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CHAPTER 2

Diagnostic performance of narrowed spectrum endoscopy, autofluorescence imaging, and confocal laser endomicroscopy for optical diagnosis of colonic polyps: a meta-analysis

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ABSTRACT

Background

Novel endoscopic technologies could allow optical diagnosis and resection of colonic polyps without histopathological testing. Our aim was to establish the sensitivity, specificity, and real-time negative predictive value of three types of narrowed spectrum endoscopy (narrow-band imaging [NBI], image-enhanced endoscopy [i-scan], and Fujinon intelligent chromoendoscopy [FICE]), confocal laser endomicroscopy (CLE), and autofluorescence imaging for differentiation between neoplastic and non-neoplastic colonic lesions.

Methods

We identified relevant studies through a search of Medline, Embase, PubMed, and the Cochrane Library. Clinical trials and observational studies were eligible for inclusion when the diagnostic performance of NBI, i-scan, FICE, autofluorescence imaging, or CLE had been assessed for differentiation, with histopathology as the reference standard, and for which a 2 × 2 contingency table of lesion diagnosis could be constructed. We did a random-effects bivariate meta-analysis using a non-linear mixed model approach to calculate summary estimates of sensitivity and specificity, and plotted estimates in a summary receiver-operating characteristic curve.

Findings

We included 91 studies in our analysis: 56 were of NBI, ten of i-scan, 14 of FICE, 11 of CLE, and 11 of autofluorescence imaging (more than one of the investigated modalities assessed in eight studies). For NBI, overall sensitivity was 91.0% (95% CI 88.6–93.0), specificity 85.6% (81.3–89.0), and real-time negative predictive value 82.5% (75.4–87.9). For i-scan, overall sensitivity was 89.3% (83.3–93.3), specificity 88.2% (80.3–93.2), and real-time negative predictive value 86.5% (78.0–92.1). For FICE, overall sensitivity was 91.8% (87.1–94.9), specificity 83.5% (77.2–88.3), and real-time negative predictive value 86.5% (78.0–92.1). For FICE, overall sensitivity was 91.8% (87.1–94.9), specificity 83.5% (77.2–88.3), and real-time negative predictive value 83.7% (77.5–88.4). For autofluorescence imaging, overall sensitivity was 86.7% (79.5–91.6), specificity 65.9% (50.9–78.2), and real-time negative predictive value 81.5% (54.0–94.3). For CLE, overall sensitivity was 93.3% (88.4–96.2), specificity 89.9% (81.8–94.6), and real-time negative predictive value 94.8% (86.6–98.1).

Interpretation

All endoscopic imaging techniques other than autofluorescence imaging could be used by appropriately trained endoscopists to make a reliable optical diagnosis for colonic lesions in daily practice. Further research should be focused on whether training could help to improve negative predictive values.

INTRODUCTION

Colorectal cancer develops from precursor lesions called colorectal polyps, which can be detected during colonoscopy. Removal of these lesions can prevent the development of the disease.¹ Colorectal polyps can be neoplastic, adenomas, or non-neoplastic (e.g., serrated polyps and inflammatory polyps). Neoplastic lesions can become malignant, but the risk of non-neoplastic lesions other than serrated polyps becoming cancerous is negligible.²³ Accurate in-vivo differentiation between the types of lesions would assist decision making about endoscopic treatment, especially in the distal colon, where non-neoplastic diminutive polyps (<5 mm) can be left in situ. Additionally, such differentiation would mean that the so-called resect and discard strategy could be implemented (ie, not all lesions would need histo-pathological tests after removal) and decisions about appropriate surveillance intervals could be made directly after colonoscopy.⁴

In 2011, the Preservation and Incorporation of Valuable Endoscopic Innovations (PIVI) statement about real-time endoscopic assessment of the histology of diminutive colorectal polyps⁵ introduced two criteria for assessments of whether a technique or device could replace histopathological assessment (the gold standard). First, when the technology for optimum diagnosis is used to make an in-situ endoscopic diagnosis for diminutive polyps with high confidence, this technology should result in the same surveillance interval that would have been assigned after pathological assessment of polyps at least 90% of the time. Second, for a technology to be used to guide the decision to leave suspected rectosigmoid hyperplastic polyps of less than 5 mm in size in place (without resection), the technology should provide 90% or greater negative predictive value, when used with high confidence, for adenomatous histology. Practically, if an endoscopist uses a specific technology and achieves a negative predictive value of at least 90%, diminutive lesions in the rectosigmoid

colon could be left in situ if they are deemed to be non-adenomatous, other lesions could be resected but not sent in for pathology, and the surveillance interval could be established immediately.⁵

In the past two decades, several new endoscopic imaging techniques have been developed to improve endoscopic differentiation between neoplastic and non-neoplastic colonic lesions beyond standard white light assessment. Narrow-band imaging (NBI, Olympus, Japan), image-enhanced endoscopy (i-scan, Pentax, Japan), and Fujinon intelligent chromoendoscopy (FICE, Fujinon, Japan)—which are also all called virtual, digital, or electronic chromoendoscopy are all built-in endoscopic imaging techniques. NBI is a blue light technology that highlights superficial mucosal vasculature and enhances surface patterns through illumination via narrowed bandwidth filters. Both i-scan and FICE use spectral-estimation technology to re construct images at different wavelengths on the basis of white light images.

Other new image enhanced techniques that are widely commercially available are confocal laser endomicroscopy (CLE) and autofluorescence imaging. CLE is a system that can provide highly magnified images of gastro intestinal epithelium that are similar to histopathological images through a miniaturised confocal laser endo-microscope, either integrated into the endoscope (Pentax, Japan), or via a probe introduced down the working channel of the endoscope (Mauna Kea Technologies, France). Autofluorescence imaging makes use of differences in mucosal blood flow and endogenous fluorophores (eg, collagen, flavins, and NADPH), which change the autofluorescence signal emitted after short wavelength illumination. The signal is processed to create a false-colour image to assist differentiation between neoplastic and non-neoplastic colonic lesions.

The diagnostic performance of these techniques has been widely studied both in single studies and in single-modality meta-analyses.⁶⁻⁸ However, a comprehensive overview of the accuracy and precision for all available techniques has not been combined in one meta-analysis with standardised inclusion criteria, data extraction, and statistical approach. Our aim was to establish the sensitivity, specificity, and real-time negative predictive value of NBI, i-scan, FICE, autofluorescence imaging, and CLE for differentiation between

neoplastic and non-neoplastic colonic lesions, with histopathology as the reference standard.

METHODS

Search strategy and selection criteria

We did a meta-analysis in accordance with the Preferred Reporting Items for Systematic reviews and Meta-Analyses (PRISMA) guidelines.⁹ Under the supervision of a librarian at the University of Amsterdam (Amsterdam, Netherlands), we searched Medline from Jan 1, 1966, to Jan 14, 2013, Embase from Jan 1, 1986, to Jan 14, 2013, and PubMed from inception to Jan 14, 2013. We used the search term "NBI' [Mesh] OR NBI [tiab] OR i-SCAN [tiab] OR FICE [tiab] OR confocal OR CLE [tiab] OR autofluorescence [tiab] OR AFI [tia] OR Fujinon intelligent chromo endoscopy [tiab] OR Flexible spectral imaging color enhancement [tiab] OR confocal [tiab] OR real time histology [tiab] AND'Colonoscopy' [Mesh] OR colonoscop* [tiab] OR colon imag* [tiab] OR intestinal imag* [tiab]". We also searched the Cochrane Library for any relevant additional review with data that was published before Jan 14, 2013. We used no language restrictions. We then selected suitable studies for inclusion in our analysis on the basis of the abstracts of the selected reports. We checked reference lists of the reports identified in the original search to identify studies that had been missed.

LKW and SEU reviewed the identified studies to assess whether they were eligible for inclusion. Clinical trials and observational studies were eligible for inclusion when the performance of NBI, i-scan, FICE, autofluorescence imaging, or CLE, or any combination of the five, had been assessed for differentiation between non-neoplastic and neoplastic lesions in the colon, with histopathology as the reference standard, and for which a 2 × 2 contingency table of lesion diagnosis could be constructed. We included conference abstracts when they contained relevant data. We approached authors of abstracts to receive relevant unpublished data. We excluded studies that were focused on surveillance in patients with inflammatory bowel diseases or polyposis syndromes, or on lesion detection only. We also excluded those for which inadequate data for histopathology were available. We examined studies for overlapping data and made contact with the relevant investigators when necessary.

Procedures

Because all the included studies were diagnostic in nature, we assessed their guality and risk of bias with QUADAS-2 (Quality Assessment of Diagnostic Accuracy Studies).¹⁰ We measured overall sensitivity, specificity, and negative predictive value for real-time assessments of NBI, CLE, autofluorescence imaging, FICE, and i-scan to differentiate between neoplastic and non-neoplastic lesions. Additionally, we investigated heterogeneity by assessing sensitivity and specificity of NBI, i-scan, FICE, autofluorescence imaging, and CLE for real-time (in-vivo) diagnosis versus post-procedure image-based diagnosis, polyp size, use of highdefinition technology, magnification, high confidence assessments, and type of colonoscopy system. LKW and SEU independently extracted relevant data from the selected studies. They solved discrepancies by discussion either with each other or with JEE and ED. For the core analysis, we constructed 2 \times 2 contingency tables of true positives (neoplastic lesions predicted to be neoplastic), false positives (non-neoplastic predicted to be neoplastic), true negatives (non-neoplastic predicted to be non-neoplastic), and false negatives (neoplastic predicted to be non-neoplastic). When possible, we used raw data for numbers of lesions. Otherwise, we calculated the number in each category from the numbers of neoplastic and non-neoplastic lesions and the sensitivity and specificity.

Additionally, LKW and SEU extracted data from each trial for country of origin, study year, study design, number of patients, sex ratio, indication for colonoscopy, number of endoscopists and their experience, number of lesions, and features of the modalities used (including high definition, magnification, and type of endoscopy system for NBI [Exera **vs** Lucera]).

