN to SWN. (A) BWN-B32: BWN with blocks of  $32 \,\mu\text{m}$ . In the BWN case, the resolution of the RF is the same as the size of the blocks. (B) BWN-B4: same as (A) but with blocks of size  $4 \,\mu\text{m}$ . (C) and (D): Series of BWN-B32-like stimuli based on example (A), with an additional fixed spatial shift. The spatial shift is represented by a red square showing how the upper-left block has been moved. The resolution of the STA is still the same as the size of the blocks. However, the different shifts will infer different samplings of the RF that could be combined to achieve a high-resolution RF (see text for details). Note that starting from (A) was chosen only for explanatory purposes so that readers could compare both conditions. The binary patterns should be a priori random. (E) SWN-B32-S4: SWN with blocks of 32 µm and random spatial shifts using a baseline shift of 4 µm. With this condition, it is the baseline shift that defines the resolution of the RF.

population. In other words, we expect not only to obtain RFs of high-resolution, but also to be able to define the RFs for more cells over shorter stimulation trials. <sup>1</sup>

The usual BWN has spatial correlations. To measure these correlations, we estimated the average spatial power spectrum of 10000 images of  $100 \times 100$  pixels varying the block size. These correlations depend on the block size as shown in Fig. 2 A. The power spectrum is flat, meaning the stimulus is white, only for block size 1 pixel, while for the others it has shape of a sinc function whose bandwidth increases with the inverse of the block size. The SWN has correlations as well. As the power spectrum is shift-invariant, the power spectrum of a SWN with a given block size is equal to the power spectrum of a BWN of the same size. Similarly, to measure these correlations, we estimated the average spatial power spectrum of 10000 images  $100 \times 1000$  pixels varying the block and the shift size. The power spectrum of SWN has shape of a sinc and, as for the BWN, the bandwidth increases with the inverse of the block size. However, when looking in detail, there are differences between the power spectrum of BWN and SWN at low frequencies. The BWN is closer to zero than the SWN. These correlations depend on the block but not on the shift size as illustrated in Fig. 2 B.

For the remainder of this paper, the stimuli are named as follows: BWN-B $\beta$  denotes a BWN with a block size  $\beta$ ; SWN-B $\beta$ -S $\alpha$  denotes a SWN with a block size  $\beta$  and a baseline shift  $\alpha$ . Sizes are expressed in µm.

# Methods

#### Synthetic data: Stimuli

In the experimental analysis we consider three stimuli: two BWNs of low and high resolution  $(32 \,\mu\text{m} \text{ and } 4 \,\mu\text{m}, \text{ respectively})$  and one SWNs whose STA is of resolution  $4 \,\mu\text{m}$  (with blocks of  $160 \,\mu\text{m}$  and shifts of  $4 \,\mu\text{m}$ ).

For each stimulus we generated 27,000 images of  $88 \times 88$  pixels, where one pixel corresponds to a square of size  $16 \,\mu\text{m}^2$ . Here we arbitrarily set the origin at the central pixel of the spatial domain, which will be more convenient to specify neurons' population. Images were refreshed at 30.3Hz, or in other words, each image was presented for 33 ms.

Each stimulus was fed independently to our artificial population of neurons described below to obtain the simulated spiking output and then the RF estimation using STA.

#### Synthetic data: Artificial retinal Ganglion cell model

We defined a population of neurons described by linear-Nonlinear Poisson (LNP) models [20, 7]. These functional models are widely used by experimentalists to characterize the cells that they record, map their RFs, and characterize their spatio-temporal feature selectivities [13, 8, 5, 26, 2]. The STA of an LNP neuron stimulated with random white noise converges to the RF of the neuron, up to a multiplicative constant [21]. In its simplest form, an LNP model is a convolution of the stimulus L with a spatio-temporal kernel K followed by a static non-linearity and stochastic (Poisson-like) mechanisms of spikes generation. Here we use this model to simulate RGCs' spiking activity in response to our stimuli. We consider that K is a Difference-of-Gaussians centered at  $(c_x, c_y)$ , center size  $\sigma_c$  and surround size  $\sigma_s$ . The detailed definition is given below.

The LNP model has three stages:

**Stage 1** describes how the neuron integrates stimulus intensity over space and time. The stimulus is denoted by S(x, y, t) where  $(x, y) \in \{0 \dots M, 0 \dots N\}$  are the space coordinates in

<sup>&</sup>lt;sup>1</sup>A python script to generate the SWN is available at: https://team.inria.fr/biovision/swn/



Figure 2: Power spectra of BWN and SWN under various conditions. For visualization purposes only the Power Spectra profile on the x-axis is showed. Both block and shift sizes are in pixels. (A) BWN with various blocks sizes. Only with block size 1 pixel the power spectrum is white. The power spectrum bandwidth increases with the inverse of the block size. (B) left SWN of various block sizes and fixed shift 5 pixels. In this case the power spectrum is very similar to the BWN's. (B) right SWN of various sift sizes and fixed block of 20 pixels. Note that if block and shift sizes are equal, then the SWN is resumed to the BWN.

 $\mu m^2$  and  $t \in \{0, 1, 2, ...\}$  is the time in ms. Space was discretized in bins of  $1 \mu m^2$ , time was discretized in bins of 1 ms.

