

UvA-DARE (Digital Academic Repository)

Direct cardiac and endothelial effects of sodium/glucose cotransporter 2 inhibitors and the role of the sodium/hydrogen exchanger 1

Uthman, L.

Publication date 2020 Document Version Other version License Other

Link to publication

Citation for published version (APA):

Uthman, L. (2020). Direct cardiac and endothelial effects of sodium/glucose cotransporter 2 inhibitors and the role of the sodium/hydrogen exchanger 1. [Thesis, fully internal, Universiteit van Amsterdam].

General rights

It is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), other than for strictly personal, individual use, unless the work is under an open content license (like Creative Commons).

Disclaimer/Complaints regulations

If you believe that digital publication of certain material infringes any of your rights or (privacy) interests, please let the Library know, stating your reasons. In case of a legitimate complaint, the Library will make the material inaccessible and/or remove it from the website. Please Ask the Library: https://uba.uva.nl/en/contact, or a letter to: Library of the University of Amsterdam, Secretariat, Singel 425, 1012 WP Amsterdam, The Netherlands. You will be contacted as soon as possible.



General introduction

Laween Uthman

DIABETES AND CARDIOVASCULAR DISEASE

Type 2 diabetes mellitus (T2DM) is a widespread welfare disease and is accountable for a large proportion of all deaths worldwide. T2DM is typically described as insulin resistance, leading to the reduced ability to control plasma glucose levels. A major morbidity burden that patients with T2DM have is heart failure ^{1,2}, which is characterized by the inadequacy of the heart to pump and/or fill with blood. The cardiac pathologies in heart failure and T2DM have several overlapping metabolic, structural and functional features relevant for the heart ³. First, impaired cardiac pump (contraction/relaxation) function, mitochondrial function and vascular function are considered as key hallmarks of T2DM and heart failure ^{8,9}. Second, the structural alterations of the failing and diabetic heart include cardiac stiffness, hypertrophy, fibrosis, cell death and chronic low-grade inflammation 6-8. Third, metabolic systemic changes covered in both heart failure and T2DM are hyperglycemia, hyperinsulinemia and increased circulating free fatty acids ^{4,5}. However, the metabolic changes in the myocardium are opposing for heart failure and T2DM. Heart failure signifies by increased glucose oxidation and reduced fatty acid oxidation, while glucose oxidation is decreased and fatty acid oxidation is elevated in T2DM. Since patients with T2DM and heart failure have a severe and worsened prognosis, it is important to understand which contributing factors could be targeted to reduce the pathologies of both diseases.

SGLT2 inhibitors

The sodium/glucose cotransporter 2 (SGLT2) inhibitors are a class of agents currently implemented as second line therapy in the treatment of T2DM. SGLT2 inhibitors, among which empagliflozin, dapagliflozin and canagliflozin figure as the more prominent members, have exhibited marked reductions in heart failure related hospitalization and cardiovascular death in patients with T2DM ¹⁰⁻¹². SGLT2 inhibitors are designed to inhibit the kidney specific SGLT2, which is at best responsible for 90% of the glucose reabsorption from the kidney filtrate. However, it has become apparent that the cardiovascular benefits of SGLT2 inhibitors cannot be explained through the sole palliating of glycaemia. The possibility of direct cardiovascular effects of SGLT2 inhibitors, regardless of renal SGLT2 inhibitors could target mechanisms underlying the pathogenesis of T2DM and heart failure, including disturbed cardiac sodium/calcium balance via activation of the sodium hydrogen exchanger 1 (NHE1),

chronic low-grade inflammation and metabolic irregularities ¹³. These pathologic mechanisms are separately described below.

