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RESEARCH ARTICLE

Carboplatin sensitivity in epithelial ovarian cancer cell lines: The impact of model systems

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Abstract

Epithelial ovarian cancer (EOC) is the most lethal gynecologic malignancy in North America, underscoring the need for the development of new therapeutic strategies for the management of this disease. Although many drugs are pre-clinically tested every year, only a few are selected to be evaluated in clinical trials, and only a small number of these are successfully incorporated into standard care. Inaccuracies with the initial *in vitro* drug testing may be responsible for some of these failures. Drug testing is often performed using 2D monolayer cultures or 3D spheroid models. Here, we investigate the impact that these different *in vitro* models have on the carboplatin response of four EOC cell lines, and in particular how different 3D models (polydimethylsiloxane-based microfluidic chips and ultra low attachment plates) influence drug sensitivity within the same cell line. Our results show that carboplatin responses were observed in both the 3D spheroid models tested using apoptosis/cell death markers by flow cytometry. Contrary to previously reported observations, these were not associated with a significant decrease in spheroid size. For the majority of the EOC cell lines (3 out of 4) a similar carboplatin response was observed when comparing both spheroid methods. Interestingly, two cell lines classified as resistant to carboplatin in 2D cultures became sensitive in the 3D models, and one sensitive cell line in 2D culture showed resistance in 3D spheroids. Our results highlight the challenges of choosing the appropriate pre-clinical models for drug testing.

Introduction

Epithelial ovarian cancer (EOC) is the 5th cause of cancer-related deaths in North American women with a 5-year survival of 45% [1]. The standard front-line treatment for EOC includes cytoreductive surgery and treatment with platinum DNA alkylating agents such as carboplatin or cisplatin combined with the anti-microtubule drug paclitaxel [2]. Although initial response

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rates are high (>70%), the disease eventually recurs in most patients, who will then develop chemoresistance [2–4]. To improve EOC survival, new therapies and their combinations are under investigation, and preclinical models are used to test their efficacy prior to eventual human trials. Because of time and cost efficiency, these models are often in the form of 2D or 3D *in vitro* assays. However, optimizing and translating the results seen in 2D *in vitro* models to an *in vivo* response is challenging, as these models do not take into account important tumor characteristics such as hypoxia, drug transport restriction, extra cellular matrix (ECM), anchorage-independent growth as well as interactions with other cell types in the tumor, including stroma/fibroblast and immune cells [5–8]. Alternatively, 3D spheroid models have been proposed as an attractive alternative as they are thought to more closely resemble the tumor tissue architecture [9–11].

There are several methods to prepare 3D spheroids from cell lines (reviewed elsewhere [11, 12]) including hanging-droplets [13–16], rotating wall vessel cultures [14, 17], and ultra-low attachment (ULA) 96-well round-bottom plates [11, 18, 19]. In ovarian cancer, several groups have described the capacity of EOC cell lines to form spheroids by different methodologies [20–26], including our work using hanging droplets [16, 27–29]. These different techniques each have their own challenges. On one hand, hanging-droplet spheroids take longer to form, are difficult to manipulate for spheroid transfer steps, and they present a challenge for experiments requiring medium changes [30, 31] or treatments. On the other hand, spheroids formed in ULA plates can be readily used in high-throughput assays [18, 19] but many cell lines do not form spheroids under these conditions. Considering the importance of the tissue microenvironment in the establishment of 3D models, components of the ECM have been used to assist spheroid formations [11]. Matrigel, consisting of collagen type IV, perlecan/heparin sulfate proteoglycan 2 and laminin, is one of the most commonly used ECM mimics [11, 32] and has been shown to improve spheroid formation in ULA plates for cell lines that otherwise would not have the capacity to aggregate [18, 19]. More recently, many bioengineered microfluidic devices have been developed and shown to be effective in forming spheroids [33–39]. In addition, the use of these microfluidic devices has gained popularity as a 3D model system for drug screening [12, 33–44]. In ovarian cancer, our group has been a pioneer in the application of this technology to evaluate drug response in EOC cell lines [37, 38, 41].

Although several studies have compared the chemosensitivity of ovarian cancer cell lines grown as monolayers with 3D-spheroid cultures [18, 41, 45, 46], little is known about the impact that different 3D spheroid model systems have on their sensitivity. In the present study, we investigated the ability of four different EOC cell lines to form 3D spheroids following different methodologies (hanging-droplets, ULA plates and microfluidic chips), and their carboplatin sensitivity in monolayer culture versus spheroids formed within microfluidic devices or round-bottom ULA plates (the latter using Matrigel as an ECM) (Fig 1). Response was measured by flow cytometry for apoptosis/cell death and spheroid size measurement. Our results show that spheroid formation of EOC cell lines is significantly faster and more uniform in polydimethylsiloxane (PDMS) microfluidic devices and Matrigel-assisted ULA plates than in hanging-droplets or ULA plates without Matrigel. Carboplatin responses were observed in both 3D spheroid models using flow cytometric analysis, but no significant decrease in spheroid size was detected. For the majority of the EOC cell lines (3 out of 4) a similar response to carboplatin treatment was observed by both spheroid methods. Interestingly, two cell lines classified as resistant to carboplatin in 2D cultures responded as sensitive in 3D models, and one sensitive cell line in 2D culture showed resistance in the 3D spheroids. Our results highlight the challenges in choosing appropriate pre-clinical models for empirical drug testing.