The spatio-temporal kernel of the neuron is denoted by K(x, y, t) where  $(x, y) \in \{0 \dots M, 0 \dots N\}$ and  $t \in \{0, \dots, T\}$  (i.e., T is its temporal support) The resulting integration denoted by L(t) is defined by an inner product in space and a convolution in time:

$$L(t) = \sum_{\tau=0}^{T} \sum_{x=0}^{N} \sum_{y=0}^{M} K(x, y, \tau) S(x, y, t - \tau).$$
(1)

Then we assume that the kernel K is separable is space and time, i.e.:

$$K(x, y, t) = K_S(x, y)K_T(t),$$
(2)

where each part of the kernel is defined according to classical models of retinal processing [6, 25], namely difference of Gaussian (DOG) for the spatial part and a polynomial multiplied by a decaying exponential for the temporal part:

$$K_{S}(x,y) = 16 \frac{1}{2\pi\sigma_{c}^{2}} \exp\left(-\frac{1}{2\sigma_{c}^{2}}\left((x-c_{x})^{2}+(y-c_{y})^{2}\right)\right)$$
(3)  
$$-8 \frac{1}{2\pi\sigma_{s}^{2}} \exp\left(-\frac{1}{2\sigma_{s}^{2}}\left((x-c_{x})^{2}+(y-c_{y})^{2}\right)\right),$$
  
$$K_{T}(t) = \left(-\frac{(0.7t)^{7}}{7!} + \frac{(0.7t)^{5}}{5!}\right) \exp\left(-0.7t\right),$$
(4)

where parameters  $\sigma_c, \sigma_s$  define the spatial integration properties of the neuron (for center and surround) and  $(c_x, c_y)$  is the position of its center. These are the parameters that we vary to define the population.

**Stage 2** gives the instantaneous spike rate  $\lambda(t)$  in spikes per bin by passing the output of the first stage by a non linearity:

$$\lambda(t) = f(L(t)), \text{ where } f(L) = \frac{1}{1 + \exp\left(-0.05L - 100\right)}.$$
(5)

**Stage 3** converts the spike rate into a series of spikes using an inhomogeneous Poisson process. Since time was discretized in 1 ms bins, the Poisson process is approximated by a Bernoulli process in each bin.

#### Synthetic data: Neural population construction

Given the neuron's model described above, our goal was to define an heterogeneous population of such neurons that will cover the different possible experimental scenarios but, for the sake of simplicity, we do not consider orientation or direction selective cells. Heterogeneity stems from the parameters' choice in the spatial part of kernel K (see (3)), namely the center of the RF,  $(c_x, c_y)$ , and the size of the central Gaussian,  $\sigma_c$ .

The generation process is illustrated in Fig. 3. First, we choose as a reference grid the grid provided by the BWNs of low resolution, i.e., with blocks of size 32 µm. The idea is to define a population of neurons where we vary positions and kernel sizes. More precisely, we define a population of 216 neurons as follows:



Figure 3: Stages to build the artificial RGC population. (A) We defined nine positions equally distributed between the center of the central block  $(c_0)$  and the center of the adjacent block in the diagonal direction  $(c_M)$ . (B) For the spatial kernel, we defined 24 possible values of  $\sigma_c$ , so that the positive part radius vary from 0.5 pixel to 1.5 blocks in steps of 0.5 pixel. The smallest RFs have a radius of  $2 \,\mu$ m and the largest  $24 \,\mu$ m. (C) We assumed here that the surround variance is three times higher than center variance.

• We first define a set of positions to evaluate the consequences of the alignment of the RF with the block center. Starting from position  $c_0 = (0,0)$  (at the center of the spatial domain), we define a family of neurons equally sampled along the diagonal direction in steps of one pixel ( $\delta_c = 4 \,\mu$ m) until the next block center  $c_{max}$  (see Fig. 3(A)). Doing so, we define nine positions:

$$(c_x, c_y) = \{(0, 0), (\delta_c, \delta_c), \dots, (8\delta_c, 8\delta_c)\} = \{(0, 0), (4, 4), \dots, (32, 32)\}.$$

Note that one parameter is sufficient to describe the neuron's position, namely we use  $c_x$  in the remainder of this paper.

• Then, for each neuron position, we defined a family of neurons with varying spatial kernel sizes (see Eq. (3)). The smallest radius  $r_0$  corresponds to a center standard deviation  $\delta_{\sigma} = 0.784 \,\mu\text{m}$ . In this way, the size of the center Gaussian (the circumference radius where the DOG changes from positive to negative) is 2 µm. The remaining  $\sigma_c$  were defined as a multiple of this one, increasing in steps of 2 µm (meaning, half pixel) the center radius (see Fig. 3(B)). Doing so, we defined 24 possible values of  $\sigma_c$ :

$$\sigma_c = \{\delta_{\sigma}, 2\delta_{\sigma}, \dots, 24\delta_{\sigma}\} = \{0.784, 1.568, \dots, 18.824\}.$$

Concerning the surround standard deviation,  $\sigma_s$ , it was set to three times  $\sigma_c$  as usually fixed in the literature (see Fig. 3(C)). Thus only one parameter is sufficient to describe the spatial kernel amplitude, i.e.,  $\sigma_c$ .

#### Synthetic data: STA

The STA is a reverse-correlation technique commonly used to estimate the RF of neurons that relies on the assumption that the neuron has a linear receptive field and the stimulus has no spatio-temporal correlations [5, 26, 21, 10]. The STA corresponds to the average sequence of images preceding spikes. It is defined as follows. Consider a neuron (model or experimental) that spiked at times  $t_1, t_2, \ldots, t_n$  when stimulated by a spatio-temporal stimulus S(x, y, t), then the STA of this neuron, denoted by  $A(x, y, \tau)$ , is given by:

$$A(x, y, \tau) = \frac{1}{n} \sum_{i=1}^{n} S(x, y, t_i - \tau)$$
(6)

with x and y belonging to the same spatial domain as the stimulus S and  $\tau$  is on  $\{-\mathcal{T}...0\}$ where  $-\mathcal{T}$  defines the temporal support of the STA.