Cardiac sodium/calcium balance and NHE1 activity

Sodium (Na*) is an element that is essential for maintenance of membrane potential in excitable cells, intracellular acid-base balance, fluid volume homeostasis and osmotic regulation between cells and extracellular fluid. In the heart, sodium is an important regulator of cardiac cell contraction, by facilitating the transport of numerous solutes and ions across the cell membrane during an action potential, including calcium (Ca²⁺) and potassium (K⁺). This transport is fueled by the potential energy that arises from the influx of Na⁺ that is favored by the steady-state electrochemical gradient in a healthy cardiomyocyte ¹⁶. Cardiac contractility highly depends on the cytoplasmatic Ca²⁺ concentration ([Ca²⁺],). [Na⁺], positively regulates [Ca²⁺], via activation of the sodium/calcium exchanger (NCX). The NCX (transmembrane and mitochondrial) exchanges one [Ca²⁺], with three extracellular/mitochondrial Na⁺ in its forwards mode to regain low [Ca²⁺], after a contraction. However, in cases where [Na⁺], is elevated (figure 1) ¹⁷⁻¹⁹, NCX becomes less active in forward mode and may even operate in a reversed mode, thereby increasing [Ca²⁺], The elevation in [Ca²⁺], includes systolic and diastolic $[Ca^{2+}]_{,}$ which means that both the contraction and relaxation status of the heart become disturbed. The depolarizing current of Ca²⁺ release via the NCX can generate delayed afterdepolarizations and arrhythmias ^{20,21}. In addition, increased [Na⁺], stimulates the lowering of mitochondrial calcium ([Ca²⁺]_m) through the mitochondrial NCX, possibly leading to decreased mitochondrial energy production and decreased activity of the mitochondrial anti-oxidant system, thereby increasing the net reactive oxygen species (ROS) balance. Thus, by controlling the $[Ca^{2+}]_{m'}$ [Na⁺]_c creates a connection between cellular ion homeostasis and metabolism ^{22,23}. Collectively, the alterations in $[Ca^{2+}]_r$ and $[Ca^{2+}]_m$ can cause contractile impairments and mitochondrial dysfunction and may ultimately result in (ir)reversible myocardial injury.

Chapter 1



Figure 1 Consequences of increased cytoplasmatic sodium ([Na⁺]_c) in the cardiac cell. Increased [Na⁺]_c is a prerequisite for elevated $[Ca^{2+}]_c$ and reduced $[Ca^{2+}]_m$, resulting in contractile impairments, energy deficits and enhanced ROS formation. Elevated cardiac $[Na^+]_c$ is seen in T2DM and several cardiac diseases, including heart failure, ischemia reperfusion injury and cardiac arrhythmias. The red marks in the figure show the actions that occur in heart failure and diabetes. NHE Na⁺/H⁺ exchanger, NCX Na⁺/Ca²⁺ exchanger, Na⁺/K⁺ pump, ATP adenosine triphosphate, ROS reactive oxygen species, DAD delayed afterdepolarizations. Image was made using Servier Medical Art (http://smart.servier.com/).

Cardiac [Na⁺]_c is tightly controlled by membrane transporters, pumps and exchangers ¹⁶. One of those transporters is the sodium/hydrogen exchanger 1 (NHE1). The NHE1 is a ubiquitously expressed membrane protein that enables the exchange of one extracellular Na⁺ ion for one intracellular H⁺ ion, thereby counteracting intracellular acidification. The activation of NHE1 will result in the increase of [Na⁺]_c. NHE1 is the major NHE isoform that is expressed in the heart. In healthy conditions, the NHE is minimally active and intracellular pH is primarily regulated via the sodium/bicarbonate cotransporters. However, cardiac NHE1 becomes stimulated during pathological conditions such as T2DM, heart failure and ischemia-reperfusion injury ^{14,24,25}. Moreover, NHE1 gets activated by inflammatory and hyperglycemic insults in vascular endothelial cells ^{26,27}. While the role of increased [Na⁺]_c in the pathogenesis of endothelial dysfunction is not fully understood, inhibition of NHE1 has been shown to block or reverse the impairment attained in immunostimulated or diabetic endothelial cells ^{26,27,29-33}.



Figure 2 Increased circulatory stimuli in T2DM and heart failure cause endothelial dysfunction via inflammation. In T2DM and heart failure, high levels of plasma glucose, fatty acids and pro-inflammatory molecules stimulate inflammatory signaling in endothelial cells. This will result in reduced NO bioavailability, increased adhesion of circulating leukocytes and the remodeling of cellcell junctions causing enhanced endothelial cell permeability. The endothelial dysfunction caused by these processes gives rise to impaired vasodilation, atherosclerotic plaque formation and disrupted endothelial barrier function. LPS lipopolysaccharide, FFA free fatty acids, NO nitric oxide, ROS reactive oxygen species. Image was made using Servier Medical Art (http://smart.servier.com/).