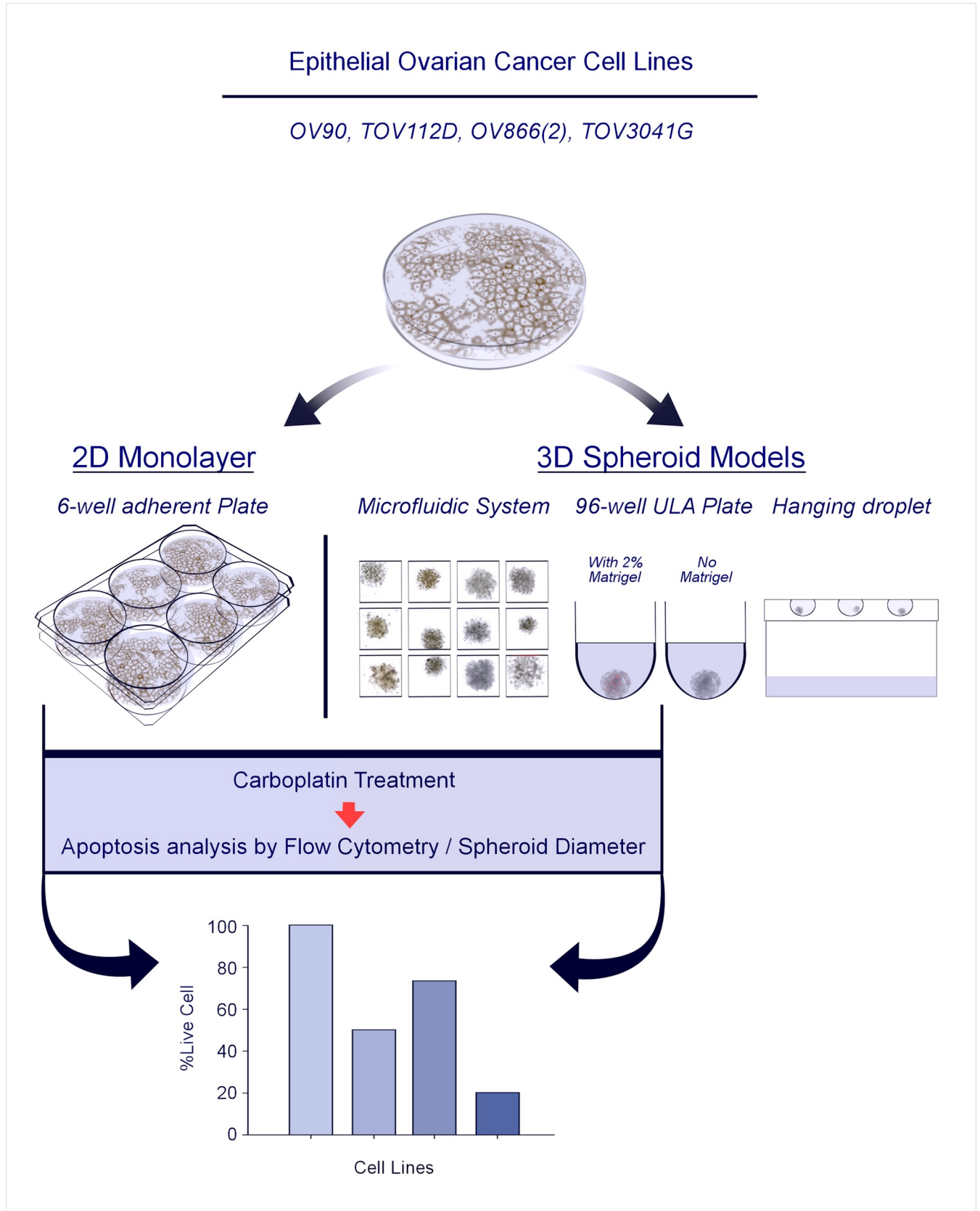


Fig 1. Schematic illustration of the study strategy. For each cell line used, 3D spheroid formation was evaluated by four different methods, i.e., PDMS-based microfluidic systems, ultra-low attachment (ULA) plates in the presence or not of 2% Matrigel, and hanging droplets. The two most effective 3D spheroid methods (microfluidic chips and Matrigel-assisted ULA plates) were used to define carboplatin sensitivity of each cell line that was then compared to responses in the 2D monolayer assay.

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Materials and methods

Cell lines

Four different human EOC cell lines were used in this study, TOV3041G, TOV112D, OV90 and OV866(2) (all BRCA1/2 wild-type). Three cell lines [TOV112D, OV90, OV866(2)] are classified as resistant to carboplatin therapy in 2D culture as measured by clonogenic assay, and one (TOV3041G) is classified as sensitive (Table 1). These cell lines were derived in our laboratory from patient tumor (TOV) or ascites (OV) [27, 47] samples. All cell lines were authenticated at the beginning of this study using STR profiling by the McGill University Genome Center (Montreal, Canada). Cells were cultured in complete OSE medium (Wisent, QC, Canada) supplemented with 10% fetal bovine serum (Wisent), 2.5 µg/mL of amphotericin B (Wisent) and 50 µg/mL of gentamicin (Wisent). Cells were maintained in 100 mm petri dishes with 5% CO₂ at 37°C.

Clonogenic survival assays

The IC₅₀'s for carboplatin, as determined by clonogenic survival assay, have previously been reported for TOV3041G, TOV112D, and OV866(2) [27, 28]. Carboplatin sensitivity for the OV90 cell line was determined in this study using the same clonogenic assay [27]. Briefly, cells were seeded in a 6-well plate at a density of 750 cells/well that allowed the formation of individual colonies. After seeding, cells were allowed to adhere for 16 hours in a 37°C, 5% CO₂ incubator after which the medium was removed and replaced with OSE complete medium containing carboplatin (0–100µM) (Hospira Healthcare Corporation, Saint-Laurent, QC). Cells were incubated with the drug for 24 hours. The drug was then removed and OSE complete medium was added to each well. When colonies became visible at a 2X magnification plates were fixed with cold methanol and stained with a solution of 0.5% blue methylene (Sigma–Aldrich Inc., St. Louis, MO) in 50% methanol. Colonies were counted under a stereomicroscope and reported as percent of control. IC₅₀ values were determined using Graph Pad Prism 5 software (GraphPad Software Inc., San Diego, CA). Each individual experiment was performed in triplicate and repeated three times.

Spheroid formation

Spheroids were formed using three different methods: hanging-droplet, PDMS-based microfluidic chips and ULA 96-well plates (with and without Matrigel). Hanging-droplet spheroids were generated as previously described [16]. Briefly, 16 µL droplets containing 4000 cells each were placed on the inside of the lid of a 150 mm cell culture plate. The lid was then gently

Table 1. Carboplatin sensitivity (IC₅₀ by clonogenic assay) of the EOC cell lines used in this study.

	Carboplatin IC ₅₀ (µM)	Reference
TOV3041G	5.2 ± 1.1	Fleury et al. 2015
TOV112D	13.4	Letourneau et al. 2012
OV90	31.8 ± 5.4	This study
OV866(2)	32.1 ± 7.1	Fleury et al. 2015

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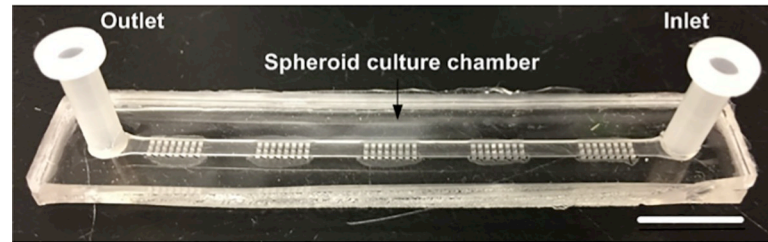


Fig 2. Design of PDMS microfluidic device developed for the culture of 3D spheroids and drug testing. Each chip is comprised of five sections of 24 wells, containing 120 wells. Each well has a dimension of $500 \times 500 \times 500 \mu\text{m}^3$. The scale bar is 1 cm.

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inverted and replaced on the base of the plate containing sterile PBS for humidification and the plate was put in the incubator with the droplets hanging. Spheroids were allowed to grow for up to 6 days. For the microfluidic method, each device contained 120 spheroid culture chambers, with one inlet and one outlet (Fig 2, see fabrication section below). A cell suspension of 1×10^6 cells in 1 mL of complete OSE medium was used. The device was seeded with $100 \mu\text{L}$ of this suspension in the inlet, and the same amount of medium was removed from the outlet. This process was repeated by seeding in the outlet and removing medium from the inlet and performed five times in each direction (total 10 times, using all of the 1×10^6 cells) to allow uniform distribution of cells across the wells. Spheroids were allowed to form over 48 hours. Finally, spheroids were also prepared using 96-well round-bottom ULA plates (Corning 4515). Wells were seeded with 2000 cells in $100 \mu\text{L}$ of cooled complete OSE medium with or without 2% Matrigel (BD, Bioscience, Canada). This was done for each cell line and condition. Spheroids were allowed to form over 48 hours.

Microfluidic device fabrication

The PDMS microfluidic device used in this work is a modification of a previously described device [35]. It was made of PDMS and used the basic principle of gravity trapping [34, 41, 48]. The device layout was developed using CATIA (Dassault Systèmes, France) and the mold was carved out of polymethyl methacrylate. Using this mold, our device was made of two bonded layers of PDMS (Fig 2); 1) the bottom later contains five segments of spheroid culture chambers, each of which contains 24 wells of $500 \mu\text{m} \times 500 \mu\text{m} \times 500 \mu\text{m}$, and 2) the top layer contains a straight channel with a height of $500 \mu\text{m}$ covering the entirety of the spheroid culture chambers with 3 mm holes for the inlet and outlet at the ends of the straight channel. The device was bonded using an upright microscope to align the layers after 30 sec of atmospheric plasma surface treatment. After bonding, the device was put in an oven at 80°C for two hours. The dimension of the device was 7.5 cm long x 1 cm wide with of 120 spheroid culture chambers (5 blocks of 24 chambers each). Nine devices could be culture at a time in an incubator using a simple pipet tip box. Prior to use, the devices were treated with 10 mg/ml Pluronic (Sigma-Aldrich, St Louis, USA) as previously described [48], to make the PDMS surface less adherent to cells. This treatment also prevents chemo-absorption by the PDMS [49].

