brought to you by I CORE



MINIREVIEW

Reactive oxygen species, ageing and the hormesis police

Paula Ludovico^{1,2} & William C. Burhans³

¹Life and Health Sciences Research Institute (ICVS), School of Health Sciences, University of Minho, Braga, Portugal; ²ICVS/3B's – PT Government Associate Laboratory, Braga/Guimarães, Portugal; and ³Department of Molecular and Cellular Biology, Roswell Park Cancer Institute, Buffalo,

Correspondence: Paula Ludovico, School of Health Sciences, University of Minho, Campus de Gualtar, 4710-057 Braga, Portugal.

Tel.: +351253604812;

fax: +351253604809;

e-mail: pludovico@ecsaude.uminho.pt

Received 29 May 2013; revised 8 July 2013; accepted 13 August 2013.

DOI: 10 1111/1567-1364 12070

Editor: Dina Petranovic

Keywords

hormesis; ROS signalling; oxidative damage; caloric restriction; longevity; ageing.

Abstract

For more than 50 years, the free radical theory served as the paradigm guiding most investigations of ageing. However, recent studies in a variety of organisms have identified conceptual and practical limitations to this theory. Some of these limitations are related to the recent discovery that caloric restriction and other experimental manipulations promote longevity by inducing hormesis effects in association with increased reactive oxygen species (ROS). The beneficial role of ROS in lifespan extension is consistent with the essential role of these molecules in cell signalling. However, the identity of specific forms of ROS that promote longevity remains unclear. In this article, we argue that in several model systems, hydrogen peroxide plays a crucial role in the induction of hormesis.

Introduction

Hormesis is an adaptive response to a variety of oxidative and other stresses that renders cells/organisms resistant to higher (and normally harmful) doses of the same stressing agent (Martins et al., 2011). The hormesis effects of reactive oxygen species (ROS) and their impact on ageing have emerged as one of the most important topics for understanding ageing as reflected in a number of recently published review articles (Hekimi et al., 2011; Ristow & Schmeisser, 2011; Calabrese, 2012; Labunskyv & Gladvshev, 2012; Maryanovich & Gross, 2013). This review will address the question of the specific form of ROS that trigger hormesis. Recently, we established that hydrogen peroxide as a hormesis trigger and extend chronological lifespan (CLS) in budding yeast (Mesquita et al., 2010). Nevertheless, more recent work on hormesis claims that superoxide anions are the triggers of hormesis effects. In this review, we decided to 'bring the case to the court' and to present the evidence supporting superoxide anions or hydrogen peroxide as triggers of hormetic effects. Arguments will be presented showing that hydrogen peroxide may be a general inducer of hormesis effects downstream of the superoxide anions through their conversion

to hydrogen peroxide by superoxide dismutases. The evidence that hydrogen peroxide may induce quiescence, which is emerging as an important determinant of longevity, will be also discussed.

ROS as 'bad guys' - guilty as charged?

In the never-ending battle between processes that maintain cellular homoeostasis and factors that disrupt this equilibrium, ROS usually have been viewed as 'bad guys'. The deleterious effects of ROS are particularly relevant to the longstanding 'free radical theory of ageing', which posits that oxidative stress and oxidative damage to macromolecules are primary determinants of ageing. Despite the prominent position this theory has attained in more than 50 years of efforts to understand ageing, as has also been noted by others recently (Lapointe & Hekimi, 2010; Hekimi et al., 2011), mounting evidence suggests that this theory is at best incomplete.

It has long been recognized that ROS cause complex and irreversible damage to cellular constituents that impair cellular homoeostasis (reviewed in de Grey, 2006). Oxidative damage is related to the high reactivity of molecular oxygen and its intermediates, which can lead

P. Ludovico & W.C. Burhans

to oxidative modifications of proteins, lipids and DNA. One of the primary sources of ROS is mitochondria, and ROS damage to mtDNA has been implicated in a number of pathological conditions, including ageing. mtDNA damage can lead to aberrant mitochondrial function that can cause 'vicious cycles' of ROS production, which amplify damage. ROS-induced damage to mitochondria eventually became the basis for reformulation of this theory as the 'mitochondrial free radical theory'.