Statistical analysis

We did a random-effects bivariate meta-analysis using a non-linear mixed model approach to calculate summary estimates of sensitivity and specificity.¹¹ This approach models the sensitivity and specificity, while accounting for the correlation between the two because of threshold effects. We used the same approach to calculate summary estimates of positive predictive values and negative predictive values.¹² For all modalities, we plotted study estimates and a summary point with its 95% Cls in a summary receiver-operating characteristic plot. In this plot, all studies are presented, with the size of the study points representing the sample

size of the study. The summary point is represented by a dot, surrounded by a 95% confidence region (appendix).

For the overall analysis, we used both real-time and post-procedure studies. To estimate the negative predictive value, we used only real-time studies, because images used in post-procedure studies would have been selected for inclusion on the basis of quality, which would mean the ratio of neoplastic to nonneoplastic lesions in included images would not be representative of the ratio recorded in patients. Because the predictive values depend directly on this ratio, they cannot be estimated from these studies.

The correlation between sensitivity and specificity makes tests for heterogeneity difficult; most variation in sensitivity will be explained by variation in specificity, and vice versa.^{13,14} Therefore, statistical tests and *I*² values as used in meta-analyses of interventions are not helpful. We assumed that heterogeneity was present in our data and planned to deal with it with random effects modeling and by investigation of the sources of heterogeneity. To investigate the potential sources, we assessed the effects of type of assessment (real-time **vs** post-procedure), magnification, high definition, type of endoscope, and polyp size (overall **vs** diminutive) on the summary estimates. We included these variables one by one as covariates in the bivariate model. We included covariates if at least three studies were available for each value of the covariate. The difference between a subgroup and the group without the feature is shown by a delta estimate. We did not assess publication bias, because no proven statistical method exists for this type of meta-analysis.¹³ We used SAS (version 9.2) with NLMIXED for all statistical analyses.

Role of the funding source

There was no funding source for this study. All authors had full access to all the data in the study and final responsibility for the decision to submit for publication.

RESULTS

From the initial keyword search, we identified 390 separate reports (figure 1). 91 studies were included in our analysis, of which 90^{15–103} had been reported; details of another were provided by Arthur Hoff man (Johannes Gutenberg University of Mainz, personal communication). More than one imaging modality was assessed in some of the studies.^{16,17,27,29,33,38,52,53}



The 56 included NBI studies^{4,15-69} were mostly done in Asia, Europe, and the USA, and were generally reported between 2006 and 2013. We included 13 NBI studies that had been reported in abstracts; we obtained additional raw data directly from the investigators. A high proportion of NBI studies consisted of real-time assessments of colorectal lesions (table 1). Two studies consisted of both post-procedure and real-time assessments. 28 NBI studies (50%) were done with Exera series endoscopy systems, 24 (43%) with Lucera spectrum endoscopy systems, and four (7%) with unknown systems. Sensitivity for differentiation of colorectal lesions in all NBI studies varied from 60.0% to 100%, and specificity from 31.5% to 100%.

					Subgro	up propoi	rtion (%)			
		Real-time	MPP (range)	MAP (range)	Diminutive	+ High Confidence	Magnification	High- Definition	Typ endo syst	e of scopy tem
	NBI	39/56	1.9	1.1	16/56	5/56	24/56	27/56	Exera	Lucera
dality		(69.6)	(0.9-2.9)	(0.5-1.5)	(28.6)	(8.8)	(42.9)	(48·2)	28/56 (50·0)	27/56 (48·2)
Moc	iSCAN	8/9 (88·9)	1.7 (0.9-2.1)	0·9 (0·6-1·3)	0/9 (0·0)	0/9 (0·0)	0/9 (0·0)	9/9 (100·0)	N/A	N/A
	FICE	13/14 (92·9)	1.6 (0.7-2.4)	1·1 (0·6-1·7)	5/14 (35·7)	0/14 (0·0)	10/14 (71·4)	11/14 (78·6)	N/A	N/A
	AFI	9/11 (81·8)	1·9 (1·4-2·1)	1.0 (0.9-1.4)	0/11 (0·0)	0/11 (0·0)	1/11 (9·1)	N/A	N/A	N/A
	CLE	6/11	2.6	1.2	0/11	0/11	11/11	N/A	iCLE	pCLE
		(54.5)	(1.6-3.6)	(1.0-1.4)	(0.0)	(0.0)	(100.0)		5/11	6/11
									(45.5)	(54.5)

 Table 1 Subgroup study characteristics by imaging modality

MPP = mean polyps per patient, MAP = mean adenomas per patient, range = interval between 10% and 90% of study means. N/A = not applicable

Of the 56 NBI studies, seven had at least one item scored as high risk in QUADAS-2 all for selection of patients—suggesting a high risk of bias (appendix). The sensitivity and specificity values for real-time NBI assessments were not significantly different from those for post-procedure assessments (sensitivity: p=0.69; specificity: p=0.56; table 2). The negative predictive value was significantly lower in studies that included a higher proportion of neoplastic lesions (p=0.00026).

NBI had significantly lower sensitivity and specificity when high-definition assessments were done than when high definition was not used (p<0.0001 for both; table 2). Sensitivity did not vary according to whether magnification was used (p=0.24), but specificity increased significantly with magnification (p=0.032; table 2). High-definition technology was used in 20 of 33 studies in which magnification was not used, and five of 21 in which magnification was used (p=0.0082). This difference could have affected the effect of magnification on performance. To test this hypothesis, we included both magnification and high definition in the same model. We noted that magnification again had no significant effect on sensitivity (p=0.82) and no longer had a significant effect on specificity (p=0.42), but the effect of high definition on sensitivity (p=0.00017) and specificity (p=0.00013) was significant.

NBI studies in which only diminutive polyps were assessed and those in which all sizes of polyps were assessed did not differ significantly in terms of sensitivity (p=0.12) or specificity (p=0.98; table 2). Similarly, studies in which an Exera endoscopy system was used and those in which a Lucera system was used did not differ significantly in terms of sensitivity (p=0.18) or specificity (p=0.11; table 2).

The ten included i-scan studies—of which nine had been reported^{37,70-76} and one communicated to us (Hoffman A, Johannes Gutenberg University of Mainz, personal communication)—were reported between 2009 and 2013. Four studies had been done in Germany, three of which were done by the same research group. Two generations of i-scan were compared in one study.76 Almost all i-scan studies were real-time assessments of colorectal lesions (table 1). Sensitivity in i-scan studies varied from 54.5% to 94.6%, and specificity from 64.0% to 100%.

None of the i-scan studies had a high-risk item in QUADAS-2 (appendix). The sensitivity and specificity values for real-time i-scan assessments were similar to those for all i-scan assessments (table 2). Because of the small number of i-scan studies, we could not do sub-analyses. All were done with high-definition colonoscopy; therefore the estimate of diagnostic performance of real-time i-scan is a high-definition estimate.

The 14 included FICE studies were reported between 2007 and 2012.^{17,31,77-88} One study consisted of only post-procedure assessments, one of both post-procedure and real-time assessments, and the rest of only real-time assessments (table 1). Sensitivity in FICE studies varied from 73.9% to 100%, and specificity from 61.2% to 96.4%.

None of the FICE studies had a high-risk item in QUADAS-2 (appendix). The diagnostic performance of real-time assessments was similar to that in the overall analysis (table 2). Sensitivity increased significantly when magnification was used (p=0.0081), but specificity did not vary (p=0.64; table 2). Sensitivity (p=0.58) and specificity (p=0.12) did not vary according to whether high definition was used (table 2). Finally, assessment of only diminutive polyps did not significantly affect sensitivity (p=0.93) or specificity (p=0.17; table 2).

The 11 included studies of autofluorescence imaging were reported between 1998 and 2012.^{15,29,33,38,52,53,89–93} Most consisted of real-time assessments (table 1). No studies were focused on differentiation of diminutive lesions alone (table 1). Sensitivity varied from 57.5% to 98.9%, and specificity from 7.5% to 90.9%.

Five studies of autofluorescence imaging had at least one item scored as high risk in QUADAS-2 for selection of patients, suggesting a high risk of bias (appendix). We also deemed one study to have a high risk for bias related to the interpretation of the index test (appendix). The sensitivity (p=0.31) and specificity (p=0.48) values for real-time AFI assessments were not significantly different from those for post-procedure assessments (table 2). Because of the small number of studies of autofluorescence imaging, we could not do sub-analyses.

The 11 included CLE studies were reported between 2004 and 2012.^{17,94–103} Five consisted of only post-procedure assessments, one of both post-procedure and real-time assessments, and the rest of only real-time assessments (table 1). No studies were focused on differentiation of diminutive lesions alone (table 1). Five CLE studies (45%) were done with integrated techniques and six (55%) with probe-based techniques. Sensitivity in CLE studies varied from 76.0% to 100%, and specificity from 68.0% to 99.1%.

None of the CLE studies had a high-risk item in QUADAS-2 (appendix). Specificity increased significantly with real-time assessments when compared with post-procedure assessments (p=0.0048), but sensitivity did not (p=0.34; table 2). The only sub-analysis that we could do for CLE was the comparison between probe-based and integrated techniques. Specificity was significantly higher for integrated techniques than for the probe-based techniques (p=0.011), but no significant effect on sensitivity was recorded (p=0.34; table 2). However, when adjusted for real-time assessment, the difference was no longer significant (p=0.53).

