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Deng, S.; de Hoog, G.S.; Badali, H.; Yang, L.; Najafzadeh, M.J.; Pan, B.; Curfs-Breuker, I.; Meis, J.F.; Liao, W. DOI 10.1128/AAC.02114-12

Publication date 2013 **Document Version** Final published version Published in

Antimicrobial Agents and Chemotherapy

Link to publication

Citation for published version (APA): Deng, S., de Hoog, G. S., Badali, H., Yang, L., Najafzadeh, M. J., Pan, B., Curfs-Breuker, I., Meis, J. F., & Liao, W. (2013). In vitro antifungal susceptibility of Cladophialophora carrionii, agent of human chromoblastomycosis. Antimicrobial Agents and Chemotherapy, 57(4), 1974-1977. https://doi.org/10.1128/AAC.02114-12

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In Vitro Antifungal Susceptibility of Cladophialophora carrionii, an Agent of Human Chromoblastomycosis

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A global collection of *Cladophialophora carrionii* strains (n = 81) was tested against nine antifungal drugs. MIC₉₀s of all strains were as follows in increasing order: itraconazole and posaconazole, 0.063 µg/ml; terbinafine, 0.125 µg/ml; isavu-conazole and voriconazole, 0.25 µg/ml; caspofungin, 2 µg/ml; micafungin, 4 µg/ml; amphotericin B, 8 µg/ml; and fluconazole, 64 µg/ml.

Chromoblastomycosis is a chronic, progressive, polymorphic implantation mycosis. Lesions are limited to cutaneous and subcutaneous tissues, causing hyperproliferation leading to verrucous or nodular clinical features (1–3). Two genera of melanized hyphomycetes, *Cladophialophora* and *Fonsecaea*, both belonging to the family *Herpotrichiellaceae* in the order *Chaetothyriales*, are common causes. They have in common that a pathogenic invasive phase is formed in skin with the expression of muriform cells. Occasional cases have been reported due to species of *Phialophora, Exophiala*, and *Rhinocladiella*, which also belong to this family (4). The disease is encountered worldwide in subtropical and tropical climate zones, with a clear distinction between the vicarious species of *Cladophialophora* in arid climates and *Fonsecaea* and *Rhinocladiella* in humid, tropical climates (5).

Cladophialophora carrionii is a relatively frequent etiologic agent of chromoblastomycosis in arid and semiarid climate zones of South and Central America (6, 7), Australia (8), and Asia (9, 10). The infection is very difficult to treat. Several therapies have been applied, but there is no standard for treatment (3). Small series of *in vitro* susceptibility studies with itraconazole, voriconazole, and terbinafine have been published showing considerable variation between and within genera and species (11, 12).

The aim of the present study was to determine the susceptibility profiles of a large collection of C. carrionii strains to nine antifungal agents, including isavuconazole (13). Isolates were taken from the reference collections of the CBS-KNAW Fungal Biodiversity Centre (CBS, Utrecht, The Netherlands) or the Institute Pasteur (CNRMA/IP, Paris, France). The set comprised isolates from Venezuela (n = 46), China (n = 20), Madagascar (n = 9), and Australia (n = 6). Seventy-five clinical isolates originated from patients with chromoblastomycosis, and six environmental isolates were from dry plant debris in Venezuela (Table 1). All strains were identified to the species level by sequencing of the internal transcribed spacer of the ribosomal DNA (rDNA) region and partial translation of the elongation factor 1- α and β -tubulin genes (S. Deng, A. H. G. Gerrits van den Ende, L. Yang, H. Badali, M. J. Najafzadeh, R. Y. Li, C. H. Klaassen, F. Hagen, J. F. Meis, B. Papierok, J. Sun, W. D. Liu, G. S. De Hoog, submitted for publication). In vitro activities of nine antifungal agents were determined with the reference guideline M38-A2 (14). Three reference strains, *Paecilomyces variotii* (ATCC 22319), *Candida parapsilosis* (ATCC 22019), and *Candida krusei* (ATCC 6258) were included as quality controls. Kruskal-Wallis and Mann-Whitney U tests were used for comparison of the MICs of all antifungal agents among strains from four groups (Latin America, Asia, Africa, and Australia).