The results of numerous studies performed over a period of decades provide support for the free radical theory. For example, oxidatively damaged molecules accumulate in aged postmitotic cells in species as diverse as yeast and humans. These molecules include misfolded proteins, protein carbonyls and lipofuscin all of which slowly accumulate with age. Lipid peroxidation also increases with age and disrupts cellular functions occurring in biomembranes. A role for oxidative damage to DNA in ageing is consistent with accumulating evidence that genome instability and the rate at which this instability occurs increase with age. The free radical theory of ageing is also consistent with numerous studies in a variety of organisms showing that inhibition of antioxidant defences shortens lifespan. Additional support for the theory is provided by the observation that ROS inhibit telomerase and promote telomere erosion, a well-recognized component of ageing in eukaryotes (Passos et al., 2007a). In the aggregate, the evidence that ROS and oxidative damage are important factors in ageing is compelling in many respects.

The case against the 'free radical theory of ageing'

In spite of the compelling evidence that ROS are guilty of disrupting cellular homoeostasis, in recent years, major cracks have appeared in the defence of the free radical theory. Evidence against the free radical theory has surfaced in studies of a variety of model organisms employed to investigate ageing. For example, we have reported the surprising observation that in the model organism Saccharomyces cerevisiae (budding yeast), hydrogen peroxide promotes chronological longevity, which is defined as the length of time that yeast cells survive in stationary phase cultures. The longevity effects of hydrogen peroxide occur in response to caloric restriction or inactivation of catalases and in cells ectopically exposed to low levels of hydrogen peroxide (Mesquita et al., 2010). Our experiments provided additional evidence against the free radical theory as well – the longevity phenotype of catalase mutants was accompanied by elevated damage in the form of protein carbonyls and lipofuscin (Mesquita et al., 2010). Similarly, elevated intracellular levels of ROS induced by 2-deoxy-D-glucose, which mimics caloric restriction by inhibiting glucose metabolism, were reported to extend the lifespan of worms (*Caenorhabditis elegans*; Schulz *et al.*, 2007; Hekimi *et al.*, 2011). Impaired insulin/ IGF-1 signalling was also recently reported to induce a transient increase in ROS that extends worm lifespan (Zarse *et al.*, 2012). These reports are relevant to the well-established role that growth signalling by glucose and other factors plays in ageing in all organisms.

Additional evidence against the free radical theory was provided by the observation that exposure of long-lived age-1 worms, but not wild-type worms, to high oxygen tension extends lifespan in parallel with decreased mitochondrial ROS and fewer protein carbonyls (Yanase et al., 2002). These findings imply that the longevity effects of high oxygen in age-1 worms are related to an initial increase in superoxide levels. Increased levels of superoxide anions have also been implicated in longevity effects in wild-type worms. For example, the lifespan of wild-type worms is extended by exposure to low concentrations of the superoxide-generating compound juglone (Cypser & Johnson, 2002). Elevated superoxide anion levels induced by mutations in the nuo-6 and isp-1 genes encoding subunits of complex I and III of the respiratory chain or by exposing worms to the superoxide-generating compound paraguat also extend the lifespan of wild-type worms (Yang & Hekimi, 2010). The absence of an effect in wild-type worms exposed to hyperoxic conditions may reflect an upper limit of superoxide anions that promote longevity, which may have been exceeded in these conditions. These and other findings recently reported in studies of more complex organisms (Perez et al., 2009) argue for acquittal of the charges against ROS brought by the free radical theory as 'bud guys' that promote ageing.

The verdict: guilty of some, but not all charges

Although hydrogen peroxide enhances chronological lifespan (CLS) in calorie-restricted budding yeast cells, this occurs in parallel with a decrease in superoxide anions. This decrease is caused by induction by hydrogen peroxide of the activity of the cytosolic Cu/Zn-dependent and the mitochondrial Mn-dependent superoxide dismutases, Sod1p and Sod2p, respectively (Mesquita et al., 2010). The induction of SOD activity by hydrogen peroxide is consistent with the earlier demonstration that budding yeast CLS is extended by SOD overexpression (Fabrizio et al., 2004; Harris et al., 2005). These findings are also consistent with earlier reports that sublethal concentrations of hydrogen peroxide induce transcription of both the SOD1 and SOD2 genes (Godon et al., 1998; Gasch et al., 2000) as well as an increase in levels of the corresponding proteins (Godon et al., 1998). Transactivation of SOD1 and SOD2 is mediated in part by the transcriptional regulator Yap1p (Yahara, 1996). It is well documented that hydrogen peroxide modifies Yap1p as part of a regulatory mechanism associated with Yap1p induction of a large number of genes (Delaunay *et al.*, 2000, 2002).