Modality	Study characteristics	No. of studies (lesions)	Summary estimates	(95% CI)		Delta per subgroup i	n percentage (95% Cl)
			Sensitivity	Specificity	NPV	Sensitivity	Specificity
NBI	Overall	56 (18,051)	91.0 (88.6 to 93.0)	85.6 (81.3 to 89.0)	1		
	Real-time	39	91.5 (88.2 to 93.9)	85.2 (80.0 to 89.3)	82.5 (75.4 to 87.9)	1.0 (-3.9 to 5.9)	-2·4 (-10·5 to 5·6)
	Polyps < 6mm	16	86-9 (81-0 to 92-8)	84.4 (76.7 to 92.1)	I	-4·6 (-10·9 to 1·8)	0.0 (-8.9 to 9.0)
	+ high confidence	5	87.1 (77.8 to 92.9)	85.3 (74.2 to 92.1)	I	I	I
	Magnification	24	92.0 (89.0 to 95.0)	89-0 (84-3 to 93-7)	I	2.6 (-1.7 to 7.0)	8.1 (0.9 to 15.3)
	No magnification	35	89.4 (86.3 to 92.6)	80-9 (75-4 to 86-4)		ı	ı
	High Definition	27	85.2 (81.1 to 89.4)	74.6 (68.0 to 81.3)	I	-8·4 (-13·0 to 3·9)	-15·7 (-23·1 to -8·3)
	Exera	28	89.4 (86.0 to 92.8)	85.7 (80.9 to 90.4)	ı	-1.4 (-6.1 to 3.3)	6.1 (-2.3 to 14.5)
	Lucera	24	90.8 (87.6 to 94.0)	79.6 (72.6 to 86.5)	ı	I	ı
iSCAN	Overall	9 (1,143)	89-3 (83-3 to 93-3)	88-2 (80-3 to 93-2)	ı	I	I
	Real-time	00	89.5 (82.7 to 93.8)	89.3 (81.0 to 94.2)	86.5 (78.0 to 92.1)	I	I
FICE	Overall	14 (4,824)	91.8 (87.1 to 94.9)	83·5 (77·2 to 88·3)	I	I	I
	Real-time	13	92·5 (87·6 to 95·6)	85.1 (78.7 to 89.8)	83·7 (77·5 to 88·4)	I	I
	Polyps < 6mm	5	83·6 (72·8 to 94·4)	86.5 (79.4 to 93.6)	ı	-1.7 (-12.0 to 8.8)	5.6 (-2.1 to 13.3)
	Magnification	10	93.7 (90.6 to 96.8)	82.6 (76.9 to 88.2)	ı	8·5 (1·4 to 15·6)	1.7 (-6.3 to 9.7)
	High Definition	10	92·6 (88·2 to 97·0)	79.8 (72.2 to 87.4)	ı	2·2 (-6·0 to 10·3)	-7.7 (-17.4 to 2.0)
AFI	Overall	11 (1,670)	86-7 (79-5 to 91-6)	65-9 (50-9 to 78-2)	ı	I	I
	Real-time	6	88.0 (80.5 to 92.8)	69.2 (51.7 to 82.4)	81.5 (54.0 to 94.3)	8.5 (-10.2 to 27.3)	12.9 (-24.0 to 49.7)
CLE	Overall	11 (1,372)	93·3 (88·4 to 96·2)	89-9 (81-8 to 94-6)	ı	I	I
	Real-time	9	94.3 (88.1 to 97.3)	94.8 (87.3 to 98.1)	94.8 (86.6 to 98.1)	5·1 (1·5 to 8·7)	12·7 (6·0 to 19·4)
	iCLE	5	94.8 (90.6 to 98.9)	94.4 (90.7 to 99.2)	ı	I	I
	pCLE	9	91·5 (86·0 to 97·0)	80-9 (69-4 to 92-4)		5.2 (1.9 to 8.4)	11.1 (5.1 to 17.2)

Table 2 Diagnostic performance of optical diagnosis of NBI, iSCAN, FICE, CLE and AFI

CI: confidence interval, NPV: Negative Predictive Value, Delta: difference between subgroup and non-subgroup

DISCUSSION

We have shown that built-in endoscopic imaging techniques have overall negative predictive values of greater than 80% in the differentiation between neoplastic and non-neoplastic lesions. NBI, i-scan, FICE, and CLE have similar sensitivity and specificity overall. Autofluorescence imaging had a sensitivity of more than 85%, but had a much lower specificity than did the other investigated modalities. In the overall analysis, only CLE had a negative predictive value of more than 90%.

To our knowledge, ours is the first meta-analysis to give an overview of the accuracy of all available built-in image-enhanced techniques for optical diagnosis of colorectal lesions. The diagnostic performance of NBI and CLE have been assessed separately in meta-analyses, 6–8 although fewer studies were included. Our analysis provides an overview of optical diagnosis with all available techniques, analysed with a standardised approach. We hope that the outcomes reported here will support future guidelines and research.

With the results of our meta-analysis, we can compare the real-time negative predictive values of the different modalities with the criteria in the PIVI statement. However, we calculated the overall negative predictive value, not just that for diminutive rectosigmoid lesions as in the PIVI statement. Because the prevalence of non-neoplastic lesions is increased in the rectosigmoid area, studies of only rectosigmoid negative predictive value are likely to show a good diagnostic performance.

To provide a complete overview of all data available for differentiation of colonic lesions and to maximise the precision of estimates, we did an overall analysis of both real-time and post-procedure assessments for all five modalities. Real-time assessment is the optimum situation to investigate performance, because it avoids bias of photographic selection, simulates an in-vivo optical diagnosis, and allows calculation of a negative predictive value on the basis of the real-life lesion prevalence. However, if we were to have included only real-time assessments, the number of included studies and reports would have substantially reduced and potentially important data would have been missed. Studies with real-time data could be analysed separately from post-procedure photographic studies, rather than using real time in the analysis. In most studies, either

a real-time or post-procedure approach was used, but not both. We did not expect characteristics of real-time studies to be different from those of postprocedure studies, and almost all real-time analyses were similar to the overall analyses. The only significant difference was that real-time specificity of CLE was significantly better than the overall specificity (p=0.0048). Stratification for real time would have reduced numbers, making investigation of other sources of heterogeneity (e.g., magnification) more difficult. When we had enough data, we did extra subgroup analyses. However, because the number of NBI studies was more than three times that of any other modality, most subgroup analysis was restricted to this technique.

Of all advanced imaging techniques, NBI is the most studied. McGill and colleagues⁷ reported that real-time assessment of colorectal lesions had a sensitivity of 91.0% (95% CI 87.6–93.5) and a specificity of 82.6% (79.0–85.7)—i.e., similar values to those in our analysis. However, the CIs reported by McGill and colleagues are wider than ours were, presumably because they used a smaller sample (28 studies).⁷

Counter intuitively, we recorded that high definition significantly decreased the performance of NBI. A possible explanation is that high-definition techniques are used by endoscopists who are less experienced than are those who use magnification. Before high-definition technology was widely available, standard-definition colonoscopes incorporated magnification to allow detailed examination of minute mucosal structures, and were mainly used by experts; the image is more detailed than with high definition but no magnification. However, we could not test this hypothesis with additional analyses.

Our sub-analysis of high confidence assessments did not improve diagnostic performance for diminutive lesions. Although it is recommended in the PIVI statement that only assessments made with high confidence should be used to make optical diagnoses, confidence is subjective and could be less important than has been previously supposed. Several studies of how individuals learn to differentiate with NBI have been done.^{22,30,35,44,51,58} Three studies^{22,44,58} of the accuracy before and after training showed that gastroenterology trainees and less experienced endoscopists (e.g., community-based endoscopists) can learn to predict histology with an accuracy of at least 90%. Therefore, an endoscopist

needs to be trained before he or she can implement the resect and discard strategy in daily clinical practice. Nevertheless, results from community based studies of NBI or i-scan suggest that training in community clinical practice could be substantially more difficult than would be assumed from training studies.^{34,35,104}

Several classification methods for differentiation of colorectal lesions have been developed to guide endoscopists in optical diagnosis of colorectal lesions, most commonly for NBI. One of the most recently developed is the NBI international colorectal endoscopic classification, which uses colour, vessels, and surface pattern to help endoscopists to distinguish between hyperplastic and adenomatous lesions.²⁷

FICE and i-scan have similar diagnostic performance to NBI, although their confidence regions are notably larger than for NBI, because of the small number of studies. CLE had a high sensitivity and specificity for real-time diagnosis of colorectal lesions. Integrated techniques had a significantly better diagnostic performance than did probe-based techniques in our initial analysis, but the difference was no longer significant after adjustment for real time. The studies of probe-based CLE could have assessed patients or lesions that were different from those of integrated CLE—i.e., spectrum bias might have been an issue. The outcome for real-time CLE is similar to the results of Su and colleagues' meta-analysis,⁸ which showed a pooled sensitivity of 0.94 (95% CI 0.88–0.97) and a pooled specificity of 0.95 (0.89–0.97). However, CLE can only be done by highly specialized endoscopists with expensive and fragile equipment—factors that might have caused the high accuracy in our analysis and make CLE less suitable for daily clinical practice than the other modalities. Nevertheless, CLE might be an interesting technique to use for specific indications in tertiary referral centres.

Autofluorescence imaging had a fairly good sensitivity, but a substantially lower specificity than did the other modalities. Autofluorescence imaging differentiates between neoplastic and non-neoplastic lesions by colour of the lesion when compared with the surrounding mucosa. This technique is accurate when the colour is either clear green or purple, but difficulties arise when the colour is not clear enough to distinguish. The fairly low specificity of autofluorescence imaging might be explained by the fact that endoscopists prefer not to misinterpret neoplastic lesions and therefore are more likely to classify lesions as

neoplastic than non-neoplastic when uncertain. However, none of the included studies of autofluorescence imaging incorporated measures of confidence in their study design. Initially, autofluorescence imaging was studied as a technique on its own for optical diagnosis, but in most studies in the past 2 years, autofluorescence imaging was combined with NBI and high-definition white light endoscopy—the so-called endoscopic trimodal imaging approach. Rotondano and colleagues' study⁵² indicated that autofluorescence imaging is of additional value to NBI for both the detection and the differentiation of colorectal lesions.