Vascular inflammation and endothelial dysfunction

Inflammation is a hallmark of T2DM and heart failure appearing in cardiac cells, including endothelial cells ³⁴⁻³⁶. The pathogenesis of inflammation in endothelial cells initiates when an extracellular toxin activates pro-inflammatory signaling cascades, leading to endothelial dysfunction (figure 2). Endothelial cells increase their release of pro-inflammatory cytokines, which enhances the expression of leukocyte attracting adhesion molecules and endothelial cell permeability. These actions aggravate the endothelial barrier function and therefore facilitate leukocyte translocation through the vessel wall. There, the leukocytes differentiate into foam cells and form atherosclerotic plaques, induce fibrosis and further stimulate endothelial dysfunction. Endothelial dysfunction also describes a process of impaired vasodilation capacity through reduced nitric oxide (NO) production or availability⁷. A reduction of NO happens when the activity of the enzyme endothelial nitric oxide synthase (eNOS) is hindered, leading to reduced

production of NO. In addition, increased levels of oxidative stress could directly and indirectly reduce NO. First, ROS can react directly with NO resulting in the production of peroxynitrite. Second, ROS can indirectly lower NO when ROS uncouples eNOS during its dimerization process, turning the monomer eNOS into a ROS producing unit, thereby further deteriorating endothelial function. Targeting endothelial inflammation in T2DM and heart failure has been extensively investigated and proposed as a promising strategy to attenuate disease development and progression in T2DM and heart failure ^{37,38}.

Metabolic disturbances

While the healthy heart preferably consumes free fatty acids (FFA, \sim 60%) as opposed to glucose and other carbohydrates (~20-30%), it is an organ with high metabolic flexibility depending on the availability of substrates ^{39,40}. In T2DM, the combination of increased FFA and insulin resistance lowers glucose utilization and the heart will predominantly depend on FFA metabolism ⁴¹. In heart failure, an opposite shift occurs, in that now the heart increases its preference for glucose, which is often associated with a buildup of metabolic glucose intermediates in the cytoplasm. The accumulation of glycolytic metabolic intermediates, such as glucose-6-phosphate (G6P) or increased channeling of glucose into aspartate can drive hypertrophy and consequently the development of heart failure ⁴²⁻⁴⁴. In addition, insulin resistance is associated with an increased risk for heart failure even in the absence of T2DM ⁴⁵. In both conditions of T2DM and heart failure, cardiac substrate utilization is insufficient, leading to the reduced production of adenosine triphosphate (ATP) while simultaneously increasing adenosine diphosphate (ADP) and adenosine monophosphate (AMP) ⁴⁶⁻⁴⁸. Increased levels AMP is sensed by adenosine monophosphate-activated protein kinase (AMPK). AMPK activation improves cardiac energy metabolism, enhances vascular function and reduces levels of oxidative stress ^{49–51}. Therefore, activation of AMPK could be a potential approach to reverse the consequences of the metabolic disarrangements employed in T2DM and heart failure. The metabolic alterations observed in T2DM and heart failure described here are summarized in figure 3.

Aim of this thesis

The general aim of this thesis is to investigate the direct cardiovascular actions of SGLT2 inhibitors in healthy and disease conditions. In **chapter 2**, an extensive literature review of the direct cellular effects of SGLT2 inhibitors is given. In **chapter 3**, we investigated the effects of SGLT2 inhibitors empagliflozin, canagliflozin and dapagliflozin in the