The induction of superoxide dismutase transcription and/or activity by hydrogen peroxide is a highly conserved regulatory mechanism. For example, hydrogen peroxide induces sodA transcription in Escherichia coli (Semchyshyn, 2009), as well as the transcription and activity of MnSOD in rat cells (Yoshioka et al., 1994). Recent data, obtained in S. cerevisiae, also suggest that not only the interaction between hydrogen peroxide and cellular proteins plays a role in signalling transduction but also thiol peroxidases can sense and transfer oxidative signals to signalling proteins regulating transcription (Fomenko et al., 2011). Thus, not only the regulatory mechanism of hydrogen peroxide seems to be conserved, but also its role in promoting longevity may be conserved as well. Consistent with this possibility, the replicative lifespan of human skin keratinocytes is extended by ectopic exposure to low concentrations of hydrogen peroxide (Yokoo et al., 2004). This effect is accompanied by an increase in telomere length (Yokoo et al., 2004). In mammalian cells, telomere elongation is inhibited by superoxide anions (Passos et al., 2007b), and telomere maintenance is enhanced by SOD activity (Serra et al., 2003). It is possible, therefore, that the replicative lifespan-extending effect of hydrogen peroxide in human keratinocytes is partially related to hydrogen peroxideinduced SOD activity that reduces superoxide anions.

A transient increase in intracellular hydrogen peroxide has also been implicated in lifespan extension associated with impaired insulin/IGF-1 signalling in C. elegans, which disrupts glucose uptake and up regulates superoxide dismutases and other oxidative stress defences, leading to a subsequent decline in overall levels of ROS (Zarse et al., 2012). Increased hydrogen peroxide also underlies the pro-survival effects of inactivating catalases or ectopic exposure to hydrogen peroxide in mutant C. elegans dauer larvae (Xie & Roy, 2012). The beneficial effects of hydrogen peroxide reported in this latter study were attributed to the induction of HIF-1-dependent alterations in lipid metabolism. Elevated intracellular levels of hydrogen peroxide may also underlie the switch from short-lived 'worker' ants to longer-lived reproductive gamergates in the eusocial insect Harpegnathos saltator. This switch is accompanied by a substantial decrease in catalase activity, which is expected to elevate intracellular levels of hydrogen peroxide (Schneider et al., 2011).

The longevity-promoting effects of caloric restriction induced by hydrogen peroxide in stationary phase budding yeast cells are accompanied by an enhanced

quiescence state indicated by the reduced frequency of budded cells (Weinberger et al., 2007, 2010). This reflects the less frequent entry of calorie-restricted stationary phase cells into S phase (Weinberger et al., 2013), where they suffer replication stress (Murakami et al., 2012; Weinberger et al., 2013). Even in the absence of caloric restriction, hydrogen peroxide levels increase in parallel with the reduction in superoxide levels as budding yeast cells approach stationary phase (Mesquita et al., 2010; Pan et al., 2011). This suggests that the regulation of SODs by hydrogen peroxide is a general feature of quiescence in budding yeast that is enhanced by caloric restriction. A similar reciprocal relationship between increased levels of hydrogen peroxide and decreased levels of superoxide anions has been reported in cultured mouse cells as they approach quiescence due to contact inhibition (Sarsour et al., 2008). Furthermore, disrupting this relationship by inactivating MnSOD, which elevates levels of superoxide and decreases levels of hydrogen peroxide, inhibits quiescence as indicated by a higher fraction of cells that remained in S phase followed by cell death (Sarsour et al., 2008). Quiescence is also emerging as an important component of longevity in mammalian cells (Chakkalakal et al., 2012). Although a causal role for hydrogen peroxide in maintaining quiescence in mammalian cells has not been addressed, the studies described above clearly suggest this possibility.

Hydrogen peroxide and/or superoxide anions – the hormesis police?

