Physiological and vegetative changes in grapevines

View metadata, citation and similar papers at core.ac.uk

provided by Universidade do Minho: RepositoriUM

(BCAs): preliminary results

Felgueiras¹, M.L.; Moutinho-Pereira², J.M.; Dias¹, A.C.P.

1 - Biology Department, Minho University, Campus de Gualtar, 4710-057 Braga - Portugal (acpdias@bio.uminho.pt) 2 – CETA/Trás-os-Montes e Alto Douro University, Quinta de Prados, Apartado 1013 – 5001-911 Vila Real – Portugal

INTRODUCTION

In recent years much research has focused on developing alternative control methods against pre and post harvest decay in grapes (Zahavi et al., 2000). The induction of a natural resistance in grape using several biological and/or chemical elicitors, known as Biological Control Agents (BCAs), has received increasing attention over the last years, and is now considered the elected strategy for disease management (Leon & Joyce, 2004). In this work we evaluate vegetative and physiological changes promoted by BCAs verus conventional chemical products (CQs) in potted grapevines (cv. Alvarinho).

MATERIALS AND METHODS

Plant material – Several grapevine plants (cv. Alvarinho) were established during the spring of 2007. Nine groups of twelve plants were defined for each condition tested: BCAs – Best Cure (BC) [FuturtEco]; Mycotric (My) [FuturtEco]; FitoAlgas (Fito) [SAPEC]; Bio Clean [NutriField]; QCs – Flint (Fit) [Bayer]; Alicite (Ali) [Bayer]; Horizon (Hrz) [Bayer]; Maestro M (MM) [SAPEC]. Each compound was applied, independently, every fifteen days, during July and August of 2007.

Photosynthetic pigments quantification – For five leaves, of each group of twelve plants, photosynthetic pigments were extracted with methanol 90%. Chlorophyll a (clor a), chlorophyll b (clor b), and carotenoids (carot) were quantified based on the specific absorption coefficients: [Clor a = 16,82.4665,2 - 9,28.4652,4; [Clor b = 36,92.4652,4 - 16,54.4665,2]; [Carot = (1000.4470 – 1,91Clor a – Clor b)/225] (Lichtenthaler, 1987). All the results obtained underwent a -test analysis.

Gas exchange measurements – Leaf gas exchange measurements were made, by a portable gas exchange system (LCA-4, Analytical Development Co. Ltd., Hoddesdon, U.K.). Parameters studied were net photosynthesis – A; transpiration – E; stomatal conductance – g.; intercellular concentration of CO₂ – ci, and were calculated according to the approach proposed by Flexas et al. (1998). All the results obtained underwent a t-test analysis.

RESULTS

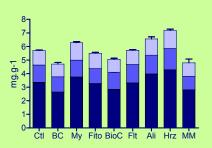


FIGURE 1: Concentration of chlorophyll a (\blacksquare Chl a), chlorophyll b (\blacksquare Chl b) and carotenoids, (\blacksquare Carot). The means are not significantly different.

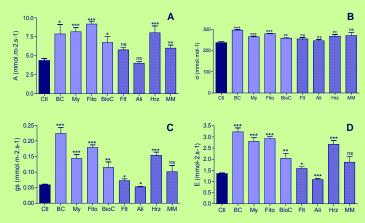
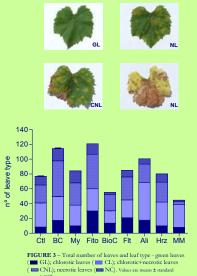


FIGURE 2 – A: Net photosynthesis (A); B: Intercellular concentration of CO₂ (ci); C: Stomatal conductance (gs); D: Transpiration (E): Significant differences between Cd and all the testiments at P < 0.05 are marked by asterisk (ns - not significant; * - significant, ** - very instances of the property of the prope



Ctl Filo BC MM Hrz

FIGURE 4 - One year old potted grapevines (stems and correspondent roots)

MAIN CONCLUSIONS

- ▶ The results of photosynthetic pigments together with the ones of gas exchange suggest a positive effect of BCAs comparatively to CQs. That is particularly noticed for the A values, which were significantly higher in all grapevines treated with BCAs. However the results are not statistically significative.
- ▶ In respect to vegetative parameters, the results suggest that in grapevines treated with BCAs, there is an increase in the total number of leaves, as well as an increase in stem and roots length, comparatively to CQs treated plants. Still, the means are not significantly different.
- ▶ In spite of the erratic results from the first year of experiments (eventually due to the fact that the plants had just been established) the trials will proceed for a 2nd and a 3rd year (with more grown plants) in order to have conclusive results

REFERENCES

LEON & JOYCE (2004) Postharvest Biology and Tecnology. 32: 1-13.

LICHTENTHALER, H. K. (1987) Methos of Enzymology. Academic Press. New York.

FLEXAS, J.; ESCALONA, J. M.; MEDRANO, H. (1998) Australian Journal of Plant Physiology. 25: 893-900.

ZAHAVI, T.; COHEN, L.; WEISS, B.; SCHENA, L.; DAUS, A.; KAPLUNOV, T.; JOHANAN, Z.; BEMARIE, R.; DROBY, S. (2000) Postharvest Biology and Tecnology. 20: 115-124.

ACKNOWLEDGMENT

Eng. Gisela Chicau – D.R.A.P-Norte – for all the plant material.
SAPEC, BAYER, FUTURECO and NUTRIFIELD companies for providing products used for the study.