Every meta-analysis has limitations due to the extraction of data from many different reports. A major limitation of our study was that we could often make no clear distinction between experts and non-experts. There is no unambiguous definition of expert; in some studies, an endoscopist was defined as an expert after a specific number of colonoscopies, but in others, individuals had to have had specific training. Generally, experts did most of the studies, which reduces the value of the outcomes for daily practice. Besides the inequality of expertise and the absence of a validated training programme, several classification systems were used in the included studies, reducing the generalisability of the overall performance. Levels of confidence, and especially whether assessments were made with high confidence should be necessary before a method of optical diagnosis can be applied in daily practice. Gupta and colleagues¹⁰⁵ showed that, besides confidence, how long it takes someone to make a diagnosis affects the accuracy; they advised a wait of 5 s before a diagnosis is made.

Other limitations are that polyp size and location were most commonly not clearly described in individual studies, and serrated lesions were subsequently not sub-classified as sessile serrated adenomas or polyps, traditional serrated adenomas, or hyperplastic lesions. Therefore, sessile serrated adenomas or polyps might have previously been put in non-neoplastic groups, whereas nowadays most clinicians deem these lesions to be neoplastic according to the serrated neoplasia pathway, especially outside the rectosigmoid.² Potential bias also exists when specific populations of patients were excluded. Studies that included patients with either inflammatory bowel disease or polyposis syndromes were excluded from our analysis because lesions in these disorders are fundamentally different in phenotype—e.g., dysplasia-associated lesions or masses in colitis

or hamartomatous polyps in polyposis syndromes. Studies in patients with hereditary non-polyposis colorectal cancer were included in the analysis because there is no reason to suspect that the endoscopic characteristics of polyps in these patients are different from those of patients with an average risk for polyps.

Another limitation is that we did not formally compare the differences between the five modalities assessed. The use of network meta-analyses for diagnostic accuracy data is not yet reliable. As yet, only one method has been reported that combines both direct and indirect comparisons; a real network meta-analysis of diagnostic accuracy has not yet been reported, although such an analysis could be possible in the future.¹⁰⁶ If formal comparisons are to be made, studies that assess all (or at least two) modalities against the same reference standard in the same patients or in a randomised design would be ideal.¹³ However, these studies are rare; we identified eight, which were all about different pairs of modalities.^{16,17,27,29,33,38,52,53} Without these so-called direct comparisons, comparisons of the accuracy of one modality with the accuracy of another could result in biased conclusions, because both modalities could be assessed in a different population. This source of bias (spectrum bias) is the reason that we did not make formal comparisons between the investigated modalities.¹⁰⁷ We did create a table with the proportions by subgroup per modality, but comparisons of the different modalities should be approached with caution.

In our meta-analysis, we calculated negative predictive values for all modalities, but negative predictive values are strongly dependent on the ratio between neoplastic lesions and non-neoplastic lesions in the patients. Care should be taken in comparisons of the accuracy of these modalities against each other and when the negative predictive values from our review are applied in practice.

In conclusion, all endoscopic imaging techniques other than autofluorescence imaging could be used by appropriately trained endoscopists to make a reliable optical diagnosis for colonic lesions in daily practice. Further research should be focused on whether training in narrow-spectrum endoscopy (ie, NBI, i-scan, and FICE) will help community-based gastroenterologists to reach a negative predictive value of at least 90% for diminutive rectosigmoid adenomas and maintain that value.

APPENDIX A SROC-plots

Summary receiver-operating characteristic (SROC) plots of overall performance per modality



	Author	Year	Country	Publication	Patients	Neo- plasia	Non- neoplasia	Magnification	High Definition	Evaluation	<6mm	Colonoscope
	Broek vd ¹⁵	2009	Netherlands	Full text	100	22	28	no	yes	РР	no	Lucera
2	Broek vd ¹⁶	2009	Netherlands	Full text	107	88	118	no	yes	RT	no	Lucera
m	Buchner ¹⁷	2010	USA	Full text	28	25	16	no	yes	РР	no	Exera
4	Canales ¹⁸	2011	Peru	Full text	134	76	43	yes	,	RT	no	Exera
5	Chang ¹⁹	2009	Taiwan	Full text	104	82	81	yes	ou	РР	no	Lucera
9	Chiu ²⁰	2012	Taiwan	Full text	133	141	39	yes	no	РР	no	Lucera
7	Coe ²¹	2012	USA	Full text	654	467	307	no	yes	RT	no	Exera
00	Dai ²²	2012	China	Full text	326	30	10	yes	yes	RT	no	Lucera
6	East ²³	2008	UK	Full text	62	50	66	yes	yes	RT	yes	Lucera
10	Gross 24	2011	Germany	Full text	214	120	135	yes	no	RT	yes	Exera
11	Heller ²⁵	2011	I	Abstract	100	27	23	no		РР	yes	UK
12	Henry ²⁶	2010	USA	Full text	52	67	59	no	yes	РР	yes	Exera
13	Hewett ²⁷	2012	USA	Full text	31	39	197	no	ou	RT/PP	no	Exera
14	Hirata ²⁸	2007	Japan	Full text	66	132	16	yes	no	RT	no	Lucera
15	Ignjatovic⁴	2009	UK	Full text	130	198	80	no	yes	RT/PP	yes	Lucera
16	Ignjatovic ²⁹	2011	UK	Full text	48	40	40	both	yes	РР	no	Lucera
17	<i>lgnjatovic</i> ³⁰	2011	UK	Full text	ı	15	15	no	ou	РР	no	Lucera
18	Kang ³¹	2012	Korea	Abstract	821	943	667	no	no	PP	no	Lucera
19	Katagiri ³²	2008	Japan	Full text	104	134	5	yes	no	РР	no	Lucera
20	Kuiper ³³	2011	Netherlands	Full text	118	115	122	no	yes	RT	no	Exera
21	Kuiper ³⁴	2012	Netherlands	Full text	108	141	167	no	yes	RT	no	Exera
22	Ladabaum ³⁵	2013	USA	Full text	ı	1541	1055	no	yes	RT	yes	Exera
23	Lau ³⁶	2012	China	Abstract	441	341	178	yes		RT	no	Exera
24	Lee 37	2011	Korea	Full text	70	80	76	no	yes	RT	yes	Lucera
25	Lin ³⁸	2013	Taiwan	Abstract	66	84	37	yes	yes	RT	no	Lucera

APPENDIX B Study characteristics

Table 3 Study characteristics NBI, iSCAN, FICE, CLE and AFI NBI

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	Author	Year	Country	Publication	Patients	Neo- plasia	Non- neoplasia	Magnification	High Definition	Evaluation	<6mm	Colonoscope
26	Lopata ³⁹	2012	USA	Abstract	482	405	401	no	yes	RT	ou	Lucera
27	Machida ⁴⁰	2004	Japan	Full text	34	34	6	no	no	RT	no	Lucera
28	Occhipinti ⁴¹	2012	Italy	Abstract	93	120	100	yes	no	РР	no	Exera
29	Oka ⁴²	2011	Japan	Full text	ı	645	53	yes	no	RT	no	UK
30	Paggi ⁴³	2012	Italy	Full text	286	350	161	no	yes	RT	yes	Exera
31	Patel ⁴⁴	2013	USA	Full text	ı	52	28	no	yes	РР	yes	Exera
32	Pohl 45	2012	Germany	Abstract	608	616	883	no	no	RT	yes	Exera
33	Ramirez ⁴⁶	2009	USA	Abstract	56	5	22	no	ı	RT	no	UK
34	Rastogi 47	2008	USA	Full text	40	43	28	no	yes	RT	yes*	Exera
35	Rastogi ⁴⁸	2009	USA	Full text	100	143	93	no	yes	RT	yes*	Exera
36	Rastogi ⁴⁹	2011	USA	Full text	210	147	237	no	yes	RT	yes*	Exera
37	Rex ⁵⁰	2009	NSA	Full text	136	230	221	yes	no	RT	yes	Exera
38	Rogart ⁵¹	2008	USA	Full text	131	131	134	yes	yes	RT	no	Exera
39	Rotondano ⁵²	2011	Italy	Full text	94	141	140	yes	yes	RT	no	Lucera
40	Sato ⁵³	2011	Japan	Full text	183	339	85	no	no	РР	no	Lucera
41	Sakamoto ⁵⁴	2012	Japan	Full text	151	52	42	yes	no	RT	no	Lucera
42	Sano ⁵⁵	2008	Japan	Full text	92	111	39	yes	no	RT	no	Exera
43	Shahid ⁵⁶	2011	NSA	Full text	65	58	72	no	yes	RT	yes	Exera
44	Sikka 57	2008	USA	Full text	63	49	31	no	no	РР	ou	Exera
45	Singh ⁵⁸	2010	Australia	Full text	32	30	20	no	yes	РР	yes	Exera
46	Soto 59	2012	Venezuela	Abstract	85	60	66	no	no	RT	no	Exera
47	Su 60	2006	Taiwan	Full text	78	65	40	no	no	RT	ou	Lucera
48	Takemura ⁶¹	2012	Japan	Full text	ı	324	47	yes	no	РР	no	Lucera
49	Tischendorf ⁶²	2007	Germany	Full text	52	63	37	yes	no	RT	no	Exera
50	Tischendorf ⁶³	2010	Germany	Full text	131	121	79	both	yes	РР	ou	Exera

Table 3 Continued NBI

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3 Conti	
Table	NBI

	Author	Year	Country	Publication	Patients	Neo- plasia	Non- neoplasia	Magnification	High Definition	Evaluation	<6mm	Colonoscope
51	Valiante ⁶⁴	2011	Italy	Abstract	65	57	47	no	yes	RT	no	Exera
52	Wada ⁶⁵	2012	Japan	Abstract	495	1420	53	yes	no	RT	no	Lucera
53	Wang 🕫	2009	NSA	Abstract	15	15	10	no	no	RT	no	
54	Yague ⁶⁷	2011	Spain	Abstract	75	107	108	no	yes	RT	no	Exera
55	Yoo ⁶⁸	2011	Korea	Full text	68	89	18	yes	no	RT	no	Lucera
56	Zhou 69	2011	China	Full text	118	118	109	both	no	RT	no	Lucera
iscan												
	Author	Year	Country	Publication	Patients	Neo- plasia	Non- neoplasia	Magnification	High Definition	Evaluation	<6mm	Colonoscope
-	Chan ⁷⁰	2012	USA	Full text	43	54	49	ou	yes	RT	no	Pentax
2	Han ^{zi}	2012	Taiwan	Full text	54	57	44	no	yes	RT	no	Pentax
m	Hoffman ⁷²	2010	Germany	Full text	100	11	117	no	yes	RT	no	Pentax
4	Hoffman ⁷³	2010	Germany	Full text	69	82	63	no	yes	RT	no	Pentax
5	Hoffman ⁷⁴	2013	Germany	Abstract		23	28	no	yes	RT	no	Pentax