STA allows a parameter free estimation of the RF, it is easy to design and use experimentally in any biological sensory modality. To estimate the STA, we use the PRANAS, an open and free platform for retinal analysis and simulation [4]. Independently of the stimulus resolution, STAs were estimated at the highest resolution possible, meaning the same resolution as the ground truth (GT) RFs.

#### Synthetic data: Evaluation

Considering the description above, a synthetic RF is 3D tensor corresponding to the space-time dimensions. To extract the spatial component, we consider the 2D slice taken at the time when the absolute RF is maximal. Similarly, a neuron STA is a 3D tensor and we extract the spatial component in the same way. The performance evaluation was computed always on the RF's and STA's spatial components. We propose three criteria to assess the validity of the SWN.

**Criterion 1 (Number of mapped RFs)** We consider that a RF was mapped if the STA absolute peak has strong deviations from the spatial profile average value. Assuming that the values are Gaussian distributed, we assess this criterion automatically, with a one sample Z-test with a maximum p-value, meaning, significance level of 10e-8 on the spatial profile's peak of each STA. If the peak of the 2D-spatial profile is accepted as the expected value of the population, then it is not statistically different from the population, thus it is Gaussian noise and the RF was not mapped. But otherwise, if the peak is rejected as the expected value of the population, then it is statistically different from the population, thus it is not noise, which means that the RF was mapped. Note that, this method allows to consistently evaluate all the STAs depending only on peak amplitude with respect to noise. Despite of its simplicity, we verified that this test is accurate by visual inspection. Please see the p-values below the spatial STAs of Fig. 4 and 6 and some extra examples on the supplementary material.

**Criterion 2 (RF parametric description)** For a mapped RF, we fit the spatial RF with a DOG as defined in eqn. (3). For that, we used the Trust Region Reflective method [19, 17], which is a bounded minimization algorithm. In practice, in order to balance between parameters variability and algorithm efficiency, we defined large bounds for each parameter:  $\sigma_c$  lower value is 0.1 and higher value is three times the image size in µm;  $c_x$  and  $c_y$  lower value is equal to 1µm and higher value the image length. To avoid local minima each STA was fitted 12 times with different initializations uniformly sampled within the bounds. Then, for the analysis, we selected the parameters that minimize the fitting error.

**Criterion 3 (STA error)** In the synthetic case, we can compare the STAs with the GTs pixelby-pixel. To do so, we measure the angle between the two vectors using the cosine similarity as suggested in [21]. If we denote by A the output of the STA, this angle is given by:

$$E(\bar{K}, A) = \cos^{-1} \frac{\langle \bar{K}, A \rangle}{\|\bar{K}\| \|A\|},$$
(7)

where  $\bar{K}$  is the true neuron's kernel defined by (2), and  $\|.\|$  the usual euclidean norm.

#### Experimental data: Stimuli

We have used four stimuli to map RGC RFs in mouse retinal wholemounts: two BWNs of low and high resolution (160  $\mu$ m and 40  $\mu$ m, respectively) and two SWNs with STA of high and super high resolution (for both stimuli blocks of 160  $\mu$ m and shifts of 40  $\mu$ m and 4  $\mu$ m, respectively).

We generated 60,000 images of each stimulus grouped in 20 blocks of 3000. These blocks were randomly sorted before projecting onto the retina to avoid response bleaching bias for certain stimulus conditions. Each image was  $664 \times 664$  pixels, where one pixel corresponds to a square of size  $4 \,\mu\text{m}^2$ . Images were refreshed at 30.3Hz (presented during 33 ms).

Light stimuli were projected onto the retina as described previously in [24] and attenuated using neutral density filters to high mesopic light levels (mean luminance 11 cd/m2).

#### Experimental data: MEA recordings

All experimental procedures were approved by the ethics committee at Newcastle University and carried out in accordance with the guidelines of the UK Home Office, under control of the Animals (Scientific Procedures) Act 1986. Experiments were performed as described previously in [12].

Briefly, a female mouse (aged 42 days) was dark-adapted overnight and killed by cervical dislocation. Eyes were enucleated, and following removal of the cornea, lens, and vitreous body, they were placed in artificial cerebrospinal fluid (aCSF) containing the following (in mM): 118 NaCl, 25 NaHCO<sub>3</sub>, 1 NaH<sub>2</sub>PO<sub>4</sub>, 3 KCl, 1 MgCl<sub>2</sub>, 2 CaCl<sub>2</sub>, 10 glucose, and 0.5 l-Glutamine, equilibrated with 95% O<sub>2</sub> and 5% CO<sub>2</sub>. The retina was isolated from the eye cup and flattened for MEA recordings. All procedures were performed in dim red light, and the room was maintained in darkness throughout the experiment. Retinal recordings were performed on the BioCam4096 platform with BioChips 4096S+ (3Brain GmbH, Lanquart, Switzerland), integrating 4096 square microelectrodes (21 × 21µm, pitch 42 µm) on an active area of  $2.67 \times 2.67$ mm.

The spatial extent  $(7.12 \text{ mm}^2)$  of the MEA chip allowed us to record simultaneously from large retinal areas (see Fig. 7). The small electrode pitch  $(42 \,\mu\text{m})$  enables sampling from many individual RGCs from these areas, providing us with an unbiased very large analytical sample size. After recording, spikes were sorted and the activity raster plots were generated. Single-unit spikes were sorted using the T-Distribution Expectation-Maximisation algorithm in Offline Sorter (Plexon Inc, Dallas, USA).