Carboplatin treatment

Previously, we have shown that the carboplatin IC_{50} for spheroids from a specific EOC cell line (TOV112D) is approximately 10x the corresponding monolayer IC_{50} value [41]. Therefore, as the most resistant cell line in this study has an IC_{50} of $30 \mu\text{M}$ (Table 1), we treated the spheroids formed in the PDMS-based microfluidic systems and in the ULA plates with $300 \mu\text{M}$

carboplatin. Moreover, in our earlier pilot studies using concentrations 10 times lower (30 μM) and 10 times higher (3000 μM), no differential results among the cell lines were observed, as they all survived at the lower dose and all died at the higher. Interestingly, the 300 μM carboplatin concentration is similar to that found in the plasma of ovarian cancer patients undergoing chemotherapy [48]. Spheroids formed 48 hours after cell seeding were treated with fresh medium (controls) or fresh medium containing 300 μM carboplatin (Hospira Healthcare Corporation) for a period of 48 hours. Carboplatin treatment was also performed on monolayer cultures prepared with 5,000 cells/well of each cell line seeded into 6-well plates. Consistent with the method for spheroids, 48 hours after inoculation, the cultures were treated with fresh medium (controls) or fresh medium containing 300 μM carboplatin and cultures were incubated for 48 hours. After treatment samples were immediately used in subsequent analyses.

Microscopy and imaging

Microscopy images were taken using an inverted microscope (Nikon TS100) at 10X or 4X magnification two and four days after seeding as well as 48 hours after treatment prior to flow cytometry analysis. PDMS microfluidic devices were put into a 100 mm petri dish for imaging purposes to maintain sterile conditions. Size estimation was done using ImageJ software (Version 1.49, National Institute of Health, Bethesda, MD).

Flow cytometry analysis

After carboplatin treatment, monolayer cells from each well of the 6-well plates were harvested by trypsin-EDTA (0.05%) as a single cell suspension. For the spheroids treated in the ULA plates, 10 were collected, pooled and dissociated with trypsin-EDTA (0.05%) for 5 minutes to obtain single-cell suspensions. For those treated in the PDMS-based microfluidic devices, 48 were pooled and dissociated as above. Cells were then labelled with Annexin-V (3:100 dilution) and 7-Amino-ActinomycinD (7-AAD, 5:100 dilution) to detect apoptotic and dead cells, respectively (PE Annexin V apoptosis Detection Kit I, BD Biosciences). After a 15-minute incubation at RT (25°C) in the dark, the stained cells were analyzed by flow cytometry (LSR-Fortessa, BD Biosciences), within 2 hours of staining. The data collected from each acquisition was analyzed using the FlowJo software (FlowJo, LLC, Ashland, USA). After excluding cell debris, viable cells were selected based on the absence of 7-AAD and/or Annexin-V markers (see example on [S1 Fig](#)).

Statistical analysis

Values are expressed as the mean \pm SEM (standard error of the mean), derived from at least three independent experiments. Single comparisons between two groups were determined by Student's t-test (paired, two-tailed). Comparisons between multiple groups were determined by Tukey ANOVA multi comparison test. *P* values < 0.05 were considered significant. All statistical analyses were done using GraphPad Prism 6 software (GraphPad Software Inc., San Diego, CA).

Results

Comparison of spheroid formation using different methods

In order to compare carboplatin sensitivity of different EOC cell lines in 2D monolayers and in 3D spheroids obtained by different methods, we first evaluated the efficiency of each 3D technique (hanging-droplets, ULA plates, and PDMS-based microfluidic chips) to form

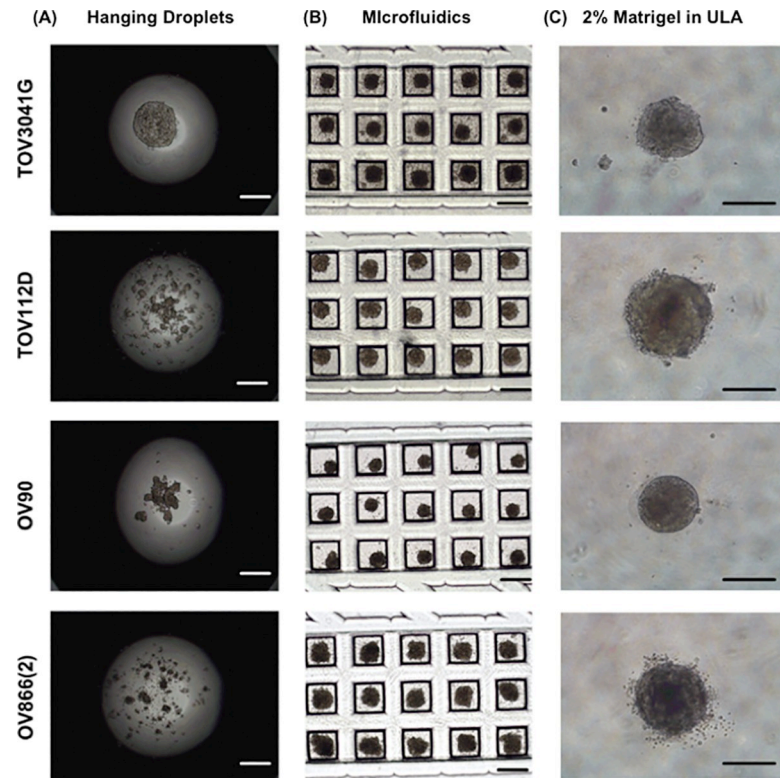


Fig 3. Capacity of EOC cell lines to form spheroids in three different culture conditions. Representative microscopic bright field images of TOV3041G, TOV112D, OV90 and OV866(2) cells cultured 48 hours in hanging droplets (A), within the custom PDMS-based microfluidic chips, (B) and in 96-well round-bottom ULA plates with 2% Matrigel (C). The scale bar is 500 μ m.

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spheroids using four EOC cell lines [TOV3041G, TOV112D, OV90 and OV866(2)]. Fig 3A shows the hanging-droplet cultures at day 2 with the four EOC cell lines. Only TOV3041G formed spheroids within 48 hours. Careful observation showed that, at this time point, TOV112D and OV90 had started to form multiple smaller spheroids. However, OV866(2) had not formed spheroids at all. Culturing the same cell lines up to day 4 and day 6, we observed that the smaller spheroids of TOV112D and OV90 had fused together to form larger spheroids (S2 Fig). However, OV866(2) did not show any sign of spheroid formation. Fig 3B shows bright field images of 3D cultures at day 2 using the PDMS-based microfluidic device. All cell lines formed spheroids in the microfluidic device within 48 hours. Fig 3C shows the bright field images of Matrigel-assisted spheroids using ULA wells at 48 hours. Again, all the cell lines formed spheroid within 48 hours. However, less compact spheroids were obtained when EOC cell lines were grown in the ULA plates in the absence of Matrigel (S3 Fig). Therefore, carboplatin sensitivity was evaluated in 3D spheroids formed in the PDMS microfluidic device and the Matrigel-assisted ULA plate since the latter is widely used for drug sensitivity assays [11, 18, 19, 45, 50].

Carboplatin sensitivity of EOC cell line spheroids obtained by two different methods

Sensitivity of the spheroids was evaluated at a concentration of 300 μ M carboplatin, a concentration similar to that found in the plasma of ovarian cancer patients undergoing chemotherapy [48]. This concentration is in range of what has been used in other studies (100 μ M,

330 μM) to evaluate platinum sensitivity of ovarian cancer spheroids [18, 46]. It is also 10 times higher than the 2D IC_{50} (by clonogenic assay) of the most resistant cell lines OV90 and OV866(2) (see Table 1). Then, the effect of carboplatin on our EOC cell lines was evaluated by measuring the size of the spheroids, as is commonly used in the literature [18, 19, 45, 51], and by measuring live/apoptotic/dead cells by flow cytometry using Annexin V/7-AAD staining of the dissociated spheroids. For the spheroids grown in the PDMS-based microfluidic chips, we observed a significant decrease in the percentage of live cells following treatment with 300 μM carboplatin for the OV90 and OV866(2) spheroids, but not for the TOV3041G and TOV112D cells, the latter being the most resistant in this assay (Fig 4B). However, no significant decrease