All of these findings point to superoxide anions as a guilty party in the loss of cellular homoeostasis that underlies ageing and to low levels of hydrogen peroxide as the 'sheriff' that keeps superoxide anions in check. The critical role of hydrogen peroxide in policing levels of superoxide anions conforms to current concepts of a phenomenon called hormesis (Calabrese, 2012). However, the results of some studies suggest that not all superoxide anions are bad guys that need to be restrained by oxidative stress defences - similar to hydrogen peroxide, superoxide anions are also capable of enhancing lifespan (Fig. 1). This includes, for example, the longevity-promoting effect of hyperoxia in age-1 worms, as well as the lifespan-extending effects in wild-type worms of the compounds juglone and paraguat and of mutations in the nuo-6 and isp-1 genes described above (Yang & Hekimi, 2010). In budding yeast, inactivation of TOR signalling by deletion of the TOR1 gene extends CLS by increasing superoxide anions early in CLS experiments (Pan et al., 2011). Consistent with a role for superoxide anions in lifespan extension of tor1∆ cells, the superoxide-generating compound menadione mimics the longevity effects of P. Ludovico & W.C. Burhans

TOR1 inactivation. Also consistent with this model, uncoupling of mitochondrial respiration from oxidative phosphorylation with dinitrophenol, which inhibits the

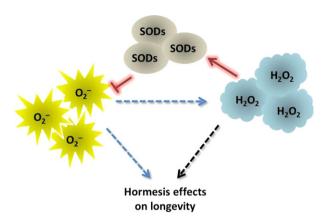
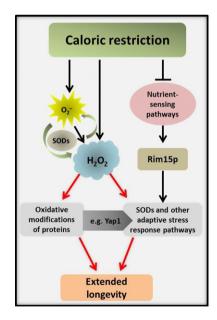


Fig. 1. Both hydrogen peroxide and superoxide anions have been implicated in hormesis effects that promote longevity. Whether superoxide anions can induce hormesis effects directly or indirectly (blue dashed arrows) via their conversion into hydrogen peroxide by SODs remains unclear. Importantly, the levels of superoxide anions and hydrogen peroxide are regulated by SOD activity. Hydrogen peroxide increases and promotes SOD activity that decreases superoxide levels (red arrows), thus creating a feedback regulatory mechanism that promotes hormesis effects by maintaining hydrogen peroxide at homoeostatic levels.

production of superoxide anions, also attenuates the life-span-extending effect of *TOR1* deletion (Pan *et al.*, 2011).

The lifespan-extending effects of ectopic exposure to hydrogen peroxide detected in budding yeast (Mesquita et al., 2010) and worms (Xie & Roy, 2012) definitively establish this form of ROS as a bona fide inducer of hormesis. Although it has been suggested that the effects of ROS on lifespan are conceptually quite distinct from hormesis effects (Hekimi et al., 2011), the effects of hydrogen peroxide on lifespan were detected in parallel with adaptive responses that elevated SOD activity (Mesquita et al., 2010) or altered lipid metabolism (Xie & Roy, 2012), which clearly meets the definition of hormesis. It is important to keep in mind that one source of intracellular hydrogen peroxide is the dismutation of superoxide anions by SODs. Therefore, the hormesis effects of superoxide anions reported in some of the studies cited above may in fact be triggered by hydrogen peroxide produced from superoxide anions by SOD activity (Fig. 1). Consistent with this possibility, caloric restriction induced by switching exponentially proliferating cultures to a lower concentration of glucose at the beginning of yeast chronological ageing experiments induces an initial increase in superoxide anions (Weinberger et al., 2010), which is followed by the sustained increase in hydrogen peroxide that triggers SOD activity and reduced levels of superoxide



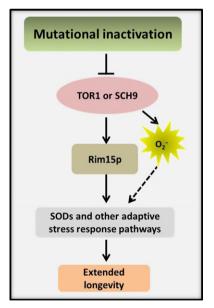


Fig. 2. Pathways mediating hormesis extension of CLS in budding yeast triggered by caloric restriction or mutational inactivation of growth signalling pathways. Dotted lines represent hypothetical pathways. Although O_2^- is dismutated to H_2O_2 , whether this is the source of H_2O_2 in calorie-restricted cells has not yet been tested and other potential sources exist, such as peroxisomes. Hydrogen peroxide modification of the transcriptional activator Yap1p (Georgiou, 2002) likely contributes to the induction of SODs (Mesquita *et al.*, 2010) and other adaptive stress response pathways. It is likely that H_2O_2 also induces oxidative stress responses by modifying other proteins, including those regulating epigenetic effects that also contribute to longevity, as described in mammalian cells (reviewed in Maryanovich & Gross, 2013). Although indirect evidence suggests that O_2^- triggers oxidative stress defences when Tor1p-Sch9p signalling is mutational inactivated, this possibility also has not been tested directly.

anions (Mesquita *et al.*, 2010). The early burst of superoxide anions induced by caloric restriction in these experiments likely reflects an increase in respiration rate first reported in calorie-restricted cells several years ago (Lin *et al.*, 2002).