#### Experimental data: STA

Before computing the STAs, the stimulus images (independently of the stimulus) were cropped at  $(640 \times 640)$  pixels to remove the partial blocks and resized to the smallest size possible without compromising the results or, in other words, to reduce to the highest resolution possible without loss of information. Precisely, the images of BWN-B160 were reduced to  $(16 \times 16)$  pixels, BWN-B40 and SWN-B160-S40 were reduced to  $(64 \times 64)$  pixels, while SWN-B160-S4 was not resized. This allowed to reduce the population STA computation time from days to several minutes (in BWN-B160) or hours (BWN-B40 and SWN-B160-S40). The STA estimation was performed as for the synthetic data. To reduce the image size before the STA analysis to the highest possible resolution without loss of information is a common trick on spiking data analysis. For comparison, in [1] only 90 stimulus dimensions were used (3 × 3 on space and 10 on time). If we do not use this trick, the stimulus dimensions in our case would be 4408960. In addition we did a pan-retinal study with 4798 responsive neurons and tested 4 stimuli. Using this trick allowed us to reduce the stimulus dimension to 2560 in the BWN-B160, to 40960 in the BWN-B40 and SWN-B160-S40, and there was no reduction in the SWN-B160-S4.

#### **Experimental data: Evaluation**

All responsive cells were considered for evaluation. The evaluation was performed similarly to the synthetic data. First the STA was estimated and analyzed using Criterion 1 (Number of mapped RFs). For the neurons whose RF was successfully mapped, Criterion 2 (RF parametric description) was considered. However, here only the center Gaussian was considered, as is usually done when analyzing mouse retinal RFs [12]. Criterion 3 could not be used since GT was not available.

## Results

#### Synthetic data: Single neuron level

Here we focus on only one neuron from the entire population. This neuron is located in  $(c_x, c_y) = (4\delta_c, 4\delta_c)$  which is at the intersection of blocks, and with a central variance kernel  $\sigma_c = 24\delta_{\sigma}$ . Note that this was chosen to have the most favorable situation for BWN-B32 (as shown later in Fig. 5).

For each of the three stimulus types, we used ten instances of each stimulus to generate rasters, 30 rasters in total. Each instance of the stimulus represents 20,000 images, which generated rasters of 11 minutes long.

Neural responses for each stimulus are given in Tab. 1. As expected, stimuli with larger blocks (BWN-B32 and SWN-B32-S4) induced stronger responses. In addition, note that SWN-B32-S4 generated the strongest response. This fact is relevant because, as shown in [22], the STA error decreases as a function of the number of spikes.

Stimulus	# spikes	spike rate
BWN-B32	9108	13.8 Hz
BWN-B4	6204	$9.4 \mathrm{Hz}$
SWN-B32-S4	9438	14.3 Hz

Table 1: Average neuron response to each stimuli across trials, with the number of spikes and the corresponding spike rate. Note that, for all the stimuli, the spike rate is high because we do not considered a refractory period in the neuron model. The spike rate using SWN-B32-S4 is 1.5 times higher than using BWN-B4

Spatio-temporal kernels estimated with each stimulus are show in Fig. 4(A). These results can be compared qualitatively with the ground truth, here represented with a spatial discretization of 4 µm. With BWN-B32, the result lacks precision in space. It has strong vertical and horizontal edges corresponding to the stimulus block size. The temporal part, however, is properly estimated. With BWN-B4, when the block size is smaller, the results are very noisy in both space and time, and no relevant information can be detected. With SWN-B32-S4, the spatial aspect has the same resolution as the ground truth, and this method gives good results in terms of precision too, with good estimate of the RF shape, even if some weak, noisy patterns remain in the periphery. The temporal part is accurate as well, as with BWN-B32. These observations suggest that SWN allows increasing the spatial part of the kernel's spatial resolution with the same stimulation time.



Figure 4: Single neuron analysis. (A) Comparison of spatio-temporal RF estimations. (2D) Spatial slice taken at the point with maximal amplitude of the STA. (1D) Horizontal cut from 2D slice passing through the point with maximal amplitude. (Temporal) Temporal cut from the spatial-temporal STA passing through the point with maximal amplitude. Below each STA's spatial slice, we provide the p-value on the mapping test (Criterion 1). Green font (resp. red front) is used to indicate when RF is mapped (resp. not mapped). A RF is mapped when p-value is less than 10e-8. The SWN allows for increasing the spatial part quality while preserving the quality of the temporal profile. (B) Error as a function of time between the RF estimate and ground truth. An estimate is done at each minute (circles, for each trial) and we fit this error with a power law (continuous line). The SWN always performs better than BWN, and the convergence rate is faster. (C) Proportion of mapped RFs at each minute. It is noticeable that very early, from the first minute, 100% of RF can be mapped with SWN. To reach 100%, one must wait seven minutes with BWN-B32. For BWN-B4, convergence is much slower so that no RF can be fit before 11 minutes.