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Pentax Pentax Pentax Pentax Pentax

RT RT RT RT

yes yes yes

25 43 66 33 32

81 66 74 77 118

1115 1118 72 -78

> Full text Full text Full text Full text

> > Germany

Neumann 77

Pigo 78

2012 2012 2011 2013 2012

Lee 76

6 × 8 01

Hong* 76

Hong 75

Italy

Full text

Korea Korea Korea

Table	3 Continued											
FICE												
	Author	Year	Country	Publication	Patients	Neoplasia	Non- neoplasia	Magnification	High Definition	Evaluation	<6mm	Colonoscope
	Buchner ¹⁷	2010	USA	Full text	47	81	38	no	yes	РР	no	Fujinon
2	Kang ³¹	2012	Korea	Abstract	821	943	667	yes	ı	RT	no	Fujinon
m	Kim ⁷⁹	2011	Korea	Full text	361	325	210	both	yes	RT	yes	Fujinon
4	Liu ⁸⁰	2008	China	Full text	223	209	242	yes	no	RT	no	Fujinon
Ŝ	Longcroft ⁸¹	2011	UK	Full text	89	155	77	no	no	RT	yes	Fujinon
9	Longcroft ⁸²	2012	UK	Full text	170	96	54	no	both	RT	no	Fujinon
7	Pohl ⁸³	2008	Germany	Full text	63	89	61	both	yes	RT	no	Fujinon
00	Pohl ⁸⁴	2009	Germany	Full text	368	236	85	yes	yes	RT	no	Fujinon
6	Santos ⁸⁵	2009	Brazil	Full text	75	124	33	yes	yes	RT	no	Fujinon
10	Santos ⁸⁶	2010	Brazil	Full text	72	82	29	yes	yes	RT	no	Fujinon
11	Santos ⁸⁷	2012	Brazil	Full text	65	67	28	yes	yes	RT	yes	Fujinon
12	Teixeira ⁸⁸	2009	Brazil	Full text	148	250	59	yes	no	RT	yes	Fujinon
13	Togashi ⁸⁹	2009	Japan	Full text	133	80	27	yes	both	RT	yes	Fujinon
1 4	Yoshida 90	2011	Japan	Full text	I	114	6	yes	yes	RT	no	Fujinon

CHAPTER 2

	Author	Year	Country	Publication	Patients	Neoplasia	Non- neoplasia	Magnification	High Definition	Evaluation	<6mm	Colonoscope
	Broek vd ¹⁵	2009	Netherlands	Full text	50	88	118	no	yes	RT	no	Olympus
2	Ignjatovic ²⁹	2011	UK	Full text	48	40	40	no	yes	РР	no	Olympus
m	Kuiper ³³	2011	Netherlands	Full text	118	116	123	no	yes	RT	no	Olympus
4	Lin ³⁸	2011	Germany	Full text	66	84	37	yes	yes	RT	no	Olympus
Ŝ	McCallum ⁹¹	2008	UK	Full text	107	54	21	no	no	RT	no	Olympus
9	Mycek ⁹²	1998	USA	Full text	17	13	11	no	no	RT	no	Xillix
7	Nakaniwa ⁹³	2005	Japan	Full text		125	43	no	no	RT	no	Superguide
œ	Rotondano ⁵²	2012	Italy	Full text	47	66	7	no	yes	RT	no	Olympus
6	Sato 53	2011	Japan	Full text	183	339	85	no	yes	РР	no	Olympus
10	Shao 94	2011	Singapore	Full text	96	34	164	no	no	RT	no	Olympus
11	Uedo 95	2007	Japan	Full text	32	26	32	no	no	RT	no	Olympus
CLE												
	Author	Year	Country	Publication	Patients	Neoplasia	Non- neoplasia	Magnification	High Definition	Evaluation	<6mm	Colonoscope
-	Andre 96	2012	France	Full text	71	93	42	yes	yes	РР	no	Mauna Kea
2	Buchner ¹⁷	2010	USA	Full text		81	38	yes	yes	РР	no	Mauna Kea
m	Buchner 97	2011	USA	Full text	54	44	25	yes	yes	РР	no	Mauna Kea
4	Gomez ⁹⁸	2010	USA	Full text	53	50	25	yes	no	РР	no	Mauna Kea
2	Hurlstone ⁹⁹	2008	UK	Full text	40	55	107	yes	no	RT	no	Pentax
9	Kiesslich 100	2004	Germany	Full text	42	38	96	yes	no	RT	no	Pentax
7	DePalma ¹⁰¹	2010	Italy	Full text	20	21	11	yes	no	РР	no	Mauna Kea
œ	Sanduleanu ¹⁰²	2010	Netherlands	Full text	72	74	42	yes	no	RT	no	Pentax
6	Shahid ¹⁰³	2012	USA	Full text	74	80	74	yes	yes	RT/PP	no	Pentax
10	Singson 104	2012	I	Abstract	30	63	44	yes	yes	RT	no	Mauna Kea
11	<i>Xie</i> ¹⁰⁵	2011	China	Full text	115	66	49	yes	no	RT	no	Pentax

Table 3 Continued AFI 2

UK = United Kingdom, USA = United States of America, PP = post-procedure, RT = real-time

APPENDIX C Quality assessment

Table 4 Quality assessment of diagnostic accuracy studies (QUADAS) – 2 tool, for quality assessment of the included studies

NBI

Study	Risk of bias				Applicabi	lity concern	S
	Patient	Index test	Reference	Flow and	Patient	Index test	Reference
Due also d 54	selection	Laterna ell'ete	standara	unnig	Selection		Standard
Broek Va 34	Intermediate	Intermediate	LOW	LOW	INO	Yes	INO
Broek va 18	Yes	Low	LOW	LOW	NO	NO	NO
Buchner ¹⁷	Low	Low	Low	Low	No	No	No
Canales ¹⁸	Intermediate	Low	Low	Low	UK	No	No
Chang ¹⁹	High	Low	Low	Low	Yes	No	No
Chiu 20	High	Low	Low	Low	Yes	No	No
Coe ²¹	Low	Low	Low	Low	No	No	No
Dai ²²	Low	Low	Low	Low	No	No	No
East 23	Intermediate	Low	Low	Low	UK	No	No
Gross ²⁴	Low	Low	Low	Low	No	No	No
Heller ²⁵	UK	UK	Low	Low	UK	UK	No
Henry ²⁶	Low	Low	Low	Low	No	No	UK
Hewett 27	UK	Low	Low	Low	UK	No	No
Hirata ²⁸	High	Low	Low	Low	Yes	No	No
lgnjatovic ⁴	Low	Low	Low	Low	No	No	No
Ignjatovic 29	Low	Low	Low	Low	No	No	No
Ignjatovic 30	High	Low	Low	Low	Yes	No	No
Kang ³¹	UK	Low	Low	Low	UK	No	No
Katagiri ³²	High	Low	Low	Low	Yes	No	No
Kuiper 33	Low	Low	Low	Low	No	No	No
Kuiper ³⁴	Low	Low	Low	Low	No	No	No
Ladabaum 35	UK	Low	Low	Low	UK	No	No
Lau 36	UK	Low	Low	Low	UK	No	No
Lee 37	Low	Low	Low	Low	No	No	No
Lin 38	UK	Low	Low	Low	No	No	No
Lopata 39	UK	Low	Low	Low	No	No	No
Machida 40	UK	Low	Low	Low	No	No	No
Occhipinti 41	UK	Low	Low	Low	No	No	No
Oka ⁴²	Low	Low	Low	Low	No	No	No
Paggi 43	Low	Low	Low	Low	No	No	No
Patel ⁴⁴	Low	Low	Low	Low	No	No	No
Pohl 45	Low	Low	Low	Low	No	No	No
Ramirez 46	UK	UK	Low	Low	UK	UK	No
Rastoqi 47	Low	Low	Low	Low	No	No	No
Rastoqi 48	Low	Low	Low	Low	No	No	No
Rastoqi 49	Low	Low	Low	Low	No	No	No
Rex 50	UK	Low	Low	Low	UK	No	No

Table 4 Continued

NBI

Study	Risk of bias				Applicabi	lity concern	S
	Patient selection	Index test	Reference standard	Flow and timing	Patient selection	Index test	Reference standard
Rogart ⁵¹	Low	Low	Low	Low	No	No	No
Rotondano ⁵²	Low	Intermediate	Low	Low	No	No	No
Sato 53	UK	Low	Low	Low	UK	No	No
Sakamoto ⁵⁴	Low	Low	Low	Low	No	No	No
Sano 55	Low	Low	Low	Low	No	No	No
Shahid 56	Low	Low	Low	Low	No	No	No
Sikka 57	Low	Low	Low	Low	No	No	No
Singh ⁵ ⁸	UK	Low	Low	Low	UK	No	No
Soto 59	UK	Low	Low	Low	UK	No	No
Su 60	UK	Low	Low	Low	UK	No	No
Takemura 61	High	Low	Low	Low	Yes	No	No
Tischendorf 62	Intermediate	Low	Low	Low	No	No	No
Tischendorf 63	High	Low	Low	Low	Yes	No	No
Valiante ⁶⁴	Low	Low	Low	Low	No	No	No
Wada 65	UK	Low	Low	Low	UK	No	No
Wang ⁶⁶	UK	Low	Low	Low	UK	No	No
Yague ⁶⁷	UK	Low	Low	Low	UK	No	No
Yoo 68	UK	Low	Low	Low	UK	No	No
Zhou ⁶⁹	Low	Low	Low	Low	No	No	No