However, the increased CLS triggered by inactivating *TOR1* is accompanied by a decrease, rather than increase, in hydrogen peroxide levels (Pan *et al.*, 2011). A decrease in intracellular hydrogen peroxide also occurs in concert with chronological lifespan extension in stationary phase cells when *SCH9*, a downstream effector of *TOR1* signalling, is inactivated (Weinberger *et al.*, 2010). Similarly, the increase in superoxide anions in long-lived *nuo-6* and *isp-1* mutant worms described above was detected in the absence of an increase in hydrogen peroxide or other forms of ROS (Yang & Hekimi, 2010). These results point to superoxide anions as a different branch of the hormesis police that extends lifespan in the absence of hydrogen peroxide effects.

The specific forms of ROS that trigger hormesis effects in laboratory experiments may be related to how specific experimental manipulations impact a myriad of signalling pathways that interact with one another in complex ways. In fact, although a number of antioxidant and repair processes have been characterized, the mechanisms whereby cells are damaged by particular oxidants or protect themselves from damage remain to be identified (Thorpe *et al.*, 2004).

The parallels between hormetic effects of caloric restriction or mutational inactivation of growth signalling pathways in budding yeast chronological ageing experiments may be informative in this regard (Fig. 2). Caloric restriction extends CLS in part by attenuating Tor1p-Sch9p-dependent glucose signalling, which leads to the activation of Rim15p and oxidative stress defences regulated by this transcriptional activator (Wei et al., 2008). As noted above, caloric restriction also increases intracellular hydrogen peroxide, which also induces oxidative stress defences (Mesquita et al., 2010), but independently of Rim15p (Weinberger et al., 2010). The increase in hydrogen peroxide in calorie-restricted cells may be triggered by the increase in superoxide anions that occurs before cells transition into stationary phase (Weinberger et al., 2010), although other possible sources of hydrogen peroxide exist. Mutational inactivation of Tor1p-Sch9p (Fig. 2) also induces Rim15p-dependent oxidative stress defences, as well as an increase in intracellular superoxide anions, which may occur independently of Rim15p. This increase may induce SODs and other oxidative stress defences and/or altered mitochondrial function (Pan et al., 2011). Ultimately, the effects of both types of experimental manipulations converge on a reduction in superoxide anions in cells in stationary phase that promotes longevity (Fig. 2).

In the context of these parallels and the fact that both types of experimental manipulations result in reduced Tor1p-Sch9p signalling, it is puzzling that mutational activation of Sch9p or Tor1p does not lead to an increase in hydrogen peroxide, similar to caloric restriction. Apparently, in addition to down regulating Tor1p and Sch9p signalling, caloric restriction triggers additional events that increase intracellular levels of hydrogen peroxide. We propose that this occurs as part of a mechanism that amplifies the hormetic effects of mutationally inactivating a subset of signalling pathways in laboratory experiments. Outside the laboratory, the depletion of nutrients likely triggers in yeast and other organisms a deeper and more sustained response that requires hydrogen peroxide to efficiently drive cells into and maintain a quiescent state, where cells are protected from deleterious effects of superoxide anions and a paucity of nutrients on DNA replication (Weinberger et al., 2010). According to this model, superoxide anions have been justly convicted as the bad guys, although the initial increase in their numbers triggered by caloric restriction may alert cells to the need to round them up. This becomes the job of hydrogen peroxide, which may serve as the true hormesis police that promotes longevity in organisms faced with stresses that more closely resemble those they encounter in their natural environment. Future studies will determine whether this verdict is corrected.

Acknowledgements

The authors wish to thank Molly Burhans for preparing the figures. This work was supported by Fundação para a Ciência e Tecnologia (FCT) and COMPETE/QREN/EU (PTDC/BIA-MIC/114116/2009), a grant from the Roswell Park Alliance Foundation and by a National Cancer Institute Support Grant (P30CA016056) to Roswell Park Cancer Institute. Authors have no conflict of interest to declare.

References

Calabrese EJ (2012) Hormesis: improving predictions in the low-dose zone. *EXS* **101**: 551–564.