In Fig. 4(B) we show how the error (eqn. 7) evolves in time for each condition (Criterion 3). Estimates are done at every minute of stimulation time, i.e., at time t, STA is estimated based upon spikes recorded in the time window [0, t]. The SWN error is always the smallest followed by the BWN-B32 and then by BWN-B4. Note that, even when using the BWN-B32 for an extremely long time, the STA error will not converge to 0, due to the different resolutions between the STA and the ground truth. in Finally, falling back on Criterion 1, Fig. 4(C) shows how the proportion of trials whose RF was mapped evolves in time for each condition. The STA estimates are the same as in Fig. 4(B). With the BWN-B4 the RF was never mapped. In contrast, the proportion of times that the RF was mapped with BWN-B32 increases with the stimulation time to reach 100% at seven minutes. With SWN-B32-S4, this number rapidly grows to reach 100% at one minute already. In other words, successful RF mapping of single neurons was achieved 10 times faster with the SWN than with BWN. In other words, as the number of spikes increases, the p-value decreases and more neurons are mapped. However, for the same roughly the same number of spikes, the p-value using the SWN is lower than using the BWN. More details follow Fig.S1 of the supplementary material ( https://doi.org/10.6084/m9.figshare.16867135).

Taken together, these observations on a single neuron suggest that our approach allows better quality RF estimates and that these estimates need less stimulation time. In the next section, we consider an entire population of neurons to verify whether these observations hold in more generally.

#### Synthetic data: Population level

For the population of 216 neurons defined in the methods Section, the three stimuli were presented for 11 minutes. Note that since inter-trial variability is low (see Fig. 4(B)), we use only one trial in this section.

In Fig. 5(A), we show the error between STA and the ground truth (Criterion 3), after 11 minutes over the whole population. This compact representation gives very instructive insights into the precision reached by each stimulus and how such performance depends on the neurons' characteristics. First, globally, the average error over the entire population is 55.6, 83.6, and 48.7 degrees for BWN-B32, BWN-B4, and SWN-B32-S4, respectively. In other words, for the same resolution,  $4\,\mu\text{m}$ , the average error using SWN is 1.7 times smaller than using BWN. Then, when delving into more details, starting with BWN-B4, we observe that strong errors are made for most neurons in the population, due to the slow rate of convergence with this stimulus, as observed in the single neuron case. We note a minor exception for the smallest RF, which seems to be better-captured when using small block sizes. Considering larger block size, namely BWN-B32. the error decreases as the RFs become larger, which reflects matching between block and RF sizes. Another effect emerges, namely some level of dependence on the neurons' position relative to the block. When the position of a neurons is "in-between" blocks, errors become larger. Which is presumably related to the fact that neurons in such positions experience less spatial average variations. This effect completely disappears with SWN-B32-S4. Indeed, although errors keep decreasing with increasing RF sizes, the neuron's position does not affect the accuracy of the responses anymore, which offers great advantages when dealing with experimental data.

From the 216 neurons, applying the Criterion 1 at the end of stimulation, all neurons were mapped with both BWN-B32 and SWN-B32-S4, but only 90 neurons were mapped with BWN-B4. In other words, for the same resolution, the SWN mapped 2.3 times more neurons than the BWN.

In Figs. 5(B)-(D), we compare the parameters of the fitted kernel with the original ones (Criterion 2), illustrating kernel parameters at 5 (left) and 11 minutes (right) respectively. These panels complement Fig. 5(A) in two ways: (1) they provide an interpretation of the nature of



Figure 5: Population analysis. (A) Error between the RF estimate and ground truth, as a function of position and size. (B)–(D) Comparing the fitted kernel parameters with the original ones. (B) Estimated sizes against the ground truth sizes for all RFs. On the left at five min of stimulation, on the right at 11 min of stimulation. (C) Estimated sizes in function of the RF position for the neurons with  $\sigma_c = 17.25$ . On the left at five min of stimulation, on the right at 11 min of stimulation. (D) Top panels shows estimated center positions against the ground truth positions for all RFs. The bottom panels shows, the same as top but for neurons with  $c_x = 34$ . Results show that SWN-B32-S4 offers the best performance at the population level, with no dependence on the neurons' position.

the error, and (2) they give an idea of the evolution in time of this error. Both BWN-B32 and SWN-B32-S4 result in biased fittings, towards a larger center size than the ground truth's. For BWN-B32, this bias essentially remains over time, while for SWN-B32-S4, it decreases for many of the larger RFs(Figs. 4(B) and 4(C)). Furthermore, BWN-B32 bias depends on the RF position, while with the increase of the raster length the SWN-B32-S4 bias becomes independent of the position (Fig. 5(C)). Concerning BWN-B4, there is no bias, at least for the very small RFs that could be mapped. At the same time, no medium and large RFs could be mapped with this stimulus. The classical STA analysis with (Gaussian) white noise has the important property that it yields a consistent and unbiased estimator for the RF (when defined as the linear filter, e.g., in the context of a linear-nonlinear model; see, e.g., [21]). In contrast, the SWN method shows spatial correlations (see Fig. 2) larger than the final resolution of the analysis, which introduces a bias towards overestimating receptive field sizes, even in the limit of infinite data. We indeed see, in Fig. 5 (C), that the estimated  $\sigma_c$  is above the ground truth and is independent of  $c_x$  (for 11 min of stimulation). In contrast the BWN-B32 shows a clear dependence on  $c_x$ . It gives a better estimation for small or large  $c_x$  and a worse estimation for  $c_x \sim 16 \mu m$ . Taken together (i.e. taking the average over  $c_x$  SWN gives a better estimation but one has to keep this overestimation effect in mind when using SWN.

On the contrary, the RFs' center estimation is unbiased with the high resolution stimuli, BWN-B4 and SWN-B32-S4, as shows Fig. 5(D). As the RF' center approximates the block border, meaning as  $c_x$  approximates 16 µm, the center estimation becomes more biased, especially for the smaller RFs. In addition this bias is systematically towards the block center. Further analysis on the estimated RFs extend, particularly the spatial autocorrelation, are available Fig.S2 of the supplementary material (see https://doi.org/10.6084/m9.figshare.16867135).