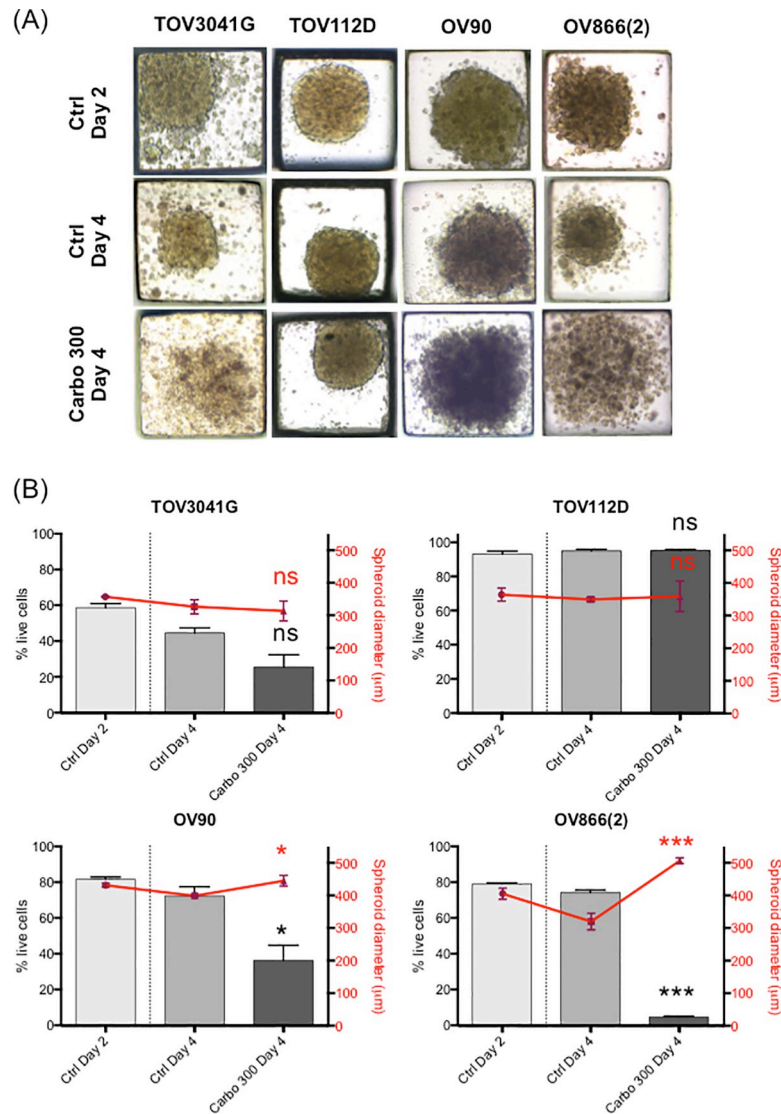


Fig 4. Carboplatin response of EOC spheroids cultured in PDMS microfluidic devices. 3D spheroids of four EOC cell lines [TOV3041G, TOV112D, OV90, OV866(2)] were formed and cultured in microfluidic devices for 2 and 4 days (Ctrl Day 2 and Ctrl Day 4) or treated at day 2 with 300 μM of carboplatin and assessed at day 4 (Carbo 300 Day 4). A) Representative microscopic bright field images at 10X magnification. B) Comparison of carboplatin treatment on EOC cell survival (grey bars) and spheroid diameters (red points). 48 spheroids were analyzed (in duplicate) from each set of experiment readouts. Error bars indicate the standard errors of the mean (SEM) of three independent experiments. Statistical analyses: Ctrl Day 4 vs Carbo 300 Day 4, * $p < 0.05$; *** $p < 0.0005$; ns, non-significant (Student's t-test).

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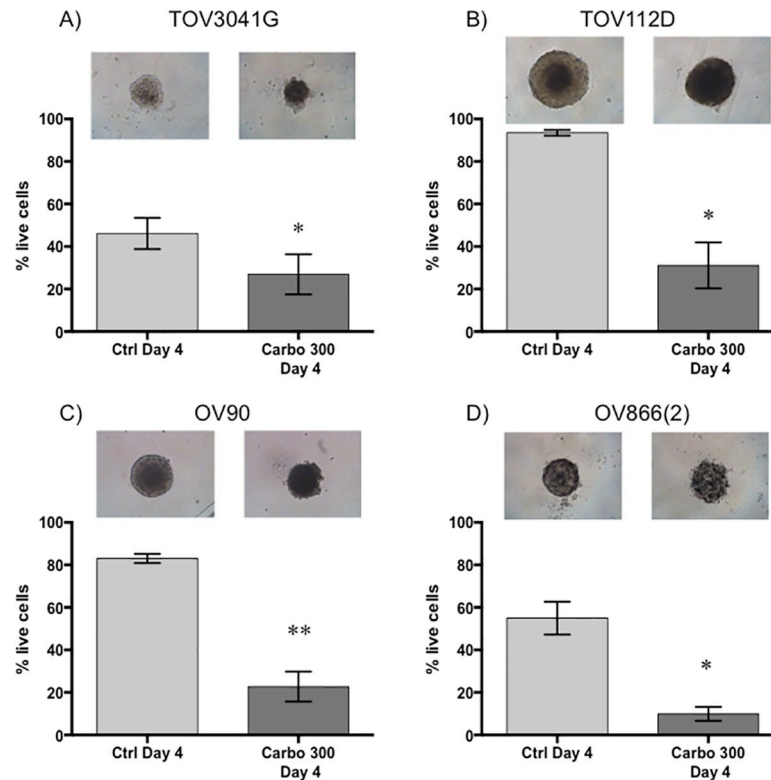


Fig 5. Carboplatin response of EOC spheroids cultured with 2% matrigel in ultra-low attachment wells. 3D spheroids of TOV3041G (A), TOV112D (B), OV90 (C) and OV866(2) (D) were formed and cultured in concave-bottom ULA plates in the presence of 2% Matrigel for 4 days (Ctrl Day 4) or treated at day 2 with 300 μ M of carboplatin and assessed at day 4 (Carbo 300 Day 4). Top panels are representative microscopic bright field images of each condition. Bar graphs represent cell survival analysed by flow cytometry of control (light grey) and treated (dark grey) spheroids. Data are shown as the mean \pm SEM of three independent experiments. * $p < 0.05$; ** $p < 0.005$ (Student's t-test).

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in the size of spheroids was observed, and on the contrary, sizes of the OV90 and OV866(2) spheroids were significantly increased, likely due to spheroid disaggregation (Fig 4A and 4B). For spheroids grown in the Matrigel-assisted ULA plates, our results showed that all four cell lines presented decreased viability after incubation with 300 μ M carboplatin, including TOV112D, as assessed by Annexin V/7-AAD staining and flow cytometry, but we did not notice a change in size when visually inspecting these spheroids (Fig 5 inserts). In the Matrigel-assisted ULA plates the highest resistance to carboplatin was observed in the TOV3041G cell line. Of note, sizes of non-treated spheroids were less than 500 μ m in diameter in days 2 and 4 in both methods (Figs 3–5), which is important to avoid extensive internal necrotic core.

Comparative carboplatin response of 3D spheroids with that of 2D monolayers

The carboplatin sensitivity of cell lines in 2D culture was originally obtained using the clonogenic assay (Table 1), where cells are plated at a low dilution in order to obtain colonies from single cells. However, in our 3D spheroid assays (both methods), a large number of cells are used to form the spheroids and carboplatin sensitivity is determined by the percentage of residual live cells. In order to approximate analysis methods used for the 2D and 3D drug sensitivity assays, we treated cell monolayers at 80% confluence with carboplatin (300 μ M), and