Table 2 summarizes the MIC results in terms of the MIC ranges, geometric mean (GM) MIC, and MIC₅₀ and MIC₉₀ values of nine antifungal agents for 81 C. carrionii strains. All strains had low MICs of itraconazole, voriconazole, posaconazole, isavuconazole, and terbinafine, while the highest MICs were consistently found with fluconazole, amphotericin B, micafungin, and caspofungin. The MIC₉₀s of fluconazole, amphotericin B, micafungin, and caspofungin were 64 μ g/ml, 8 μ g/ml, 4 μ g/ml, and 2 μ g/ml, respectively. These data are in agreement with previously reported findings for Cladophialophora (11, 15), Rhinocladiella (16), and Fonsecaea (17). No difference was found in the activities between voriconazole and isavuconazole against C. carrionii (MIC range, 0.016 to 1 μg/ml; GM, 0.148/0.136 μg/ml; MIC₉₀, 0.25 μg/ml). The MIC range and MIC₉₀ of voriconazole were 2 log₂-dilution steps more active than values found in C. bantiana (range, 0.125 to 4 µg/ml; MIC₉₀, 2 µg/ml) (15) and in Phialophora and Cyphello*phora* (MIC range, 0.125 to 4 μg/ml; MIC₉₀, 1 μg/ml) (18). Table 3 shows rare *Cladophialophora* species causing (sub)cutaneous disorders but which are related to Fonsecaea (19) and to C. yegresii, an environmental sibling of C. carrionii. The values were in the same range, with the exception of lower MICs of caspofungin and micafungin in the cutaneous species C. immunda and C. saturnica and of voriconazole in C. yegresii and C. samoensis.

Received 21 October 2012 Returned for modification 29 November 2012 Accepted 15 December 2012

Published ahead of print 4 February 2013

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TABLE 1 Cladophialophora strains used in this study