#### Experimental data: Single neuron level

In Fig. 6 we show four representative cases of estimated RFs. For these four neurons, we observe different situations concerning the number of mapped RFs measured with Criterion 1. In all cases selected, SWN-B160-S4 was mapped. No striking difference appears for SWN stimuli with shifts of  $40 \,\mu\text{m}$  and  $4 \,\mu\text{m}$  (when both were mapped). Results with BWN-B40 are always noisy, even when the RF is mapped. We also found several analogies with the synthetic data. RFs estimated with the SWN stimuli were smoother than RFs estimated with the BWN. The RF temporal profile, on the other hand, was not altered by the shifting process.

#### **Experimental data: Neural Population**

4978 Neurons spiked at least one time for at least one stimulus condition. Fig. 7(A) illustrates the log spiking activity of the retina recorded from the RGC layer. Responses to light occurred across the entire active area of the MEA, with particular emphasis on the dorsal-lateral axis. As Tab. 2 shows, the number of spikes was similar for the stimulus of large blocks - BWN-B160, SWN-B160-S40 and SWN-B160-S4 - independently of shifting or not.

Stimulus	# spikes	spike rate
BWN-B160	2683	$1.4 \mathrm{Hz}$
BWN-B40	2070	1.0 Hz
SWN-B160-S40	2690	$1.4 \mathrm{Hz}$
SWN-B160-S4	2774	$1.4 \mathrm{Hz}$

Table 2: Average neuron response to each stimuli, with the number of spikes and the corresponding spike rate.



Figure 6: STAs representatives examples for foor neurons, showing different situations in terms of RF mapping depending on the stimulus. More examples are available in the supplementary material (see Fig.S3 of https://doi.org/10.6084/m9.figshare.16867135). The representation is the same as in Fig. 4. In general, SWN yields smoother spatial STAs than the BWN, without modifying the temporal STA.



Figure 7: Retinal activity pan-retinal view. (A) The Log spike count during the entire experiment for each retinal channel. This results in a visualisation of the retina outline and gives an overall estimation of the number of active channels. (B) Number of RFs distributed over the MEA. Retina orientated identically to panel (A). For visualization purposes the values of all sub-panels were smoothed with a Gaussian kernel. RFs' positions are in agreement with the activity map. In general, SWN-B160-S4 centers were more uniformly distributed than with the other stimuli, covering almost the entire area of stimulation.

Fig. 7(B) shows the distribution of the centers of all mapped RFs depending on the stimulus type. As for the synthetic neurons, an experimental neuron was considered mapped if it passed Criterion 1. This plot was achieved by fitting STAs with DOGs to find their center position and size. Overall, the distribution of the STA's center was similar to the activity map for BWN-B160 and both SWN. With the SWN-B160-S4, we could map neurons on the nasal-dorsal direction, which was not possible with the remaining stimuli, thus centers were more uniformly distributed with the super high resolution SWN.

Fig. 8(A)–(B) show STA convergence properties with respect to the four stimuli. Basic count of the number of RFs mapped. Fig. 8(A) reveals that the SWNs method outperforms the BWNs method in all conditions. Interestingly, increasing the resolution leads to different behaviors. Considering the BWN, to increase the resolution from 160 to  $40 \,\mu\text{m}$  yielded a heavy reduction of mapped RFs. In contrast, considering the SWN, to increase the resolution from 40 to  $4 \,\mu\text{m}$  yielded an increased incidence of mapped RFs. When the final STA resolution is fixed to  $40 \,\mu\text{m}$  and we use the SWN, we can successfully map 21 times more numerous RFs than when using the BWN. Similarly to the synthetic data, the SWN lead to larger spike rate which resulted in lower p-values. For more details see Fig.S4 of the supplementary material (https://doi.org/10.6084/m9.figshare.16867135).

Going a step further, Fig. 8(B) shows how many cells were found for each stimulus. This is represented as an Euler diagram. Except for a few cells, we observe the following pattern of



Figure 8: Mapped RFs statistics. (A) Number of mapped RFs per stimulus. Both SWNs outperform the BWNs, for the same resolution, 40 µm, SWN mapped 21 times more RFs then BWN. (B) Euler diagram (in percentage) of mapped RFs. Cell percentages below 3% are not shown. All RFs mapped with BWN were also mapped with the SWN. (C) Distribution of RFs sizes per stimulus. Values below 40 µm are not shown due to their biological implausibility. The SWN mapped a broader range of sizes, specially when considering the very high resolution stimuli, SWN-B160-S4.

$\operatorname{stimulus}$	RF size
BWN-B160	$102.9\pm21.7$
BWN-B40	$71.7 \pm 11.8$
SWN-B160-S40	$105.3 \pm 19.5$
SWN-B160-S4	$91.5\pm25.4$

Table 3: Mean and standard deviation statistics of RFs sizes per stimulus

inclusions:

#### BWN-B40 $\subset$ BWN-B160 $\subset$ SWN-B160-S40 $\subset$ SWN-B160-S4.

Note that a small percentage of RFs' was mapped with SWN-B160-S40 and not with SWN-B160-S4, but this number is four times smaller than the reverse situation.