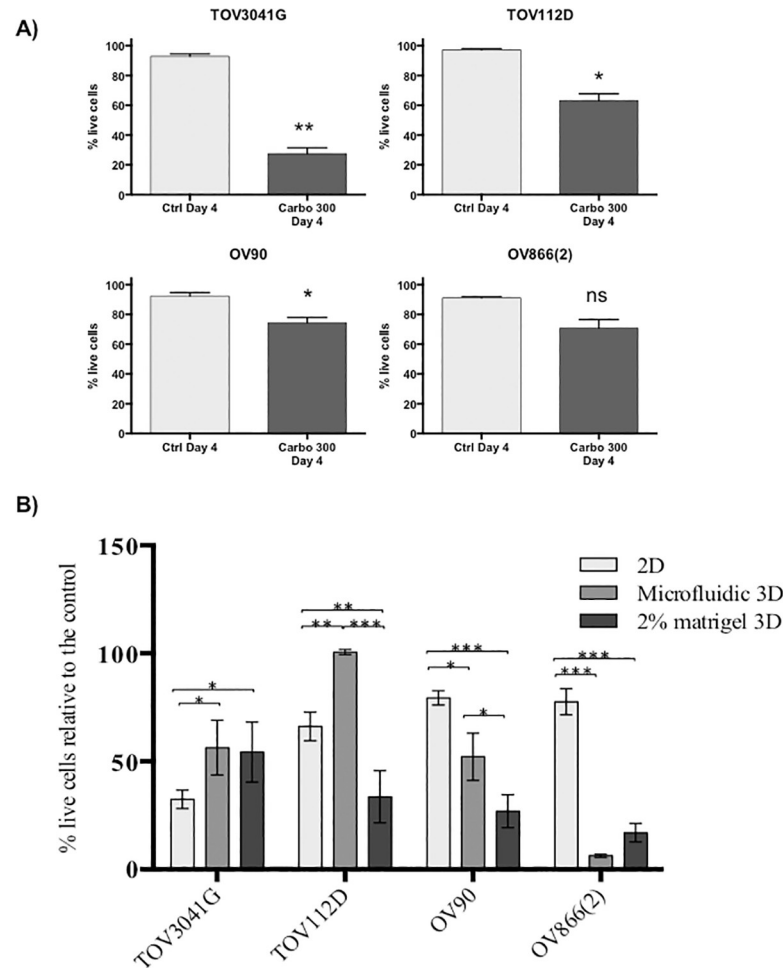


Fig 6. Comparison of carboplatin response between 2D-monolayers and two different 3D spheroid models. A) TOV3041G, TOV112D, OV90 and OV866(2) were cultured as 2D monolayers for 4 days without treatment (Ctrl Day 4) or treated at day 2 with 300 μ M of carboplatin and assessed at day 4 (Carbo 300 Day 4). Bar graphs represent cell survival analysed by flow cytometry of control (light grey) and treated (dark grey). Data are shown as the mean \pm SEM of three independent experiments * $p < 0.05$; ** $p < 0.005$; ns, non-significant (Student's t-test) B) 2D-monolayer carboplatin sensitivity was compared to that of 3D spheroid cultures. Each treatment response was normalized relative to the percentage of live cells in comparison to the appropriate control. * p -value < 0.05 ; ** p -value < 0.005 ; *** p -value < 0.0005 ; Turkey ANOVA multi comparison test of three independent experiments.

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evaluated cell viability by flow cytometry as for the spheroids. Sensitivity of 2D monolayers (Fig 6A) corresponded proportionately with that of the clonogenic assays where TOV3041G was the most sensitive and OV90 and OV866(2) the most resistant (Table 1).

When comparing the percentage of live cells by flow cytometry (normalized to controls) in the three methods tested, i.e. monolayer, spheroids from PDMS-based microfluidic devices and Matrigel-assisted spheroids, we observed that the spheroids of TOV3041G were significantly more resistant to carboplatin than in monolayer culture, and that the spheroids of OV90 and OV866(2) were significantly more sensitive than their respective monolayer cultures. This was independent of the spheroid formation method used (Fig 6B). However, when comparing both 3D cultures, OV90 spheroids formed in the microfluidic devices were significantly more resistant than those obtained using Matrigel-assisted ULA plates. However, both spheroid models were significantly more sensitive than the monolayer cultures. In contrast,

spheroids of the TOV112D cell line formed in the microfluidic device were more resistant than the monolayer culture but were more sensitive when formed by the Matrigel-assisted technique. Table 2 summarizes this comparative study, in which a cell line ranking #1 has lower viability (more sensitivity) than the cell line ranking as #2, and this approach was further applied to determine rank order. When cell lines have similar percentage of cell survival, the same ranking is given to both of them. This classification shows a clear inversion of sensitivity from 2D to 3D for two of the four cell lines studied, in which the most sensitive becomes resistant to carboplatin (TOV3041G) and the most resistant becomes the most sensitive [OV866(2)].

Discussion

In recent years, 3D cell culture models such as multi-cellular spheroids have become more widely used as pre-clinical models and are expected to bridge the gap between 2D and animal models. In the particular case of ovarian cancer, 3D spheroids are clinically relevant models since in advanced disease EOC cells can spread to the peritoneum as aggregates/spheroids [52, 53]. In the literature, several EOC cell lines have been studied by different spheroid generating methods, including hanging-droplets [16, 22, 27–29, 41, 54, 55], biomimetic hydrogels [23], plastic wells coated with matrix scaffolds [20, 46, 56], and more recently by microfluidics [33, 34, 37, 38, 42, 44] and commercially available round-bottom ULA plates [18, 45, 50, 57]. In the present study we first compared the ability of four EOC cell lines to form spheroids using these ULA 96-well plates (in the absence or presence of Matrigel) with that of hanging-droplets and PDMS-based microfluidic chips. For the latter, we used a recently developed [34, 37, 38] PDMS microfluidic device with a capacity of 120 spheroids per device. We showed that both Matrigel-assisted ULA plates and simple well-based PDMS microfluidic devices were the most robust and efficient methods to form spheroids. However, our microfluidic devices have the advantage of using much smaller volumes of media or drug, which is desirable when working with expensive compounds or extensive drug screenings.

We evaluated the carboplatin response of EOC cell lines grown as spheroids using these two methods and compared these to the respective monolayer cultures. Spheroid size is widely used as a measure of drug response in several studies, including those with EOC cells [18, 19, 45, 51], where decreased spheroid size has been interpreted as reflecting cell death [19]. In contrast, our results show that, independent of the method of spheroid formation used, no decrease in spheroid volume was observed. However, a significant increase in spheroid volume was observed for two cell lines [OV90 and OV866(2)] when using PDMS-based microfluidic devices, likely reflecting spheroid disaggregation due to cell death. Our findings indicate that spheroid size is not appropriate to be used as a measurement of carboplatin response, at least when using a short incubation time (like 48h of this study), as cell dissociation/disaggregation due to cell death/apoptosis would increase spheroid size. Therefore, the response of the spheroids and monolayers to the drug was also assessed by the broadly accepted method of flow cytometry through the analysis of apoptosis (Annexin V) and cell death (7-AAD). We

Table 2. Relative carboplatin sensitivity of four EOC cell lines in three different culture conditions.

Cell line	2D culture model	3D culture model	
		PDMS Microfluidic devices	Ultra-low attachment wells and 2% Matrigel
TOV3041G	1	2	3
TOV112D	2	3	2
OV90	3	2	2
OV866(2)	3	1	1

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observed decreased cell survival of spheroids after carboplatin treatment, with the highest level of cell death being observed in the two cell lines that showed increased spheroid volumes. Strikingly, in three of the four cell lines studied, the 3D sensitivity (independent of the spheroid formation method used) was inverted when compared with the corresponding 2D test, where two cell lines [OV90 and OV866(2)] classified as resistant in 2D cultures became sensitive in 3D models, and one sensitive cell line (TOV3041G) in 2D culture showed resistance in the 3D model. The latter result is more consistent with the literature where 3D spheroids are more resistant than 2D cultures [11, 18, 34, 41, 46, 58]. Although the exact signaling mechanism implicated in this resistance is still unclear, reports have attributed it to decreased compound access, reduced sensitivity in response to hypoxia, cells cycling more slowly, cell-cell contact influence, and an increased EMT phenotype [11, 12, 58].