TABLE 2 MIC values of nine antifungal agents against 81 C. carrionii strains

pecies ladophialophora carrionii	Accession no. CBS 108.97 CBS 109.97 CBS 164.54 CBS 165.54	Source Chromoblastomycosis, male Chromoblastomycosis, male	Origin Venezuela		MIC (µg/1	ml) ^a				
	CBS 164.54		3.7 1		MIC $(\mu g/ml)^a$					
			Venezuela	Sturin (a) and down						
		Chromoblastomycosis, male	Venezuela	Strain (n) and drug	GM	Range	50%	909		
	CBS 165.54	Chromoblastomycosis, male	Venezuela	All C. carrionii strains (81)						
	CBS 166.54 CBS 986.96	Chromoblastomycosis, male Clinical material	Venezuela Venezuela	Amphotericin B	2.643	0.5-8	2	8		
	CBS 857.96	Chromoblastomycosis, male	Venezuela	Fluconazole Itraconazole	25.04	4-64 0.008-0.125	32	64		
	CBS 858.96	Chromoblastomycosis, male	Venezuela	Voriconazole	0.03 0.148	0.008-0.125	0.031 0.125	0.0 0.2		
	CBS 114392	Chromoblastomycosis, female	Venezuela	Posaconazole	0.025	0.016-0.063	0.016	0.2		
	CBS 114393	Chromoblastomycosis, male	Venezuela	Isavuconazole	0.136	0.016-1	0.125	0.2		
	CBS 114394	Chromoblastomycosis, male	Venezuela	Caspofungin	1.367	0.25-4	2	2		
	CBS 114395	Chromoblastomycosis, female	Venezuela	Micafungin	0.296	0.016-8	0.25	4		
	CBS 114397	Chromoblastomycosis, male	Venezuela	Terbinafine	0.049	0.008 - 1	0.031	0.1		
	CBS 114398	Chromoblastomycosis, female	Venezuela							
	CBS 114399 CBS 114400	Chromoblastomycosis, female	Venezuela Venezuela	C. carrionii, Venezuela (46)						
	CBS 114400 CBS 114401	Chromoblastomycosis, male Chromoblastomycosis, female	Venezuela	Amphotericin B	2.767	0.5-8	2	8		
	CBS 114401 CBS 114402	Chromoblastomycosis, female	Venezuela	Fluconazole	31.07	8-64	32	64		
	CBS 114403	Chromoblastomycosis, male	Venezuela	Itraconazole	0.038	0.016-0.125	0.031	0.		
	CBS 114404	Chromoblastomycosis, male	Venezuela	Voriconazole Posaconazole	0.181 0.029	0.031-1	0.125 0.031	0.0		
	CBS 117889	Chromoblastomycosis, female	Venezuela	Isavuconazole	0.168	0.016-0.063 0.016-1	0.031	0.0		
	CBS 117890	Chromoblastomycosis, male	Venezuela	Caspofungin	1.363	0.25-4	1	2		
	CBS 117891	Chromoblastomycosis, male	Venezuela	Micafungin	0.206	0.016-8	0.25	0.		
	CBS 117892	Chromoblastomycosis, male	Venezuela	Terbinafine	0.053	0.016-1	0.031	0.		
	CBS 117893	Chromoblastomycosis, male	Venezuela							
	CBS 117895	Chromoblastomycosis, male	Venezuela	C. carrionii, China (20)						
	CBS 117896	Chromoblastomycosis, male	Venezuela	Amphotericin B	2.639	0.5-8	4	8		
	CBS 117897	Chromoblastomycosis, male	Venezuela	Fluconazole	19.027	8-32	16	32		
	CBS 117898 CBS 117899	Chromoblastomycosis, female	Venezuela Venezuela	Itraconazole	0.022	0.016-0.063	0.016	0.		
	CBS 117899 CBS 117900	Chromoblastomycosis, male Chromoblastomycosis, male	Venezuela	Voriconazole	0.109	0.016-0.5	0.125	0.		
	CBS 117900 CBS 117901	Chromoblastomycosis, female	Venezuela	Posaconazole	0.021	0.016-0.063	0.016	0.		
	CBS 117902	Chromoblastomycosis, male	Venezuela	Isavuconazole	0.092	0.016-0.25	0.125	0.		
	CBS 117903	Chromoblastomycosis, male	Venezuela	Caspofungin	1.625	0.25-4	2	2		
	CBS 117904	Chromoblastomycosis, male	Venezuela	Micafungin	0.342	0.063-4	0.25	1		
	CBS 117905	Chromoblastomycosis, male	Venezuela	Terbinafine	0.037	0.008-0.125	0.031	0.		
	CBS 117906	Chromoblastomycosis, male	Venezuela	Commissiii Madaaaaa (0)						
	CBS 117908	Chromoblastomycosis, male	Venezuela	C. carrionii, Madagascar (9) Amphotericin B	3.175	1-8	4	4		
	CBS 117909	Chromoblastomycosis, male	Venezuela	Fluconazole	18.664	4-64	4			
	CBS 121844	Chromoblastomycosis, male	Venezuela	Itraconazole	0.023	0.016-0.125	0.016	0.		
	CBS 859.96	Dry plant debris	Venezuela	Voriconazole	0.116	0.016-0.5	0.125	0.		
	CBS 860.96	Dry plant debris	Venezuela	Posaconazole	0.02	0.016-0.063	0.016	0.		
	CBS 861.96	Dry plant debris	Venezuela	Isavuconazole	0.107	0.031-0.5	0.063	0.		
	CBS 862.96	Dry plant debris	Venezuela	Caspofungin	1.361	0.25-4	1	4		
	CBS 863.96 CBS131736	Dry plant debris Soil	Venezuela Venezuela	Micafungin	1.47	0.125-8	2	4		
	CBS131833	Chromoblastomycosis, male	China	Terbinafine	0.053	0.008-0.125	0.063	0.		
	CBS131834	Chromoblastomycosis, male	China							
	CBS131835	Chromoblastomycosis, male	China	C. carrionii, Australia (6)						
	CBS131836	Chromoblastomycosis, male	China	Amphotericin B	1.414	0.5-4	NC	N		
	CBS131838	Chromoblastomycosis, male	China	Fluconazole	17.96	8-64	NC	N		
	CBS131839	Chromoblastomycosis, male	China	Itraconazole	0.02	0.016-0.063 0.031-0.5	NC NC	N N		
	CBS131840	Chromoblastomycosis, male	China	Voriconazole Posaconazole	0.125 0.022	0.016-0.063	NC	N		
	CBS131841	Chromoblastomycosis, male	China	Isavuconazole	0.022	0.063-0.5	NC	N		
	CBS131842	Chromoblastomycosis, male	China	Caspofungin	0.793	0.5-1	NC	N		
	CBS131843	Chromoblastomycosis, male	China	Micafungin	0.281	0.063-4	NC	N		
	CBS131844	Chromoblastomycosis, male	China	Terbinafine	0.07	0.016-0.25	NC	N		
	CBS131845	Chromoblastomycosis, male	China							
	CB\$131846	Chromoblastomycosis, male	China	^a GM, geometric mean; 50% and 90%, MIC ₅₀ and MIC ₉₀ ,		₀ and MIC ₉₀ , respec	tively; NC, no)		
	CBS131847 CBS131848	Chromoblastomycosis, male Chromoblastomycosis, male	China China	comparison because there v	vere <9 strains po	er species available.				
	CBS131848 CBS131850	Chromoblastomycosis, male	China							
	CBS131851	Chromoblastomycosis, male	China							
	CBS132096	Chromoblastomycosis, male	China							
	CBS132097	Chromoblastomycosis, male	China	The activities of i	traconazole a	and posaconazo	ole against	C		
	CBS132100	Chromoblastomycosis, male	China							
	CBS131854	Chromoblastomycosis	Madagascar	rionii were compara						
	CBS131855	Chromoblastomycosis	Madagascar	ana and of Fonsecaed	a species (15.	17). Phialopha	ora and Ci	vphe		
	CBS131856	Chromoblastomycosis	Madagascar							
	CBS131734	Chromoblastomycosis	Madagascar	phora (18) had respo						
	CBS131735	Chromoblastomycosis	Madagascar	similar to those four	nd in C. carr	<i>ionii</i> , but the it	traconazol	le va		
	CBS131857	Chromoblastomycosis	Madagascar	was different (MIC90	$0.5 \mu\sigma/ml$	Terbinafine va	ried consi	dera		
	CBS 100434	Chromoblastomycosis, male	Madagascar							
	CBS 260.83	Chromoblastomycosis, male	Madagascar	in its activity against						
	CBS 362.70	Human skin, male	Madagascar	1 µg/ml). MIC ranges and MIC ₉₀ s of posaconazole, isavucor						
	CBS 160.54	Chromoblastomycosis, male	Australia							
	CBS 162.54	Chromoblastomycosis, male	Australia	zole, voriconazole, and terbinafine showed potent activity against						
	CBS 163.54 CBS131852	Chromoblastomycosis, male Unknown	Australia	C. carrionii (Table 2). Posaconazole was the drug with the be						
	CB\$131852 CB\$131853	Unknown Unknown	Australia Australia	overall in vitro activ						
	CBS131855 CBS 406.96	Chromoblastomycosis, male	Australia	model of C. carrionia				aiiii		
	CDC 114405	Plant Castages	Vanami				m Venezu	ا دام		
adaphicter	CBS 114405 CBS 114406	Plant, Cactaceae	Venezuela Venezuela	For micafungin, most <i>C. carrionii</i> isolates from Venezuela ha low MICs. The range was 0.016 to 8 μ g/ml, the GM was 0.2						
ladophialophora yegresii	CD3 114400	Plant, Cactaceae	v chezuela	Iow MICs The ran	10 Was 0 016	to 8 µg/ml t	he (M w	as 0		