Chakkalakal JV, Jones KM, Basson MA & Brack AS (2012)
The aged niche disrupts muscle stem cell quiescence. *Nature* **490**: 355–360.

Cypser JR & Johnson TE (2002) Multiple stressors in Caenorhabditis elegans induce stress hormesis and extended longevity. J Gerontol A Biol Sci Med Sci 57: B109–B114.

Delaunay A, Isnard AD & Toledano MB (2000) H₂O₂ sensing through oxidation of the Yap1 transcription factor. *EMBO J* **19**: 5157–5166.

Delaunay A, Pflieger D, Barrault MB, Vinh J & Toledano MB (2002) A thiol peroxidase is an H₂O₂ receptor and redox-transducer in gene activation. *Cell* **111**: 471–481.

6 P. Ludovico & W.C. Burhans

- Fabrizio P, Battistella L, Vardavas R *et al.* (2004) Superoxide is a mediator of an altruistic aging program in *Saccharomyces cerevisiae. J Cell Biol* **166**: 1055–1067.
- Fomenko DE, Koc A, Agisheva N *et al.* (2011) Thiol peroxidases mediate specific genome-wide regulation of gene expression in response to hydrogen peroxide. *P Natl Acad Sci USA* **108**: 2729–2734.
- Gasch AP, Spellman PT, Kao CM et al. (2000) Genomic expression programs in the response of yeast cells to environmental changes. Mol Biol Cell 11: 4241–4257.
- Georgiou G (2002) How to flip the (redox) switch. *Cell* 111: 607–610.
- Godon C, Lagniel G, Lee J et al. (1998) The H₂O₂ stimulon in Saccharomyces cerevisiae. J Biol Chem **273**: 22480–22489.
- de Grey AD (2006) Free radicals in aging: causal complexity and its biomedical implications. *Free Radic Res* **40**: 1244–1249.
- Harris N, Bachler M, Costa V, Mollapour M, Moradas-Ferreira P & Piper PW (2005) Overexpressed Sod1p acts either to reduce or to increase the lifespans and stress resistance of yeast, depending on whether it is Cu(2+)-deficient or an active Cu, Zn-superoxide dismutase. *Aging Cell* 4: 41–52.
- Hekimi S, Lapointe J & Wen Y (2011) Taking a "good" look at free radicals in the aging process. *Trends Cell Biol* **21**: 569–576.
- Labunskyy VM & Gladyshev VN (2012) Role of reactive oxygen species-mediated signaling in aging. *Antioxid Redox Signal*. doi:10.1089/ars.2012.4891. [Epub ahead of print].
- Lapointe J & Hekimi S (2010) When a theory of aging ages badly. *Cell Mol Life Sci* **67**: 1–8.
- Lin SJ, Kaeberlein M, Andalis AA et al. (2002) Calorie restriction extends Saccharomyces cerevisiae lifespan by increasing respiration. Nature 418: 344–348.
- Martins I, Galluzzi L & Kroemer G (2011) Hormesis, cell death and aging. *Aging (Albany NY)* 3: 821–828.
- Maryanovich M & Gross A (2013) A ROS rheostat for cell fate regulation. *Trends Cell Biol* 23: 129–134.
- Mesquita A, Weinberger M, Silva A *et al.* (2010) Caloric restriction or catalase inactivation extends yeast chronological lifespan by inducing H₂O₂ and superoxide dismutase activity. *P Natl Acad Sci USA* **107**: 15123–15128.
- Murakami C, Delaney JR, Chou A *et al.* (2012) pH neutralization protects against reduction in replicative lifespan following chronological aging in yeast. *Cell Cycle* 11: 3087–3096.
- Pan Y, Schroeder EA, Ocampo A, Barrientos A & Shadel GS (2011) Regulation of yeast chronological life span by TORC1 via adaptive mitochondrial ROS signaling. *Cell Metab* 13: 668–678.
- Passos JF, Saretzki G & von Zglinicki T (2007a) DNA damage in telomeres and mitochondria during cellular senescence: is there a connection? *Nucleic Acids Res* **35**: 7505–7513.
- Passos JF, Saretzki G, Ahmed S *et al.* (2007b) Mitochondrial dysfunction accounts for the stochastic heterogeneity in telomere-dependent senescence. *PLoS Biol* 5: e110.