In a recent report [18], a comparative analysis of cisplatin response of monolayer cultures and 3D spheroids of 16 EOC cell lines showed that the majority of the spheroids (13/16) were more resistant to cisplatin than the respective 2D cultures (much as we have observed with the TOV3041G cell line), whereas two cell lines were equally sensitive and one showed higher sensitivity in 3D spheroids than 2D monolayers, the latter similar to what we observed in two of our cell lines [OV9 and OV866(2)]. Another study using 11 EOC cell lines showed increased cisplatin resistance of 3D spheroids in seven cell lines, with no change for the other four [46]. It is possible too that the read-out methods used in these studies have underestimated spheroids cell death when compared to ours. In these articles, platinum sensitivity of ovarian cancer cell lines cultured as 3D spheroids have been mainly evaluated by metabolic assay (since it is a more high-throughput assay) that would overestimate cell survival, as early apoptotic or senescent cells are still metabolically active. We think that the cell death/apoptosis/cell viability FACS method used in our work, although less high-throughput, would more accurately estimate cell survival after carboplatin treatment in 3D spheroids. Nevertheless, all together, these findings imply that the assumption that spheroids are more resistant to chemotherapy than the monolayer cultures is not always accurate and may influence the choice of EOC models used to study chemoresistance. Previous publications have attempted to correlate molecular features (i.e. epithelial and mesenchymal protein biomarkers) with platinum sensitivity of ovarian cancer cell lines grown as 2D vs 3D [18, 46]. Based in these results, it is unlikely that these factors play a role in the sensitivity inversion of our cell lines.

When comparing the two spheroid models, we generally obtained similar carboplatin responses, except for the TOV112D cell line. Spheroids of this cell line were more resistant than the monolayer when assayed using the PDMS-based microfluidic device, but more sensitive in the Matrigel-assisted ULA plates. It is possible that this cell line is more affected by the Matrigel presence than the others. It is known that ECM components, like Matrigel, can affect drug responses [11]. Although we treated our PDMS devices with pluronic to prevent cell adherence and chemo-absorption [49], performing 3D assays in both PDMS chips and conventional plates enabled us to rule out the well-known absorption of drugs by the hydrophobic porous matrix of the PDMS as a significant cause for this discrepancy [59]. While the absorbed concentration has not been directly controlled, our results (Fig 6B) do not show a systematic increase in resistance in the PDMS devices as would be suggested if the cells are experiencing an effectively lower concentration due to material absorption. Rather, resistance is slightly lower in one cell line [OV866(2)] and significantly higher in another one (TOV112D). Regarding the latter, it is possible that this cell line is more affected by components of the ECM that might influence drug efficacy. Matrigel is a mixture of different ECM components that has high variability issues due to batch effect [60], making it difficult to thoroughly investigate the role of this ECM mimic in the 3D chemosensitivity of this cell line. Therefore, because PDMS-based microfluidic devices were fast, reliable and high-throughput for the analyses of 3D

spheroids, and because they used very small media/drug volumes, they make an interesting alternative choice for drug screening either in pharmaceutical research or in clinical settings. In a very recent report, 3D spheroids from tumor tissue of ovarian cancer patients were obtained using the round-bottom ULA plates (without Matrigel) [57]. When compared to clinical outcome, response of these patient-derived 3D spheroids to chemotherapeutic drugs, including the standard of care carboplatin-paclitaxel combination therapy, showed concordance in 89% of the cases (39 of 44) [57]. However, spheroids of different sizes, shapes and compactness were observed from sample to sample and a high number of replicates were needed. The PDMS microfluidic device described here may overcome this technical difficulty by providing a large number of uniform spheroids to be tested for drug response using less patient sample, suggesting that it could easily be incorporated into a clinic setting or used for drug development in the future.

Although descriptive, the present work aims to emit a clear warning that there is no strong tendency of EOC cells to become more resistant in 3D than in 2D, and that spheroid size is an inefficient predictor of cell response as size variation can be both a change of cell number or a change in tissue compactness.

Conclusion

In drug discovery, the choice of an appropriate *in vitro* pre-clinical model can have a major influence on correctly predicting drug response in human clinical trials. Here, we evaluated carboplatin sensitivity of four epithelial ovarian cancer cell lines using both 2D and 3D *in vitro* models and tested different spheroid forming methods. Channel-based spheroid forming chips held several advantages over commercially available ultra-low attachment plates or the hanging droplet method. Irrespective of the spheroid forming method used, some striking carboplatin sensitivity inversions were noted within a given cell line when 2D and 3D results were compared. Overall, the present study highlights the challenges of *in vitro* models and discusses the different facets of 2D and 3D *in vitro* drug testing.

Supporting information

S1 Fig. 7-AAD and Annexin-V flow cytometry analysis. Graphs represent an example of flow cytometry analysis of spheroids treated with 300 μ M carboplatin and its respective control. After excluding cell debris, viable cells were selected based on the absence of 7-AAD and/or Annexin-V markers (red square). Apoptotic and/or dead cells are shown in the other quadrants.

(TIF)

S2 Fig. Spheroid formation using hanging drops with different EOC cell lines. (A) Bright field images at day 4. (B) Bright field images at day 6. Scale bar, 500 μ m.

(TIF)

S3 Fig. Spheroid formation using ULA round-bottom wells (without Matrigel) with different EOC cell lines. Bright field images at day 2. Note that TOV112D cells in these conditions form multiple spheroids of smaller size. Scale bar, 300 μ m.

(TIF)

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References

1. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2019. *CA Cancer J Clin.* 2019; 69(1):7–34. <https://doi.org/10.3322/caac.21551> PMID: 30620402
2. Lheureux S, Gourley C, Vergote I, Oza AM. Epithelial ovarian cancer. *Lancet.* 2019; 393(10177):1240–53. [https://doi.org/10.1016/S0140-6736\(18\)32552-2](https://doi.org/10.1016/S0140-6736(18)32552-2) PMID: 30910306
3. Coleman RL, Monk BJ, Sood AK, Herzog TJ. Latest research and treatment of advanced-stage epithelial ovarian cancer. *Nature reviews Clinical oncology.* 2013; 10(4):211–24. <https://doi.org/10.1038/nrclinonc.2013.5> PMID: 23381004
4. Wilson MK, Pujade-Lauraine E, Aoki D, Mirza MR, Lorusso D, Oza AM, et al. Fifth Ovarian Cancer Consensus Conference of the Gynecologic Cancer InterGroup: recurrent disease. *Annals of oncology: official journal of the European Society for Medical Oncology.* 2017; 28(4):727–32.
5. Imamura Y, Mukohara T, Shimono Y, Funakoshi Y, Chayahara N, Toyoda M, et al. Comparison of 2D- and 3D-culture models as drug-testing platforms in breast cancer. *Oncol Rep.* 2015; 33(4):1837–43. <https://doi.org/10.3892/or.2015.3767> PMID: 25634491
6. Johnson JI, Decker S, Zaharevitz D, Rubinstein LV, Venditti JM, Schepartz S, et al. Relationships between drug activity in NCI preclinical in vitro and in vivo models and early clinical trials. *British journal of cancer.* 2001; 84(10):1424–31. <https://doi.org/10.1054/bjoc.2001.1796> PMID: 11355958
7. Phillips RM, Bibby MC, Double JA. A critical appraisal of the predictive value of in vitro chemosensitivity assays. *Journal of the National Cancer Institute.* 1990; 82(18):1457–68. <https://doi.org/10.1093/jnci/82.18.1457> PMID: 2202838
8. Riedl A, Schleder M, Pudielko K, Stadler M, Walter S, Unterleuthner D, et al. Comparison of cancer cells in 2D vs 3D culture reveals differences in AKT-mTOR-S6K signaling and drug responses. *J Cell Sci.* 2017; 130(1):203–18. <https://doi.org/10.1242/jcs.188102> PMID: 27663511
9. Elliott NT, Yuan F. A review of three-dimensional in vitro tissue models for drug discovery and transport studies. *Journal of pharmaceutical sciences.* 2011; 100(1):59–74. <https://doi.org/10.1002/jps.22257> PMID: 20533556
10. Nyga A, Cheema U, Loizidou M. 3D tumour models: novel in vitro approaches to cancer studies. *Journal of cell communication and signaling.* 2011; 5(3):239–48. <https://doi.org/10.1007/s12079-011-0132-4> PMID: 21499821