healthy mouse cardiomyocyte on $[Na^+]_c$ and NHE1 activity, and assessed the acute effects of the SGLT2 inhibitors on the physiology of the isolated healthy mouse heart. We also explored the ability of empagliflozin, canagliflozin and dapagliflozin to bind to NHE1 in a computational study. In **chapter 4**, we studied whether acute treatment with empagliflozin can reduce ischemia reperfusion injury in the isolated mouse heart in the presence and absence of insulin, and in relation to NHE inhibition. In **chapter 5**, we studied the direct and acute effects of empagliflozin on cardiac glucose and fatty acid metabolism in type 2 diabetic mouse hearts. In **chapter 6**, we investigated whether empagliflozin is able to reduce $[Na^+]_c$ and NHE1 activity in human endothelial cells. Finally, we explored possible anti-inflammatory effects of empagliflozin and dapagliflozin in tumor necrosis factor α -stimulated endothelial cells (**chapter 7**) and of canagliflozin, empagliflozin and dapagliflozin in lipopolysaccharide-stimulated endothelial cells (**chapter 8**).





REFERENCES

- Cavender, M. A. *et al.* Impact of Diabetes Mellitus on Hospitalization for Heart Failure, Cardiovascular Events, and Death: Outcomes at 4 Years from the Reduction of Atherothrombosis for Continued Health (REACH) Registry. *Circulation* 132, 923–931 (2015).
- 2. Kannel, W. B., Hjortland, M. & Castelli, W. P. Role of Diabetes in Congestive Heart Failure: The Framingham Study. *Am. J. Cardiol.* **34**, 29–34 (1974).
- 3. Maack, C. *et al.* Heart failure and diabetes: Metabolic alterations and therapeutic interventions: A state-of-The-Art review from the Translational Research Committee of the Heart Failure Association-European Society of Cardiology. *Eur. Heart J.* **39**, 4243–4254 (2018).
- Amaral, N. & Okonko, D. O. Metabolic abnormalities of the heart in type II diabetes. Diabetes Vasc. Dis. Res. 12, 239–248 (2015).
- 5. Jia, G., Hill, M. A. & Sowers, J. R. Diabetic cardiomyopathy: An update of mechanisms contributing to this clinical entity. *Circ. Res.* **122**, 624–638 (2018).
- Hutchinson, K. R., Lord, C. K., West, T. A. & Stewart, J. A. Cardiac Fibroblast-Dependent Extracellular Matrix Accumulation Is Associated with Diastolic Stiffness in Type 2 Diabetes. *PLoS One* 8, (2013).
- 7. Paulus, W. J. & Tschöpe, C. A novel paradigm for heart failure with preserved ejection fraction: Comorbidities drive myocardial dysfunction and remodeling through coronary microvascular endothelial inflammation. *J. Am. Coll. Cardiol.* **62**, 263–271 (2013).
- Bugger, H. & Abel, E. D. Molecular mechanisms of diabetic cardiomyopathy. *Diabetologia* 57, 660–671 (2014).
- 9. Bugger, H. & Abel, E. D. Mitochondria in the diabetic heart. Cardiovasc. Res. 88, 229–240 (2010).
- 10. Zinman, B. *et al.* Empagliflozin, cardiovascular outcomes, and mortality in type 2 diabetes. *N. Engl. J. Med.* **373**, 2117–2128 (2015).
- Neal, B. *et al.* Canagliflozin and cardiovascular and renal events in type 2 diabetes. *N. Engl. J. Med.* 377, 644–657 (2017).
- Wiviott, S. D. *et al.* Dapagliflozin and cardiovascular outcomes in type 2 diabetes. *N. Engl. J. Med.* 380, 347–357 (2019).
- Uthman, L. *et al.* Direct cardiac actions of sodium glucose cotransporter 2 inhibitors target pathogenic mechanisms underlying heart hailure in diabetic patients. *Front. Physiol.* 9, 1575 (2018).
- Packer, M. Activation and Inhibition of Sodium-Hydrogen Exchanger Is a Mechanism That Links the Pathophysiology and Treatment of Diabetes Mellitus with That of Heart Failure. *Circulation* 136, 1548–1559 (2017).
- Packer, M. Autophagy stimulation and intracellular sodium reduction as mediators of the cardioprotective effect of sodium–glucose cotransporter 2 inhibitors. *Eur. J. Heart Fail.* ejhf.1732 (2020). doi:10.1002/ejhf.1732