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Study	Risk of bias				Applicabil	ity concern	s
	Patient selection	Index test	Reference standard	Flow and timing	Patient selection	Index Test	Reference Standard
Chan ⁷⁰	Low	Low	Low	Low	No	No	No
Han 71	Intermediate	Low	Low	Low	No	No	No
Hoffman 72	Low	Low	Low	Low	No	No	No
Hoffman ⁷³	Low	Low	Low	Low	No	No	No
Hoffman 74	Low	Low	Low	Low	No	No	No
Hong 75	Low	Low	Low	Low	No	No	No
Hong* 75	Low	Low	Low	Low	No	No	No
Lee ⁷⁶	Low	Low	Low	Low	No	No	No
Neumann ⁷⁷	Low	Low	Low	Low	No	No	No
Pigo ⁷⁸	Low	Low	Low	Low	No	No	No

Table 4 Continued

FICE

Study	Risk of bias			Applicability concerns			
	Patient selection	Index test	Reference standard	Flow and timing	Patient selection	Index Test	Reference Standard
Buchner 17	Low	Low	Low	Low	No	No	No
Kang ³¹	UK	Low	Low	Low	UK	No	No
Kim 79	Low	Low	Low	Low	No	No	No
Liu ⁸⁰	Low	Low	Low	Low	No	No	No
Longcroft ⁸¹	Low	Low	Low	Low	No	No	No
Longcroft ⁸²	Low	Low	Low	Low	No	No	No
Pohl ⁸³	Low	Low	Low	Low	No	No	No
Pohl ⁸⁴	Low	Low	Low	Low	No	No	No
Santos ⁸⁵	Low	Low	Low	Low	No	No	No
Santos ⁸⁶	Low	Low	Low	Low	No	No	No
Santos 87	Low	Low	Low	Low	No	No	No
Teixeira ⁸⁸	Low	Low	Low	Low	No	No	No
Togashi ⁸⁹	Intermediate	Low	Low	Low	Intermediate	No	No
Yoshida 90	Intermediate	Low	Low	Low	Intermediate	No	No

AFI

Study	Risk of bias	Applicability concerns					
	Patient selection	Index test	Reference standard	Flow and timing	Patient selection	Index Test	Reference Standard
Broek vd 15	Intermediate	Intermediate	Low	Low	No	Yes	No
lgnjatovic ²⁹	High	Intermediate	Low	Low	Yes	No	No
Kuiper ³³	Low	Intermediate	Low	Low	No	No	No
Lin ³⁸	Low	Low	Low	Low	No	No	No
McCallum ⁹¹	Intermediate	Low	Low	Low	Yes	No	No
Mycek ⁹²	High	High	Low	Low	Yes	Yes	No
Nakaniwa 93	High	Low	Low	Low	Yes	No	No
Rotondano ⁵²	Low	Intermediate	Low	Low	No	No	No
Sato 53	Intermediate	Low	Low	Low	UK	No	No
Shao 94	High	Low	Low	Low	UK	No	No
Uedo ⁹⁵	High	Intermediate	Low	Low	Yes	UK	No

REFERENCES

- 1. Zauber AG, Winawer SJ, O'Brien MJ, et al. Colonoscopic polypectomy and long-term prevention of colorectal-cancer deaths. N Engl J Med 2012; 366: 687–96.
- Patai AV, Molnar B, Tulassay Z, Sipos F. Serrated pathway: alternative route to colorectal cancer. World J Gastroenterol 2013; 19: 607–15.
- 3. Alvarez C, Andreu M, Castells A, et al. Relationship of colonoscopy-detected serrated polyps with synchronous advanced neoplasia in average-risk individuals. Gastrointest Endosc 2013; 78: 333–41.
- Ignjatovic A, East JE, Suzuki N, Vance M, Guenther T, Saunders BP. Optical diagnosis of small colorectal polyps at routine colonoscopy (Detect InSpect ChAracterise Resect and Discard; DISCARD trial): a prospective cohort study. Lancet Oncol 2009; 10: 1171–78.
- Rex DK, Kahi C, O'Brien M, et al. The American Society for Gastrointestinal Endoscopy PIVI (Preservation and Incorporation of Valuable Endoscopic Innovations) on real-time endoscopic assessment of the histology of diminutive colorectal polyps. Gastrointest Endosc 2011; 73: 419–22.
- 6. Wu L, Li Y, Li Z, Cao Y, Gao F. Diagnostic accuracy of narrow-band imaging for the differentiation of neoplastic from non-neoplastic colorectal polyps: a meta-analysis. Colorectal Dis 2013; 15: 3–11.
- McGill SK, Evangelou E, Ioannidis JP, Soetikno RM, Kaltenbach T. Narrow band imaging to differentiate neoplastic and non-neoplastic colorectal polyps in real time: a meta-analysis of diagnostic operating characteristics. Gut 2013; published online April 17. DOI:10.1136/gutjnl-2012-303965.
- 8. Su P, Liu Y, Lin S, et al. Efficacy of confocal laser endomicroscopy for discriminating colorectal neoplasms from non-neoplasms: a systematic review and meta-analysis. Colorectal Dis 2013; 15: e1–12.
- 9. Moher D, Liberati A, Tetzlaff J, Altman DG, The PRISMA Group. Preferred Reporting Items for Systematic Reviews and Meta- Analyses: the PRISMA statement. PLoS Med 2009; 6: e1000097.
- 10. Whiting PF, Rutjes AW, Westwood ME, et al. QUADAS-2: a revised tool for the quality assessment of diagnostic accuracy studies. Ann Intern Med 2011; 155: 529–36.
- 11. Reitsma JB, Glas AS, Rutjes AW, Scholten RJ, Bossuyt PM, Zwinderman AH. Bivariate analysis of sensitivity and specificity produces informative summary measures in diagnostic reviews. J Clin Epidemiol 2005; 58: 982–90.
- 12. Leeflang MM, Deeks JJ, Rutjes AW, Reitsma JB, Bossuyt PM. Bivariate meta-analysis of predictive values of diagnostic tests can be an alternative to bivariate meta-analysis of sensitivity and specificity. J Clin Epidemiol 2012; 65: 1088–97.
- 13. Deeks JJ, Macaskill P, Irwig L. The performance of tests of publication bias and other sample size effects in systematic reviews of diagnostic test accuracy was assessed. J Clin Epidemiol 2005; 58: 882–93.
- 14. Macaskill P, Gatsonis C, Deeks JJ, Harbord RM, Takwoingi Y. Analysing and presenting results. In: Deeks JJ, Bossuyt PM, Gatsonis C, eds. Cochrane handbook for systematic reviews of diagnostic test accuracy, version 1.0. Oxford, UK: The Cochrane Collaboration, 2010.
- 15. van den Broek FJ, Fockens P, van Eeden S, et al. Clinical evaluation of endoscopic trimodal imaging for the detection and differentiation of colonic polyps. Clin Gastroenterol Hepatol 2009; 7: 288–95.
- 16. van den Broek FJ, van Soest EJ, Naber AH, et al. Combining autofluorescence imaging and narrow-band imaging for the differentiation of adenomas from non-neoplastic colonic polyps among experienced and non-experienced endoscopists. Am J Gastroenterol 2009; 104: 1498–507.
- 17. Buchner AM, Shahid MW, Heckman MG, et al. Comparison of probe-based confocal laser endomicroscopy with virtual chromoendoscopy for classification of colon polyps. Gastroenterology 2010; 138: 834–42.
- Canales OM, Miyagui J, Takano, Poquioma E, Sano Y, Emura F. Usefulness of the Exera magnifying NBI system for differential diagnosis of small colorectal polyps using the Sano-Emura classification in Peru. Gastrointest Endosc 2011; 73 (suppl 1): AB438–39.
- 19. Chang C-C, Hsieh C-R, Lou H-Y et al. Comparative study of conventional colonoscopy, magnifying chromoendoscopy, and magnifying narrow-band imaging systems in the differential diagnosis of small colonic polyps between trainee and experienced endoscopist. Int J Colorectal Dis 2009; 24: 1413–19.
- 20. Chiu HM, Chang CY, Chen CC, et al. A prospective comparative study of narrow-band imaging, chromoendoscopy, and conventional colonoscopy in the diagnosis of colorectal neoplasia. Gut 2007; 56: 373–79.