11. Xu X, Farach-Carson MC, Jia X. Three-dimensional in vitro tumor models for cancer research and drug evaluation. *Biotechnology advances*. 2014; 32(7):1256–68. <https://doi.org/10.1016/j.biotechadv.2014.07.009> PMID: 25116894
12. Hoarau-Vechot J, Raffii A, Touboul C, Pasquier J. Halfway between 2D and Animal Models: Are 3D Cultures the Ideal Tool to Study Cancer-Microenvironment Interactions? *International journal of molecular sciences*. 2018; 19(1). <https://doi.org/10.3390/ijms19010181> PMID: 29346265
13. Frey O, Misun PM, Fluri DA, Hengstler JG, Hierlemann A. Reconfigurable microfluidic hanging drop network for multi-tissue interaction and analysis. *Nature communications*. 2014; 5:4250. <https://doi.org/10.1038/ncomms5250> PMID: 24977495
14. Pampaloni F, Reynaud EG, Stelzer EH. The third dimension bridges the gap between cell culture and live tissue. *Nature reviews Molecular cell biology*. 2007; 8(10):839–45. <https://doi.org/10.1038/nrm2236> PMID: 17684528
15. Tung YC, Hsiao AY, Allen SG, Torisawa YS, Ho M, Takayama S. High-throughput 3D spheroid culture and drug testing using a 384 hanging drop array. *Analyst*. 2011; 136(3):473–8. <https://doi.org/10.1039/c0an00609b> PMID: 20967331
16. Zietarska M, Maugard CM, Filali-Mouhim A, Alam-Fahmy M, Tonin PN, Provencher DM, et al. Molecular description of a 3D in vitro model for the study of epithelial ovarian cancer (EOC). *Molecular carcinogenesis*. 2007; 46(10):872–85. <https://doi.org/10.1002/mc.20315> PMID: 17455221
17. Castaneda F, Kinne RK. Short exposure to millimolar concentrations of ethanol induces apoptotic cell death in multicellular HepG2 spheroids. *Journal of cancer research and clinical oncology*. 2000; 126(6):305–10. <https://doi.org/10.1007/s004320050348> PMID: 10870639
18. Heredia-Soto V, Redondo A, Berjon A, Miguel-Martin M, Diaz E, Crespo R, et al. High-throughput 3-dimensional culture of epithelial ovarian cancer cells as preclinical model of disease. *Oncotarget*. 2018; 9(31):21893–903. <https://doi.org/10.18632/oncotarget.25098> PMID: 29774110
19. Vinci M, Gowan S, Boxall F, Patterson L, Zimmermann M, Court W, et al. Advances in establishment and analysis of three-dimensional tumor spheroid-based functional assays for target validation and drug evaluation. *BMC biology*. 2012; 10:29. <https://doi.org/10.1186/1741-7007-10-29> PMID: 22439642
20. Chen J, Wang J, Chen D, Yang J, Yang C, Zhang Y, et al. Evaluation of characteristics of CD44+CD117+ ovarian cancer stem cells in three dimensional basement membrane extract scaffold versus two dimensional monocultures. *BMC cell biology*. 2013; 14:7. <https://doi.org/10.1186/1471-2121-14-7> PMID: 23368632
21. Chen MW, Yang ST, Chien MH, Hua KT, Wu CJ, Hsiao SM, et al. The STAT3-miRNA-92-Wnt Signaling Pathway Regulates Spheroid Formation and Malignant Progression in Ovarian Cancer. *Cancer research*. 2017; 77(8):1955–67. <https://doi.org/10.1158/0008-5472.CAN-16-1115> PMID: 28209618
22. Dong Y, Stephens C, Walpole C, Swedberg JE, Boyle GM, Parsons PG, et al. Paclitaxel resistance and multicellular spheroid formation are induced by kallikrein-related peptidase 4 in serous ovarian cancer cells in an ascites mimicking microenvironment. *PloS one*. 2013; 8(2):e57056. <https://doi.org/10.1371/journal.pone.0057056> PMID: 23451143
23. Loessner D, Stok KS, Lutolf MP, Huttmacher DW, Clements JA, Rizzi SC. Bioengineered 3D platform to explore cell-ECM interactions and drug resistance of epithelial ovarian cancer cells. *Biomaterials*. 2010; 31(32):8494–506. <https://doi.org/10.1016/j.biomaterials.2010.07.064> PMID: 20709389
24. Masiello T, Dhall A, Hemachandra LPM, Tokranova N, Melendez JA, Castracane J. A Dynamic Culture Method to Produce Ovarian Cancer Spheroids under Physiologically-Relevant Shear Stress. *Cells*. 2018; 7(12). <https://doi.org/10.3390/cells7120277> PMID: 30572633
25. Nunes AS, Barros AS, Costa EC, Moreira AF, Correia IJ. 3D tumor spheroids as in vitro models to mimic in vivo human solid tumors resistance to therapeutic drugs. *Biotechnology and bioengineering*. 2019; 116(1):206–26. <https://doi.org/10.1002/bit.26845> PMID: 30367820
26. Rafehi S, Ramos Valdes Y, Bertrand M, McGee J, Prefontaine M, Sugimoto A, et al. TGFbeta signaling regulates epithelial-mesenchymal plasticity in ovarian cancer ascites-derived spheroids. *Endocrine-related cancer*. 2016; 23(3):147–59. <https://doi.org/10.1530/ERC-15-0383> PMID: 26647384
27. Fleury H, Communal L, Carmona E, Portelance L, Arcand SL, Rahimi K, et al. Novel high-grade serous epithelial ovarian cancer cell lines that reflect the molecular diversity of both the sporadic and hereditary disease. *Genes Cancer*. 2015; 6(9–10):378–98. <https://doi.org/10.18632/genesandcancer.76> PMID: 26622941
28. Letourneau IJ, Quinn MC, Wang LL, Portelance L, Caceres KY, Cyr L, et al. Derivation and characterization of matched cell lines from primary and recurrent serous ovarian cancer. *BMC Cancer*. 2012; 12:379. <https://doi.org/10.1186/1471-2407-12-379> PMID: 22931248
29. Ouellet V, Zietarska M, Portelance L, Lafontaine J, Madore J, Puiffe ML, et al. Characterization of three new serous epithelial ovarian cancer cell lines. *BMC Cancer*. 2008; 8:152. <https://doi.org/10.1186/1471-2407-8-152> PMID: 18507860