- Bers, D. M., Barry, W. H. & Despa, S. Intracellular Na+ regulation in cardiac myocytes. *Cardiovasc. Res.* 57, 897–912 (2003).
- Despa, S., Islam, M. A., Weber, C. R., Pogwizd, S. M. & Bers, D. M. Intracellular Na+ concentration is elevated in heart failure but Na/K pump function is unchanged. *Circulation* 105, 2543–2548 (2002).
- **18**. Lambert, R. *et al.* Intracellular Na+concentration ([Na+]i) is elevated in diabetic hearts due to enhanced Na+-glucose cotransport. *J. Am. Heart Assoc.* **4**, 1–11 (2015).
- **19**. Despa, S. Myocyte [Na+]i Dysregulation in Heart Failure and Diabetic Cardiomyopathy. *Front. Physiol.* **9**, 1303 (2018).
- Bertero, E. & Maack, C. Calcium Signaling and Reactive Oxygen Species in Mitochondria. *Circ. Res.* 122, 1460–1478 (2018).
- Baartscheer, A., Schumacher, C. a, Belterman, C. N. W., Coronel, R. & Fiolet, J. W. T. S R calcium handling and calcium after-transients in a rabbit model of heart failure. *Heart Fail.* 58, 99–108 (2003).
- Bay, J., Kohlhaas, M. & Maack, C. Intracellular Na+ and cardiac metabolism. J. Mol. Cell. Cardiol. 61, 20–27 (2013).
- 23. Sedova, M. & Blatter, L. A. Intracellular sodium modulates mitochondrial calcium signaling in vascular endothelial cells. *J. Biol. Chem.* **275**, 35402–35407 (2000).
- 24. Darmellah, A. *et al.* Enhanced activity of the myocardial Na+/H+ exchanger contributes to left ventricular hypertrophy in the Goto-Kakizaki rat model of type 2 diabetes: Critical role of Akt. *Diabetologia* **50**, 1335–1344 (2007).
- 25. Karmazyn, M. The role of the myocardial sodium-hydrogen exchanger in mediating ischemic and reperfusion injury. *Ann. N. Y. Acad. Sci.* **874**, 326–334 (1999).
- Wang, S., Peng, Q., Zhang, J. & Liu, L. Na+/H+exchanger is required for hyperglycaemiainduced endothelial dysfunction via calcium-dependent calpain. *Cardiovasc. Res.* 80, 255–262 (2008).
- Cui, G. M. *et al.* Amiloride attenuates lipopolysaccharide-accelerated atherosclerosis via inhibition of NHE1-dependent endothelial cell apoptosis. *Acta Pharmacol. Sin.* 34, 231–238 (2013).
- Németh, Z. H., Deitch, E. A., Lu, Q., Szabó, C. & Haskó, G. NHE blockade inhibits chemokine production and NF-κB activation in immunostimulated endothelial cells. *Am. J. Physiol. -Cell Physiol.* 283, 396–403 (2002).
- Baartscheer, A. *et al.* Chronic inhibition of the Na+/H+- exchanger causes regression of hypertrophy, heart failure, and ionic and electrophysiological remodelling. *Br. J. Pharmacol.* **154**, 1266–1275 (2008).
- 30. Karmazyn, M. NHE-1: Still a viable therapeutic target. J. Mol. Cell. Cardiol. 61, 77-82 (2013).
- **31.** Engelhardt, S. Inhibition of Na+-H+ Exchange Prevents Hypertrophy, Fibrosis, and Heart Failure in beta1-Adrenergic Receptor Transgenic Mice. *Circ. Res.* **90**, 814–819 (2002).