- Coe SG, Thomas C, Crook J, Ussui V, Diehl N, Wallace MB. Colorectal surveillance interval assignment based on in vivo prediction of polyp histology: impact of endoscopic quality improvement program. Gastrointest Endosc 2012; 76: 118–25.
- 22. Dai J, Shen YF, Sano Y et al. Evaluation of narrow-band imaging in the diagnosis of colorectal lesions: is a learning curve involved? Dig Endosc 2013; 25: 180–88.
- East JE, Suzuki N, Bassett P, et al. Narrow band imaging with magnification for the characterization of small and diminutive colonic polyps: pit pattern and vascular pattern intensity. Endoscopy 2008; 40: 811–17.
- 24. Gross S, Trautwein C, Behrens A, et al. Computer-based classification of small colorectal polyps by using narrow-band imaging with optical magnification. Gastrointest Endosc 2011; 74: 1354–59.
- 25. Zervos X, Heller D, Sussman DA, Barkin JS. NBI: no biopsy indicated? Gastroenterology 2011; 140 (suppl 1): 719.
- Henry ZH, Yeaton P, Shami VM, et al. Meshed capillary vessels found on narrow-band imaging without optical magnification effectively identifies colorectal neoplasia: a North American validation of the Japanese experience. Gastrointest Endosc 2010; 72: 118–26.
- 27. Hewett DG, Huff man ME, Rex DK. Leaving distal colorectal hyperplastic polyps in place can be achieved with high accuracy by using narrow-band imaging: an observational study. Gastrointest Endosc 2012; 76: 374–80.
- 28. Hirata M, Tanaka S, Oka S, et al. Magnifying endoscopy with narrow band imaging for diagnosis of colorectal tumors. Gastrointest Endosc 2007; 65: 988–95.
- 29. Ignjatovic A, East JE, Guenther T, et al. What is the most reliable imaging modality for small colonic polyp characterization? Study of white-light, autofluorescence, and narrow-band imaging. Endoscopy 2011; 43: 94–99.
- Ignjatovic A, Thomas-Gibson S, East JE, et al. Development and validation of a training module on the use of narrow-band imaging in differentiation of small adenomas from hyperplastic colorectal polyps. Gastrointest Endosc 2011; 73: 128–33.
- 31. Kang HY, Kim YS, Kang SJ, et al. Comparison of narrow band imaging (NBI) and fujinon intelligent color enhancement (FICE) in predicting colon polyp histology during a screening colonoscopy: a prospective randomized study. Gastrointest Endosc 2012; 75 (suppl): AB323.
- 32. Katagiri A, Fu KI, Sano Y, et al. Narrow band imaging with magnifying colonoscopy as diagnostic tool for predicting histology of early colorectal neoplasia. Aliment Pharmacol Ther 2008; 27: 1269–74.
- 33. Kuiper T, van den Broek FJ, Naber AH, et al. Endoscopic trimodal imaging detects colonic neoplasia as well as standard video endoscopy. Gastroenterology 2011; 140: 1887–94.
- 34. Kuiper T, Marsman WA, Jansen JM, et al. Accuracy for optical diagnosis of small colorectal polyps in nonacademic settings. Clin Gastroenterol Hepatol 2012; 10: 1016–20.
- Ladabaum U, Fioritto A, Mitani A, et al. Real-time optical biopsy of colon polyps with narrow band imaging in community practice does not yet meet key thresholds for clinical decisions. Gastroenterology 2013; 144: 81–91.
- 36. Lau JY, Teo EK, Rerknimitr R, Singh R, Bourke MJ, Ng SC. An interim analysis of an Asia-Pacific multicenter randomized study comparing colonoscopy using a new high definition system in either white light or narrow band imaging in the detection of adenomas in subjects undergoing screening. Gastrointest Endosc 2012; 75 (suppl 1): AB173.
- 37. Lee CK, Lee SH, Hwangbo Y. Narrow-band imaging versus I-Scan for the real-time histological prediction of diminutive colonic polyps: a prospective comparative study by using the simple unified endoscopic classification. Gastrointest Endosc 2011; 74: 603–09.
- Lin TL, Chen CC, Fang YJ, Han ML, Lee JY, Wang HP. Combining autofluorescence imaging, chromoendoscopy, and narrow-band imaging with magnifying methods for the differentiation of neoplastic and non-neoplastic colonic polyps—a preliminary report. J Gastroenterol Hepatol 2011; 26: 111.
- Lopata A, Mentler E, Smith J, et al. Diagnostic accuracy comparing white light to narrow band imaging in identification of non-adenomatous and adenomatous polyps in a community-based setting. Am J Gastroenterol 2012; 107: S794.
- 40. Machida H, Sano Y, Hamamoto Y, et al. Narrow-band imaging in the diagnosis of colorectal mucosal lesions: a pilot study. Endoscopy 2004; 36: 1094–98.

- Occhipinti P, Saettone S, Quaglia A, Comi G, Calcara C, Broglia L. Accuracy of predicting polyps histology using narrow band imaging (NBI) with magnification in routine clinical practice. Gastrointest Endosc 2011; 73 (suppl 1): AB444.
- 42. Oka S, Tanaka S, Takata S, Kanao H, Chayama K. Clinical usefulness of narrow band imaging magnifying classification for colorectal tumors based on both surface pattern and microvessel features. Dig Endosc 2011; 23 (suppl 1): 101–05.
- 43. Paggi S, Rondonotti E, Amato A, et al. Resect and discard strategy in clinical practice: a prospective cohort study. Endoscopy 2012; 44: 899–904.
- 44. Patel SG, Rastogi A, Austin G, et al. Gastroenterology trainees can easily learn histologic characterization of diminutive colorectal polyps with narrow band imaging. Clin Gastroenterol Hepatol 2013; 11: 997–1003.
- 45. Pohl H, Bensen SP, Berk BS, et al. Real time diminutive polyp diagnosis accurately determines colonoscopy surveillance interval in clinical practice. Gastrointest Endosc 2012; 75 (suppl 1): AB151.
- 46. Ramirez-Ramirez MA, Sobrino-Cossio SR, Hernandez-Guerrero A, et al. Narrow band imaging versus conventional colonoscopy for detection of diminutive polyps: a randomized cross-over study with histopathological confirmation. Gastrointest Endosc 2009; 69 (suppl 1): AB292.
- Rastogi A, Bansal A, Wani S, et al. Narrow-band imaging colonoscopy—a pilot feasibility study for the detection of polyps and correlation of surface patterns with polyp histologic diagnosis. Gastrointest Endosc 2008; 76: 280–86.
- 48. Rastogi A, Keighley J, Singh V, et al. High accuracy of narrow band imaging without magnification for the real-time characterization of polyp histology and its comparison with high-definition white light colonoscopy: a prospective study. Am J Gastroenterol 2009; 104: 2422–30.
- 49. Rastogi A, Early D, Gupta N, et al. Randomized, controlled trial of standard-definition white light, highdefinition white light, and narrow-band imaging colonoscopy for the detection of colon polyps and prediction of polyp histology. Gastrointest Endosc 2011; 74: 593–602.
- 50. Rex DK. Narrow-band imaging without optical magnification for histologic analysis of colorectal polyps. Gastroenterology 2009; 136: 1174–81.
- 51. Rogart JN, Jain D, Siddiqui UD, et al. Narrow-band imaging without high magnification to differentiate polyps during real-time colonoscopy: improvement with experience. Gastrointest Endosc 2008; 68: 1136–45. 52 Rotondano G, Bianco MA, Sansone S, et al. Trimodal endoscopic imaging for the detection and differentiation of colorectal adenomas: a prospective single-centre clinical evaluation. Int J Colorectal Dis 2012; 27: 331–36.
- 52. Sato R, Fujiya M, Watari J, et al. The diagnostic accuracy of high-resolution endoscopy, autofluorescence imaging and narrow-band imaging for differentially diagnosing colon adenoma. Endoscopy 2011; 43: 862–68.
- 53. Sakamoto T, Matsuda T, Aoki T, Nakajima T, Saito Y. Time saving with narrow-band imaging for distinguishing between neoplastic and non-neoplastic small colorectal lesions. J Gastroenterol Hepatol 2012; 27: 351–55.
- 54. Sano Y, Ikematsu H, Fu KI, et al. Meshed capillary vessels by use of narrow-band imaging for differential diagnosis of small colorectal polyps. Gastrointest Endosc 2009; 69: 278–83.
- 55. Shahid MW, Buchner AM, Heckman MG, et al. Diagnostic accuracy of probe-based confocal laser endomicroscopy and narrow band imaging for small colorectal polyps: a feasibility study. Am J Gastroenterol 2012; 107: 231–39.
- 56. Sikka S, Ringold DA, Jonnalagadda S, Banerjee B. Comparison of white light and narrow band high definition images in predicting colon polyp histology, using standard colonoscopes without optical magnification. Endoscopy 2008; 40: 818–22.
- 57. Singh R, Nordeen N, Mei SL, Kaff es A, Tam W, Saito Y. West meets East: preliminary results of narrow band imaging with optical magnification in the diagnosis of colorectal lesions: a multicenter Australian study using the modifi ed Sano's classifi cation. Dig Endosc 2011; 23 (suppl 1): 126–30.
- Soto JR, Emura F, Bronstein M, et al. Meshed capillary vessels found on narrow-band imaging without optical magnification effectively identifies colorectal neoplasia: a pilot study in Venezuela. Gastrointest Endosc 2011; 73 (suppl 1): AB217.