 μ g/ml, and the MIC₉₀ was 0.5 μ g/ml. Some strains deviated sig-

Drug	MIC $(\mu g/ml)^a$										
	$C. \ carrionii \ (n = 28)$					Range					
	GM	Range	50%	90%	<i>C. samoensis</i> $(n = 1)$	C. yegresii (n = 3)	C. immunda (n = 6)	C. saturnica $(n = 4)$			
Amphotericin B	2.499	0.5-8	2	4	2	0.25-0.5	0.5–4	1–2			
Fluconazole	35.33	16-64	32	64	32	16-32	16-32	8-16			
Itraconazole	0.039	0.016-0.125	0.031	0.063	0.25	0.25-0.5	0.031-0.25	0.031-0.25			
Voriconazole	0.205	0.063-1	0.25	0.5	4	2-2	0.25-1	0.5-1			
Posaconazole	0.033	0.016-0.063	0.031	0.063	0.125	0.125-0.125	0.031-0.063	0.031-0.125			
Isavuconazole	0.2	0.063-1	0.25	0.5	1	0.125-0.5	0.25-0.5	0.25-0.5			
Caspofungin	0.313	0.25-4	1	2	2	1-1	1-2	2-8			
Micafungin	0.906	0.125-4	1	2	0.25	0.25-0.25	4-8	4-8			
Terbinafine	0.05	0.016-0.25	0.063	0.125	ND	0.063-0.063	ND	ND			