- **32**. Mentzer, R. M. *et al.* Sodium-Hydrogen Exchange Inhibition by Cariporide to Reduce the Risk of Ischemic Cardiac Events in Patients Undergoing Coronary Artery Bypass Grafting: Results of the EXPEDITION Study. *Ann. Thorac. Surg.* **85**, 1261–1270 (2008).
- Vial, G. *et al.* Na+/H+ exchange inhibition with cariporide prevents alterations of coronary endothelial function in streptozotocin-induced diabetes. *Mol. Cell. Biochem.* 310, 93–102 (2008).
- **34**. Sharma, A. *et al.* Oxidative stress and NLRP3-inflammasome activity as significant drivers of diabetic cardiovascular complications: Therapeutic implications. *Front. Physiol.* **9**, 114 (2018).
- **35.** Odegaard, A. O. *et al.* Oxidative stress, inflammation, endothelial dysfunction and incidence of type 2 diabetes. *Cardiovasc. Diabetol.* **15**, 1–12 (2016).
- Natali, A., Toschi, E., Baldeweg, S., Ciociaro, D. & Favilla, S. Clustering of Insulin Resistance With Vascular Dysfunction and Low-Grade Inflammation in Type 2 Diabetes. *Diabetes* 55, 1133–1140 (2006).
- **37.** Donath, M. Y. Targeting inflammation in the treatment of type 2 diabetes: Time to start. *Nat. Rev. Drug Discov.* **13**, 465–476 (2014).
- 38. Pollack, R. M., Donath, M. Y., LeRoith, D. & Leibowitz, G. Anti-inflammatory agents in the treatment of diabetes and its vascular complications. *Diabetes Care* **39**, S244–S252 (2016).
- **39.** Bing, R. J., Siegel, A., Ungar, I. & Gilbert, M. Metabolism of the human heart. II. Studies on fat, ketone and amino acid metabolism. *Am. J. Med.* **16**, 504–515 (1954).
- Khairallah, M. *et al.* Profiling substrate fluxes in the isolated working mouse heart using 13C-labeled substrates: Focusing on the origin and fate of pyruvate and citrate carbons. *Am. J. Physiol. Hear. Circ. Physiol.* 286, H1461–H1470 (2004).
- Varma, U., Koutsifeli, P., Benson, V. L., Mellor, K. M. & Delbridge, L. M. D. Molecular mechanisms of cardiac pathology in diabetes – Experimental insights. *Biochim. Biophys. Acta - Mol. Basis Dis.* 1864, 1949–1959 (2018).
- 42. Bertero, E. & Maack, C. Metabolic remodelling in heart failure. *Nat. Rev. Cardiol.* **15**, 457–470 (2018).
- **43.** Ritterhoff, J. *et al.* Metabolic remodeling promotes cardiac hypertrophy by directing glucose to aspartate biosynthesis. *Circ. Res.* 182–196 (2020). doi:10.1161/CIRCRESAHA.119.315483
- Karlstaedt, A., Khanna, R., Thangam, M. & Taegtmeyer, H. Glucose 6-Phosphate Accumulates via Phosphoglucose Isomerase Inhibition in Heart Muscle. *Circ. Res.* 60–74 (2020). doi:10.1161/CIRCRESAHA.119.315180
- **45**. Vardeny, O. *et al.* Insulin Resistance and Incident Heart Failure. The ARIC Study (Atherosclerosis Risk in Communities). *JACC Hear. Fail.* **1**, 531–536 (2013).
- **46.** Levelt, E. *et al.* Cardiac energetics, oxygenation, and perfusion during increased workload in patients with type 2 diabetes mellitus. *Eur. Heart J.* **37**, 3461–3469 (2016).
- 47. Neubauer, S. The Failing Heart An Engine Out of Fuel. N. Engl. J. Med. 356, 1140–1151 (2007).

- **48**. Ingwall, J. S. Energy metabolism in heart failure and remodelling. *Cardiovasc. Res.* **81**, 412–419 (2009).
- **49**. Li, J. *et al.* Pharmacological activation of AMPK prevents Drp1-mediated mitochondrial fission and alleviates endoplasmic reticulum stress-associated endothelial dysfunction. *J. Mol. Cell. Cardiol.* **86**, 62–74 (2015).
- **50.** He, C., Li, H., Viollet, B., Zou, M. H. & Xie, Z. AMPK suppresses vascular inflammation in vivo by inhibiting signal transducer and activator of transcription-1. *Diabetes* **64**, 4285–4297 (2015).
- **51.** Gélinas, R. *et al.* AMPK activation counteracts cardiac hypertrophy by reducing O-GlcNAcylation. *Nat. Commun.* **9**, 374 (2018).