- Su MY, Hsu CM, Ho YP, Chen PC, Lin CJ, Chiu CT. Comparative study of conventional colonoscopy, chromoendoscopy, and narrow-band imaging systems in differential diagnosis of neoplastic and non-neoplastic colonic polyps. Am J Gastroenterol 2006; 101: 2711–16. Articles 1346 www.thelancet.com/ oncology Vol 14 December 2013
- 60. Takemura Y, Yoshida S, Tanaka S, et al. Computer-aided system for predicting the histology of colorectal tumors by using narrow-band imaging magnifying colonoscopy (with video). Gastrointest Endosc 2012; 75: 179–85.
- 61. Tischendorf JJ, Wasmuth HE, Koch A, Hecker H, Trautwein C, Winograd R. Value of magnifying chromoendoscopy and narrow band imaging (NBI) in classifying colorectal polyps: a prospective controlled study. Endoscopy 2007; 39: 1092–96.
- 62. Tischendorf JJ, Schirin-Sokhan R, Streetz K, et al. Value of magnifying endoscopy in classifying colorectal polyps based on vascular pattern. Endoscopy 2010; 42: 22–27.
- 63. Valiante F, Bellumat A, De Bona M, et al. Histological characterization of small colon polyps using NBI magnification. Dig Liver Dis 2011; 43 (suppl 3): S222.
- 64. Wada Y, Kudo SE, Kashida H, et al. Diagnosis of colorectal lesions with the magnifying narrow-band imaging system. Gastrointest Endosc 2009; 70: 522–31.
- 65. Wang AY, Shami VM, Kahaleh M, et al. Presence of meshed capillary vessels on narrow-band imaging effectively identifies colorectal neoplasia without optical magnification: a North American validation of the Japanese experience. Gastrointest Endosc 2009; 69 (suppl 1): AB297.
- 66. Yague AS, Pereda T, Cid-Manas JI, Coniniga AG, Munoz CL, Cantos AS. High definition NBI without optical magnification for real-time characterization of small colon polyps: a prospective study compared to white light endoscopy. Gastrointest Endosc 2011; 73 (suppl 1): AB306.
- 67. Yoo HY, Lee MS, Ko BM, et al. Correlation of narrow band imaging with magnifying colonoscopy and histology in colorectal tumors. Clin Endosc 2011; 44: 44–50.
- 68. Zhou Q J, Yang JM, Fei BY, Xu QS, Wu WQ, Ruan HJ. Narrow-band imaging endoscopy with and without magnification in diagnosis of colorectal neoplasia. World J Gastroenterol 2011; 17: 666–70.
- 69. Chan JL, Lin L, Feiler M, Wolf AI, Cardona DM, Gellad ZF. Comparative effectiveness of i-SCAN and highdefinition white light characterizing small colonic polyps. World J Gastroenterol 2012; 18: 5905–11.
- Han ML, Lee YC, Chen CC, et al. Computer-generated surface and tone enhancements to distinguish neoplastic from non-neoplastic colon polyps less than 1 cm in diameter. Int J Colorectal Dis 2012; 27: 337–44.
- Hoff man A, Sar F, Goetz M, et al. High definition colonoscopy combined with i-Scan is superior in the detection of colorectal neoplasias compared with standard video colonoscopy: a prospective randomized controlled trial. Endoscopy 2010; 42: 827–33.
- 72. Hoff man A, Kagel C, Goetz M, et al. Recognition and characterization of small colonic neoplasia with high-definition colonoscopy using i-Scan is as precise as chromoendoscopy. Dig Liver Dis 2010; 42: 45–50.
- 73. Hong SN, Choe WH, Lee JH, et al. Prospective, randomized, back-to-back trial evaluating the usefulness of i-SCAN in screening colonoscopy. Gastrointest Endosc 2012; 75: 1011–21.
- 74. Neumann H, Vieth M, Fry LC, et al. Learning curve of virtual chromoendoscopy for the prediction of hyperplastic and adenomatous colorectal lesions: a prospective 2-center study. Gastrointest Endosc 2013; 78: 115–20.
- Pigo F, Bertani H, Manno M, et al. i-Scan high-definition white light endoscopy and colorectal polyps: prediction of histology, interobserver and intraobserver agreement. Int J Colorectal Dis 2013; 28: 399– 406.
- 76. Kim YS, Kim D, Chung SJ, et al. Differentiating small polyp histologies using real-time screening colonoscopy with Fuji Intelligent Color Enhancement. Clin Gastroenterol Hepatol 2011; 9: 744–49.
- 77. Liu YX, Huang LY, Bian XP, Cui J, Xu N, Wu CR. Fuji Intelligent Chromo Endoscopy and staining technique for the diagnosis of colon tumor. Chin Med J (Engl) 2008; 121: 977–82.
- 78. Longcroft-Wheaton GR, Higgins B, Bhandari P. Flexible spectral imaging color enhancement and indigo carmine in neoplasia diagnosis during colonoscopy: a large prospective UK series. Eur J Gastroenterol Hepatol 2011; 23: 903–11.

- 79. Longcroft-Wheaton G, Brown J, Cowlishaw D, Higgins B, Bhandari P. High-definition vs. standard-definition colonoscopy in the characterization of small colonic polyps: results from a randomized trial. Endoscopy 2012; 44: 905–10.
- 80. Pohl J, Nguyen-Tat M, Pech O, et al. Computed virtual chromoendoscopy for classifi cation of small colorectal lesions: a prospective comparative study. Am J Gastroenterol 2008; 103: 562–69.
- 81. Pohl J, Lotterer E, Balzer C, et al. Computed virtual chromoendoscopy versus standard colonoscopy with targeted indigocarmine chromoscopy: a randomised multicentre trial. Gut 2009; 58: 73–78.
- dos Santos CE, Pereira-Lima JC, Lopes CV, Malaman D, Parada AA, Salomao AD. Comparative study between MBI (FICE) and magnification chromoendoscopy with indigo carmine in the differential diagnosis of neoplastic and non-neoplastic lesions of the colorectum. Arq Gastroenterol 2009; 46: 111–15 (in Portuguese).
- dos Santos CE, Lima JC, Lopes CV, Pereira-Lima JC, Parada AA. Computerized virtual chromoendoscopy versus indigo carmine chromoendoscopy combined with magnification for diagnosis of small colorectal lesions: a randomized and prospective study. Eur J Gastroenterol Hepatol 2010; 22: 1364–71.
- 84. dos Santos CE, Malaman D, Lopes CV, et al. Digital chromoendoscopy for diagnosis of diminutive colorectal lesions. Diagn Ther Endosc 2012; 2012: 279521.
- Teixeira CR, Torresini RS, Canali C, et al. Endoscopic classification of the capillary-vessel pattern of colorectal lesions by spectral estimation technology and magnifying zoom imaging. Gastrointest Endosc 2009; 69: 750–56.
- Togashi K, Osawa H, Koinuma K, et al. A comparison of conventional endoscopy, chromoendoscopy, and the optimal-band imaging system for the differentiation of neoplastic and non-neoplastic colonic polyps. Gastrointest Endosc 2009; 69: 734–41.
- Yoshida N, Naito Y, Inada Y, et al. The detection of surface patterns by fl exible spectral imaging color enhancement without magnification for diagnosis of colorectal polyps. Int J Colorectal Dis 2012; 27: 605–11.
- McCallum AL, Jenkins JT, Gillen D, Molloy RG. Evaluation of autofluorescence colonoscopy for the detection and diagnosis of colonic polyps. Gastrointest Endosc 2008; 68: 283–90.
- 89. Mycek MA, Schomacker KT, Nishioka NS. Colonic polyp differentiation using time-resolved autofluorescence spectroscopy. Gastrointest Endosc 1998; 48: 390–94.
- 90. Nakaniwa N, Namihisa A, Ogihara T, et al. Newly developed autofluorescence imaging videoscope system for the detection of colonic neoplasms. Dig Endosc 2005; 17: 235–40.
- 91. Shao X, Zheng W, Huang Z. Near-infrared autofluorescence spectroscopy for in vivo identification of hyperplastic and adenomatous polyps in the colon. Biosens Bioelectron 2011; 30: 118–22.
- 92. Uedo N, Higashino K, Ishihara R, Takeuchi Y, Iishi H. Diagnosis of colonic adenomas by new autofluorescence imaging system: a pilot study. Dig Endosc 2004; 19 (suppl 1): S134–38.
- Andre B, Vercauteren T, Buchner AM, Krishna M, Ayache N, Wallace MB. Software for automated classification of probe-based confocal laser endomicroscopy videos of colorectal polyps. World J Gastroenterol 2012; 18: 5560–69.
- 94. Buchner AM, Gomez V, Heckman MG, et al. The learning curve of in vivo probe-based confocal laser endomicroscopy for prediction of colorectal neoplasia. Gastrointest Endosc 2011; 73: 556–60.
- 95. Gomez V, Buchner AM, Dekker E, et al. Interobserver agreement and accuracy among international experts with probe-based confocal laser endomicroscopy in predicting colorectal neoplasia. Endoscopy 2010; 42: 286–91.
- Hurlstone DP, Baraza W, Brown S, Thomson M, Tiffin N, Cross SS. In vivo real-time confocal laser scanning endomicroscopic colonoscopy for the detection and characterization of colorectal neoplasia. Br J Surg 2008; 95: 636–45.
- 97. Kiesslich R, Burg J, Vieth M, et al. Confocal laser endoscopy for diagnosing intraepithelial neoplasias and colorectal cancer in vivo. Gastroenterology 2004; 127: 706–13.
- 98. De Palma GD, Staibano S, Siciliano S, et al. In vivo characterisation of superficial colorectal neoplastic lesions with high-resolution probe-based confocal laser endomicroscopy in combination with video-mosaicing: a feasibility study to enhance routine endoscopy. Dig Liver Dis 2010; 42: 791–97.
- 99. Sanduleanu S, Driessen A, Gomez-Garcia E, Hameeteman W, De Bruine A, Masclee A. In vivo diagnosis and classification of colorectal neoplasia by chromoendoscopy-guided confocal laser endomicroscopy. Clin Gastroenterol Hepatol 2010; 8: 371–78.

- Shahid MW, Buchner AM, Raimondo M, Woodward TA, Krishna M, Wallace MB. Accuracy of real-time vs blinded offline diagnosis of neoplastic colorectal polyps using probe-based confocal laser endomicroscopy: a pilot study. Endoscopy 2012; 44: 343–48.
- 101. Singson Z, Hashemzadeh M, Mazen JM. Utilization of probe-based confocal laser endomicroscopy in a resect and discard approach to small colon polyps. Gastrointest Endosc 2012; 75 (suppl 1): AB224.
- 102. Xie XJ, Li CQ, Zuo XL, et al. Differentiation of colonic polyps by confocal laser endomicroscopy. Endoscopy 2011; 43: 87–93.
- 103. Schachschal G, Mayr M, Treszl A, et al. Endoscopic versus histological characterisation of polyps during screening colonoscopy. Gut 2013; published online June 28. DOI:10.1136/ gutjnl-2013-304562.
- 104. Gupta N, Kaltenbach T, Sato T, et al. Diagnosis time determines the accuracy of optical diagnosis of diminutive polyp histology. Gastrointest Endosc 2013; 77 (suppl 1): AB553–54.
- 105. Trikalinos TA, Hoaglin DC, Small KM, Schmid CH. Evaluating practices and developing tools for comparative effectiveness reviews of diagnostic test accuracy: methods for the joint meta-analysis of multiple tests. January, 2013. http://www.ncbi.nlm.nih. gov/books/NBK148804/ (accessed Oct 18, 2013).
- 106. Ransohoff DF, Feinstein AR. Problems of spectrum bias and bias in evaluating the efficacy of diagnostic test. N Eng J Med 1978; 299: 926–30.