TABLE 3 MIC values of nine antifungal agents against *C. carrionii* and rare environmental *Cladophialophora* species eventually causing chromoblastomycosis or other types of skin disease

^{*a*} GM, geometric mean; 50% and 90%, MIC₅₀ and MIC₉₀, respectively; ND, not determined. Note that for *C. immunda* (n = 6) and *C. saturnica* (n = 4), only eight antifungal agents were tested.

nificantly (Table 2), and all nine strains from Madagascar had 3 \log_2 -dilution-step-higher MICs than the majority of Venezuelan strains (range, 0.125 to 8 µg/ml; GM, 1.47 µg/ml; MIC₉₀, 4 µg/ml) (P < 0.01). The activities against Chinese and Australian strains were intermediate. For amphotericin B, the MIC range (0.5 to 8 µg/ml) and MIC₉₀ (8 µg/ml) were much higher than those of *C. bantiana* (MIC range, 0.125 to 2 µg/ml; MIC₉₀, 1 µg/ml) (15) and *Fonsecaea* (MIC range, 0.5 to 2 µg/ml; MIC₉₀, 2 µg/ml) (17) and confirmed the results from a recent study (11).

The 81 investigated isolates of *C. carrionii* represented a worldwide collection from four continents: South America (n = 46), Asia (n = 20), Africa (n = 9), and Australia (n = 6). In a molecular phylogenetic analysis (Deng et al., submitted), three main populations were recognizable: an Asian group, a South American group, and a variable African/Australian group. The susceptibility against itraconazole, voriconazole, posaconazole, isavuconazole for the Latin American group was less than that of remaining groups (P < 0.05), and micafungin was active against most strains from Venezuela (GM, 0.206 µg/ml; MIC₉₀, 0.5 µg/ml), but inactive for strains from Madagascar (GM, 1.47 µg/ml; MIC₉₀, 4 µg/ ml) and some scattered isolates from other continents. There was a significant difference (P < 0.01) in the MICs of micafungin between Madagascar and Venezuelan strains, but the activity of terbinafine among these three groups showed no difference (P > 0.05).

These results suggest that *C. carrionii*, the etiologic agent of chromoblastomycosis in arid climates, is particularly susceptible *in vitro* to the newer azoles and terbinafine, but resistant to amphotericin B, fluconazole, and caspofungin. This profile is similar to that of melanized fungi studied previously (12, 16, 17). The results for micafungin are variable because all strains from Madagascar and some from other continents deviate significantly from the remaining strains. In general, these *in vitro* data still need to be verified by clinical studies.

ACKNOWLEDGMENTS

This study was funded by NSFC grant no. 81060125 from the Natural Science Foundation of China and partially supported by program 973 no. 2013CB531601 and no. 2013CB531606 from the National Basic Research Program, by the Major Infectious Disease Fund (2013ZX10004612) and the Shanghai Science and Technology Commission (no. 10dz2220100), and by an educational grant from Basilea Pharmaceutica International

AG, Basel, Switzerland. J.F.M. received grants from Astellas, Merck, Basilea, and Schering-Plough.

We acknowledge B. Papierok for making strains from Madagascar available.

J.F.M. has been a consultant to Basilea and Merck and received speaker's fees from Merck, Pfizer, Schering-Plough, Gilead, and Janssen Pharmaceutica. All other authors report they have no potential conflicts of interest.