In Fig. 8(C) we compare the estimated center sizes for the mapped RFs. RFs with radius smaller than 40 µm or larger than 180 µm were considered outliers as their size is not biologically plausible (see, for instance, [12]). Outliers are not shown in this Fig. The number of outliers depended on the resolution. While high-resolution stimuli lead to a relatively small number of outliers: 5, 7 and 6 for BWN-B40, SWN-B160-S40 and SWN-B160-S4, respectively. In contrast, using the low resolution BWN-B160 stimulus yielded to 287 outliers. As in the synthetic case, the BWN of high resolution was pruned for small centre sizes, while the BWN of low resolution and the SWN-B160-S40 stimuli were pruned to larger radii. Notably, the SWN-B160-S4 was not pruned to a specific range. Nevertheless, the shape of the radii distribution depended on the stimulus as well. The spread around the preferred value was low in the BWN-B40, medium in the BWN-B160 and SWN-B160-S40 cases and large in the SWN-B160-S4 case. These results are quantified in Tab. 3. BWN-B40 yielded the smallest RF size values (mean and standard deviation), while values are equally large when using BWN-B160 and SWN-B160-S40. Using SWN-B160-S4 yields the largest standard deviation. As for the synthetic data these differences on the fitting sizes distribution are reflected on the STAs' spatial autocorrelation. More details on the supplementary material (see Fig.S5 of https://doi.org/10.6084/m9.figshare.16867135).

## Discussion and conclusion

In summary, here we show that a SWN stimulus considerably improves RF estimation for RGCs. Using synthetic data, we demonstrate that with SWN, RF estimation is independent of the position of individual neurons relative to the stimulus, which is not the case for BWN. The resolution is always higher with SWN, since there is no compromise between responsiveness (given by the block size) and resolution, and the latter increases by reducing the stimulus baseline shift. At the population level, not only do we achieve higher mapping resolution, but we also map more RFs, with more neuronal variability.

Because neurons exhibit stronger activity when presented with larger block sizes, the use of SWN stimuli also leads to faster RF mapping, which makes the STA approach more efficient. In the case of synthetic data, we showed that the whole population can be mapped seven times faster with SWN. This is important when dealing with experimental data because mapping RFs with STA is often just one preliminary step in a much longer experimental pipeline (using various stimuli), which often leads to bleaching of light responses (see, e.g., [24]).

SWN has spatial correlations larger than the final resolution of the analysis, which introduces a bias towards overestimating RF sizes. This overestimation partially decreases over time, but it will be present even in the limit of infinite data. A possible solution is to apply a deconvolution approach as used for other non-white noise stimuli [28, 26]. If using such methods, one must be careful with the deconvolution process, as it involves the correlation matrix inversion, and SWN correlations decrease quickly to zero (this can be seen because the power spectrum bandwidth is small). As suggested in [28], to prevent numerical errors, in this case, it is necessary to determine the subset of eigenvectors of the stimulus autocorrelation matrix that have significant non-zero eigenvalues.

This size overestimation seems to decrease with stimulation time for larger RFs, but theoretically it is known that it will be always present due to the stimulus correlations. Due to the same correlations, similar bias is present in the low resolution BWN as well, but without changing significantly with time. In addition, the SWN bias is invariant to the RF position, while the BWN is larger and depends on the RF distance to the block center. Computationally, we also found that the STAs mapped with high resolution stimuli, with BWN or SWN, are aligned with the RFs centers'. In contrast, low resolution BWN shows strong biases towards the block centers'.

Visually, there were no striking differences between the RFs mapped with the SWN-B160-S40 and SWN-B160-S4. However, their fitting results were not similar. With SWN-B160-S4 a broader distribution of sizes were mapped than with SWN-B160-S40. SWN-B160-S4 mapped both small and large RFs. Remarkably, to map both small and large RFs was not possible with any other stimuli, thus with the one pixel resolution SWN we mapped the larger amount of RFs and without the sizes being biased towards a specific range. In addition, the SWN-B160-S4 centers were more uniformly distributed, covering almost the entire area of stimulation.

Another advantage of SWN is that the new stimuli are also easy to produce and the same reverse correlation methods can be used to recover the RF. Of course, in practice, one still needs to choose a suitable block size. This choice still relies on the experimenter expertise, but with SWN, it is less critical since the variability introduced by the shift will compensate for a sub-optimal block size value. A possible strategy is to use a block size smaller than the expected RF field size or the dendritic field width of the measured cell types with a shift about 1/5 of the block size. Nevertheless, this strategy must be adapted to the experimental context such as animal species, cell type, recording conditions and the study goal.

The general approach applied to STA in this paper — making use of super-resolution methods to boost the performance of RF estimation methods — will allow for more efficient stimuli design in sensory physiology. We also expect that this general approach could be applicable to other spike-triggered methods like the Spike Triggered Covariance [21, 27], since it is a good approximation of Gaussian White Noise, and easy to implement. However, further computational studies must be performed, as Spike Triggered Covariance is typically more susceptible to deviations from Gaussian white noise structure of the stimulus. Furthermore, this super-resolution idea might also be useful on other sensory modalities where the STA has been shown to be interesting.