30. Gupta N, Liu JR, Patel B, Solomon DE, Vaidya B, Gupta V. Microfluidics-based 3D cell culture models: Utility in novel drug discovery and delivery research. *Bioeng Transl Med*. 2016; 1(1):63–81. <https://doi.org/10.1002/btm2.10013> PMID: 29313007
31. Joshi P, Lee MY. High Content Imaging (HCI) on Miniaturized Three-Dimensional (3D) Cell Cultures. *Biosensors (Basel)*. 2015; 5(4):768–90. <https://doi.org/10.3390/bios5040768> PMID: 26694477
32. Fridman R, Benton G, Aranoutova I, Kleinman HK, Bonfil RD. Increased initiation and growth of tumor cell lines, cancer stem cells and biopsy material in mice using basement membrane matrix protein (Cul-trex or Matrigel) co-injection. *Nature protocols*. 2012; 7(6):1138–44. <https://doi.org/10.1038/nprot.2012.053> PMID: 22596226
33. Marimuthu M, Rousset N, St-Georges-Robillard A, Lateef MA, Ferland M, Mes-Masson AM, et al. Multi-size spheroid formation using microfluidic funnels. *Lab Chip*. 2018; 18(2):304–14. <https://doi.org/10.1039/c7lc00970d> PMID: 29211088
34. Patra B, Lafontaine J, Bavoux M, Zerouali K, Glory A, Ahanj M, et al. On-chip combined radiotherapy and chemotherapy testing on soft-tissue sarcoma spheroids to study cell death using flow cytometry and clonogenic assay. *Sci Rep*. 2019; 9(1):2214. <https://doi.org/10.1038/s41598-019-38666-9> PMID: 30778138
35. Patra B, Peng CC, Liao WH, Lee CH, Tung YC. Drug testing and flow cytometry analysis on a large number of uniform sized tumor spheroids using a microfluidic device. *Sci Rep*. 2016; 6:21061. <https://doi.org/10.1038/srep21061> PMID: 26877244
36. Sart S, Tomasi RF, Amselem G, Baroud CN. Multiscale cytometry and regulation of 3D cell cultures on a chip. *Nature communications*. 2017; 8(1):469. <https://doi.org/10.1038/s41467-017-00475-x> PMID: 28883466
37. St-Georges-Robillard A, Cahuzac M, Peant B, Fleury H, Lateef MA, Ricard A, et al. Long-term fluorescence hyperspectral imaging of on-chip treated co-culture tumour spheroids to follow clonal evolution. *Integrative biology: quantitative biosciences from nano to macro*. 2019; 11(4):130–41. <https://doi.org/10.1093/intbio/zyz012> PMID: 31172192
38. St-Georges-Robillard A, Masse M, Cahuzac M, Strupler M, Patra B, Orimoto AM, et al. Fluorescence hyperspectral imaging for live monitoring of multiple spheroids in microfluidic chips. *Analyst*. 2018; 143(16):3829–40. <https://doi.org/10.1039/c8an00536b> PMID: 29999046
39. Zuchowska A, Kwapiszewska K, Chudy M, Dybko A, Brzozka Z. Studies of anticancer drug cytotoxicity based on long-term HepG2 spheroid culture in a microfluidic system. *Electrophoresis*. 2017; 38(8):1206–16. <https://doi.org/10.1002/elps.201600417> PMID: 28090668
40. Chen YC, Lou X, Zhang Z, Ingram P, Yoon E. High-Throughput Cancer Cell Sphere Formation for Characterizing the Efficacy of Photo Dynamic Therapy in 3D Cell Cultures. *Sci Rep*. 2015; 5:12175. <https://doi.org/10.1038/srep12175> PMID: 26153550
41. Das T, Meunier L, Barbe L, Provencher D, Guenat O, Gervais T, et al. Empirical chemosensitivity testing in a spheroid model of ovarian cancer using a microfluidics-based multiplex platform. *Biomicrofluidics*. 2013; 7(1):11805. <https://doi.org/10.1063/1.4774309> PMID: 24403987
42. Li SS, Ip CK, Tang MY, Sy SK, Yung S, Chan TM, et al. Modeling Ovarian Cancer Multicellular Spheroid Behavior in a Dynamic 3D Peritoneal Microdevice. *Journal of visualized experiments: JoVE*. 2017(120).
43. Ruppen J, Wildhaber FD, Strub C, Hall SR, Schmid RA, Geiser T, et al. Towards personalized medicine: chemosensitivity assays of patient lung cancer cell spheroids in a perfused microfluidic platform. *Lab Chip*. 2015; 15(14):3076–85. <https://doi.org/10.1039/c5lc00454c> PMID: 26088102
44. Watters KM, Bajwa P, Kenny HA. Organotypic 3D Models of the Ovarian Cancer Tumor Microenvironment. *Cancers*. 2018; 10(8). <https://doi.org/10.3390/cancers10080265> PMID: 30096959
45. Eetezadi S, Evans JC, Shen YT, De Souza R, Piquette-Miller M, Allen C. Ratio-Dependent Synergism of a Doxorubicin and Olaparib Combination in 2D and Spheroid Models of Ovarian Cancer. *Molecular pharmaceutics*. 2018; 15(2):472–85. <https://doi.org/10.1021/acs.molpharmaceut.7b00843> PMID: 29283581
46. Lee JM, Mhaweche-Fauceglia P, Lee N, Parsanian LC, Lin YG, Gayther SA, et al. A three-dimensional microenvironment alters protein expression and chemosensitivity of epithelial ovarian cancer cells in vitro. *Laboratory investigation; a journal of technical methods and pathology*. 2013; 93(5):528–42. <https://doi.org/10.1038/labinvest.2013.41> PMID: 23459371
47. Provencher DM, Lounis H, Champoux L, Tétrault M, Manderson EN, Wang JC, et al. Characterization of four novel epithelial ovarian cancer cell lines. *In Vitro Cell Dev Biol Anim*. 2000; 36(6):357–61. [https://doi.org/10.1290/1071-2690\(2000\)036<0357:COFNEO>2.0.CO;2](https://doi.org/10.1290/1071-2690(2000)036<0357:COFNEO>2.0.CO;2) PMID: 10949993
48. Astolfi M, Péant B, Lateef MA, Rousset N, Kendall-Dupont J, Carmona E, et al. Micro-dissected tumor tissues on chip: an ex vivo method for drug testing and personalized therapy. *Lab Chip*. 2016; 16(2):312–25. <https://doi.org/10.1039/c5lc01108f> PMID: 26659477

49. Tanyeri M, Tay S. Viable cell culture in PDMS-based microfluidic devices. *Methods in cell biology*. 2018; 148:3–33. <https://doi.org/10.1016/bs.mcb.2018.09.007> PMID: 30473072
50. Sokolova E, Kutova O, Grishina A, Pospelov A, Guryev E, Schulga A, et al. Penetration Efficiency of Antitumor Agents in Ovarian Cancer Spheroids: The Case of Recombinant Targeted Toxin DARPIn-LoPE and the Chemotherapy Drug, Doxorubicin. *Pharmaceutics*. 2019; 11(5). <https://doi.org/10.3390/pharmaceutics11050219> PMID: 31067739
51. Mikhail AS, Eetezadi S, Allen C. Multicellular tumor spheroids for evaluation of cytotoxicity and tumor growth inhibitory effects of nanomedicines in vitro: a comparison of docetaxel-loaded block copolymer micelles and Taxotere(R). *PloS one*. 2013; 8(4):e62630. <https://doi.org/10.1371/journal.pone.0062630> PMID: 23626842
52. Kurman RJ, Shih Ie M. The Dualistic Model of Ovarian Carcinogenesis: Revisited, Revised, and Expanded. *The American journal of pathology*. 2016; 186(4):733–47. <https://doi.org/10.1016/j.ajpath.2015.11.011> PMID: 27012190
53. Naora H, Montell DJ. Ovarian cancer metastasis: integrating insights from disparate model organisms. *Nature reviews Cancer*. 2005; 5(5):355–66. <https://doi.org/10.1038/nrc1611> PMID: 15864277
54. Barbolina MV, Adley BP, Kelly DL, Shepard J, Fought AJ, Scholtens D, et al. Downregulation of connective tissue growth factor by three-dimensional matrix enhances ovarian carcinoma cell invasion. *International journal of cancer*. 2009; 125(4):816–25. <https://doi.org/10.1002/ijc.24347> PMID: 19382180
55. Sodek KL, Ringuette MJ, Brown TJ. Compact spheroid formation by ovarian cancer cells is associated with contractile behavior and an invasive phenotype. *International journal of cancer*. 2009; 124(9):2060–70. <https://doi.org/10.1002/ijc.24188> PMID: 19132753
56. Iwanicki MP, Davidowitz RA, Ng MR, Besser A, Muranen T, Merritt M, et al. Ovarian cancer spheroids use myosin-generated force to clear the mesothelium. *Cancer discovery*. 2011; 1(2):144–57. <https://doi.org/10.1158/2159-8274.CD-11-0010> PMID: 22303516
57. Shuford S, Wilhelm C, Rayner M, Elrod A, Millard M, Mattingly C, et al. Prospective Validation of an Ex Vivo, Patient-Derived 3D Spheroid Model for Response Predictions in Newly Diagnosed Ovarian Cancer. *Sci Rep*. 2019; 9(1):11153. <https://doi.org/10.1038/s41598-019-47578-7> PMID: 31371750
58. Niero EL, Rocha-Sales B, Lauand C, Cortez BA, de Souza MM, Rezende-Teixeira P, et al. The multiple facets of drug resistance: one history, different approaches. *Journal of experimental & clinical cancer research: CR*. 2014; 33:37. <https://doi.org/10.1186/1756-9966-33-37> PMID: 24775603
59. Toepke MW, Beebe DJ. PDMS absorption of small molecules and consequences in microfluidic applications. *Lab Chip*. 2006; 6(12):1484–6. <https://doi.org/10.1039/b612140c> PMID: 17203151
60. Zaman MH. The role of engineering approaches in analysing cancer invasion and metastasis. *Nature reviews Cancer*. 2013; 13(8):596–603. <https://doi.org/10.1038/nrc3564> PMID: 23864050