REFERENCES

- Carrión AL. 1950. Chromoblastomycosis. Ann. N. Y. Acad. Sci. 50:1255– 1282.
- 2. McGinnis MR. 1983. Chromoblastomycosis and phaeohyphomycosis: new concepts, diagnosis, and mycology. J. Am. Acad. Dermatol. 8:1–16.
- 3. Queiroz-Telles F, Esterre P, Perez-Blanco M, Vitale RG, Salgado CG, Bonifaz A. 2009. Chromoblastomycosis: an overview of clinical manifestations, diagnosis and treatment. Med. Mycol. 47:3–15.
- Haase G, Sonntag L, Melzer-Krick B, de Hoog GS. 1999. Phylogenetic inference by SSU gene analysis of members of the *Herpotrichiellaceae* with special reference to human pathogenic species. Stud. Mycol. 43:80–97.
- Najafzadeh MJ, Sun J, Vicente VA, Klaassen CH, Bonifaz A, Gerrits van den Ende AHG, Menken SB, de Hoog GS. 2011. Molecular epidemiology of *Fonsecaea* species. Emerg. Infect. Dis. 17:464–469.
- Lavelle P. 1980. Chromoblastomycosis in Mexico. Pan Am. Health Org. Sci. Publ. 396:235–247.
- Silva JP, De Sousa W, Rozental S. 1999. Chromoblastomycosis: a retrospective study of 325 cases on Amazonic Region (Brazil). Mycopathologia 143:171–175.
- 8. Riddley M. 1957. The natural habitat of *Cladosporium carrionii*, a cause of chromoblastomycosis. Aust. J. Dermatol. 4:23–27.
- 9. Li RY, Wang DL. 1996. Studies on phaeohyphomycosis and its causative agents in China. Jpn. J. Med. Mycol. 37:135–141.
- Rajendran C, Ramesh V, Misra RS, Kandhari S, Upreti HB, Datta KK. 1997. Chromoblastomycosis in India. Int. J. Dermatol. 36:29–33.
- González GM, Rojas OC, Bocanegra-García V, González JG, Garza-González E. 27 June 2012. Molecular diversity of *Cladophialophora carrionii* in patients with chromoblastomycosis in Venezuela. Med. Mycol. [Epub ahead of print.] doi:10.3109/13693786.2012.695457.
- 12. Vitale RG, Perez-Blanco M, de Hoog GS. 2009. *In vitro* activity of antifungal drugs against *Cladophialophora* species associated with human chromoblastomycosis. Med. Mycol. 47:35–40.
- 13. Guinea J, Bouza E. 2008. Isavuconazole: a new and promising antifungal triazole for the treatment of invasive fungal infections. Future Microbiol. 3:603–615.
- 14. Clinical and Laboratory Standards Institute. 2008. Reference method for broth dilution antifungal susceptibility testing of filamentous fungi, 2nd ed. Approved standard. CLSI document M38-A2. Clinical and Laboratory Standards Institute, Wayne, PA.

- Badali H, de Hoog GS, Curfs-Breuker I, Klaassen CH, Meis JF. 2010. Use of amplified fragment length polymorphism to identify 42 *Cladophia-lophora* strains related to cerebral phaeohyphomycosis with in vitro antifungal susceptibility. J. Clin. Microbiol. 48:2350–2356.
- Badali H, de Hoog GS, Curfs-Breuker I, Meis JF. 2010. *In vitro* activities of antifungal drugs against *Rhinocladiella mackenziei*, an agent of fatal brain infection. J. Antimicrob. Chemother. 65:175–177.
- Najafzadeh MJ, Badali H, Illnait-Zaragozi MT, de Hoog GS, Meis JF. 2010. In vitro activities of eight antifungal drugs against 55 clinical isolates of Fonsecaea spp. Antimicrob. Agents Chemother. 54:1636– 1638.
- Feng P, Najafzadeh MJ, Sun J, Ahmed S, Xi L, de Hoog GS, Lai W, Lu C, Klaassen CH, Meis JF. 2012. In vitro activities of nine antifungal drugs against 81 *Phialophora* and *Cyphellophora* isolates. Antimicrob. Agents Chemother. 56:6044–6047.
- de Hoog GS, Vicente VA, Najafzadeh MJ, Harrak MJ, Badali H, Seyedmousavi S. 2011. Waterborne *Exophiala* species causing disease in coldblooded animals. Persoonia 27:46–72.
- Calvo E, Pastor FJ, Salas V, Mayayo E, Capilla J, Guarro J. 2012. Histopathology and antifungal treatment of experimental murine chromoblastomycosis caused by *Cladophialophora carrionii*. J. Antimicrob. Chemother. 67:666–670.