# References

- 1. Ahn, J., Rueckauer, B., Yoo, Y., Goo, Y.S.: New features of receptive fields in mouse retina through spike-triggered covariance. Experimental neurobiology **29**(1), 38 (2020)
- Atencio, C.A., Sharpee, T.O., Schreiner, C.E.: Cooperative nonlinearities in auditory cortical neurons. Neuron 58(6), 956–966 (2008)
- Berdondini, L., Imfeld, K., Maccione, A., Tedesco, M., Neukom, S., Koudelka-Hep, M., Martinoia, S.: Active pixel sensor array for high spatio-temporal resolution electrophysiological recordings from single cell to large scale neuronal networks. Lab on a Chip 9(18), 2644–2651 (2009)

- 4. Cessac, B., Kornprobst, P., Kraria, S., Nasser, H., Pamplona, D., Portelli, G., Viéville, T.: Pranas: A new platform for retinal analysis and simulation. Frontiers in Neuroinformatics 11, 49 (2017). DOI 10.3389/fninf.2017.00049. URL http://journal.frontiersin.org/article/10.3389/fninf.2017.00049
- Chichilnisky, E.J.: A simple white noise analysis of neuronal light responses. Network: Comput. Neural Syst. 12, 199–213 (2001)
- Croner, L.J., Kaplan, E.: Receptive fields of P and M ganglion cells across the primate retina. Vision research 35(1), 7–24 (1995)
- 7. Dayan, P., Abbott, L.F.: Theoretical neuroscience: computational and mathematical modeling of neural systems. Computational Neuroscience Series (2001)
- DeAngelis, G.C., Ohzawa, I., Freeman, R.D.: Spatiotemporal organization of simple-cell receptive fields in the cat's striate cortex. i. general characteristics and postnatal development. Journal of neurophysiology 69(4), 1091–1117 (1993)
- Földiák, P.: Stimulus optimisation in primary visual cortex. Neurocomputing 38-40, 1217– 1222 (2001)
- 10. Gollisch, T.: Estimating receptive fields in the presence of spike-time jitter. Network: Computation in Neural Systems 17(2), 103–129 (2006)
- Greenspan, H.: Super-resolution in medical imaging. Computer Journal 52(1), 43–63 (2009). DOI 10.1093/comjnl/bxm075
- Hilgen, G., Pirmoradian, S., Pamplona, D., Kornprobst, P., Cessac, B., Hennig, M.H., Sernagor, E.: Pan-retinal characterisation of light responses from ganglion cells in the developing mouse retina. Scientific reports 7(1), 1–14 (2017)
- 13. Jones, J.P., Palmer, L.A.: The two-dimensional spatial structure of simple receptive fields in cat striate cortex. Journal of Neurophysiology **58**(6), 1187–1211 (1987)
- Liu, C., Sun, D.: A bayesian approach to adaptive video super resolution. In: IEEE Computer Society Conference on Computer Vision and Pattern Recognition (CVPR), pp. 209–216 (2011)
- 15. Machens, C.K.: Adaptive sampling by information maximization. Physical review letters **88**(2), 228104 (2002)
- MacKay, D.J.C.: Information-based objective functions for active data selection. Neural Computation 4, 590–604 (1992)
- The Mathworks, Inc., Natick, Massachusetts: MATLAB version 9.3.0.713579 (R2017b) (2017)
- Merino, M.T., Nunez, J.: Super-resolution of remotely sensed images with variable-pixel linear reconstruction. IEEE Transactions on Geoscience and Remote Sensing 45(5), 1446– 1457 (2007)
- Moré, J.J., Sorensen, D.C.: Computing a trust region step. SIAM Journal on Scientific and Statistical Computing 4(3), 553–572 (1983)

- 20. Odermatt, B., Nikolaev, A., Lagnado, L.: Encoding of luminance and contrast by linear and nonlinear synapses in the retina. Neuron **73**(4), 758–773 (2012)
- Paninski, L.: Convergence properties of three spike-triggered analysis techniques. Network 14, 437–464 (2003)
- Paninski, L.: Maximum likelihood estimation of cascade point-process neural encoding models. Network: Comput. Neural Syst. 15(04), 243–262 (2004)
- Peeters, R.R., Kornprobst, P., Nikolova, M., Sunaert, S., Vieville, T., Malandain, G., Deriche, R., Faugeras, O., Ng, M., Van Hecke, P.: The use of super-resolution techniques to reduce slice thickness in functional mri. International Journal of Imaging Systems and Technology 14(3), 131–138 (2004)
- Portelli, G., Barrett, J.M., Hilgen, G., Masquelier, T., Maccione, A., Di Marco, S., Berdondini, L., Kornprobst, P., Sernagor, E.: Rank order coding: a retinal information decoding strategy revealed by large-scale multielectrode array retinal recordings. eNeuro 3(3) (2016)
- Rodieck, R.W.: Quantitive analysis of cat retinal ganglion cell response to visual stimuli. Vision Research 5(12), 583–601 (1965)
- Schwartz, O., Pillow, J.W., Rust, N.C., Simoncelli, E.P.: Spike-triggered neural characterization. Journal of vision 6, 484–507 (2006)
- Simoncelli, E.P., Paninski, J.P., Pillow, J., Schwartz, O.: Characterization of neural responses with stochastic stimuli. The cognitive neurosciences, 3rd edition pp. 327–338 (2004)
- Theunissen, F.E., David, S.V., Singh, N.C., Hsu, A., Vinje, W.E., Gallant, J.L.: Estimating spatio-temporal receptive fields of auditory and visual neurons from their responses to natural stimuli. Network: Computation in Neural Systems 12(3), 289–316 (2001)
- 29. Wang, H., Wen, D.: The progress of sub-pixel imaging methods. In: J. Ojeda-Castaneda, S. Han, P. Jia, J. Fang, D. Fan, L. Qian, Y. Gu, X. Yan (eds.) Selected Papers from Conferences of the Photoelectronic Technology Committee of the Chinese Society of Astronautics: Optical Imaging, Remote Sensing, and Laser-Matter Interaction 2013, vol. 9142, pp. 166–170. International Society for Optics and Photonics, SPIE (2014)
- Yue, L., Shen, H., Li, J., Yuan, Q., Zhang, H., Zhang, L.: Image super-resolution: The techniques, applications, and future. Signal Processing 128, 389–408 (2016)