Perez VI, Bokov A, Van Remmen H, Mele J, Ran Q, Ikeno Y & Richardson A (2009) Is the oxidative stress theory of aging dead? *Biochim Biophys Acta* 1790: 1005–1014.

- Ristow M & Schmeisser S (2011) Extending life span by increasing oxidative stress. Free Radic Biol Med 51: 327–336.
- Sarsour EH, Venkataraman S, Kalen AL, Oberley LW & Goswami PC (2008) Manganese superoxide dismutase activity regulates transitions between quiescent and proliferative growth. *Aging Cell* 7: 405–417.
- Schneider SA, Schrader C, Wagner AE, Boesch-Saadatmandi C, Liebig J, Rimbach G & Roeder T (2011) Stress resistance and longevity are not directly linked to levels of enzymatic antioxidants in the ponerine ant *Harpegnathos saltator*. *PLoS One* 6: e14601.
- Schulz TJ, Zarse K, Voigt A, Urban N, Birringer M & Ristow M (2007) Glucose restriction extends *Caenorhabditis elegans* life span by inducing mitochondrial respiration and increasing oxidative stress. *Cell Metab* 6: 280–293.
- Semchyshyn H (2009) Hydrogen peroxide-induced response in *E. coli* and *S. cerevisiae*: different stages of the flow of the genetic information. *Cent Eur J Biol* 4: 142–153.
- Serra V, von Zglinicki T, Lorenz M & Saretzki G (2003) Extracellular superoxide dismutase is a major antioxidant in human fibroblasts and slows telomere shortening. *J Biol Chem* **278**: 6824–6830.
- Thorpe GW, Fong CS, Alic N, Higgins VJ & Dawes IW (2004) Cells have distinct mechanisms to maintain protection against different reactive oxygen species: oxidative-stress-response genes. *P Natl Acad Sci USA* **101**: 6564–6569.
- Wei M, Fabrizio P, Hu J, Ge H, Cheng C, Li L & Longo VD (2008) Life span extension by calorie restriction depends on Rim15 and transcription factors downstream of Ras/PKA, Tor, and Sch9. PLoS Genet 4: e13.
- Weinberger M, Feng L, Paul A *et al.* (2007) DNA replication stress is a determinant of chronological lifespan in budding yeast. *PLoS ONE* 2: e748.
- Weinberger M, Mesquita A, Caroll T *et al.* (2010) Growth signaling promotes chronological aging in budding yeast by inducing superoxide anions that inhibit quiescence. *Aging* (*Albany NY*) 2: 709–726.
- Weinberger M, Sampaio-Marques B, Ludovico P & Burhans WC (2013) DNA replication stress-induced loss of reproductive capacity in *S. cerevisiae* and its inhibition by caloric restriction. *Cell Cycle* **12**: 1189–1200.
- Xie M & Roy R (2012) Increased levels of hydrogen peroxide induce a HIF-1-dependent modification of lipid metabolism in AMPK compromised *C. elegans* dauer larvae. *Cell Metab* 16: 322–335.
- Yahara I (1996) Stress-inducible cellular responses. Introduction. *EXS* **77**: XI–XII.
- Yanase S, Yasuda K & Ishii N (2002) Adaptive responses to oxidative damage in three mutants of *Caenorhabditis elegans*

- (age-1, mev-1 and daf-16) that affect life span. Mech Ageing Dev 123: 1579–1587.
- Yang W & Hekimi S (2010) A mitochondrial superoxide signal triggers increased longevity in *Caenorhabditis elegans*. *PLoS Biol* 8: e1000556.
- Yokoo S, Furumoto K, Hiyama E & Miwa N (2004) Slow-down of age-dependent telomere shortening is executed in human skin keratinocytes by hormesis-like-effects of trace hydrogen peroxide or by anti-oxidative effects of pro-vitamin C in common
- concurrently with reduction of intracellular oxidative stress. *J Cell Biochem* **93**: 588–597.
- Yoshioka T, Homma T, Meyrick B, Takeda M, Moore-Jarrett T, Kon V & Ichikawa I (1994) Oxidants induce transcriptional activation of manganese superoxide dismutase in glomerular cells. *Kidney Int* **46**: 405–413.
- Zarse K, Schmeisser S, Groth M *et al.* (2012) Impaired insulin/ IGF1 signaling extends life span by promoting mitochondrial L-proline catabolism to induce a transient ROS signal. *Cell Metab* **15**: 451–465.